

Small Interfering RNA-Mediated Suppression of Fas Modulate Apoptosis and Proliferation in Rat Intervertebral Disc Cells

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Study Design: *In vitro* cell culture model.

Purpose: To investigate the effect of small interfering RNA (siRNA) on Fas expression, apoptosis, and proliferation in serum-deprived rat disc cells.

Overview of Literature: Synthetic siRNA can trigger an RNA interference (RNAi) response in mammalian cells and precipitate the inhibition of specific gene expression. However, the potential utility of siRNA technology in downregulation of specific genes associated with disc cell apoptosis remains unclear.

Methods: Rat disc cells were isolated and cultured in the presence of either 10% fetal bovine serum (FBS) (normal control) or 0% FBS (serum deprivation to induce apoptosis) for 48 hours. Fas expression, apoptosis, and proliferation were determined. Additionally, siRNA oligonucleotides against Fas (Fas siRNA) were transfected into rat disc cells to suppress Fas expression. Changes in Fas expression were assessed by reverse transcription-polymerase chain reaction and semiquantitatively analyzed using densitometry. The effect of Fas siRNA on apoptosis and proliferation of rat disc cells were also determined. Negative siRNA and transfection agent alone (Mock) were used as controls.

Results: Serum deprivation increased apoptosis by 40.3% ($p < 0.001$), decreased proliferation by 45.3% ($p < 0.001$), and upregulated Fas expression. Additionally, Fas siRNA suppressed Fas expression in serum-deprived cultures, with 68.5% reduction at the mRNA level compared to the control cultures ($p < 0.001$). Finally, Fas siRNA-mediated suppression of Fas expression significantly inhibited apoptosis by 9.3% and increased proliferation by 21% in serum-deprived cultures ($p < 0.05$ for both).

Conclusions: The observed dual positive effect of Fas siRNA might be a powerful therapeutic approach for disc degeneration by suppression of harmful gene expression.

Keywords: Disc cells; Fas; Apoptosis; Cell proliferation; Small interfering RNA

Introduction

RNA interference (RNAi) facilitates inhibition of specific genes by sequence-specific gene silencing using double-stranded RNA (dsRNA) [1,2]. This approach involves

post-transcriptional gene silencing via dsRNA-mediated inhibition of gene expression through degradation of a specific, targeted mRNA. Small interfering RNA (siRNA), a type of RNAi, is a hybrid consisting of a sense and an antisense strand homologous in sequence to the sup-

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pressed gene [3]. Therefore, synthetic siRNA can trigger an RNAi response in mammalian cells and induce inhibition of specific gene expression. The specificity and potency of synthetic siRNA facilitate elucidation of gene function and investigate novel approaches for the treatment of various diseases [4]. However, the potential utility of siRNA technology in downregulation of specific genes associated with disc cell apoptosis remains unclear.

Fas is a type-I 45-kDa transmembrane protein that belongs to the tumor necrosis factor family. A wide range of human cells express Fas on their plasma membrane [5,6]. After binding of its natural ligand (Fas ligand), Fas triggers apoptosis in susceptible cells via two major downstream pathways: caspase 9-mediated intrinsic pathway and caspase 8-mediated extrinsic pathway. Previous studies reported a higher rate of apoptosis due to Fas/Fas ligand-mediated caspase activation in degenerated or herniated discs [7-9]. In addition, the extent of Fas expression was correlated with the severity of disc degeneration [7]. Therefore, specific downregulation of Fas might represent a novel therapeutic strategy against disc degeneration due to inappropriate apoptosis of disc cells.

In the current study, we investigated the effect of siRNA-mediated suppression of Fas on apoptosis and proliferation in rat disc cells, by transfection of Fas-targeting siRNA into cells for 48 hours under conditions of serum deprivation.

Materials and Methods

The experimental methods of the current study were the same as described in the previous paper [10].

