# The rise of mechanical metamaterials: Auxetic constructs for skin wound healing

Journal of Tissue Engineering Volume 14: 1–16 © The Author(s) 2023 Article reuse guidelines: sagepub.com/journals-permissions DOI: 10.1177/20417314231177838 journals.sagepub.com/home/tej



Óscar Lecina-Tejero<sup>1</sup>, María Ángeles Pérez<sup>1,2</sup>, Elena García-Gareta<sup>1,2,3</sup> and Carlos Borau<sup>1,4</sup>

## Abstract

Auxetic materials are known for their unique ability to expand/contract in multiple directions when stretched/compressed. In other words, they exhibit a negative Poisson's ratio, which is usually positive for most of materials. This behavior appears in some biological tissues such as human skin, where it promotes wound healing by providing an enhanced mechanical support and facilitating cell migration. Skin tissue engineering has been a growing research topic in recent years, largely thanks to the rapid development of 3D printing techniques and technologies. The combination of computational studies with rapid manufacturing and tailored designs presents a huge potential for the future of personalized medicine. Overall, this review article provides a comprehensive overview of the current state of research on auxetic constructs for skin healing applications, highlighting the potential of auxetics as a promising treatment option for skin wounds. The article also identifies gaps in the current knowledge and suggests areas for future research. In particular, we discuss the designs, materials, manufacturing techniques, and also the computational and experimental studies on this topic.

## **Keywords**

Skin tissue engineering, auxetic materials, additive manufacturing

Date received: 21 February 2023; accepted: 6 May 2023

## Introduction

The importance of skin for human health is unquestionable, as it is the human body's first defense against physical and biological threats. Acting as a mechanical barrier to the outside environment, preventing the entry of pathogens and micro-organisms into the body,<sup>1</sup> thermoregulation and self-healing<sup>2</sup> are among its main functions.

This complex and large organ has two layers called epidermis and dermis, which are populated by different cell types and have different functional, mechanical, and biological characteristics.<sup>3</sup>

The epidermis is a thin, poorly vascularized layer located on the outermost part of the skin.<sup>4</sup> It is composed of keratinocytes that proliferate outwards and, as they differentiate, fill with keratin, creating a layer of dead cells that provides protection against external agents and prevents the loss of water and other substances. The dermis is the inner of the two layers of the skin. It is a thick, highly vascularized layer of connective tissue composed of fibroblasts and extracellular matrix (ECM) of collagen, elastin and glycosaminoglycans among other components.<sup>5</sup> This dermal ECM is the major contributor to the mechanical properties of the skin,<sup>6</sup> where the combination of stiff collagen fibers and flexible elastin fibers, as well as their cross-linking, results in an interesting mechanical behavior consisting of a characteristic "J-shaped" stress-strain curve<sup>7</sup> together with an auxetic behavior.<sup>8,9</sup> Skin mechanical properties may vary depending on several factors<sup>10,11</sup>

 <sup>2</sup>Aragon Institute for Health Research (IIS Aragon), Miguel Servet University Hospital, 50009 Zaragoza, Aragon, Spain
 <sup>3</sup>Division of Biomaterials & Tissue Engineering, UCL Eastman Dental Institute, University College London, London, UK
 <sup>4</sup>Centro Universitario de la Defensa de Zaragoza, Zaragoza, 50090, Spain

#### **Corresponding author:**

Carlos Borau, Multiscale in Mechanical and Biological Engineering, Aragon Institute of Engineering Research (I3A), University of Zaragoza, C/María de Luna s/n, Zaragoza, Aragon 50018, Spain. Email: cborau@unizar.es

Creative Commons Non Commercial CC BY-NC: This article is distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 License (https://creativecommons.org/licenses/by-nc/4.0/) which permits non-commercial use, reproduction and distribution of the work without further permission provided the original work is attributed as specified on the SAGE and Open Access pages (https://us.sagepub.com/en-us/nam/open-access-at-sage).

<sup>&</sup>lt;sup>1</sup>Multiscale in Mechanical and Biological Engineering, Aragon Institute of Engineering Research (I3A), University of Zaragoza, Zaragoza, Aragon, Spain

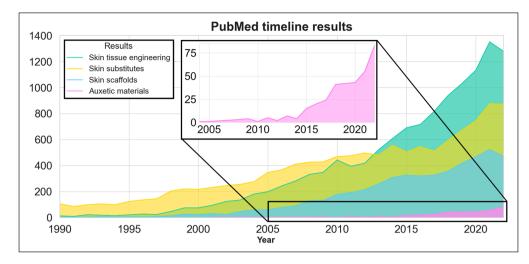


Figure 1. PubMed search results from skin tissue engineering related topics, showing the increasing interest and development in recent years.

that may be local, such as the particular location of the body being measured, its current care status or the direction of the sample fibers with respect to Langer's lines,<sup>12</sup> or global, such as the patient's age or gender.<sup>13,14</sup>

Mechanical alterations of the skin, such as those resulting from wound formation, give rise to processes in which physiological functions are also affected due to loss of tissue integrity and functionality, leading in extreme cases to disabilities or even death.<sup>15</sup> The importance of these wounds depends on their size, where small or superficial wounds quickly heal due to the high regenerative capacity of the skin,<sup>16</sup> while larger wounds (area >4 cm),<sup>17</sup> such as those caused by burn damage or acute trauma, may require more healing time and even additional surgical interventions, sometimes even requiring skin substitutes to achieve good repair and regeneration.<sup>18</sup>

In this context, many different skin substitutes have been developed in the last decades.<sup>3,6,19–23</sup> The most common and widely used solution is transplantation,<sup>20,24</sup> where the affected area is covered with tissue of the same or similar type as the missing tissue, taken from another part of the patient's body or from a donor, whether of the same species or not, known as autografts, allografts, and xenografts, respectively.

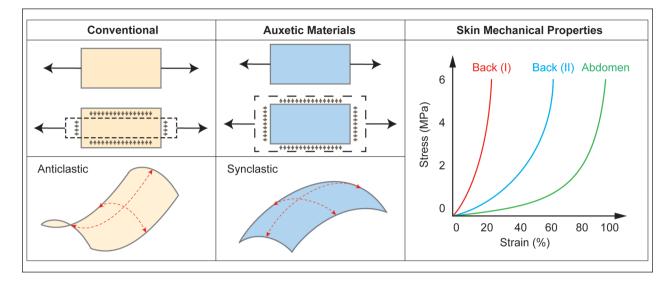
The graft transplantation method, although effective in healing the wound and giving rise to a tissue quite similar to the original one, requires a separate surgical intervention to remove the tissue to be transplanted, with all the risk that this entails, in addition to leaving a scar in another part of the body. Furthermore, it is also a procedure limited by the availability of tissue to extract,<sup>21</sup> so that in cases of exceptionally large area skin wounds, these solutions become unfeasible. In fact, besides immunoreactions,<sup>25,26</sup> there is another drawback to the use of human skin grafts that is the non-zero risk of disease transmission, as there has been at least one case of HIV transmission between donor and recipient.<sup>27</sup>

To overcome these problems, tissue engineering research has been focusing on the development of scaffolds or tissue substitutes that act as templates for cell infiltration and subsequent regeneration of damaged skin, culminating in a plethora of skin substitutes,<sup>3,28,29</sup> some of them showing promising techniques and results,<sup>30</sup> and even reaching the market and clinic.<sup>31</sup> As an example, searching on PubMed for related keywords such as "skin tissue engineering," "skin substitutes," and "skin scaf-folds" from the last few decades yields a substantial number of results, indicating an increasing growth in this area during that time period (as shown in Figure 1). Similarly, a search for "auxetic materials" demonstrates an exponential rise in results from recent years, highlighting the increased interest in auxetic behavior research.

