

Article

Design and Evaluation of Ambiphilic Aryl Thiol-Iminium-Based Molecules for Organocatalyzed Thioacyl Aminolysis

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improvements and directions toward cysteine-independent organocatalyzed native chemical ligation are discussed.

1. INTRODUCTION

Over the past several decades, advances in chemical protein synthesis (CPS) have enabled the evaluation of uniquely synthetic proteins among those accessible by traditional biochemical and recombinant technologies.¹ As the applicability of CPS methods broadens, especially due to growing interest in mirror-image and site-specifically modified proteins that are exclusive to chemical synthesis, there is an aligned need for the discovery and development of orthogonal peptide ligation strategies.² These advances should be chemoselective, providing alternative retrosynthetic ligation disconnects aimed at improving the overall efficiency of CPS.³ The ubiquity of native chemical ligation (NCL)⁴ and Ala-ligation methods⁵ used in CPS endeavors has proven their robustness, yet residue-specific limitations (i.e., the need for Cys or Ala residues) have motivated innovative developments.^{3b,6,7} A few recent NCL-inspired technologies that have enabled Cysindependent access to proteins with total atomic control include the Staudinger,⁸ α -keto acid-hydroxylamine (KAHA),⁹ and Ser/Thr ligations.¹⁰ Auxiliary-based strategies,⁶ including aldehyde-capture ligation (ACL)¹¹ and related methods,^{12,13} as well as N-terminal auxiliary ligations^{14,15} have significantly extended the scope of NCL. Although enabling, such strategies can be limited to residue-specific ligation junctions, may rely on the synthesis of reactive C- and N-terminal auxiliaries, and furthermore can involve postligation modification steps, restricting their overall generality to skilled practitioners.

studies demonstrate the ability of this designed organocatalyst to

deliver critical intermediates capable of undergoing these individual reactions necessary for the proposed process. Future design

To streamline protein retrosynthesis and develop tools to improve CPS user access, we hypothesized that a rationally designed ambiphilic organocatalyst combining nucleophilic (e.g., thiol)^{4,14,15} and electrophilic (e.g., aldehyde, iminium)^{11-13,16-18} functionalities could catalyze an aqueous thioester aminolysis reaction between solid-phase peptide synthesis accessible peptide partners (Figure 1a).¹⁹ Inspired by the mechanisms of NCL and auxiliary-based ligation reactions,^{4,6,20} we envisioned an organocatalytic process to achieve Cys-independent thioacyl aminolysis between any Cterminal thioester peptide-1 and N-terminal peptide-2. Overall, the ambiphilic organocatalyst could enable transthioesterification, amine capture, and S-to-N acyl transfer events to form a ligated peptide.

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synthesis and characterized reactivity

In a recent report, we demonstrated the dynamic properties of a series of electronically perturbed cyclic *N*,*S*-acetals (Figure 1b).²⁰ Under acidic conditions 1 undergoes C1-S bond ionization to form a ring-opened zwitterionic intermediate bearing aryl thiol(ate) and benzylic iminium functionalities that satisfied our ambiphilic organocatalyst design features. Due to the efficient synthetic access to these cyclic N,S-acetals and promising early observations that suggested transthioesterification and amine capture events could occur in a dynamic process, we explored their competency in model organocatalyzed thioacyl aminolysis reactions (Figure 1). Unfortunately, despite extensive efforts with 1, we were unable to achieve organocatalyzed thioacyl aminolysis reactions. We hypothesized that the process inefficiency may be due in part to the poor water solubility of 1. Furthermore, the proposed catalytic cycle may be hindered by the requirement for an eight-

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membered tetrahedral intermediate, which may be unfavorable for efficient S-to-N acyl transfer events.^{6b,21} Preliminary observations from the evaluation of cyclic N,S-acetals informed key features to include in a revised organocatalyst design (Figure 1c). Ongoing efforts have focused on an organocatalyst design that might exhibit improved aqueous solubility due to greater ionic character while preferentially enabling S-to-N acyl transfer events via a more favorable six-membered tetrahedral intermediate. We therefore envisioned the design and synthesis of aryl thiol–iminium-based organocatalyst **2a** (Figure 1c). We find that 2a exists in dynamic equilibrium with its dimeric form (2b). Guided by our knowledge of the dynamic reactivity of N,S-acetals, we herein describe our design logic, synthesis, and evaluation efforts toward the development of an aryl thioliminium-based ambiphile for organocatalyzed thioacyl aminolysis.

