


Review

Targeting Chaperone/Co-Chaperone Interactions with Small Molecules: A Novel Approach to Tackle Neurodegenerative Diseases

Lisha Wang ¹, Liza Bergkvist ¹, Rajnish Kumar ^{1,2}, Bengt Winblad ^{1,3} and Pavel F. Pavlov ^{1,*}

¹ Department of Neuroscience Care and Society, Division of Neurogeriatrics, Karolinska Institutet, 17164 Solna, Sweden; lisha.wang@ki.se (L.W.); lizabergkvist@gmail.com (L.B.); rajnish.kumar@ki.se (R.K.); bengt.winblad@ki.se (B.W.)

² Department of Pharmaceutical Engineering & Technology, Indian Institute of Technology (BHU), Varanasi 221005, India

³ Theme Inflammation and Aging, Karolinska University Hospital, 14186 Huddinge, Sweden

* Correspondence: pavel.pavlov@ki.se; Tel.: +46-761929700

Abstract: The dysfunction of the proteostasis network is a molecular hallmark of neurodegenerative diseases such as Alzheimer's disease, Parkinson's disease, Huntington's disease, and amyotrophic lateral sclerosis. Molecular chaperones are a major component of the proteostasis network and maintain cellular homeostasis by folding client proteins, assisting with intracellular transport, and interfering with protein aggregation or degradation. Heat shock protein 70 kDa (Hsp70) and 90 kDa (Hsp90) are two of the most important chaperones whose functions are dependent on ATP hydrolysis and collaboration with their co-chaperones. Numerous studies implicate Hsp70, Hsp90, and their co-chaperones in neurodegenerative diseases. Targeting the specific protein–protein interactions between chaperones and their particular partner co-chaperones with small molecules provides an opportunity to specifically modulate Hsp70 or Hsp90 function for neurodegenerative diseases. Here, we review the roles of co-chaperones in Hsp70 or Hsp90 chaperone cycles, the impacts of co-chaperones in neurodegenerative diseases, and the development of small molecules modulating chaperone/co-chaperone interactions. We also provide a future perspective of drug development targeting chaperone/co-chaperone interactions for neurodegenerative diseases.

Keywords: Hsp70; Hsp90; co-chaperones; neurodegenerative diseases; small molecules



Citation: Wang, L.; Bergkvist, L.; Kumar, R.; Winblad, B.; Pavlov, P.F. Targeting Chaperone/Co-Chaperone Interactions with Small Molecules: A Novel Approach to Tackle Neurodegenerative Diseases. *Cells* **2021**, *10*, 2596. <https://doi.org/10.3390/cells10102596>

Academic Editor: Alexander E. Kabakov

Received: 8 September 2021

Accepted: 25 September 2021

Published: 29 September 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Proteins perform different functions that are essential for the physiology of an organism. The proteostasis network maintains the health of the proteome by controlling the protein synthesis, folding, trafficking, disaggregation, and degradation [1]. The dysfunction of the proteostasis network is a hallmark of neurodegenerative diseases, such as Alzheimer's disease (AD), Parkinson's disease (PD), Huntington's disease (HD), and amyotrophic lateral sclerosis (ALS) [2]. Molecular chaperones play an important role in the proteostasis network. They maintain cellular homeostasis by folding client proteins, assisting with intracellular transport, interfering with protein aggregation, or directing misfolded or aggregated proteins to cellular clearance pathways. Heat shock protein 70 kDa (Hsp70) and 90 kDa (Hsp90) are two of the most important chaperone families whose functions are dependent on ATP hydrolysis to provide energy and collaboration with their co-chaperones to form the dynamic complex. Healthy proteostasis is especially important for virtually non-dividing neurons in the adult brain; it is not surprising that many studies have implicated Hsp70, Hsp90, and their co-chaperones in neurodegenerative diseases [3–5]. Modulation of the functions of Hsp70 and Hsp90 has been regarded as an attractive pharmacological intervention for neurodegenerative diseases.

Although much work has gone into the development of Hsp70 or Hsp90 inhibitors in cancer, none of them have had success in clinical trials owing to their adverse effects. Most inhibitors were discovered to completely inhibit chaperone functions by occupying the N-terminal ATP binding pocket [6,7]. However, Hsp70 and Hsp90 regulate a large spectrum of client proteins and inhibiting ATPase function can also interfere with all of their client proteins, leading to undesirable/toxic effects, making this an unsuccessful strategy. Moreover, the inhibition of Hsp90 ATPase activity triggers a compensatory mechanism resulting in a heat shock response [8]. Another option is to target the Hsp90 C-terminus, which does not induce detrimental heat shock response. The C-terminal inhibitors have been well-summarized in recent reviews and employ multiple mechanisms including inhibition of C-terminal dimerization, induction of chaperone conformational cycles, and modulation of the interaction with co-chaperones [9,10]. Among them, dihydropyridine derivatives have shown neuroprotective effects *in vivo* [11,12]. However, this approach has been limited owing to the lack of complete structural information and exact binding sites in the C-terminus [13].

Therefore, alternative approaches to modulate Hsp70 or Hsp90 are needed. Numerous co-chaperones associate with Hsp70 or Hsp90 at different stages of chaperone cycles and appear to be dependent on the substrates involved in neurodegenerative diseases. Targeting the protein–protein interactions (PPIs) between chaperones and their particular partner co-chaperones provides the opportunity to specifically modulate Hsp70 or Hsp90 function in neurodegenerative diseases. In this review, we summarize the knowledge of the involvement of co-chaperones in Hsp70 or Hsp90 chaperone cycles, the functions of co-chaperones in neurodegenerative diseases, and the development of small molecules that modulate chaperone/co-chaperone PPIs. We also discuss the dicarboxylate clamp mechanism in the interactions of Hsp90 C-terminal Met-Glu-Glu-Val-Asp (MEEVD) peptide and tetratricopeptide repeat (TPR) domain of co-chaperones and provide useful insights for future drug development targeting chaperone/co-chaperone interactions for neurodegenerative diseases.

2. Hsp70 and Its Co-Chaperones

2.1. Hsp70 Structure and Chaperone Cycle

Hsp70 consists of two domains connected by a flexible linker: an N-terminal ATP binding domain (NBD) and a C-terminal substrate/client binding domain (SBD). SBD is divided into a β subdomain (SBD β), hosting the substrate binding site, and an α subdomain (SBD α), forming a lid lock structure to trap client proteins. Hsp70 function is dependent on the coordinated activity of all three domains [14] (Figure 1). When ATP binds the NBD, the lid formed by SBD α opens and client proteins are now able to bind in the hydrophobic pocket found in the SBD β . Hydrolysis of ATP results in the closing of the C-terminal lid (SBD α) and trapping of the client protein. Release of ADP opens the C-terminal lid and client proteins are able to leave the SBD. However, the ability of Hsp70 to hydrolyze ATP is intrinsically low. To facilitate this process, the co-chaperones of Hsp70 step in, such as J-proteins (also called Hsp40s) and nucleotide exchange factors (NEFs). J-proteins all share a 70 amino acid stretch, the J-domain, which interacts with Hsp70 NBD, the inter-domain linker, and SBD β to stimulate its ATPase activity and facilitate client capture [15,16]. NEFs can be categorized into four distinct families: GrpE, Hsp110, HspBP1, and Bag proteins [17]. They bind to the NBD of Hsp70 to accelerate the release of ADP and, in some cases, foster client dissociation. Humans have 13 Hsp70s, 41 J-proteins, and 13 NEFs, which play a major role in driving the multiple functions of Hsp70 [18]. Taken together, the co-chaperones of Hsp70 are thought to modulate the enzymatic activity of the chaperone and help to guide its interactions with client proteins. Therefore, inhibiting the interaction between Hsp70 and its specific co-chaperones could be a therapeutic strategy to target specific Hsp70 functions, thus avoiding adverse effects arising from the general inhibition of Hsp70 function.

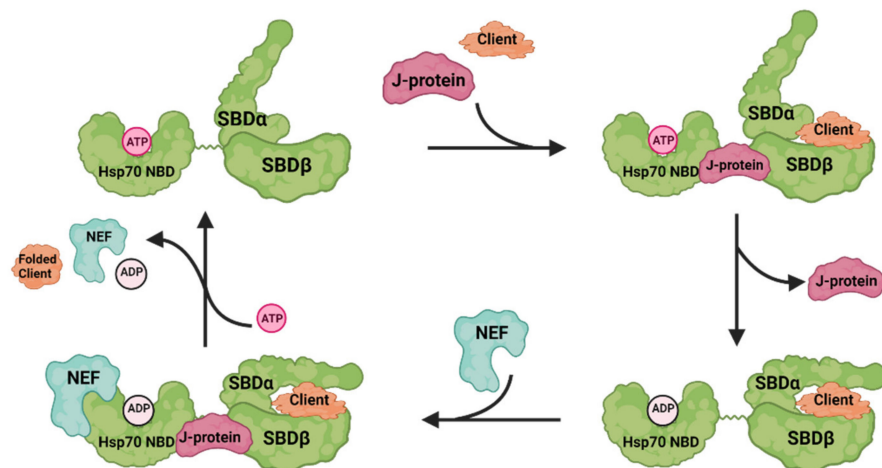


Figure 1. A schematic illustration of the Hsp70 chaperone cycle. Hsp70 consists of two domains connected by a flexible linker: an N-terminal ATP binding domain (NBD) and a C-terminal substrate/client binding domain (SBD), which is divided into a β subdomain (SBD β), hosting the substrate binding site, and an α subdomain (SBD α), forming a lid lock structure to trap clients. When ATP binds to the NBD, the lid formed by SBD α opens. Co-chaperone J-protein binds to Hsp70 to stimulate its ATPase activity and facilitate client binding in the hydrophobic pocket of SBD β . Hydrolysis of ATP results in the closing of the C-terminal lid (SBD α) and trapping of the client. Another co-chaperone, nucleotide exchange factor (NEF), binds to the NBD of Hsp70 to accelerate the release of ADP. Then, SBD α opens and the folded client is able to leave Hsp70. Created with BioRender.com.

2.2. Hsp70 and Its Co-Chaperones in Neurodegenerative Diseases

The role of Hsp70 in protein homeostasis connects it closely to protein misfolding and neurodegenerative diseases like AD, PD, HD, and prion disease [5]. In addition to re-folding misfolded proteins, Hsp70 has been shown to inhibit A β aggregation, as well as promote degradation of A β and tau aggregates via the proteasomal system [5]. On the contrary, the constitutively expressed Hsc70, one of Hsp70 family members, prevents tau degradation, and an imbalance in the Hsp70 family and their co-chaperones promotes tau pathology [19]. Additional potential pathogenic involvement of Hsp70 in protein misfolding diseases was shown by Fontaine and colleagues: Hsp70 in complex with its co-chaperone J-protein DnaJC5 controls the extracellular release of disease-associated proteins like tau, α -synuclein, and transactive response DNA-binding protein 43 (TDP-43), providing a mechanism for how aggregation-prone proteins are able to leave the cell and possibly propagate [20]. It was recently shown that Hsp70 in complex with its co-chaperones, the J-proteins DNAJB1 and HSPA4 and an Hsp110-type NEF, disassembles tau fibrils *in vitro*, which results in the release of seeding potent oligomeric tau [21]. This suggests that the anti-aggregation properties of Hsp70 are a two-edged sword, as it eliminates tau fibrils at the cost of generating new seeds. Therefore, potential future treatment strategies for neurodegenerative, protein misfolding diseases could be identifying small molecules targeting specific Hsp70 isoforms and their co-chaperones that contribute to pathogenesis.

2.3. Hsp70 Co-Chaperone Interaction Inhibitors

No specific, competitive inhibitors have been developed for Hsp70 and its co-chaperones. Thus far, the Hsp70 inhibitors explored have either been targeting ATPase activity or resulted in allosteric inhibition. Allosteric inhibitors, i.e., non-competitive inhibitors, of Hsp70 can be divided into three classes [22]. For two of these classes of Hsp70 allosteric inhibitors (exemplified by VER-155008 and YK5), few studies have shown any effect on Hsp70 co-chaperone interactions. The third class of allosteric inhibitors are based on MKT-077 (1, Figure 2) (with next generation molecules like YM-1 and JG-98) and have been shown

to impact chaperone/co-chaperone interactions. Structural studies showed that MKT-077 and analogues bind to a highly conserved hydrophobic pocket next to the NBD, with binding being favoured in the ADP-bound state of Hsp70. Although MKT-077 analogues bind $>20\text{\AA}$ away from the Hsp70–NEF interaction surface, they trap a conformational state that disfavors specific co-chaperone interaction, preventing the NEF family of Bag proteins from binding [22]. **YM-1** (2, Figure 2) and **JG-98** (3, Figure 2) were both shown to inhibit Hsp70–Bag3 interactions by a pull-down assay, and their treatments are sufficient to suppress tumor growth in mice that mirrors the effects of Hsp70 depletion [23,24].

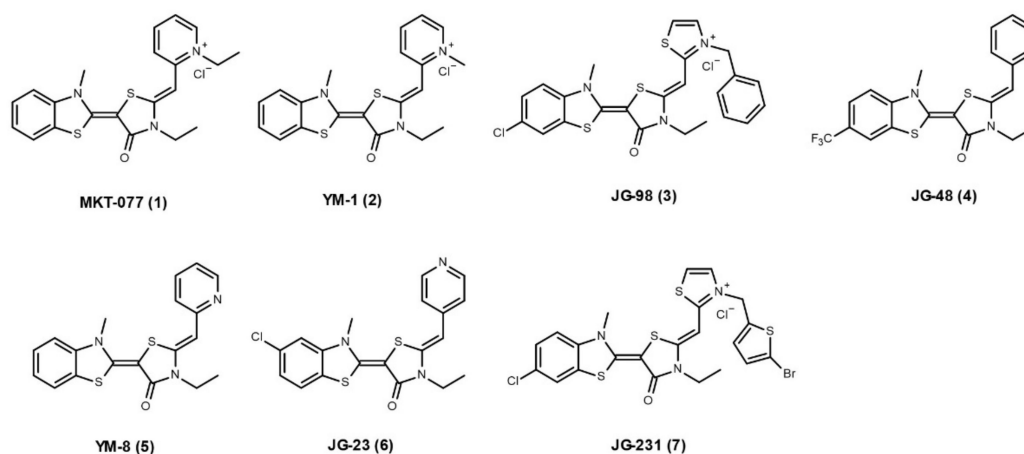


Figure 2. Small molecules with the ability to disrupt Hsp70/co-chaperone interactions.

The role of inhibiting the interactions between Hsp70 and its co-chaperones in the context of neurodegenerative diseases has also been investigated. **JG-48** (4, Figure 2) is an analogue of YM-1, developed to not interfere with Hsp70 chaperone activity. Using this compound, Young et al., observed an increase in tau turnover *in vitro* [25]. JG-48 is partially able to inhibit the NEF family of Bag proteins from interacting with Hsp70. This results in a stabilized Hsp70–tau complex, subsequently initiating degradation of tau. Another YM-1 analogue, **YM-8** (5, Figure 2), is able to penetrate the blood–brain barrier and decrease tau phosphorylation in cultured brain slices [26]. In a recent study, Shao et al. further improved the pharmacokinetic properties of YM-8, resulting in the new compound **JG-23** (6, Figure 2) [27]. JG-23 is 12 times more stable than YM-8, while still retaining promoting tau degradation *in vitro*.

Other MKT-077 analogues, e.g., **JG-231** (7, Figure 2), have also been shown to inhibit the interaction between Hsp70 and J-proteins. In a recent study by Bengoechea and colleagues, they showed that both genetical and pharmacological inhibition (JG-231) of the interaction between Hsp70 and DnaJB6 was beneficial in a mouse model of muscular dystrophy [28]. However, none of these allosteric inhibitors have been translated into the clinic; only MKT-077 has been tested in a clinical trial setting, where it failed owing to renal toxicity in phase I.

3. Hsp90 and Its Co-Chaperones

3.1. Hsp90 Structure and Chaperone Cycles

Hsp90 serves as a platform for folding and maturation of many client proteins, such as steroid hormone receptors (SHRs), protein kinases, transcription factors, and E3 ubiquitin ligases. To fulfil its function, Hsp90 homodimer undergoes ATP-regulated conformational rearrangements. Hsp90 is composed of three domains: the N-terminal domain (NTD), the middle domain (MD), and the C-terminal domain (CTD). The Hsp90 NTD has the ATP binding pocket, the MD is important for ATP hydrolysis and binding with the client proteins, while the CTD ends in an MEEVD motif that is responsible for interacting with the TPR domains present in a subgroup of Hsp90 co-chaperones [29]. In the absence of

ATP, Hsp90 homodimer adopts an open V-shaped conformation. Upon ATP binding to the NTD, the N-terminal lids close over the bound ATP, leading to the intermediate state. Further structural rearrangements induce the NTD dimerize to form the closed 1 state and then associate with the MD to form the closed 2 state. After ATP hydrolysis by the residues from Hsp90 MD, Hsp90 returns to the open conformation and ADP and inorganic phosphate (Pi) are released.

The dynamic conformational changes of Hsp90 are regulated by a set of Hsp90 co-chaperones (Table 1). Complex formation between Hsp90 and its clients has been studied most extensively for SHRs [30] (Figure 3A). At first, Hsp70–Hsp90 organizing protein (Hop, also known as stress-inducible phosphoprotein 1, Sti1) facilitates unfolded SHRs transferred from Hsp70 to Hsp90 and binds to the open conformation of Hsp90 and inhibits Hsp90 ATPase activity. The co-chaperone with a peptidyl-prolyl *cis-trans* isomerase (PPIase) domain, like the FK506 binding protein (FKBP) 51 or FKBP52, binds to the other Hsp90 TPR-acceptor site to form an asymmetric complex. The binding of ATP results in the intermediate state of Hsp90. Activator of Hsp90 ATPase homolog 1 (Aha1) promotes the formation of the closed 1 state and accelerates Hsp90 ATPase activity. Co-chaperone p23 (Sba1 in yeast) competes with Aha1 for binding to Hsp90 and stabilizes the closed 2 state by reducing Hsp90 ATPase activity. After ATP hydrolysis, p23, the PPIase, and the folded client are released from Hsp90. For protein kinases, the Hsp90 cycle (Figure 3B) is described specifically below (see Section 3.2.1). Cell division cycle 37 (Cdc37) recognizes and delivers an extensive range of protein kinases to Hsp90, which mediates kinase maturation. Protein phosphatase 5 (PP5, also known as Ppt1 in yeast) dephosphorylates Cdc37 at the late stage, leading Hsp90 to adopt an open state and repeat the chaperone cycle.