Moreover, the search of "auxetic materials" also reveals an exponential increase in the results from the last years, bringing to the board the rise of interest that auxetic behavior research has been experiencing.

Further research is still needed, and expected, as currently available materials present disadvantages such as high costs, issues of take and integration, and non-satisfactory esthetic outcomes.<sup>32</sup> Therefore, new strategies are needed in the design of novel and effective skin substitutes.

In this context, auxetic materials have gained attention for their potential use in skin wound healing due to their unique mechanical properties. Moreover, skin itself shows an auxetic behavior, as previously mentioned. As shown in Figure 2, auxetic structures can stretch/contract in multiple directions when they are subjected to external forces, allowing them to conform to the shape of the wound bed and provide good coverage and protection to the damaged tissue. In the context of stretching, auxetic structures exhibit mechanical behavior similar to that of skin. Both materials can withstand high levels of strain before experiencing stiffening at a certain point.



**Figure 2.** Left panels: schematic diagram comparing conventional and auxetic material behaviors. In addition to their negative Poisson's ratio, auxetic materials exhibit synclastic deformation which allows for more even distribution of pressure across the surface, potentially benefiting wound healing and reducing the risk of pressure sores in skin healing applications. Right panel: skin stress-strain J-shaped relationship for different parts of the body (Adapted from Jang et al.<sup>34</sup>).

Additionally, auxetic materials have exceptional out-ofplane bending behavior, allowing them to conform to synclastic surfaces and exert stable amounts of pressure along their shape when bent.<sup>33</sup>

This property may be very useful in wound healing applications, especially in areas such as joints, where the wound shape may be affected by the movement. The bending behavior of the scaffold may help maintain contact between the scaffold and the wound in these sites. This is an important function for skin repair, as it may help to prevent further damage and provide a suitable environment for cell proliferation and tissue remodeling. The porous structure also enables cell migration and infiltration,<sup>35</sup> which could enhance the formation of new tissue. Additionally, auxetic structures are known to have good mechanical strength and stability, which could help supporting the wound area during the healing process, preventing the scaffold from collapsing or deforming under the loads applied by the surrounding tissue. These properties, together with the developments of 3D printing technologies that allow both flexibility and a tight control of microgeometries, are rising auxetic scaffolds as a promising option for skin repair.

The aim of this review is to provide an overview of the current state of the art on research on auxetic constructs for skin tissue engineering. To this end, the review summarizes the auxetic designs that have been studied or proposed for skin wound healing, and outlines the manufacturing techniques applied to produce them, as well as the computational and experimental studies that have been conducted to evaluate their performance, from a critical point of view in order to understand their potential and limitations.

## **Auxetic materials**

## Auxetic properties and biological tissues

Auxeticity is a mechanical characteristic associated to materials exhibiting a negative Poisson's ratio, which means that they grow transversely when stretched longitudinally and vice versa, they shrink in the transverse direction when compressed longitudinally.<sup>36</sup> Materials exhibiting this behavior are included in a recently considered material group called "mechanical metamaterials," which englobes materials that have unusual mechanical properties, not commonly found in natural materials, due to their particular micro-scale structure rather than the properties of their constituents. These materials can be engineered to have particular properties such as negative stiffness,<sup>37</sup> negative Poisson's ratios, which are the auxetic materials, or negative mass density.<sup>38</sup>

However, despite this classification, auxetic properties have actually been found in natural materials for years. Actually, there are a lot of auxetic crystalline materials<sup>39</sup> and auxetic foams,<sup>40</sup> as their inner pore-based structure can develop an auxetic configuration. Even there are some biological tissues that have also been shown to develop auxetic behavior, as it has been reported in cow teat,<sup>8</sup> cat,<sup>7</sup> pig,<sup>9,41</sup> and salamander<sup>42</sup> skins, arteries,<sup>43,44</sup> tendons,<sup>45</sup> cancellous bone,<sup>46</sup> embryonic epithelia,<sup>47</sup> cornea,<sup>48</sup> and more recently, in marine sponges.<sup>49</sup>

These biological tissues develop the auxetic behavior mainly because of the structural organization and crosslinking of their ECM fibers, demonstrating the importance of ECM structure in the mechanical behavior of the tissue, and also leading to the idea of using auxetic scaffolds to mimic the mechanical properties of the ECM in order to provide better support for the implanted cells.

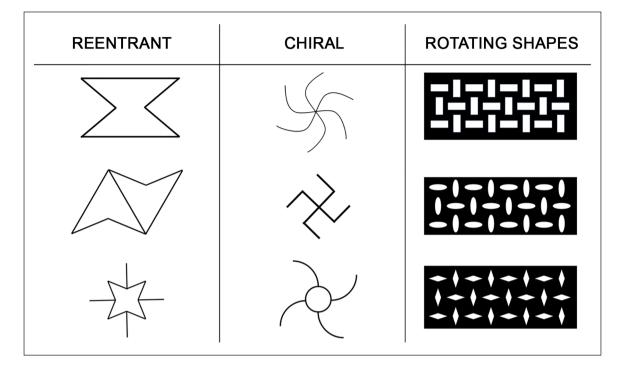


Figure 3. Examples of 2D geometry patterns with auxeticity development.

There are some reviews<sup>50,51</sup> on the topic of auxetic scaffolds for general tissue engineering, in which the collected studies show some encouraging results on cell migration, proliferation and differentiation enhancement due to auxetic scaffolds, in addition to studying the cyto-compatibility and the survival viability of different cell types when cultured in this kind of particular mechanical environments.

Nowadays, there are various paths to follow in order to achieve auxeticity in tissue engineering constructs. Apart from the auxetic foams already commented,<sup>40</sup> there are three main geometry design techniques based on repeatable basic patterns that are suitable for their application in manufacturing methods and could be useful to study the influence on cell behavior of this mechanical issue: reentrant, chiral, and rotating shape designs (Figure 3).

Reentrant designs consist of common polygons with an inversion in some of their angles, which gives the auxetic behavior to the structure. Controlling these angles and the size of the designs is crucial to set the structural auxetic behavior.

Chiral designs are similar fiber designs that are characterized by having a center, or axis, around which a radial fiber distribution coil. Thus, the coiling degree and fiber amount would be the main parameters to control the auxetic development of the structure.

