

A Simple Near-Infrared Fluorescent Probe for the Detection of Peroxynitrite

Luling Wu^{+, [a]}, Xue Tian^{+, [a]}, Hai-Hao Han,^[b] Jie Wang,^[b] Robin R. Groleau,^[a] Paramabhorn Tosuwan,^[c] Boontana Wannalarse,^[c] Adam C. Sedgwick,^[d] Steven D. Bull,^[a] Xiao-Peng He,^{*, [b]} and Tony D. James^{*, [a]}

Herein, we report the evaluation and synthesis of a reaction based fluorescent probe **DCM-Bpin** for the detection of Peroxynitrite (ONOO⁻). **DCM-Bpin** exhibits selective fluorescence off-on response for ONOO⁻ over other reactive oxygen species, including H₂O₂. Moreover, **DCM-Bpin** is biocompatible and has been used to visualize exogenous ONOO⁻ in HeLa cells.

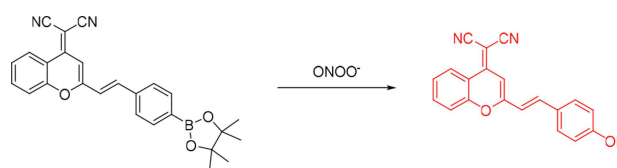
Peroxynitrite (ONOO⁻) is a key intracellular signaling molecule in both physiological and pathological processes, formed *in vivo* by the very fast radical coupling reaction between nitric oxide (NO^{*}) and superoxide (O₂^{*-}).^[1] The high reactivity of ONOO⁻ leads to the reaction with almost all types of biomolecules, such as proteins, lipids and DNA, causing oxidative stress and deleterious effects on cellular function.^[2,3] As such, ONOO⁻ has been implicated as a key pathogenic factor for a number of diseases, including inflammatory, ischemia-reperfusion and neurodegenerative diseases,^[4,5] therefore new and effective technologies for ONOO⁻ detection are of prime importance. Of particular interest is the development of novel small molecule fluorescent probes for the detection of such species, as

fluorescence probes are often more selective, less invasive and more convenient than many other methods for the detection of biologically relevant analytes in cells.^[6-8] Although a wide array of sensors have been developed with absorption and emission peaks in the visible range (400–650 nm), near-infrared (NIR) fluorescence sensors (emission in 650–900 nm NIR region) are still rare, despite distinct advantages for *in vitro* and *in vivo* tracing of molecular processes.^[9] Not only does NIR fluorescence avoid interference with the auto-fluorescence of indigenous molecules, near-infrared light results in less scattering and deeper tissue penetration, limiting the damage to living cells.^[10,11] As such, the development of NIR fluorescent sensors is of growing interest to the sensing community.^[12-16]

Herein we report the evaluation of a D-π-A-based^[17] fluorescent probe **DCM-Bpin**, in which a 2-(2-methyl)-4*H*-chromen-4-ylidene)malononitrile (**DCM**) serves as the NIR fluorescence acceptor, and the boronate ester Bpin moiety acts as an ONOO⁻ reporter. The fluorescence of the **DCM** system is quenched by Bpin, and so its rapid oxidation to the corresponding phenol (donor) by ROS species can be used to trigger a turn-on response, resulting in a peroxynitrite-activated “turn-on” probe (Scheme 1). **DCM-Bpin** has previously been reported as part of upconversion nanoparticles (UCNPs) for the detection of hydrogen peroxide, however, the **DCM-Bpin** probe was not evaluated for the detection of ONOO⁻,^[18] and given that peroxynitrite reacts with boronates much more rapidly,^[19] we reasoned this system would be suitable for the detection of peroxynitrite.

The synthesis of **DCM-Bpin** was successfully achieved in one step by the condensation of 4-formylphenylboronic acid, pinacol ester and **DCM** (using the previously described procedure).^[20] Heating to reflux in ethanol and piperidine for 4.5 hours, followed by filtration and washing with cold ethanol produced **DCM-Bpin** as a yellow solid in 62% yield (Scheme 2).^[18]

The UV-Vis (Figure S1) and fluorescence (Figure 1) behavior of **DCM-Bpin** was evaluated in pH 7.40 buffer solution (5%



Scheme 1. Fluorescence “turn on” mechanism of **DCM-Bpin** with the addition of ONOO⁻.

