

in the context of this work and other respiratory diseases (10)? Graham Hall and Charlie Irvin recently challenged the profession by asking, “As health professionals, do we have an obligation to reflect on what the actual pathophysiology for a specific lung disease is and considering this, look at different physiological outcomes beyond FEV<sub>1</sub>?” (15). Indeed, within the realm of lung transplantation, should we call time on our obedience to a test that is “too little, too late”? Being cognizant of the devastating potential consequences for the transplanted patient, we need to embrace and incorporate a more physiologically relevant and structurally accurate noninvasive test to detect the site of pathology early, to finally unleash the noise of the silent zone. ■

**Author disclosures** are available with the text of this article at [www.atsjournals.org](http://www.atsjournals.org).

Omar S. Usmani, M.B. B.S., Ph.D.  
National Heart and Lung Institute  
Imperial College London  
London, United Kingdom

ORCID ID: 0000-0002-4367-254X (O.S.U.).

## References

- Chambers DC, Yusen RD, Cherikh WS, Goldfarb SB, Kucheryavaya AY, Khusch K, *et al.*; International Society for Heart and Lung Transplantation. The registry of the International Society for Heart and Lung Transplantation: thirty-fourth adult lung and heart-lung transplantation report-2017; focus theme: allograft ischemic time. *J Heart Lung Transplant* 2017;36:1047–1059.
- Roden AC, Aisner DL, Allen TC, Aubry MC, Barrios RJ, Beasley MB, *et al.* Diagnosis of acute cellular rejection and antibody-mediated rejection on lung transplant biopsies: a perspective from members of the Pulmonary Pathology Society. *Arch Pathol Lab Med* 2017;141:437–444.
- Usmani OS, Biddiscombe MF, Barnes PJ. Regional lung deposition and bronchodilator response as a function of beta2-agonist particle size. *Am J Respir Crit Care Med* 2005;172:1497–1504.
- Hogg JC, Macklem PT, Thurlbeck WM. Site and nature of airway obstruction in chronic obstructive lung disease. *N Engl J Med* 1968;278:1355–1360.
- Benzimra M. Surveillance bronchoscopy: is it still relevant? *Semin Respir Crit Care Med* 2018;39:219–226.
- McNulty W, Usmani OS. Techniques of assessing small airways dysfunction. *Eur Clin Respir J* 2014;1:10.3402/ecrj.v1.25898.
- Estenne M, Van Muylem A, Knoop C, Antoine M. Detection of obliterative bronchiolitis after lung transplantation by indexes of ventilation distribution. *Am J Respir Crit Care Med* 2000;162:1047–1051.
- Thompson BR, Hodgson YM, Kotsimbos T, Liakakos P, Ellis MJ, Snell GI, *et al.* Bronchiolitis obliterans syndrome leads to a functional deterioration of the acinus post lung transplant. *Thorax* 2014;69:487–488.
- Reynaud-Gaubert M, Thomas P, Badier M, Cau P, Giudicelli R, Fuentes P. Early detection of airway involvement in obliterative bronchiolitis after lung transplantation: functional and bronchoalveolar lavage cell findings. *Am J Respir Crit Care Med* 2000;161:1924–1929.
- Usmani OS, Barnes PJ. Assessing and treating small airways disease in asthma and chronic obstructive pulmonary disease. *Ann Med* 2012;44:146–156.
- Foy BH, Soares M, Bordas R, Richardson M, Bell A, Singapuri A, *et al.* Lung computational models and the role of the small airways in asthma. *Am J Respir Crit Care Med* 2019;200:982–991.
- Cho E, Wu JKY, Birriel DC, Matelski J, Nadj R, DeHaas E, *et al.* Airway oscillometry detects spirometric-silent episodes of acute cellular rejection. *Am J Respir Crit Care Med* 2020;201:1536–1544.
- Agusti A, Calverley PM, Celli B, Coxson HO, Edwards LD, Lomas DA, *et al.*; Evaluation of COPD Longitudinally to Identify Predictive Surrogate Endpoints (ECLIPSE) investigators. Characterisation of COPD heterogeneity in the ECLIPSE cohort. *Respir Res* 2010;11:122.
- Hutchinson J. On the capacity of the lungs, and on the respiratory functions, with a view of establishing a precise and easy method of detecting disease by the spirometer. *Med Chir Trans* 1846;29:137–252.
- Hall GL, Irvin CG. Using lung function measurements to greater advantage in patients with lung disease: which test and when? *Respirology* 2014;19:780–781.

Copyright © 2020 by the American Thoracic Society



## suPAR Surprises as a Biomarker of Invasive Outcomes in Pleural Infection

Pleural infection is a major healthcare burden worldwide. Adhesions and loculations, often present in pleural infection, have fascinated pulmonologists for centuries. Their pathogenesis and best management are still debated.