Rotating shape designs are based on a pattern of cuts made on a membrane that results on a connected-shapes distribution that, when stretched, makes the shapes rotate around each other expanding the total area occupied by the membrane. The shape, size and distribution of the cuts are the main parameters to set the auxetic behavior of these designs.

To obtain reentrant and chiral designs, it is common to 3D print a scaffold by controlling the spatial distribution and orientation of the fibers to draw the 2D or 3D auxetic design. A widely used application of these techniques is to develop fiber reinforced scaffolds, where these fibers, made by biopolymers, enhance the mechanical behavior of other materials, such as hydrogel, gelatin, bio-inks, or tighter and thinner fiber structures. Hence, this reinforcing structure would be added to give physical support to the cell culture, and also affect synergistically the mechanical behavior as reported in some studies.<sup>52–55</sup>

In the case of rotating shape designs, the most common technique is to first manufacture the membrane, which could be made by both stiff materials,<sup>56</sup> as electrospun biopolymers, or soft materials,<sup>57</sup> as hydrogels, and then apply the cut pattern with other techniques such as laser cut, which has high precision and control. When applying this technique, the whole membrane acts as cell support and the mechanical behavior is more influenced by the properties of the constituent material. The existence of the cuts implies that there would be void spaces inside the scaffold when the membrane is stretched, so it is definitely something to consider when designing the scaffold application. More details regarding the manufacturing of this kind of scaffolds are described later on in the *Section 3.2. Fabrication technologies*.

Publication	Material	Fabrication method	Auxetic design	Cell culture	Particular details	Main conclusions
Chansoria et al. <sup>58</sup>	Bilayer patch of GelMA and PEGDA	Digital light processing	Reentrant, chiral, and rotating shape 2D macro designs	3T3 Fibroblasts	Patch design oriented to cover organs and to protect the bottom cell layer	Obtained good cell proliferation, area coverage and adaptation to dynamic organ mechanics
Flamourakis et al. <sup>59</sup>	Photopolymer SZ2080	Multiphoton stereolithography	Reentrant 3D micro designs	NIH-3T3 Fibroblasts	Cell-sized pore auxetic scaffold to study auxetic influence at cell level	Good cell penetration, proliferation, directionality, and scaffold shape adaptation to cell requirements
Jin et al. <sup>60</sup>	PCL fibers	Melt electro- writing	Reentrant 2D multiscale design	Human umbilical vein endothelial cells (HUVECs) and bone marrow stem cells (BMSCs)	Auxetic thick fiber macro design under a thin fiber layer that provides cell support. Intricate pore size scaffold with versatile mechanical properties	Different cell proliferation depending on cell type, solves the problem of biocompatibility and mechanical strength simultaneously

Table 1. Publications about auxetic scaffolds related to skin tissue engineering.

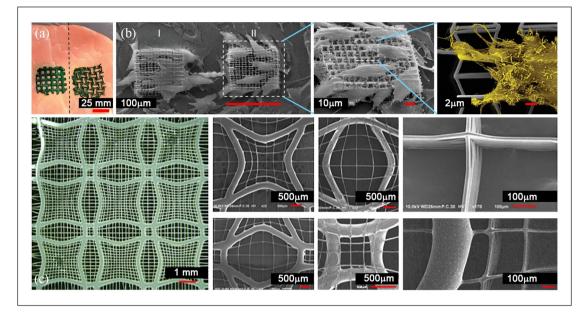
## Auxetic scaffolds in tissue engineering

Despite the existence of various different auxetic scaffolds in the literature, to the best of our knowledge, a specific application of these structures to skin repair has not been reported. Table 1 summarizes the main characteristics of the studies where a link between the auxetic scaffolds and a possible application for skin tissue engineering could be suggested, while Figure 4 shows details of the auxetic constructs developed.

Although these studies greatly differ from each other, all of them show encouraging results about the use of auxetic scaffolds. For example, Chansoria et al.58 developed and studied auxetic hydrogel patches designed for conforming to the complex mechanics of dynamic organs, such as heart, lungs, bladder, or skin. Their patches were composed of two layers obtained by photocuring simultaneously with 2D auxetic macro patterns, in the order of centimeters, both the non-fouling polyethylene glycoldiacrylate (PEGDA) top layer and the gelatin methacryloyl (GelMA) bottom layer, which allowed cell adhesion and proliferation. They applied different reentrant, chiral and rotating shapes designs to their patches and studied and tested them both numerically and experimentally, which they did by performing a parametric study that allowed them to relate the mechanical properties of the patches and the organs, and then select the ones they wanted to test and study their behavior and possible application. In this context, they were able to relate some of these designs with the skin mechanical behavior, but they did not further explore its application.

Flamourakis et al.<sup>59</sup> laser-fabricated small auxetic scaffolds with SZ2080, which is a hybrid organic-inorganic photoresist.<sup>61</sup> The high precision of the two-photonpolymerization technique, which they used to manufacture the scaffolds, allowed them to obtain 3D reentrant auxetic structures in the order of units of microns, which were about the same size of the cells, in order to better study the auxetic influence in cell response. After culturing mouse NIH-3T3 fibroblasts, they found that cells were able to penetrate and proliferate through the scaffold, and they also observed that cells were aligned along the scaffold structure and also adapting the scaffold shape to suit their requirements.

Jin et al.<sup>60</sup> went into the multiscale design of a fibrous poly(caprolactone) (PCL) scaffold composed by an 2D reentrant auxetic macro-structure, with fiber diameters of 400 microns, and a 2D web-like micro-structure that was composed of 10-micron diameter fibers. These multiscale scaffolds were designed not only to have an auxetic mechanical behavior due to their macro-scale design, but also to give physical support to the cells at the microscale. They seeded human umbilical vein endothelial cells (HUVECs) and bone marrow stem cells (BMSCs) which showed different behaviors, where HUVECs proliferated along the direction of the fibers, while BMSCs proliferated by filling the void spaces between the fibers. These scaffolds have an intricate pore size due to the micro-scale design, and their macro-scale design gives them a tunable mechanical behavior, thus it is a versatile way to develop auxetic scaffolds. However, they only studied the cell responses in static conditions, so dynamic testing of these structures would be interesting for observing the mechanobiological influence that auxetic behavior could have in the cells.



**Figure 4.** Details of the auxetic scaffold studies conducted. (a) Bilayer hydrogel non-auxetic and auxetic patches showing its in vivo application in pig lung. (b) Laser-made auxetic scaffolds showing NIH-3T3 fibroblast culture with cell scaffold directionality and attachment. (c) Multiscale auxetic scaffold where both macro and micro designs and fiber sizes are visible. Illustrations adapted with permission from Chansoria et al.,<sup>58</sup> Flamourakis et al.,<sup>59</sup> and Jin et al.<sup>60</sup> respectively.

The potential applications for skin tissue engineering of these publications diverge due to differences in sizes, scales, materials, and study subjects, although they could be adapted. For example, the hydrogel patches<sup>58</sup> could be manufactured in a reduced scale to decrease void spaces within the hydrogel, thus enhancing the coverage for large area wounds. In the same way, the 3D laser-manufactured<sup>59</sup> and the multiscale<sup>60</sup> fiber scaffolds could be an option to improve the effectiveness and mechanical and biological properties of fiber reinforced hydrogel scaffolds that could also be applied in large wounds.

