Accuracy of the *Helicobacter pylori* diagnostic tests in patients with peptic ulcer bleeding: a systematic review and network meta-analysis

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Abstract

Introduction: Some studies suggest that the accuracy of *Helicobacter pylori* diagnostic tests is decreased in peptic ulcer bleeding (PUB). We aimed to assess the accuracy of diagnostic tests for *H. pylori* in patients with PUB in a diagnostic test accuracy (DTA) network meta-analysis. **Methods:** A systematic search was carried out in seven databases until November 2019. We collected or calculated true and false positive and negative values, and constructed 2×2 diagnostic contingency tables with reference standards including histology, rapid urease test, urea breath test, serology, stool antigen test, culture, and polymerase chain reaction. We ranked the index tests by the superiority indices (SI) and calculated pooled sensitivity and specificity of each test.

Discussion: Our search yielded 40 eligible studies with 27 different diagnostic strategies for *H. pylori.* In 32 articles, the reference standard was a combination of multiple tests. In 12 studies, the index tests were compared with a single testing method. We analyzed seven networks with the reference standards against a single or a combination of diagnostic index tests. None of the index tests had better diagnostic accuracy (SI between 9.94 and 2.17) compared with the individual index tests as all the confidence intervals included 1. Combined testing strategies had higher sensitivities (0.92–0.62) and lower specificities (0.85–0.46) while single tests proved to have higher specificities (0.83–0.77) and lower sensitivities (0.73–0.42).

Conclusion: Use of combined tests may have a rationale in clinical practice due to their higher sensitivities. The differences between the included DTA studies limited the comparison of the testing strategies.

Keywords: accuracy, bleeding, diagnostic, Helicobacter, histology, peptic, pylori, tests, UGIB, ulcer

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Introduction

Peptic ulcer bleeding (PUB) is the most frequent cause of acute nonvariceal upper gastrointestinal (GI) bleeding,^{1–4} and has a reported mortality of between 11% and 13.1%.^{5,6} *Helicobacter pylori* infection (HPI) remains one of the common causes of peptic ulcer disease.^{7,8} In the case of underlying HPI, untoward outcomes of the PUB episode such as rebleeding, need for repeat endoscopy and transfusion are better if the bacteria are

eradicated.⁹ Gisbert *et al.* found that eradication of HPI can reduce rebleeding episodes from 23.7% to 4.5%.¹⁰

An optimal testing strategy would be desirable; however, the international guidelines on PUB do not provide clear guidance for clinicians concerning HPI testing in the acute setting. The American College of Gastroenterology (ACG) guidelines recommend biopsy-based testing,⁴ whereas the

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European Society of Gastroenterological Endoscopy (ESGE)^{1,2} does not specify a diagnostic method. The American Society of Gastroenterological Endoscopy (ASGE) recommends the eradication of HPI, but it does not determine the method of testing.³

There is a lack of guidance in the Maastricht/ Florence V guideline on the ideal timing of eradication therapy after an acute episode of PUB,¹¹ and there are multiple logistical factors that hamper early testing and eradication.

Multiple previous studies and reviews proved a decreased diagnostic performance of HPI tests in PUB. The reasons include recent proton pump inhibitor (PPI) use, which can change the number and load of detectable organisms. Intragastric blood contains albumin and human plasma with killing factors, which can interfere with bacteria.^{11–13} A meta-regression reported that testing on the index admission underestimates the true prevalence of HPI in PUB, likely due to the decreased diagnostic performance.¹⁴ Therefore, an optimal strategy or preference for the available tests is needed.

The meta-analysis of Gisbert *et al.* in 2006 on the diagnostic accuracy of *H. pylori* testing in the setting of acute PUB drew many conclusions on the diagnostic performance of single tests compared with 'gold standards'.¹⁵ They found that biopsybased tests had low sensitivity and high specificity in PUB. The urea breath test remained highly accurate, but the stool antigen test was less reliable. They did not recommend serology as the first test. Since then, multiple studies have been completed, the genre of diagnostic test accuracy (DTA) meta-analysis improved significantly, with detailed recommendations in the Cochrane Handbook for DTA reviews.¹⁶ Also, the method of network meta-analysis for DTA studies was developed.¹⁷

We aimed to assess the diagnostic performance of all HPI testing strategies in PUB in a diagnostic test accuracy network meta-analysis.

