



## **Central Venous Disease Increases the Risk of Microbial Colonization in Hemodialysis Catheters**

Xianhui Liang<sup>1,2</sup>, Yamin Liu<sup>1</sup>, Bohan Chen<sup>1</sup>, Ping Li<sup>1</sup>, Peixiang Zhao<sup>1</sup>, Zhangsuo Liu<sup>1,2\*</sup> and Pei Wang<sup>1,2\*</sup>

<sup>1</sup> Blood Purification Center, Institute of Nephrology, the First Affiliated Hospital of Zhengzhou University, Zhengzhou, China, <sup>2</sup> Research Institute of Nephrology, Zhengzhou University, Zhengzhou, China

**Objectives:** Tunneled-cuffed catheters (TCCs) are widely used in maintenance hemodialysis patients. However, microbial colonization in catheters increases the likelihood of developing various complications, such as catheter-related infection (CRI), catheter failure, hospitalization, and death. Identification of the risk factors related to microorganism colonization may help us reduce the incidence of these adverse events. Therefore, a retrospective analysis of patients who underwent TCC removal was conducted.

#### **OPEN ACCESS**

#### Edited by:

Ping Fu, Sichuan University, China

#### Reviewed by:

Tomasz Tomasz Gołebiowski, Wroclaw Medical University, Poland Fan He, Huazhong University of Science and Technology, China

#### \*Correspondence:

Zhangsuo Liu zhangsuoliu@zzu.edu.cn Pei Wang wangpei146@hotmail.com

#### Specialty section:

This article was submitted to Nephrology, a section of the journal Frontiers in Medicine

Received: 23 December 2020 Accepted: 21 July 2021 Published: 23 August 2021

#### Citation:

Liang X, Liu Y, Chen B, Li P, Zhao P, Liu Z and Wang P (2021) Central Venous Disease Increases the Risk of Microbial Colonization in Hemodialysis Catheters. Front. Med. 8:645539. doi: 10.3389/fmed.2021.645539 **Methods:** From a pool of 389 adult patients, 145 were selected for inclusion in the study. None of the patients met the diagnostic criteria for CRI within 30 days before recruitment. The right internal jugular vein was the unique route evaluated. The catheter removal procedure was guided by digital subtraction angiography. Catheter tips were collected for culture. Biochemical and clinical parameters were collected at the time of catheter removal.

**Results:** The average age of this cohort was 55.46  $\pm$  17.25 years. A total of 45/145 (31.03%) patients were verified to have a positive catheter culture. The proportions of gram-positive bacteria, gram-negative bacteria, and fungi were 57.8, 28.9, and 13.3%, respectively. History of CRI [odds ratio (OR) = 2.44, 95% confidence interval (CI) 1.09 to 5.49], fibrin sheath (OR = 2.93, 95% CI 1.39-6.19), white blood cell (WBC) count >5.9  $\times$  10<sup>9</sup>/l (OR = 2.31, 95% Cl 1.12-4.77), moderate (OR = 4.87, 95% Cl 1.61-14.78) or severe central venous stenosis (CVS) (OR = 4.74, 95% CI 1.16-19.38), and central venous thrombosis (CVT) (OR = 3.41, 95% Cl 1.51-7.69) were associated with a significantly increased incidence of microbial colonization in a univariate analysis. Central venous disease (CVD) elevated the risk of microbial colonization, with an OR of 3.37 (1.47–7.71, P = 0.004). A multivariate analysis showed that both CVS and CVT were strongly associated with catheter microbial colonization, with ORs of 3.06 (1.20–7.78, P = 0.019) and 4.13 (1.21– 14.05, P = 0.023, respectively. As the extent of stenosis increased, the relative risk of catheter microbial colonization also increased. In patients with moderate and severe stenosis, a sustained and significant increase in OR from 5.13 to 5.77 was observed.

1

**Conclusions:** An elevated WBC count and CVD can put hemodialysis patients with TCCs at a higher risk of microbial colonization, even if these patients do not have the relevant symptoms of infection. Avoiding indwelling catheters is still the primary method for preventing CRI.

Keywords: tunneled cuffed catheter, catheter-related infection, central venous stenosis, central venous thrombosis, microbial colonization

## INTRODUCTION

Hemodialysis remains the main component of renal replacement therapy (RRT) worldwide. Well-functioning vascular access, a fundamental aspect of hemodialysis, has always been an area of great concern (1-3). Arteriovenous access, including arteriovenous fistulas (AVFs) and arteriovenous grafts (AVGs), is often characterized by longer patency and fewer complications and is also considered to be more ideal than central venous catheters (CVCs) (4, 5). However, a large proportion of patients (~45%) maintain their treatment through catheters based on the following considerations: late referral for dialysis, critical illness, poor peripheral vascular conditions, or low acceptance of arteriovenous access (6). Indwelling catheters are believed to be the primary source of systemic infection, aptly referred to as catheter-related infections (CRIs); therefore, indwelling catheters put patients at increased risks for catheter failure, death, hospitalization, and other adverse events (7-9). Hence, efforts to limit the use of CVCs and lessen the incidence of CRIs have become a focus of research (10, 11). Increasing evidence indicates that microbial colonization of a catheter is closely related to but often precedes bloodstream infection in hemodialysis patients (12-15). The risk of developing CRIs is dramatically increased in patients with positive catheter cultures (16). Strategies intended to prevent or eliminate microorganism colonization may be able to provide a substantial breakthrough in the care of patients with catheters (17). In this retrospective study, we collected the clinical data of patients with positive catheter cultures who had no evidence of a catheter-related blood stream infection (CRBSI); we also examined the factors involved in microbial colonization and central venous disease (CVD).