2. RESULTS AND DISCUSSION

2.1. Reactivity of Cyclic N,S-Acetals from Initial Thioacyl Aminolysis Studies. To better understand the reactivity of aryl thiol-iminium-based ambiphiles as designed

organocatalysts for NCL, we directed our efforts to evaluate the reactions of 1 with model peptide thioesters. The N,Sacetal moiety of 1 provides the latent nucleophilic thiol and electrophilic iminium¹⁸ functionalities required for respective transthioesterification and amine capture events (Figure 1a). Ideally, once both peptide fragments are captured, an S-to-N acyl transfer would occur to complete the peptide ligation process. Interestingly, we discovered that 1 incorporated several dynamic properties that allowed us to develop and evaluate its ability to undergo transthioesterification (Figure 2). The dynamic reactivity observed in our first-generation N,S-acetals includes ring-opening and ring-closing behavior.²⁰ The treatment of 1 in excess CF₃CO₂D resulted in rapid and quantitative C1-S bond ionization, enabling interchangeable ring-opened and ring-closed forms under equilibrium control by titration with triethylamine (Figure 2a). With an operationally reversible system in hand, we proposed that in situ formed ring-opened species (e.g., 3) could participate in transthioesterification reactions with model peptide thioesters (4), followed by a subsequent amine capture and S-to-N acyl transfer event to achieve an overall organocatalyzed thioacyl aminolysis process (Figure 2b). Despite the observed production of transthioesterification intermediates such as adduct-1 (5), these intermediates do not appear to undergo efficient amine capture events. Therefore, informed by shortcomings with 1, we set out to prepare revised organocatalyst 2a (Scheme 1).

2.2. Synthesis of Organocatalyst 2a and Its Corresponding N,S-Acetal Dimer 2b. In the design stage, we reasoned that improving the likelihood of the amine capture step would be integral to balance the relative rates of transthioesterification, amine capture, and S-to-N acyl transfer events. In NCL reactions the transthioesterification step is ratelimiting, whereas in N-terminal auxiliary-based NCLs, the ratelimiting step is the S-to-N acyl transfer event.²² We presume that the tethered nature of the thiol in 1, which predominately exists in the ring-closed state, precludes amine capture events. Building from this evaluation, we revised our synthetic efforts to design a system that would avoid trapping by an intramolecular thiol and improve the likelihood of S-to-N acyl transfer events via a hypothesized six-membered tetrahedral intermediate. These informed observations resulted in the design of the second-generation organocatalyst 2a and its corresponding N,S-acetal dimer 2b.

We developed an efficient route to prepare organocatalyst 2a in six steps from commercially available 8-bromoisoquinoline (Scheme 1). We found that the proposed monomeric form 2a exists in equilibrium with the functional N,S-acetal dimer 2b. The synthesis begins with the chemoselective reduction of 8bromoisoquinoline 6 upon treatment with sodium cyanoborohydride in the presence of boron trifluoride diethyl etherate to yield 8-bromo-1,2,3,4-tetrahydroisoquinoline 7.23 The resultant amine 7 is arylated with phenylboronic acid using Chan-Lam coupling conditions to provide 8-bromo-2-phenyl-1,2,3,4-tetrahydroisoquinoline 8 in 39% yield over two steps.²⁴ Initial observations suggest the N-phenyl tertiary amine 8 is susceptible to aerobic benzylic oxidation under ambient conditions. Therefore, a careful two-step procedure to convert aryl bromide 8 into dithiocarbamate 10 was developed. Using Buchwald's copper-catalyzed halogen exchange chemistry,² the aryl bromide 8 can be efficiently converted into the requisite aryl iodide 9 in 92% isolated yield. This aryl iodide (9) provides a handle to examine conditions for the installation

Scheme 1. Synthesis of a Functional Aryl Thiol-iminium Organocatalyst: (a) Synthesis Route toward an Organocatalyst Precursor; (b) Equilibration Enabling Access to an N,S-Acetal Dimer (2b) and an Alternative Disulfide-Based Oxidized Dimer (14)



a) dynamic reactivity: ring-opening and ring-closing via C1–S bond ionization



Figure 2. Cyclic *N*,*S*-acetals are dynamic in solution and reactive toward thioesters: (a) C1-S bond ionization of 1; (2) addition steps. Transthioesterification events occur, but amine capture events are not observable.

