

Lack of TYK2 Signaling Enhances Host Resistance to *Candida albicans* Skin Infection

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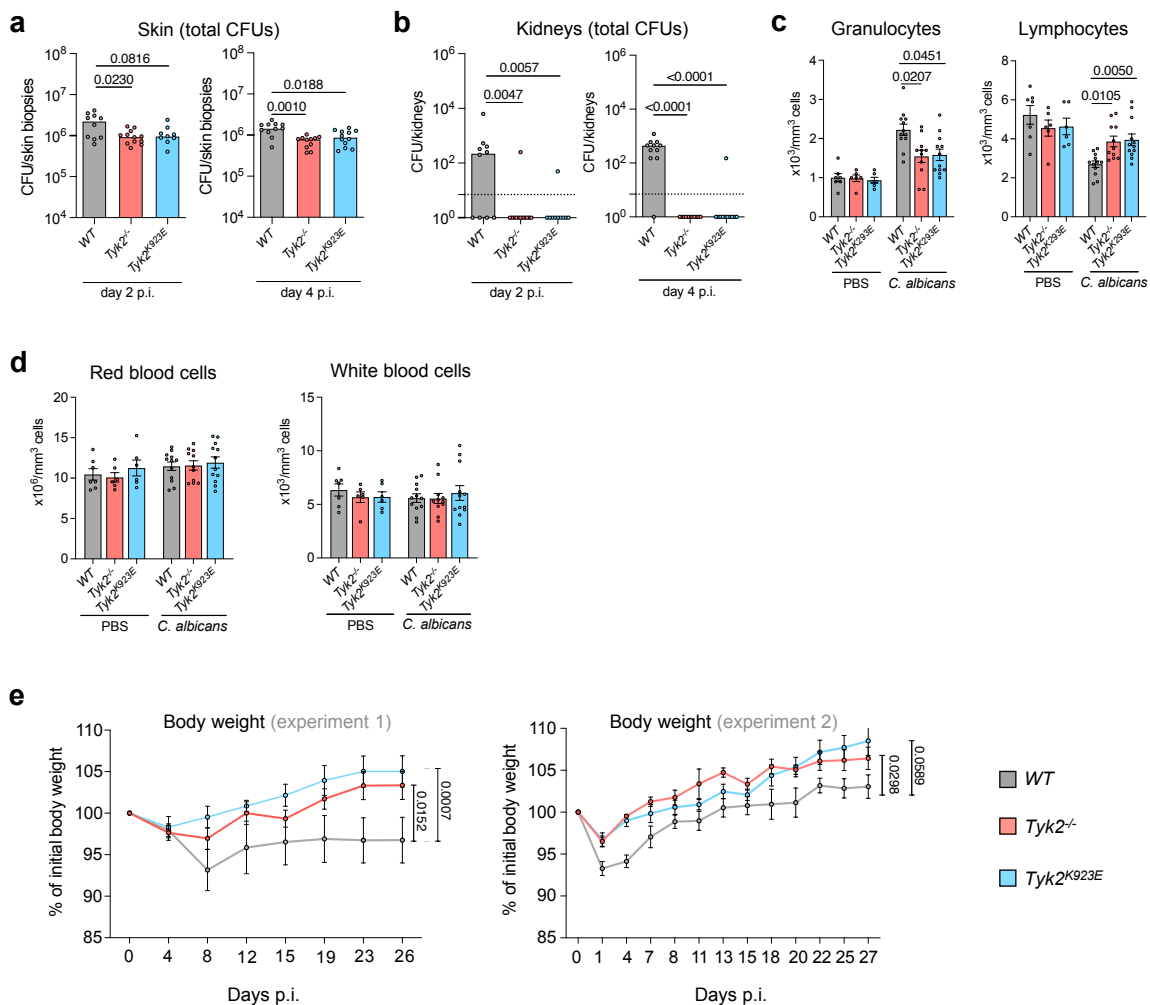
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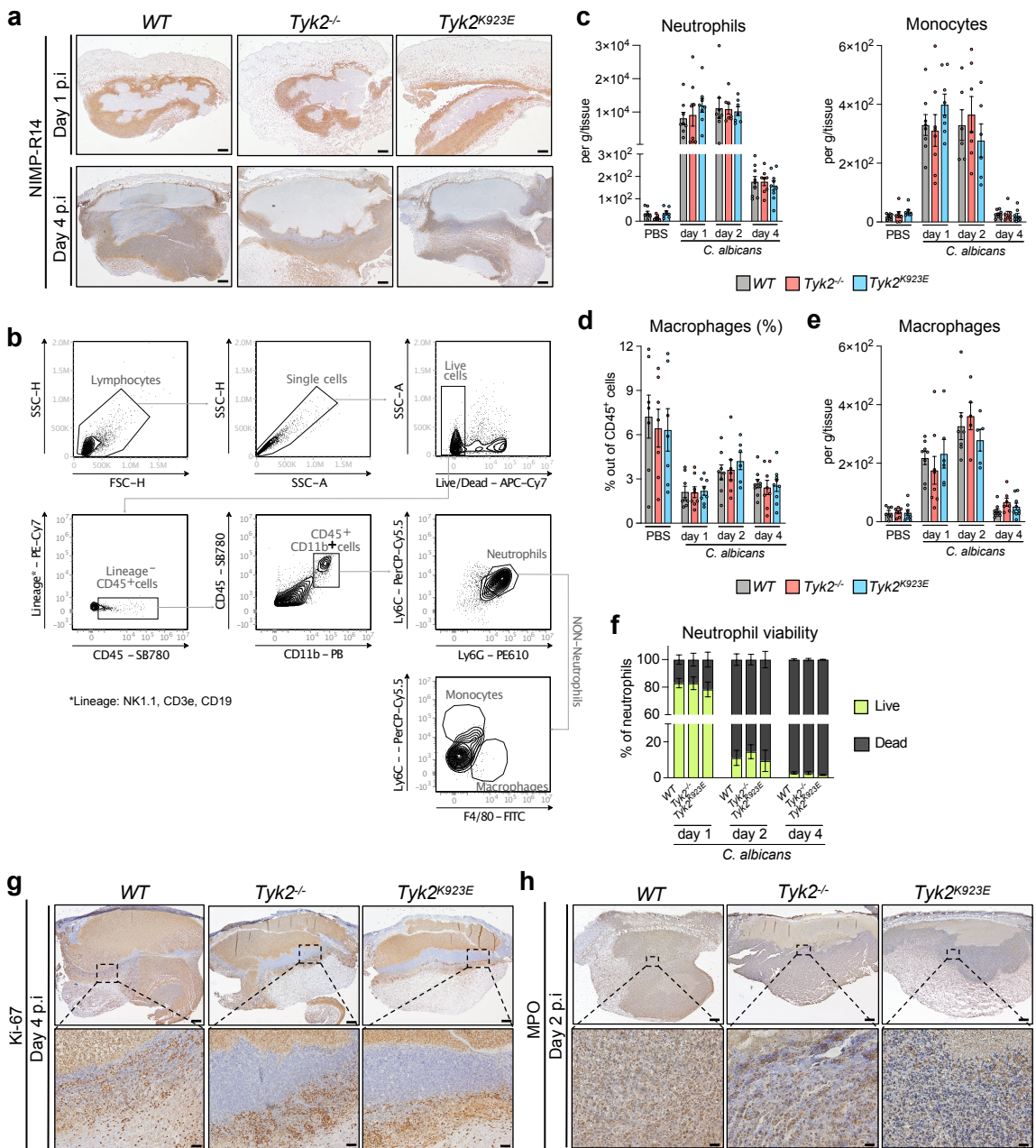
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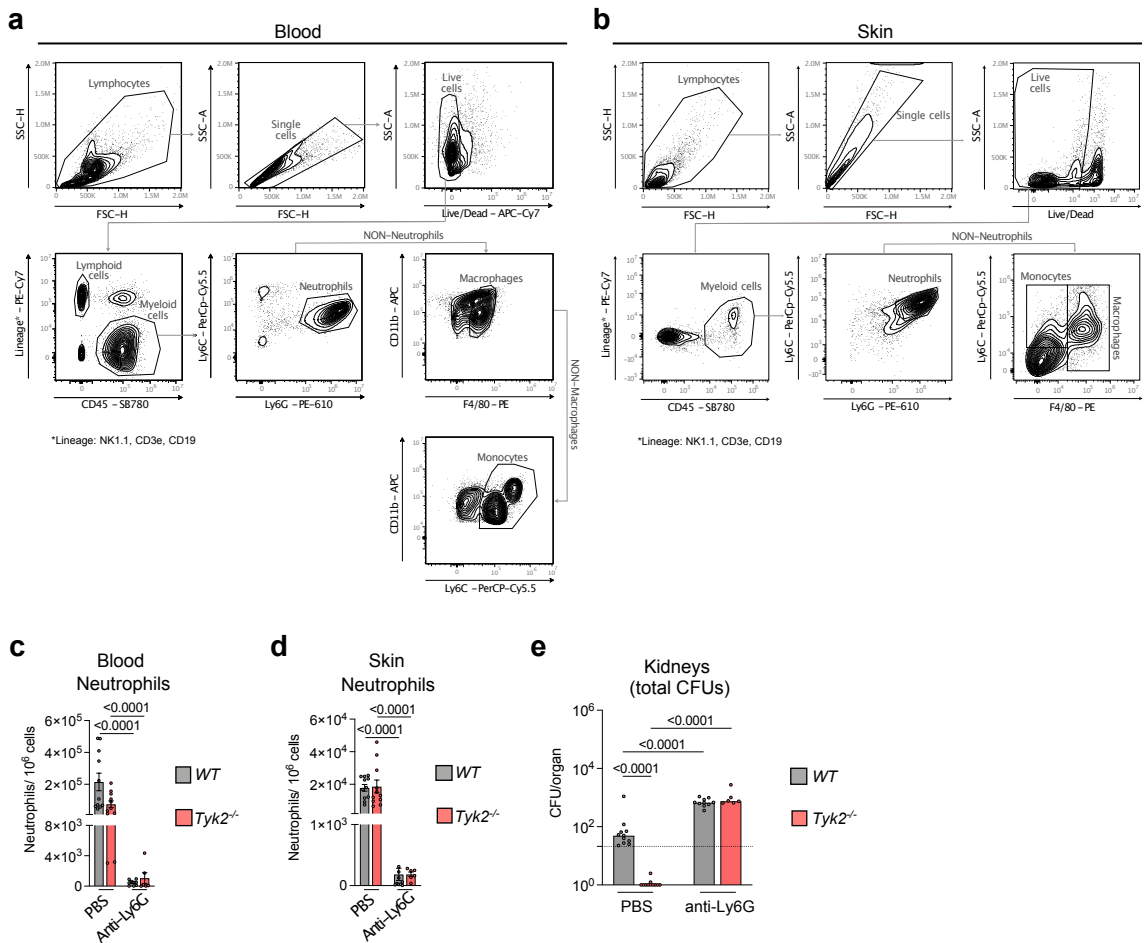
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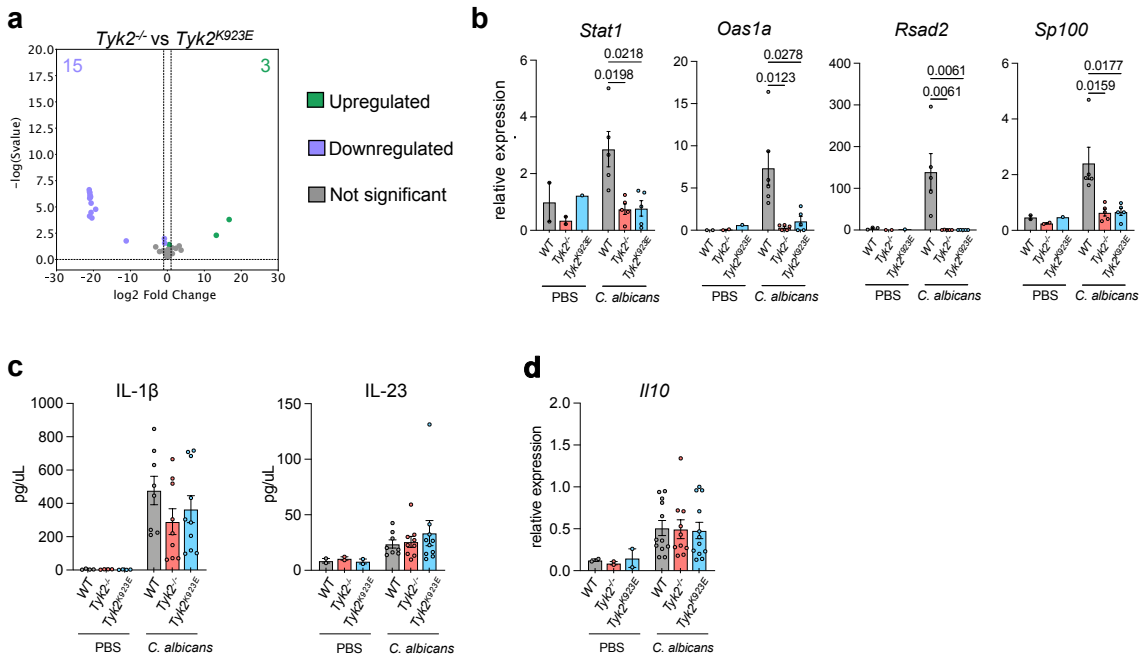
Supplementary Fig. S1. a-b Fungal load in the skin (a) and in the kidneys (b) was measured on days 2 and day 4 p.i. The total number of CFUs per skin (4 biopsies of 4 mm) and kidneys is represented. For each time-point, pooled data from 2 independent experiments are shown. Skin - day 2: n=10 (WT), n=12 (*Tyk2*^{-/-}), n=9 (*Tyk2*^{K923E}); day 4: n=11 (WT), n=11 (*Tyk2*^{-/-}), n=12 (*Tyk2*^{K923E}); Kidneys - day 2: n=10 (WT), n=12 (*Tyk2*^{-/-}), n=10 (*Tyk2*^{K923E}); day 4: n=11 (WT), n=11 (*Tyk2*^{-/-}), n=11 (*Tyk2*^{K923E}); Median values are given; n: biological replicates. The dotted line indicates the assay detection limit. **c-d** Blood cell composition on day 4 p.i was determined with a Vet ABC analyzer. The total numbers of granulocytes and lymphocytes (c) and of red and white blood cells (d) are shown. For each cell population, pooled data from 2 independent experiments are shown. PBS: n=7 (WT) and n=6 (*Tyk2*^{-/-} and *Tyk2*^{K923E}); *C. albicans*: n=12 (WT and *Tyk2*^{K923E}) and n=11 (*Tyk2*^{-/-}); Mean values ± SEM are given; n: biological replicates. **e** Body weight of infected WT, *Tyk2*^{-/-} and *Tyk2*^{K923E} mice was measured overtime and is shown as percentage relative to the weight on the day before the infection (set as 100%). Two independent experiments are separately represented. Experiment 1: n=6 (WT and *Tyk2*^{-/-}) and n=5 (*Tyk2*^{K923E}); Experiment 2: n=7 (WT) and n=6 (*Tyk2*^{-/-} and *Tyk2*^{K923E}); Mean values ± SEM are given; n: biological replicates. Statistical analysis was conducted using One-way ANOVA followed by Tukey's multiple comparison test (a-e). Statistical significance is only given for the comparison between the genotypes (c,d). Source data are provided as a Source Data file.