[a] L. Wu,⁺ X. Tian,⁺ R. R. Groleau, Prof. S. D. Bull, Prof. T. D. James
Department of Chemistry
University of Bath, BA2 7AY, UK
E-mail: t.d.james@bath.ac.uk

[b] H.-H. Han, J. Wang, Prof. X.-P. He
Key Laboratory for Advanced Materials and Joint International Research
Laboratory of Precision Chemistry and Molecular Engineering, Feringa
Nobel Prize Scientist Joint Research Center, School of Chemistry and
Molecular Engineering, East China University of Science and Technology,
130 Meilong Rd., Shanghai 200237, China
E-mail: xphe@ecust.edu.cn

[c] P. Tosuwan, Dr. B. Wannalarse
Paramabhorn Tosuwan and Boontana Wannalarse
Department of Chemistry, Faculty of Science
Kasetsart University, Bangkok 10900, Thailand

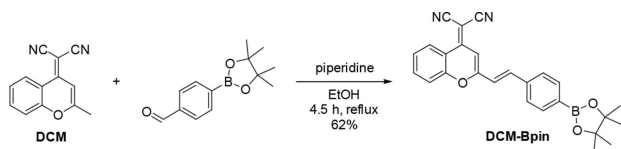
[d] Dr. A. C. Sedgwick
Department of Chemistry
University of Texas at Austin, 105 E 24th street A5300, Autin, TX 78712-
1224, USA

[†] equal contribution

Supporting information for this article is available on the WWW under
<https://doi.org/10.1002/open.201900301>

An invited contribution to a Special Collection dedicated to Functional Supramolecular Systems

© 2019 The Authors. Published by Wiley-VCH Verlag GmbH & Co. KGaA. This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.



Scheme 2. Synthesis of target probe DCM-Bpin.

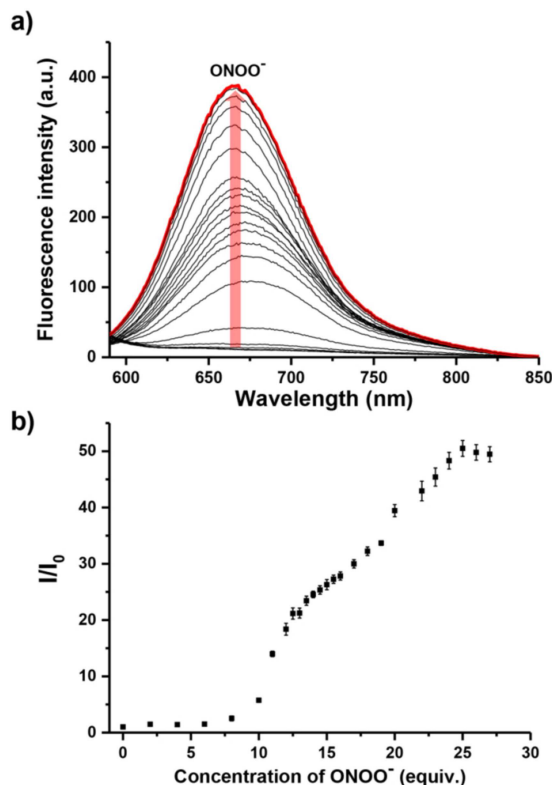


Figure 1. (a) Fluorescence spectra of DCM-Bpin (10 μM) with addition of ONOO $^-$ (from 0 to 27 equiv.) in PBS buffer solution (10 mM, containing 5% DMSO, pH = 7.40). The red line shows the highest intensity after addition of ONOO $^-$ (25 equiv.); (b) Fluorescence intensity changes (I/I_0) of probe DCM-Bpin (10 μM) with addition of ONOO $^-$ (from 0 to 27 equiv.) in PBS buffer solution (10 mM containing 5% DMSO, pH = 7.40) after 5 min. $\lambda_{\text{ex}} = 560 \text{ nm}$ / $\lambda_{\text{em}} = 667 \text{ nm}$. Slit widths: ex = 10 nm, em = 20 nm.

DMSO was required to improve solubility). As shown in Figure S1, the maximum absorption of DCM-Bpin at 434 nm shifted to 530 nm with the addition of ONOO $^-$. As expected, DCM-Bpin was initially non-fluorescent, and upon addition of ONOO $^-$ (0–27 equiv.) a “turn-on” fluorescence response of up to 50-fold was observed at 667 nm using an excitation wavelength of 560 nm (Figure 1).

Subsequently, we compared the response of DCM-Bpin to ONOO $^-$ over other reactive oxygen species (ROS), including ClO $^-$, $\cdot\text{OH}$, O $_2^{\cdot-}$, $^1\text{O}_2$, ROO \cdot , H $_2\text{O}_2$ in PBS buffer (10 mM, pH 7.40). As shown in Figures 2 and S2, only the addition of ONOO $^-$ to the probe led to any distinct optical spectral changes, with no “turn-on” response observed for any of the other ROS species. This indicated that DCM-Bpin has highly selective optical detection for ONOO $^-$.