### MATERIALS AND METHODS

#### **Study Design**

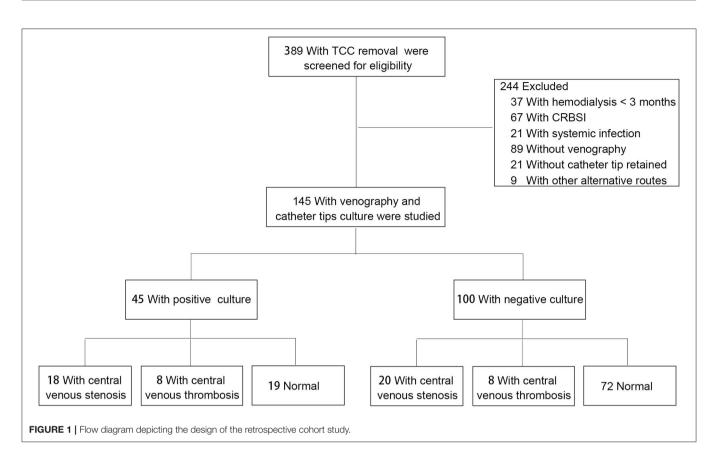
For this retrospective cohort study, a total of 389 adult patients who underwent tunneled-cuffed catheter (TCC) removal in our center between June 2019 and June 2020 were screened for inclusion (**Figure 1**). None of the patients met the diagnostic criteria for CRIs within 30 days before recruitment. The right internal jugular vein was the unique route chosen for analysis; patients with access through other alternative routes were excluded. Patients with incomplete clinical data, systemic infection, and exit-site or tunnel infection were not included. The procedure was guided by digital subtraction angiography and performed with strict aseptic techniques under local anesthesia. Antimicrobial lock solutions are not routinely used to prevent CRBSI; however, we usually use a 1,500-U/ml heparin sodium lock solution, and urokinase is used every 2–4 weeks to prevent catheter-associated thrombus. When anticoagulation was contraindicated, the patients received sodium chloride (10%). Finally, 145 patients were enrolled in this study for further assessment. Demographic data as well as clinical parameters were documented, including sex, age, primary cause of end-stage kidney disease, history of CRI, catheter indwelling time, presence of CVD, hemoglobin, platelet count, etc.

Catheter removal was performed when the catheter was no longer needed or the catheter was no longer able to be used. Successful kidney transplantation, maturation of arteriovenous access, external catheter breakage, catheter loosening or even falling out, and catheter dysfunction were common reasons for removal. After TCC removal, the follow process was performed: (1) A 5-cm segment of each catheter tip was collected and cultured. (2) Patients with an AVF or AVG that was ready for use were changed to arteriovenous access. (3) New peripheral vascular access was established for patients who preferred AVF or AVG. Usually, a new TCC was implanted through the femoral vein to maintain dialysis treatment before a patient was converted to arteriovenous access. Early cannulation grafts were used in patients when suitable. (4) For patients who still needed a catheter, a new TCC could be inserted over a guidewire during in situ exchange or after repuncture. Biochemical and clinical parameters were collected at the time of catheter removal. Catheter tip cultures were performed by semi-quantitative culture on blood agar plates in the Microbiology Laboratory of the First Affiliated Hospital of Zhengzhou University. The plate was incubated in a 35°C incubator for 48 h. The catheter tip cultures were considered positive if 15 or more colony-forming units (CFUs) were observed.

CVD was defined as central venous stenosis (CVS) or central venous thrombosis (CVT), both of which were diagnosed with digital subtraction angiography by two or more experienced vascular access physicians. CVT was defined as a filling defect in the lumen of the vessel, and CVS was defined as a narrow lumen. Stenosis was classified as mild, moderate, or severe according to the degree of stenosis: <50, 50–70, or  $\geq$ 70%, respectively.

#### **Statistical Analysis**

Continuous variables are summarized using the mean  $\pm$  SD or median (interquartile range). Categorical variables are summarized using numbers and percentages; continuous variables were analyzed by using *t*-tests and Mann–Whitney *U* tests, as appropriate. Categorical variables were compared using the  $\chi^2$  test or Fisher's exact test. Logistic binary regression was



applied to examine the association between catheter microbial colonization and the resulting variables. P < 0.05 was considered statistically significant.

