



Research article

Sensitive sandwich-type electrochemical immunosensing of p53 protein based on $Ti_3C_2T_x$ MXene nanoribbons and ferrocene/gold

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ARTICLE INFO

Keywords:

p53 protein
Tumor
Electrochemical immunosensor
MXene
 $Ti_3C_2T_x$
Ferrocene

ABSTRACT

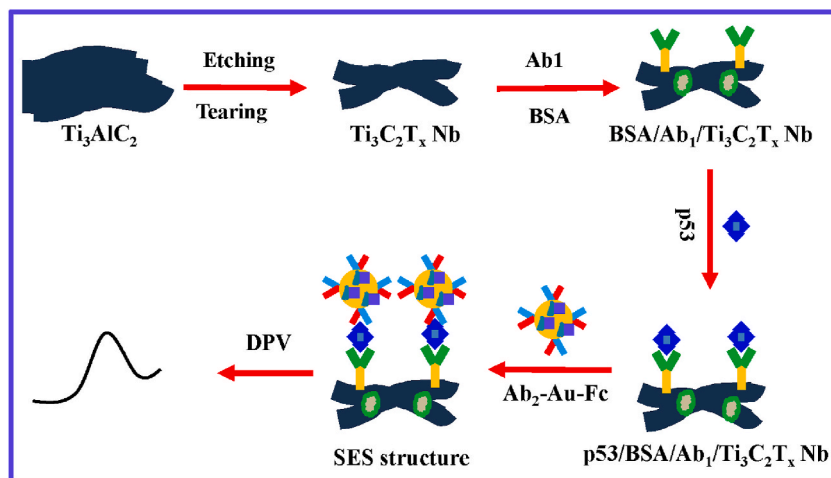
Since the p53 protein is an important promising biomarker of lung tumor and colorectal tumor, it is very essential to design a highly effective mean to monitor the degree of p53 for the early clinical analysis/therapy of the related tumors. In this work, a sandwich-type electrochemical immunosensing (SES) platform is proposed for the first time to detect p53 via synthesizing $Ti_3C_2T_x$ MXene nanoribbons ($Ti_3C_2T_x$ Nb) and ferrocene/gold nanoparticles (Fc/Au) respectively as the sensing substrate and signal-amplifier. The superior electrical property and large surface area of $Ti_3C_2T_x$ Nb are beneficial to assemble the initial p53-antibodies (Ab_1), while the synthesized Fc/Au is devoted to assemble the secondary p53 antibodies (Ab_2) and gives a magnified signal. By adopting the Fc molecules as the probes, the experiments reveal the response current of Fc resulted from the SES structure increases along with the p53 increase from 1.0 to 200.0 $pg\ mL^{-1}$. A considerable low detection limit (1.0 $pg\ mL^{-1}$) is achieved after optimizing several key conditions, it is thus confirmed the as-proposed SES mean exhibits significant application in the detection of p53 protein and other targets.

1. Introduction

The malignant tumors are a kind of diseases with extremely high mortality, generally the early monitor towards the related biomarkers has important significance to offer vital information for the timely and efficient therapy [1–3]. As a reported tumor inhibitor, the p53 protein plays key roles to prevent the gene mutations and maintain the genome integrity [4,5], and generally the physiological degree of p53 in normal human is considerable low ($<10\ pg\ mL^{-1}$) [6–8]. In such case, the accurate quantification of p53 is extremely useful for the early clinical analysis and therapy of the malignant tumors. Traditional means for the detection of p53 mainly include enzyme-linked immunosorbent assay, chromatography-mass spectrometry and western blotting, immunohistochemistry [9–12], but which were restricted resulted from their cost, operation and analysis times. Sequentially significant efforts have been motivated to design low-cost, simple and rapid methodologies. Recently, the electrochemical immunoassays received some attentions since they exhibit some unique superiorities such as simple operation, inexpensive and time-saving [6,13]. In addition, it has been reported, attributed to the advantages in the feature of signal-amplification, the sandwich-type electrochemical immunosensing (SES) means have aroused considerable attentions to detect various analytes in the past years [14–16]. Therefore, it is very promising to design a sensitive and effective SES platform for p53 detection.