observab

for details

of a sulfur-containing functionality at the 8-position. We found that 9 can be successfully converted into dithiocarbamate 10 in 49% yield using tetramethylthiuram disulfide and zinc in the presence of catalytic copper(II) chloride.²⁶ Notably, these reductive conditions prevent any undesired aerobic benzylic oxidation from occurring prematurely. Controlled benzylic oxidation (C1) using diethyl azodicarboxylate (DEAD) converts 10 into *N*,*O*-acetal 11.²⁷ This benzylic oxidation

may proceed via iminium ion formation followed by trapping with methanol. Optimization efforts led to the development of an efficient procedure to convert 10 into 11 in 69% isolated yield. The use of DEAD to affect this oxidation is superior to other oxidation chemistries that were specifically developed for 2-aryl-1,2,3,4-tetrahydroisoquinoline systems. The crystallization of 11 in 2-propanol produced derivative 12, which was suitable for structure determination using X-ray diffraction (CCDC 2222419).²⁸ This suggests that 11 readily engages in a dynamic equilibrium exchange process, via the intermediate iminium ion, to provide 12. Next, we evaluated a variety of conditions to cleave the dithiocarbamate moiety (11) and anticipated the isolation of aryl thiol 13. We found that it was difficult to purify the cleavage byproducts away from presumed product 13. We hypothesized using ethylenediamine would facilitate purification, as the cleavage byproducts would be acyclic or cyclic thioureas. Upon workup, we observed both the acyclic and cyclic thiourea byproducts, and their polarity differences allowed us to routinely isolate a stable product, first presumed to be 13, in 22% yield by column chromatography. A thorough characterization of the stable product revealed its constitution to be most consistent with an *N*,*S*-acetal dimer **2b**, which we presume to be derived via the intermediacy of reactive monomer 2a (Scheme 1b). The presumed aryl thiol 13 is not observed and likely converts into 2b via reactive monomer 2a.

2.3. Evaluation of *N*,*S*-Acetal Dimer 2b in a Dipeptide-Forming Model System. Intrigued by the preference for a dimeric state, we further studied the equilibria of *N*,*S*-acetal 2b in aqueous solution (Scheme 1b). Under aqueous acidic conditions $(3:1 \text{ CH}_3\text{CN}:\text{H}_2\text{O}, 0.1\%)$

 CF_3CO_2H), the monomeric **2a** and dimeric **2b** forms likely exist in equilibrium and are mostly indistinguishable by standard ESI mass spectrometry due to a similar dissociation to their preferred iminium states. However, ¹H and ¹³C NMR spectroscopic data are most consistent with *N*,*S*-acetal dimer **2b**. The prolonged exposure of **2b** under aqueous conditions leads to the formation of an oxidized species that we assign to disulfide dimer **14**. We reason that **14** could be used as a precatalyst for organocatalyzed thioacyl aminolysis reactions in the presence of a suitable disulfide reductant (e.g., tris(2carboxyethyl)phosphine). However, we find **2b** to be a more convenient, latent form of the designed reactive organocatalyst **2a**.

To elucidate the reactivity of 2a, we combined 2b with a single residue thioester Ac-Ala-SPh (15) (Figure 3a). An adduct, 16 ($t_r = 4.2 \text{ min}$), consistent with transthioesterification is observable by UPLC-MS analysis. Encouraged by this, we added H-Gly-OMe (4 equiv) and allowed the aqueous mixture to react for 18 h. Analysis by UPLC-MS showed three distinct intermediates that we tentatively assign as N_iN-acetal adducts (17a,b) (Figure 3b). These adducts are observed in iminium ion forms with different retention times (17a, $t_r = 3.9$ min; 17b, $t_r = 4.0$ min; see the Supporting Information). Their distinct elution times suggest that each adduct is a unique N,Nacetal (17a,b), where attack of 16 by H-Gly-OMe yields diastereoisomers 17a,b. We note that attack by water (X = OH) or thiophenol (X = SPh) may yield other possible adducts (17c). While the preference for ionization of 17a, b to the common iminium ion (16) masks their structural identity, this result is consistent with our previous studies using 1 and supports the efficiency of the transthioesterification event.

