


LETTER TO THE EDITOR

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IRF4 expression is low in Philadelphia negative myeloproliferative neoplasms and is associated with a worse prognosis

Cosimo Cumbo^{1†}, Francesco Tarantini^{1†}, Luisa Anelli¹, Antonella Zagaria¹, Immacolata Redavid¹, Crescenzo Francesco Minervini¹, Nicoletta Coccaro¹, Giuseppina Tota¹, Alessandra Ricco¹, Elisa Parciante¹, Maria Rosa Conserva¹, Giorgina Specchia², Pellegrino Musto¹ and Francesco Albano^{1*} 

Abstract

Interferon regulatory factor 4 (*IRF4*) is involved in the pathogenesis of various hematologic malignancies. Its expression has been related to the negative regulation of myeloid-derived suppressor cells (MDSCs) and the polarization of anti-inflammatory M2 macrophages, thereby altering immunosurveillance and inflammatory mechanisms. An abnormal inflammatory status in the bone marrow microenvironment of myeloproliferative neoplasms (MPNs) has recently been demonstrated; moreover, in chronic myeloid leukemia a downregulated expression of *IRF4* has been found. In this context, we evaluated the *IRF4* expression in 119 newly diagnosed consecutive Philadelphia negative MPNs (Ph-MPNs), showing a low expression among the MPNs phenotypes with a more significant decrease in primary myelofibrosis patients. Lower *IRF4* levels were associated with *JAK2* + and triple negatives cases carrying the worst prognosis. Furthermore, the *IRF4* levels were related to leukemic transformation and a shorter leukemia-free survival; moreover, the risk of myelofibrosis transformation in polycythemia vera and essential thrombocythemia patients was more frequent in cases with lower *IRF4* levels. Overall, our study demonstrates an *IRF4* dysregulated expression in MPNs patients and its association with a worse prognosis. Further studies could validate these data, to improve our knowledge of the MPNs pathogenesis and confirm the *IRF4* role as a new prognostic factor.

Keywords: *IRF4* expression, Philadelphia negative MPNs, Prognosis

To the Editor

Interferon regulatory factor 4 (*IRF4*) is a transcription factor with an established role in the pathogenesis of various hematologic malignancies [1]. *IRF4* expression has been related to the negative regulation of myeloid-derived suppressor cells (MDSCs), thereby altering immunosurveillance; moreover, its expression drives inflammation through the polarization of

anti-inflammatory M2 macrophages [2, 3]. In chronic myeloid leukemia a downregulated expression of *IRF4* has been found [4] and recent evidence supports its prognostic role in *JAK2V617F* mutated myeloproliferative neoplasms (MPNs) [5].

With these premises, we evaluated *IRF4* expression in 119 Philadelphia negative MPNs (Ph-MPNs) patients (Additional file 2: Table S1) to verify its role on clinical outcome (median follow-up: 61.5 months, range: 1–238). The quantification was calculated as the ratio between *IRF4* and *GUSB* number of copies (I/G) (Additional file 1).

The bone marrow (BM) *IRF4* median value was 0.11 I/G (min. 0.11- max. 0.12) and 0.04 I/G (min. 0.001

*Correspondence: francesco.albano@uniba.it

[†]Cosimo Cumbo and Francesco Tarantini contributed equally to this work

¹ Department of Emergency and Organ Transplantation (D.E.T.O.), Hematology and Stem Cell Transplantation Unit, University of Bari "Aldo Moro", P.Zza G. Cesare, 11, 70124 Bari, Italy