1. Intervertebral disc cell culture

The Animal Care and Use Committee of the author's institution approved all experiments. Lumbar intervertebral discs (L1-L6) were harvested from five 4-week-old male Sprague Dawley rats immediately after sacrifice. The discs were carefully dissected under a microscope to isolate nucleus pulposus tissues, which were cultured in Dulbecco's modified Eagle's medium (DMEM; Gibco BRL, Grand Island, NY, USA) supplemented with 10% fetal bovine serum (FBS; Hyclone, Ottawa, Ontario, Canada), 100 U/mL penicillin (Gibco BRL), and 100 mg/mL streptomycin (Gibco BRL) in a humidified 5% CO₂ incubator at 37°C for 12 hours. To isolate disc cells, nucleus pulposus tis-

sues in DMEM medium were digested with 0.2% pronase (Sigma Chemical, St. Louis, MO, USA) for 4 hours. After enzymatic digestion, the suspension was filtered through a 70- μ m mesh (Falcon, Franklin Lakes, NJ, USA). Filtered cells were then washed with DMEM and grown as primary cultures. After seven passages, rat disc cells were trypsinized, subcultured into six-well plates at 1×10^6 cells/well, and cultured in the presence of either 10% FBS (normal control) or 0% FBS (to induce apoptosis) for 48 hours.

2. Terminal deoxynucleotidyl transferase-mediated dUTP nick-end labeling assay

Apoptosis of disc cells was determined using a terminal deoxynucleotidyl transferase-mediated dUTP nick-end labeling (TUNEL) assay. Briefly, 15 μ L APOPercentage dye (Biocolor Life Science, Carrickfergus, UK) was added to culture media to stain cells for 30 minutes. After removal of the culture medium containing the dye, the cells were gently washed twice with 500 μ L/well phosphate-buffered saline (PBS) and transferred to an inverted microscope to capture images. Disc cells cultured for 48 hours were used as controls.

3. Flow cytometry

Apoptosis of disc cells was also determined by double staining with fluorescein isothiocyanate (FITC)-conjugated Annexin V and propidium iodide (PI) (PharMingen, San Diego, CA, USA) according to the manufacturer's instructions. Briefly, cells were washed with cold PBS and suspended in binding buffer (10 mM HEPES-NaOH pH 7.4, 140 mM NaCl, 2.5 mM CaCl₂) at 1×10^6 cells/well and stained with 5 μ L Annexin V-FITC and 5 μ L PI. Cells were analyzed using a FACScan flow cytometer (Becton Dickinson, San Jose, CA, USA) at 48 hours.

4. Cell proliferation assay

Proliferation of disc cells was determined by the colorimetric CellTiter 96 Aqueous One Solution cell proliferation assay (Promega, Madison, WI, USA). Briefly, disc cell cultures in media supplement with 10% FBS or 0% FBS were added to the wells of a 96-well plate, and the medium was allowed to equilibrate for 1 hour. Next, 20 μ L 3-(4,5-dimethylthiazol-2-yl)-5-(3-

carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium, inner salt (MTS), and phenazine methosulfate were added to each well. After incubation for 2 hours at 37°C in a humidified 5% CO₂ incubator, absorbance of the plate at 490 nm was recorded using a spectrophotometric plate reader. Data are presented as means±standard deviation from six replicates.

5. Western blotting

Fas protein levels in disc cells were determined by Western blot analysis according to the manufacturer's instructions (Santa Cruz Biotechnology, Paso Robles, CA, USA). Briefly, disc cells were washed with ice-cold PBS and lysed with protein lysis buffer containing 50 mM HEPES pH 7.5, 150 mM NaCl, 1.5 mM MgCl₂, 1 mM EGTA, 10% glycerol, 1% Triton X-100, and 1 μM phenylmethylsulfonyl fluoride. Cell lysates were centrifuged at 12,000 g for 15 minutes, and protein concentrations were measured using the bicinchoninic acid method (Thermo Fisher Scientific, Pittsburgh, PA, USA). A total of 50 μg protein per sample was electrophoresed on 10% sodium dodecyl sulfate-polyacrylamide gels and transferred onto nitrocellulose membranes. The membranes were incubated with a primary antibody against Fas (Santa Cruz Biotechnology), followed by incubation with a horseradish peroxidase-linked IgG secondary antibody (BioRad, Richmond, CA, USA). Immunoreactive bands were visualized using an enhanced chemiluminescence detection kit (Santa Cruz Biotechnology).