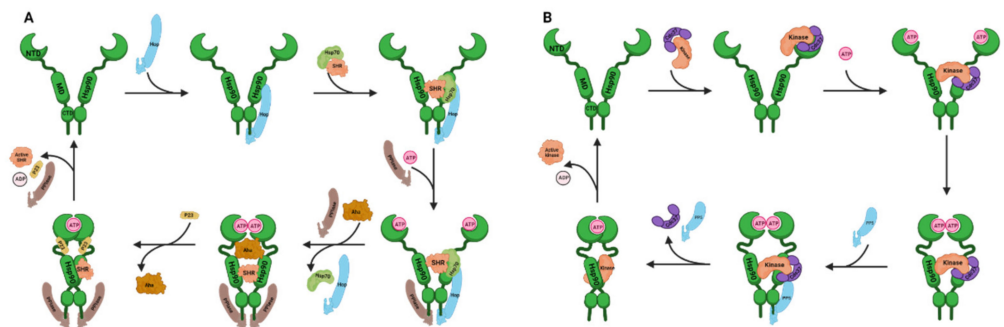


Figure 3. (A) Schematic illustration of Hsp90 chaperone cycles. Hsp90 includes three domains: the N-terminal domain (NTD), the middle domain (MD), and the C-terminal domain (CTD). Hsp90 homodimer undergoes ATP-regulated conformational rearrangements. (A) Hsp90-driven activation of steroid hormone receptors (SHRs). Hsp70–Hsp90 organizing protein (Hop, also known as stress-inducible phosphoprotein 1, Sti1) transfers unfolded SHRs from Hsp70 to Hsp90 and binds to one of the tetratricopeptide repeat (TPR)-acceptor sites of Hsp90 in the open state. The co-chaperone with peptidyl-prolyl *cis-trans* isomerase (PPIase) domain binds to the other Hsp90 TPR-acceptor site to form an asymmetric complex, and the binding of ATP leads to the Hsp90 intermediate state. Activator of Hsp90 ATPase homolog 1 (Aha1) promotes the formation of Hsp90 closed 1 state and accelerates ATPase activity. Co-chaperone p23 competes with Aha1 for binding to Hsp90 and stabilizes Hsp90 closed 2 state by reducing ATPase activity. After ATP hydrolysis, p23, the PPIase, and the active SHR are released. (B) Hsp90-driven activation of the protein kinases. After phosphorylation, Cdc37 captures the kinase to form a Cdc37–kinase complex that binds to Hsp90 NTD. In the presence of ATP, the Cdc37–kinase complex moves to Hsp90 MD, and Hsp90 transforms from an open state to a closed state. Then, protein phosphatase 5 (PP5) binds to Hsp90 CTD and dephosphorylates Cdc37. Finally, Cdc37, PP5, and active kinase are released, and Hsp90 returns to the open state for another cycle. Created with BioRender.com.

Table 1. Hsp90 co-chaperones, their regulation of Hsp90 function, and involvement in processes related to neurodegeneration.

Co-Chaperone	Full Name	Interacting Domain in Co-Chaperone	Binding Site in HSP90	Function	Disease	Cellular Processes
CDC37	Cell division cycle 37	MD, NTD	NTD, MD	Prevents closure of the “lid” in HSP90; Specific for maturation of kinases	AD, PD, ALS, FTL	Stabilizes tau via Hsp90 and regulates the stability of distinct tau kinases, specifically Cdk5 and Akt [31]; Preserves TDP-43 [32]; Its client kinases include DYRK1A [33] Stabilizes LRRK2 [34]; Stabilizes PINK1 and influences its subcellular distribution [35,36]
Aha1	Activator of Hsp90 ATPase homolog 1	NTD, CTD	NTD, MD	Stimulates ATPase activity of HSP90	AD	Increases tau fibril formation, Aha1 overexpression in rTg4510 mouse increases tau accumulation, leading to both neuron loss and cognitive deficits [37]. Aha1 overexpression in aged wild-type mice impairs associative learning and promotes tau phosphorylation [38].
p23 (Sba1 in yeast)	Co-chaperone p23	NTD	NTD, MD	Stabilizes the HSP90 closed 2 state; Inhibits Hsp90 ATPase activity	AD, PD	Knockdown of p23 reduces both total and phosphorylated tau levels [39]. Contributes to neurotoxicity in PD [40].
Hop (Sti1)	Hsp70-Hsp90 organizing protein (stress-inducible phosphoprotein 1)	TPR	CTD, MD	Transfers clients from Hsp70 to Hsp90; Inhibits Hsp90 ATPase activity	HD, AD, Prion diseases	Hop overexpression in yeast inhibits the toxicity of HTT103Q and reorganizes small HTT103Q foci into larger assemblies [41]. Hop downregulation enhances tau toxicity in the fly model of tauopathy [42]. Binds to PrPC and promotes calcium influx through α 7nAChRs [43]. Inhibits A β oligomers’ binding to PrPC and prevents synaptic loss, neuronal death, and depression of long-term potentiation induced by A β oligomers [44].
PP5 (Ppt1 in yeast)	Protein phosphatase 5	TPR	CTD	Dephosphorylates Hsp90; Dephosphorylates Cdc37	AD	Dephosphorylates tau and its activity decreases in AD neocortex [45]. Protects primary neuron death induced by A β [46].

Table 1. Cont.

Co-Chaperone	Full Name	Interacting Domain in Co-Chaperone	Binding Site in HSP90	Function	Disease	Cellular Processes
CHIP	C terminus of Hsp70-interacting protein	TPR	CTD	E3 ubiquitin ligase	AD, PD, HD	<p>Promotes the degradation of phosphorylated tau [39,47].</p> <p>Reduces α-synuclein oligomerization and mediates α-synuclein degradation [48,49].</p> <p>Reduces the uptake of α-synuclein fibrils by neuro-2a cells [50].</p> <p>Promotes the degradation of LRRK2 [51,52].</p> <p>CHIP overexpression promotes the degradation of polyglutamine-expanded HTT or ataxin-3 [53].</p> <p>Suppresses polyglutamine aggregation and toxicity [54].</p>
FKBP51	FK506 binding protein 51 kDa	TPR	CTD	<p>Peptidyl-prolyl isomerase activity;</p> <p>Participates in Hsp90-steroid receptor complex;</p> <p>Generally regulates Hsp90 conformational cycle</p>	AD, PD, HD	<p>Enhances the production of tau oligomers and prevents tau degradation [55].</p> <p>Increases with age in the mouse brain, and its expression is higher in AD patients [55,56].</p> <p>Involved in Pink1's regulation of AKT on neuronal survival [57].</p> <p>FKBP51 downregulation reduces mutant HTT levels in HD models both in vitro and in vivo [58].</p>
FKBP52	FK506 binding protein 52 kDa	TPR	CTD	<p>Peptidyl-prolyl isomerase activity;</p> <p>Participates in Hsp90-steroid receptor complex;</p> <p>Generally regulates Hsp90 conformational cycle</p>	AD, PD	<p>Induces aggregation of multiple tau species in vitro [59–61].</p> <p>FKBP52 overexpression in the hippocampus leads to cognitive impairments and neurotoxicity in aged wild-type mice and rTg4510 transgenic mice [38,62].</p> <p>FKBP52 levels are abnormally low in the frontal cortex of AD brains [63].</p> <p>Suppresses Aβ toxicity and increases the lifespan of <i>Drosophila</i>, which expresses Aβ peptides [64].</p> <p>Accelerates α-synuclein aggregation and neuronal cell death [65].</p> <p>Generates immune responses to α-synuclein-based immunizations in mice [66].</p>

3.2. Hsp90 and Its Co-Chaperones in Neurodegenerative Diseases and Their PPI Inhibitors

Extensive research supports the important roles of Hsp90 co-chaperones in various neurodegenerative diseases, which are summarized in Table 1 and described in the following subsections. Unlike directly inhibiting Hsp90, specifically targeting these Hsp90/co-chaperones PPIs can modulate Hsp90 function by “fine-tuning” and provide an alternative therapeutic strategy for neurodegenerative diseases. Until now, many small molecules have been reported to inhibit the interactions between Hsp90 and Cdc37, Aha1, p23, Hop, or other TPR co-chaperones. The following subsections highlight PPIs of interest and the developed PPI inhibitors, as well as their binding sites, drug development, and chemical structures.

3.2.1. Hsp90–Cdc37 Interaction

Cdc37 specifically recognizes and delivers an extensive range of protein kinases to Hsp90, which forms an Hsp90–Cdc37–kinase complex that mediates kinase maturation [7]. Several lines of evidence prove the roles of the Hsp90–Cdc37 complex in neurodegeneration. Cdc37 stabilizes tau via Hsp90 and regulates the stability of distinct tau kinases, specifically cyclin-dependent kinase 5 (Cdk5) and Akt [31]. The Hsp90–Cdc37 complex also preserves TDP-43, which mislocalizes and accumulates in the cytoplasm in the ALS, frontal temporal dementia (FTLD), and some cases of AD [32]. Moreover, the client kinases of the Hsp90–Cdc37 chaperone system also include dual-specificity tyrosine-phosphorylation-regulated kinase 1A (DYRK1A), leucine-rich repeat kinase 2 (LRRK2), and PTEN-induced kinase 1 (PINK1), which are involved in AD and PD [33–36]. Unlike directly inhibiting Hsp90 ATPase, blocking the Hsp90–Cdc37–kinase chaperone cycle could selectively modulate Hsp90 kinase clients, rather than inhibiting all the Hsp90 client proteins. Therefore, disrupting Hsp90–Cdc37–kinase interactions has emerged as a potential alternative therapeutic strategy for neurodegenerative diseases.

Two different states of Hsp90–Cdc37 complex structures (PDB: 1US7 and PDB: 5FWL) have been identified by Roe et al., in 2004 and Verba et al., in 2016, respectively [67,68]. These structures provide essential clues to understand the binding between Hsp90 and Cdc37. Before ATP binding, Hsp90 exhibits a clamp-like open state. After phosphorylation by CK2, Cdc37 can capture the client kinase to form a Cdc37–kinase complex that binds to the NTD of Hsp90 via Cdc37 MD. When the NTD of Hsp90 binds ATP, Cdc37–kinase complex moves to the MD of Hsp90, which interacts with the NTD of Cdc37 and the N-lobe of the kinase. Then, Hsp90 transforms from an open state to a closed state. After that, PP5 binds to the CTD of Hsp90 and dephosphorylates Cdc37. Then, Cdc37, PP5, and active kinase are released and Hsp90 returns to the open state for another chaperone cycle [7] (Figure 3B). This Hsp90–Cdc37 binding process makes it possible to discover compounds disrupting Hsp90–Cdc37–kinase interactions.

The developed inhibitors targeting Hsp90–Cdc37 interactions include naturally derived products (Figure 4), Cdc37 peptides, and small molecular inhibitors (Figure 5).

Natural Products

Some natural products have been found to inhibit the Hsp90–Cdc37 interaction, such as celastrol, withaferin A, sulforaphane, FW-04-806, kongensin A, platycodin D, apigenin, and 18 β -glycyrrhetic acid derivatives. Although these natural products are nonspecific and have poor druggability, their drug-like derivatives could be developed as a therapy for neurodegenerative diseases.

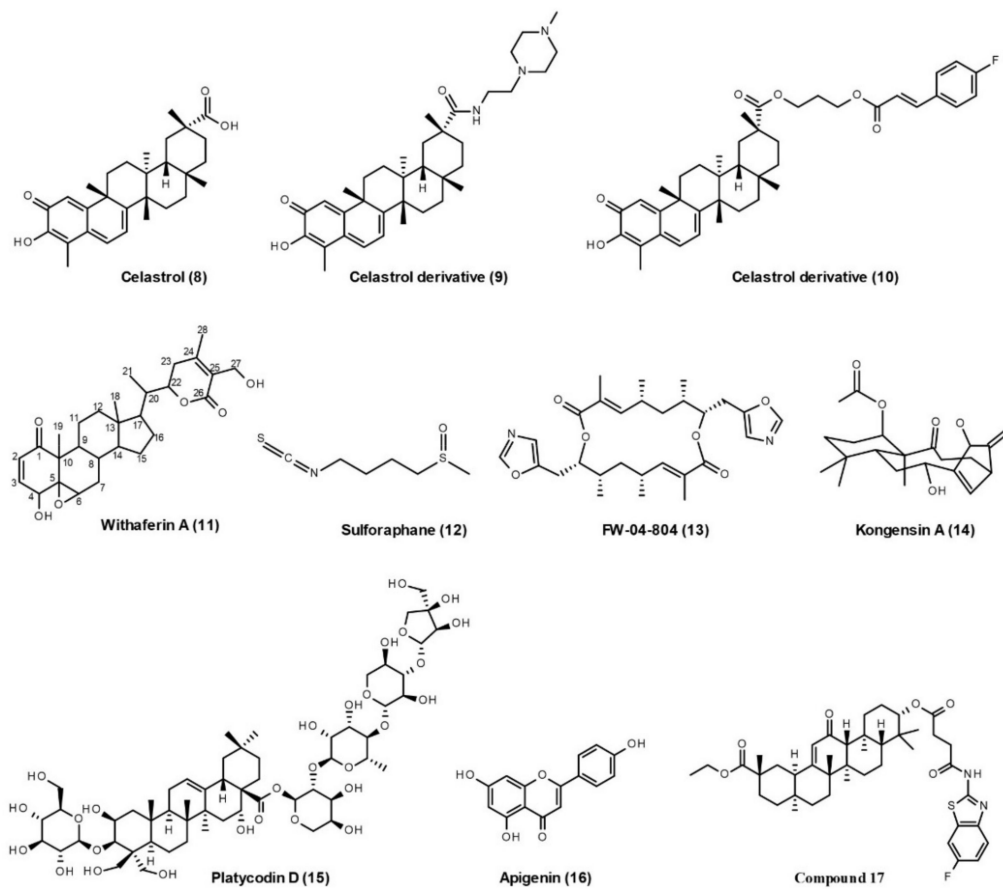


Figure 4. Natural products and their derivatives with the ability to disrupt Hsp90–Cdc37 interactions.

Celastrol (8, Figure 4) is a quinone methide triterpene isolated from the *Tripterygium wilfordii* Hook F. In pancreatic cancer cells, celastrol disrupts the Hsp90–Cdc37 interaction, but does not disrupt Hsp90–Hop and Hsp90–p23 interactions [69]. Several studies have investigated the binding sites of celastrol on the Hsp90–Cdc37 complex, although with conflicting results. In 2009, proteolytic fingerprinting indicated that celastrol binds to the CTD of Hsp90 to protect it from trypsin digestion [70], while HSQC NMR studies show that the quinone methide of celastrol reacts with the thiol group of Cdc37 in the NTD through a Michael addition, thereby disrupting the Hsp90–Cdc37 complex [71]. In 2014, multiple techniques, such as size-exclusion chromatography coupled to multi-angle laser light scattering, native-PAGE, dynamic light scattering, differential scanning calorimetry, differential scanning fluorescence, and chaperone and PPI assays, have been applied to characterize the interaction, and it was found that celastrol affects the oligomeric state of Hsp90 by binding to its CTD [72]. Thus, the action mode of celastrol is not only via its “Michael acceptor” functionality, but also involves other mechanisms that need to be clarified in further studies. To improve the drug-like properties of celastrol, many celastrol derivatives have recently been produced. Two of them (9 and 10) have been selected for improved Hsp90–Cdc37 disruption activity and antiproliferative activity [73,74]. Although celastrol has non-negligible anti-tumor efficacy [75], it possesses an extensive medical value in the treatment of neurodegenerative diseases such as AD, PD, ALS, cerebral ischemia, multiple sclerosis, and spinal cord injury, which has been well-summarized in a recently published review [76]. The celastrol derivatives targeting neurodegenerative diseases need to be developed in the future.

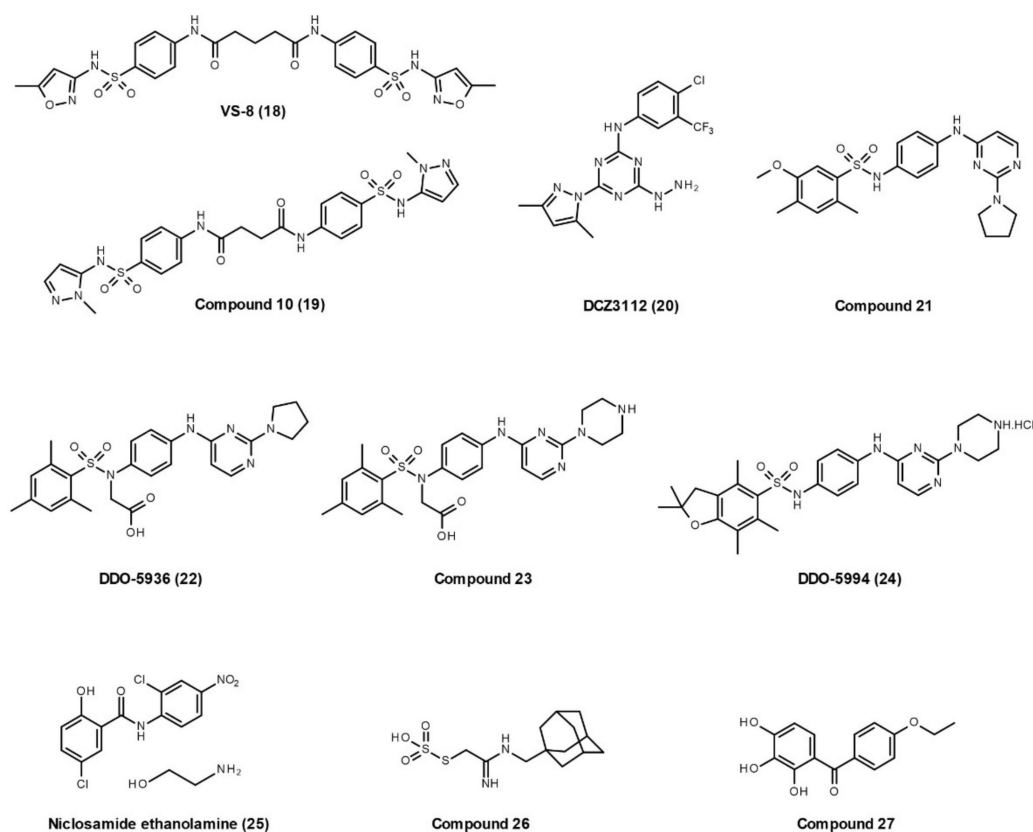


Figure 5. Developed small molecules with the ability to disrupt Hsp90–Cdc37 interactions.

