

# Equilibrium Studies of Dibutyltin(IV)–Zwitterionic Buffer Complexation

M. A. El-Gahami · H. M. Albishri

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**Abstract** Equilibrium studies in aqueous solution are reported for dibutyltin(IV) (DBT) complexes of the zwitterionic buffers “Good’s buffers” Mes and Mops. Stoichiometric and formation constants of the complexes formed were determined at different temperatures and ionic strength  $0.1 \text{ mol}\cdot\text{L}^{-1} \text{ NaNO}_3$ . The results show that the best fit of the titration curves were obtained when the complexes  $\text{ML}$ ,  $\text{MLH}_{-1}$ ,  $\text{MLH}_{-2}$  and  $\text{MLH}_{-3}$  were considered beside the hydrolysis product of the dibutyltin(IV) cation. The thermodynamic parameters  $\Delta H^\circ$ ,  $\Delta S^\circ$  and  $\Delta G^\circ$  calculated from the temperature dependence of the formation constant of the dibutyltin(IV) complexes with 2-(*N*-morpholino)ethanesulfonic acid (Mes) and 3-(*N*-morpholino)-propanesulfonic acid (Mops) were investigated. The effect of dioxane as a solvent on the formation constants of DBT–Mes and DBT–Mops complexes decrease linearly with the increase of dioxane proportion in the medium. The concentration distribution of the various complexes species was evaluated as a function of pH.

**Keywords** Equilibrium studies · Dibutyltin(IV) complexes · “Good’s buffers” · Effect of solvent and temperature · Thermodynamic parameters

## 1 Introduction

Organotin(IV) compounds are receiving increasing interest because of their biological and potential pharmaceutical applications as antitumor agents [1–6]. On the other hand, in recent years many organotin(IV) compounds have been tested for their *in vitro* activity

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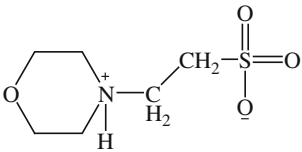
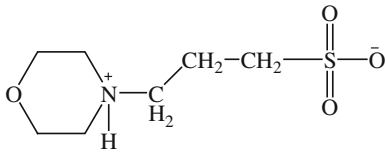
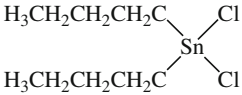
against a large variety of tumor lines and have been found to be as effective as, or better than, traditional heavy metal anticancer drugs such as *cis*-platin [6]. In addition, organotin(IV) compounds are also of interest in view of their considerable structural diversity.

Zwitterionic buffers [7–9] are currently used routinely in biochemical and speciation studies under the assumption that they undergo little if any interaction with biologically (or environmentally) important metal ions. However, there has been an increase in reports on buffer interferences when most of Good's buffers were used in the presence of metal ions [10–16]. Many conflicting data and conclusions that were reported by investigators studying identical metal cation protein systems (at the same pH in carefully executed experiments) might be due in part to the failure to include the presence of metal–buffer complexes.

These compounds (Good's buffers) contain hydroxyl, carboxylate or sulfhydryl groups in addition to a primary nitrogen donor atom. Therefore, one might expect that metal complexes of these buffers would form in biochemical/environmental systems containing metal ions, when these compounds are used to control the pH. On the other hand, it is known that 2-(*N*-morpholino)ethanesulfonic acid (Mes) and 3-(*N*-morpholino)-propanesulfonic acid (Mops) are important hydrogen ion buffers for biological media.

This paper describes an investigation of the different complex species of the two biologically important compounds (dibutyltin(IV), M and zwitterionic buffers, L). A systematic study of this complex system has been carried out. The effect of solvent and temperature on the complex formation equilibrium were also investigated. Stoichiometric and stability of the various complex species in the complex system have been examined by pH-metric techniques.