6. Transfection with siRNA against Fas

Disc cells were grown to reach 80%–85% confluency on the day of transfection. GeneBank accession no. for Fas was NM139194. The following siRNA constructs were used in this study: Fas siRNA sense, 5'-GAAAC-GAACUGCACCCGGAU-3'; negative siRNA sense, 5'-UAGCGACUAAACACAUCAA-3' (Dharmacon,

Thermo Fisher Scientific). Briefly, cells were transfected with siRNA using the DharmaFECT siRNA transfection reagent (Dharmacon) according to the manufacturer's instructions. Transfections were performed with serum-free α-modified Eagle's medium containing 200 nmol siRNA and 6 μL DharmaFECT. After 18 hours, the transfection medium was replaced with a complete medium, and the cells were collected at 48 hours for analysis of Fas silencing by reverse transcription polymerase chain reaction (RT-PCR).

7. Reverse transcription polymerase chain reaction

Total RNA from disc cells was extracted using TRIzol reagent (Invitrogen, Grand Island, NY, USA) according to the manufacturer's instructions. For cDNA synthesis, 2 μg total RNA was reverse transcribed in a reaction mixture containing 25 units of ribonuclease inhibitor, 15 units of reverse transcriptase, 500 ng Oligo (dT) primers, 3 mM MgCl₂, 0.5 mM dNTP, and 1× RT buffer (Promega). Synthesized cDNA was used as a template for polymerase chain reaction (PCR) amplification with GoTaq polymerase (Promega) and primers for Fas (Bioneer, Cheongwon, Korea). PCR was performed with the Mycycler thermal cycler (BioRad, Hercules, CA, USA) according to the manufacturer's instructions. Analysis of the PCR products in 1% agarose gels showed single-band amplification products of expected sizes. Primer sequences and experimental conditions of PCR are summarized in Tables 1 and 2. Blots were semiquantitatively analyzed using

Table 1. The primer sequences of polymerase chain reaction

Primer	Sequence	Size (bp)
Fas	5'-GATGTGGAAGTGGCAGAG-3'	487
	5'-GGCTGACTTTCTCCTGGT-3'	
GAPDH	5'-ATCATCTCCGCCCTTCTGC-3'	437
	5'-GCCTGCTTCACCACCTTCTT-3'	

GAPDH, glyceraldehyde-3-phosphate dehydrogenase.

Table 2. The experimental conditions of polymerase chain reaction

Primer	Experimental conditions			Cycle
	Denaturation	Annealing	Polymerization	
Fas	94°C 5 min	94°C 30 sec → 46.3°C 30 sec → 72°C 30 sec	72°C 5 min	35
GAPDH	94°C 5 min	94°C 30 sec → 56°C 30 sec → 72°C 30 sec	72°C 5 min	25

GAPDH, glyceraldehyde-3-phosphate dehydrogenase.