Withaferin A (11, Figure 4) is steroidal lactone isolated from the *Withania somnifera*. In pancreatic cancer cells, withaferin A disrupts the Hsp90–Cdc37 interaction, while it neither blocks ATP binding to Hsp90 nor disrupts the Hsp90–p23 interaction [77]. A withaferin A-biotin pull-down assay shows that Withaferin A binds to the CTD of Hsp90 [77], suggesting that targeting the Hsp90 CTD may disrupt the Hsp90–Cdc37 interaction by allosteric regulation. Later, the computational docking results indicate that withaferin A has the potential to inhibit the Hsp90–Cdc37 interaction by disrupting the stability of the Hsp90–Cdc37 complex [78]. Structure–activity relationship studies show that the C-5(6) epoxy functional group of withaferin A is required for binding with Hsp90, the substitution of C-2,3 position may hinder its inhibition on Hsp90 activity, while the C-4 hydroxyl group in its A ring may enhance the inhibition on Hsp90 and disruption of the Hsp90–Cdc37 complex [79]. Withaferin A is not only a promising anticancer compound, but also has many other therapeutic benefits, including neuroprotective (AD, PD, and ALS), cardioprotective, anti-viral (COVID-19 and Hepatitis), and osteoporotic effects [80].

Sulforaphane (12, Figure 4) is an isothiocyanate found in cruciferous vegetables, such as cabbage, broccoli, cauliflower, and kale. In pancreatic cancer cells, sulforaphane induces the Hsp90 degradation and blocks the Hsp90–Cdc37 interaction without affecting the ATP binding pocket of Hsp90 [81]. NMR and liquid chromatography coupled to mass spectrometry studies reveal that sulforaphane binds to the NTD of Hsp90 [81]. Sulforaphane has shown efficacy in a wide range of human-related neurological pathologies, such as AD, PD, HD, ALS, multiple sclerosis, autism spectrum disorder, and schizophrenia [82,83]. There is now a clinical study recruiting participants to investigate the effects of sulforaphane in patients with prodromal to mild AD (NCT04213391).

FW-04-806 (13, Figure 4), also known as conglobatin A, is extracted from the Chinese native *Streptomyces* FIM-04-806. Immunoprecipitation confirms that FW-04-806 disrupts the Hsp90–Cdc37 interaction, leading to enhanced tumor-arresting activity and the degradation of Hsp90 clients in breast cancer cells [84]. Chemoproteomics and computational

approaches together reveal that FW-04-806 binds to the NTD of Hsp90 without affecting ATP binding of Hsp90 [84]. Recent computational docking results indicate that FW-04-806 sterically blocks and disturbs critical interactions, notably between Glu47 of Hsp90 NTD and Arg167 of Cdc37 at the interface of the complex (PDB: 1US7) [85].

Kongensin A (14, Figure 4) is isolated from *Croton kongensis* and has the potential to inhibit necroptosis and induce apoptosis. The bio-orthogonal ligation method reveals that Hsp90 is the direct cellular target of kongensin A, and further studies demonstrate that kongensin A binds covalently to Cys420 on the MD of Hsp90, dissociates Hsp90 from Cdc37, and subsequently inhibits necroptosis [86,87].

Platycodin D (15, Figure 4) is an effective triterpenesaponin isolated from the roots of *Platycodon grandiflorus*. It is shown that platycodin D disrupts the Hsp90–Cdc37 interaction and, subsequently, leads to the degradation of multiple Hsp90 client proteins without affecting Hsp90 ATPase activity [88]. However, its binding sites on the Hsp90–Cdc37 complex are unclear and need further studies.

Apigenin (16, Figure 4) is a natural product belonging to the flavone class and is abundant in common fruits and vegetables. Apigenin not only possesses anti-carcinogenic effects [89], but also plays important roles in pathogenesis of neurodegenerative diseases, such as AD, PD, multiple sclerosis, epilepsy, and stroke [90]. In multiple myeloma cells, apigenin disrupts the Hsp90–Cdc37–client complex, induces the degradation of multiple kinase clients, and decreases phosphorylation of Cdc37 [91]. Its binding sites on the Hsp90–Cdc37 complex also require investigation in further studies.

18 β -Glycyrrhetic acid is a pentacyclic triterpenoid found in *Glycyrrhiza glabra* L. (licorice) roots. In 2018, a series of aminobenzothiazole derivatives of 18 β -Glycyrrhetic acid were designed and synthesized as Hsp90–Cdc37 disruptors with the ability to inhibit cell migration and drug-resistance [92]. Among them, **compound 17** (Figure 4) exhibits the most potent activity to disrupt the Hsp90–Cdc37 interaction (IC₅₀, 0.14 μ M). Its docking study suggested that compound 17 tightly binds to the active site of the Hsp90–Cdc37 complex and the small bulky and strongly electrophilic group of aminobenzothiazole side chains at its C-30 position is crucial for improving its activity.

Cdc37 Peptides

Another way to develop PPI inhibitors is to locate endogenous peptides at the binding epitopes of participating proteins. In 2015, the first small peptide, Pep-1 (Ac-KHFGMLRRWDD-NH₂), was derived with high potency to disrupt the Hsp90–Cdc37 interaction [93]. The hot-spot Arg167 has been revealed as one of the most important binding determinants. Pep-1 not only occupies the Cdc37 binding site, but also interferes with Hsp90 ATPase activity. Later, more peptides were designed for verification. In 2017, a shorter peptide, Pep-5 (Ac-HFGMLRR-NH₂), was found, which exhibited more stable binding and better ligand properties than Pep-1 [94]. In a recent study, five new peptides (NYSVWDHIEVSDDLKDGFSKSMVN, NYSVWDHIEVDDDLKDGFSKSMVN, NYSVWDHIEVEDDLKDGFSKSMVN, LSKDGFSKSMVN, and PSKDIFLKSMIN) were designed and synthesized with the ability to disrupt the Hsp90–Cdc37–Cdk4 complex in co-immunoprecipitation experiments [95]. The discovery of these peptides improved our understanding of the Hsp90–Cdc37 binding interface. However, they still need further optimization to improve their membrane permeability, proteolytic stability, and drug-like potential.

Small-Molecule Inhibitors

Several critical residues at the Hsp90–Cdc37 binding interface have been identified by Jiang et al. in 2010 [96]. The Split Renilla luciferase protein fragment-assisted complementation bioluminescence shows that mutations in Hsp90 (Q133A, F134A, and A121N) and mutations in Cdc37 (M164A, R167A, L205A, and Q208A) reduce the Hsp90/Cdc37 interaction by 70–95%, and mutations in Hsp90 (E47A and S113A) and a mutation in Cdc37 (A204E) decrease the Hsp90/Cdc37 interaction by 50%. This study provides the “hot-spots”

at the Hsp90–Cdc37 binding interface to develop small-molecule inhibitors. Several small-molecule inhibitors targeting the Hsp90–Cdc37 interaction have been developed recently. However, their effects on neurodegeneration have not yet been investigated.

In 2017, Wang et al., identified small molecules inhibiting the Hsp90–Cdc37 interaction by utilizing a structure-based virtual screening workflow, derivatives synthesis, and their biological evaluation [97]. The chemical databases (Specs and NCI database) with 500,000 molecules were screened by pharmacophore and cross-docking filtrations. After that, 31 compounds were obtained and purchased for further *in vitro* identification, including homogeneous time-resolved fluorescence assay, ATPase inhibition assay, fluorescence polarization assay, and direct binding assay (biolayer interferometry). Finally, **VS-8 (18, Figure 5)** reveals moderate binding ability to Hsp90 (K_d , 80 μ M) and disrupts the Hsp90–Cdc37 interaction (IC_{50} , 77 μ M) without effects on Hsp90 ATPase activity. To enhance the potency, 16 derivatives were designed and synthesized. Among them, **compound 19**, with N-methylpyrazole substitution and the central linker of two alkyl carbons, exhibits better binding capacity (K_d , 40 μ M), a more promising inhibitory effect (IC_{50} , 27 μ M), and preferable antiproliferative activity against multiple cancer cell lines.

In 2018, Chen et al. found that **DCZ3112 (20, Figure 5)**, a novel derivative of triazine, directly binds to the NTD of Hsp90, inhibits the Hsp90–Cdc37 interaction without inhibiting ATPase activity, and leads to the degradation of Hsp90 client proteins [98]. DCZ3112 predominantly acts in HER2-positive breast cancer, exerts synergistic effects when applied in combination with anti-HER2 antibodies, and overcomes trastuzumab resistance [98].

In 2019, Wang et al. identified polar interactions between Arg167 on Cdc37 and Glu47 and Gln133 on Hsp90 as the most important binding determinants in recognition during the dynamic cycle of the Hsp90–Cdc37 interaction [99]. Then, they designed a screening workflow that identified **compound 21 (Figure 5)**, which has a moderate binding affinity to Hsp90. Based on the hit compound 21, **DDO-5936 (22)** was identified as an active inhibitor disrupting the Hsp90–Cdc37 interaction, with no effects on Hsp90 ATPase activity. Although they failed to obtain the co-crystal structure of DDO-5936–Hsp90, the NMR detection and mutagenesis validation demonstrated that DDO-5936 specifically binds to the Glu47 on Hsp90 NTD to block the critical interaction between Hsp90 and Cdc37. In addition, inhibition of the Hsp90–Cdc37 complex by DDO-5936 results in downregulation of cyclin-dependent kinase 4 and consequent inhibition of cell proliferation through Cdc37-dependent cell cycle arrest, as well as *in vivo* antitumor potency in a xenograft model [99]. Later, the same group developed **compound 23** with improved binding affinity and antiproliferative activity, preferable stability in plasma and microsomes, and oral efficacy *in vivo*, compared with DDO-5936 [100]. Based on the binding mode of compound 21, they recently discovered a hydrophobic pocket centered on Phe213 of Hsp90 that contributes to the binding affinity of Hsp90–Cdc37 interaction inhibitors [101]. An optimum compound **DDO-5994 (24)** was identified with an ideal binding on a Phe213 hydrophobic core. DDO-5994 has improved the binding affinity, antiproliferative activity, and antitumor potency in mice bearing HCT116 xenograft tumors.

Niclosamide ethanolamine (NEN, 25, Figure 5) is an anthelmintic drug approved by the United States Food and Drug Administration for the treatment of parasitic infections. Several studies have shown that NEN can be used for the treatment of hepatocellular carcinoma [102], lipotoxicity [103], systemic lupus erythematosus [104], and artery constriction [105]. Chen et al. found NEN inhibits multiple kinases that are regulated by the Hsp90–Cdc37 complex, such as AKT, EGFR, STAT3, LRP6, and Raf family [102]. Using purified recombinant Hsp90 and Cdc37 in ELISA and co-immunoprecipitation assays, they found that NEN disrupts the Hsp90–Cdc37 interaction. Their results also showed that NEN binds to Cdc37 in pull-down assay and thermal shift assay, but its specific binding site on Cdc37 is unknown.

To identify novel Hsp90–Cdc37 interaction inhibitors, Siddiqui et al. recently established a mammalian cell lysate-based, medium-throughput amenable split *Renilla* luciferase assay, which employs N-terminal and C-terminal fragments of *Renilla* luciferase fused to

full-length human Hsp90 and Cdc37, respectively [106]. Later, the same group screened more than 120,000 compounds via an FW-04-806 (conglobatin A)-based pharmacophore model and molecular docking, and confirmed the hits' *in vitro* effects using their formerly established mammalian cell lysate-based split *Renilla* luciferase assay [85]. The active **compounds 26** and **27** were identified with K-Ras selectivity. Both compounds potently decrease the Hsp90 client protein levels without affecting Hsp90 ATPase activity, and inhibit the cancer cell proliferation and microtumor formation.

3.2.2. Hsp90–Aha1 Interaction

Aha1 accelerates Hsp90 ATPase activity and has two domains, an NTD and a CTD, connected by a flexible linker [107,108]. In 2004, Meyer et al. reported a fragment-based crystal structure of Aha1 NTD in complex with Hsp90 MD that provides a model for Aha1 recruitment to Hsp90 [109]. In 2010, Koulov et al. have demonstrated that Aha1 NTD and CTD cooperatively bind across the dimer interface of Hsp90 to modulate the ATP hydrolysis cycle and Aha1 CTD binds to Hsp90 NTD, promoting ATP hydrolysis by stabilization of the N-terminal dimer interface [110]. At the same time, Retzlaff et al. also proved that acceleration of the Hsp90 ATPase cycle requires the interaction of both Aha1 NTD and CTD in a cooperative manner with both Hsp90 NTD and MD in an asymmetric activation mechanism [111]. In 2020, Liu et al. revealed that the Hsp90–Aha1 complex has six different states by cryo-electron microscopy [112]. Combining with previous data, they proposed a multistep activation model: Aha1 is firstly recruited to Hsp90 through interactions between Aha1 NTD and Hsp90 MD; then Aha1 CTD binds to Hsp90, which leads to a structural transition of Hsp90 from its open state to a semi-closed state and undocks Hsp90 NTD from Hsp90 MD; in the presence of ATP, Aha1 CTD rearranges its binding interface with Hsp90 MD, stabilizing a fully closed state followed by Hsp90 NTD dimerization; finally, Aha1 NTD tilts up to interact with dimerized Hsp90 NTD and facilitates ATP hydrolysis [112,113].

Aha1 has an important role in AD tauopathy. Aha1 increased tau fibril formation in the presence of Hsp90 *in vitro*. Overexpression of Aha1 in the rTg4510 tau transgenic mouse model increased neurotoxic oligomeric and insoluble tau accumulation, leading to both neuron loss and cognitive deficits [37]. Moreover, overexpression of Aha1 in aged wild-type mice impaired associative learning and promoted tau phosphorylation [38]. Therefore, small molecules inhibiting Hsp90–Aha1 PPI can be developed for the treatment of AD.

In 2017, Ihrig et al. first screened a collection of 14,400 drug-like compounds for Hsp90–Aha1 PPI inhibitors by AlphaScreen assay, and identified two drug-like inhibitors, **compounds 28** and **29** (Figure 6), that show positive effects on cystic fibrosis [114]. In the same year, Stiegler et al. screened about 15,000 chemical compounds by resonance energy transfer assay and selected six inhibitors with promising effects on the Hsp90–Aha1 interaction [115]. The most effective inhibitor, **HAM-1 (30, Figure 6)**, specifically prevents the interaction between Hsp90 NTD and Aha1 CTD, without dissociating the binding of Hsp90 MD and Aha1 NTD. Thus, HAM-1 does not change Aha1 affinity, but can abrogate the Aha1-induced ATPase stimulation of Hsp90 [115]. In 2020, Singh et al. built a modified quinaldine red-based high-throughput assay to screen compounds that inhibit Aha1-stimulated Hsp90 ATPase activity [116]. They identified a novel inhibitor of the Aha1-stimulated Hsp90 ATPase activity, **SEW84 (31, Figure 6)**, and revealed that it binds to the CTD of Aha1 to weaken its asymmetric binding to Hsp90. However, SEW84 does not affect the basal ATPase activity of Hsp90, avoiding the toxic effects of common Hsp90 inhibitors. Importantly, SEW84 reduces the tau phosphorylation in HEK293 cells expressing RFP-tagged 0N4R-tau, primary rat cortical neurons expressing endogenous WT-tau, and cultured brain slices from rTg4510 transgenic mouse [116]. SEW84 needs further *in vivo* experiments to verify its effect on tauopathies.

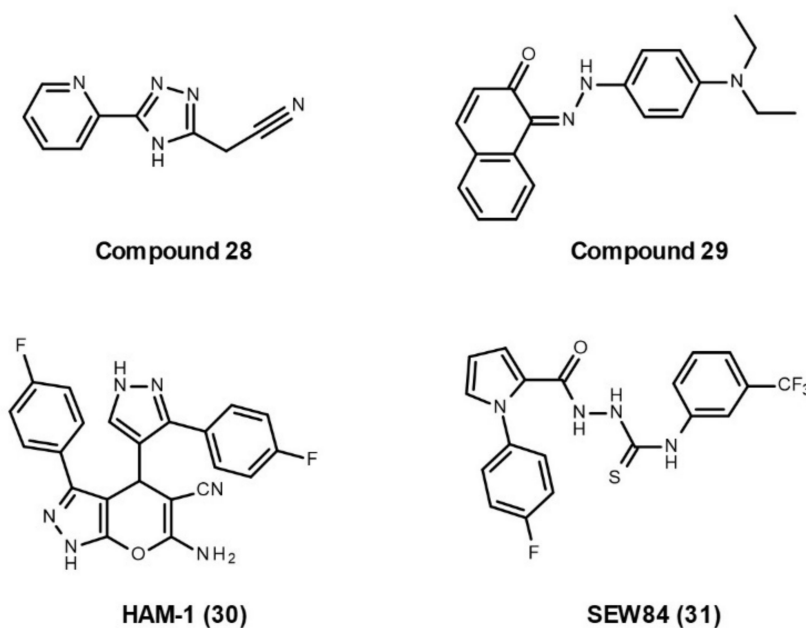


Figure 6. Developed small molecules with the ability to disrupt Hsp90–Aha1 interactions.

3.2.3. Hsp90–p23 Interaction

Co-chaperone p23 preferentially interacts with Hsp90 and stabilizes the closed 2 state of Hsp90 [117]. The crystal structure of p23 and Hsp90 shows that the folded NTD of p23 binds to the junction of two NTDs of Hsp90. Another identified interaction site is located in the MD of Hsp90, which stabilizes the interaction with p23 [118,119]. The interaction between Hsp90 and p23 leads to the inhibition of Hsp90 ATPase activity [107,120,121]; that is, p23 either inhibits the hydrolysis process or impedes the subsequent release of ADP and Pi, and hence the re-opening of Hsp90 [122]. There is some evidence showing that p23 has an important role in AD and PD. Knockdown of p23 via siRNA transfection reduces both total and phosphorylated tau levels [39]. It is also proposed that the Hsp90–p23 complex contributes to neurotoxicity in PD. Mitochondrial stress increases the association of the Hsp90–p23 complex and the enzyme prolyl hydroxylase domain 2 in cultured dopaminergic neurons, while p23 inhibition prevents mitochondrial stress-induced neurotoxicity [40]. Therefore, inhibition of the Hsp90–p23 complex can provide a new strategy for AD or PD treatment.