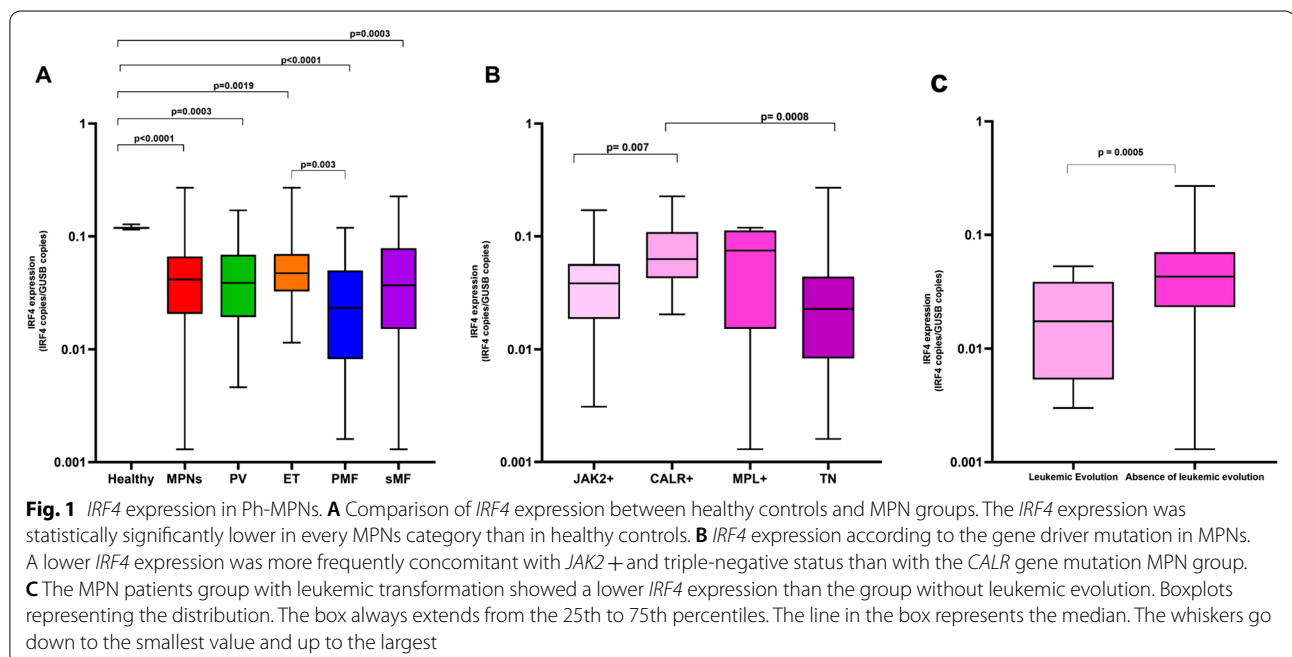
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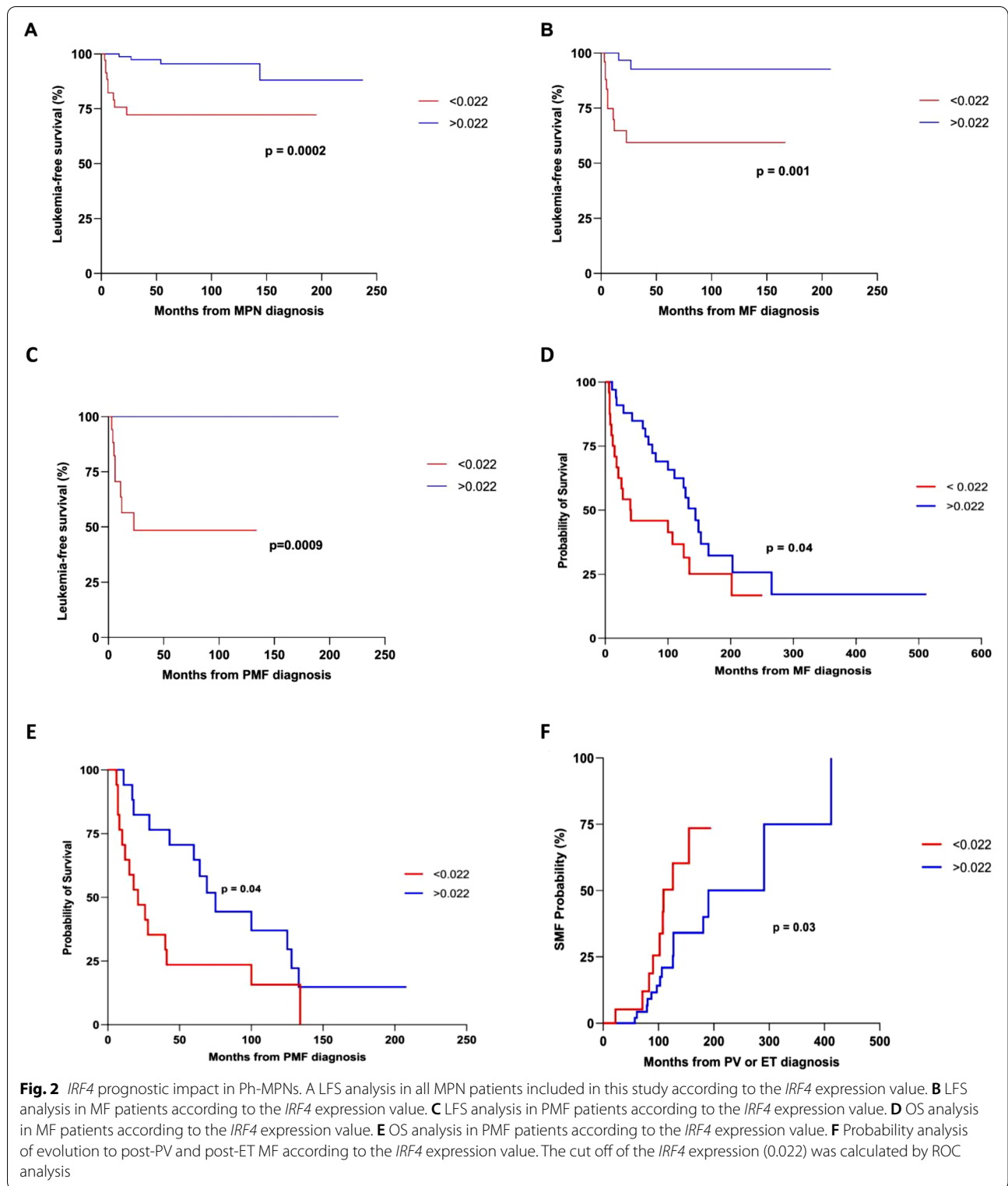