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Scheme 1. Illustrations for constructing the as-designed SES sensing platform of p53.

As one of the two key factors in constructing SES platform, designing or selecting appropriate nanomaterial as the substrate of sensor is very important [17–19]. As a type of emerging-representative 2D materials with high electrical conductivity and large surface area recently, the compounds of transition metal carbides/carbonitrides (denoted as MXenes) have been utilized widely in many research fields such as energy storage and conversion, sensors and biomedicine [20–24]. For instance, by growing CoFe-PBA on ferrocene functionalized $\text{Ti}_3\text{C}_2\text{T}_x$ MXene, Wang et al. [25] developed a sensitive electrochemical sensor of xanthine. In this sensor, the introduction of $\text{Ti}_3\text{C}_2\text{T}_x$ MXene is dedicated to accommodate ferrocene and give higher conductivity. In general, MXenes are achieved by exfoliating the “A” element in the MAX phase, where “M” is the transition metal and “A” is the element in the IIIA/IVA group, while “X” refers to C or/and N [26–28]. In addition, the MXenes-based nanomaterials can offer good biocompatibility and be easy to functionalizing. These superiorities of MXenes devoted themselves to be a desirable candidate as the substrate to construct a SES platform for p53. In addition, there is no doubt the construction of signal amplifier is also considerable important for the sensing performances of SES platform. Nanogold (Au), as an universally accessible precious metal exhibiting superior conductivity and biocompatibility, is very easy to be functionalized [29,30]. Meanwhile, the ferrocene (Fc) molecule is a common and acceptable signal-probe since it can exhibit high electroactivity [31,32].

Inspired by the above insights, in this work, by synthesizing $\text{Ti}_3\text{C}_2\text{T}_x$ MXene (a typical MXene material) nanoribbons ($\text{Ti}_3\text{C}_2\text{T}_x \text{Nb}$) as the substrate to assemble Ab_1 of p53 ($\text{Ab}_1\text{-Ti}_3\text{C}_2\text{T}_x \text{Nb}$), and functionalizing Fc coupled with Ab_2 of p53 to the Au surface as the specific signal-amplifier ($\text{Ab}_2\text{-Au-Fc}$), we proposed a novel SES mean to detect the degree of p53 (Scheme 1). In the existence of p53 protein, a SES structure consisted of $\text{Ab}_2\text{-Au-Fc/p53/Ab}_1\text{-Ti}_3\text{C}_2\text{T}_x \text{Nb}$ would be formed and the relative response signal of the Fc probe is proportional and sensitive to the p53 level, hence achieving a highly sensitive and effective monitoring for the p53 protein. This works provided a reliable and sensitive mean for p53 detection, it's expected which can offer significant real application.

2. Experimental section

2.1. Reagents and equipment

The Ti_3AlC_2 raw material was purchased from Forsman Technology Company (China). HF ($\geq 40.0\%$), KOH , $\text{HAuCl}_4 \cdot 3\text{H}_2\text{O}$, NaBH_4 , Fc , NaOH , 4-aminobenzoic acid (PABA), 4-aminobenzoic acid (ABZ) 1-Ethyl-3-[3-dimethylaminopropyl]carbodiimide hydrochloride (EDC), NaNO_2 and N-hydroxysuccinimide (NHS) were obtained from Aladdin Biotechnology Co., Ltd (China). Bovine serum albumin (BSA), carcinoembryonic antigen (CEA), alpha fetoprotein (AFP), IgG antigen and anti-p53 antibodies were offered by Abcam Co., Ltd. The phosphate buffer solution (PBS) were prepared with KH_2PO_4 (0.1 M), K_2HPO_4 (0.1 M) and KCl (0.1 M). Electrochemical CHI 660E Workstation was utilized for all the electrochemical measurements.