To better understand the presumed formation of the two amine capture products 17a,b, we considered an earlier spectroscopic observation where *N*,*O*-acetal 11 exhibits solvent-dependent interconversion of anti- and synperiplanar diastereomeric forms in chloroform- d_1 . Interestingly, the ¹H



b) proposed nucleophile capture N,X-acetals (17):

all N,X-acetal adducts are observed as iminium ions by UPLC-MS analysis



Figure 3. N,S-acetal dimer (2b) reacts with thioesters to form adduct-1 (16) and adduct-2 (17a,b).

NMR spectrum of **11** in benzene- d_6 shows a preference for the one diastereomeric form. This type of structural dynamicity via C1–O bond ionization is akin to similar observations made for cyclic *N*,*S*-acetal **1**, where interconversion occurs via C1–S bond ionization.²⁰ Taken together, we assign **17a**,**b** as diastereomeric Gly-OMe adducts and assign the third peak to the iminium ion intermediate **16**. The poor solubility of **2b** under buffered aqueous conditions limited our ability to quantify this complex process. Further studies are needed to understand the interconversion of **17a**,**b** as well as the implications of stereochemistry on the efficiency of productive *N*,*S*-acyl transfer events. We anticipate that solvent-dependent effects will play a critical role in tuning the reactivity of **2b**, as well as downstream reactivities of adduct-1 and adduct-2 type intermediates.

2.4. Evaluation of *N*,*S*-Acetal Dimer 2b in a Peptide-Forming Model System. Analytical and solubility complications with the dipeptide system led us to evaluate a more compatible model system using 2a, peptide thioester 18, and benzyl amine (Figure 4a). These experiments allowed us to





Figure 4. *N*,*S*-acetal dimer (2b) reacts with thioester peptide (18) to form adduct-1 (19) and adduct-2 (20).

observe transthioesterifcation and amine capture events. The reaction of 2b and Ac-LYRAG-SPh (18) yields transthioesterification product 19 in good conversion. We added an excess of benzylamine to this reaction mixture and observed a ternary adduct (20) that is consistent with a stepwise process involving transthioesterification followed by amine capture. The adduct (20) was characterized using UPLC-MS (Figure 4b). Again, TIC extraction (m/z 475) shows two peaks with distinct elution times (20a, $t_r = 3.1$ min; 20b, $t_r = 3.2$ min; see the Supporting Information), which we attribute to the two diastereomeric benzyl amine adducts. When examining the MS data for each unique peak, we observe four ions (Figure 4b). Two ions correspond to the $[M + 2H^+]^{2+}$ and $[M + H^+]^+$ ions of 20. Interestingly, the other ions correspond to ligated peptide 21 ($[M + H^+]^+$) and monomer 2a ($[M^+]^+$). It is unusual to detect the product (21) and monomer (2a) in this region ($t_r \approx 3.2 \text{ min}$), as they are respectively found at $t_r = 0.7$ min (21) and $t_r = 4.4 \min (2a)$. Therefore, we propose that the ionization of ternary adduct 20 produces 21 and regenerates monomer 2a. Despite the encouraging observation of these adducts, organocatalyzed and organopromoted experiments using 2b did not show an indication of improved reactivity when compared with the background aminolysis experiments without 2b.²⁹ Ongoing efforts are focused on the development of improved analytical methods to better characterize stepwise events toward organocatalyzed NCL.

3. CONCLUSIONS

Our objective to understand the reactivity of N,S-acetals for use in organocatalyzed thioacyl aminolysis has led to several discoveries. These underexplored ambiphilic molecules are dynamic in solution and exhibit the ability to reversibly exchange adducts via C1-S bond ionization to produce observable reactive benzylic iminium ion intermediates. With our first-generation design (1) we characterized dynamic C1-S bond ionization and transthioesterification reactivity. However, amine capture and N-to-S acyl transfer events remained elusive, leading us to prepare the dimeric N,S-acetal system 2b that can undergo transthioesterification and amine capture reactions via the monomeric intermediate 2a. The transthioesterification process using 2b is more efficient than the corresponding reactivity of 1. While amine adducts at C1 are observable and are supported by an amine capture experiment using benzylamine, they do not appear to be the predominate species under the evaluated conditions and are elusive when using amino ester model systems under buffered conditions.^{30,31} Although further development of conditions and organocatalyst refinement are needed, we are encouraged by the initial reactivity of these novel N,S-acetal-based systems toward realizing organocatalyzed thioacyl aminolysis procedures.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acsomega.2c07586.