– max. 0.27) in the healthy and in the MPNs groups, respectively ($p < 0.0001$). Considering the *IRF4* median value for every MPN type, the difference compared to the healthy controls (HC) remained statistically significant (Fig. 1A). In particular, the primary myelofibrosis (PMF) patients showed a lower *IRF4* median value than that of the other groups compared to the HC. Among the MPNs, the PMF *IRF4* median value was lower than in the essential thrombocythemia (ET) group ($p = 0.003$). *CALR* mutated cases showed a higher *IRF4* expression than those with *JAK2* (0.06 vs 0.03, $p = 0.007$) or triple-negative (TN) (0.06 I/G vs 0.02 I/G, $p = 0.0008$) (Fig. 1B). *IRF4* expression was not associated with variables as sex, age, risk group [6]. Fourteen (11.7%) patients showed leukemic transformation (LT): 8 PMF, 4 secondary myelofibrosis (SMF), and 2 polycythemia vera (PV); they had a lower *IRF4* expression at diagnosis compared to the other MPN patients (0.01 I/G vs 0.04 I/G, $p = 0.0005$) (Fig. 1C). An optimal cutoff of the *IRF4* expression value best identifying the possibility of MPN leukemic transformation was defined by ROC analysis. The area under the curve was 0.79 (95% CI 0.71–0.86; $p < 0.0001$). Representative cutoff values for sensitivity and specificity were calculated and plotted on the curve. An optimal value of 0.022 I/G was obtained, with a sensitivity of 76.9% (95% CI 46.2–95.0) and a specificity of 76.1% (95% CI 66.9–84.0). This value distinguished MPN patients with a higher probability of LT; in fact, the group with an *IRF4* value < 0.022 I/G had shorter leukemia-free survival

(LFS) (Fig. 2A). The LFS analysis was also considered only for the myelofibrosis (MF) group; patients with an *IRF4* value < 0.022 I/G showed a shorter LFS ($p = 0.001$, Fig. 2B). In another LFS analysis, this difference was confirmed when considering only the PMF group: the median LFS for PMF patients with *IRF4* < 0.022 I/G was 23 months, whereas LFS was not reached for the > 0.022 I/G group ($p = 0.0009$) (Fig. 2C). Overall survival (OS) analysis in the MF group showed that patients with *IRF4* > 0.002 I/G had longer median survival than those in the < 0.022 I/G group (143.9 mo. versus 40.5 mo., $p = 0.04$) (Fig. 2D). Also, in the PMF group, the *IRF4* value > 0.022 I/G was associated with a longer OS (75 mo versus 21 mo, $p = 0.04$) (Fig. 2E). Moreover, PV and ET patients with an *IRF4* value < 0.022 I/G showed a shorter time to MF transformation (109 mo. versus 190 mo., $p = 0.03$) (Fig. 2F).