2.2. Synthesis of $\text{Ti}_3\text{C}_2\text{T}_x \text{Nb}$

The $\text{Ti}_3\text{C}_2\text{T}_x$ MXene nanosheets ($\text{Ti}_3\text{C}_2\text{T}_x \text{NS}$) were firstly synthesized via etching the Al layer in the Ti_3AlC_2 powder according to the reported method [33]. Briefly, under stirring at $\sim 35^\circ\text{C}$, 05 g raw material of Ti_3AlC_2 was slowly added into a HF solution of 10.0 mL. After holding for 20 h, the $\text{Ti}_3\text{C}_2\text{T}_x \text{NS}$ can be obtained via washing and drying.

Next, $\text{Ti}_3\text{C}_2\text{T}_x \text{Nb}$ were synthesized via tearing the above obtained $\text{Ti}_3\text{C}_2\text{T}_x \text{NS}$ as follows: the 0.1 g $\text{Ti}_3\text{C}_2\text{T}_x \text{NS}$ were added into a 3.0 M KOH solution (25.0 mL) under continuous stirring under for 2 days at room temperature, thus the designed $\text{Ti}_3\text{C}_2\text{T}_x \text{Nb}$ can be collected after washing and drying. For further functionalizing $\text{Ti}_3\text{C}_2\text{T}_x \text{Nb}$ with carboxyl dedicated to immobilize Ab_1 , 0.08 g ABZ and 0.07g NaOH were firstly added to 25.0 mL watery. Next, 3.0 mL HCl solution was added quickly to the solution and maintained for