Materials and methods, general experimental procedures, characterization data and spectra for associated molecules, and reaction analyses (PDF)

Crystallographic data for compound 12 (CIF)

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Notes

The authors declare no competing financial interest.

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REFERENCES

(1) For selected examples, see: (a) Werner, H. M.; Cabalteja, C. C.; Horne, W. S. Peptide Backbone Composition and Protease Susceptibility: Impact of Modification Type, Position, and Tandem Substitution. *ChemBioChem.* **2016**, *17*, 712–718. (b) Stephens, O. M.; Kim, S.; Welch, B. D.; Hodsdon, M. E.; Kay, M. S.; Schepartz, A. Inhibiting HIV fusion with a beta-peptide foldamer. *J. Am. Chem. Soc.* **2005**, *127*, 13126–13127. (c) Noguchi, T.; Ishiba, H.; Honda, K.; Kondoh, Y.; Osada, H.; Ohno, H.; Fujii, N.; Oishi, S. Synthesis of Grb2 SH2 Domain Proteins for Mirror-Image Screening Systems. *Bioconjugate Chem.* **2017**, *28*, 609–619.

(2) For some notable orthogonal ligation strategies, see selected examples: (a) Nilsson, B. L.; Hondal, R. J.; Soellner, M. B.; Raines, R.

(3) (a) Isidro-Llobet, A.; Kenworthy, M. N.; Mukherjee, S.; Kopach, M. E.; Wegner, K.; Gallou, F.; Smith, A. G.; Roschangar, F. Sustainability Challenges in Peptide Synthesis and Purification: From R&D to Production. J. Org. Chem. 2019, 84, 4615-4628.
(b) Agouridas, V.; El Mahdi, O.; Cargoët, M.; Melnyk, O. A statistical view of protein chemical synthesis using NCL and extended methodologies. Bioorg. Med. Chem. 2017, 25, 4938-4945.

(4) (a) Dawson, P. E.; Muir, T. W.; Clark-Lewis, I.; Kent, S. B. Synthesis of proteins by native chemical ligation. *Science* **1994**, *266*, 776–779. (b) Hackeng, T. M.; Griffin, J. H.; Dawson, P. E. Protein synthesis by native chemical ligation: expanded scope by using straightforward methodology. *Proc. Natl. Acad. Sci. U.S.A.* **1999**, *96*, 10068–10073.

(5) (a) Yan, L. Z.; Dawson, P. E. Synthesis of peptides and proteins without cysteine residues by native chemical ligation combined with desulfurization. *J. Am. Chem. Soc.* 2001, 123, 526–533. (b) Wan, Q.; Danishefsky, S. J. Free-radical-based, specific desulfurization of cysteine: a powerful advance in the synthesis of polypeptides and glycopolypeptides. *Angew. Chem., Int. Ed. Engl.* 2007, 46, 9248–9252.

(6) For selected reviews, see: Chow, Y.; Li, X. Development of thiolindependent peptide ligations for protein chemical synthesis. Tetrahedron Lett. 2015, 56, 3715-3720. (b) Burke, H. M.; McSweeney, L.; Scanlan, E. M. Exploring chemoselective S-to-N acyl transfer reactions in synthesis and chemical biology. Nat. Commun. 2017, 8, 15655. (c) Yang, J.; Zhao, J. Recent developments in peptide ligation independent of amino acid side-chain functional group. Sci. China: Chem. 2018, 61, 97-112. (d) Agouridas, V.; El Mahdi, O.; Diemer, V.; Cargoët, M.; Monbaliu, J.-C. M.; Melnyk, O. Native Chemical Ligation and Extended Methods: Mechanisms, Catalysis, Scope, and Limitations. Chem. Rev. 2019, 119, 7328-7443. (e) Pattabiraman, V. R.; Ogunkoya, A. O.; Bode, J. W. Amide-Forming Ligation Reactions. Organic Reactions 2019, 231-592. (f) Giesler, R. J.; Erickson, P. W.; Kay, M. S. Enhancing native chemical ligation for challenging chemical protein syntheses. Curr. Opin. Chem. Biol. 2020, 58, 37-44.