Celastrol (8), gedunin (32), cucurbitacin D (33), and ailanthon (34) are the four natural products that have been proved to inhibit Hsp90–p23 PPIs (Figure 7). Contrary to the former results [69], Chadli et al. reported that **celastrol** (8) can disrupt the Hsp90–p23 complex by altering the structure of p23, causing it to polymerize into amyloid-like fibrils [123]. **Gedunin** (32) is a tetranortriterpenoid natural product isolated from the Indian neem tree. Patwardhan et al. revealed that gedunin inhibits the p23 chaperoning activity, blocks its cellular interaction with Hsp90, prevents glucocorticoid receptor nuclear localization, and interferes with p23-mediated gene regulation [124]. Using molecular docking and mutational analysis, they identified three amino acids (Thr90, Ala94, and Lys95) that mediate noncovalent interactions of p23 with gedunin. In human breast and cervical cancer cell lines, gedunin induces apoptotic cell death and caspase-7-dependent cleavage of p23. Gedunin has been shown to have a potential therapeutic role in neurodegenerative diseases. It enhances tau degradation [125], inhibits oligomeric A β -induced microglia activation [126], and degrades the abnormal mutant huntingtin (HTT) aggregates and intranuclear inclusions in cells from HD patients [127]. Deoxygedunin also protects nigrostriatal dopaminergic neurons and improves the behavioral performance in PD animal models [128]. However, these studies do not evaluate the effects of gedunin on Hsp90–p23 PPIs in these neurodegenerative conditions. The underlying link between gedunin’s neuro-

protective effects and its inhibition of Hsp90–p23 PPIs need to be investigated in the future. **Cucurbitacin D (33)** is an active component in *Cucurbita texana*. Co-immunoprecipitation showed that cucurbitacin D disrupts interactions between Hsp90 and two co-chaperones, p23 and Cdc37, in MCF7 cell lysates [129]. In 2016, He et al. identified a natural compound, **ailanthone (34)**, that prevents Hsp90–p23 PPIs and decreases the interaction between chaperones and the androgen receptor (AR) followed by ubiquitin/proteasome-mediated degradation of AR as well as other p23 clients [130]. Ailanthone does not interact with Hsp90, but directly binds to p23 on the surface formed by Ser100, Val101, Lys95, Arg93, Pro87, and Trp8. Ailanthone not only blocks tumor growth and metastasis of castration-resistant prostate cancer, but also possesses favourable drug-like properties such as good bioavailability, high solubility, lack of CYP inhibition, and low hepatotoxicity [130]. Its effects on neurodegeneration need to be studied later.

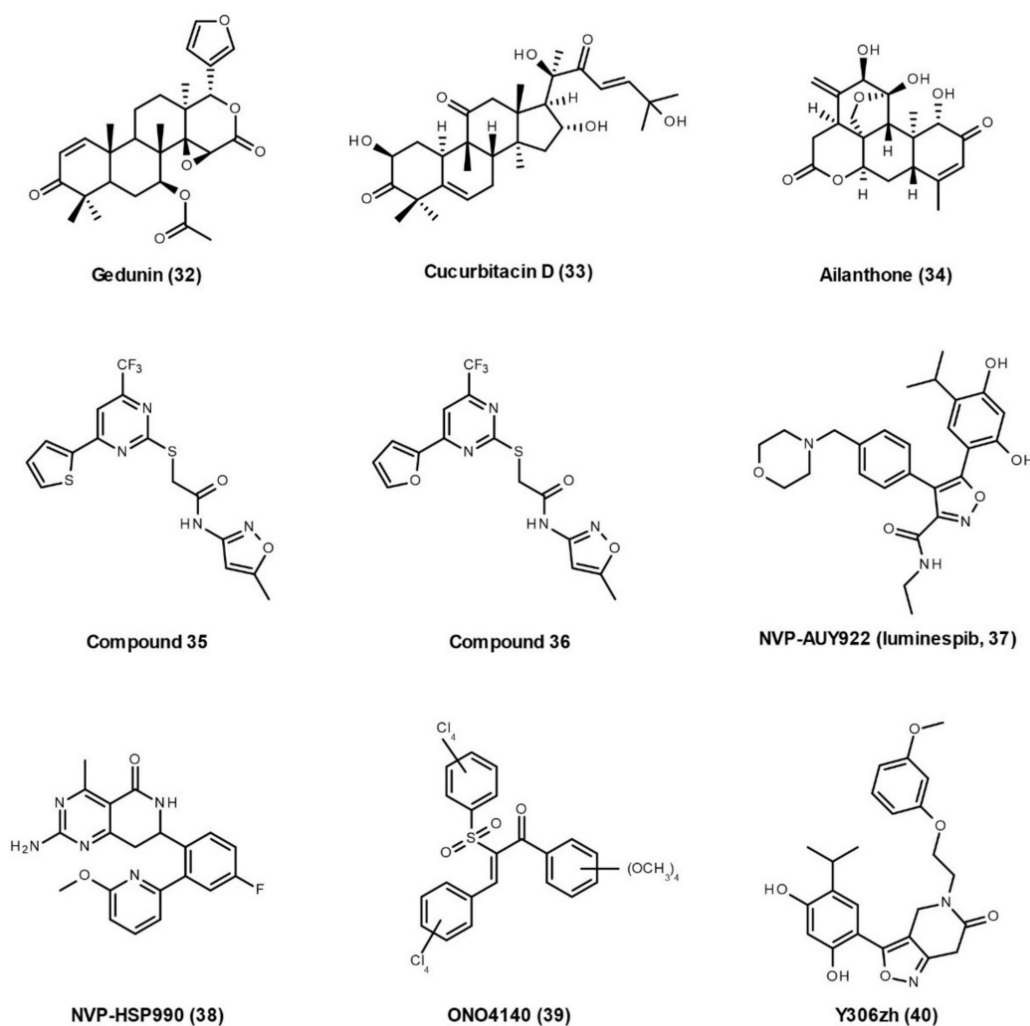


Figure 7. Developed small molecules with the ability to disrupt Hsp90–p23 interactions.

In addition, Chan et al. developed a dual luciferase (Renilla and Firefly) reporter system for high-throughput screening and identifying inhibitors of Hsp90–p23 interactions [131]. Using this method, they identified a potent compound, *N*-(5-methylisoxazol-3-yl)-2-[4-(thiophen-2-yl)-6-(trifluoromethyl)pyrimidin-2-ylthio]acetamide (35, Figure 7), that inhibits Hsp90–p23 bioluminescence imaging signals and leads to the degradation of Hsp90 clients. They also performed a structural activity relationship study with 62 analogs of compound 35, and identified **compound 36** as the lead compound that outperformed compound 35 in inhibiting Hsp90–p23 interactions. Some Hsp90 inhibitors with anti-

cancer activities also induce Hsp90–p23 dissociation, such as **NVP-AUY922 (luminespib, 37)**, **NVP-HSP990 (38)**, **ONO4140 (39)**, and **Y306zh (40)** [132–135]. Y306zh binds to the NTD of Hsp90, causes ATP to be incapable of attaching to Hsp90 NTD, and thus disrupts Hsp90–p23 PPIs [135].

3.2.4. Hsp90–Hop Interaction

Several Hsp90 co-chaperones contain the TPR domain that mediates the binding to the conserved MEEVD pentapeptide at the CTD of Hsp90 [136]. One of the best characterized TPR-containing co-chaperones is Hop, which facilitates unfolded client proteins transferred from Hsp70 to Hsp90 for maturation. Hop contains three TPR domains, TPR1, TPR2A, and TPR2B. Its TPR1 and TPR2B domains are responsible for Hsp70 binding, while the TPR2A domain is for Hsp90 binding [137,138]. The TPR2A and TPR2B domains of Hop also interact with the MD of Hsp90 [138].

Hop is important in HD, AD, and prion diseases [139]. Hop overexpression in yeast inhibits the toxicity of HTT with 103Q glutamine stretch (HTT103Q) and reorganizes small HTT103Q foci into larger assemblies [41]. Downregulation of Hop enhances tau toxicity in a fly model of tauopathy [42]. Secreted Hop binds to the cellular prion protein (PrP^C) and promotes calcium influx through $\alpha 7$ nicotinic acetylcholine receptors ($\alpha 7$ nAChRs), which modulates neuronal differentiation and survival [43]. As the PrP^C ligand, Hop also inhibits A β oligomers' binding to PrP^C and prevents synaptic loss, neuronal death, and depression of long-term potentiation induced by A β oligomers [44]. The TPR1 and TPR2A of Hop contribute to the binding of C-terminal of PrP^C and can directly inhibit both A β oligomers' binding to PrP^C and neuronal toxicity [140]. Excess Hsp90 can disrupt the Hop–PrP^C interaction by interfering with the TPR2A domain of Hop and blocks Hop neuroprotective functions [140]. Therefore, disrupting the Hsp90–Hop interaction can promote the degradation of Hsp90 clients and provide more Hop to bind with PrP^C, leading to a new therapeutic strategy for neurodegenerative disease.

In 2008, Yi and Regan first screened more than 97,000 compounds by AlphaScreen assay to identify a new class of small molecules that inhibit the interaction of the C-terminal peptide of Hsp90 and the TPR2A domain of Hop [141]. Following structure clustering analysis, competition confirmation, counter screen for false positives, fluorescence polarization assay, and isothermal titration calorimetry, they have identified six active compounds, **41–46** (Figure 8), with a 7-azapteridine ring system that binds at a key position on the TPR2A interaction interface. These compounds can reduce the levels of the Hsp90-dependent client protein HER2 in human breast cancer cell lines without inducing Hsp70 overexpression. Later, the same group characterized the anticancer activity of **compound 42** (Figure 8) and showed that this compound is effective in killing different breast cancer cell lines including the highly metastatic MDA-MB-231 [142].

In 2011, Horibe et al. designed the hybrid Antp-TPR peptide (KAYARIGNSYFK) based on the structure of the Hsp90–TPR2A complex [143]. This peptide includes the conserved amino acids Lys301 and Arg305 that donate hydrogen bonds to the Hsp90 C-terminal region. The surface plasmon resonance experiments showed that the hybrid Antp-TPR peptide specifically inhibits the interaction of Hsp90 with Hop, rather than the interaction of Hsp70 with Hop and the interaction of Hsp90 with FKBP5 or PP5. They also proved the antitumor activity of this peptide both in various cancer cell lines and in a xenograft model of human pancreatic cancer in mice. Later, another peptide PEP73 (INSAYKLYARG) was designed based on the *in silico* docking of Hsp90 α and Hop, but its inhibition effects need to be further verified [144].

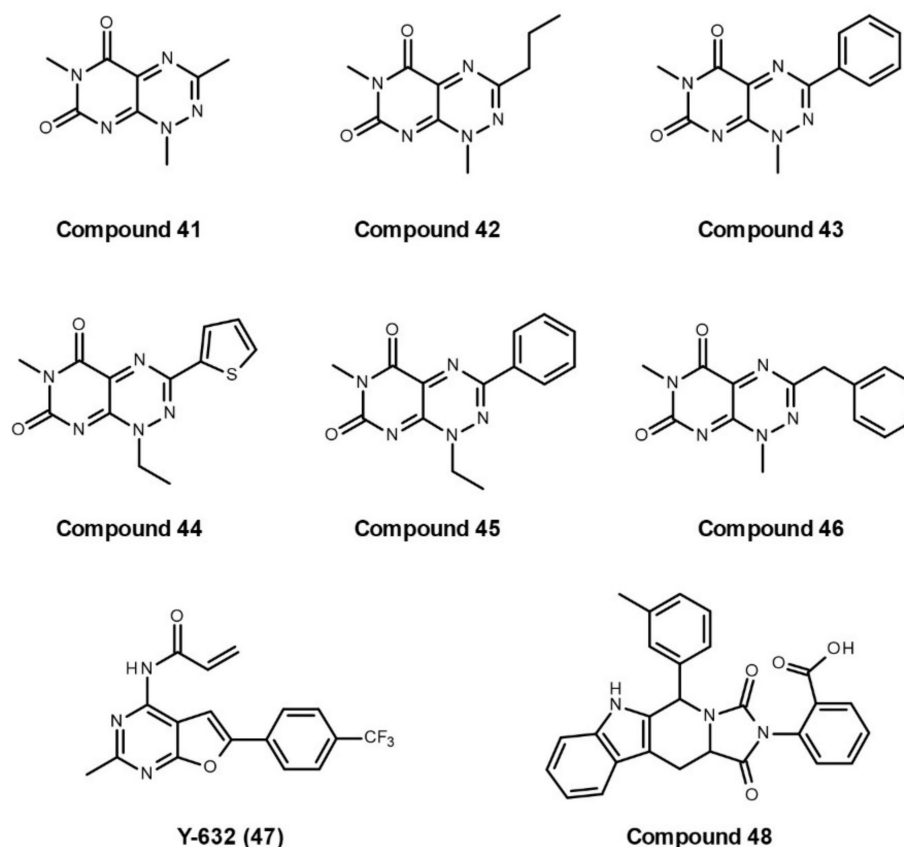


Figure 8. Developed small molecules with the ability to disrupt Hsp90–Hop interactions.

In 2016, Wang et al. found a pyrimidine derivative, **Y-632 (47)** (Figure 8), as a novel inhibitor disrupting the interaction between Hsp90 and Hop [145]. Y-632 neither binds to Hsp90 nor inhibits Hsp90 ATPase activity. It inhibits Hsp90 function by intracellular thiol oxidation, thereby disturbing the Hsp90–Hop interaction. Y-632 induces the degradation of diverse Hsp90 client proteins through the ubiquitin-proteasome pathway and can efficiently overcome imatinib resistance mediated by Bcr-Abl point mutations.

Recently, Darby et al. mimicked key native “carboxylate clamp” interactions between Hsp90 and its TPR co-chaperones and designed several compounds that block the interaction between Hop TPR2A and Hsp90 C-terminal MEEVD peptide after the failure of AlphaScreen high-throughput screening [146]. The binding of these compounds to the Hop TPR2A domain was confirmed by mapping ^1H - ^{15}N HSQC chemical shift perturbations to their new reported NMR solution-state structure of Hop TPR2A. Co-immunoprecipitation was also used to prove the Hsp90–Hop PPI inhibition effects of a selected compound, **48** (Figure 8), in human cancer cells. However, these compounds have relatively low potency to disrupt Hsp90–Hop PPI. Much effort is still required to overcome the challenge for finding efficient small-molecule ligands disrupting the Hop TPR2A–Hsp90 MEEVD interface.

3.2.5. Hsp90 and Other TPR Co-Chaperones Interactions

In addition to the aforementioned Hop, there are several other important TPR containing co-chaperones of Hsp90, such as PP5, C terminus of Hsp70-interacting protein (CHIP), FKBP 51, and FKBP52. PP5 contains the phosphatase domain in the C-terminal region and three consecutive TPR domains in the N-terminal region for PPIs. Its TPR domains interact with its extreme C-terminal alpha J helix (αJ) in the auto-inhibited state, which blocks substrate access to the catalytic groove [147]. This auto-inhibition can be broken by interacting the PP5 TPR domains with PP5 activators such as chaperones Hsp70 and Hsp90. PP5 can directly dephosphorylate Hsp90, which modulates Hsp90 conformational cycle

and client maturation [148,149]. Additionally, binding of PP5 to the Hsp90–Cdc37–kinase heterocomplex leads to Cdc37 dephosphorylation and subsequent release of Cdc37 and mature kinase. CHIP contains an N terminal TPR domain together with a U-box domain linked via a long helical region [150]. CHIP binds to both Hsp70 and Hsp90 via its TPR domain and interacts with the proteasome by acting as an E3 ligase using its U-box domain, which effectively crosslinks the chaperones to the ubiquitin–proteasome system for substrate degradation [151]. FKBP51 and FKBP52 share high homology and the same domain structure. They contain the PPIase domain (FK1), as well as FK2 and TPR domains. With the PPIase activity, they were classified as immunophilins owing to their tight binding to the immunosuppressants FK506 and rapamycin. FKBP51 and FKBP52 have been shown to participate in the Hsp90–steroid receptor complex and regulate the progression of the Hsp90 conformational cycle [152].

Hsp90 TPR Co-Chaperones in Neurodegenerative Diseases

These TPR co-chaperones have important roles in neurodegenerative diseases. PP5 is a serine/threonine protein phosphatase that can dephosphorylate tau at AD-associated abnormal phosphorylation sites and its activity decreases in AD neocortex [45]. PP5 also protects primary rat cortical neurons from cell death induced by A β [46]. Acting as an E3 ligase for ubiquitination degradation, CHIP has been involved in several neurodegenerative diseases where protein aggregates are a hallmark, such as AD, PD, and HD. Specifically, the Hsp90–CHIP complex is essential for the degradation of phosphorylated tau in AD, and the deletion of CHIP in mice leads to the accumulation of hyperphosphorylated and caspase-3 cleaved tau species [39,47]. In PD, CHIP reduces α -synuclein oligomerization and mediates α -synuclein degradation [48,49]. The overexpression of CHIP in cell culture inhibits α -synuclein inclusion formation and reduces α -synuclein levels [49]. CHIP also reduces the uptake of α -synuclein fibrils by Neuro-2a cells that interfere with the propagation of pathogenic α -synuclein assemblies [50]. Moreover, CHIP binds, ubiquitinates, and promotes the ubiquitin proteasomal degradation of LRRK2 [51,52]. Overexpression of CHIP protects against and knockdown of CHIP exacerbates the toxicity of mutant LRRK2 [51]. In HD, transient overexpression of CHIP increases the ubiquitination and the rate of degradation of polyglutamine-expanded HTT or ataxin-3 [53]. CHIP suppresses polyglutamine aggregation and toxicity, and its haploinsufficiency markedly accelerates disease phenotype in an HD transgenic mouse model [54]. FKBP51, in complex with Hsp90, enhances the production of tau oligomers and prevents tau degradation by the 20S proteasome [55]. The levels of FKBP51 increase with age in the mouse brain, and AD patients have a higher expression of FKBP51 than age-matched controls [55,56]. FKBP51 is also involved in Pink1's regulation of AKT on neuronal survival [57]. Moreover, downregulation of FKBP51 reduces mutant HTT levels in HD models both *in vitro* and *in vivo* [58]. The function of FKBP52 in AD is complicated. FKBP52 induces aggregation of multiple tau species *in vitro* [59–61]. Overexpression of FKBP52 in the hippocampus leads to cognitive impairments and neurotoxicity in aged wild-type mice and rTg4510 transgenic mice [38,62]. However, FKBP52 levels are abnormally low in the frontal cortex of AD brains, as compared with controls [63]. It is suggested that this abnormal decrease of FKBP52 levels in the AD brain hinders autophagy efficiency and contributes to tau pathology [153]. In *Drosophila*, which expresses A β peptides, downregulation of FKBP52 potentiates A β toxicity, while FKBP52 overexpression suppresses A β toxicity and increases the lifespan [64]. Thus, further studies are required to elucidate the roles of FKBP52 in the progression of AD. In PD, FKBP52 overexpression accelerates α -synuclein aggregation and neuronal cell death, whereas knockdown of FKBP52 reduces α -synuclein aggregation and prevents cell death [65]. Moreover, FKBP52 can generate immune responses to α -synuclein-based immunizations in mice [66]. Therefore, modulating the interaction between Hsp90 and these TPR co-chaperones can represent a therapeutic strategy for neurodegenerative diseases.