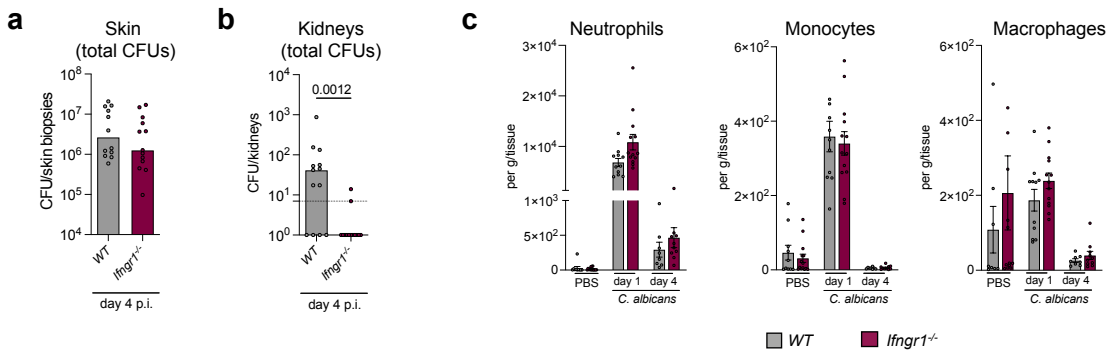
Supplementary Fig. S2. **a** Skin sections obtained from infected WT, *Tyk2*^{-/-} and *Tyk2*^{K923E} mice on day 1 and 4 p.i. were stained with an anti-NIMP-R14 antibody. Data are representative of sections from 5 mice per genotype (day 1) and 4 WT and 5 *Tyk2*^{-/-} and *Tyk2*^{K923E} mice (day 4). Scale bar: 200 μ m. **b** The gating strategy used to define skin-infiltrating neutrophils, monocytes and macrophages is shown. **c-e** Skin-infiltrating neutrophils (gated as CD45⁺CD11b⁺Ly6C⁺Ly6G⁺ cells) and monocytes (gated as CD45⁺CD11b⁺Ly6G⁺Ly6C^{high}F4/80⁺ cells) (c) and macrophages (gated as CD45⁺CD11b⁺Ly6G⁺F4/80⁺ cells) (d, e) were quantified by flow cytometry on days 1, 2 and 4 p.i. The number of these cells per gram of skin (c, e) or the percentage of cells out of myeloid CD45⁺CD11b⁺ cells (d) is shown. For each time-point, pooled data from 2 independent experiments are shown. **c** PBS: n=7 (WT, *Tyk2*^{-/-}, *Tyk2*^{K923E}); C. albicans day 1: n=8 (WT, *Tyk2*^{-/-}, *Tyk2*^{K923E}); C. albicans day 2: n=8 (WT) and n=7 (*Tyk2*^{-/-}, *Tyk2*^{K923E}); C. albicans day 4: n=9 (WT, *Tyk2*^{-/-}) and n=8 (*Tyk2*^{K923E}); **d** PBS: n=7 (WT, *Tyk2*^{-/-}, *Tyk2*^{K923E}); C. albicans day 1: n=8 (WT, *Tyk2*^{-/-}, *Tyk2*^{K923E}); C. albicans day 2: n=8 (WT), n=7 (*Tyk2*^{-/-}) and n=6 (*Tyk2*^{K923E}); C. albicans day 4: n=9 (WT, *Tyk2*^{K923E}) and n=8 (*Tyk2*^{-/-}); **e** PBS: n=7 (WT, *Tyk2*^{-/-}, *Tyk2*^{K923E}); C. albicans day 1: n=8 (WT, *Tyk2*^{-/-}, n=6 (*Tyk2*^{K923E}); C. albicans day 2: n=7 (WT), n=5 (*Tyk2*^{-/-}, *Tyk2*^{K923E}), day 4: n=9 (WT, *Tyk2*^{K923E}) and n=7 (*Tyk2*^{-/-}); Mean values \pm SEM are given; n: biological replicates. **f** The viability of skin infiltrating neutrophils was quantified based on flow cytometry analysis with live/dead staining. For each time-point, pooled data from 2 experiments is shown. C. albicans day 1: n=8 (WT, *Tyk2*^{-/-}, *Tyk2*^{K923E}); C. albicans day 2: n=8 (WT), n=7 (*Tyk2*^{-/-}, *Tyk2*^{K923E}); C. albicans day 4: n=9 (WT, *Tyk2*^{K923E}) and n=8 (*Tyk2*^{-/-}); Mean values \pm SEM are given; n: biological replicates. **g** Skin sections obtained from infected WT, *Tyk2*^{-/-} and *Tyk2*^{K923E} mice on day 4 p.i. were stained with an anti-Ki-67 antibody. Scale bar: 200 μ m (top), 50 μ m (bottom). **h** Skin sections obtained from infected WT, *Tyk2*^{-/-} and *Tyk2*^{K923E} mice on day 2 p.i. were stained with an anti-MPO antibody. Scale bar: 200 μ m (top), 20 μ m (bottom). Data are representative of sections from 4 WT and 5 *Tyk2*^{-/-} and *Tyk2*^{K923E} mice (g), and 7 mice per genotype (h). Statistical analysis was conducted using One-way ANOVA followed by Tukey's multiple comparison test (c-f). Statistical significance is only given for the comparison between the genotypes (c-f). Source data are provided as a Source Data file.