(7) Schmidt, M.; Toplak, A.; Quaedflieg, P. J.; Nuijens, T. Enzymemediated ligation technologies for peptides and proteins. *Curr. Opin. Chem. Biol.* **2017**, *38*, 1–7.

(8) (a) Nilsson, B. L.; Kiessling, L. L.; Raines, R. T. Staudinger ligation: a peptide from a thioester and azide. *Org. Lett.* **2000**, *2*, 1939–1941. (b) Saxon, E.; Armstrong, J. I.; Bertozzi, C. R. A "traceless" Staudinger ligation for the chemoselective synthesis of amide bonds. *Org. Lett.* **2000**, *2*, 2141–2143.

(9) Bode, J. W. Chemical Protein Synthesis with the alpha-Ketoacid-Hydroxylamine Ligation. *Acc. Chem. Res.* **201**7, *50*, 2104–2115.

(10) Lee, C. L.; Li, X. Serine/threonine ligation for the chemical synthesis of proteins. *Curr. Opin. Chem. Biol.* **2014**, *22*, 108–114.

(11) Raj, M.; Wu, H.; Blosser, S. L.; Vittoria, M. A.; Arora, P. S. Aldehyde capture ligation for synthesis of native peptide bonds. *J. Am. Chem. Soc.* **2015**, *137*, 6932–6940.

(12) Tung, C. L.; Wong, C. T. T.; Li, X. Peptide 2-formylthiophenol esters do not proceed through a Ser/Thr ligation pathway, but participate in a peptide aminolysis to enable peptide condensation and cyclization. *Org. Biomol. Chem.* **2015**, *13*, 6922–6926.

(13) Fouché, M.; Masse, F.; Roth, H.-J. Hydroxymethyl Salicylaldehyde Auxiliary for a Glycine-Dependent Amide-Forming Ligation. *Org. Lett.* **2015**, *17*, 4936–4939.

(14) Offer, J. Native chemical ligation with N α acyl transfer auxiliaries. *Pept. Sci.* **2010**, *94*, 530–541.

(15) For the most recent and general N-terminal-based ligation auxiliaries, see: Trunschke, S.; Piemontese, E.; Fuchs, O.; Abboud, S.; Seitz, O. Enhancing Auxiliary-Mediated Native Chemical Ligation at Challenging Junctions with Pyridine Scaffolds. *Chem. - Eur. J.* 2022, 28, No. e202202065, and references cited therein.

(16) Kemp, D. S. The amine capture strategy for peptide bond formation—an outline of progress. *Biopolymers* **1981**, *20*, 1793–1804.

(17) (a) Liu, C.-F.; Tam, J. P. Chemical Ligation Approach To Form a Peptide Bond between Unprotected Peptide Segments. Concept and Model Study. J. Am. Chem. Soc. **1994**, 116, 4149-4153. (b) Liu, C.-F.; Tam, J. P. Peptide segment ligation strategy without use of protecting groups. Proc. Natl. Acad. Sci. U.S.A. **1994**, 91, 6584-6588. (18) Leleu, S.; Penhoat, M.; Bouet, A.; Dupas, G.; Papamicaël, C.; Marsais, F.; Levacher, V. Amine Capture Strategy for Peptide Bond Formation by Means of Quinolinium Thioester Salts. J. Am. Chem. Soc. **2005**, 127, 15668-15669.

(19) Related approaches for peptide bond formation: (a) Wu, H.; Handoko; Raj, M.; Arora, P. S. Iterative Design of a Biomimetic Catalyst for Amino Acid Thioester Condensation. Org. Lett. 2017, 19, 5122-5125. (b) Handoko; Satishkumar, S.; Panigrahi, N. R.; Arora, P. S. Rational Design of an Organocatalyst for Peptide Bond Formation. J. Am. Chem. Soc. 2019, 141, 15977-15985. (c) Handoko; Panigrahi, N. R.; Arora, P. S. Two-Component Redox Organocatalyst for Peptide Bond Formation. J. Am. Chem. Soc. 2022, 144, 3637-3643.