Among the 14 patients with LT, 9 (64.2%), 7 PMF and 2 SMF, were analyzed in next-generation sequencing [7] to detect the high molecular risk (HMR) mutations (Additional file 3: Table S2). In 5 PMF patients, HMR mutations were found; moreover, all 9 patients showed at least one additional genetic lesion besides the gene driver mutation (Additional file 4: Figure S1). Despite the data paucity, just under half of the patients with MF who had undergone LT (44.4%) did not have HRM mutations. In three PMF cases, the *IRF4* expression was evaluated during ruxolitinib treatment. All cases exhibited an increased *IRF4* expression compared to the value at diagnosis ($p = 0.003$); in the two patients with a longer





duration of ruxolitinib treatment (59 mo. and 72 mo., respectively), there was a one-log increment of *IRF4* expression (Additional file 5: Figure S2).

Despite the main limits of our study (relatively small number of patients in each group, few NGS data, inability to determine the *IRF4* production source)

we demonstrate an *IRF4* dysregulated expression in MPNs patients, particularly in PMF and in *JAK2*+ and *TN*+ cases, distinguishing those with a higher probability of SMF. Furthermore, the *IRF4* expression was associated with LT and a shorter LFS. Further studies are warranted to validate these data to confirm this biomarker as a new prognostic factor.

Abbreviations

IRF4: Interferon regulatory factor 4; MDSCs: Myeloid-derived suppressor cells; MPNs: Myeloproliferative neoplasms; Ph-MPNs: Philadelphia negative myeloproliferative neoplasms; BM: Bone marrow; HC: Healthy controls; PMF: Primary myelofibrosis; ET: Essential thrombocythemia; TN: Triple negative; LT: Leukemic transformation; SMF: Secondary myelofibrosis; PV: Polycythemia vera; LFS: Leukemia-free survival; MF: Myelofibrosis; OS: Overall survival; HMR: High molecular risk.

Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s40164-021-00253-y>.

Additional file 1: Supplementary methods. Detailed experimental procedures for droplet digital PCR, NGS and statistical analysis.

Additional file 2: Table S1. Main patients' data. MPN patients biological and clinical characteristics.

Additional file 3: Table S2. NGS results. VCF file reporting the annotation of variants identified by NGS analysis.

Additional file 4: Figure S1. Oncoprinter visualization of NGS results. Variants identified for all cases analyzed (columns) are reported. The percentage value associated with each gene indicates its variants occurring in the cohort analyzed.

Additional file 5: Figure S2. Expression analysis of the *IRF4* gene transcript at diagnosis and during ruxolitinib treatment in three myelofibrosis patients. The amount of *IRF4* gene transcript was significantly increased during ruxolitinib therapy in all cases analyzed.

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Authors' contributions

Conception and design of the study: CC, FT and FA. Acquisition of data and/or analysis and interpretation of data: CC, FT, LA, AZ, IR, CFM, NC, GT, AR, EP, MRC, GS, PM and FA. Drafting of the manuscript: FA. All authors revised the manuscript for important intellectual content and approved the final version submitted for publication. All authors read and approved the final manuscript.

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Availability of data and materials

Not applicable.

Declarations

Ethics approval and consent to participate

The local ethics committee approved the study. Informed consent was obtained from all patients before study inclusion, in accordance with the Declaration of Helsinki. Patients' records/information were anonymized and de-identified before analysis.

Consent for publication

Consent for publication was obtained from patients before their enrolment in the present study.

Competing interests

The authors declare that they have no competing interests.

Author details

¹Department of Emergency and Organ Transplantation (D.E.T.O.), Hematology and Stem Cell Transplantation Unit, University of Bari "Aldo Moro", P.Zza G. Cesare, 11, 70124 Bari, Italy. ²School of Medicine, University of Bari "Aldo Moro", 70124 Bari, Italy.

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