Materials and methods

Protocol

A diagnostic accuracy meta-analysis and systematic review were planned using the Preferred Reporting Items for Systematic Review and Meta-Analysis (PRISMA) protocols for DTA studies.¹⁸ The analysis was registered in advance on PROSPERO with registration number CRD42019113083, and the protocol was later updated as a network metaanalysis due to the significant variation between the comparisons of testing strategies.¹⁹

Data sources and searches

We included studies from adult populations with PUB where index tests were compared with reference tests for identifying HPI. The outcomes were the diagnostic performance measures of the different diagnostic tests.

A systematic search was conducted in seven databases: Medline *via* PubMed, Embase, Cochrane Central Register of Controlled Trials, Clinical Trials. gov, WHO Global Health Library, Web of Science, Scopus, from inception to 1 November 2019. There were no restrictions or filters imposed on the search strategy apart from human studies.

Keywords for the computer-aided search were (bleed* OR haemorrhage OR hemorrhage OR haematemesis OR hematemesis OR melaena OR melena) AND ('upper gastrointestinal' OR 'upper GI' OR nonvariceal OR peptic OR gastric OR duodenal OR gastroduodenal OR antrum OR antral OR pylorus OR pyloric OR GU OR DU OR PU OR ulcer OR stomach OR curling) AND (helicobacter OR pylori). Additional articles were identified from the reference lists of primarily eligible studies.

Study selection

Records were managed by EndNote X7.4, software (Clarivate Analytics, Philadelphia, PA, USA). After exclusion of duplicates, the remaining studies were screened by title, abstract and finally by full text by two independent authors (NV, ME). Additional articles were searched manually and identified from the reference lists of primarily eligible studies. We calculated Cohen's kappa coefficient to measure the agreement between two raters (NV, ME) in three levels (title, abstract and full text) of the selection process.²⁰ Disagreements were resolved by consensus and the involvement of the senior reviewer (BE).

Eligibility criteria

All prospective, cross-sectional DTA studies with relevant information about the accuracy of any HPI diagnostic test without language restriction were included in our analysis. Articles without direct or indirect information on the true positives (TP), true negatives (TN), false positives (FP) and false negatives (FN) were excluded from the analysis. Conference abstracts were excluded after we discovered they did not contain enough information.

Data extraction

Data were extracted independently by two investigators (NV, ME) and populated manually into a purpose-designed Excel 2016 sheet (Office 365, Microsoft, Redmond, WA, USA). Data were collected on the year of publication, geographical location, study type, number of enrolled patients, and basic demographics (age, sex ratio). Most importantly, the raw data (TP, TN, FP, FN), the name, manufacturer, cut off value, biopsy site and timing of both index tests and reference standards were collected. Data about therapy after admission before a diagnosis of HPI, the timing of endoscopic examination, the bleeding source and the risk factors (smoking, alcohol consumption, nonsteroidal anti-inflammatory drug and aspirin use, history of GI bleeding and PUB) were also collected. Other relevant findings were mentioned in an additional column as free text. Disagreements were resolved by consensus and the involvement of the corresponding author.

Risk of bias and applicability

A Revised Tool for the Quality Assessment of Diagnostic Accuracy Studies (QUADAS-2) was used for the quality assessment of the DTA studies, and the result of the assessment was graphically demonstrated.²¹ Risk of bias was assessed independently by two investigators (NV, ME). Disagreements were resolved by consensus and the involvement of the corresponding author.

Statistical analysis

We performed a DTA network meta-analysis to investigate which diagnostic method can be the best choice to detect HPI in PUB. This method allows us to make direct as well as indirect comparisons through a common comparator (i.e. the reference standard).