The fibrinolytic pathway has been the subject of active research, and derangements of the local fibrinolytic system can influence the pathogenesis of pleural organization and fibrotic repair, as reviewed

elsewhere (1). The suPAR (soluble urokinase-type plasminogen activator receptor) represents a potential clinical application of the cumulative knowledge gained to date. In this issue of the *Journal* (p. 1545–1553), Arnold and colleagues (2) explore the measurement of suPAR as a new biomarker in pleural infection. Their findings merit further investigation.

suPAR occurs in biologic fluids, including plasma, urine, and pleural fluids, and is proteolytically cleaved from the surface of cells bearing the uPAR (urokinase-type plasminogen activator receptor), which regulates cellular proteolysis, viability, movement, and proliferation (3). It is also possible that an alternatively spliced variant of suPAR may contribute to the suPAR concentrations seen in pleural fluids, as has previously been demonstrated in cancer cell lines (4). suPAR concentrations increase in

Ⓜ This article is open access and distributed under the terms of the Creative Commons Attribution Non-Commercial No Derivatives License 4.0 (<http://creativecommons.org/licenses/by-nc-nd/4.0/>). For commercial usage and reprints, please contact Diane Gern ([dgern@thoracic.org](mailto:dgern@thoracic.org)).

Originally Published in Press as DOI: 10.1164/rccm.202003-0525ED on March 16, 2020

inflammatory conditions, including bacterial, viral, and mycobacterial infections and cancer (3). Pleural fluid suPAR concentrations have been shown to discriminate between effusions attributable to congestive heart failure of inflammatory or malignant causes (5, 6). Traditional biomarkers such as pleural fluid pH, glucose, and lactate dehydrogenase often influence decisions on chest tube placement in the setting of pleural infection, but they do not predict the need for intrapleural fibrinolytic therapy or surgery.

Arnold and colleagues found that pleural fluid suPAR concentrations were increased in 37 patients with pleural infections versus 47 control subjects who had either transudative effusions or malignancy (2). The pleural fluids were removed at presentation and archived, after which suPAR concentrations were measured using a commercially available assay. The assay detects suPAR (consisting of domains I–III) and the suPAR (or uPAR) domains II and III, so that the readout of the assay reflects intact suPAR and a fragment that may be present in solution. In addition, the assay may also detect the membrane-bound cell receptor uPAR because microvesicles or cellular fragments may be present in pleural fluids.

Pleural fluid suPAR concentrations were significantly elevated in pleural fluids obtained from patients with loculated pleural infection versus those with infection who did not have progression to loculation. Although traditional biomarker trends were similar, pleural fluid suPAR concentrations predicted subsequent chest tube insertion more accurately than did pleural fluid pH. Pleural fluid suPAR also more accurately predicted the need for more invasive management, as assessed by referral for intrapleural fibrinolytic therapy or thoracic surgery.

Clinician behavior in deciding on chest tube drainage, and more so in employing fibrinolytic therapy and surgery, is notoriously variable among or even within medical centers. It is therefore most intriguing that pleural fluid suPAR concentrations are able to predict these rather difficult clinical decisions. Can this be chance, or is suPAR causally related to more severe (or worsening of) parapneumonic effusions? To make a firm statement about the clinical relevance of suPAR will require a prospective validation cohort from different healthcare systems with assessment against clinical data relating to patient progress (e.g., ongoing fever, leukocyte counts, and C-reactive protein) and predetermined criteria for intrapleural fibrinolytic therapy and/or surgery.

Is it possible that suPAR is a marker of pleural loculation not specific to pleural infection? Pleural loculation is usually an indirect reflection of the degree of inflammation and not solely found in parapneumonic effusions. The authors provide data showing that similar results were found within an albeit small malignant effusion cohort, supporting this hypothesis. The findings suggest that high-grade inflammation that progresses to intrapleural organization results in elevations in pleural fluid suPAR that in turn can predict the need for interventions to expedite drainage in patients with pleural infection.

Loculations, however, are not good predictors of ability to drain pleural effusions; they are often (but not necessarily correctly) the reason why chest tubes are inserted and intrapleural therapy and/or surgery is initiated. Currently, there is no consensus about the definition of “loculation,” let alone a validated quantification method of loculation in the literature. In daily practice, a loculated effusion can range from a few septations to extensive

“honeycombing.” In the study by Arnold and colleagues, loculation was graded as “yes” or “no” without a preset definition, often by junior staff. This highlights the need for a way to evaluate the severity of loculation within the pleural cavity, and suPAR may serve such a role. If so, the next step will be to determine what outcome suPAR (or degree of loculation) accurately predicts various pleural diseases extending beyond infection.