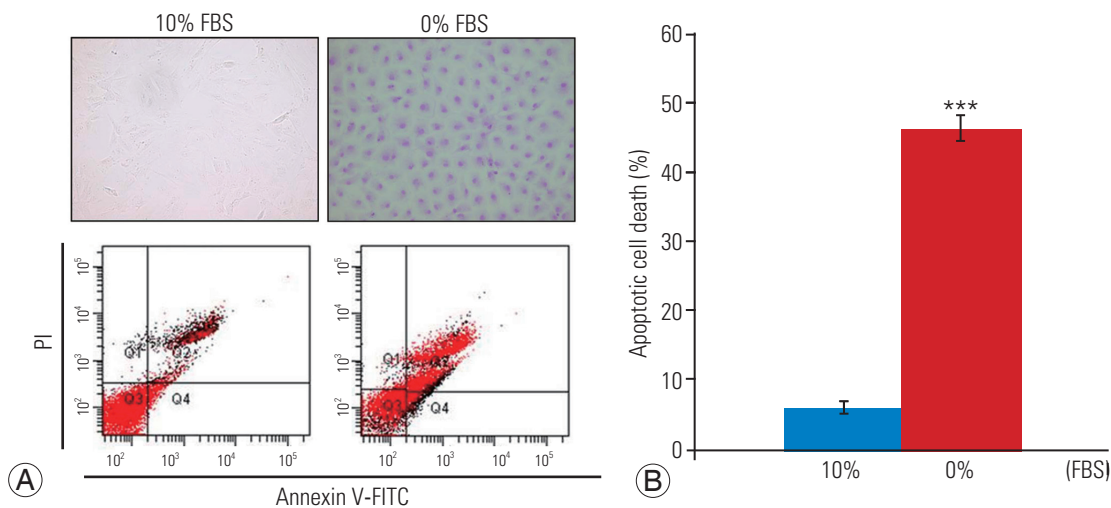


Fig. 1. (A) Terminal deoxynucleotidyl transferase-mediated dUTP nick-end labeling and flow cytometric analysis. PI, propidium iodide; V-FITC, Fluorescein isothiocyanate-conjugated Annexin V. (B) Disc cells incubated in 0% fetal bovine serum (FBS) for 48 hours displayed a significantly greater rate 40.3% for apoptotic cell death as compared to cells incubated in 10% FBS (5.9%±1.1% vs. 46.2%±3.5%). *** p <0.001.

GF670 imaging densitometer and the Molecular Analyst software (Bio-Rad), and results were expressed relative to the expression of *Gapdh* housekeeping gene. All experiments were repeated three times per sample, and average of three technical replicates was used as the final band density.

8. Effect of siRNA-mediated suppression of Fas on apoptosis and proliferation

TUNEL, flow cytometry, and MTS assay were used to investigate the effect of siRNA-mediated suppression of Fas expression on apoptosis and proliferation of disc cells using the methods described above. Statistical analyses were performed using Student's *t*-test. A p -values of <0.05 were considered to indicate statistical significance.

Results

1. Serum deprivation induces Fas upregulation and apoptosis and inhibits proliferation of disc cells

TUNEL assay and flow cytometry results are presented in Fig. 1A. There was a significant increase in apoptotic death in disc cells incubated in a serum-deprived medium than in those incubated in a medium containing 10% FBS for 48 hours (46.2%±3.5% vs. 5.9%±1.1%, p <0.001) (Fig. 1B). Similarly, the MTS assay revealed that the proliferation rate of disc cells cultured in the serum-deprived medium

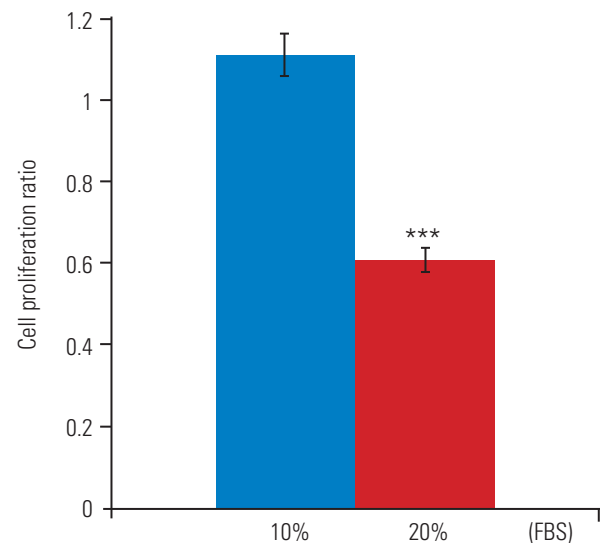


Fig. 2. MTS proliferation assay showing 45.3% reduction in proliferation of disc cells incubated in 0% fetal bovine serum (FBS) compared to those incubated in 10% FBS (0.608±0.12 vs. 1.111±0.24). MTS, 3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium. *** p <0.001.