## RESULTS

## **Patient Characteristics**

The average age of this cohort was 55.46  $\pm$  17.25 years. The male-to-female ratio was 1.46:1. Diabetes mellitus, chronic glomerulonephritis, and hypertension were the top three reasons for RRT. Positive catheter cultures were detected in a total of 45/145 (31.03%) patients, whereas 100/145 (68.97%) control patients had negative cultures. The mean indwelling catheter times for patients with and without microbial colonization were  $25.6 \pm 20.30$  months and  $26.91 \pm 24.61$  months, respectively. The patient characteristics (Table 1) revealed that, compared with the negative group, the positive group had a higher prevalence of CVD (57.8 vs. 28%, *P* = 0.001), more CVT (17.8 vs. 8.0%), more serious CVS (moderate stenosis, 24.3 vs. 7.6%; serious stenosis 13.5 vs. 4.3%), and a higher percentage of fibrin sheaths (46.7 vs. 23%, P = 0.004). Previous CRIs, a white blood cell (WBC) count over 5.9  $\times$  10<sup>9</sup>/l, and elevated C-reactive protein also increased the risk for microbial colonization.

## **Microbiological Characteristics**

Among the 45 positive cultures, the proportion due to grampositive bacteria was comparatively higher, and the proportions of gram-positive bacteria, gram-negative bacteria, and fungi were 57.8, 28.9, and 13.3% (**Table 2**), respectively. *Staphylococcus epidermidis* was the overwhelming primary pathogen, accounting for 65.4% of gram-positive bacteria, followed by *Pseudomonas aeruginosa, Escherichia coli*, and *Staphylococcus aureus*. All six strains of fungi were *Candida parapsilosis*.

# Analysis of Risk Factors for Microbial Colonization

As shown in **Table 1**, the number of patients with a history of CRI, fibrin sheath, elevated WBC count, and moderate to severe CVS and CVT was substantially different between the positive and negative groups. To determine whether these variables gave rise to microbial colonization of the catheter, we further conducted univariate logistic regression analyses. The results are shown in the right column of **Table 1**. History of CRI [odds ratio (OR) = 2.44, 95% confidence interval (CI) 1.09–5.49], fibrin sheath (OR = 2.93, 95% CI 1.39–6.19), WBC count  $\geq$ 5.9 × 10<sup>9</sup>/l (OR = 2.31, 95% CI 1.12–4.77), moderate CVS (OR = 4.87, 95% CI 1.61–14.78) or severe CVS (OR = 4.74, 95% CI 1.16–19.38), and CVT (OR = 3.41, 95% CI 1.51–7.69) were associated with a significantly increased incidence of microbial colonization.

Multivariate logistic regression (**Table 3**, **Figure 2**) was adjusted for age, sex, presence of diabetes mellitus, catheter indwelling time, fibrin sheath, history of CRI, and elevated WBC count and indicated that CVD elevated the risk of microbial colonization with an OR of 3.37 (1.47–7.71, P = 0.004). In model 2, when CVD was classified as thrombus or stenosis, the results

TABLE 1 | Clinical and laboratory data of patients in this retrospective cohort study.

Variable	Positive group (N = 45, %)	Negative group $(N = 100, \%)$	Р	OR (95%CI)	Р
Sex			0.632		0.632
Male	28 (62.2)	58 (58)		1	
Female	17 (37.8)	42 (42)		0.84 (0.41-1.73)	
Age			0.537		0.537
<60	25 (55.6)	61 (61.0)		1	
≥60	20 (44.4)	39 (39.0)		1.25 (0.61–2.55)	
Primary disease			0.277		0.288
Others	3 (6.7)	8 (8)		1	
Diabetic nephropathy	22 (48.8)	30 (30.0)		1.96 (0.46-8.22)	0.360
Hypertension	7 (15.6)	18 (18)		1.04 (0.21–5.08)	0.964
Chronic glomerulonephritis	10 (22.2)	35 (35)		0.76 (0.17–3.42)	0.723
Polycystic kidney	3 (6.7)	9 (9)		0.89 (0.14–5.72)	0.901
Diabetes mellitus	22 (48.9)	39 (39.0)	0.264	1.50 (0.74–3.04)	0.266
History of CRI	15 (33.3)	17 (17.0)	0.028	2.44 (1.09-5.49)	0.031
Reason for catheter removal			0.067	1	0.075
AVF, AVG maturation	9 (20)	34 (34.0)			
Kidney transplantation	4 (8.9)	18 (18.0)		0.84 (0.23–3.11)	0.793
Mechanical complication	4 (8.9)	4 (4.0)		3.78 (0.79–18.13)	0.097
Catheter dysfunction	28 (62.2)	44 (44.0)		2.40 (1.0-5.76)	0.049
Catheter indwelling time			0.276		0.277
<20 months	19 (42.2)	52 (52.0)		1	
≥20 months	26 (57.8)	48 (48.0)		1.48 (0.73–3.01)	
Central venous disease	26 (57.8)	28 (28.0)	0.001	3.52 (1.69–7.34)	0.001
Type of disease			0.003		0.004
Normal	19 (42.2)	72 (72.0)		1	
CVT	8 (17.8)	8 (8.0)		3.41 (1.51–7.69)	0.003
CVS	18 (40)	20 (20.0)		3.79 (1.26–11.41)	0.018
Degree of stenosis#			0.007		0.013
Normal	19 (51.4)	72 (78.3)		1	
Mild	4 (10.8)	9 (9.8)		1.68 (1.47–6.07)	0.425
Moderate	9 (24.3)	7 (7.6)		4.87 (1.61–14.78)	0.005
Severe	5 (13.5)	4 (4.3)		4.74 (1.16–19.38)	0.030
Fibrin sheath	21 (46.7)	23 (23.0)	0.004	2.93 (1.39–6.19)	0.005
WBC count (10 <sup>9</sup> /l)			0.023		0.024
<5.9	16 (35.6)	56 (56.0)		1	
≥5.9	29 (65.4)	44 (44.0)		2.31 (1.12–4.77)	
Hemoglobin (g/l)	$103.87 \pm 19.28$	$105.64 \pm 18.78$	0.604	1.00 (0.98–1.01)	0.601
Serum albumin (g/l)	$38.87 \pm 4.09$	$40.32\pm6.13$	0.097	0.95 (0.90–1.02)	0.153
Serum ferritin (ng/ml)	275.70 (179.02, 465.80)	246.40 (109.88, 418.70)	0.054	1.00	0.852
C-reactive protein ( $\mu$ g/ml)	13.10 (7.05, 20.49)	5.43 (2.46, 12.42)	<0.001	1.02 (1.00-1.05)	0.079