We collected the raw data of diagnostic tests, TP, TN, FP, and FN values and created 2×2

tables for each study. If raw data on the diagnostic accuracy were not provided, but detailed indirect data on the diagnostic performance were available, TP, TN, FP, FN were calculated. To assess the relative performance of a diagnostic test, we calculated pooled sensitivity and specificity for the index tests compared with the reference standard, and ranked them according to superiority indices (SI). The larger the SI, the more accurately a test is expected to predict the targeted condition compared with other tests. The network meta-analytical calculations were performed by the R programming language (R Core Team 2019, Vienna, Austria, R version 3.6.1) developed by Nyaga *et al.*,¹⁷ with the use of the ANOVA arm-based model. Publication bias was not assessed as the Handbook for DTA Reviews of Cochrane Methods argues that conventional ways of assessment are not reliable and can lead to misinterpretation of the publication bias.16

To display the network, we constructed a graph where nodes represent different screening methods, and edges represent head-to-head comparisons. The size of nodes correlates with the number of studies. The thickness of edges represents the number of comparisons between the two tests. The potential nodes of the network were the single tests that had enough connections with other tests to allow statistical analysis in networks.

Since we performed a network meta-analysis of diagnostic accuracy studies in R software, inconsistency should have been calculated instead of heterogeneity. However, since the conditions of node splitting analysis were not met, we were unable to assess inconsistency.

Results

Study selection

Our final statistical analyses included 40 observational cross-sectional studies.^{12,22–60} The study selection process with the Cohen's kappa values is shown in Figure 1. Eligible studies were reported between 1998 and 2016 from four continents. The number of study participants ranged between 32 and 324. Characteristics of the studies included in our analysis are shown in Table 1.

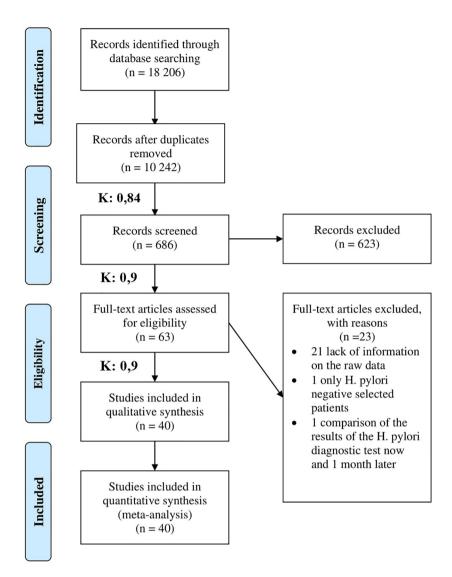


Figure 1. PRISMA flow chart for the study selection procedure. K value is the Cohen's kappa coefficient, if K is between 0.81 and 1.00 it means an almost perfect or perfect agreement. PRISMA, preferred reporting items for systematic review and meta-analysis.

Results of meta-analysis

The included 40 studies used 27 different definitions of the reference standard. In 32 articles, the reference standard was a combination of multiple tests. In 12 studies, the index tests were compared with a single testing method. We could form seven networks with the single tests (histology, rapid urease test, urea breath test, serology, stool antigen test, culture, polymerase chain reaction) serving as gold standards of the following networks. In the seven networks, the top three index tests based on their SI are shown in Table 2. None of the index tests had better diagnostic accuracy (SI between 9.94 and 2.17) compared with the individual index tests as all the confidence intervals included 1. Combined testing strategies had higher sensitivities (0.92–0.62) and lower specificities (0.85–0.46), while single tests proved to have higher specificities (0.83–0.77) and lower sensitivities (0.73–0.42). Out of the single tests, only the urea breath test against histology,