The structure of the compounds under investigations can be represented diagrammatically as follows:

		
<p>2-(<i>N</i>-morpholino)ethanesulfonic acid (Mes) <math>pK_{a2} = 6.06</math> at 25.00 °C physiological pH = 5.50–6.50</p>	<p>3-(<i>N</i>-morpholino)propanesulfonic acid (Mops) <math>pK_{a2} = 7.12</math> at 25.00 °C physiological pH = 6.50–7.90</p>	
		
	<p>Dibutyltin(IV) (DBT)</p>	

## 2 Experimental

### 2.1 Material and Reagents

Dibutyltin(IV) dichloride was received from Aldrich Chemicals Co., Germany (purity 99 %). The ligands (L) used were 2-(*N*-morpholino) ethanesulfonic acid (Mes) and 3-(*N*-morpholino) propanesulfonic acid (Mops); their purities were 98 and 99 %, respectively. These materials were supplied from Flucka Chemicals Co., Germany. Carbonate-free sodium hydroxide stock solutions were prepared by diluting the contents of concentrated volumetric solution vials. These solutions were systematically checked by titration against potassium hydrogen phthalate. These materials (sodium hydroxide, potassium hydrogen phthalate and NaNO<sub>3</sub>) were received from BDH and purified according to literature methods [14, 16]. All solutions were prepared in deionized water.

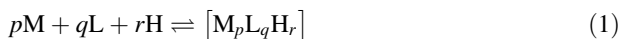
### 2.2 Apparatus

pH titrations were performed using a Metrohm 751 GPD titrino instrument. The titro-processor was calibrated with standard buffer solutions prepared according to [17]. The titrations were carried out in a purified nitrogen atmosphere using a titration vessel described previously [18]. The temperature was maintained constant by a Color Ultra Thermostat.

### 2.3 Procedure and Measuring Techniques

The protonation constants of the ligands were determined by titrating a 40 mL of ligand solution (2.50 mol·L<sup>-1</sup>). The hydrolysis constants of dibutyltin(IV) were determined by titrating a 40 mL of dibutyltin(IV) dichloride (2.50 mol·L<sup>-1</sup>). The formation constant of organotin(IV) complexes was determined by titrating a 40 mL of solution containing the ligand (2.50 mol·L<sup>-1</sup>) and dibutyltin(IV) with concentrations 2.50 and 1.25 mol·L<sup>-1</sup>. The titrations were performed at different temperatures and in dioxane–water mixtures of different compositions. The ionic strength was adjusted to 0.10 mol·L<sup>-1</sup> with NaNO<sub>3</sub>. p*K*<sub>w</sub>s at different temperatures and in dioxane–water solutions were determined as described previously [19]. For this purpose, various amounts of standard NaOH solution were added to a solution containing 0.10 mol·L<sup>-1</sup> NaNO<sub>3</sub>. The [OH<sup>-</sup>] was calculated from the amount of base added. The [H<sup>+</sup>] was calculated from the pH value. The product of [OR] and [H<sup>+</sup>] was taken and the mean values obtained in this way for the log<sub>10</sub> concentration product are log<sub>10</sub> *K*<sub>w</sub> = -14.28, -14.72, -15.13, -15.46, -15.70, -16.21 for 12.50, 25.00, 37.50, 50.00, 62.50 and 75.00 % by mass dioxane–water solutions, and log<sub>10</sub> *K*<sub>w</sub> = -14.25, -14.07, -13.89, -13.74 and -13.69 for 15.00, 20.00, 25.00, 30.00 and 35.00 °C for water solutions, respectively.

The species formed in the systems were characterized by the general equilibrium processes are given by Eq. 1, while the formation constants for these generalized species are given by Eq. 2:



$$\beta_{pqr} = \frac{[M_pL_qH_r]}{[M]^p [L]^q [H]^r} \quad (2)$$

The calculations were performed using the computer program [20] MINIQUAD-75 loaded on a Pentium II–233 computer. The stoichiometric and stability constants of the complexes formed were determined by trying various possible composition models for the system. The model was selected to give the best statistical fit and be chemically consistent with the titration data without giving any systematic drifts in the magnitudes of various residuals, as described elsewhere [20]. The concentration distribution diagrams were obtained using the program SPECIES [21].

### 3 Results and Discussion

#### 3.1 Solution-Phase Investigations

The dibutyltin(IV) cation is known [19, 22–26] to form stable water soluble mono- and polynuclear hydroxide species (Tables 1 and 2) in the whole pH range studied. Since the hydroxide ion and the ligands are in strong competition for the metal ion, these species were always taken into consideration in the equilibrium systems.