<sup>#</sup>CVS is graded according to the degree of stenosis, <50%, 50%−70%, ≥70% are mild, moderate, and severe stenosis, respectively. The catheter indwelling time and WBC count are divided into two groups according to the median. Data are expressed as mean ± SD, median (interquartile range), or number.

suggested that both CVT and CVS were strongly associated with catheter microbial colonization, with ORs of 3.06 (1.20–7.78, P = 0.019) and 4.13 (1.21–14.05, P = 0.023), respectively. In model 3, CVS was further divided into mild, moderate, and severe stenosis. As the extent of stenosis increased, the relative risk of catheter microbial colonization also increased. In patients with moderate and severe stenosis, a sustained and significant increase in risk was found, with an OR ranging from 5.13 to 5.77.

### DISCUSSION

It is widely accepted that arteriovenous access is preferential to CVCs in most incident and prevalent hemodialysis patients. Several clinical practice guidelines on vascular access, including the Kidney Disease Outcome Quality Initiative (K/DOQI), and expert consensus indicate that catheters should be used in certain clinical circumstances, such as failure to establish arteriovenous access, patient preference considered valid after proper assessment, and some specific medical conditions (4, 18). According to the Chinese Expert Consensus on Hemodialysis Vascular Access (second edition), the proportion of catheters should be limited to within 10% (19). Current evidence suggests

TABLE 2 | Distribution of positive pathogens.

Pathogens	Numbers	Proportion (%)
Gram-positive bacteria	26	57.8
Staphylococcus epidermidis	17	
Staphylococcus aureus	2	
Brevibacterium casei	1	
Staphylococcus hominis	1	
Staphylococcus haemolyticus	1	
Staphylococcus capitis	1	
Corynebacterium striata	1	
Staphylococcus wokerii	1	
Streptococcus angina	1	
Gram-negative bacteria	13	28.9
Pseudomonas aeruginosa	3	
Escherichia coli	2	
Actinobacillus rhizobium	1	
Acinetobacter lophi	1	
Achromobacter xylosoxidans	1	
Proteus mirabilis	1	
Stenotrophomonas maltophilia	1	
Enterobacter cloacae complex	1	
Enterobacter cloacae subsp. Cloacae	1	
Klebsiella oxytoca	1	
Fungus	6	
Candida parapsilosis	6	13.3

TABLE 3 | Multivariate logistic regression analysis of microbial colonization of TCC.

that catheters are associated with lower patency and higher complications than other vascular access types (20). CRI, for instance, is a well-known reason for all-cause mortality, hospitalization, and catheter dysfunction (21, 22). A number of recent publications reported that hemodialysis CVCs are associated with a two- to five-fold higher risk of infection-related hospitalization and a heavier economic burden due to CRBSI (23, 24). Understanding the pathogenesis of CRI can help us optimize catheter use. During catheter implantation, neglecting aseptic principles may result in microorganism migration. Relatedly, organisms transfer from the surrounding skin, health providers' hands, draping cloth next to the catheter exit site (known as an extraluminal pathway), or external or internal surfaces of catheter hubs or caps (extraluminal or intraluminal pathways) (17). Once organisms are attached to the catheter or accumulate in the fibrin sheath, biofilm production, a crucial and prerequisite step toward infection, is initiated. A mature biofilm is a unique self-sustaining colony of microorganisms protected by an exopolysaccharide matrix (25). The biofilm provides a suitable environment for the colonization and growth of organisms and helps them protect against antibiotics (17, 26, 27). In addition, a mature biofilm presents a hidden danger for subsequent disseminated infection and bloodstream infection. Studies have revealed a clear correlation between catheter microorganism colonization and CRBSI (12-14). The incidence of microorganism colonization in hemodialysis patients varies from 16.5 to 68% (12, 25, 28). From our observations, the rate of positive TCC tip cultures in patients without definite evidence of infection was 31.03%. The most common organisms (57.8%) detected were gram-positive bacteria, especially S. epidermidis.