Author	Country	Index tests	Definition of reference standards	N° of patients
Archimandritis <i>et al.</i> ²²	Greece	Rapid urease test	Positive histology	72
Bravo Paredes <i>et al.</i> ²³	Peru	Rapid urease test	Positive histology	93
Castro-Fernandez et al. ²⁴	Spain	Rapid urease test	Assumed 100% infection	120
Castro-Fernandez et al. ²⁵	Spain	Rapid urease test, histology	Positive at least one of the followings: rapid urease test and histology or urea breath test	173
Chandrasakha <i>et al</i> . ²⁶	Thailand	Polymerase chain reaction, rapid urease test, histology, culture	Positive culture or a positive rapid urease test and histology	64
Choi <i>et al.²⁷</i>	Korea	Culture, rapid urease test, histology	Positive culture or any two of the followings: rapid urease test, histology, serology	157
Chung <i>et al.</i> ²⁸	Korea	Histology, rapid urease test, urea breath test, serology	At least two positive of the followings: rapid urease test, histology, urea breath test, serology	32
Chung <i>et al.</i> ²⁹	Korea	Polymerase chain reaction	Positive in two of the followings: histology, rapid urease test, serology	170
Colin <i>et al.</i> ³⁰	France	Rapid urease test, histology, culture	Positive serology	182
Demiray <i>et al.</i> ³¹	Turkey	Stool antigen test	Positive urea breath test or histology and/or rapid urease test	22
Garcia-Diaz <i>et al.</i> ³²	Spain	Serology	Positive in for at least one of the followings: rapid urease test, histology, culture or urea breath test	214
Gisbert <i>et al.</i> ³⁴	Spain	Stool antigen test	Positive rapid urease test or histology or urea breath test	34
Gisbert <i>et al.</i> ³³	Spain	Urea breath test	Positive urea breath test	131
Grino <i>et al.</i> ³⁵	Spain	Rapid urease test, urea breath test serology, histology	Positive histology or positive serology and urea breath test	78
Grino <i>et al.³⁶</i>	Spain	Stool antigen test, rapid urease test, histology, urea breath test, serology	Positive histology or positive two of the followings: rapid urease test, serology, urea breath test	68
Hanvivatong <i>et al.³⁷</i>	Taiwan	Stool antigen test, urea breath test serology, rapid urease test, histology	Positive culture or two of histology, rapid urease test, serology, urea breath test	56
Lahmidani <i>et al.</i> ³⁸	Marocco	Rapid urease test	Positive histology	116
Lee <i>et al.</i> ³⁹	Taiwan	Rapid urease test	Positive culture or histology	55
Lee et al. ⁴⁰	Taiwan	Rapid urease test	Positive urea breath test	116
Liao <i>et al.</i> 41	China	Rapid urease test, histology, urea breath test	At least two positive of the followings: rapid urease test, histology, urea breath test	57

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TaiwanRapid urease test, culture, histology, urea breath test, serologyRositive histology and culture or any two of the followings: rapid ureaseet al. ⁵⁵ The NetherlandsStool antigen testPositive culture or histology and rapid urease test ⁵⁷ SpainUrea breath testPositive histology or rapid urease test or urea breath test ⁵⁷ SpainUrea breath testPositive histology or rapid urease test or urea breath test ⁵⁷ SpainUrea breath testPositive histology or rapid urease test or urea breath test ⁵⁹ PolandRapid urease test, urea breath test, serologyPositive histology or rapid urease test or urea breath test1, ⁵⁹ PolandUrea breath test, urea breath test, serologyPositive histology1, ⁵⁹ PolandUrea breath testJapanSerologyPositive rapid urease test or culture or histology	Tang <i>et al.</i> ⁵⁵	Taiwan	Rapid urease test	Positive histology	324
et al.56The NetherlandsStool antigen test57NetherlandsPositive histology and rapid urease test57SpainUrea breath test57SpainUrea breath test58DenmarkRapid urease test, urea breath test, serology59PolandUrea breath test, serology59PolandUrea breath test, serology59DapanSerology59DapanSerology59DapanSerology50DoilDestive rapid urease test or culture or histology	Tu et al. ¹²	Taiwan	Rapid urease test, culture, histology, urea breath test, serology	Positive histology and culture or any two of the followings: rapid urease test, urea breath test, serology	77
⁵⁷ Spain Urea breath test stensen Denmark Positive histology or rapid urease test or urea breath test stensen Denmark Rapid urease test, urea breath test, serology ol. ⁵⁹ Poland Urea breath test Japan Serology Positive rapid urease test or culture or histology	van Leerdam <i>et al.</i> ⁵⁶	The Netherlands	Stool antigen test	Positive culture or histology and rapid urease test	36
stensen Denmark Rapid urease test, urea breath test, serology Positive histology ol. ⁵⁹ Poland Urea breath test Positive serology Japan Serology Positive rapid urease test or culture or histology	Velayos <i>et al.</i> ⁵⁷	Spain	Urea breath test	Positive histology or rapid urease test or urea breath test	70
 Poland Urea breath test Positive serology Japan Serology Positive rapid urease test or culture or histology 	Wildner-Christensen et al. ⁵⁸	Denmark	Rapid urease test, urea breath test, serology	Positive histology	95
Japan Serology Positive rapid urease test or culture or histology	Winiarski <i>et al.</i> ⁵⁹	Poland	Urea breath test	Positive serology	81
	Wong <i>et al.</i> ⁶⁰	Japan	Serology	Positive rapid urease test or culture or histology	59