Hsp90–TPR Co-Chaperone Interaction Inhibitors

Currently, there are only a few publications reporting the inhibitors of the interactions between Hsp90 and TPR co-chaperones. In cell-based studies, **arachidonic acid** (**49**, Figure 9) or **nocodazole** (**50**, Figure 9) can inhibit the interaction of Hsp90 with FLAG-tagged PP5 via the TPR domain of PP5 and increase the phosphatase activity of PP5 [154]. As a derivative of the natural product Sansalvamide A, **SM145** (**51**, Figure 9) binds between the NTD and MD of Hsp90 and allosterically disrupts the interaction between Hsp90 and all tested TPR co-chaperones, such as Hop, FKBP51, FKBP52, FKBP38, and cyclophilin 40 (Cyp40) [155,156]. It is speculated that SM145 induces or stabilizes an Hsp90 conformation that hinders access to MEEVD residues where TPR domains interact [155]. SM145 also induces a caspase-3 dependent apoptotic event and leads to a decrease in hormone receptor protein levels without triggering the heat shock response [155,156]. However, SM145 has poor overall synthetic yields and low solubility. Later, a new analogue, **SM253** (**52**, Figure 9), was synthesized with relatively high overall yields and good aqueous solubility [157]. Another group designed a new compound, **LB76** (**53**, Figure 9), that binds directly to the CTD of Hsp90 and interacts with its MEEVD residues, thereby disrupting the interactions between Hsp90 and TPR co-chaperones, such as Hop, Cyp40, FKBP51, and FKBP38 [158]. LB76 blocked the interaction between Hsp90 CTD and TPR co-chaperones with different IC₅₀ values: 4 μM for Hop, 7.2 μM for Cyp40, 48 μM for FKBP38, and 100 μM for FKBP51. This work provides the possibility to design small molecules that selectively inhibit the interaction between Hsp90 and TPR proteins. In addition, our group has identified a novel compound, **GMP-1** (**54**, Figure 9), that disrupts interactions between Hsp90/70 and the TPR protein, mitochondrial protein import receptor Tom70 [159]. Treatment with GMP-1 demonstrated its neuroprotective effects in mouse and *Drosophila* models of AD.

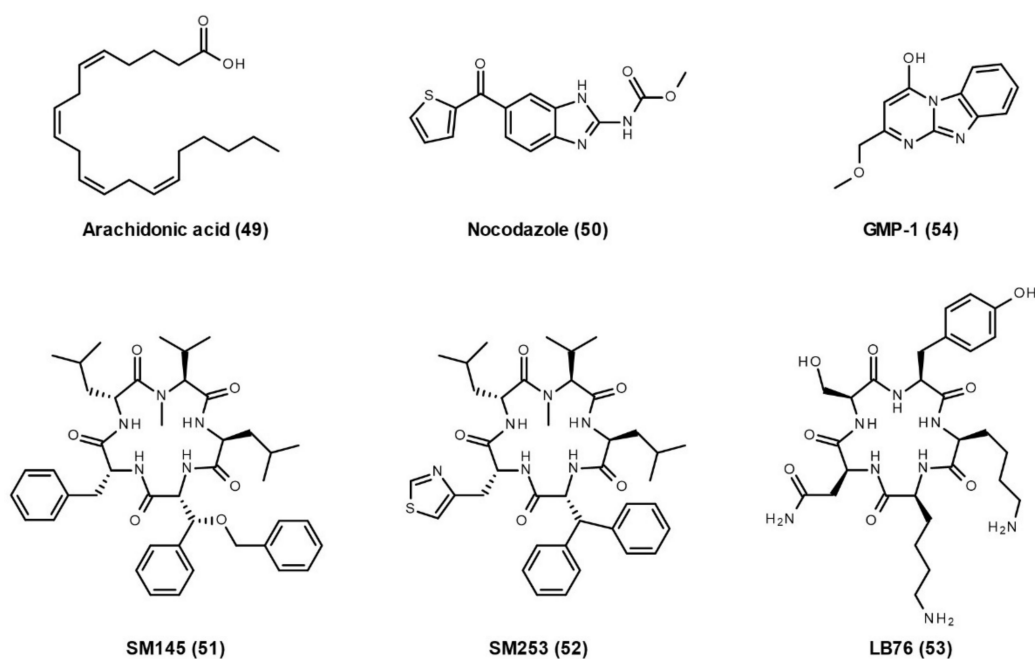


Figure 9. Developed small molecules with the ability to disrupt interactions between Hsp90 and TPR co-chaperones.

Hsp90/Hsp70 Interaction with Dicarboxylate Clamp TPR (dcTPR) Co-Chaperones

Molecular co-chaperones containing the TPR domain interact with Hsp90/Hsp70 via the formation of a dicarboxylate clamp between side chains of the residues in the TPR domain (dcTPR) and two carboxylic acid groups of the C-terminal aspartate or glutamate of the Hsp90/Hsp70 [160]. The residues involved in dicarboxylate clamp formation are conserved in most of the TPR-Hsp90/Hsp70 interactions [161]. In the case of the interaction

between the dcTPR domain of FKBP51 and Hsp90, residues Lys272 and Lys352 are involved in the formation of dicarboxylate clamp with the C-terminal aspartate of Hsp90 through hydrogen bonding and salt bridges (Figure 10). We have recently reported a conservational profile of several dcTPR domain-containing proteins (HOP, FKBP52, CHIP, AIP, FKBP38, FKBP51, and Tah1), which supports the conservative nature of dicarboxylate clamp forming residues. It was found that Lys272, Lys352, Asn322, Glu273, and Lys329 were conserved throughout these proteins, out of which Lys272 and Lys352 are the two residues involved in the formation of the dicarboxylate clamp [161]. In another study, we have identified dcTPR interacting proteins having an acidic C-terminal domain similar to Hsp90/Hsp70 [162].

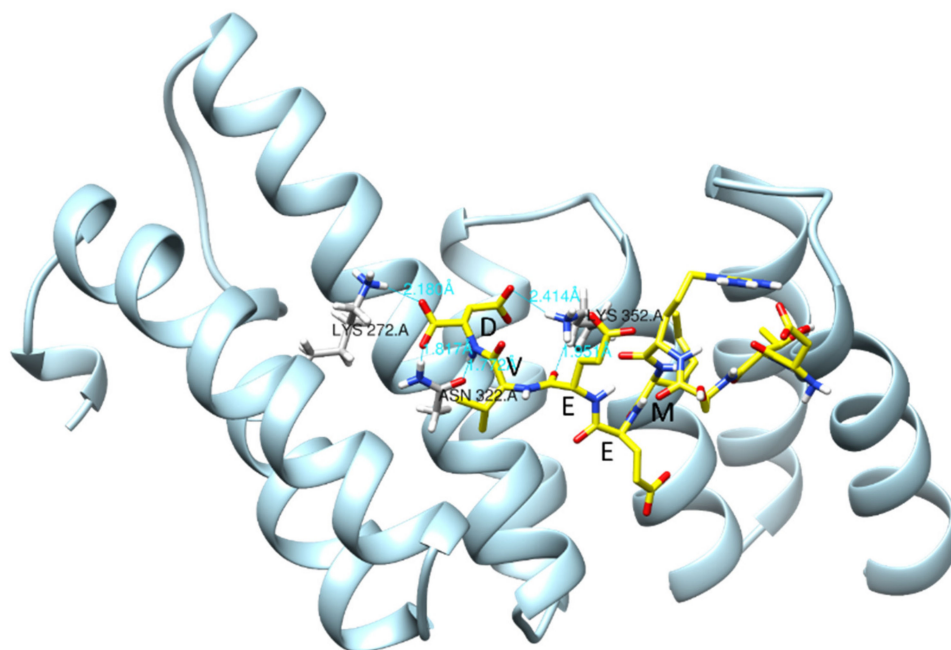


Figure 10. Dicarboxylate clamp mechanism in the interaction of the TPR domain of FKBP51 (shown as ribbon) and C-terminal MEEVD peptide of Hsp90 (shown as bonds with carbons in yellow colour). The two carboxylic acid groups of terminal aspartate of Hsp90 form hydrogen bonds with Lys272 at a distance of 2.18 Å, with Asn322 at a distance of 1.817 Å, and with Lys352 at a distance of 2.414 Å. The figure is generated using PDB id: 5NJX in UCSF-Chimera [163]. For clarity purposes, the FK1 and FK2 domains of FKBP51 are omitted.

The development of ligands targeting the dcTPR domain for inhibition of Hsp90/co-chaperone interaction is considered as an alternate strategy for drug development against these targets. However, there are some limitations such as the charged nature of compounds with which to compete and disruption of the dicarboxylate clamp along a poor pharmacokinetic profile of such compounds [164]. Another challenge in the development of these compounds is the selective inhibition of Hsp90/co-chaperone interactions because of its highly conservative nature. However, our lab is continuing the development of ligands targeting the dcTPR domain using a combined in silico and structural biology approach. To expedite the identification of hits as inhibitors of the dcTPR-Hsp90/Hsp70 interaction, we have developed and validated a high throughput AlphaScreen assay to screen compound libraries [165]. We have obtained several selective and potent inhibitors of the Hsp90–FKBP51 interaction (unpublished results).

4. Conclusions

For most neurodegenerative disorders, only symptomatic treatment is available. There is an urgent need for the development of new therapeutics with disease-modifying properties. Because protein misfolding and aggregation is a major molecular hallmark of these

brain disorders, strategies to modulate Hsp70/Hsp90 molecular chaperone networks are particularly attractive. In addition to general difficulties associated with the development of central nervous system (CNS) drugs, life-long treatment will set an additional hurdle for such molecules in terms of their potential toxicity. Therefore, modulation of the Hsp70/Hsp90 network by interference with small molecules affecting interactions between chaperones and their specific co-chaperones can change neuronal proteostasis and ultimately modify the course of the disease.

Modulation of PPIs through small molecules is generally considered difficult for the following reasons. PPIs often occur at the protein surface interfaces of a flat shape and large area (1500–3000 Å²). This makes it difficult for small molecules to bind with high affinity breaking selective PPIs. Moreover, drugs acting on PPIs tend to have a higher molecular weight (>400 Da) as compared with traditional small molecule drugs (200–500 Da), making it challenging to meet the criteria like Lipinski's "rule of 5" required for CNS drugs.

With regards to chaperone/co-chaperone interactions, two types of PPIs can be observed: PPIs where the interaction area is large, and PPI inhibitors that would allosterically interact with their targets. For example, only allosteric inhibitors of Hsp70 interactions with J-proteins and NEFs have been developed so far. Such PPI inhibitors would lack selectivity towards particular co-chaperones; however, developing Hsp70 isoform-specific molecules can provide sufficient selectivity for modulation of biological processes. On the other side, PPIs of dcTPR co-chaperones with Hsp70/Hsp90 are mediated by a short C-terminal -EEVD peptide. The -EEVD peptide-binding groove of dcTPR proteins has an area of 650–750 Å² [137]. Moreover, several X-ray structures of dcTPR proteins with or without C-terminal peptide of Hsp90/Hsp70 have been solved. This allows to implement computer-aided drug design to obtain small inhibitors of chaperone/co-chaperone PPIs. Again, the selectivity of the inhibitors is a major issue. The binding site for the -EEVD peptide has a similar tertiary structure and conserved amino acids responsible for the interaction, but the overall homology between different dcTPR domains of co-chaperones is below 50% [137]. Theoretically, reasonable specificity for small molecules (modulators) competing for -EEVD peptide can be achieved, as reported for the FKBP51/FKBP52 pair by our group [165]. Most likely, however, it will be impossible to achieve high specificity towards each member of the dcTPR protein family, which, in humans, consists of more than 20 proteins.

Finally, modulation of the molecular chaperone network by inhibition of particular chaperone/co-chaperone interactions with small molecules is an attractive strategy to treat neurodegenerative disorders. However, the development of such PPIs as CNS drugs poses great challenges that need to be resolved at the level of drug potency, selectivity, toxicity, and CNS permeability.

Author Contributions: L.W. was responsible for the Hsp90 review and drafting of the manuscript. L.B. was responsible for the Hsp70 review and drafting of the manuscript. R.K. was responsible for the Hsp90-TPR co-chaperones review and drafting of the manuscript. B.W. and P.F.P. were responsible for the conclusion section and provided mentorship for the manuscript project. All authors have read and agreed to the published version of the manuscript.

Funding: This review was supported by grants from the Swedish Research Council (2018-02843), Alzheimer Foundation (AF940014), Brain Foundation (Fo 2019-0140), Foundation for Geriatric Diseases at Karolinska Institutet (2021-00681), Gunvor and Josef Anérs Foundation, Magnus Bergvalls Foundation, Gun and Bertil Stohnes Foundation, Tore Nilssons Foundation for medical research, Margaretha af Ugglas Foundation, and the Foundation for Old Servants (2020-01028).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Acknowledgments: The authors would like to thank the guest editor (Alexander E. Kabakov) for inviting them to participate in the Special Issue “Role of Heat Shock Proteins: From Molecular Mechanisms to Therapeutic Opportunities”.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Balch, W.E.; Morimoto, R.I.; Dillin, A.; Kelly, J.W. Adapting proteostasis for disease intervention. *Science* **2008**, *319*, 916–919. [[CrossRef](#)]
2. Labbadia, J.; Morimoto, R.I. The biology of proteostasis in aging and disease. *Annu. Rev. Biochem.* **2015**, *84*, 435–464. [[CrossRef](#)] [[PubMed](#)]
3. Zatsepina, O.G.; Evgen'ev, M.B.; Garbuz, D.G. Role of a Heat Shock Transcription Factor and the Major Heat Shock Protein Hsp70 in Memory Formation and Neuroprotection. *Cells* **2021**, *10*, 1638. [[CrossRef](#)] [[PubMed](#)]
4. Bohush, A.; Bieganowski, P.; Filipek, A. Hsp90 and Its Co-Chaperones in Neurodegenerative Diseases. *Int. J. Mol. Sci.* **2019**, *20*, 4976. [[CrossRef](#)] [[PubMed](#)]
5. Maiti, P.; Manna, J.; Veleri, S.; Frautschy, S. Molecular chaperone dysfunction in neurodegenerative diseases and effects of curcumin. *BioMed Res. Int.* **2014**, *2014*, 495091. [[CrossRef](#)]
6. Ambrose, A.J.; Chapman, E. Function, Therapeutic Potential, and Inhibition of Hsp70 Chaperones. *J. Med. Chem.* **2021**, *64*, 7060–7082. [[CrossRef](#)]
7. Wang, L.; Zhang, Q.; You, Q. Targeting the HSP90-CDC37-kinase chaperone cycle: A promising therapeutic strategy for cancer. *Med. Res. Rev.* **2021**. Online ahead of print. [[CrossRef](#)]
8. Bagatell, R.; Paine-Murrieta, G.D.; Taylor, C.W.; Pulcini, E.J.; Akinaga, S.; Benjamin, I.J.; Whitesell, L. Induction of a heat shock factor 1-dependent stress response alters the cytotoxic activity of hsp90-binding agents. *Clin. Cancer Res. Off. J. Am. Assoc. Cancer Res.* **2000**, *6*, 3312–3318.
9. Bickel, D.; Gohlke, H. C-terminal modulators of heat shock protein of 90kDa (HSP90): State of development and modes of action. *Bioorganic Med. Chem.* **2019**, *27*, 115080. [[CrossRef](#)]
10. Dutta Gupta, S.; Bommaka, M.K.; Banerjee, A. Inhibiting protein-protein interactions of Hsp90 as a novel approach for targeting cancer. *Eur. J. Med. Chem.* **2019**, *178*, 48–63. [[CrossRef](#)]
11. Kasza, A.; Hunya, A.; Frank, Z.; Fulop, F.; Torok, Z.; Balogh, G.; Santha, M.; Balind, A.; Bernath, S.; Blundell, K.L.; et al. Dihydropyridine Derivatives Modulate Heat Shock Responses and have a Neuroprotective Effect in a Transgenic Mouse Model of Alzheimer's Disease. *J. Alzheimer's Dis. JAD* **2016**, *53*, 557–571. [[CrossRef](#)]
12. Roe, M.S.; Wahab, B.; Torok, Z.; Horvath, I.; Vigh, L.; Prodromou, C. Dihydropyridines Allosterically Modulate Hsp90 Providing a Novel Mechanism for Heat Shock Protein Co-induction and Neuroprotection. *Front. Mol. Biosci.* **2018**, *5*, 51. [[CrossRef](#)]
13. Mak, O.W.; Sharma, N.; Reynisson, J.; Leung, I.K.H. Discovery of novel Hsp90 C-terminal domain inhibitors that disrupt co-chaperone binding. *Bioorganic Med. Chem. Lett.* **2021**, *38*, 127857. [[CrossRef](#)]
14. Sharma, D.; Masion, D.C. Hsp70 structure, function, regulation and influence on yeast prions. *Protein Pept. Lett.* **2009**, *16*, 571–581. [[CrossRef](#)]
15. Malinverni, D.; Jost Lopez, A.; De Los Rios, P.; Hummer, G.; Barducci, A. Modeling Hsp70/Hsp40 interaction by multi-scale molecular simulations and coevolutionary sequence analysis. *eLife* **2017**, *6*, e23471. [[CrossRef](#)]
16. Kityk, R.; Kopp, J.; Mayer, M.P. Molecular Mechanism of J-Domain-Triggered ATP Hydrolysis by Hsp70 Chaperones. *Mol. Cell* **2018**, *69*, 227–237. [[CrossRef](#)] [[PubMed](#)]
17. Bracher, A.; Verghese, J. The nucleotide exchange factors of Hsp70 molecular chaperones. *Front. Mol. Biosci.* **2015**, *2*, 10. [[CrossRef](#)] [[PubMed](#)]
18. Kampinga, H.H.; Craig, E.A. The HSP70 chaperone machinery: J proteins as drivers of functional specificity. *Nat. Reviews. Mol. Cell Biol.* **2010**, *11*, 579–592. [[CrossRef](#)] [[PubMed](#)]
19. Jinwal, U.K.; Akoury, E.; Abisambra, J.F.; O'Leary, J.C., 3rd; Thompson, A.D.; Blair, L.J.; Jin, Y.; Bacon, J.; Nordhues, B.A.; Cockman, M.; et al. Imbalance of Hsp70 family variants fosters tau accumulation. *FASEB J.: Off. Publ. Fed. Am. Soc. Exp. Biol.* **2013**, *27*, 1450–1459. [[CrossRef](#)] [[PubMed](#)]
20. Fontaine, S.N.; Zheng, D.; Sabbagh, J.J.; Martin, M.D.; Chaput, D.; Darling, A.; Trotter, J.H.; Stothert, A.R.; Nordhues, B.A.; Lussier, A.; et al. DnaJ/Hsc70 chaperone complexes control the extracellular release of neurodegenerative-associated proteins. *EMBO J.* **2016**, *35*, 1537–1549. [[CrossRef](#)] [[PubMed](#)]
21. Nachman, E.; Wentink, A.S.; Maciona, K.; Bousset, L.; Katsinelos, T.; Allinson, K.; Kampinga, H.; McEwan, W.A.; Jahn, T.R.; Melki, R.; et al. Disassembly of Tau fibrils by the human Hsp70 disaggregation machinery generates small seeding-competent species. *J. Biol. Chem.* **2020**, *295*, 9676–9690. [[CrossRef](#)] [[PubMed](#)]
22. Li, X.; Shao, H.; Taylor, I.R.; Gestwicki, J.E. Targeting Allosteric Control Mechanisms in Heat Shock Protein 70 (Hsp70). *Curr. Top. Med. Chem.* **2016**, *16*, 2729–2740. [[CrossRef](#)] [[PubMed](#)]
23. Colvin, T.A.; Gabai, V.L.; Gong, J.; Calderwood, S.K.; Li, H.; Gummuluru, S.; Matchuk, O.N.; Smirnova, S.G.; Orlova, N.V.; Zamulaeva, I.A.; et al. Hsp70-Bag3 interactions regulate cancer-related signaling networks. *Cancer Res.* **2014**, *74*, 4731–4740. [[CrossRef](#)] [[PubMed](#)]