was significantly lower than that of disc cells cultured in 10% FBS after 48 hours (0.608±0.12 vs. 1.111±0.24, p <0.001) (Fig. 2). Furthermore, Western blot analysis demonstrated that Fas expression was upregulated in serum-deprived disc cells compared to those incubated under normal serum conditions (Fig. 3). These results strongly suggested that the upregulation of Fas expression was responsible for the increased apoptosis and decreased proliferation of disc cells induced by serum deprivation.

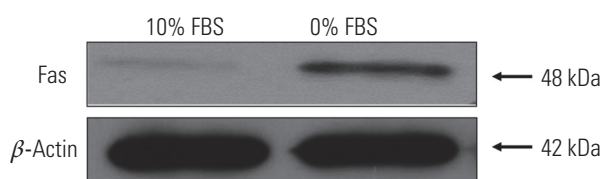


Fig. 3. Western blot analysis demonstrating that Fas expression was upregulated in disc cells incubated in 0% fetal bovine serum (FBS) compared to those cultured in 10% FBS.

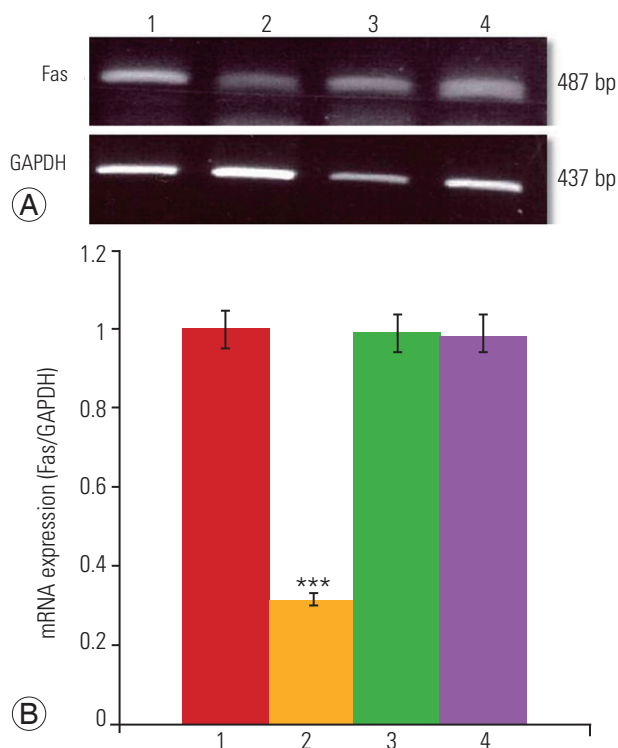


Fig. 4. (A) Analysis of *Fas* levels by reverse transcription-polymerase chain reaction. *Fas* siRNA significantly suppressed *Fas* mRNA levels in disc cells cultured in 0% fetal bovine serum (FBS) compared to those transfected with negative siRNA or the transfection agent alone (Mock). *Fas* mRNA was reduced by 68.5% with *Fas* siRNA ($p < 0.001$). (B) Note that *Fas* mRNA levels did not change in negative siRNA- or Mock-transfected cultures under conditions of serum deprivation. siRNA, small interfering RNA; 1, 0% FBS; 2, 0% FBS+Fas siRNA; 3, 0% FBS+negative siRNA; 4, 0% FBS+Mock. GAPDH, glyceraldehyde-3-phosphate dehydrogenase. *** $p < 0.001$.