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Gold standards	Index tests	N° of patients	Superiority index (95% CI)	Pooled sensitivity (95% CI)	Pooled specificity (95% CI)
Histology	Positive culture or two of the followings: histology, rapid urease test, serology, urea breath test	41	7.51 (0.05–25)	0.79 (0.33–1.00)	0.70 (0.22-0.96)
	Positive culture or two of the followings: rapid urease test, histology, serology	157	6.52 (0.05–23)	0.81 (0.29–1.00)	0.67 (0.17–0.96)
	Positive urea breath test	95	5.98 (0.05–21)	0.73 (0.23-0.97)	0.77 (0.27–0.99)
Rapid urease test	Positive histology and/or rapid urease test	81	9.94 (0.05–33)	0.84 (0.32–1.00)	0.59 (0.14–0.95)
	Positive histology and urea breath test	96	7.89 (0.04–31)	0.81 (0.33-1.00)	0.61 (0.14–0.95)
	Positive at least two of the followings: rapid urease test, histology, urea breath test	57	7.24 (0.04–33)	0.86 (0.36–1.00)	0.46 [0.10-0.76]
Urea breath test	Positive histology and rapid urease test	81	7.56 (0.07–23)	0.81 (0.34–1.00)	0.73 (0.23–0.99)
	Positive at least two of the followings: rapid urease test, histology, urea breath test, serology	64	6.53 (0.08–21)	0.89 (0.48–1.00)	0.61 (0.16–0.97)
	Positive at least two of the followings: rapid urease test, histology, urea breath test	57	6.48 (0.07–21)	0.76 [0.24–0.99]	0.79 (0.31–1.00)
Serology	Positive at least two of the followings: rapid urease test, histology, urea breath test, serology	32	7.81 (0.08–21)	0.72 (0.35–0.97)	0.80 (0.23–1.00)
	Positive histology or positive serology and urea breath test	68	5.69 (0.06–19)	0.83 (0.36–0.99)	0.57 (0.12–0.90)
	Positive histology or two of the followings: rapid urease test, serology, urea breath test	78	5.06 (0.06–17)	0.82 (0.29–0.99)	0.52 (0.10–0.88)
Stool antigen test	Positive culture or positive rapid urease test and histology	93	3.87 (0.33–9)	0.76 (0.56–0.89)	0.73 (0.49–0.89)
	Positive culture or two of the followings: histology, rapid urease test, serology, urea breath test	34	3.77 (0.11–11)	0.80 (0.32–1.00)	0.55 (0.13–0.93)
	Positive histology or two of the followings: rapid urease test, serology, urea breath test	68	2.66 (0.11–9.05)	0.72 (0.23–0.98)	0.56 [0.12-0.96]
Culture	Positive culture or positive rapid urease test and histology	199	7.35 (0.71–11)	0.94 (0.83–1.00)	0.77 [0.45-0.94]
	Positive culture or two of the followings: rapid urease test, histology, serology	157	2.28 (0.11–9)	0.84 (0.35–1.00)	0.46 [0.08–0.91]
	Positive culture or two of the followings: rapid urease test, histology, serology	55	2.17 (0.11–9)	0.82 (0.36–1.00)	0.49 (0.10–0.88)
Polymerase chain reaction	Positive culture or rapid urease test and histology	152	4.99 (0.11–13)	0.62 (0.34–0.88)	0.85 (0.48-0.96)
	Positive culture	79	4.47 (0.07–11)	0.42 [0.07–0.79]	0.83 (0.32–1.00)
	Positive culture or positive three of the followings: rapid urease test, histology, urea breath test, serology	55	4.44 (0.14–15)	0.68 (0.34–1.00)	0.80 (0.27–0.98)
Cl, confidence interval.	rval.				