24. Li, X.; Colvin, T.; Rauch, J.N.; Acosta-Alvear, D.; Kampmann, M.; Dunyak, B.; Hann, B.; Aftab, B.T.; Murnane, M.; Cho, M.; et al. Validation of the Hsp70-Bag3 protein-protein interaction as a potential therapeutic target in cancer. *Mol. Cancer Ther.* **2015**, *14*, 642–648. [[CrossRef](#)]
25. Young, Z.T.; Rauch, J.N.; Assimon, V.A.; Jinwal, U.K.; Ahn, M.; Li, X.; Dunyak, B.M.; Ahmad, A.; Carlson, G.A.; Srinivasan, S.R.; et al. Stabilizing the Hsp70-Tau Complex Promotes Turnover in Models of Tauopathy. *Cell Chem. Biol.* **2016**, *23*, 992–1001. [[CrossRef](#)]
26. Miyata, Y.; Li, X.; Lee, H.F.; Jinwal, U.K.; Srinivasan, S.R.; Seguin, S.P.; Young, Z.T.; Brodsky, J.L.; Dickey, C.A.; Sun, D.; et al. Synthesis and initial evaluation of YM-08, a blood-brain barrier permeable derivative of the heat shock protein 70 (Hsp70) inhibitor MKT-077, which reduces tau levels. *ACS Chem. Neurosci.* **2013**, *4*, 930–939. [[CrossRef](#)]
27. Shao, H.; Li, X.; Hayashi, S.; Bertron, J.L.; Schwarz, D.M.C.; Tang, B.C.; Gestwicki, J.E. Inhibitors of heat shock protein 70 (Hsp70) with enhanced metabolic stability reduce tau levels. *Bioorganic Med. Chem. Lett.* **2021**, *41*, 128025. [[CrossRef](#)]
28. Bengoechea, R.; Findlay, A.R.; Bhadra, A.K.; Shao, H.; Stein, K.C.; Pittman, S.K.; Daw, J.A.; Gestwicki, J.E.; True, H.L.; Weihl, C.C. Inhibition of DNAJ-HSP70 interaction improves strength in muscular dystrophy. *J. Clin. Investig.* **2020**, *130*, 4470–4485. [[CrossRef](#)]
29. Schopf, F.H.; Biebl, M.M.; Buchner, J. The HSP90 chaperone machinery. *Nat. Reviews. Mol. Cell Biol.* **2017**, *18*, 345–360. [[CrossRef](#)]
30. Eckl, J.M.; Richter, K. Functions of the Hsp90 chaperone system: Lifting client proteins to new heights. *Int. J. Biochem. Mol. Biol.* **2013**, *4*, 157–165.
31. Jinwal, U.K.; Trotter, J.H.; Abisambra, J.F.; Koren, J., 3rd; Lawson, L.Y.; Vestal, G.D.; O’Leary, J.C., 3rd; Johnson, A.G.; Jin, Y.; Jones, J.R.; et al. The Hsp90 kinase co-chaperone Cdc37 regulates tau stability and phosphorylation dynamics. *J. Biol. Chem.* **2011**, *286*, 16976–16983. [[CrossRef](#)]
32. Jinwal, U.K.; Abisambra, J.F.; Zhang, J.; Dharia, S.; O’Leary, J.C.; Patel, T.; Braswell, K.; Jani, T.; Gestwicki, J.E.; Dickey, C.A. Cdc37/Hsp90 protein complex disruption triggers an autophagic clearance cascade for TDP-43 protein. *J. Biol. Chem.* **2012**, *287*, 24814–24820. [[CrossRef](#)]
33. Sonamoto, R.; Kii, I.; Koike, Y.; Sumida, Y.; Kato-Sumida, T.; Okuno, Y.; Hosoya, T.; Hagiwara, M. Identification of a DYRK1A Inhibitor that Induces Degradation of the Target Kinase using Co-chaperone CDC37 fused with Luciferase nanoKAZ. *Sci. Rep.* **2015**, *5*, 12728. [[CrossRef](#)] [[PubMed](#)]
34. Narayan, M.; Zhang, J.; Braswell, K.; Gibson, C.; Zitnyar, A.; Lee, D.C.; Varghese-Gupta, S.; Jinwal, U.K. Withaferin A Regulates LRRK2 Levels by Interfering with the Hsp90- Cdc37 Chaperone Complex. *Curr. Aging Sci.* **2015**, *8*, 259–265. [[CrossRef](#)] [[PubMed](#)]
35. Moriwaki, Y.; Kim, Y.J.; Ido, Y.; Misawa, H.; Kawashima, K.; Endo, S.; Takahashi, R. L347P PINK1 mutant that fails to bind to Hsp90/Cdc37 chaperones is rapidly degraded in a proteasome-dependent manner. *Neurosci. Res.* **2008**, *61*, 43–48. [[CrossRef](#)] [[PubMed](#)]
36. Weihofen, A.; Ostaszewski, B.; Minami, Y.; Selkoe, D.J. Pink1 Parkinson mutations, the Cdc37/Hsp90 chaperones and Parkin all influence the maturation or subcellular distribution of Pink1. *Hum. Mol. Genet.* **2008**, *17*, 602–616. [[CrossRef](#)]
37. Shelton, L.B.; Baker, J.D.; Zheng, D.; Sullivan, L.E.; Solanki, P.K.; Webster, J.M.; Sun, Z.; Sabbagh, J.J.; Nordhues, B.A.; Koren, J., 3rd; et al. Hsp90 activator Aha1 drives production of pathological tau aggregates. *Proc. Natl. Acad. Sci. USA.* **2017**, *114*, 9707–9712. [[CrossRef](#)] [[PubMed](#)]
38. Criado-Marrero, M.; Gebru, N.T.; Blazier, D.M.; Gould, L.A.; Baker, J.D.; Beaulieu-Abdelahad, D.; Blair, L.J. Hsp90 co-chaperones, FKBP52 and Aha1, promote tau pathogenesis in aged wild-type mice. *Acta Neuropathol. Commun.* **2021**, *9*, 65. [[CrossRef](#)] [[PubMed](#)]
39. Dickey, C.A.; Kamal, A.; Lundgren, K.; Klosak, N.; Bailey, R.M.; Dunmore, J.; Ash, P.; Shoraka, S.; Zlatkovic, J.; Eckman, C.B.; et al. The high-affinity HSP90-CHIP complex recognizes and selectively degrades phosphorylated tau client proteins. *J. Clin. Investig.* **2007**, *117*, 648–658. [[CrossRef](#)] [[PubMed](#)]
40. Rane, A.; Rajagopalan, S.; Ahuja, M.; Thomas, B.; Chinta, S.J.; Andersen, J.K. Hsp90 Co-chaperone p23 contributes to dopaminergic mitochondrial stress via stabilization of PHD2: Implications for Parkinson’s disease. *Neurotoxicology* **2018**, *65*, 166–173. [[CrossRef](#)]
41. Wolfe, K.J.; Ren, H.Y.; Trepte, P.; Cyr, D.M. The Hsp70/90 cochaperone, Sti1, suppresses proteotoxicity by regulating spatial quality control of amyloid-like proteins. *Mol. Biol. Cell* **2013**, *24*, 3588–3602. [[CrossRef](#)] [[PubMed](#)]
42. Ambegaokar, S.S.; Jackson, G.R. Functional genomic screen and network analysis reveal novel modifiers of tauopathy dissociated from tau phosphorylation. *Hum. Mol. Genet.* **2011**, *20*, 4947–4977. [[CrossRef](#)] [[PubMed](#)]
43. Beraldo, F.H.; Arantes, C.P.; Santos, T.G.; Queiroz, N.G.; Young, K.; Rylett, R.J.; Markus, R.P.; Prado, M.A.; Martins, V.R. Role of alpha7 nicotinic acetylcholine receptor in calcium signaling induced by prion protein interaction with stress-inducible protein 1. *J. Biol. Chem.* **2010**, *285*, 36542–36550. [[CrossRef](#)] [[PubMed](#)]
44. Ostapchenko, V.G.; Beraldo, F.H.; Mohammad, A.H.; Xie, Y.F.; Hirata, P.H.; Magalhaes, A.C.; Lamour, G.; Li, H.; Maciejewski, A.; Belrose, J.C.; et al. The prion protein ligand, stress-inducible phosphoprotein 1, regulates amyloid-beta oligomer toxicity. *J. Neurosci. Off. J. Soc. Neurosci.* **2013**, *33*, 16552–16564. [[CrossRef](#)] [[PubMed](#)]
45. Liu, F.; Iqbal, K.; Grundke-Iqbal, I.; Rossie, S.; Gong, C.X. Dephosphorylation of tau by protein phosphatase 5: Impairment in Alzheimer’s disease. *J. Biol. Chem.* **2005**, *280*, 1790–1796. [[CrossRef](#)] [[PubMed](#)]
46. Sanchez-Ortiz, E.; Hahm, B.K.; Armstrong, D.L.; Rossie, S. Protein phosphatase 5 protects neurons against amyloid-beta toxicity. *J. Neurochem.* **2009**, *111*, 391–402. [[CrossRef](#)]

47. Dickey, C.A.; Yue, M.; Lin, W.L.; Dickson, D.W.; Dunmore, J.H.; Lee, W.C.; Zehr, C.; West, G.; Cao, S.; Clark, A.M.; et al. Deletion of the ubiquitin ligase CHIP leads to the accumulation, but not the aggregation, of both endogenous phospho- and caspase-3-cleaved tau species. *J. Neurosci. Off. J. Soc. Neurosci.* **2006**, *26*, 6985–6996. [[CrossRef](#)]
48. Kalia, L.V.; Kalia, S.K.; Chau, H.; Lozano, A.M.; Hyman, B.T.; McLean, P.J. Ubiquitinylation of alpha-synuclein by carboxyl terminus Hsp70-interacting protein (CHIP) is regulated by Bcl-2-associated athanogene 5 (BAG5). *PLoS ONE* **2011**, *6*, e14695. [[CrossRef](#)]
49. Shin, Y.; Klucken, J.; Patterson, C.; Hyman, B.T.; McLean, P.J. The co-chaperone carboxyl terminus of Hsp70-interacting protein (CHIP) mediates alpha-synuclein degradation decisions between proteasomal and lysosomal pathways. *J. Biol. Chem.* **2005**, *280*, 23727–23734. [[CrossRef](#)]
50. Bendifallah, M.; Redeker, V.; Monsellier, E.; Bousset, L.; Bellande, T.; Melki, R. Interaction of the chaperones alpha B-crystallin and CHIP with fibrillar alpha-synuclein: Effects on internalization by cells and identification of interacting interfaces. *Biochem. Biophys. Res. Commun.* **2020**, *527*, 760–769. [[CrossRef](#)]
51. Ko, H.S.; Bailey, R.; Smith, W.W.; Liu, Z.; Shin, J.H.; Lee, Y.I.; Zhang, Y.J.; Jiang, H.; Ross, C.A.; Moore, D.J.; et al. CHIP regulates leucine-rich repeat kinase-2 ubiquitination, degradation, and toxicity. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 2897–2902. [[CrossRef](#)]
52. Ding, X.; Goldberg, M.S. Regulation of LRRK2 stability by the E3 ubiquitin ligase CHIP. *PLoS ONE* **2009**, *4*, e5949. [[CrossRef](#)]
53. Jana, N.R.; Dikshit, P.; Goswami, A.; Kotliarova, S.; Murata, S.; Tanaka, K.; Nukina, N. Co-chaperone CHIP associates with expanded polyglutamine protein and promotes their degradation by proteasomes. *J. Biol. Chem.* **2005**, *280*, 11635–11640. [[CrossRef](#)]
54. Miller, V.M.; Nelson, R.F.; Gouvion, C.M.; Williams, A.; Rodriguez-Lebron, E.; Harper, S.Q.; Davidson, B.L.; Rebagliati, M.R.; Paulson, H.L. CHIP suppresses polyglutamine aggregation and toxicity in vitro and in vivo. *J. Neurosci. Off. J. Soc. Neurosci.* **2005**, *25*, 9152–9161. [[CrossRef](#)]
55. Blair, L.J.; Nordhues, B.A.; Hill, S.E.; Scaglione, K.M.; O’Leary, J.C., 3rd; Fontaine, S.N.; Breydo, L.; Zhang, B.; Li, P.; Wang, L.; et al. Accelerated neurodegeneration through chaperone-mediated oligomerization of tau. *J. Clin. Investig.* **2013**, *123*, 4158–4169. [[CrossRef](#)] [[PubMed](#)]
56. Jinwal, U.K.; Koren, J., 3rd; Borysov, S.I.; Schmid, A.B.; Abisambra, J.F.; Blair, L.J.; Johnson, A.G.; Jones, J.R.; Shults, C.L.; O’Leary, J.C., 3rd; et al. The Hsp90 cochaperone, FKBP51, increases Tau stability and polymerizes microtubules. *J. Neurosci. Off. J. Soc. Neurosci.* **2010**, *30*, 591–599. [[CrossRef](#)] [[PubMed](#)]
57. Boonying, W.; Joselin, A.; Huang, E.; Qu, D.; Safarpour, F.; Iyirhiaro, G.O.; Gonzalez, Y.R.; Callaghan, S.M.; Slack, R.S.; Figeys, D.; et al. Pink1 regulates FKBP5 interaction with AKT/PHLPP and protects neurons from neurotoxin stress induced by MPP(.). *J. Neurochem.* **2019**, *150*, 312–329. [[CrossRef](#)] [[PubMed](#)]
58. Bailus, B.J.; Scheeler, S.M.; Simons, J.; Sanchez, M.A.; Tshilenge, K.T.; Creus-Muncunill, J.; Naphade, S.; Lopez-Ramirez, A.; Zhang, N.; Lakshika Madushani, K.; et al. Modulating FKBP5/FKBP51 and autophagy lowers HTT (huntingtin) levels. *Autophagy* **2021**, *1–22*, Online ahead of print. [[CrossRef](#)] [[PubMed](#)]
59. Giustiniani, J.; Chambraud, B.; Sardin, E.; Dounane, O.; Guillemeau, K.; Nakatani, H.; Paquet, D.; Kamah, A.; Landrieu, I.; Lippens, G.; et al. Immunophilin FKBP52 induces Tau-P301L filamentous assembly in vitro and modulates its activity in a model of tauopathy. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, 4584–4589. [[CrossRef](#)]
60. Giustiniani, J.; Guillemeau, K.; Dounane, O.; Sardin, E.; Huvent, I.; Schmitt, A.; Hamdane, M.; Buee, L.; Landrieu, I.; Lippens, G.; et al. The FK506-binding protein FKBP52 in vitro induces aggregation of truncated Tau forms with prion-like behavior. *FASEB J. Off. Publ. Fed. Am. Soc. Exp. Biol.* **2015**, *29*, 3171–3181. [[CrossRef](#)]
61. Kamah, A.; Cantrelle, F.X.; Huvent, I.; Giustiniani, J.; Guillemeau, K.; Byrne, C.; Jacquot, Y.; Landrieu, I.; Baulieu, E.E.; Smet, C.; et al. Isomerization and Oligomerization of Truncated and Mutated Tau Forms by FKBP52 are Independent Processes. *J. Mol. Biol.* **2016**, *428*, 1080–1090. [[CrossRef](#)] [[PubMed](#)]
62. Criado-Marrero, M.; Gebru, N.T.; Gould, L.A.; Blazier, D.M.; Vidal-Aguilar, Y.; Smith, T.M.; Abdelmaboud, S.S.; Shelton, L.B.; Wang, X.; Dahrendorff, J.; et al. FKBP52 overexpression accelerates hippocampal-dependent memory impairments in a tau transgenic mouse model. *NPJ Aging Mech. Dis.* **2021**, *7*, 9. [[CrossRef](#)] [[PubMed](#)]
63. Giustiniani, J.; Sineus, M.; Sardin, E.; Dounane, O.; Panchal, M.; Sazdovitch, V.; Duyckaerts, C.; Chambraud, B.; Baulieu, E.E. Decrease of the immunophilin FKBP52 accumulation in human brains of Alzheimer’s disease and FTDP-17. *J. Alzheimer’s Dis. JAD* **2012**, *29*, 471–483. [[CrossRef](#)] [[PubMed](#)]
64. Sanokawa-Akakura, R.; Cao, W.; Allan, K.; Patel, K.; Ganesh, A.; Heiman, G.; Burke, R.; Kemp, F.W.; Bogden, J.D.; Camakaris, J.; et al. Control of Alzheimer’s amyloid beta toxicity by the high molecular weight immunophilin FKBP52 and copper homeostasis in *Drosophila*. *PLoS ONE* **2010**, *5*, e8626. [[CrossRef](#)]
65. Gerard, M.; Deleersnijder, A.; Daniels, V.; Schreurs, S.; Munck, S.; Reumers, V.; Pottel, H.; Engelborghs, Y.; Van den Haute, C.; Taymans, J.M.; et al. Inhibition of FK506 binding proteins reduces alpha-synuclein aggregation and Parkinson’s disease-like pathology. *J. Neurosci. Off. J. Soc. Neurosci.* **2010**, *30*, 2454–2463. [[CrossRef](#)]
66. Labrador-Garrido, A.; Cejudo-Guillen, M.; Daturpalli, S.; Leal, M.M.; Klippstein, R.; De Genst, E.J.; Villadiego, J.; Toledo-Aral, J.J.; Dobson, C.M.; Jackson, S.E.; et al. Chaperome screening leads to identification of Grp94/Gp96 and FKBP4/52 as modulators of the alpha-synuclein-elicited immune response. *FASEB J. Off. Publ. Fed. Am. Soc. Exp. Biol.* **2016**, *30*, 564–577. [[CrossRef](#)]
67. Roe, S.M.; Ali, M.M.; Meyer, P.; Vaughan, C.K.; Panaretou, B.; Piper, P.W.; Prodromou, C.; Pearl, L.H. The Mechanism of Hsp90 regulation by the protein kinase-specific cochaperone p50(cdc37). *Cell* **2004**, *116*, 87–98. [[CrossRef](#)]