The results of the work of Arnold and colleagues add credence to the possibility that suPAR may contribute to the regulation of pleural loculation in addition to its role as a biomarker. suPAR is known to bind scuPA (single-chain urokinase plasminogen activator), which increases its ability to exhibit plasminogen activator activity (7). suPAR can also bind the more active two-chain urokinase (tcuPA) that derives from plasmin-mediated cleavage of scuPA and thus could localize plasminogen activator activity within pleural fluids. These effects could support fibrinolysis in the presence of low concentrations of PAI-1 (plasminogen activator inhibitor) that may occur after intrapleural administration of fibrinolytics, as occurs when scuPA is administered intrapleurally (8). However, suPAR-bound single- or two-chain uPA is susceptible to PAI-1, and increments of PAI-1 are generally seen in pleural loculation in pleural infection (1, 9). Thus, the role of suPAR in the regulation of intrapleural fibrinolytic therapy remains unclear and is worthy of further investigation.

Apart from suPAR, other new inflammation- and fibrinolysis-related biomarkers of the outcomes of pleural infection may soon emerge. For instance, PAI-1 and its activity have likewise been strongly implicated in the pathogenesis of pleural injury outcomes (1, 10). The ability of baseline pleural fluids to support fibrinolytic activity, called the “fibrinolytic potential,” is another candidate biomarker (11). The clinical implication of these markers as predictors of treatment or prognosis provides a new and exciting area of pleural disease research. ■

---

**Author disclosures** are available with the text of this article at [www.atsjournals.org](http://www.atsjournals.org).

Steven Idell, M.D., Ph.D.  
*Department of Cellular and Molecular Biology*  
*University of Texas Health Science Center at Tyler*  
*Tyler, Texas*

Y. C. Gary Lee, M.B. Ch.B., Ph.D.  
*Sir Charles Gairdner Hospital and University of Western Australia*  
*Perth, Western Australia, Australia*

ORCID IDs: 0000-0003-4389-2152 (S.I.);  
 0000-0002-0036-511X (Y.C.G.L.).

---

## References

1. Komissarov AA, Rahman N, Lee YCG, Florova G, Shetty S, Idell R, *et al*. Fibrin turnover and pleural organization: bench to bedside. *Am J Physiol Lung Cell Mol Physiol* 2018;314:L757–L768.
2. Arnold DT, Hamilton FW, Elvers KT, Frankland SW, Zahan-Evans N, Patole S, *et al*. Pleural fluid suPAR levels predict the need for invasive management in parapneumonic effusions. *Am J Respir Crit Care Med* 2020;201:1545–1553.
3. Thunø M, Macho B, Eugen-Olsen J. suPAR: the molecular crystal ball. *Dis Markers* 2009;27:157–172.

4. Grismayer B, Sato S, Kopitz C, Ries C, Soelch S, Schmitt M, *et al*. Overexpression of the urokinase receptor splice variant uPAR-del4/5 in breast cancer cells affects cell adhesion and invasion in a dose-dependent manner and modulates transcription of tumor-associated genes. *Biol Chem* 2012;393:1449–1455.
5. Matzkies LM, Raggam RB, Flick H, Rabensteiner J, Feierl G, Hoenigl M, *et al*. Prognostic and diagnostic potential of suPAR levels in pleural effusion. *J Infect* 2017;75:465–467.
6. Ozsu S, Oztuna F, Mentese A, Abul Y, Ozlu T. Diagnostic value of suPAR in differentiating noncardiac pleural effusions from cardiac pleural effusions. *Clin Respir J* 2016;10:61–66.
7. Higazi A, Cohen RL, Henkin J, Kniss D, Schwartz BS, Cines DB. Enhancement of the enzymatic activity of single-chain urokinase plasminogen activator by soluble urokinase receptor. *J Biol Chem* 1995;270:17375–17380.
8. Beckert L, Brockway B, Simpson G, Southcott AM, Lee YCG, Rahman N, *et al*. Phase 1 trial of intrapleural LTI-01; single chain urokinase in complicated parapneumonic effusions or empyema. *JCI Insight* 2019; 5:e127470.
9. Komissarov AA, Florova G, Azghani AO, Buchanan A, Boren J, Allen T, *et al*. Dose dependency of outcomes of intrapleural fibrinolytic therapy in new rabbit empyema models. *Am J Physiol Lung Cell Mol Physiol* 2016;311:L389–L399.
10. Florova G, Azghani AO, Karandashova S, Schaefer C, Yarovoi SV, Declerck PJ, *et al*. Targeting plasminogen activator inhibitor-1 in tetracycline-induced pleural injury in rabbits. *Am J Physiol Lung Cell Mol Physiol* 2018;314:L54–L68.
11. Idell S, Florova G, Shetty S, Tucker T, Idell R, Koenig K, *et al*. Precision-guided, personalized intrapleural fibrinolytic therapy for empyema and complicated parapneumonic pleural effusions: the case for the fibrinolytic potential. *Clin Pulm Med* 2017;24: 163–169.

Copyright © 2020 by the American Thoracic Society