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and culture against polymerase chain reaction, had SI values ranked within the top three. The graphically displayed networks, results of the full analysis and ranking are detailed in Supplementary Files S1–S7. When we ranked the index tests based on their pooled sensitivity, only combinations of tests ranked in the top three in all seven networks. Ranking of index tests based on their pooled specificity identified nine single tests among all 21 top three ranks. However, all sensitivity and specificity values had wide 95% confidence intervals (CIs) ranging between 0.0 and 1. The pooled specificity values with a corresponding CIs and top three specificity and sensitivity values highlighted are shown in Supplementary Files S1–S7.

Risk of bias and applicability assessment

With the use of QUADAS-2 assessment tool, overall, only two studies^{56,58} proved to be free of an unclear or high risk of bias. Five studies^{42,43,45,47,51} were found to carry an unclear risk of bias. The remaining 33 articles carried a high risk of bias. Detailed results of the risk assessment are shown in Table 3.

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Study	Risk of bias	Risk of bias				Applicability concerns		
	Patient selection	Index test	Reference standard	Flow and timing	Patient selection	Index test	Reference standard	
Archimandritis et al. ²²	8	0	8	0	0	0	0	
Bravo Paredes <i>et al.</i> ²³	8	?	?	?		0	0	
Castro-Fernandez et al. ²⁴	8	?	8	0	?	?	8	
Castro-Fernandez <i>et al.</i> ²⁵	?		8	8		0	0	
Chandrasakha <i>et al.</i> ²⁶			0	8	0	0	<u></u>	
Choi <i>et al.</i> ²⁷		?	0	8	0	0		
Chung et al. ²⁸	8	?	?	8	8	?	0	
Chung et al. ²⁹	8	?	?	8	0	٢	0	
Colin <i>et al.</i> ³⁰	8	0	8	?	٢	0	0	
Demiray et al. ³¹	?	?	8	8	?	?	?	
Garcia-Diaz et al. ³²	?	?	8	8	٢	0	0	
Gisbert <i>et al.</i> ³⁴	8	?	8	8	٢	٢	٢	
Gisbert <i>et al.</i> ³³	©	?	8	8	٢	8	8	
Grino <i>et al.</i> ³⁵	٢	?	?	8	٢	٢	٢	
Grino et al. ³⁶	0	0	0	8	٢	0	0	
Hanvivatong <i>et al.</i> ³⁷	٢	?	8	8	0	٢	0	
Lahmidani <i>et al.</i> ³⁸	8	?	?	٢	٢	٢	٢	
Lee et al. ³⁹	?	?	?	8	٢	٢	٢	
Lee et al. ⁴⁰	?	?	8	٢	٢	٢	٢	
Liao <i>et al.</i> ⁴¹			0	8		0	0	

(Continued)

Table 3. (Continued)

