

Supplementary Material

Three aphid-transmitted viruses encourage vector migration from infected common bean (*Phaseolus vulgaris*) plants through a combination of volatile and surface cues

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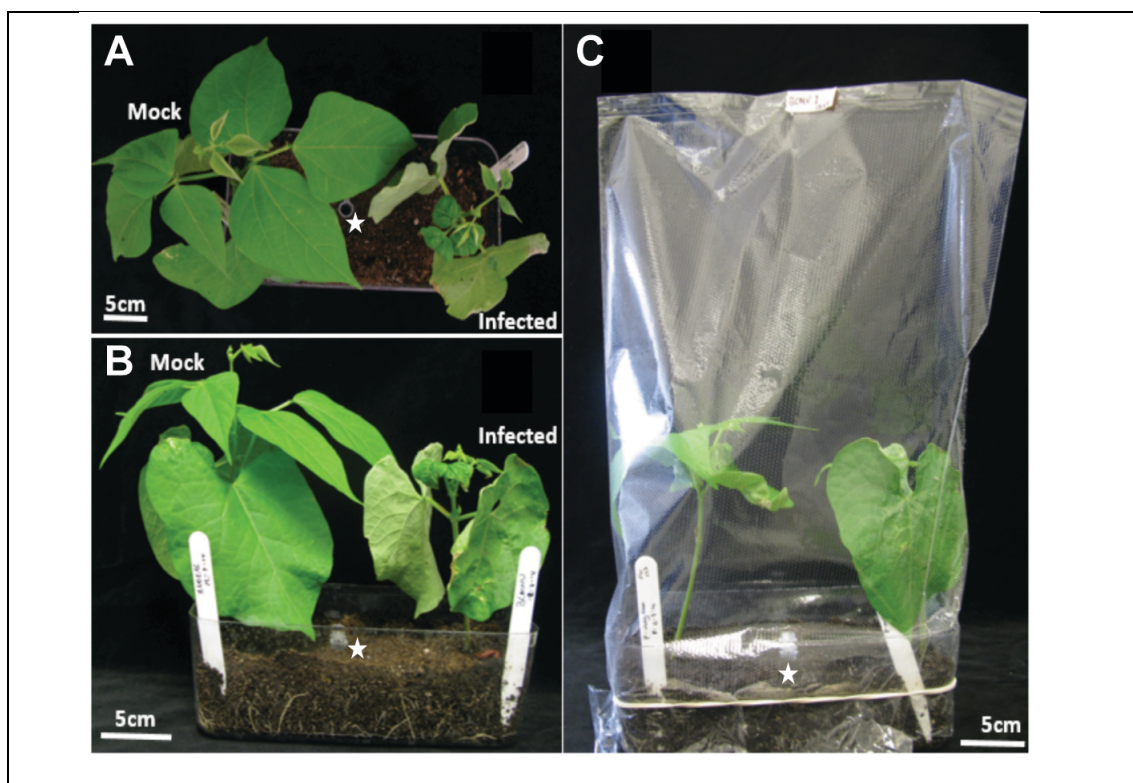
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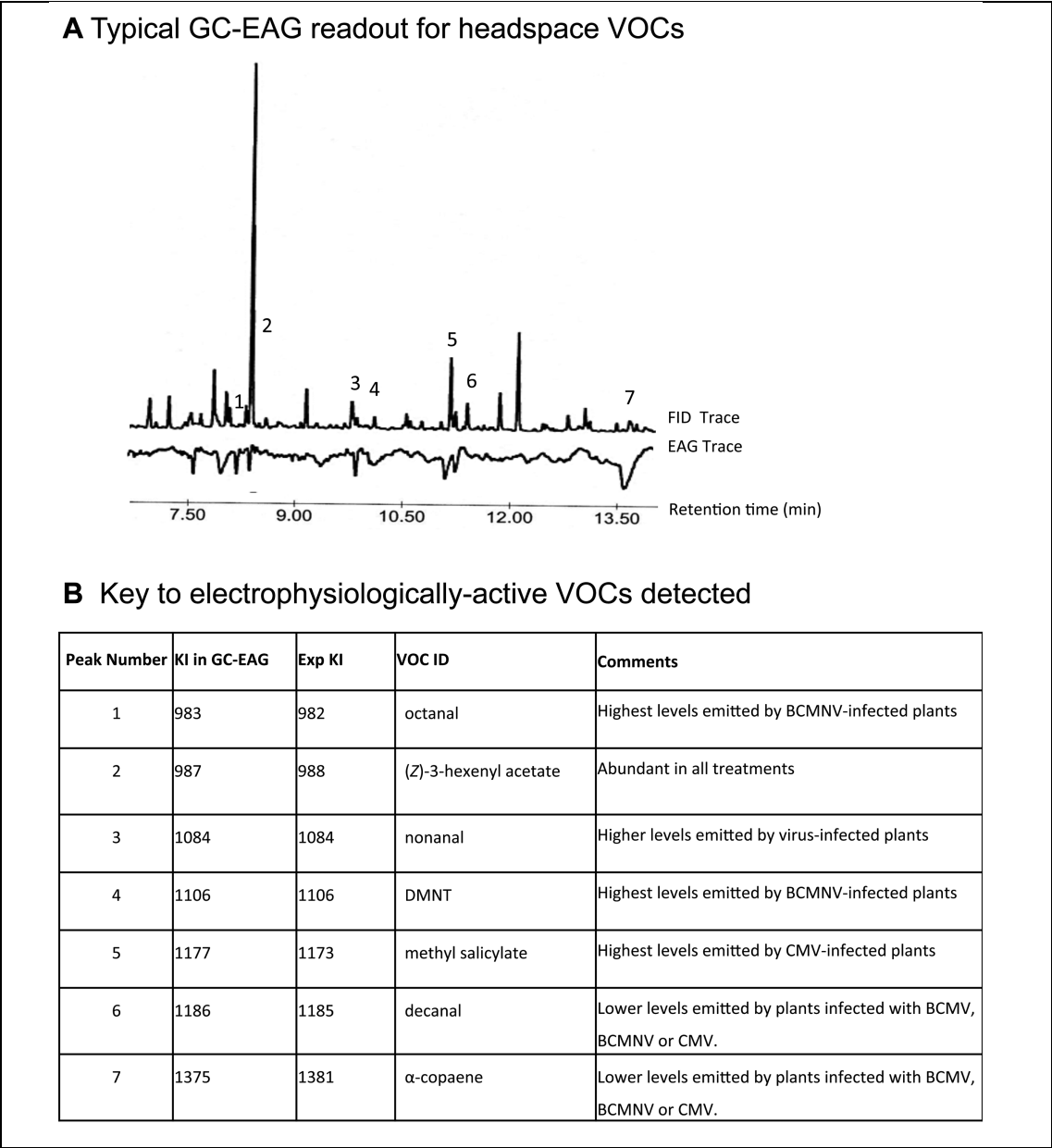
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Supplementary Figures

SUPPLEMENTARY FIGURE 1. Experimental set up for aphid free-choice tests. In this example the bean plant labeled ‘Infected’ had been inoculated with BCMNV 10 days previously at the two-leaf stage, while sterile water had been used for mock-inoculation (plant labeled ‘Mock’). Panels **A** and **B** show, respectively, views from above and from the side of the experimental set up. Aphids (*Myzus persicae* or *Aphis fabae*) were released from a microfuge tube placed equidistantly between the plants (indicated by a white star) and their settling behavior observed for up to 24 h post-release. A micro-perforated bag, which is porous to air but which confines aphids, was secured around the set up so as to prevent the aphids from escaping (**C**).



SUPPLEMENTARY FIGURE 2. Coupled gas chromatography–electroantennography (GC-EAG). Example of an amalgamated GC-EAG recording for an *A. fabae* antennal preparation responding to headspace volatile organic compounds (VOCs) emitted by CMV-infected bean plants (a). The upper trace represents the flame ionization detector response following GC and the lower trace represents the aphid electroantennography (EAG) response. The key (b) indicates the electrophysiologically active VOC peaks, 1-7, as well as the actual and expected Kovats retention index (KI) for these VOCs.