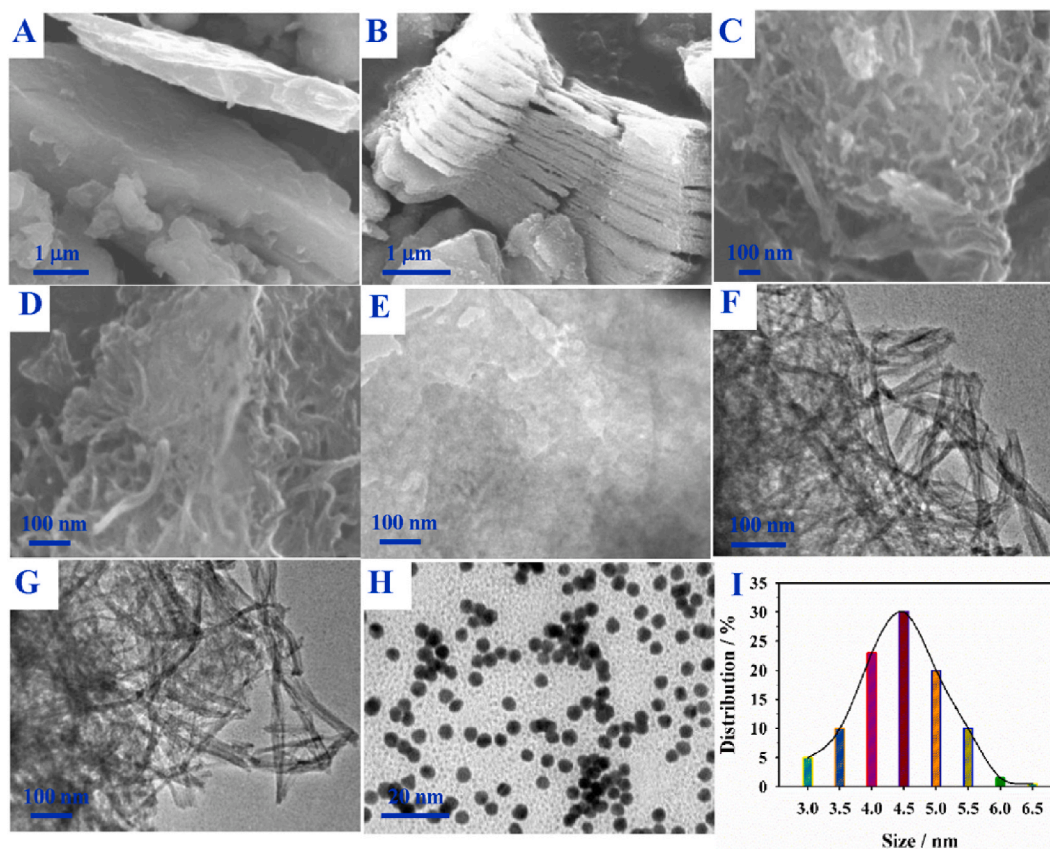


Fig. 1. The SEM images of (A) Ti₃AlC₂, (B) Ti₃C₂T_x NS, (C) Ti₃C₂T_x Nb and (D) carboxylated Ti₃C₂T_x Nb; the TEM images of (E) Ti₃C₂T_x NS, (F) Ti₃C₂T_x Nb, (G) carboxylated Ti₃C₂T_x Nb and (H) Au particles; (I) the particle size distribution of Au.

~30 min under stirring. In final, the formed mixture were centrifuged and dried to achieve Ti₃C₂T_x Nb product.

2.3. Synthesis for Au and immobilizing Ab₂ coupled with Fc

For preparing Ab₂-Au-Fc, the tiny Au nanoparticles were firstly synthesized: 0.1 mL sodium citrate solution (0.2 M) was added dropwise to a 3.0 mL HAuCl₄ solution and heated at about 60 °C for 2 h, thus obtaining the Au nanoparticles. Then, 20.0 μL Ab₂ solution of 60.0 μg mL⁻¹ and 20.0 μL Fc solution of 0.4 mM were added into the obtained Au solution (1.0 mL), and this mixture was incubated for 1 h at room temperature to obtain Ab₂-Au-Fc signaling amplifier.

2.4. Fabrication of the SES structure

Firstly, a definite of Ti₃C₂T_x Nb suspension of (1.0 mg mL⁻¹) was dropped to the surface of SPCE electrode and a 1:1 mixture solution containing EDC/NHS (0.1 M/0.2 M) were used to activate carboxyls. In sequentially, 8.0 μL solution of Ab₁ (10.0 μg mL⁻¹) was coated and immobilized to the activated surface of Ti₃C₂T_x Nb/SPCE as Ab₁/Ti₃C₂T_x Nb/SPCE. Then, 3.0 μL BSA (100.0 mg mL⁻¹) was utilized to restrict the non-specific immobilization. For sensing p53, the fabricated BSA/Ab₁/Ti₃C₂T_x Nb/SPCE was interacted with p53 at various degrees, and a SES structure would be formed by combining p53/BSA/Ab₁/Ti₃C₂T_x Nb/SPCE with Ab₂-Au-Fc, which was then monitored via differential pulse voltammetry (DPV).

3. Results and discussion

3.1. Characterization of the synthesized Ti₃C₂T_x Nb and Au

Fig. 1 displayed the SEM and TEM images of the as-synthesized Ti₃C₂T_x Nb and related materials. It was noted the original Ti₃AlC₂ materials show smooth surface with aligned densely layered-structures (Fig. 1A). After HF etching, the synthesized Ti₃C₂T_x NS (Fig. 1B and E) layered-structures were very evident and displayed packed loosely accordion-type morphology compared to the original Ti₃AlC₂ sample. Interestingly, via tearing Ti₃C₂T_x NS, Ti₃C₂T_x Nb (Fig. 1C and F) could be well evidenced with minor diameter (~28

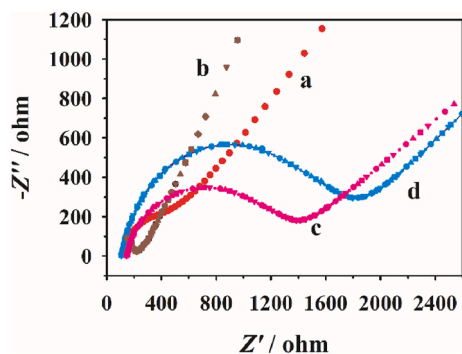


Fig. 2. The EIS plots of SPCE (a), $\text{Ti}_3\text{C}_2\text{T}_x$ Nb/SPCE (b), BSA/Ab₁/ $\text{Ti}_3\text{C}_2\text{T}_x$ Nb/SPCE (c), and Ab₂-Au-Fc/p53/BSA/Ab₁/ $\text{Ti}_3\text{C}_2\text{T}_x$ Nb/SPCE (d).