A multivariate logistic regression analysis indicated that elevated WBC count, moderate to severe CVS, and CVT were independent risk factors for microbial colonization in catheters. CVS is often experienced by hemodialysis patients with arteriovenous access (29). In many cases, CVS is associated

Variable	Model 1		Model 2*		Model 3 <sup>#</sup>	
	OR	Р	OR	Р	OR	Р
Age	1.30 (0.58–2.92)	0.522	1.29 (0.57–2.89)	0.544	1.55 (0.63–3.80)	0.340
Sex	1.32 (0.57–3.05)	0.516	1.36 (0.58–3.19)	0.476	1.44 (0.59–3.53)	0.427
Diabetes mellitus	1.62 (0.70-3.74)	0.261	1.58 (0.68–3.68)	0.286	1.72 (0.68-4.35)	0.250
History of CRI	2.09 (0.83-5.23)	0.116	2.13 (0.85–5.35)	0.108	1.89 (0.71–5.08)	0.205
Catheter indwelling time	1.63 (0.72–3.67)	0.241	1.65 (0.73–3.73)	0.231	1.19 (0.49–2.91)	0.697
Fibrin sheath	2.22 (0.96–5.14)	0.062	2.31 (0.98-5.43)	0.056	1.99 (0.76–5.21)	0.162
WBC count (10 <sup>9</sup> /l)	2.47 (1.10-5.58)	0.029	2.42 (1.07-5.47)	0.034	2.59 (1.05-6.42)	0.039
Central venous disease	3.37 (1.47-7.71)	0.004	3.06 (1.20-7.78)	0.019 <sup>a</sup>	1.76 (0.44-7.02)	0.425 <sup>aa</sup>
			4.13 (1.21–14.05)	0.023 <sup>b</sup>	5.13 (1.41–18.63) 5.77 (1.04–32.04)	0.013 <sup>bi</sup> 0.045 <sup>co</sup>

The three models were adjusted for age, sex, diabetes mellitus, catheter indwelling time, fibrin sheath, history of CRI, and WBC count.

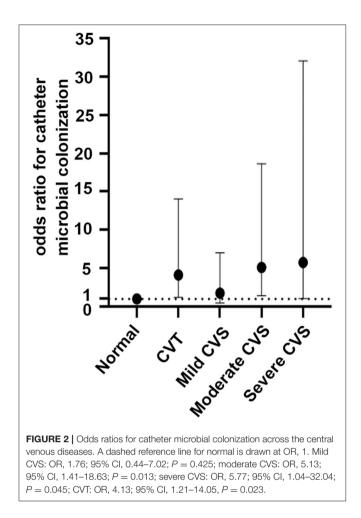
\*Model 2 central venous disease is stratified into CVS and CVT.

 ${}^{b}P = 0.023$  for CVT.

<sup>#</sup>Model 3 CVS is graded into mild, moderate, and severe stenosis according to the degree of stenosis.

 $^{aa}P = 0.425$  for mild CVS,  $^{bb}P = 0.013$  for mild CVS,  $^{cc}P = 0.045$  for CVS.

 $<sup>^{</sup>a}P = 0.019$  for CVS.



with indwelling catheters or other endovascular devices. The incidence of CVS varies greatly among different routes (30). The right internal jugular vein is preferable to the femoral vein for catheterization because of the lower risks for microbial colonization, CVS, and thrombotic complications (31). The incidence of CVS with a right internal jugular vein TCC was 26.2% in our study, which was relatively close to the 18-41% (29, 32, 33) reported in previous literature. Patients with CVS might have a higher chance of experiencing catheter dysfunction; therefore, more frequent nursing interventions were needed during the treatment and interval periods. This would undoubtedly increase the risk of microbes entering the catheter. Hemodynamic changes might be involved in the colonization process. Local lumen stenosis could result in greater blood flow resistance or even obstruction, which facilitates microorganism colonization in catheters. Furthermore, for reasons related or unrelated to CVS, hemodialysis patients with arteriovenous access often require percutaneous intracavitary treatment or surgical treatment to maintain access patency (34, 35). When a patient undergoes an operation or procedure, there is an additional opportunity for microorganisms to enter and spread, thereby amplifying the risk of microbial colonization in patients with stenotic lesions. In a retrospective study, 54 hemodialysis patients underwent venography at least half a year after catheter removal; the authors found that the incidence of CVS in patients with previous CRI was threefold higher than that in the control group (75 vs. 28%) (36), which indicated that CVS and CRI interact and have a synergistic relationship.