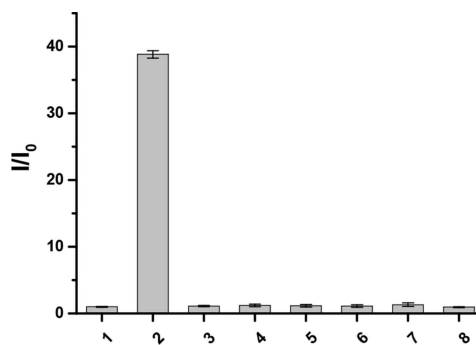


Figure 2. Selectivity bar chart of probe DCM-Bpin (10 μM) with addition of various ROS in PBS buffer solution. 1. Blank, 2. ONOO $^-$ (200 μM), 3. $\cdot\text{OH}$ (500 μM), 4. O $_2^{\cdot-}$ (500 μM), 5. $^1\text{O}_2$ (500 μM), 6. H $_2\text{O}_2$ (500 μM), 7. ROO \cdot (500 μM), 8. ClO $^-$ (500 μM). ONOO $^-$ were measured after 5 min, other ROS were measured after 1 h. Error bar represents s.d. $\lambda_{\text{ex}} = 560 \text{ nm}$ / $\lambda_{\text{em}} = 667 \text{ nm}$. Slit widths: ex = 10 nm, em = 20 nm.

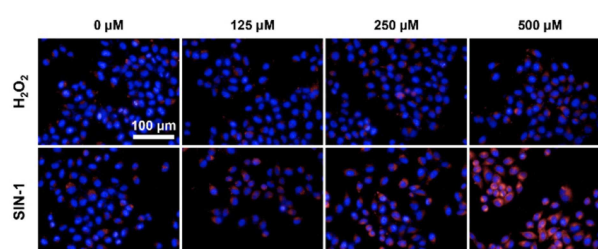


Figure 3. Fluorescence imaging of HeLa cells with DCM-Bpin (20 μM) in the absence or presence of H $_2\text{O}_2$ or SIN-1. Excitation channel = 560–580 nm, emission channel = 650–760 nm. Cell nuclei were stained with Hoechst 33342.

Having determined the selectivity of DCM-Bpin, we evaluated its ability to image exogenous ONOO $^-$ at the cellular level using live HeLa cells. As shown in Figure 3, DCM-Bpin exhibited a good “turn on” response with the addition of peroxynitrite donor SIN-1.^[21] No fluorescent response was observed in the presence of up to 25 equivalents of H $_2\text{O}_2$, confirming the high selectivity of the DCM-Bpin system to ONOO $^-$ over other ROS species. Moreover, MTS cell proliferation assays indicated that DCM-Bpin was not toxic towards HeLa cells (Figure S5).

In conclusion, DCM-Bpin, a near-infrared fluorescent probe, was shown to be suitable for the detection of ONOO $^-$. This probe not only exhibits an excellent “turn-on” response when exposed to peroxynitrite, it also exhibits high selectivity towards ONOO $^-$ over other common reactive oxygen species. DCM-Bpin is biocompatible and displays good sensitivity and selectivity towards ONOO $^-$ in HeLa cells.

Experimental Section

General Methods

All starting materials and reagents were purchased from Sigma Aldrich, Alfa Aesar, Fluorochem, or Acros Organics, and used as received without any further purification. Unless otherwise stated, all solvents used were reagent grade and were used without distillation. All water was distilled. Thin-layer chromatography was

performed by using commercially available Fluorochem aluminum-backed plates coated with a layer of silica gel (60 Å) with fluorescent indicator UV254. These plates were visualized by using ultraviolet light with a wavelength of either 254 or 365 nm. Silica gel column chromatography was carried out by using Sigma Aldrich 60 Å silica gel (200–400 mesh).

All NMR spectra were obtained using an Agilent ProPulse 500 with all spectra recorded in chloroform-*d*. LC–MS analyses were performed using an Agilent QTOF 6545 with Jetstream ESI spray source coupled to an Agilent 1260 Infinity II Quat pump HPLC with 1260 autosampler, column oven compartment and variable wavelength detector (VWD). All pH measurements taken during fluorescence/absorption experiments were recorded on a Hanna Instruments HI 9321 microprocessor pH meter, which was routinely calibrated by using Fisher Chemicals standard buffer solutions (pH 4.0: phthalate; 7.0: phosphate; 10.0: borate). UV-Vis measurements were performed on an Agilent Cary 60 UV-Vis Spectrophotometer, utilizing a Hellma silica (quartz) cuvette with a 10 mm path length (provides photometry with two windows). Fluorescence study was performed an Agilent Cary Eclipse Fluorescence Spectrophotometer.

Full synthetic procedures, characterisation data, and fluorescence analysis protocols can be found in the Supporting Information.