The acid dissociation constants of Mes and Mops were determined under the same experimental conditions of ionic strength and temperature (Tables 1 and 2). These constants should be taken into consideration during the evaluation of the pH-metric data. It is found that  $\log_{10} \beta_{011} = 6.06$  and  $7.12$  for Mes and Mops, respectively [27]. The protonation constant for the zwitterionic buffers (Good's buffers) clearly indicate the different basicity of tertiary amine. This is due to the substitution of increasingly larger alkyl chains between the tertiary amine group and the sulfonate group, the addition of hydroxyl groups on the molecule, or other modifications to the backbone N-substituted structure.

The titration curves for the complex solutions with (1:1 and 2:1) ligand-to-metal ratio clearly indicate the formation of different kinds of complexes (Fig. 1). The best fit of the titration curves was obtained when complexes ML, MLH<sub>-1</sub>, MLH<sub>-2</sub> and MLH<sub>-3</sub> were included beside the hydrolysis products of the dibutyltin(IV) cation (Tables 1 and 2).

The potentiometric titration curves of dibutyltin(IV) complexes against both of Mes and Mops with different stoichiometric species (1:1 and 1:2, metal:ligand) of these systems started at pH  $\sim 3.0$ . At this pH the sulphonate group was already deprotonated and does not play a role in the mode of coordination with the DBT(IV) anion. With increasing pH Mes and Mops react with DBT(IV) as monodentate ligands forming deprotonated complexes ML. ML competes with the hydroxo complex at lower pH  $\sim 3.0$ . At higher pHs further deprotonation was observed leading to form mixed-hydroxo complex species MLH<sub>-1</sub>, MLH<sub>-2</sub>, and MLH<sub>-3</sub>.

#### 3.2 Effect of Temperature

The thermodynamic parameters  $\Delta H^*$ ,  $\Delta S^*$  and  $\Delta G^*$  were obtained by a linear least-squares fit of  $\log_{10} K$  versus  $1/T$  leading to an intercept  $\Delta S^*/R$  and a slope  $-\Delta H^*/R$ . The results obtained are summarized in Table 3 and explained as follows:

- (a) The formation constants for the hydrolyzed species of dibutyltin(IV) are discussed in the previous work [19].
- (b) The protonation reactions of the tertiary amine site of Mes are exothermic and the entropy is positive. This means that the protonation constant of Mes is ordered and favored with increasing temperature.

**Table 1** Formation constants of dibutyltin(IV)–Mes complexes in water at different temperatures

System <sup>a</sup>	Temperature (°C)	$p^b$	$q^b$	$r^b$	$\log_{10} \beta^c$	$S^d$	
DBT	15.00	1	0	–1	–2.50 (0.01)	$4.4 \times 10^{-8}$	
		1	0	–2	–8.00 (0.01)		
		1	0	–3	–29.50 (0.03)		
		2	0	–4	–9.10 (0.01)		
		2	0	3	–14.21 (0.02)		
DBT–Mes	15.00	0	1	1	4.30 (0.01)	$2.1 \times 10^{-8}$	
		1	1	0	6.51 (0.01)	$1.6 \times 10^{-8}$	
		1	1	–1	2.16 (0.01)		
		1	1	–2	–3.10 (0.02)		
		1	1	–3	–11.60 (0.01)		
DBT	20.00	1	1	–1	–3.81 (0.01)	$4.2 \times 10^{-8}$	
		1	1	–2	–7.81 (0.01)		
		1	1	–3	–28.10 (0.01)		
		2	1	–4	–8.91 (0.03)		
		2	1	–2	–13.98 (0.01)		
DBT–Mes	20.00	1	1	–1	4.60 (0.01)	$1.6 \times 10^{-8}$	
		1	1	–2	6.83 (0.01)	$1.4 \times 10^{-8}$	
		1	1	–3	2.50 (0.01)		
		1	1	–4	–3.61 (0.03)		
DBT	25.00	1	1	–2	–12.00 (0.02)	$3.7 \times 10^{-8}$	
		1	0	–1	–4.01 (0.01)		
		1	0	–2	–8.30 (0.01)		
		1	0	–3	–28.62 (0.02)		
		1	0	–4	–9.30 (0.01)		
DBT–Mes	25.00	1	0	–3	–14.61 (0.02)	$1.3 \times 10^{-8}$	
		0	1	–1	4.81 (0.01)		$1.3 \times 10^{-8}$
		1	1	–2	7.30 (0.01)		$1.0 \times 10^{-8}$
		1	1	–3	3.22 (0.01)		
		1	1	–4	–4.21 (0.02)		
DBT	30.00	2	1	–3	–12.45 (0.02)	$3.3 \times 10^{-8}$	
		1	0	5	–4.83 (0.01)		
		1	0	–1	–9.21 (0.01)		
		1	0	–2	–28.80 (0.02)		
		2	0	–3	–9.62 (0.03)		
DBT–Mes	30.00	2	0	–4	–15.00 (0.01)	$7.3 \times 10^{-9}$	
		1	1	–1	5.20 (0.01)		$7.3 \times 10^{-9}$
		1	1	–2	7.60 (0.01)		$8.8 \times 10^{-9}$
		1	1	–3	3.82 (0.02)		
		1	1	–4	–4.50 (0.03)		
DBT–Mes	30.00	1	1	–3	–12.88 (0.02)		
		1	1	–3	–12.88 (0.02)		