2. siRNA-mediated suppression of Fas in disc cells

The results of RT-PCR analysis are presented in Fig. 4. As shown in Fig. 4A, *Fas* siRNA significantly suppressed *Fas* mRNA levels in disc cells cultured in 0% FBS compared to those transfected with negative siRNA or the transfection reagent alone (Mock). Quantification showed that

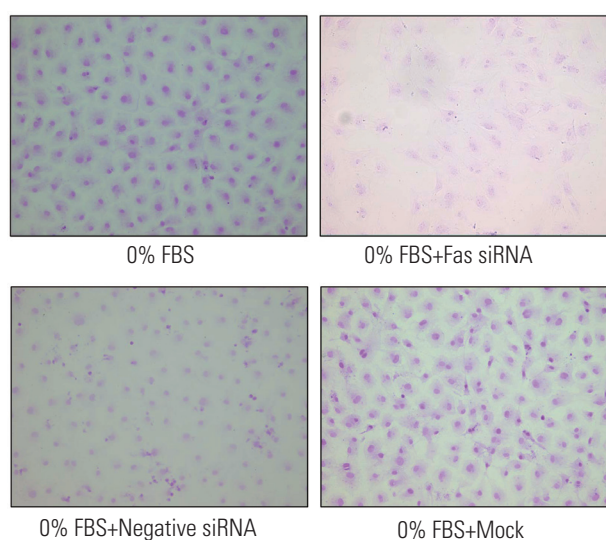


Fig. 5. Terminal deoxynucleotidyl transferase-mediated dUTP nick-end labeling assay (x400) demonstrating significantly decreased apoptosis of *Fas* small interfering RNA (siRNA)-transfected disc cells compared to negative siRNA- and Mock-transfected disc cells in conditions of serum deprivation. FBS, fetal bovine serum.

there was a 68.5% reduction in *Fas* mRNA in serum-deprived *Fas* siRNA-transfected disc cells ($p < 0.001$) (Fig. 4B). Conversely, neither negative siRNA nor Mock led to a reduction in *Fas* mRNA levels in cells cultured under conditions of serum deprivation.

3. Effect of siRNA-mediated Fas suppression on apoptosis and proliferation of disc cells

The TUNEL assay demonstrated that under conditions of serum deprivation, *Fas* siRNA significantly decreased apoptotic death of disc cells compared to those transfected with negative siRNA or Mock-transfected cells (Fig. 5). As presented in Fig. 6, flow cytometry analysis revealed that *Fas* siRNA significantly inhibited apoptosis by 9.3% in cells cultured in 0% FBS ($36.9\% \pm 2.7\%$ vs. $46.2\% \pm 3.5\%$, $p < 0.05$). In contrast, neither negative siRNA nor Mock-inhibited serum deprivation-induced apoptosis of disc cells. Finally, *Fas* siRNA led to a significant increase of 21% in proliferation in cells cultured in 0% FBS (0.736 ± 0.17 vs. 0.608 ± 0.12 , $p < 0.05$), whereas increased proliferation was not observed in serum-deprived cultures transfected with Mock-transfected or negative siRNA (Fig. 7).