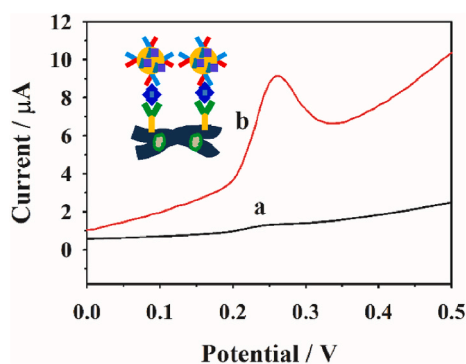


Fig. 3. DPV response of (a) BSA/Ab₁/ $\text{Ti}_3\text{C}_2\text{T}_x$ Nb/SPCE and (b) p53/BSA/Ab₁/ $\text{Ti}_3\text{C}_2\text{T}_x$ Nb/SPCE after their incubation in Ab₂-Au-Fc solution.

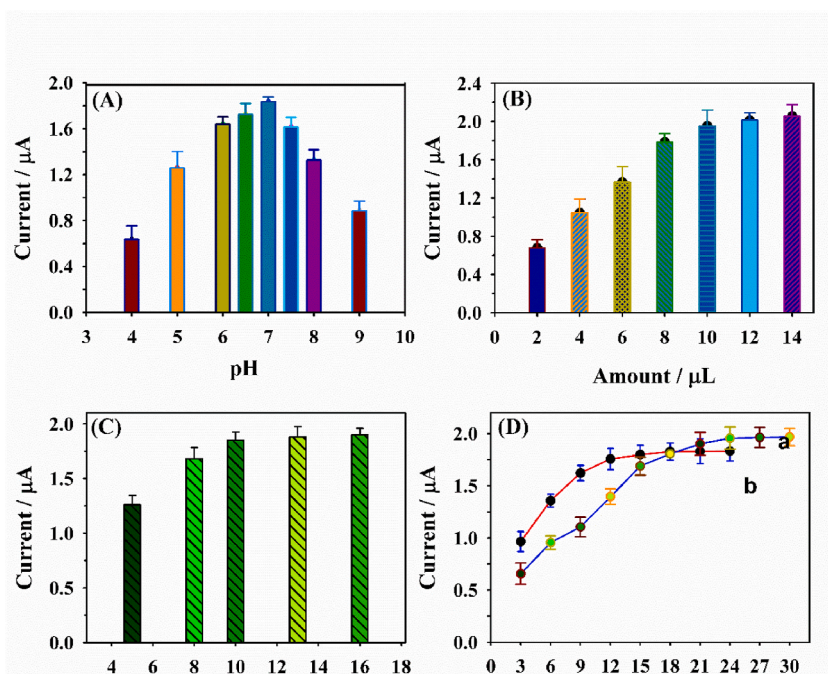


Fig. 4. Influences of the (A) pH value of PBS; (B) amount of $\text{Ti}_3\text{C}_2\text{T}_x$ Nb; (C) Ab₁ concentration; (D) interaction time of p53 with BSA/Ab₁/ $\text{Ti}_3\text{C}_2\text{T}_x$ Nb (a) and p53/BSA/Ab₁/ $\text{Ti}_3\text{C}_2\text{T}_x$ Nb with Ab₂-Au-Fc (b).