## Computational studies of auxetic materials

Simulations have several advantages over experimental studies among which we can highlight the cost and time efficiency and the ability to perform broad sensitivity analysis at different scales (from micro to macro) which can provide a more complete understanding of the behavior of studied systems. Computational studies on auxetic structures and designs have gained attention in recent years and have been used to predict mechanical behaviors and to optimize the material properties and geometries of the constructs.

For example, Jang et al.<sup>34</sup> did a numerical analysis of auxetic 2D chiral patterns designed for an application as a structural reinforcement for skin-mounted electro-physiological sensors. They ran a parametric study of the geometry and its different combinations, they fabricated some design samples with polymethylmethacrylate (PMMA) by

photolithography and tested them experimentally, validating the numerical results. The mechanical properties measured were analogous to those of collagen and elastin in biological tissues and exhibited three phases of tensile loading: the first phase was characterized by bendingdominated deformation, followed by a second phase where fibers rotated, twisted, and aligned with the direction of stress, and finally, a stretching-dominated deformation mode in the last phase. During the final phase of mechanical loading, the mechanical modulus was observed to increase by several orders of magnitude compared to the initial phase. Additionally, when the auxetic designs were subjected to low strains, they exhibited a negative Poisson's ratio effect in that region. However, this behavior disappeared as the shapes were fully extended. They proposed their constructs to be used as sensors, but both the numerical analysis and the auxetic design could be scaled and the materials made biocompatible to fabricate fiber reinforced scaffolds for skin implantation.

A similar study was conducted by Liu and Zhang<sup>62</sup> where auxetic 2D chiral patterns but with straighter fibers were analyzed numerically. In this case, the parametric study was related to cat skin behavior, and the application was to develop architected cylindrical shells with shape memory effects. Results showed that the chiral patterns had the potential to exhibit isotropic Poisson's ratios ranging from -1 to 1, even over large strains. Furthermore, the developed design methods could identify appropriate geometries to achieve specific Poisson's ratios while also matching the mechanical properties of cat skin. Once

again, the analysis and designs could be adapted to the necessary scale and materials to also be applied in wound healing applications.

Lastly, there are two publications<sup>63,64</sup> where auxetic 2D patterns combining reentrant and rotating shapes designs as cuts in bilayer membranes with different properties simulating the epidermis and dermis layers of the skin were analyzed with a parametric study in order to understand the effect of the auxetic patterns on split thickness skin graft expansion. The results of this study showed that all auxetic graft designs confirmed the negative Poisson's effect. Moreover, when models were subjected to uniaxial strain, the meshing ratios of auxetic grafts exceeded 30, which was significantly higher than traditional grafts with ratios around 3. However, the study also identified some limitations, such as the assumption of skin as an isotropic and elastic material model, or the constant thickness assumption. Despite these limitations, further research and experimental investigations could enhance our understanding and contribute to developing a large skin graft area using a small size donor skin. This is particularly significant in skin transplantation and burn surgery.

In summary, computational studies provide a costeffective and efficient way to analyze and optimize auxetic structures and designs, and to predict their behavior under different conditions, which can greatly assist experimental studies and the overall development of auxetic materials for skin tissue engineering applications. Further research is needed to develop constructs that can be easily integrated with other wound healing therapies and techniques.

# Fabrication of auxetic scaffolds

## Materials

Scaffolds are obviously a critical component in tissue engineering, as they provide the necessary support and microenvironment for cells to grow and differentiate into functional tissue. The materials used for manufacturing scaffolds must have both a combination of mechanical properties that makes them suitable for the corresponding manufacturing technology and also biological properties to not cause an adverse reaction in the body when it comes into contact with living tissue. The ideal scaffold material should be biocompatible, biodegradable, and should support cell attachment, proliferation, and differentiation.

A wide range of materials have been used to fabricate scaffolds for tissue engineering, including natural, synthetic, and composite materials.<sup>65–67</sup> Natural materials, such as collagen, gelatin, and chitosan,<sup>68–73</sup> are biocompatible and biodegradable, but they can be difficult to process and may lack the mechanical strength for some applications. Synthetic materials, such as poly(lactic acid) (PLA), poly(glycolic acid) (PGA), and PCL,<sup>74,75</sup> are easy to process and have well-defined mechanical properties, but they may be less biocompatible. Composite materials, such as

those made from a combination of natural and synthetic materials, can offer the best of both worlds, providing the necessary mechanical properties while also being biocompatible and biodegradable.

Examples of new materials that have been used in scaffold fabrication are hydrogels, which have gained attention for their ability to mimic the mechanical properties of the native ECM.<sup>76–78</sup> Hydrogels can be made from natural or synthetic polymers and can be tailored to mimic the mechanical properties of the tissue it is supposed to replace, which can greatly improve the tissue regeneration process.

The auxetic constructs developed in the studies summarized in Table 1 showcase a diverse range of biomaterials, each with distinct properties and potential applications in skin tissue engineering. In order to provide a better comprehensive understanding of these materials, Table 2 outlines their main properties and composition, among other critical factors that influence their performance in tissue engineering applications.

In conclusion, the choice of materials for scaffold fabrication is a critical aspect in tissue engineering, as it affects the scaffold's ability to be adapted to the particular mechanical properties and microarchitecture of the tissue to replace.

## Fabrication technologies

3D printing techniques are currently among the most promising and viable approach to manufacture functional and custom-fit scaffolds capable of promoting tissue regeneration. Such techniques can help in the challenge of accurately controlling the spatial distribution of pores and structures within the scaffold, which is critical for a proper cell development. Thus, depending on some factors such as the tissue to be replaced, the material utilized or the type of scaffold, there exist multiple ways to manufacture these tissue-engineered constructs.<sup>87</sup>

As the constructs may need specific requirements in their microstructures, not every 3D printing technique is suitable to be used in tissue-engineered scaffolds manufacturing. Thus, Figure 5 gathers details of the different microstructures achieved by some of the most used 3D printing techniques for the fabrication of tissue engineering scaffolds,<sup>88–95</sup> which can be grouped as electro-printing techniques, digital light processing (DLP) techniques, and 3D-bioprinting. Note that the geometries depicted are non-auxetic, as the figure focuses on the achieved scale and fiber organization.