(20) Kirkeby, E. K.; Roberts, A. G. Design, synthesis and characterization of structurally dynamic cyclic N,S-acetals. *Chem. Commun.* 2020, *56*, 9118–9121.

(21) Monbaliu, J. C.; Dive, G.; Stevens, C. V.; Katritzky, A. R. Governing Parameters of Long-Range Intramolecular S-to-N Acyl Transfers within (S)-Acyl Isopeptides. *J. Chem. Theory. Comput.* **2013**, *9*, 927–934.

(22) Wang, C.; Guo, Q. X.; Fu, Y. Theoretical analysis of the detailed mechanism of native chemical ligation reactions. *Chem. Asian. J.* **2011**, *6*, 1241–1251.

(23) Gribble, G. W.; Heald, P. W. Reactions of Sodium-Borohydride in Acidic Media III. Reduction and Alkylation of Quinoline and Isoquinoline with Carboxylic-Acids. *Synthesis (Stuttg)*. **1975**, *1975*, 650–652.

(24) (a) Derosa, J.; O'Duill, M. L.; Holcomb, M.; Boulous, M. N.; Patman, R. L.; Wang, F.; Tran-Dube, M.; McAlpine, I.; Engle, K. M. Copper-Catalyzed Chan-Lam Cyclopropylation of Phenols and Azaheterocycles. J. Org. Chem. 2018, 83, 3417–3425. (b) Vantourout, J. C.; Miras, H. N.; Isidro-Llobet, A.; Sproules, S.; Watson, A. J. Spectroscopic Studies of the Chan-Lam Amination: A Mechanism-Inspired Solution to Boronic Ester Reactivity. J. Am. Chem. Soc. 2017, 139, 4769–4779.

(25) Klapars, A.; Buchwald, S. L. Copper-catalyzed halogen exchange in aryl halides: an aromatic Finkelstein reaction. *J. Am. Chem. Soc.* **2002**, *124*, 14844–14845.

(26) Wu, X. M.; Yan, G. B. Copper-Catalyzed Synthesis of S-Aryl Dithiocarbamates from Tetraalkylthiuram Disulfides and Aryl Iodides in Water. *Synlett* **2019**, *30*, 610–614.

(27) Suga, T.; Iizuka, S.; Akiyama, T. Versatile and highly efficient oxidative C(sp(3))-H bond functionalization of tetrahydroisoquinoline promoted by bifunctional diethyl azodicarboxylate (DEAD): scope and mechanistic insights. *Org. Chem. Front.* **2016**, *3*, 1259–1264.

(28) Cambridge Crystallographic Data Centre (CCDC) accession number for compound 12: CCDC 2222419.

(29) Some unhindered thioacyl aminolysis reactions can proceed uncatalyzed, and intramolecular thioacyl aminolysis reactions can be efficient. See: (a) Payne, R. J.; Ficht, S.; Greenberg, W. A.; Wong, C.-H. Angew. Chem., Int. Ed. 2008, 47, 4411-4415. (b) Li, Y.; Yongye, A.; Giulianotti, M.; Martinez-Mayorga, K.; Yu, Y.; Houghten, R. A. J. Comb. Chem. 2009, 11, 1066-1072.

(30) Additional studies with substrates bearing lysine ε -amino sidechains are needed to establish *N*-terminal amine vs lysine ε -amine selectivity. We propose that *N*-terminal selective thioacyl aminolysis reactions may be possible based on pK differences, where on average *N*-terminal amines (pK = 7.7 ± 0.5) have lower values than lysine ε -amines (pK = 10.5 ± 1.1). See: Grimsley, G. R.; Scholtz, J. M.; Pace,

C. N. A summary of the measured pK values of the ionizable groups in folded proteins. *Protein Sci.* 2008, *18*, 247–251.

(31) In a related process involving intermediate imine formation, Chou and co-workers demonstrated that the *N*-terminal amine of peptides and proteins can be modified at pH = 6.1 with excellent selectivity of >99/1 α -amino/ ε -amino. See: Chen, D.; Disotaur, M. M.; Xiong, X.; Wang, Y.; Chou, D. H.-C. Selective N-terminal functionalization of native peptides and proteins. *Chem. Sci.* **2017**, *8*, 2717–2722.