**Table 1** continued

System <sup>a</sup>	Temperature (°C)	$p^b$	$q^b$	$r^b$	$\log_{10} \beta^c$	$S^d$
DBT	35.00	1	1	-1	-6.20 (0.01)	$3.7 \times 10^{-8}$
		1	0	3	-8.30 (0.01)	
		1	0	-1	-25.30 (0.01)	
		2	0	-3	-8.65 (0.02)	
		1	0	-1	14.30 (0.01)	
DBT–Mes		1	1	-4	6.20 (0.01)	$7.9 \times 10^{-9}$
		1	1	-1	8.00 (0.01)	
		1	1	-2	-5.20 (0.01)	
		1	1	-3	-12.40 (0.01)	
		1	1	-4	-6.00 (0.00)	

<sup>a</sup> Mes is 2-(*N*-morpholino)ethanesulfonic acid

<sup>b</sup>  $p$ ,  $q$ ,  $r$  are the stoichiometric coefficient corresponding to dibutyltin(IV), Good's buffers and  $H^+$ , respectively

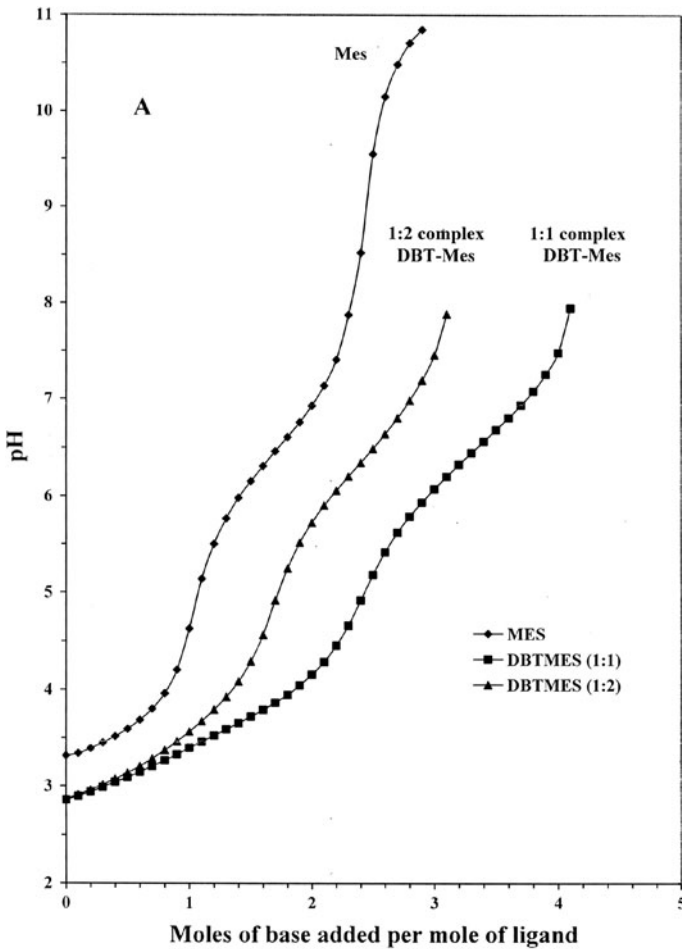
<sup>c</sup> Standard deviations are given in parentheses