CVT is also common in patients receiving hemodialysis through a CVC. The incidence of CVT was 11% in this study. However, Wilkin et al. (37) confirmed CVT by ultrasonography in 2003 and observed a higher incidence of 25.9% in 143 hemodialysis patients with a history of an indwelling TCC. Importantly, various factors, such as different assessment techniques, catheter tip positions, frequencies of catheterization, and lock solutions, contribute to the variations in CVT incidence. On the one hand, CVT promotes colonization by blocking blood flow and aggravating vortex flow; on the other hand, thrombi composed mainly of fibrin, fibronectin, collagen, laminin, and several types of immunoglobulin provide abundant nutrition and a suitable area for microbial colonization. Microorganisms represented by S. aureus and S. epidermidis are prone to adhering in this area (38). Moreover, coagulase released by these microorganisms activates the coagulation system and accelerates thrombosis, thereby promoting bacterial colonization and biofilm growth, ultimately resulting in the spread of bacteria in the blood (39).

The association between other factors (age, sex, diabetes mellitus, history of CRIs, and catheter indwelling time) and microbial colonization in the catheter was not demonstrated in our study. Among these characteristics, the effect of age on CRIs is controversial. Older age was considered a protective factor for CRBSI in the report of Murea et al. (40). In their opinion, the worse physical condition of elderly patients makes them less active than younger patients, placing less external mechanical stress on the catheter, which in turn maintains the integrity of the subcutaneous tunnel and reduces the introduction of skin microorganisms and the formation of catheter biofilms. Conversely, older age was a risk factor for microbial colonization and bloodstream infection in our previous study (41) because it led to more possibilities of malnutrition, impaired immunity, and frailty. Moreover, our present findings did not support a correlation between history of CRIs and microbial colonization of the catheter, which was similar to the finding of Delistefani et al. (42) that past infection was related to recurrent local catheter infection (exit or tunnel infection) but not to CRBSI.

#### CONCLUSION

Our study indicates that an elevated WBC count and CVD were independent risk factors for TCC microbial colonization and may pave the way for large-scale, in-depth research on the association between CVD and CRI. Avoiding central venous catheterization and preventing catheter-related central venous lesions are crucial steps to reducing the incidence of microbial colonization in catheters.

## DATA AVAILABILITY STATEMENT

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

### **ETHICS STATEMENT**

The studies involving human participants were reviewed and approved by the Institutional Review Board of the First Affiliated Hospital of Zhengzhou University. Written informed consent for participation was not required for this study in accordance with the national legislation and the institutional requirements.

#### **AUTHOR CONTRIBUTIONS**

XL devised the conceptual ideas and took the lead in writing the manuscript. XL, PW, and ZL contributed to the study design. YL, PZ, and PL collected the data from tunneled-cuffed catheter removal patients. XL and PW performed catheter removal and

#### REFERENCES

- Cortez AJ, Paulson WD, Schwab SJ. Vascular access as a determinant of adequacy of dialysis. *Semin Nephrol.* (2005) 25:96–101. doi: 10.1016/j.semnephrol.2004.09.016
- Schwab SJ. Assessing the adequacy of vascular access and its relationship to patient outcome. Am J Kidney Dis. (1994) 24:316–20. doi: 10.1016/S0272-6386(12)80197-1
- Mandolfo S, Borlandelli S, Ravani P, Imbasciati E. How to improve dialysis adequacy in patients with vascular access problems. J Vasc Access. (2006) 7:53–9. doi: 10.1177/112972980600700203
- Lok CE, Huber TS, Lee T, Shenoy S, Yevzlin AS, Abreo K, et al. KDOQI Clinical practice guideline for vascular access: 2019 update. Am J Kidney Dis. (2020) 75:S1–S164. doi: 10.1053/j.ajkd.2019. 12.001
- Schmidli J, Widmer MK, Basile C, de Donato G, Gallieni M, Gibbons CP, et al. Editor's choice-vascular access: 2018 clinical practice guidelines of the european society for vascular surgery (ESVS). *Eur J Vasc Endovasc Surg.* (2018) 55:757–818. doi: 10.1016/j.ejvs.2018.02.001
- Pisoni RL, Zepel L, Port FK, Robinson BM. Trends in US vascular access use, patient preferences, and related practices: an update from the US DOPPS practice monitor with international comparisons. *Am J Kidney Dis.* (2015) 65:905–15. doi: 10.1053/j.ajkd.2014. 12.014
- Maraj S, Jacobs LE, Kung SC, Raja R, Krishnasamy P, Maraj R, et al. Epidemiology and outcome of infective endocarditis in hemodialysis patients. *Am J Med Sci.* (2002) 324:254–60. doi: 10.1097/00000441-200211000-00004
- Xue H, Ix JH, Wang W, Brunelli SM, Lazarus M, Hakim R, et al. Hemodialysis access usage patterns in the incident dialysis year and associated catheter-related complications. *Am J Kidney Dis.* (2013) 61:123– 30. doi: 10.1053/j.ajkd.2012.09.006
- Lok CE, Mokrzycki MH. Prevention and management of catheterrelated infection in hemodialysis patients. *Kidney Int.* (2011) 79:587– 98. doi: 10.1038/ki.2010.471
- Mermel LA, Allon M, Bouza E, Craven DE, Flynn P, O'Grady NP, et al. Clinical practice guidelines for the diagnosis and management of intravascular catheter-related infection: 2009 Update by the Infectious Diseases Society of America. *Clin Infect Dis.* (2009) 49:1–45. doi: 10.1086/599376
- Vanholder R, Canaud B, Fluck R, Jadoul M, Labriola L, Marti-Monros A, et al. Catheter-related blood stream infections (CRBSI): a European view. *Nephrol Dial Transplant*. (2010) 25:1753–6. doi: 10.1093/ndt/gfq205