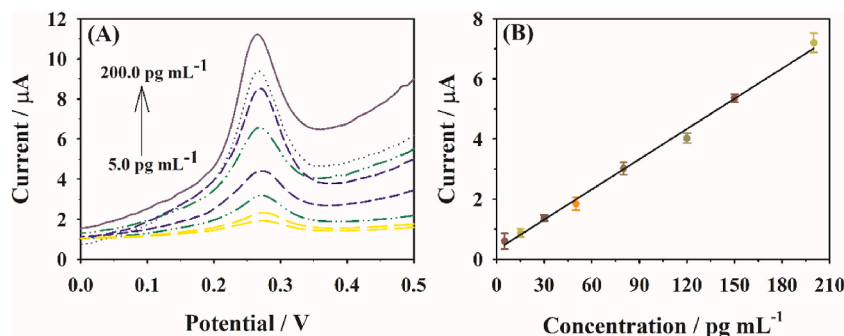


Fig. 5. (A) The DPV responses of p53 with various concentrations (5.0, 15.0, 30.0, 50.0, 80.0, 120.0, 150.0 and 200.0 pg mL^{-1}) based on the developed SES sensor; (B) the calibration plot of p53 detection.

Table 1

Comparisons of various immunosensing platform with the present method for p53 protein.

Electrode	Method	Linearity (pg mL^{-1})	LOD (pg mL^{-1})	Ref.
AuNPs	LSV	87000.0–210000.0	2100.0	[5]
PEDOT:PSS/AuNPs	DPV	1000.0–120000.0	90.0	[4]
Au/NiPc	DPV	0.1–500.0	–	[34]
SAM	LSV	4.3–4370.0	4.3	[35]
PEG	CV	10.0–10000.0	10.0	[36]
PTAA/Au/PMB	SWV	1000.0–100000.0	650.0	[37]
$\text{Ti}_3\text{C}_2\text{T}_x$ Nb	DPV	5.0–200.0	1.0	This work

* PEDOT:PSS (poly (3, 4-ethylenedioxythiophene):polystyrenesulfonate); NiPc: nickel phthalocyanine; SAM: Self-assembled monolayer; PEG: pencil graphite electrode; PTAA: poly(thiophene acetic acid); PMB: poly(thiophene acetic acid); LSV: linear sweep voltammograms; CV: cyclic voltammetry; SWV: square wave voltammetry.

nm). As is evident, the $\text{Ti}_3\text{C}_2\text{T}_x$ Nb with carboxyl groups have well-defined structures that were almost the same as displayed for $\text{Ti}_3\text{C}_2\text{T}_x$ Nb (Fig. 1D and G). This indicates that the carboxyl functionalization does not change the microstructure and morphology of the materials, thus which were further characterized by Fourier transform infrared (FT-IR) spectrum (Fig. S1). Compared with the pure $\text{Ti}_3\text{C}_2\text{T}_x$ Nb, the carboxylated $\text{Ti}_3\text{C}_2\text{T}_x$ Nb gave a strong and typical peak of the carboxyl at $\sim 1637 \text{ cm}^{-1}$.

Next, the used Au particles were characterized by TEM. As shown in Fig. 1H and I, the as-synthesized Au nanoparticles exhibited very uniform size, and the diameter is tiny to about 4.5 nm through statistical investigation, such small size is beneficial to immobilize Ab₂ and Fc molecules.

3.2. Electrochemical evaluation of the SES platform

The impedance spectroscopy (EIS) was firstly utilized to characterize the processes in fabricating SES structure, which was conducted in a $[\text{Fe}(\text{CN})_6]^{3-/4-}$ solution (5.0 mM) (Fig. 2). In general, the common biomolecules exhibit poor conductive and pose inhibiting effect to the electron channel, thus increasing the impedance values. As displayed in Fig. 2, the charge-transfer resistance (R_{CT}) of $\text{Ti}_3\text{C}_2\text{T}_x$ Nb/SPCE was smaller compared to the bare SPCE resulted from the superior conductivity of the $\text{Ti}_3\text{C}_2\text{T}_x$ MXene. When Ab₁, p53 and Ab₂-Au-Fc were assembled in successive to the surface of $\text{Ti}_3\text{C}_2\text{T}_x$ Nb/SPCE, the R_{CT} values at the fabricated BSA/Ab₁/ $\text{Ti}_3\text{C}_2\text{T}_x$ Nb/SPCE and Ab₂-Au-Fc/p53/BSA/Ab₁/ $\text{Ti}_3\text{C}_2\text{T}_x$ Nb/SPCE showed an increase in turn. These indicated that the fabricated SES layer was achieved successfully.