Discussion

Selective silencing of endogenous genes by siRNA is uti-

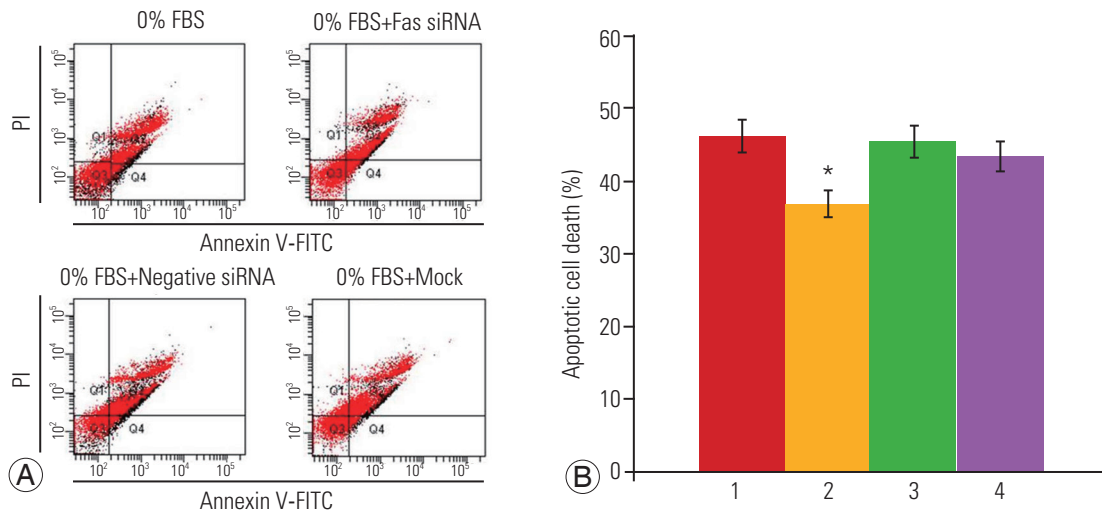


Fig. 6. (A) Flow cytometric analysis. Fas siRNA reduced apoptosis by 9.3% in disc cells cultured in 0% fetal bovine serum (FBS) ($36.9\% \pm 2.7\%$ vs. $46.2\% \pm 3.5\%$). PI, propidium iodide; V-FITC, Fluorescein isothiocyanate-conjugated Annexin V. **(B)** Note that the rate of apoptotic death of cell cultures in 0% FBS was not inhibited in negative siRNA- or Mock-transfected cultures. siRNA, small interfering RNA; siRNA, small interfering RNA; 1, 0% FBS; 2, 0% FBS+Fas siRNA; 3, 0% FBS+negative siRNA; 4, 0% FBS+Mock. * $p < 0.05$.

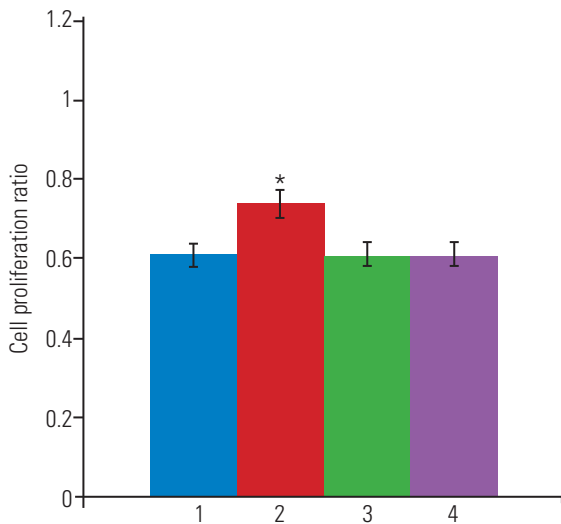


Fig. 7. MTS proliferation assay showing that Fas siRNA significantly increased proliferation by 21% in disc cells cultured in 0% fetal bovine serum (FBS) (0.736 ± 0.17 vs. 0.608 ± 0.12). Note that the rate of proliferation of cells cultured in 0% FBS was not increased in negative siRNA- or Mock-transfected cells. siRNA, small interfering RNA; 1, 0% FBS; 2, 0% FBS+Fas siRNA; 3, 0% FBS+negative siRNA; 4, 0% FBS+Mock; MTS, 3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium. * $p < 0.05$.

lized extensively [1-4]. Among numerous advantages of this technology are the relative ease of synthetic siRNA construction and application. Moreover, siRNA is efficient: a single dose of siRNA can sustain RNAi sufficiently long to allow recovery of cellular regulatory systems [3,4]. In the current study, we demonstrated that serum deprivation

