



# Genome Sequence of a Single-Stranded DNA Virus Identified in Gila Monster Feces

Vasishtha Somayaji,<sup>a</sup> Dale DeNardo,<sup>b</sup> Melissa A. Wilson Sayres,<sup>b,c</sup> Mellecha Blake,<sup>a,b</sup> Kara Waits,<sup>a</sup> Rafaela S. Fontenele,<sup>a</sup> Simona Kraberger,<sup>a</sup> Arvind Varsani<sup>a,b,c,d</sup>

<sup>a</sup>Biodesign Center for Fundamental and Applied Microbiomics, Arizona State University, Tempe, Arizona, USA

<sup>b</sup>School of Life Sciences, Arizona State University, Tempe, Arizona, USA

<sup>c</sup>Center for Evolution and Medicine, Arizona State University, Tempe, Arizona, USA

<sup>d</sup>Structural Biology Research Unit, Department of Clinical Laboratory Sciences, University of Cape Town, Observatory, Cape Town, South Africa

**ABSTRACT** The Gila monster (*Heloderma suspectum*) is native to the Sonoran Desert. Metagenomic analyses of a Gila monster fecal sample revealed the presence of a small, circular, single-stranded DNA virus that is most closely related to a gemyrovirus (family *Genomoviridae*) genome from caribou feces sharing 88% genome-wide pairwise identity.

The Gila monster (*Heloderma suspectum*) is a venomous lizard native to the Sonoran Desert. Despite significant activity in recovering a broad diversity of viruses associated with various reptiles (1) in the last decade, only viruses from the *Adenoviridae* family (2, 3) have been detected in Gila monsters to date. Here, we sampled the feces of a Gila monster to identify associated DNA viruses. The freshly deposited fecal sample (5 g) was homogenized in 20-ml SM buffer (0.1 M NaCl, 50 mM Tris HCl [pH 7.4]) and centrifuged at 10,000 × *g* for 10 min. The resulting supernatant was filtered sequentially through a 0.45- $\mu$ m and 0.2- $\mu$ m syringe filter. We used 200  $\mu$ l of the filtrate to extract viral DNA using the High Pure viral nucleic acid kit (Roche Diagnostics, USA). Circular viral DNA was amplified using rolling circle amplification (RCA) with the Illustra TempliPhi 100 amplification kit (GE Healthcare, USA). The RCA products were used to prepare a 2 × 100-bp paired-end library and sequenced on a HiSeq 4000 instrument. Paired-end reads (24,473,832 reads, read length 101) were trimmed using Trimmomatic v0.36 (4) and then *de novo* assembled using ABySS 2.0 (5) with a k-mer of 64 resulting in 304,270 contigs ( $N_{50}$ , 1,868 nucleotides). All *de novo* assembled contigs of >500 nucleotides (nt) were analyzed using a BLASTx search (6) against a local viral protein database compiled from GenBank. A contig of ~2,000 nt was identified as having similarity to the replication-associated protein (Rep) encoded by viruses in the family *Genomoviridae*. Genomoviruses have small, circular, single-stranded DNA genomes that encode two main proteins, the Rep and a capsid protein (CP) (7). Genomoviruses ( $n = 196$ ) have been detected from various samples, including plant leaves, blood, serum, cerebrospinal fluid, feces, and the whole body of various insects, wastewater, and sediments; however, only one genomovirus (*Sclerotinia sclerotiorum* hypovirulence-associated DNA virus 1) infecting the fungus *Sclerotinia sclerotiorum* has been identified (8).

Based on the genomovirus-like contig, a pair of abutting primers (Gila\_283F, 5'-GTATTGTATCTAGCTAGCCTCATGCCAATATGTTG-3', and Gila\_283R, 5'-AAATTGGATC TATCACATTACAATTACGCAGTTG-3') were designed to recover the complete genome. The genome was amplified with PCR using the Gila\_283F/R primer pair with KAPA HiFi HotStart DNA polymerase (Roche, USA), cloned, and Sanger sequenced.

The Gila monster-associated virus 1 (GmaV1; GenBank accession number [MH378453](https://doi.org/10.1128/MRA.00925-18))

Received 5 July 2018 Accepted 24 July 2018 Published 23 August 2018

**Citation** Somayaji V, DeNardo D, Wilson Sayres MA, Blake M, Waits K, Fontenele RS, Kraberger S, Varsani A. 2018. Genome sequence of a single-stranded DNA virus identified in Gila monster feces. *Microbiol Resour Announc* 7:e00925-18. <https://doi.org/10.1128/MRA.00925-18>.