68. Verba, K.A.; Wang, R.Y.; Arakawa, A.; Liu, Y.; Shirouzu, M.; Yokoyama, S.; Agard, D.A. Atomic structure of Hsp90-Cdc37-Cdk4 reveals that Hsp90 traps and stabilizes an unfolded kinase. *Science* **2016**, *352*, 1542–1547. [[CrossRef](#)]
69. Zhang, T.; Hamza, A.; Cao, X.; Wang, B.; Yu, S.; Zhan, C.G.; Sun, D. A novel Hsp90 inhibitor to disrupt Hsp90/Cdc37 complex against pancreatic cancer cells. *Mol. Cancer Ther.* **2008**, *7*, 162–170. [[CrossRef](#)] [[PubMed](#)]
70. Zhang, T.; Li, Y.; Yu, Y.; Zou, P.; Jiang, Y.; Sun, D. Characterization of celastrol to inhibit hsp90 and cdc37 interaction. *J. Biol. Chem.* **2009**, *284*, 35381–35389. [[CrossRef](#)] [[PubMed](#)]
71. Sreeramulu, S.; Gande, S.L.; Gobel, M.; Schwalbe, H. Molecular mechanism of inhibition of the human protein complex Hsp90-Cdc37, a kinome chaperone-cochaperone, by triterpene celastrol. *Angew. Chem.* **2009**, *48*, 5853–5855. [[CrossRef](#)] [[PubMed](#)]
72. Zanphorlin, L.M.; Alves, F.R.; Ramos, C.H. The effect of celastrol, a triterpene with antitumorigenic activity, on conformational and functional aspects of the human 90kDa heat shock protein Hsp90alpha, a chaperone implicated in the stabilization of the tumor phenotype. *Biochim. Et Biophys. Acta* **2014**, *1840*, 3145–3152. [[CrossRef](#)] [[PubMed](#)]
73. Jiang, F.; Wang, H.J.; Bao, Q.C.; Wang, L.; Jin, Y.H.; Zhang, Q.; Jiang, D.; You, Q.D.; Xu, X.L. Optimization and biological evaluation of celastrol derivatives as Hsp90-Cdc37 interaction disruptors with improved druglike properties. *Bioorganic Med. Chem.* **2016**, *24*, 5431–5439. [[CrossRef](#)] [[PubMed](#)]
74. Li, N.; Xu, M.; Wang, B.; Shi, Z.; Zhao, Z.; Tang, Y.; Wang, X.; Sun, J.; Chen, L. Discovery of Novel Celastrol Derivatives as Hsp90-Cdc37 Interaction Disruptors with Antitumor Activity. *J. Med. Chem.* **2019**, *62*, 10798–10815. [[CrossRef](#)] [[PubMed](#)]
75. Shi, J.; Li, J.; Xu, Z.; Chen, L.; Luo, R.; Zhang, C.; Gao, F.; Zhang, J.; Fu, C. Celastrol: A Review of Useful Strategies Overcoming its Limitation in Anticancer Application. *Front. Pharmacol.* **2020**, *11*, 558741. [[CrossRef](#)] [[PubMed](#)]
76. Bai, X.; Fu, R.J.; Zhang, S.; Yue, S.J.; Chen, Y.Y.; Xu, D.Q.; Tang, Y.P. Potential medicinal value of celastrol and its synthesized analogues for central nervous system diseases. *Biomed. Pharmacother. Biomed. Pharmacother.* **2021**, *139*, 111551. [[CrossRef](#)] [[PubMed](#)]
77. Yu, Y.; Hamza, A.; Zhang, T.; Gu, M.; Zou, P.; Newman, B.; Li, Y.; Gunatilaka, A.A.; Zhan, C.G.; Sun, D. Withaferin A targets heat shock protein 90 in pancreatic cancer cells. *Biochem. Pharmacol.* **2010**, *79*, 542–551. [[CrossRef](#)]
78. Grover, A.; Shandilya, A.; Agrawal, V.; Pratik, P.; Bhasme, D.; Bisaria, V.S.; Sundar, D. Hsp90/Cdc37 chaperone/co-chaperone complex, a novel junction anticancer target elucidated by the mode of action of herbal drug Withaferin A. *BMC Bioinform.* **2011**, *12* (Suppl. S1), S30. [[CrossRef](#)] [[PubMed](#)]
79. Gu, M.; Yu, Y.; Gunaherath, G.M.; Gunatilaka, A.A.; Li, D.; Sun, D. Structure-activity relationship (SAR) of withanolides to inhibit Hsp90 for its activity in pancreatic cancer cells. *Investig. New Drugs* **2014**, *32*, 68–74. [[CrossRef](#)]
80. Behl, T.; Sharma, A.; Sharma, L.; Sehgal, A.; Zengin, G.; Brata, R.; Fratila, O.; Bungau, S. Exploring the Multifaceted Therapeutic Potential of Withaferin A and Its Derivatives. *Biomedicines* **2020**, *8*, 571. [[CrossRef](#)] [[PubMed](#)]
81. Li, Y.; Karagoz, G.E.; Seo, Y.H.; Zhang, T.; Jiang, Y.; Yu, Y.; Duarte, A.M.; Schwartz, S.J.; Boelens, R.; Carroll, K.; et al. Sulforaphane inhibits pancreatic cancer through disrupting Hsp90-p50(Cdc37) complex and direct interactions with amino acids residues of Hsp90. *J. Nutr. Biochem.* **2012**, *23*, 1617–1626. [[CrossRef](#)]
82. Kim, J. Pre-Clinical Neuroprotective Evidences and Plausible Mechanisms of Sulforaphane in Alzheimer's Disease. *Int. J. Mol. Sci.* **2021**, *22*, 2929. [[CrossRef](#)] [[PubMed](#)]
83. Schepici, G.; Bramanti, P.; Mazzon, E. Efficacy of Sulforaphane in Neurodegenerative Diseases. *Int. J. Mol. Sci.* **2020**, *21*, 8637. [[CrossRef](#)] [[PubMed](#)]
84. Huang, W.; Ye, M.; Zhang, L.R.; Wu, Q.D.; Zhang, M.; Xu, J.H.; Zheng, W. FW-04-806 inhibits proliferation and induces apoptosis in human breast cancer cells by binding to N-terminus of Hsp90 and disrupting Hsp90-Cdc37 complex formation. *Mol. Cancer* **2014**, *13*, 150. [[CrossRef](#)] [[PubMed](#)]
85. Siddiqui, F.A.; Parkkola, H.; Vukic, V.; Oetken-Lindholm, C.; Jaiswal, A.; Kiriazis, A.; Pavic, K.; Aittokallio, T.; Salminen, T.A.; Abankwa, D. Novel Small Molecule Hsp90/Cdc37 Interface Inhibitors Indirectly Target K-Ras-Signaling. *Cancers* **2021**, *13*, 927. [[CrossRef](#)] [[PubMed](#)]
86. Li, D.; Li, C.; Li, L.; Chen, S.; Wang, L.; Li, Q.; Wang, X.; Lei, X.; Shen, Z. Natural Product Kongensin A is a Non-Canonical HSP90 Inhibitor that Blocks RIP3-dependent Necroptosis. *Cell Chem. Biol.* **2016**, *23*, 257–266. [[CrossRef](#)]
87. Bai, Y.; Lam, H.C.; Lei, X. Dissecting Programmed Cell Death with Small Molecules. *Acc. Chem. Res.* **2020**, *53*, 1034–1045. [[CrossRef](#)]
88. Li, T.; Chen, X.; Dai, X.Y.; Wei, B.; Weng, Q.J.; Chen, X.; Ouyang, D.F.; Yan, R.; Huang, Z.J.; Jiang, H.L.; et al. Novel Hsp90 inhibitor platycodin D disrupts Hsp90/Cdc37 complex and enhances the anticancer effect of mTOR inhibitor. *Toxicol. Appl. Pharmacol.* **2017**, *330*, 65–73. [[CrossRef](#)]
89. Patel, D.; Shukla, S.; Gupta, S. Apigenin and cancer chemoprevention: Progress, potential and promise (review). *Int. J. Oncol.* **2007**, *30*, 233–245. [[CrossRef](#)] [[PubMed](#)]
90. Ayooobi, F.; Shamsizadeh, A.; Fatemi, I.; Vakilian, A.; Allahtavakoli, M.; Hassanshahi, G.; Moghadam-Ahmadi, A. Bio-effectiveness of the main flavonoids of *Achillea millefolium* in the pathophysiology of neurodegenerative disorders- a review. *Iran. J. Basic Med. Sci.* **2017**, *20*, 604–612. [[CrossRef](#)]
91. Zhao, M.; Ma, J.; Zhu, H.Y.; Zhang, X.H.; Du, Z.Y.; Xu, Y.J.; Yu, X.D. Apigenin inhibits proliferation and induces apoptosis in human multiple myeloma cells through targeting the trinity of CK2, Cdc37 and Hsp90. *Mol. Cancer* **2011**, *10*, 104. [[CrossRef](#)]

92. Jin, L.; Huang, R.; Huang, X.; Zhang, B.; Ji, M.; Wang, H. Discovery of 18beta-glycyrrhetic acid conjugated aminobenzothiazole derivatives as Hsp90-Cdc37 interaction disruptors that inhibit cell migration and reverse drug resistance. *Bioorganic Med. Chem.* **2018**, *26*, 1759–1775. [[CrossRef](#)] [[PubMed](#)]
93. Wang, L.; Bao, Q.-C.; Xu, X.-L.; Jiang, F.; Gu, K.; Jiang, Z.-Y.; Zhang, X.-J.; Guo, X.-K.; You, Q.-D.; Sun, H.-P. Discovery and identification of Cdc37-derived peptides targeting the Hsp90–Cdc37 protein–protein interaction. *RSC Adv.* **2015**, *5*, 96138–96145. [[CrossRef](#)]
94. Wang, L.; Li, L.; Fu, W.T.; Jiang, Z.Y.; You, Q.D.; Xu, X.L. Optimization and bioevaluation of Cdc37-derived peptides: An insight into Hsp90–Cdc37 protein–protein interaction modulators. *Bioorganic Med. Chem.* **2017**, *25*, 233–240. [[CrossRef](#)] [[PubMed](#)]
95. D’Annessa, I.; Hurwitz, N.; Pirota, V.; Beretta, G.L.; Tinelli, S.; Woodford, M.; Freccero, M.; Mollapour, M.; Zaffaroni, N.; Wolfson, H.; et al. Design of Disruptors of the Hsp90–Cdc37 Interface. *Molecules* **2020**, *25*, 360. [[CrossRef](#)] [[PubMed](#)]
96. Jiang, Y.; Bernard, D.; Yu, Y.; Xie, Y.; Zhang, T.; Li, Y.; Burnett, J.P.; Fu, X.; Wang, S.; Sun, D. Split Renilla luciferase protein fragment-assisted complementation (SRL-PFAC) to characterize Hsp90–Cdc37 complex and identify critical residues in protein/protein interactions. *J. Biol. Chem.* **2010**, *285*, 21023–21036. [[CrossRef](#)]
97. Wang, L.; Li, L.; Zhou, Z.H.; Jiang, Z.Y.; You, Q.D.; Xu, X.L. Structure-based virtual screening and optimization of modulators targeting Hsp90–Cdc37 interaction. *Eur. J. Med. Chem.* **2017**, *136*, 63–73. [[CrossRef](#)]
98. Chen, X.; Liu, P.; Wang, Q.; Li, Y.; Fu, L.; Fu, H.; Zhu, J.; Chen, Z.; Zhu, W.; Xie, C.; et al. DCZ3112, a novel Hsp90 inhibitor, exerts potent antitumor activity against HER2-positive breast cancer through disruption of Hsp90–Cdc37 interaction. *Cancer Lett.* **2018**, *434*, 70–80. [[CrossRef](#)]
99. Wang, L.; Zhang, L.; Li, L.; Jiang, J.; Zheng, Z.; Shang, J.; Wang, C.; Chen, W.; Bao, Q.; Xu, X.; et al. Small-molecule inhibitor targeting the Hsp90–Cdc37 protein–protein interaction in colorectal cancer. *Sci. Adv.* **2019**, *5*, eaax2277. [[CrossRef](#)]
100. Wang, L.; Jiang, J.; Zhang, L.; Zhang, Q.; Zhou, J.; Li, L.; Xu, X.; You, Q. Discovery and Optimization of Small Molecules Targeting the Protein–Protein Interaction of Heat Shock Protein 90 (Hsp90) and Cell Division Cycle 37 as Orally Active Inhibitors for the Treatment of Colorectal Cancer. *J. Med. Chem.* **2020**, *63*, 1281–1297. [[CrossRef](#)]
101. Zhang, Q.; Wu, X.; Zhou, J.; Zhang, L.; Xu, X.; Zhang, L.; You, Q.; Wang, L. Design, synthesis and bioevaluation of inhibitors targeting HSP90–CDC37 protein–protein interaction based on a hydrophobic core. *Eur. J. Med. Chem.* **2021**, *210*, 112959. [[CrossRef](#)]
102. Chen, B.; Wei, W.; Ma, L.; Yang, B.; Gill, R.M.; Chua, M.S.; Butte, A.J.; So, S. Computational Discovery of Niclosamide Ethanolamine, a Repurposed Drug Candidate That Reduces Growth of Hepatocellular Carcinoma Cells In Vitro and in Mice by Inhibiting Cell Division Cycle 37 Signaling. *Gastroenterology* **2017**, *152*, 2022–2036. [[CrossRef](#)]
103. Park, J.S.; Lee, Y.S.; Lee, D.H.; Bae, S.H. Repositioning of niclosamide ethanolamine (NEN), an anthelmintic drug, for the treatment of lipotoxicity. *Free Radic. Biol. Med.* **2019**, *137*, 143–157. [[CrossRef](#)]
104. Han, P.; Weng, W.; Chen, Y.; Cai, Y.; Wang, Y.; Wang, M.; Zhan, H.; Yuan, C.; Yu, X.; Shao, M.; et al. Niclosamide ethanolamine attenuates systemic lupus erythematosus and lupus nephritis in MRL/lpr mice. *Am. J. Transl. Res.* **2020**, *12*, 5015–5031.
105. Li, S.L.; Yan, J.; Zhang, Y.Q.; Zhen, C.L.; Liu, M.Y.; Jin, J.; Gao, J.L.; Xiao, X.L.; Shen, X.; Tai, Y.; et al. Niclosamide ethanolamine inhibits artery constriction. *Pharmacol. Res.* **2017**, *115*, 78–86. [[CrossRef](#)] [[PubMed](#)]
106. Siddiqui, F.A.; Parkkola, H.; Manoharan, G.B.; Abankwa, D. Medium-Throughput Detection of Hsp90/Cdc37 Protein–Protein Interaction Inhibitors Using a Split Renilla Luciferase-Based Assay. *SLAS Discov. Adv. Life Sci. R D* **2020**, *25*, 195–206. [[CrossRef](#)]
107. Panaretou, B.; Siligardi, G.; Meyer, P.; Maloney, A.; Sullivan, J.K.; Singh, S.; Millson, S.H.; Clarke, P.A.; Naaby-Hansen, S.; Stein, R.; et al. Activation of the ATPase activity of hsp90 by the stress-regulated cochaperone aha1. *Mol. Cell* **2002**, *10*, 1307–1318. [[CrossRef](#)]
108. Wolmarans, A.; Lee, B.; Spyropoulos, L.; LaPointe, P. The Mechanism of Hsp90 ATPase Stimulation by Aha1. *Sci. Rep.* **2016**, *6*, 33179. [[CrossRef](#)] [[PubMed](#)]
109. Meyer, P.; Prodromou, C.; Liao, C.; Hu, B.; Roe, S.M.; Vaughan, C.K.; Vlastic, I.; Panaretou, B.; Piper, P.W.; Pearl, L.H. Structural basis for recruitment of the ATPase activator Aha1 to the Hsp90 chaperone machinery. *EMBO J.* **2004**, *23*, 1402–1410. [[CrossRef](#)]
110. Koulov, A.V.; LaPointe, P.; Lu, B.; Razvi, A.; Coppinger, J.; Dong, M.Q.; Matteson, J.; Laister, R.; Arrowsmith, C.; Yates, J.R., 3rd; et al. Biological and structural basis for Aha1 regulation of Hsp90 ATPase activity in maintaining proteostasis in the human disease cystic fibrosis. *Mol. Biol. Cell* **2010**, *21*, 871–884. [[CrossRef](#)] [[PubMed](#)]
111. Retzlaff, M.; Hagn, F.; Mitschke, L.; Hessling, M.; Gugel, F.; Kessler, H.; Richter, K.; Buchner, J. Asymmetric activation of the hsp90 dimer by its cochaperone aha1. *Mol. Cell* **2010**, *37*, 344–354. [[CrossRef](#)]
112. Liu, Y.; Sun, M.; Myasnikov, A.G.; Elnatan, D.; Delaeter, N.; Nguyenquang, M.; Agard, D. Cryo-EM structures reveal a multistep mechanism of Hsp90 activation by co-chaperone Aha1. *Biorxiv* **2020**. [[CrossRef](#)]
113. Oroz, J.; Blair, L.J.; Zweckstetter, M. Dynamic Aha1 co-chaperone binding to human Hsp90. *Protein Sci. A Publ. Protein Soc.* **2019**, *28*, 1545–1551. [[CrossRef](#)]
114. Ihrig, V.; Obermann, W.M.J. Identifying Inhibitors of the Hsp90–Aha1 Protein Complex, a Potential Target to Drug Cystic Fibrosis, by Alpha Technology. *SLAS Discov. Adv. Life Sci. R D* **2017**, *22*, 923–928. [[CrossRef](#)]
115. Stiegler, S.C.; Rubbelke, M.; Korotkov, V.S.; Weiwad, M.; John, C.; Fischer, G.; Sieber, S.A.; Sattler, M.; Buchner, J. A chemical compound inhibiting the Aha1–Hsp90 chaperone complex. *J. Biol. Chem.* **2017**, *292*, 17073–17083. [[CrossRef](#)]
116. Singh, J.K.; Hutt, D.M.; Tait, B.; Guy, N.C.; Sivils, J.C.; Ortiz, N.R.; Payan, A.N.; Komaragiri, S.K.; Owens, J.J.; Culbertson, D.; et al. Management of Hsp90-Dependent Protein Folding by Small Molecules Targeting the Aha1 Co-Chaperone. *Cell Chem. Biol.* **2020**, *27*, 292–305.e296. [[CrossRef](#)]