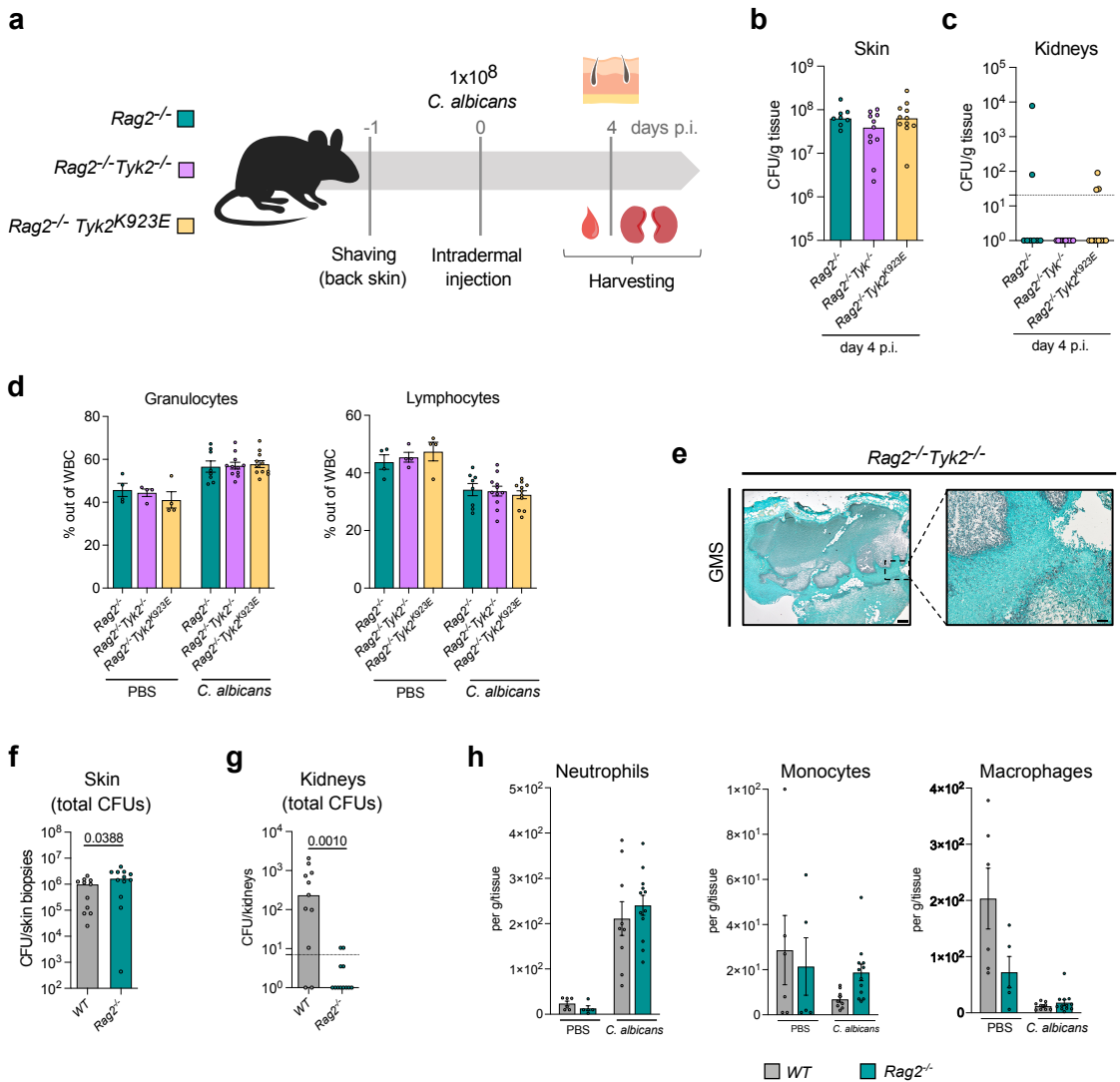
Supplementary Fig. S3. a-b The gating strategies used to define neutrophils, monocytes and macrophages in the blood (**a**) and the skin (**b**) are shown. **c-d** The numbers of neutrophils in the blood (**c**) and in the skin (**d**) are shown. Pooled data from 2 independent experiments are shown. **c** PBS: n=11 (WT, *Tyk2*^{-/-}); Anti-Ly6G: n=9 (WT), n=6 (*Tyk2*^{-/-}); **d** PBS: n=11 (WT, *Tyk2*^{-/-}); Anti-Ly6G: n=10 (WT), n=6 (*Tyk2*^{-/-}); Mean values \pm SEM are given; n: biological replicates. **e** Fungal load in the kidneys of mice injected with anti-Ly6G or PBS was measured on day 2 p.i. PBS: n=11 (WT, *Tyk2*^{-/-}); Anti-Ly6G: n=11 (WT), n=6 (*Tyk2*^{-/-}); Mean values \pm SEM are given; n: biological replicates. The dotted line indicates the assay detection limit. Statistical analysis was conducted using One-way ANOVA followed by Tukey's multiple comparison test (**c-e**). Source data are provided as a Source Data file.