angiography diagnosis. YL, PW, and XL contributed to the discussion and interpretation of the results. YL, BC, and XL contributed to manuscript editing. All authors agreed that the entire concept and ownership of this work belong to PW. All authors approved the final manuscript.

#### FUNDING

This study is supported by grants from the National Science and Technology Major Project of China (Grant No. 2020ZX09201-009).

#### ACKNOWLEDGMENTS

We thank Dr. Chunhua Song for assistance in data processing. We are grateful to all medical staff and staff nurses in the Blood Purification Center, Institute of Nephrology, the First Affiliated Hospital of Zhengzhou University, who strongly contributed to the success of the study.

- Dittmer ID, Sharp D, McNulty CA, Williams AJ, Banks RA, A. prospective study of central venous hemodialysis catheter colonization and peripheral bacteremia. *Clin Nephrol.* (1999) 51:34–9.
- Salzman MB, Rubin LG. Relevance of the catheter hub as a portal for microorganisms causing catheter-related bloodstream infections. *Nutrition*. (1997) 13:155-17S. doi: 10.1016/S0899-9007(97)00217-7
- 14. Mermel LA. What is the predominant source of intravascular catheter infections. *Clin Infect Dis.* (2011) 52:211–2. doi: 10.1093/cid/ciq108
- Fux CA, Uehlinger D, Bodmer T, Droz S, Zellweger C, Mühlemann K. Dynamics of hemodialysis catheter colonization by coagulasenegative staphylococci. *Infect Control Hosp Epidemiol.* (2005) 26:567–74. doi: 10.1086/502586
- Rodríguez-Aranda A, Alcazar JM, Sanz F, García-Martín F, Otero JR, Aguado JM, et al. Endoluminal colonization as a risk factor for coagulase-negative staphylococcal catheter-related bloodstream infections in haemodialysis patients. *Nephrol Dial Transplant*. (2011) 26:948–55. doi: 10.1093/ndt/gfq481
- Mermel LA. What is the evidence for intraluminal colonization of hemodialysis catheters. *Kidney Int.* (2014) 86:28–33. doi: 10.1038/ki.2013.527
- Kukita K, Ohira S, Amano I, Naito H, Azuma N, Ikeda K, et al. 2011 update Japanese society for dialysis therapy guidelines of vascular access construction and repair for chronic hemodialysis. *Ther Apher Dial.* (2015) 19:1–39. doi: 10.1111/1744-9987.12296
- Working Group of vascular Access, Blood Purification Center branch, Chinese Hospital Association. Expert Consensus on vascular access for hemodialysis in China (2nd Edition). *Chin J Blood Purif.* (2019) 18:365–81. doi: 10.3969/j.issn.1671-4091.2019.06.001
- Ethier J, Mendelssohn DC, Elder SJ, Hasegawa T, Akizawa T, Akiba T, et al. Vascular access use and outcomes: an international perspective from the Dialysis Outcomes and Practice Patterns Study. *Nephrol Dial Transplant*. (2008) 23:3219–26. doi: 10.1093/ndt/gfn261
- 21. Allon M. Dialysis catheter-related bacteremia: treatment and prophylaxis. *Am J Kidney Dis.* (2004) 44:779–91. doi: 10.1016/S0272-6386(04)01078-9
- Allon M, Daugirdas J, Depner TA, Greene T, Ornt D, Schwab SJ. Effect of change in vascular access on patient mortality in hemodialysis patients. *Am J Kidney Dis.* (2006) 47:469–77. doi: 10.1053/j.ajkd.2005.11.023
- Thomson PC, Stirling CM, Geddes CC, Morris ST, Mactier RA. Vascular access in haemodialysis patients: a modifiable risk factor for bacteraemia and death. QJM. (2007) 100:415–22. doi: 10.1093/qjmed/hcm040
- Taylor G, Gravel D, Johnston L, Embil J, Holton D, Paton S. Canadian Nosocomial Infection Surveillance Program; Canadian Hospital Epidemiology Committee. Incidence of bloodstream infection in multicenter

inception cohorts of hemodialysis patients. Am J Infect Control. (2004) 32:155-60. doi: 10.1016/j.ajic.2003.05.007