117. Ali, M.M.; Roe, S.M.; Vaughan, C.K.; Meyer, P.; Panaretou, B.; Piper, P.W.; Prodromou, C.; Pearl, L.H. Crystal structure of an Hsp90-nucleotide-p23/Sba1 closed chaperone complex. *Nature* **2006**, *440*, 1013–1017. [[CrossRef](#)]
118. Martinez-Yamout, M.A.; Venkitakrishnan, R.P.; Preece, N.E.; Kroon, G.; Wright, P.E.; Dyson, H.J. Localization of sites of interaction between p23 and Hsp90 in solution. *J. Biol. Chem.* **2006**, *281*, 14457–14464. [[CrossRef](#)] [[PubMed](#)]
119. Karagoz, G.E.; Duarte, A.M.; Ippel, H.; Uetrecht, C.; Sinnige, T.; van Rosmalen, M.; Hausmann, J.; Heck, A.J.; Boelens, R.; Rudiger, S.G. N-terminal domain of human Hsp90 triggers binding to the cochaperone p23. *Proc. Natl. Acad. Sci. USA* **2011**, *108*, 580–585. [[CrossRef](#)] [[PubMed](#)]
120. McLaughlin, S.H.; Sobott, F.; Yao, Z.P.; Zhang, W.; Nielsen, P.R.; Grossmann, J.G.; Laue, E.D.; Robinson, C.V.; Jackson, S.E. The co-chaperone p23 arrests the Hsp90 ATPase cycle to trap client proteins. *J. Mol. Biol.* **2006**, *356*, 746–758. [[CrossRef](#)] [[PubMed](#)]
121. Richter, K.; Walter, S.; Buchner, J. The Co-chaperone Sba1 connects the ATPase reaction of Hsp90 to the progression of the chaperone cycle. *J. Mol. Biol.* **2004**, *342*, 1403–1413. [[CrossRef](#)]
122. Rehn, A.B.; Buchner, J. p23 and Aha1. *Sub-Cell. Biochem.* **2015**, *78*, 113–131. [[CrossRef](#)]
123. Chadli, A.; Felts, S.J.; Wang, Q.; Sullivan, W.P.; Botuyan, M.V.; Fauq, A.; Ramirez-Alvarado, M.; Mer, G. Celastrol inhibits Hsp90 chaperoning of steroid receptors by inducing fibrillization of the Co-chaperone p23. *J. Biol. Chem.* **2010**, *285*, 4224–4231. [[CrossRef](#)]
124. Patwardhan, C.A.; Fauq, A.; Peterson, L.B.; Miller, C.; Blagg, B.S.; Chadli, A. Gedunin inactivates the co-chaperone p23 protein causing cancer cell death by apoptosis. *J. Biol. Chem.* **2013**, *288*, 7313–7325. [[CrossRef](#)]
125. Gorantla, N.V.; Das, R.; Chidambaram, H.; Dubey, T.; Mulani, F.A.; Thulasiram, H.V.; Chinnathambi, S. Basic Limonoid modulates Chaperone-mediated Proteostasis and dissolve Tau fibrils. *Sci. Rep.* **2020**, *10*, 4023. [[CrossRef](#)]
126. Tom, S.; Rane, A.; Katewa, A.S.; Chamoli, M.; Matsumoto, R.R.; Andersen, J.K.; Chinta, S.J. Gedunin Inhibits Oligomeric Abeta1-42-Induced Microglia Activation Via Modulation of Nrf2-NF-kappaB Signaling. *Mol. Neurobiol.* **2019**, *56*, 7851–7862. [[CrossRef](#)]
127. Yang, W.; Xie, J.; Qiang, Q.; Li, L.; Lin, X.; Ren, Y.; Ren, W.; Liu, Q.; Zhou, G.; Wei, W.; et al. Gedunin Degrades Aggregates of Mutant Huntingtin Protein and Intranuclear Inclusions via the Proteasomal Pathway in Neurons and Fibroblasts from Patients with Huntington’s Disease. *Neurosci. Bull.* **2019**, *35*, 1024–1034. [[CrossRef](#)]
128. Nie, S.; Xu, Y.; Chen, G.; Ma, K.; Han, C.; Guo, Z.; Zhang, Z.; Ye, K.; Cao, X. Small molecule TrkB agonist deoxygedunin protects nigrostriatal dopaminergic neurons from 6-OHDA and MPTP induced neurotoxicity in rodents. *Neuropharmacology* **2015**, *99*, 448–458. [[CrossRef](#)]
129. Hall, J.A.; Seedarala, S.; Rice, N.; Kopel, L.; Halaweish, F.; Blagg, B.S. Cucurbitacin D Is a Disruptor of the HSP90 Chaperone Machinery. *J. Nat. Prod.* **2015**, *78*, 873–879. [[CrossRef](#)]
130. He, Y.; Peng, S.; Wang, J.; Chen, H.; Cong, X.; Chen, A.; Hu, M.; Qin, M.; Wu, H.; Gao, S.; et al. Ailanthone targets p23 to overcome MDV3100 resistance in castration-resistant prostate cancer. *Nat. Commun.* **2016**, *7*, 13122. [[CrossRef](#)]
131. Chan, C.T.; Reeves, R.E.; Geller, R.; Yaghoubi, S.S.; Hoehne, A.; Solow-Cordero, D.E.; Chiosis, G.; Massoud, T.F.; Paulmurugan, R.; Gambhir, S.S. Discovery and validation of small-molecule heat-shock protein 90 inhibitors through multimodality molecular imaging in living subjects. *Proc. Natl. Acad. Sci. USA* **2012**, *109*, E2476–E2485. [[CrossRef](#)]
132. Jensen, M.R.; Schoepfer, J.; Radimerski, T.; Massey, A.; Guy, C.T.; Brueggen, J.; Quadt, C.; Buckler, A.; Cozens, R.; Drysdale, M.J.; et al. NVP-AUY922: A small molecule HSP90 inhibitor with potent antitumor activity in preclinical breast cancer models. *Breast Cancer Res. BCR* **2008**, *10*, R33. [[CrossRef](#)]
133. Menezes, D.L.; Taverna, P.; Jensen, M.R.; Abrams, T.; Stuart, D.; Yu, G.K.; Duhl, D.; Machajewski, T.; Sellers, W.R.; Pryer, N.K.; et al. The novel oral Hsp90 inhibitor NVP-HSP990 exhibits potent and broad-spectrum antitumor activities in vitro and in vivo. *Mol. Cancer Ther.* **2012**, *11*, 730–739. [[CrossRef](#)]
134. Eachkoti, R.; Reddy, M.V.; Lieu, Y.K.; Cosenza, S.C.; Reddy, E.P. Identification and characterisation of a novel heat shock protein 90 inhibitor ONO4140. *Eur. J. Cancer* **2014**, *50*, 1982–1992. [[CrossRef](#)]
135. Xue, N.; Jin, J.; Liu, D.; Yan, R.; Zhang, S.; Yu, X.; Chen, X. Antiproliferative effect of HSP90 inhibitor Y306zh against pancreatic cancer is mediated by interruption of AKT and MAPK signaling pathways. *Curr. Cancer Drug Targets* **2014**, *14*, 671–683. [[CrossRef](#)]
136. Blatch, G.L.; Lassel, M. The tetratricopeptide repeat: A structural motif mediating protein-protein interactions. *BioEssays News Rev. Mol. Cell. Dev. Biol.* **1999**, *21*, 932–939. [[CrossRef](#)]
137. Scheufler, C.; Brinker, A.; Bourenkov, G.; Pegoraro, S.; Moroder, L.; Bartunik, H.; Hartl, F.U.; Moarefi, I. Structure of TPR domain-peptide complexes: Critical elements in the assembly of the Hsp70-Hsp90 multichaperone machine. *Cell* **2000**, *101*, 199–210. [[CrossRef](#)]
138. Schmid, A.B.; Lagleder, S.; Grawert, M.A.; Rohl, A.; Hagn, F.; Wandinger, S.K.; Cox, M.B.; Demmer, O.; Richter, K.; Groll, M.; et al. The architecture of functional modules in the Hsp90 co-chaperone Sti1/Hop. *EMBO J.* **2012**, *31*, 1506–1517. [[CrossRef](#)]
139. Lackie, R.E.; Maciejewski, A.; Ostapchenko, V.G.; Marques-Lopes, J.; Choy, W.Y.; Duennwald, M.L.; Prado, V.F.; Prado, M.A.M. The Hsp70/Hsp90 Chaperone Machinery in Neurodegenerative Diseases. *Front. Neurosci.* **2017**, *11*, 254. [[CrossRef](#)]
140. Maciejewski, A.; Ostapchenko, V.G.; Beraldo, F.H.; Prado, V.F.; Prado, M.A.; Choy, W.Y. Domains of STIP1 responsible for regulating PrPC-dependent amyloid-beta oligomer toxicity. *Biochem. J.* **2016**, *473*, 2119–2130. [[CrossRef](#)]
141. Yi, F.; Regan, L. A novel class of small molecule inhibitors of Hsp90. *ACS Chem. Biol.* **2008**, *3*, 645–654. [[CrossRef](#)] [[PubMed](#)]
142. Pimienta, G.; Herbert, K.M.; Regan, L. A compound that inhibits the HOP-Hsp90 complex formation and has unique killing effects in breast cancer cell lines. *Mol. Pharm.* **2011**, *8*, 2252–2261. [[CrossRef](#)] [[PubMed](#)]

143. Horibe, T.; Kohno, M.; Haramoto, M.; Ohara, K.; Kawakami, K. Designed hybrid TPR peptide targeting Hsp90 as a novel anticancer agent. *J. Transl. Med.* **2011**, *9*, 8. [[CrossRef](#)] [[PubMed](#)]
144. Gupta, U.K.; Mahanta, S.; Paul, S. In silico design of small peptide-based Hsp90 inhibitor: A novel anticancer agent. *Med Hypotheses* **2013**, *81*, 853–861. [[CrossRef](#)] [[PubMed](#)]
145. Wang, W.; Liu, Y.; Zhao, Z.; Xie, C.; Xu, Y.; Hu, Y.; Quan, H.; Lou, L. Y-632 inhibits heat shock protein 90 (Hsp90) function by disrupting the interaction between Hsp90 and Hsp70/Hsp90 organizing protein, and exerts antitumor activity in vitro and in vivo. *Cancer Sci.* **2016**, *107*, 782–790. [[CrossRef](#)]
146. Darby, J.F.; Vidler, L.R.; Simpson, P.J.; Al-Lazikani, B.; Matthews, S.J.; Sharp, S.Y.; Pearl, L.H.; Hoelder, S.; Workman, P. Solution structure of the Hop TPR2A domain and investigation of target druggability by NMR, biochemical and in silico approaches. *Sci. Rep.* **2020**, *10*, 16000. [[CrossRef](#)]
147. Kang, H.; Sayner, S.L.; Gross, K.L.; Russell, L.C.; Chinkers, M. Identification of amino acids in the tetratricopeptide repeat and C-terminal domains of protein phosphatase 5 involved in autoinhibition and lipid activation. *Biochemistry* **2001**, *40*, 10485–10490. [[CrossRef](#)]
148. Wandinger, S.K.; Suhre, M.H.; Wegele, H.; Buchner, J. The phosphatase Ppt1 is a dedicated regulator of the molecular chaperone Hsp90. *EMBO J.* **2006**, *25*, 367–376. [[CrossRef](#)]
149. Soroka, J.; Wandinger, S.K.; Mausbacher, N.; Schreiber, T.; Richter, K.; Daub, H.; Buchner, J. Conformational switching of the molecular chaperone Hsp90 via regulated phosphorylation. *Mol. Cell* **2012**, *45*, 517–528. [[CrossRef](#)]
150. Ballinger, C.A.; Connell, P.; Wu, Y.; Hu, Z.; Thompson, L.J.; Yin, L.Y.; Patterson, C. Identification of CHIP, a novel tetratricopeptide repeat-containing protein that interacts with heat shock proteins and negatively regulates chaperone functions. *Mol. Cell. Biol.* **1999**, *19*, 4535–4545. [[CrossRef](#)]
151. Edkins, A.L. CHIP: A co-chaperone for degradation by the proteasome. *Sub-Cell. Biochem.* **2015**, *78*, 219–242. [[CrossRef](#)]
152. Storer, C.L.; Dickey, C.A.; Galigniana, M.D.; Rein, T.; Cox, M.B. FKBP51 and FKBP52 in signaling and disease. *Trends Endocrinol. Metab. TEM* **2011**, *22*, 481–490. [[CrossRef](#)] [[PubMed](#)]
153. Chambraud, B.; Daguinot, C.; Guillemeau, K.; Genet, M.; Dounane, O.; Meduri, G.; Pous, C.; Baulieu, E.E.; Giustiniani, J. Decrease of neuronal FKBP4/FKBP52 modulates perinuclear lysosomal positioning and MAPT/Tau behavior during MAPT/Tau-induced proteotoxic stress. *Autophagy* **2021**, 1–20. [[CrossRef](#)] [[PubMed](#)]
154. Zeke, T.; Morrice, N.; Vazquez-Martin, C.; Cohen, P.T. Human protein phosphatase 5 dissociates from heat-shock proteins and is proteolytically activated in response to arachidonic acid and the microtubule-depolymerizing drug nocodazole. *Biochem. J.* **2005**, *385*, 45–56. [[CrossRef](#)] [[PubMed](#)]
155. Ardi, V.C.; Alexander, L.D.; Johnson, V.A.; McAlpine, S.R. Macrocycles that inhibit the binding between heat shock protein 90 and TPR-containing proteins. *ACS Chem. Biol.* **2011**, *6*, 1357–1366. [[CrossRef](#)] [[PubMed](#)]
156. McConnell, J.R.; Alexander, L.A.; McAlpine, S.R. A heat shock protein 90 inhibitor that modulates the immunophilins and regulates hormone receptors without inducing the heat shock response. *Bioorganic Med. Chem. Lett.* **2014**, *24*, 661–666. [[CrossRef](#)]
157. Koay, Y.C.; McConnell, J.R.; Wang, Y.; Kim, S.J.; Buckton, L.K.; Mansour, F.; McAlpine, S.R. Chemically accessible hsp90 inhibitor that does not induce a heat shock response. *ACS Med. Chem. Lett.* **2014**, *5*, 771–776. [[CrossRef](#)]
158. Rahimi, M.N.; McAlpine, S.R. Protein-protein inhibitor designed de novo to target the MEEVD region on the C-terminus of Hsp90 and block co-chaperone activity. *Chem. Commun.* **2019**, *55*, 846–849. [[CrossRef](#)]
159. Pavlov, P.F.; Hutter-Paier, B.; Havas, D.; Windisch, M.; Winblad, B. Development of GMP-1 a molecular chaperone network modulator protecting mitochondrial function and its assessment in fly and mice models of Alzheimer’s disease. *J. Cell. Mol. Med.* **2018**, *22*, 3464–3474. [[CrossRef](#)]
160. Kumar, R.; Winblad, B.; Pavlov, P.F. Hsp90 as a Member of Dicarboxylate Clamp TPR Protein Interaction Network: Implication in Human Diseases and Prospect as a Drug Target. In *Heat Shock Protein 90 in Human Diseases and Disorders*; Kaur, P., Asea, A.A.A., Eds.; Springer: Cham, Switzerland, 2019; pp. 281–295.
161. Kumar, R.; Moche, M.; Winblad, B.; Pavlov, P.F. Combined x-ray crystallography and computational modeling approach to investigate the Hsp90 C-terminal peptide binding to FKBP51. *Sci. Rep.* **2017**, *7*, 14288. [[CrossRef](#)]
162. Bernadotte, A.; Kumar, R.; Winblad, B.; Pavlov, P.F. In silico identification and biochemical characterization of the human dicarboxylate clamp TPR protein interaction network. *FEBS Open Bio.* **2018**, *8*, 1830–1843. [[CrossRef](#)] [[PubMed](#)]
163. Pettersen, E.F.; Goddard, T.D.; Huang, C.C.; Couch, G.S.; Greenblatt, D.M.; Meng, E.C.; Ferrin, T.E. UCSF Chimera—a visualization system for exploratory research and analysis. *J. Comput. Chem.* **2004**, *25*, 1605–1612. [[CrossRef](#)]
164. Kolos, J.M.; Voll, A.M.; Bauder, M.; Hausch, F. FKBP Ligands-Where We Are and Where to Go? *Front. Pharmacol.* **2018**, *9*, 1425. [[CrossRef](#)] [[PubMed](#)]
165. Wang, L.; Bergkvist, L.; Kumar, R.; Winblad, B.; Pavlov, P.F. Studies of Chaperone-Cochaperone Interactions using Homogenous Bead-Based Assay. *J. Vis. Exp. JoVE* **2021**. [[CrossRef](#)] [[PubMed](#)]