The feasibility about the as-proposed SES sensor was characterized by DPV technology as it can offer superior sensitivity. Fig. 3 displayed the DPV curves of BSA/Ab₁/ $\text{Ti}_3\text{C}_2\text{T}_x$ Nb/SPCE and p53/BSA/Ab₁/ $\text{Ti}_3\text{C}_2\text{T}_x$ Nb/SPCE after the incubation with Ab₂-Au-Fc. The achieved results revealed there is no DPV signal presented without p53 (300.0 pg mL^{-1}), that is BSA/Ab₁/ $\text{Ti}_3\text{C}_2\text{T}_x$ Nb/SPCE. Nevertheless, a strong DPV current of Fc could be noted at p53/BSA/Ab₁/ $\text{Ti}_3\text{C}_2\text{T}_x$ Nb/SPCE after incubating with Ab₂-Au-Fc to generate a SES structure (Ab₂-Au-Fc/p53/BSA/Ab₁/ $\text{Ti}_3\text{C}_2\text{T}_x$ Nb). The related reason was that the specific antibody-antigen recognition between p53-antibodies and p53 could induce the Ab₂-Au-Fc capture to p53/BSA/Ab₁/ $\text{Ti}_3\text{C}_2\text{T}_x$ Nb. These suggested that the as-proposed SES proposal for p53 was feasible.

In order to achieve the sensitive determination for p53, several key experiments need to be optimized. Firstly, there is no doubt that the acid-base property of sensing system is very important. Fig. 4A gave the effect of the different pH values of PBS to the DPV signal of Ab₂-Au-Fc/p53/BSA/Ab₁/ $\text{Ti}_3\text{C}_2\text{T}_x$ Nb/SPCE, it's noted the signals increased along with the pH value increase and the maximum was at 7.0. As for the influence from the dropped amount of $\text{Ti}_3\text{C}_2\text{T}_x$ Nb to the surface of electrode (Fig. 4B), the peak current increases with the amount increase. Whereas when exceeding 10.0 μL , the DPV signal showed little change, so 10.0 μL $\text{Ti}_3\text{C}_2\text{T}_x$ Nb dispersion was the amount that coated to the SPCE surface. Meanwhile, Fig. 4C exhibited the effect of Ab₁ concentration, the results revealed 10.0 μg

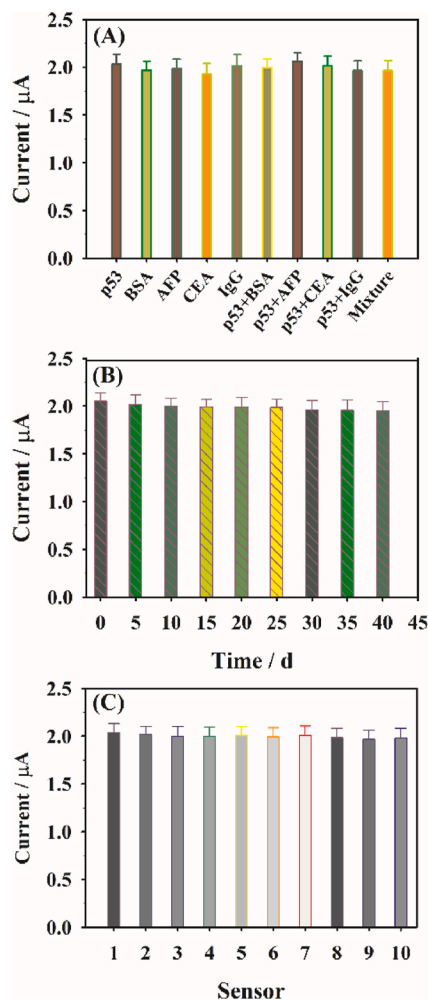


Fig. 6. (A) Selectivity, (B) reproducibility and (C) stability for p53 detection based on the constructed SES sensor.

mL^{-1} is enough, thus this concentration was used for the experiments. Fig. 4D displayed the effect about the interaction time between p53 and its antibodies. It can be found the desirable time between p53 and Ab_1 was 12 min, while that was 21 min between p53 and Ab_2 .