increased apoptosis and decreased proliferation of disc cells, with upregulation of Fas. These results strongly suggested that Fas upregulation was responsible for the increased apoptosis and decreased proliferation of disc cells, resulting in disc degeneration. Therefore, we believe that specific downregulation of Fas by siRNA is a potential therapeutic approach to disc degeneration due to inappropriate apoptosis and proliferation of disc cells. To that end, we utilized siRNA oligonucleotides targeting Fas to transfect into disc cells for 48 hours under conditions of serum deprivation. Fas siRNA inhibited Fas expression in disc cells by approximately 68.5% at the mRNA level. We believe that the inhibitory effect of siRNA on Fas was more rapid and effective than expected, as complete serum deprivation inhibited cell survival. Therefore, our results suggested that siRNA silencing of Fas should facilitate the investigation of apoptotic pathways in disc cells in models of disc degeneration. According to the literature review, this is the first study to demonstrate siRNA-mediated suppression of Fas expression in disc cells.

In studies to characterize biologic repair of degenerated discs, several anti-apoptotic agents were shown to attenuate or prevent apoptosis of disc cells under various experimental conditions [11-14]. Growth factors such as platelet-derived growth factor, insulin-like growth factor-1, and nerve growth factor exert prosurvival, anti-apoptotic effects on disc cells [15,16]. However, the range of anti-apoptotic effects of growth factors varies from 0.5%

to 5% depending on the experimental conditions and dosages. Although cocktail therapy with growth factors was shown to induce the anti-apoptotic effect by 9% [16], the optimal dosage of individual growth factors for reducing apoptosis of disc cells is at present unclear. Moreover, the specific mechanisms and/or downstream pathways related to the anti-apoptotic effect of each growth factor should be investigated.

Apoptosis is mediated by the activation of caspases. Caspases act as either initiators (caspase 8 or caspase 9) or a common executioner (caspase 3) of apoptosis. Therefore, an alternative strategy involves interference with caspase activation. Previous studies reported that caspase inhibitors attenuated apoptosis of disc cells [17,18]. However, caspase inhibitors block apoptosis after its initiation or at a later stage. Therefore, targeting caspase activation may not be effective, as activation of early apoptotic signals might have detrimental effects on disc cell metabolism and activity. Moreover, even in the same cell type, caspase activation differs depending on the apoptotic stimulus. These limitations provide further support for the utilization of siRNA technology to target early stages of apoptosis and to act prior to caspase activation.

Finally, we investigated the effect of siRNA-mediated Fas suppression on the apoptosis and proliferation of disc cells under conditions of serum deprivation. The efficacy of therapeutic apoptosis inhibition by Fas siRNA (approximately 10%) was superior to that reported for nerve growth factor and insulin-like growth factor 1 (2% and 5%, respectively) [15]. In addition, siRNA-mediated suppression of Fas expression significantly increased cell proliferation by approximately 21%. These results suggested that downregulation of Fas expression during the initiation stage of apoptosis can attenuate or delay the onset of apoptosis and induce proliferation, leading to enhanced viability of disc cells. Therefore, we believe that this dual positive effect of Fas siRNA is a powerful therapeutic approach for disc degeneration.

There were several limitations to the current study. Percentage of cell death was used as the primary outcome measure; however, assessment of cell metabolism and activity as well as the ability to produce extracellular matrix remain critical measures. For biologic repair of degenerated discs, survival and proliferation of disc cells must be followed by a corresponding increase in extracellular matrix synthesis. Disc cell survival and proliferation without positive anabolic function does not truly lead to the re-

generation of a degenerated disc. Therefore, extracellular matrix synthesis should be investigated to maximize the efficacy of therapeutic siRNA targeting Fas in disc cells. Finally, while Fas siRNA led to more than 50% reduction in Fas expression, comparable changes in apoptosis and proliferation were not observed. Therefore, the efficacy of Fas siRNA on disc cell viability should be enhanced for robust regeneration of degenerated discs.

Conclusions

The observed dual positive effect of Fas siRNA might be a powerful therapeutic approach for disc degeneration by suppression of harmful gene expression. Furthermore, these findings suggest that siRNA can be used in gene function studies on disc degeneration.

Conflict of Interest

No potential conflict of interest relevant to this article was reported.

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