**Editor** John J. Dennehy, Queens College

**Copyright** © 2018 Somayaji et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Arvind Varsani, [arvind.varsani@asu.edu](mailto:arvind.varsani@asu.edu).

genome sequence is 2,191 nt (GC content, 50.5%), has a “TAATATTAT” nonanucleotide motif, and encodes three proteins, the CP in the virion sense and the Rep and RepA on the complementary sense. The GmaV1 genome shares 88%, 75%, and 66% genome pairwise identity, respectively, with a caribou feces-associated genomovirus sampled in Canada (GenBank accession number [KJ938717](#)) (9), a bovine blood-associated genomovirus from Germany ([LK931484](#)) (10), and a sewage-associated genomovirus from a sewage oxidation pond in New Zealand ([KJ547634](#)) (11). The CP and Rep amino acid sequences of GmaV1 share between 69 and 84% and 39 and 90% pairwise identity, respectively, with those of the genomoviruses from caribou feces ([KJ938717](#)) (9), bovine blood ([LK931484](#)) (10), and the sewage oxidation pond ([KJ547634](#)) (11). Based on the *Genomoviridae* classification (12), GmaV1 would belong to the genus *Gemykrogvirus*, which currently has three species. GmaV1 represents the third documented virus to be associated with the Gila monster; however, it is unknown whether this virus is associated with its gut flora or diet or whether it is pathogenic to the animal.

**Data availability.** The complete genome sequence of Gila monster-associated virus 1 has been deposited in GenBank with accession number [MH378453](#).

## ACKNOWLEDGMENTS

This work was supported by funds from the Biodesign Institute and School of Life Sciences at Arizona State University awarded to A.V.

## REFERENCES

- Marschang RE. 2011. Viruses infecting reptiles. *Viruses* 3:2087–2126. <https://doi.org/10.3390/v3112087>.
- Papp T, Fledelius B, Schmidt V, Kaján GL, Marschang RE. 2009. PCR-sequence characterization of new adenoviruses found in reptiles and the first successful isolation of a lizard adenovirus. *Vet Microbiol* 134: 233–240. <https://doi.org/10.1016/j.vetmic.2008.08.003>.
- Wellehan JFX, Johnson AJ, Harrach B, Benkő M, Pessier AP, Johnson CM, Garner MM, Childress A, Jacobson ER. 2004. Detection and analysis of six lizard adenoviruses by consensus primer PCR provides further evidence of a reptilian origin for the atadenoviruses. *J Virol* 78:13366–13369. <https://doi.org/10.1128/JVI.78.23.13366-13369.2004>.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>.
- Jackman SD, Vandervalk BP, Mohamadi H, Chu J, Yeo S, Hammond SA, Jahesh G, Khan H, Coombe L, Warren RL, Biro I. 2017. ABySS 2.0: resource-efficient assembly of large genomes using a Bloom filter. *Genome Res* 27:768–777. <https://doi.org/10.1101/gr.214346.116>.
- Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. *J Mol Biol* 215:403–410. [https://doi.org/10.1016/S0022-2836\(05\)80360-2](https://doi.org/10.1016/S0022-2836(05)80360-2).
- Krupovic M, Ghabrial SA, Jiang D, Varsani A. 2016. *Genomoviridae*: a new family of widespread single-stranded DNA viruses. *Arch Virol* 161: 2633–2643. <https://doi.org/10.1007/s00705-016-2943-3>.
- Yu X, Li B, Fu YP, Jiang DH, Ghabrial SA, Li GQ, Peng YL, Xie JT, Cheng JS, Huang JB, Yi XH. 2010. A geminivirus-related DNA mycovirus that confers hypovirulence to a plant pathogenic fungus. *Proc Natl Acad Sci U S A* 107:8387–8392. <https://doi.org/10.1073/pnas.0913535107>.
- Ng TFF, Chen L-F, Zhou Y, Shapiro B, Stiller M, Heintzman PD, Varsani A, Kondov NO, Wong W, Deng X, Andrews TD, Moorman BJ, Meulendyk T, MacKay G, Gilbertson RL, Delwart E. 2014. Preservation of viral genomes in 700-y-old caribou feces from a subarctic ice patch. *Proc Natl Acad Sci U S A* 111:16842–16847. <https://doi.org/10.1073/pnas.1410429111>.
- Lamberto I, Gunst K, Muller H, zur Hausen H, de Villiers E-M. 2014. Mycovirus-like DNA virus sequences from cattle serum and human brain and serum samples from multiple sclerosis patients. *Genome Announc* 2:e00848-14. <https://doi.org/10.1128/genomeA.00848-14>.
- Kraberger S, Arguello-Astorga GR, Greenfield LG, Galilee C, Law D, Martin DP, Varsani A. 2015. Characterisation of a diverse range of circular replication-associated protein encoding DNA viruses recovered from a sewage treatment oxidation pond. *Infect Genet Evol* 31:73–86. <https://doi.org/10.1016/j.meegid.2015.01.001>.
- Varsani A, Krupovic M. 2017. Sequence-based taxonomic framework for the classification of uncultured single-stranded DNA viruses of the family *Genomoviridae*. *Virus Evol* 3:vev037. <https://doi.org/10.1093/ve/vev037>.