Acknowledgements

LW wishes to thank China Scholarship Council and the University of Bath for supporting his PhD work in the UK. We would like to thank the EPSRC and the University of Bath for funding. TDJ wishes to thank the Royal Society for a Wolfson Research Merit Award. XPH thanks the National Natural Science Foundation of China (21722801 and 21572058), the Programme of Introducing Talents of Discipline to Universities (B16017) and the Shanghai Rising-Star Program (16QA1401400). RRG wishes to thank the EPSRC Centre for Doctoral Training in Catalysis (EP/L016443/1). PT thanks the Development and Promotion of Science and Technology Talents Project (DPST) for a Royal Government of Thailand Scholarship. NMR characterisation facilities were provided by the Material and Chemical Characterisation Facility (MC2) at the University of Bath (<http://go.bath.ac.uk/mc2>). The EPSRC UK National Mass Spectrometry Facility at Swansea University is thanked for analyses.

Conflict of Interest

The authors declare no conflict of interest.

Keywords: near-infrared · fluorescence · boronate · peroxyxynitrite · probe

- [1] P. Pacher, J. S. Beckman, L. Liaudet, *Physiol. Rev.* **2007**, *87*, 315–424.
- [2] A. Weidinger, A. V. Kozlov, *Biomolecules* **2015**, *5*, 472–484.
- [3] R. Radi, *Proc. Natl. Acad. Sci. USA* **2018**, *115*, 5839–5848.
- [4] O. I. Aruoma, *J. Am. Oil Chem. Soc.* **1998**, *75*, 199–212.
- [5] P. Poprac, K. Jomova, M. Simunkova, V. Kollar, C. J. Rhodes, M. Valko, *Trends Pharmacol. Sci.* **2017**, *38*, 592–607.
- [6] H. R. Kermis, Y. Kostov, P. Harms, G. Rao, *Biotechnol. Prog.* **2002**, *18*, 1047–1053.
- [7] L. Wu, A. C. Sedgwick, X. Sun, S. D. Bull, X.-P. He, T. D. James, *Acc. Chem. Res.* **2019**, *52*, 2582–2597.
- [8] J. Chan, S. C. Dodani, C. J. Chang, *Nat. Chem.* **2012**, *4*, 973–984.
- [9] D. Wu, A. C. Sedgwick, T. Gunnlaugsson, E. U. Akkaya, J. Yoon, T. D. James, *Chem. Soc. Rev.* **2017**, *46*, 7105–7123.
- [10] L. Yuan, W. Lin, K. Zheng, L. He, W. Huang, *Chem. Soc. Rev.* **2013**, *42*, 622–661.
- [11] R. Weissleder, *Nat. Biotechnol.* **2001**, *19*, 316–317.
- [12] X. Zhang, L. Zhang, W.-W. Ma, Y. Zhou, Z.-N. Lu, S. Xu, *Front. Chem.* **2019**, DOI:10.3389/fchem.2019.00032.
- [13] J. Ning, T. Liu, P. Dong, W. Wang, G. Ge, B. Wang, Z. Yu, L. Shi, X. Tian, X. Huo, L. Feng, C. Wang, C. Sun, J. Cui, T. D. James, X. Ma, *J. Am. Chem. Soc.* **2019**, *141*, 1126–1134.
- [14] E. A. Owens, M. Henary, G. El Fakhri, H. S. Choi, *Acc. Chem. Res.* **2016**, *49*, 1731–1740.
- [15] A. T. Wrobel, T. C. Johnstone, A. Deliz Liang, S. J. Lippard, P. Rivera-Fuentes, *J. Am. Chem. Soc.* **2014**, *136*, 4697–4705.
- [16] X. Wu, X. Sun, Z. Guo, J. Tang, Y. Shen, T. D. James, H. Tian, W. Zhu, *J. Am. Chem. Soc.* **2014**, *136*, 3579–3588.
- [17] N. Karton-Lifshin, L. Albertazzi, M. Bendikov, P. S. Baran, D. Shabat, *J. Am. Chem. Soc.* **2012**, *134*, 20412–20420.
- [18] H. Wang, Y. Li, M. Yang, P. Wang, Y. Gu, *ACS Appl. Mater. Interfaces* **2019**, *11*, 7441–7449.
- [19] A. Sikora, J. Zielonka, M. Lopez, J. Joseph, B. Kalyanaraman, *Free Radical Biol. Med.* **2009**, *47*, 1401–1407.
- [20] W. Sun, J. Fan, C. Hu, J. Cao, H. Zhang, X. Xiong, J. Wang, S. Cui, S. Sun, X. Peng, *Chem. Commun.* **2013**, *49*, 3890–3892.
- [21] F. J. Martin-Romero, Y. Gutiérrez-Martin, F. Henao, C. Gutiérrez-Merino, *J. Fluoresc.* **2004**, *14*, 17–23.

Manuscript received: October 3, 2019

Revised manuscript received: November 8, 2019