Study	Risk of bias	Risk of bias				Applicability concerns		
	Patient selection	Index test	Reference standard	Flow and timing	Patient selection	Index test	Reference standard	
Lin et al. ⁴²	?	0	٢	?	0	٢	٢	
Lin et al. ⁴³	?	0	0	?	0	0	0	
Lo et al. ⁴⁴	0	0	٢	8	0	٢	٢	
López Peñas <i>et al.</i> ⁴⁵	?	?	?	?	?	?	?	
Manguso <i>et al.</i> 46	8	0	٢	8	0	٢	0	
Pascual et al.47	0	?	?	?	0	٢	٢	
Peitz <i>et al.</i> ⁴⁸	8	0	٢	8	٢	٢	٢	
Peitz <i>et al.</i> ⁴⁹	0	0	٢	8	٢	٢	0	
Ramírez-Lázaro <i>et al.</i> 50	8	0	8	٢	8	0	8	
Romero Gómez <i>et al.</i> ⁵¹	?	?	?	?	?	?	?	
Saez et al. ⁵²	8	?	?	8	٢	0	٢	
Shilling et al. ⁵³	0	?	?	8	0	٢	٢	
Sfarti <i>et al.</i> ⁵⁴	?	?	8	8	٢	٢		
Tang et al.55	0	0	٢	0	٢	٢	0	
Tu <i>et al.</i> ¹²	?	?	?	8	٢	٢	٢	
van Leerdam <i>et al.</i> 56	0	0	٢	8	٢	٢	٢	
Velayos <i>et al.</i> ⁵⁷	8	?	8	8	?	?	?	
Wildner-Christensen <i>et al.</i> ⁵⁸	0	0	0	٢	0	0	٢	
Winiarski <i>et al.</i> ⁵⁹	?	?	8	?	٢	٢	٢	
Wong et al. ⁶⁰	8	?	?	8	?	?	?	

Discussion

Our results from the network meta-analysis of the eligible DTA studies could order the index tests for the detection of HPI in PUB based on their superiority index. Still, their wide confidence intervals could not prove this order beyond doubt. Combined index tests showed a tendency of higher sensitivity, while single index tests had higher specificity values when ranked.

Reasons for combined tests as diagnostic gold standards in the included studies

The majority of the included DTA studies (28 of 40) used a combination of multiple testing methods for HPI as a gold standard. None of these studies gave specific reasons for the approach above. This seems to be an established strategy across the studies without sufficient evidence. With the use of the combined tests, one can increase the sensitivity of the testing. The use of this strategy in DTA studies is controversial, as it identifies more patients with HPI but compromises the validity of the results of the DTA – even more so, as 15 of 30 DTA studies included the index test in their combined method of the gold standard.

We believe that HPI in PUB should be detected as soon as possible and it is a preferable strategy over delayed testing. With this approach, clinicians can maximise the number of patients who genuinely need eradication and, at the same time, a small but increased proportion of patients will receive eradication unnecessarily (FNs). Given the significant risks of untreated HPI after PUB and the low risks of potential side effects (diarrhoea 1.6%; bloating or abdominal pain 1.3%; nausea or vomiting 0.4%) from the unnecessary antibiotic therapy,⁶¹ the clinical approach of combined testing seems reasonable.

Another reason for the combined and more aggressive testing approach can be the risk of loss to follow up after hospitalisation for PUB. The previous Maastricht/Florence IV guideline recommended the initiation of eradication at the time of introduction of oral feeding,62 arguing that a proportion of patients would be lost to follow up. In the study of Yoon et al., results showed that 13.3% of the patients who were lost to follow up before the confirmation of HPI, and 41.4% of the patients confirmed to have HPI after discharge, did not receive eradication.63 In 2014, as part of a retrospective analysis, Kim et al. reported that only 47% of PUB patients had HPI testing during their index admission, less than 10% had any testing after discharge, and 15% were lost to follow up.64 Given the findings above, the identification of HPI in the acute setting is of utmost importance.

Reasons for missing the opportunity to test HPI during admission for PUB

Invasive tests requiring tissue sampling during endoscopy. The endoscopic procedures for patients presenting with PUB are often stressful and are done out of hours, when access to diagnostic tools may be limited. Endoscopic management of acute GI bleeding is likely associated with patient and operator fatigue, and results in poorer adherence to guidelines. After the successful endoscopic termination of an acute PUB, the endoscopist may feel that histological sampling could contribute to a recurrent episode of bleeding. Finally, poor visualisation due to residual blood during the intervention may prevent safe sampling.