- Lok CE. Avoiding trouble down the line: the management and prevention of hemodialysis catheter-related infections. *Adv Chronic Kidney Dis.* (2006) 13:225–44. doi: 10.1053/j.ackd.2006.04.009
- Dasgupta MK. Biofilms and infection in dialysis patients. Semin Dial. (2002) 15:338–46. doi: 10.1046/j.1525-139X.2002.00084.x
- Marcus RJ, Post JC, Stoodley P, Hall-Stoodley L, McGill RL, Sureshkumar KK, et al. Biofilms in nephrology. *Expert Opin Biol Ther.* (2008) 8:1159– 66. doi: 10.1517/14712598.8.8.1159
- Kang JS, Jang HR, Lee JE, Park YJ, Rhee H, Seong EY, et al. The bacterial colonization in tunneled cuffed dialysis catheter and its effects on residual renal function in incident hemodialysis patients. *Clin Exp Nephrol.* (2016) 20:294–301. doi: 10.1007/s10157-015-1148-5
- Agarwal AK. Central vein stenosis. Am J Kidney Dis. (2013) 61:1001– 15. doi: 10.1053/j.ajkd.2012.10.024
- Ge X, Cavallazzi R, Li C, Pan SM, Wang YW, Wang FL. Central venous access sites for the prevention of venous thrombosis, stenosis and infection. *Cochrane Database Syst Rev.* (2012) 2012:CD004084.
- Maya ID, Allon M. Outcomes of tunneled femoral hemodialysis catheters: comparison with internal jugular vein catheters. *Kidney Int.* (2005) 68:2886– 9. doi: 10.1111/j.1523-1755.2005.00762.x
- Naroienejad M, Saedi D, Rezvani A. Prevalence of central vein stenosis following catheterization in patients with end-stage renal disease. Saudi J Kidney Dis Transpl. (2010) 21:975–8.
- MacRae JM, Ahmed A, Johnson N, Levin A, Kiaii M. Central vein stenosis: a common problem in patients on hemodialysis. ASAIOJ. (2005) 51:77– 81. doi: 10.1097/01.MAT.0000151921.95165.1E
- Saeed M, Newman GE, McCann RL, Sussman SK, Braun SD, Dunnick NR. Stenoses in dialysis fistulas: treatment with percutaneous angioplasty. *Radiology*. (1987) 164:693–7. doi: 10.1148/radiology.164.3.2956626
- Tordoir JHM, Zonnebeld N, van Loon MM, Gallieni M, Hollenbeck M. Surgical and endovascular intervention for dialysis access maturation failure during and after arteriovenous fistula surgery: review of the evidence. *Eur J Vasc Endovasc Surg.* (2018) 55:240–8. doi: 10.1016/j.ejvs.2017. 12.001
- Hernández D, Díaz F, Suria S, Machado M, Lorenzo V, Losada M, et al. Subclavian catheter-related infection is a major risk factor for the late development of subclavian vein stenosis. *Nephrol Dial Transplant*. (1993) 8:227–30. doi: 10.1093/oxfordjournals.ndt.a092436

- Wilkin TD, Kraus MA, Lane KA, Trerotola SO. Internal jugular vein thrombosis associated with hemodialysis catheters. *Radiology*. (2003) 228:697–700. doi: 10.1148/radiol.2283020681
- Herrmann M, Vaudaux PE, Pittet D, Auckenthaler R, Lew PD, Schumacher-Perdreau F, et al. Fibronectin, fibrinogen, and laminin act as mediators of adherence of clinical staphylococcal isolates to foreign material. *J Infect Dis.* (1988) 158:693–701. doi: 10.1093/infdis/158.4.693
- Raad II, Luna M, Khalil SA, Costerton JW, Lam C, Bodey GP. The relationship between the thrombotic and infectious complications of central venous catheters. JAMA. (1994) 271:1014–6. doi: 10.1001/jama.271.13.1014
- Murea M, James KM, Russell GB, Byrum GV. 3rd, Yates JE, Tuttle NS, et al. Freedman BI Risk of catheter-related bloodstream infection in elderly patients on hemodialysis. *Clin J Am Soc Nephrol.* (2014) 9:764– 70. doi: 10.2215/CJN.07710713
- Wang K, Wang P, Liang X, Lu X, Liu Z. Epidemiology of haemodialysis catheter complications: a survey of 865 dialysis patients from 14 haemodialysis centres in Henan province in China. *BMJ Open.* (2015) 5:e007136. doi: 10.1136/bmjopen-2014-007136
- Delistefani F, Wallbach M, Müller GA, Koziolek MJ, Grupp C. Risk factors for catheter-related infections in patients receiving permanent dialysis catheter. *BMC Nephrol.* (2019) 20:199. doi: 10.1186/s12882-019-1392-0

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

**Publisher's Note:** All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2021 Liang, Liu, Chen, Li, Zhao, Liu and Wang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.