The electro-printing category includes widely used and interesting techniques, such as electrospinning (ES) and melt electro-writing (MEW), which have been extensively studied for scaffold fabrication.<sup>56,89,98,99</sup>

ES, widely described in publications since the end of the last century,<sup>100–102</sup> is a fiber deposition process where an electrical charge is used to extrude a melt polymer solution into thin fibers, which are collected onto a grounded

Biomaterial		Description	<ul> <li>Main Properties</li> <li>Biocompatibility Biodegradability</li> <li>Hydrophobicity Semi-crystallinity Low meting point (~60°C) Elasticity FDA approved</li> </ul>	
Poly-ε-caprolactone (Po	CL) <sup>75,79</sup>	Synthetic polymer: a polyester synthesized by ring-opening polymerization of ε- caprolactone using different catalysts or by 2-methylene-1-3-dioxepane.		
SZ2080 <sup>61,80–83</sup>		Resin: a hybrid organic/inorganic photoresist made of two components: methacryloxypropil trimethoxysylane and zirconium propoxide.	<ul> <li>Biocompatibility</li> <li>Long-term stability</li> <li>Chemical and electrochemical inertia</li> <li>Transparency</li> <li>Photopolymerizable</li> <li>Ultra-low shrinkage during polymerization</li> </ul>	
Gelatin methacryloyl (GelMA) / Polyethylene glycol diacrylate (PEGDA)	GelMA <sup>84,85</sup>	Modified natural hydrogel produced by the reaction of gelatin with methacrylic anhydride, whereby the amino groups on the side chains of gelatin are replaced by methacryloyl groups, thus forming modified gelatin.	<ul> <li>Biocompatibility</li> <li>Biodegradability</li> <li>Photopolymerizable</li> <li>Thermostability</li> <li>Tunable physicochemical properties</li> </ul>	
	PEGDA <sup>83,85,86</sup>	Synthetic hydrogel that is a PEG derivative fabricated through substituting terminal hydroxyl groups of PEG with acrylates.	<ul> <li>Biocompatibility</li> <li>Biodegradability</li> <li>Hydrophilicity</li> <li>Photopolymerizable</li> <li>Elasticity</li> <li>Tunable physicochemical properties</li> </ul>	

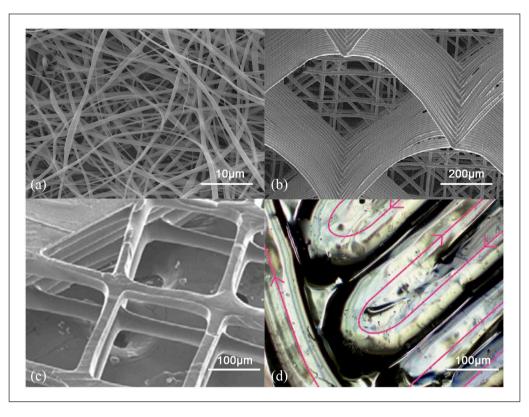
Table 2. Descriptive list of biomaterials suitable for auxetic constructs for skin tissue engineering applications.

collector in a spun way, resulting in a scaffold composed of a dense network of interconnected fibers. This technique allows for the production of fibers with diameters in the nanometer and micrometer range, which is compatible with the natural ECM fiber size range<sup>103</sup> and could promote cell attachment and proliferation. It also allows for the production of scaffolds with a high surface area, which can increase the interactions between cells and the scaffold. However, ES presents some limitations, as it can be challenging to achieve a uniform fiber diameter and distribution, and the fibers may have a smaller size than those produced by other techniques, which could result in weaker structures.<sup>104</sup> Furthermore, ES may not be suitable for certain types of polymers, as they may not be able to be stable at the high voltages required.

On the other hand, MEW, which has been under extensive development since 2011,<sup>105</sup> is a similar process that allows fabricating the polymer fibers with a higher resolution and precision where both position and orientation of the deposited fiber can be defined,<sup>106</sup> allowing a more precise control in the design of the object to be printed. It also involves using an electrical current to melt and extrude a polymer material through a fine nozzle, and then it is cooled and solidified as it is extruded, forming fibers and structures. In this case, the nozzle or the collector can be moved spatially to form the desired pattern or shape as the fiber is solidifying. MEW offers some advantages over other 3D printing techniques, such as its ability to print at high resolutions with high precision, which allows for the creation of scaffolds with a high degree of control over the microarchitecture,<sup>107</sup> as the set up can be moved in the three dimensions to create complex shapes. It also can be used to create scaffolds with a wide range of pore sizes and architectures, which can be tailored to meet the specific requirements of the application. Nevertheless, MEW also has some limitations due to the specialized equipment required to develop this technology.

Both ES and MEW processes work by using, in addition to the application of the electric field, mechanical means to encourage the material deposition using a syringe pump that controls the polymer solution feed rate.<sup>108</sup> The ratio between the electrical and the mechanical parameters must be considered, as it defines the amount of polymer delivered to the jet that can be accepted for the flow rate provided by the electric field, seeking to avoid defects such as ribbon-like structures or spherical drops.<sup>98,109,110</sup>

ES and MEW techniques have been used to manufacture numerous types of biomedical devices applicable in various fields, such as multi-purpose biocompatible scaffolds,<sup>55,111–115</sup> cartilage,<sup>116</sup> and cardiac<sup>117–119</sup> tissue engineering scaffolds, constructs for stem cell therapy<sup>120</sup> or drug delivery devices.<sup>121–124</sup> Their application in skin tissue engineering scaffold manufacturing is also wide. For



**Figure 5.** Different 3D printed structures achievable by: (a) Electrospinning. (b) Melt electro-writing. (c) Digital light processing. (d) 3D-bioprinting. Illustrations adapted with permission from Bhullar et al.,<sup>56</sup> Castilho et al.,<sup>53</sup> Soman et al.,<sup>96</sup> and Pourchet et al.,<sup>97</sup> respectively.

instance, it has been proposed the use of scaffolds with nanometric fiber size<sup>125–128</sup> or combined materials<sup>129</sup> for skin healing.

DLP techniques are an illumination method. Their application to additive manufacturing has been growing and becoming common in recent years due to its high performance in terms of resolution, printing speed, scalability, and material versatility, as it can be applied to soft materials like polymers and hydrogels, but also to metals and ceramics.<sup>130–136</sup>

In DLP applied to scaffold fabrication, a digital light projector is used to shine a pattern of light onto a photosensitive material solution, which contains a base-material, such as a hydrogel or a gelatin, and a photoinitiator, which reacts when exposed to the light, thus controlling the base-material solidification.<sup>137</sup> The light pattern is produced by a digital micromirror device (DMD) consisting of an array of micromirrors capable of changing their orientation via micro-actuators controlled by computer that allow them to reflect the light onto the material or onto an outer zone, where it is absorbed to prevent other surface reflections that could affect the precision of the fabrication process.<sup>138</sup> This precise control can be applied to obtain a wide range of shapes and geometries.

Besides the light pattern, other parameters, such as light intensity or wavelength, must also be controlled when using this process depending on the application, the photoinitiator, or the presence of cells, as they could be affected by the nature of the light applied.

The most commonly used polymers in DLP are acrylates, which are a class of monomers that can be polymerized to form polymers through photopolymerization. The use of these monomers is due to their excellent optical properties, low toxicity, and easy handling.