<sup>d</sup> Sum of square of residuals

**Table 2** Formation constants of dibutyltin(IV)–Mops complexes in water at different temperature

System	Temperature (°C)	$p$	$q$	$r$	$\log_{10} \beta$	$S$
DBT–Mops	15.00	1	0	1	-3.46 (0.01)	$1.8 \times 10^{-7}$
		1	0	2	-8.01 (0.01)	
		1	0	-3	-25.40 (0.03)	
		2	0	-2	-4.52 (0.01)	
		2	0	-4	-14.11 (0.01)	
DBT–Mops	20.00	0	0	-1	-3.30 (0.01)	$2.4 \times 10^{-7}$
		1	0	-2	-7.60 (0.01)	
		1	0	-3	-23.65 (0.02)	
		1	0	-2	-3.38 (0.01)	
		1	0	-4	-12.98 (0.01)	
DBT–Mops	25.00	0	1	1	-3.03 (0.01)	$7.5 \times 10^{-8}$
		1	1	0	-6.68 (0.01)	
		1	1	-1	-23.06 (0.01)	
		1	1	-2	-3.08 (0.01)	
		1	1	-3	-12.61 (0.01)	
DBT–Mops	30.00	0	1	1	-2.56 (0.01)	$8.3 \times 10^{-8}$
		1	1	0	-6.12 (0.03)	
		1	1	-1	-22.66 (0.01)	
		1	1	-2	-2.76 (0.01)	
		1	1	-3	-11.51 (0.01)	

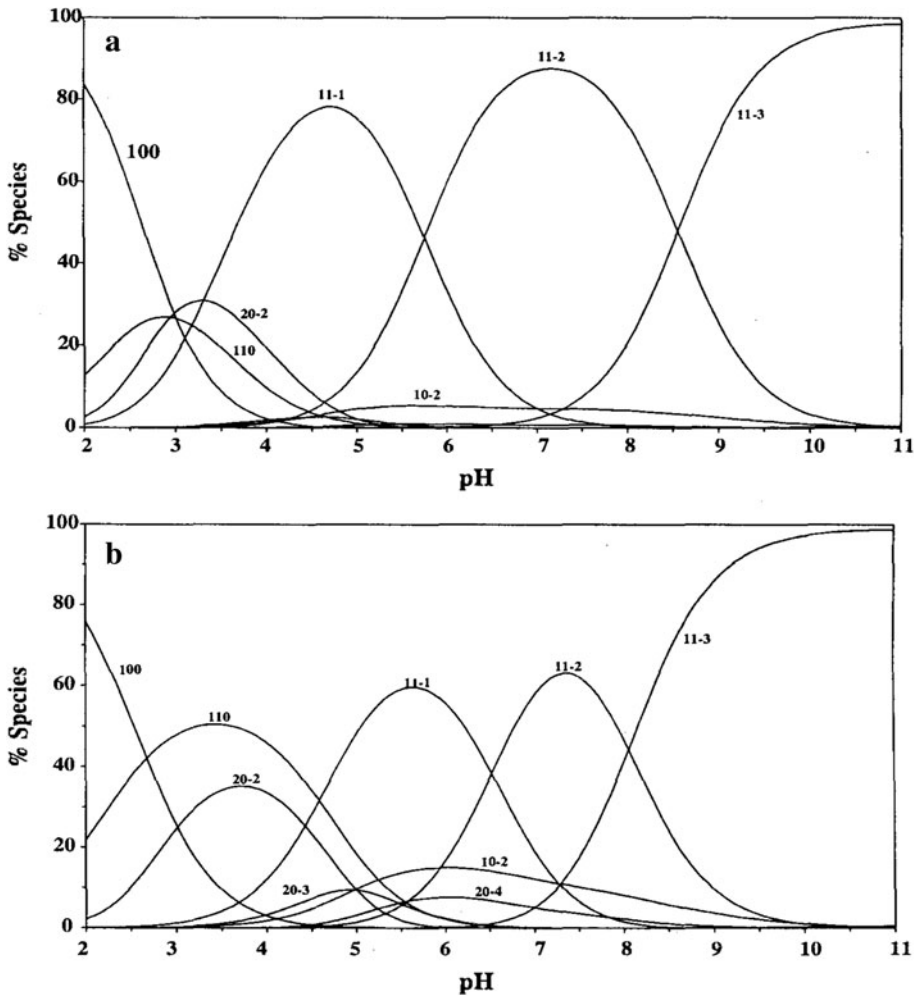
(c) The complex-formation of Mes with DBT is exothermic for ML,  $MLH_{-1}$  and  $MLH_{-2}$  species and endothermic for  $MLH_{-3}$  species. This may be explained statistically based on the presumption that more coordination sites are available for binding ML,  $MLH_{-1}$  and  $MLH_{-2}$  species (Fig. 2). The formation of the  $MLH_{-1}$  species is more exothermic than for the ML species. This means that the hydroxide complex is more