Another essential clinical problem leading to reluctance to take biopsies is PUB aggravated by anticoagulant and antiplatelet treatment. A recent multicentric retrospective study from France on upper GI bleeding found that 475 of 2498 patients (19%) took oral anticoagulants, either Vitamin K antagonist or direct oral anticoagulant.⁶⁵ A French prospective multicenter study in 2011 reported 8.1% antiplatelet use in upper GI bleeding.⁶⁶

Non-invasive tests. The urea breath test has very low feasibility in the acute setting of PUB, as patients have to fast before and often after the index endoscopy.

Stool antigen testing has a similar problem concerning feasibility; the opportunity of stool sampling for HPI testing in the admission department is often missed. Also, a Dutch study revealed a high rate of false-positive results in PUB patients explained by a cross-reaction with the blood.⁵⁶

Also, patients with PUB receive, and are committed to, long-term PPI treatment before a urea breath test and stool antigen test is performed.¹ The current guideline suggests a 14 day PPI free period before a urea breath test can be performed.¹¹ Even 3 days of high dose PPI treatment reduces the detection of *H. pylori* significantly, and patients presenting with PUB would often receive PPI treatment before the index endoscopy.⁶⁷

Serology would seem the most feasible out of all non-invasive tests, but the presence of antibodies may only indicate a previous infection instead of an acute one.⁶⁸

Strengths of the study

The update of the most recent DTA meta-analysis on the same topic was published in 2006,¹⁵ with the then most recent study published in 2004; therefore, an update was necessary. We used a comprehensive and rigorous search strategy in seven databases. Detailed data extraction covered 70 items, which are shown in Supplementary file S8. We used a new statistical method of network meta-analysis developed for DTA studies. Assessment of the risks of bias was performed by the purpose-designed tool of QUADAS-2. Due to the clear study designs of the included reports, the patient population matched the review question.

Limitations of the study

As detailed in Table 3, the overall quality of the included studies was suboptimal, with high and unclear risks of bias. In many studies, it was unclear whether enrollment was consecutive, and inappropriate exclusion of subjects often occurred. Blinding the interpretation of the index tests and threshold of cut off value were not defined in many studies. Blinding the interpretation of the reference tests were not pre-specified. Another significant limitation was the unclear or prolonged interval between the index and reference tests. Not all patients received the same reference standard test when a combination was used. In some studies, a few participants were excluded from the final analysis.

Not only the combination of tests and index tests, but also the actual tests differed (manufacturer, methodology, etc.). In some articles, PPI use preceded either or both index and reference tests. The tissue sampling was not uniform across studies: some used antral, others antral and gastric body mucosal samples.

Implications for clinical practice

Combined tests may have a role in HPI testing in PUB as they have higher sensitivities. Endoscopic and gastroenterology units should have a tailored approach based on the availability of the individual tests.

Implication for research

Future DTA studies should use uniform gold standards. Also, they should focus on the feasibility and cost-effectiveness of the combined testing strategies.

In conclusion, our network meta-analysis demonstrated that none of the individual tests or the strategy of combined tests is superior in the detection of HPI. The combined tests have an increased sensitivity, which can translate to an optimized eradication strategy as it can result in the identification of most patients needing eradication therapy.

Author contributions

NV, BT, AS and PH conceptualized and designed the study in cooperation with BE; DP, SP, and

ZS constructed the search query. NV, AM, and ZS carried out the search process. NV and ME screened the articles for eligibility. NV, ME, and BE performed the data extraction; NV, ME and BE conducted the quality assessment. NV, BE, BT and ZS wrote the article. AS carried out the statistical analysis. DP, PS, AM, BT, ME, and PH provided valuable feedback after critically reviewing the first drafts of the manuscript. All authors contributed and approved the final manuscript for publication.

Conflict of interest statement

The authors declare that there is no conflict of interest.

Ethical approval

No ethical approval was required for this review as all data were already published in peer reviewed journals. No patients were involved in the design, conduct or interpretation of our review.

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Supplemental material

Supplemental material for this article is available online.

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