Photoinitiators are compounds that, upon exposure to light, generate free radicals or other reactive species that can initiate a chain reaction leading to the solidification of the material. When the photoinitiator is added to the acrylate, it serves as a catalyst that initiates the photopolymerization process when the acrylate is exposed to light. This process causes the acrylate molecules to form chemical bonds with one another, creating the solid scaffold. The photoinitiator is a crucial component of this process because it ensures that the polymerization reaction starts quickly and efficiently when the acrylate is exposed to light. Without it, the reaction would proceed at a much slower rate, making it difficult to form solid structures. Additionally, the properties of the photoinitiator can be tuned, such as its absorption wavelength, to match the light source and the rate of polymerization to control the polymerization kinetics of the acrylate.

DLP has several advantages over traditional scaffold fabrication methods, such as being able to create complex and highly porous structures with sub-millimeter resolution and high precision, as well as include gradients of mechanical and chemical properties in the scaffold.

DLP applications in the medical field are vast. In fact, this process has been used for medical devices such as implants, scaffold manufacturing,<sup>118,139,140</sup> or to study biological mechanisms as drug delivery.<sup>141</sup> Furthermore, its application into skin tissue engineering is promising. For instance, Zhou et al. have developed a functional living skin combining this technique with a bioprinting bioink that shows a superior performance in promoting dermal regeneration and also mimicking the physiological structure of natural skin.<sup>142</sup>

Finally, 3D bioprinting techniques are novel and also promising fabrication technologies developed in recent years that has been applied in several biomedical applications, including some encouraging results in skin regeneration studies.<sup>143–150</sup>

3D bioprinting involves a variety of techniques to control the printing process. Some of these techniques use extrusion-based methods, others rely on laser-assisted methods, and stereolithography is also a common option used for different applications.<sup>151–153</sup> Each technique offers a unique set of advantages and disadvantages, allowing researchers to choose the most appropriate approach for their specific application depending on the media deposited, which should also be carefully selected. The media printed in 3D bioprinting are bioinks, which are complex solutions composed of living cells, biomaterials and biological substances that can enhance the viability of the cells in its environment.<sup>154,155</sup>

These techniques allow the creation of geometrically biomimetic constructs suitable for patient-specific applications due to the control over localization and composition of the deposited solution, which can be designed to be similar to native tissue.<sup>97,156</sup>

Among the advantages of these techniques are their speed and efficiency, scalability, high degree of control and precision over each deposition step and the high cell density obtainable. However, they have some disadvantages such as complexity of the factors that must be reproduced and the difficulty in achieving a really close resemblance to the native tissue, due to its high intricacy. Some of the most commonly used biomaterials for 3D bioprinting are gelatin, collagen, and alginate, as these biopolymers are appropriate for mimicking the ECM due to their prominent level of cross-linked fibers and they also have suitable mechanical properties that support their printability.<sup>35,157,158</sup>

In the future, bioprinting is expected to become an increasingly important tool for creating functional, threedimensional tissue structures that could perceive and respond to their surroundings.<sup>159</sup> Advancements in materials, printing techniques, and bioprinting methods are expected to lead to the creation of more complex and anatomically accurate tissue structures. Furthermore, the integration of bioprinting with other technologies such as stem cell research and bio-fabrication, is expected to enable the creation of functional, living tissue structures that can be used for tissue repair and regeneration. Additionally, the integration of bioprinting with computational techniques, such as computer-aided design and simulation, is also expected to play a significant role, as it has the potential to change the way we think about tissue engineering and regenerative medicine.

Figure 6 presents a schematic diagram of the described fabrication techniques to summarize some of their main features to better understand their potential use for different skin tissue engineering applications.

## **Conclusions and future perspectives**

This review article has provided a comprehensive overview of the current state of research on the use of auxetic constructs for skin healing applications. The unique mechanical properties of auxetic materials, such as negative Poisson's ratio and their ability to accommodate large deformations both uniaxially and biaxially, have been found to have a positive impact on cell infiltration and proliferation,<sup>58–60</sup> making them a promising treatment option for skin tissue engineering scaffolds.

This review has examined various types of auxetic materials that have been tested numerically and experimentally, where studies have shown that auxetic scaffolds could promote wound healing by enhancing the cell microenvironment and being more adjustable to the complex body geometries. Additionally, the review has identified different biomaterials and fabrication techniques available to develop auxetic micro and macro structures, each presenting different application possibilities.

Advanced fabrication technologies such as 3D printing techniques or electrospinning can contribute significantly to the development of auxetic scaffolds. For example, 3D printing enables the fabrication of complex structures with tailored properties, while electrospinning can be used to create nanofibrous scaffolds with high surface area-to-volume ratios.

Computational studies have proven to be an important tool in predicting the mechanical behavior of auxetic structures and designs, aiding in the optimization of their geometry designs and material properties for skin tissue engineering applications. The analysis of auxetic material designs through computational studies also contributes to obtain and adjust key parameters, such as strain rates or stiffness, to mimic the mechanical properties of the native ECM.

Nevertheless, several hurdles and challenges remain in the development and application of auxetic materials. These include the need to improve our understanding of their behavior and mechanical properties under different loading conditions, as well as a proper evaluation of the

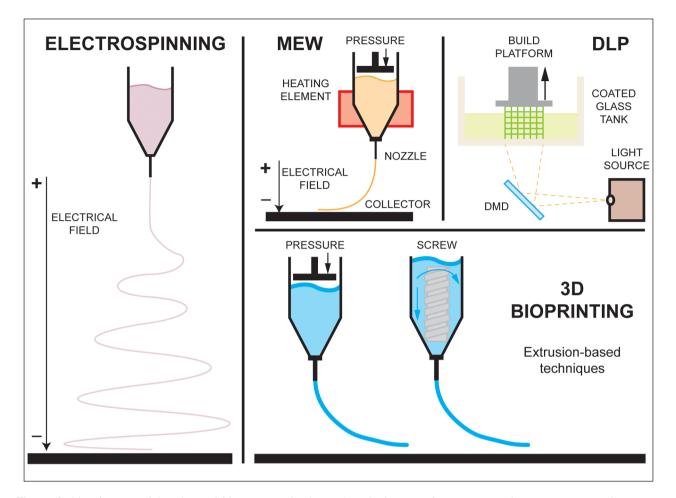


Figure 6. Main features of the advanced fabrication technologies described to manufacture auxetic skin tissue engineered constructs.

biocompatibility and long-term stability of these materials to ensure their safety and efficacy. To this end, relevant experiments would need to be performed, evaluating their quality and performance. These experiments should include tests to measure their tensile and compressive properties or their biocompatibility and degradation behavior. Furthermore, it is important to consider the practical implications of using these materials in a clinical setting, including regulatory compliance, cost-effectiveness, and scalability. This may require conducting extensive preclinical testing, such as in vitro and animal studies, in order to establish the safety of the materials. Also, while these materials have the potential to offer significant benefits over traditional skin grafts, they may also be more expensive to produce due to the use of advanced fabrication technologies or specialized biomaterials, which is intrinsically linked to the ability to manufacture these materials on a larger scale to meet the demands of the clinical market. To address these challenges, researchers and manufacturers may need to explore new approaches to manufacturing that can increase efficiency and reduce costs, for example using automation or high-throughput processing methods that may help to streamline the production process and reduce both time and costs involved.