Finally, the detection capabilities of the as-fabricated SES mean for p53 were evaluated. As displayed in Fig. 5, the DPV signals gradually increased with the p53 level increased ranging from 5.0 to 200.0 pg mL^{-1} (Fig. 5A), and the linear equation can be written down as $\Delta I (\mu\text{A}) = 0.314 + 0.034C (\text{pg mL}^{-1})$ (Fig. 5B). Based on $S/N = 3$, the limit of detection (LOD) was 1.0 pg mL^{-1} . In order to further understand the as-designed SES mean, the comparisons between this work and other sensors for p53 detection were also performed. From Table 1, it was assured the detection performances of the SES mean were comparable to the previous immunosensors with lower LOD value. In addition, it should be pointed out that the p53 protein can be divided into wild type and mutant type. The wild-type p53 protein is extremely unstable, with a half-life of only a few minutes, while the mutant p53 proteins are more stable, have a longer half-life. Therefore, the p53 protein detected by this sensor is mutant p53 protein.

3.3. Reliability and real samples analysis

Firstly, the proposed SES mean's selectivity was confirmed after the addition of various interfering substances to the detection system. As displayed in Fig. 6A, it was observed there was little change observed for the signal of Fc after the addition of BSA, AFP, CEA and IgG (10.0 times of p53 level). The stability of the SES mean was also evaluated (Fig. 6B), and the signal responses of SES structure remained almost unchanged via storing the sensing platform for 40 days. Next, the reproducibility was investigated by measuring the signals of Fc at 10-independently-fabricated SES structures. As displayed in Fig. 6C, the RSD value between them was 3.68 %. These indicated the as-fabricated SES mean exhibited desirable reliability.

Finally, the standard addition technique was adopted to validate the as-proposed SES mean in real application. Various degrees of p53 protein were added in turn to the real human serum samples, and the response signals of the SES sensor towards these samples

were recorded. Then, the recoveries were calculated to reveal the capability in the real applications. From Table S1, the obtained recoveries were ranging from 94.7 % to 97.0 %, confirming the as-proposed SES sensing platform exhibited satisfactory real ability for p53 detection.

4. Conclusion

In summary, a novel SES sensing platform for p53 was designed in this work by preparing $\text{Ti}_3\text{C}_2\text{T}_x\text{Nb}$ as sensing platform and Au particles to immobilize Fc and Ab₂ as signal amplifier. Owing to the excellent properties (e.g., high electrical conductivity and large surface area) of $\text{Ti}_3\text{C}_2\text{T}_x\text{Nb}$ which are beneficial to immobilize Ab₁ and improve conductivity, and the signal amplification function of Ab₂-Au-Fc, the as-designed SES sensing platform displayed superior analytical performances under the optimized conditions: wider linearity (5.0–200.0 pg mL⁻¹) coupled with lower LOD (1.0 pg mL⁻¹). Additionally, the as-proposed SES mean displayed good stability, selectivity and reproducibility as well as real applications, suggesting this SES mean for p53 showed important potential application. Through adopting other appropriate antibodies instead of p53 antibodies, we expected the as-designed SES sensing model has a broad application in the detection for other analytes.

Data availability statement

Data will be made available on request.

CRediT authorship contribution statement

Song Lin: Writing – original draft, Visualization, Validation, Methodology, Investigation, Data curation. **Lixin Wen:** Writing – review & editing, Validation, Supervision, Software, Resources. **Hong Zhao:** Writing – review & editing, Validation, Resources, Data curation. **Donghua Huang:** Validation, Resources, Methodology. **Zuwei Yang:** Validation, Investigation. **Qinge Zou:** Data curation, Conceptualization. **Ling Jiang:** Writing – review & editing, Validation, Supervision, Project administration, Methodology, Investigation, Funding acquisition.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgment

This work was financially supported by Fujian Province Natural Science Foundation (No.: 2023J011759).

Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.heliyon.2024.e36910>.

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