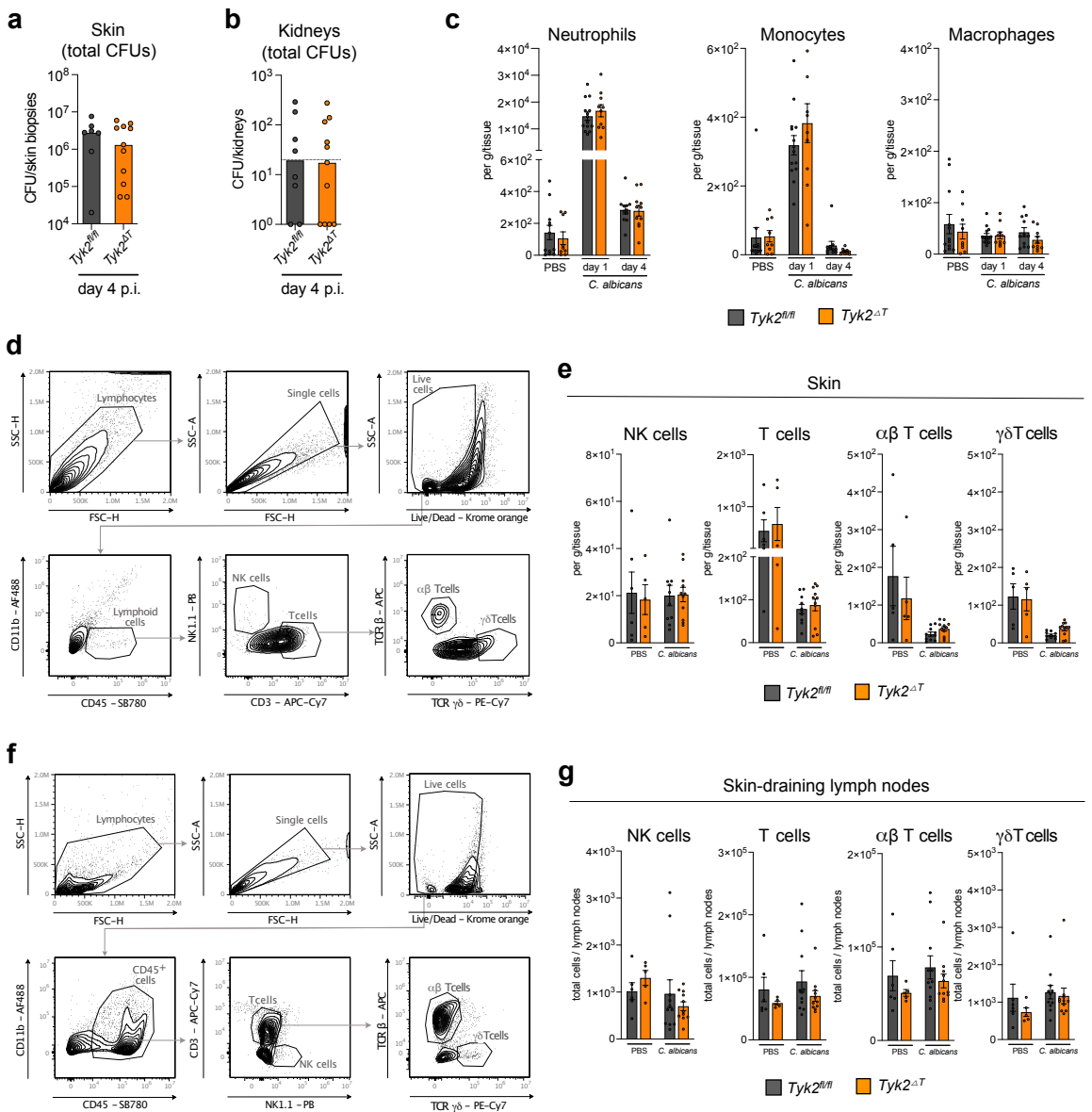
Supplementary Fig. S4. **a** Volcano plot showing the differentially expressed genes between *Tyk2*^{-/-} and *Tyk2*^{K923E}. Genes were identified from DESeq2-normalized read counts of genes with a threshold of $p_{adj} < 0.05$ between *Tyk2*^{-/-} and *Tyk2*^{K923E} cells. Data are from one experiment (n=3 per genotype). **b** mRNA levels of *Stat1*, *Oas1a*, *Rsd2*, and *Sp100* in the skin on day 1 p.i. were measured by RT-qPCR. Data were normalized to the housekeeping gene *Ube2d2*. One experiment is shown. PBS: n=2 (WT, *Tyk2*^{-/-}), n=1 (*Tyk2*^{K923E}); *C. albicans*: n=5 (WT, *Tyk2*^{-/-}, *Tyk2*^{K923E}); Mean values \pm SEM are given; n: biological replicates. **c** IL-1β and IL-23 in the skin on day 2 p.i. were measured using a Luminex assay. Pooled data from 2 independent experiments are shown. IL-1β – PBS: n=4/genotype; *C. albicans*, n=8 (WT), n=10 (*Tyk2*^{-/-}), n=9 (*Tyk2*^{K923E}); IL-23 – PBS: n=2/genotype (PBS); *C. albicans*: n=8 (WT), n=10 (*Tyk2*^{-/-}), n=9 (*Tyk2*^{K923E}); Mean values \pm SEM are given; n: biological replicates. **d** mRNA levels of *Il10* in the infected skin on day 4 p.i. were measured by RT-qPCR. Data were normalized to the housekeeping gene *Ube2d2*. Pooled data from 2 independent experiments are shown. PBS: n=2 (PBS); *C. albicans*: n=12 (WT), n=10 (*Tyk2*^{-/-}), n=12 (*Tyk2*^{K923E}); Mean values \pm SEM are given; n: biological replicates. Statistical analysis was conducted using One-way ANOVA followed by Tukey's multiple comparison test (**b-d**). Statistical significance is only given for the comparison between the genotypes (**b-d**). Source data are provided as a Source Data file.