In conclusion, the use of auxetic materials in skin tissue engineering holds great promise, and advanced fabrication technologies are rapidly advancing this field. Nonetheless, a more comprehensive understanding of their properties and behavior, coupled with well-designed experiments, is needed to overcome remaining hurdles and facilitate their successful translation into clinical applications.

### **Declaration of conflicting interests**

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

## Funding

The author(s) disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: The Authors acknowledge the project LMP 176\_21 "Design, 3D printing, and predictive modeling of wound healing dressings based on hydrogels reinforced with auxetic structures" funded by the Department of Science, University and Knowledge Society of the Government of Aragon. E.G-G is funded by a Ramon & Cajal Fellowship (RYC2021-033490-I, funded by MCIN/ AEI/10.13039/501100011033 and the EU "NextGenerationEU/ PRTR"). Authors would also like to acknowledge the Spanish Ministry of Economy and Competitiveness through the project PID2020-113819RB-I00.

## ORCID iD

Óscar Lecina-Tejero 问 https://orcid.org/0000-0001-7996-5646

#### References

- Groeber F, Holeiter M, Hampel M, et al. Skin tissue engineering–in vivo and in vitro applications. *Adv Drug Deliv Rev* 2011; 63: 352–366.
- Schulz JT, Tompkins RG and Burke JF. Artificial skin. Annu Rev Med 2000; 51: 231–244.
- Ahmadi Ashtiani HR, Akaberi M, Nilforoushzadeh MA, et al. Repairing injured skin: biologics, skin substitutes, and scaffolds: review. *J Skin Stem Cell* 2019; 5(4): e86162.
- Bhardwaj N, Chouhan D and Mandal BB. 3D functional scaffolds for skin tissue engineering. In: Deng Y and Kuiper J (eds) *Functional 3D tissue engineering scaffolds: materials, technologies, and applications.* Amsterdam: Elsevier, 2018, pp.345–365.
- Metcalfe AD and Ferguson MW. Tissue engineering of replacement skin: the crossroads of biomaterials, wound healing, embryonic development, stem cells and regeneration. *J R Soc Interface* 2007; 4: 413–437.
- Supp DM and Boyce ST. Engineered skin substitutes: practices and potentials. *Clin Dermatol* 2005; 23(4): 403–412.
- Veronda DR and Westmann RA. Mechanical characterization of skin-finite deformations. J Biomech 1970; 3(1): 111–124.
- Lees C, Vincent JF and Hillerton JE. Poisson's ratio in skin. Biomed Mater Eng 1991; 1(1): 19–23.
- Pissarenko A, Yang W, Quan H, et al. Tensile behavior and structural characterization of pig dermis. *Acta Biomater* 2019; 86: 77–95.
- Joodaki H and Panzer MB. Skin mechanical properties and modeling: A review. *Proc IMechE, Part H: J Engineering in Medicine* 2018; 232: 323–343.
- 11. Zahouani H, Pailler-Mattei C, Sohm B, et al. Characterization of the mechanical properties of a dermal equivalent compared with human skin in vivo by indentation and static friction tests. *Skin Res Technol* 2009; 15(1): 68–76.
- Ní Annaidh A, Bruyère K, Destrade M, et al. Characterization of the anisotropic mechanical properties of excised human skin. *J Mech Behav Biomed Mater* 2012; 5(1): 139–148.
- Agache PG, Monneur C, Leveque JL, et al. Mechanical properties and Young's modulus of human skin in vivo. *Arch Dermatol Res* 1980; 269: 221–232.
- 14. Elsner P, Barel AO, Berardesca E, et al. *Mechanical function* of the skin: State of the art. Basel: Karger, 1998. Vol. 26.
- 15. Sree D. Artificial skin scaffold to treat burn scars and it's other applications. *Int J Pharm Biol Sci* 5(2): 11–24.
- Bacakova M, Pajorova J, Broz A, et al. A two-layer skin construct consisting of a collagen hydrogel reinforced by a fibrin-coated polylactide nanofibrous membrane. *Int J Nanomedicine* 2019; 14: 5033–5050.

- Herndon DN, Barrow RE, Rutan RL, et al. A comparison of conservative versus early excision. *Ann Surg* 1989; 209: 547–553.
- MacNeil S. Progress and opportunities for tissue-engineered skin. *Nature* 2007; 445: 874–880.
- Tavakoli S and Klar AS. Bioengineered skin substitutes: Advances and future trends. *Appl Sci* 2021; 11(4): 11493– 11518.
- Nyame TT, Chiang HA, Leavitt T, et al. Tissue-engineered skin substitutes. *Plast Reconstr Surg* 2015; 136(6): 1379– 1388.
- Boyce ST. Design principles for composition and performance of cultured skin substitutes. *Burns* 2001; 27: 523–533.
- Davison-Kotler E, Sharma V, Kang NV, et al. A universal classification system of skin substitutes inspired by factorial design. *Tiss Eng Part B Rev* 2018; 24: 279–288.
- Kyriakidis C, Lali F, Greco KV, et al. Chronic leg ulcers: are tissue engineering and biomaterials science the solution? *Bioengineering* 2021; 8: 62.
- Adams DC and Ramsey ML. Grafts in dermatologic surgery: review and update on full- and split-thickness skin grafts, free cartilage grafts, and composite grafts. *Dermatol Surg* 2006; 31: 1055–1067.
- Centanni JM, Straseski JA, Wicks A, et al. Stratagraft skin substitute is well-tolerated and is not acutely immunogenic in patients with traumatic wounds: results from a prospective, randomized, controlled dose escalation trial. *Ann Surg* 2011; 253(4): 672–683.
- Falanga V and Sabolinski M. A bilayered living skin construct (APLIGRAF®) accelerates complete closure of hardto-heal venous ulcers. *Wound Repair Regen* 1999; 7(4): 201–207.
- 27. Clarke JA. HIV transmission and skin grafts. *Lancet* 1987; 1: 983.
- Chogan F, Chen Y, Wood F, et al. Skin tissue engineering advances in burns: a brief introduction to the past, the present, and the future potential. *J Burn Care Res* 2023; 44(Supplement 1): S1–S4.
- Jorgensen AM, Mahajan N, Atala A, et al. Advances in skin tissue engineering and regenerative medicine. *J Burn Care Res* 2023; 44(Suppl 1): S33–S41.
- Chang P, Li S, Sun Q, et al. Large full-thickness wounded skin regeneration using 3D-printed elastic scaffold with minimal functional unit of skin. *J Tissue Eng* 2022; 13: 20417314211063022.
- Shevchenko RV, James SL and James SE. A review of tissue-engineered skin bioconstructs available for skin reconstruction. J R Soc Interface 2010; 7: 229–258.
- Hama R, Reinhardt JW, Ulziibayar A, et al. Recent tissue engineering approaches to mimicking the extracellular matrix structure for skin regeneration. *Biomimetics* 2023; 8: 130.
- Chow L, Yick K, Wong KH, et al. 3D printing auxetic architectures for hypertrophic scar therapy. *Macromol Mater Eng* 2022; 307(5). DOI: 10.1002/mame.202100866
- Jang KI, Chung HU, Xu S, et al. Soft network composite materials with deterministic and bio-inspired designs. *Nat Commun* 2015; 6: 6566.
- Choi DJ, Park SJ, Gu BK, et al. Effect of the pore size in a 3D bioprinted gelatin scaffold on fibroblast proliferation. J Ind Eng Chem 2018; 67: 388–395.