**Fig. 1** Potentiometric titration curves of DBT-Mes in water

**Table 3** Thermodynamic parameters for the equilibrium of dibutyltin(IV) complexes in water

Equilibrium	$\Delta H^\circ$ (kJ·mol <sup>-1</sup> )	$\Delta S^\circ$ (J·K <sup>-1</sup> ·mol <sup>-1</sup> )	$\Delta G^\circ$ (kJ·mol <sup>-1</sup> )
DBT-Mes			
$M^{2+} + L^- \rightleftharpoons ML^+$	-37.5 (0.30)	-25 (1)	-30.6 (0.6)
$ML^+ + OH^- \rightleftharpoons MLH_{-1}$	-58.2 (0.01)	-160 (1)	-20.5 (0.5)
$MLH_{-1} + OH^- \rightleftharpoons MLH_{-2}$	-25.3 (0.01)	36 (1)	-32.8 (0.3)
$MLH_{-2} + OH^- \rightleftharpoons MLH_{-3}$	-34.2 (0.03)	230 (1)	-40.1 (0.1)
DBT-Mops			
$M^{2+} + L^- \rightleftharpoons ML^+$	-40.2 (0.30)	-31 (2)	-40.1 (0.3)
$ML^+ + OH^- \rightleftharpoons MLH_{-1}$	-65.3 (0.10)	-172 (1)	-25.3 (0.1)
$MLH_{-1} + OH^- \rightleftharpoons MLH_{-2}$	-36.5 (0.30)	25 (2)	-35.6 (0.5)
$MLH_{-2} \rightleftharpoons MLH_{-3}$	-35.2 (0.10)	220 (1)	-50.3 (0.1)



**Fig. 2** Species distribution curves in the DBT(IV)-Mes (a) and DBT(IV)-Mops (b) systems.  $[M] = 2.50 \text{ mol}\cdot\text{L}^{-1}$ ,  $[L] = 2.50 \text{ mol}\cdot\text{L}^{-1}$ . The notation of the different species corresponds to the  $pqr$  values of the corresponding complex  $MpLqHr$

favored than the deprotonated complex species, due to the high affinity of dibutyltin(IV) to form hydroxide complex.

### 3.3 Effect of Solvent

Traditionally, water has been considered as the solvent which best represents biological conditions. Although this is a general assumption, a lower polarity has been detected in some biochemical micro-environments, such as active sites of enzymes and side chains in proteins, sometimes hidden in lower dielectric constant cavities [28–32]. In these cases, the selection of other solvents seems more appropriate in order to emulate properly the medium's real features. Careful examination of the medium effect on the equilibrium constants, Table 4, is summarized in the following:



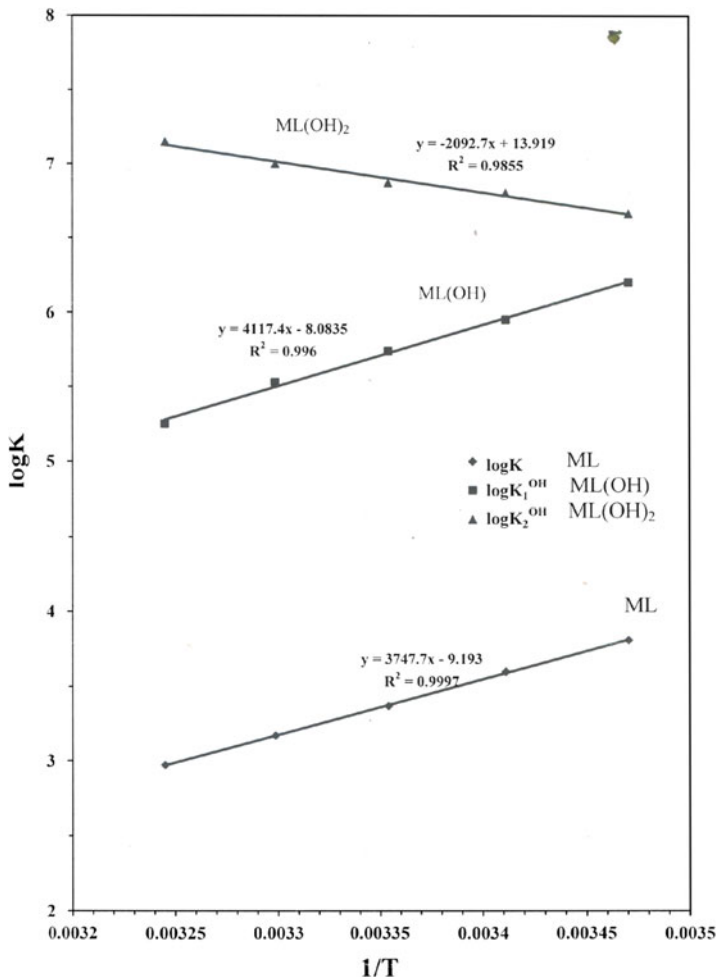
**Table 4** Formation constants of dibutyltin(IV) complexes in 75.00 mass-% dioxane–25.00 mass-% water solutions at different compositions

System	Dioxane (%)	<i>p</i>	<i>q</i>	<i>r</i>	$\log_{10} \beta$	<i>S</i>
DBT	12.50	1	0	-1	-3.1 (0.01)	$1.2 \times 10^{-7}$
		1	0	-2	-8.30 (0.01)	
		1	0	-4	-29.01 (0.02)	
		2	0	-2	-3.50 (0.01)	
		2	0	-3	-7.88 (0.02)	
DBT–Mes		0	1	1	5.12 (0.01)	$3.1 \times 10^{-8}$
		1	1	0	4.31 (0.01)	$2.8 \times 10^{-9}$
		1	1	-1	1.98 (0.01)	
		1	1	-2	-3.40 (0.01)	
		1	1	-3	-10.98 (0.02)	
DBT–Mops		0	1	-1	7.63 (0.01)	$3.6 \times 10^{-8}$
		1	1	-2	4.68 (0.01)	$2.1 \times 10^{-9}$
		1	1	-3	2.10 (0.01)	
		1	1	-4	-4.61 (0.02)	
DBT	25.00	1	0	-1	-3.65 (0.01)	$1.0 \times 10^{-8}$
		1	0	-2	-8.87 (0.01)	
		1	0	-3	-30.20 (0.02)	
		2	0	-4	-3.81 (0.03)	
		2	0	-2	-8.20 (0.02)	
DBT–Mes		0	1	1	6.12 (0.01)	$4.2 \times 10^{-8}$
		1	1	0	5.30 (0.01)	$3.6 \times 10^{-9}$
		1	1	-1	2.12 (0.01)	
		1	1	-2	-4.5 (0.01)	
		1	1	-3	-11.12 (0.03)	
DBT–Mops		1	1	0	7.66 (0.01)	$5.68 \times 10^{-8}$
		1	1	-1	6.12 (0.01)	$3.8 \times 10^{-9}$
		1	1	-2	3.12 (0.01)	
		1	1	-3	-6.13 (0.02)	
DBT	37.50	1	0	-1	-4.20 (0.01)	$1.5 \times 10^{-8}$
		1	0	-2	-9.10 (0.01)	
		1	0	-3	-31.0 (0.02)	
		1	0	-4	-4.21 (0.03)	
		2	0	-2	-8.41 (0.03)	
DBT–Mes		1	1	1	6.50 (0.01)	$4.1 \times 10^{-8}$
		1	1	0	6.12 (0.01)	$2.1 \times 10^{-9}$
		1	1	-1	3.0 (0.02)	
		1	1	-2	4.80 (0.01)	
		1	1	-3	11.50 (0.03)	
DBT – Mops		1	1	0	7.80 (0.01)	$4.8 \times 10^{-8}$
		1	1	-1	6.40 (0.01)	$3.6 \times 10^{-9}$
		1	1	-2	3.16 (0.02)	
		1	1	-3	-6.90 (0.03)	
		1	1	-4	-13.70 (0.01)	