Supplementary Fig. S5. a-b Fungal load in the skin (**a**) and in the kidneys (**b**) was measured on day 4 p.i. The total number of CFUs per skin (4 biopsies of 4 mm) and kidneys is represented. Pooled data from 2 independent experiments are shown. **a** $n=12$ (WT), $n=13$ (*Ifngr1^{-/-}*); **b** $n=13$ (WT, *Ifngr1^{-/-}*); Median values are given; n : biological replicates. The dotted line indicates the assay detection limit. **c** Skin-infiltrating neutrophils (gated as CD45⁺CD11b⁺Ly6C⁺Ly6G⁺ cells), monocytes (gated as CD45⁺CD11b⁺Ly6G⁺Ly6C^{high}F4/80⁻ cells) and macrophages (gated as CD45⁺CD11b⁺Ly6G⁺F4/80⁺ cells) were quantified based on flow cytometry analysis on day 1 and 4 p.i. For each time-point, pooled data from 2 independent experiments are shown. Neutrophils and Monocytes – PBS: $n=10$ (WT, *Ifngr1^{-/-}*); *C. albicans* day 1: $n=11$ (WT), $n=13$ (*Ifngr1^{-/-}*); *C. albicans* day 4: $n=8$ (WT), $n=10$ (*Ifngr1^{-/-}*); Macrophages – PBS: $n=8$ (WT), $n=13$ (*Ifngr1^{-/-}*); *C. albicans* day 1: $n=11$ (WT), $n=13$ (*Ifngr1^{-/-}*); *C. albicans* day 4: $n=8$ (WT), $n=10$ (*Ifngr1^{-/-}*); Mean values \pm SEM are given; n : biological replicates. Statistical analysis was conducted using a two-tailed Mann Whitney test (**a**, **b**) and One-way ANOVA followed by Tukey's multiple comparison test (**c**). Statistical significance is only given for the comparison between the genotypes (**c**). Source data are provided as a Source Data file.