**Table 4** continued

System	Dioxane (%)	$p$	$q$	$r$	$\log_{10} \beta$	$S$
DBT	50.00	0	1	1	-4.6 (0.01)	$1.8 \times 10^{-8}$
		1	1	0	-9.36 (0.01)	
		1	1	-1	-32.10 (0.01)	
		1	1	-2	-4.61 (0.02)	
		1	1	-3	-8.80 (0.01)	
DBT–Mes		1	1	-1	6.80 (0.01)	$5.1 \times 10^{-8}$
		1	1	-2	6.88 (0.01)	$2.4 \times 10^{-9}$
		2	1	-3	3.61 (0.02)	
		1	1	-4	-5.20 (0.01)	
		1	1	-2	-12.10 (0.03)	
DBT–Mops		1	1	-1	6.85 (0.01)	$5.3 \times 10^{-8}$
		1	1	-2	7.10 (0.01)	$3.8 \times 10^{-9}$
		1	1	-3	3.91 (0.02)	
		2	1	-4	-5.50 (0.01)	
		2	1	-1	-12.60 (0.03)	
DBT	62.50	1	0	-1	-4.88 (0.01)	$2.2 \times 10^{-8}$
		1	0	-2	-9.62 (0.01)	
		1	0	-3	-32.33 (0.02)	
		2	0	-4	-4.31 (0.01)	
		2	0	-2	-9.12 (0.03)	
DBT–Mes		0	1	1	6.88 (0.01)	$5.4 \times 10^{-8}$
		1	1	0	7.12 (0.01)	$2.8 \times 10^{-9}$
		1	1	-1	3.82 (0.02)	
		1	1	-2	-5.51 (0.01)	
		1	1	-3	-12.31 (0.03)	
DBT–Mops		1	1	1	7.21 (0.01)	$6.1 \times 10^{-8}$
		1	1	0	7.56 (0.01)	$4.0 \times 10^{-9}$
		1	1	-1	4.20 (0.02)	
		2	1	-2	-5.81 (0.03)	
		2	1	-3	-2.81 (0.02)	
DBT	75.00	1	0	-1	-5.10 (0.01)	$2.6 \times 10^{-8}$
		1	0	-2	-9.82 (0.01)	
		1	0	-3	-32.50 (0.02)	
		2	0	-4	-4.66 (0.01)	
		2	0	-2	-9.54 (0.03)	
DBT–Mes		0	1	-1	7.20 (0.01)	$6.1 \times 10^{-8}$
		1	1	-2	7.51 (0.01)	$3.2 \times 10^{-9}$
		1	1	-3	4.12 (0.01)	
		1	1	-4	-5.60 (0.02)	
		2	1	-3	-12.65 (0.02)	
DBT–Mops		1	1	-1	7.80 (0.01)	$6.6 \times 10^{-8}$
		1	1	-2	8.20 (0.01)	
		1	1	-3	4.65 (0.01)	
		1	1	-4	-6.20 (0.02)	
		1	1	-2	-13.0 (0.01)	

- (a)  $\log_{10} \beta_{011}$  of Mes and Mops (tertiary amine), as well as the hydrolysis constants of dibutyltin(IV), decrease linearly with the increase of dioxane proportion in the medium, This may be correlated with the ability of a solvent of relatively low dielectric constant to decrease the electrostatic forces between the proton and ligand anions in the case of ligand dissociation and that between the proton and hydrolyzed form of organotin(IV).
- (b) The variation of the stability constants of the DBT complexes with Mes as a function of solvent composition is shown in Fig. 2. The stability constants of the  $MLH_{-1}$  species with Mes and Mops increase with increasing dioxane proportion in the mixed solvent. On the other hand the amount of  $ML$ ,  $MLH_{-2}$ ,  $MLH_{-3}$  species decrease with increasing dioxane proportion in the mixed solvent. This may be explained on the premise that the  $MLH_{-1}$  species involves neutralization of charges.



**Fig. 3** Effect of temperature on  $\log_{10} K$  of the DBT–Mes complex in aqueous solutions

## 4 Conclusion

The formation equilibrium of DBT(IV) complexes involving zwitterionic buffer (Mes and Mops) were investigated at different temperatures and variation of water–dioxane proportion. The effect of dioxane as a solvent on the formation constants of DBT–Mes and DBT–Mops complexes decrease linearly with the increase of temperature in the medium (Fig. 3).

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