Supplementary Fig. S6. a *Rag2*^{-/-}, *Rag2*^{-/-}*Tyk2*^{-/-} and *Rag2*^{-/-}*Tyk2*^{K923E} mice were infected as described in the legend to Fig. 1. **b-c** Fungal load in the skin (**b**) and kidneys (**c**) was measured on day 4 p.i. Pooled data from 2 independent experiments are shown. *n*=8 (*Rag2*^{-/-}), *n*=11 (*Rag2*^{-/-}*Tyk2*^{-/-}, *Rag2*^{-/-}*Tyk2*^{K923E}); Median values are given; *n*: biological replicates. The dotted line indicates the assay detection limit. **d** Blood cell composition on day 4 p.i. was determined with a Vet ABC analyzer. The percentage of granulocytes and lymphocytes out of total white blood cells (WBC) is shown. For each cell population, pooled data from 2 independent experiments are shown. PBS: *n*=4 (*Rag2*^{-/-}, *Rag2*^{-/-}*Tyk2*^{-/-}, *Rag2*^{-/-}*Tyk2*^{K923E}); *C. albicans*: *n*=8 (*Rag2*^{-/-}), *n*=11 (*Rag2*^{-/-}*Tyk2*^{-/-}, *Rag2*^{-/-}*Tyk2*^{K923E}); Mean values \pm SEM are given; *n*: biological replicates. **e** Representative pictures of the infected skin on day 4 p.i. A GMS staining of the skin sections is shown. Data are representative of sections from 4 mice per genotype. Scale bar represents 200 μ m (left) or 50 μ m (right). **f-g** Fungal load in the skin (**f**) and in the kidneys (**g**) was measured on day 4 p.i. The total number of CFUs per skin (4 biopsies of 4 mm) and kidneys and pooled data from 2 independent experiments are shown. *n*=11 (WT), *n*=12 (*Rag2*^{-/-}); Median values are given; *n*: biological replicates. The dotted line indicates the assay detection limit. **h** Skin-infiltrating neutrophils (gated as CD45⁺CD11b⁺Ly6C⁺Ly6G⁺ cells), monocytes (gated as CD45⁺CD11b⁺Ly6G⁺Ly6C^{high}F4/80⁻ cells) and macrophages (gated as CD45⁺CD11b⁺Ly6G⁺F4/80⁺ cells) were quantified based on flow cytometry analysis on day 4 p.i. Pooled data from 2 independent experiments are shown. PBS: *n*=6 (WT), *n*=5 (*Rag2*^{-/-}); *C. albicans*: *n*=9 (WT), *n*=12 (*Rag2*^{-/-}); Mean values \pm SEM are given; *n*: biological replicates. Statistical analysis was conducted using One-way ANOVA followed by Tukey's multiple comparison test (**b-d**, **h**) or two-tailed Mann Whitney test (**f-g**). Statistical significance is only given for the comparison between the genotypes (**b-d**, **h**). Source data are provided as a Source Data file.



Supplementary Fig. S7. a-b Fungal load in the skin (**a**) and in the kidneys (**b**) was measured on day 4 p.i. The total number of CFUs per skin (4 biopsies of 4 mm) and kidneys is represented. Pooled data from 2 independent experiments are shown. **a** $n=7$ ($Tyk2^{fl/m}$), $n=11$ ($Tyk2^{\Delta T}$); **b** $n=8$ ($Tyk2^{fl/m}$), $n=11$ ($Tyk2^{\Delta T}$); Median values are given; n : biological replicates. The dotted line indicates the assay detection limit. **c** Skin-infiltrating neutrophils (gated as $CD45^+CD11b^+Ly6C^+Ly6G^+$ cells), monocytes (gated as $CD45^+CD11b^+Ly6G-Ly6C^{high}F4/80^-$ cells) and macrophages (gated as $CD45^+CD11b^+Ly6G-F4/80^+$ cells) were quantified by flow cytometry analysis on 4 p.i. Pooled data from 2 independent experiments are shown. PBS: $n=12$ ($Tyk2^{fl/m}$), $n=9$ ($Tyk2^{\Delta T}$); *C. albicans* day 1: $n=14$ ($Tyk2^{fl/m}$), $n=10$ ($Tyk2^{\Delta T}$); *C. albicans* day 4: $n=11$ ($Tyk2^{fl/m}$), $n=11$ ($Tyk2^{\Delta T}$, Neutrophils), $n=10$ ($Tyk2^{\Delta T}$, Monocytes, Macrophages); Mean values \pm SEM are given; n : biological replicates. **d-g** The gating strategies used to identify NK cells, T cells, MMT cells and MMT cells in the skin (**d**) and in the skin-draining lymph nodes (**f**) on day 4 p.i. and the total numbers of each of these populations per gram of skin (**e**) or in the lymph nodes (**g**) are presented. Pooled data from 2 independent experiments are shown. **e** PBS: $n=6$ ($Tyk2^{fl/m}$, NK cells and T cells), $n=5$ ($Tyk2^{fl/m}$, MMT cells and MMT cells), $n=5$ ($Tyk2^{\Delta T}$); *C. albicans*: $n=10$ ($Tyk2^{fl/m}$), $n=11$ ($Tyk2^{\Delta T}$); **g** PBS: $n=6$ ($Tyk2^{fl/m}$), $n=5$ ($Tyk2^{\Delta T}$); *C. albicans*: $n=11$ ($Tyk2^{fl/m}$, $Tyk2^{\Delta T}$); Mean values \pm SEM are given; n : biological replicates. Statistical analysis was conducted using two-tailed Mann Whitney test (**a,b**) or One-way ANOVA followed by Tukey's multiple comparison test (**c, e, g**). Statistical significance is only given for the comparison between the genotypes (**c, e, g**). Source data are provided as a Source Data file.