- Mir M, Ali MN, Sami J, et al. Review of mechanics and applications of auxetic structures. *Adv Mater Sci Eng* 2014; 2014: 1–17.
- Yu X, Zhou J, Liang H, et al. Mechanical metamaterials associated with stiffness, rigidity and compressibility: A brief review. *Prog Mater Sci* 2018; 94: 114–173.
- Kadic M, Milton GW, van Hecke M, et al. 3D metamaterials. *Nat Rev Phys* 2019; 1: 198–210.
- Baughman RH, Shacklette JM, Zakhidov AA, et al. Negative Poisson's ratios as a common feature of cubic metals. *Nature* 1998; 392: 362–365.
- Critchley R, Corni I, Wharton JA, et al. A review of the manufacture, mechanical properties and potential applications of auxetic foams. *Phys Status Solidi B Basic Res* 2013; 250(10): 1963–1982.
- Dwivedi KK, Lakhani P, Kumar S, et al. Effect of collagen fibre orientation on the Poisson's ratio and stress relaxation of skin: an ex vivo and in vivo study. *R Soc Open Sci* 2022; 9(3): 211301.
- Frolich LM, LaBarbera M and Stevens WP. Poisson's ratio of a crossed fibre sheath: the skin of aquatic salamanders. J Zool 1994; 232: 231–252.
- Timmins LH, Wu Q, Yeh AT, et al. Structural inhomogeneity and fiber orientation in the inner arterial media. *Am J Physiol Heart Circ Physiol* 2010; 298: 1537H1537–1537H1545.
- 44. Skacel P and Bursa J. Poisson's ratio of arterial wall inconsistency of constitutive models with experimental data. J Mech Behav Biomed Mater 2016; 54: 316–327.
- Gatt R, Vella Wood M, Gatt A, et al. Negative Poisson's ratios in tendons: an unexpected mechanical response. *Acta Biomater* 2015; 24: 201–208.
- Williams JL and Lewis JL. Properties and an anisotropic model of cancellous bone from the proximal tibial epiphysis. *J Biomech Eng* 1982; 104(1): 50–56.
- Wiebe C and Brodland GW. Tensile properties of embryonic epithelia measured using a novel instrument. *J Biomech* 2005; 38(10): 2087–2094.
- Patten K and Wess T. Suprafibrillar structures of collagen, evidence for local organization and auxetic behaviour in architectures. *J Biophys Chem* 2013; 04(03): 103–109.
- Kraus EA, Mellenthin LE, Siwiecki SA, et al. Rheology of marine sponges reveals anisotropic mechanics and tuned dynamics. *J R Soc Interface* 2022; 19(195): 20220476.
- Kim Y, Son KH and Lee JW. Auxetic structures for tissue engineering scaffolds and biomedical devices. *Materials* 2021; 14: 6821.
- Mardling P, Alderson A, Jordan-Mahy N, et al. The use of auxetic materials in tissue engineering. *Biomater Sci* 2020; 8: 2074–2083.
- Beckett LE, Lewis JT, Tonge TK, et al. Enhancement of the mechanical properties of hydrogels with continuous fibrous reinforcement. ACS Biomater Sci Eng 2020; 6: 5453–5473.
- Castilho M, Mouser V, Chen M, et al. Bi-layered microfibre reinforced hydrogels for articular cartilage regeneration. *Acta Biomater* 2019; 95: 297–306.
- Afghah F, Iyison NB, Nadernezhad A, et al. 3D fiber reinforced hydrogel scaffolds by melt electrowriting and gel casting as a hybrid design for wound healing. *Adv Healthc Mater* 2022; 11(11): e2102068.

- Visser J, Melchels FP, Jeon JE, et al. Reinforcement of hydrogels using three-dimensionally printed microfibres. *Nat Commun* 2015; 6: 6933.
- Bhullar SK, Rana D, Lekesiz H, et al. Design and fabrication of auxetic PCL nanofiber membranes for biomedical applications. *Mater Sci Eng C Mater Biol Appl* 2017; 81: 334–340.
- Chen YW, Wang K, Ho CC, et al. Cyclic tensile stimulation enrichment of Schwann cell-laden auxetic hydrogel scaffolds towards peripheral nerve tissue engineering. *Mater Des* 2020; 195: 195.
- Chansoria P, Blackwell J, Etter EL, et al. Rationally designed anisotropic and Auxetic hydrogel patches for adaptation to dynamic organs. *Adv Funct Mater* 2022; 32: 2207590.
- Flamourakis G, Spanos I, Vangelatos Z, et al. Laser-made 3D auxetic metamaterial scaffolds for tissue engineering applications. *Macromol Mater Eng* 2020; 305(7): 305.
- Jin Y, Xie C, Gao Q, et al. Fabrication of multi-scale and tunable auxetic scaffolds for tissue engineering. *Mater Des* 2021; 197: 197.
- Pertoldi L, Zega V, Comi C, et al. Dynamic mechanical characterization of two-photon-polymerized SZ2080 photoresist. *J Appl Phys* 2020; 128(17): 175102.
- Liu J and Zhang Y. Soft network materials with isotropic negative Poisson's ratios over large strains. *Soft Matter* 2018; 14(5): 693–703.
- 63. Gupta S, Gupta V and Chanda A. Biomechanical modeling of novel high expansion auxetic skin grafts. *Int J Numer Methods Biomed Eng* 2022; 38(5): e3586.
- Gupta V and Chanda A. Expansion potential of skin grafts with novel I-shaped auxetic incisions. *Biomed Phys Eng Express* 2021; 8(1): 015016.
- Bi H and Jin Y. Current progress of skin tissue engineering: seed cells, bioscaffolds, and construction strategies. *Burns Trauma* 2013; 1: 63–72.
- Mogoşanu GD and Grumezescu AM. Natural and synthetic polymers for wounds and burns dressing. *Int J Pharm* 2014; 463(2): 127–136.
- Sahana TG and Rekha PD. Biopolymers: applications in wound healing and skin tissue engineering. *Mol Biol Rep* 2018; 45: 2857–2867.
- Ma L, Gao C, Mao Z, et al. Collagen/chitosan porous scaffolds with improved biostability for skin tissue engineering. *Biomaterials* 2003; 24(26): 4833–4841.
- Aboulgheit S, Abdelkader S, Aboushelib M, et al. Collagen chitosan scaffolds on induced skin defect in a rat model (an experimental study). *Alex Dent J* 2021; 46(1): 136–143.
- Ansari M, Kordestani SS, Nazralizadeh S, et al. Biodegradable cell-seeded collagen based polymer scaffolds for wound healing and skin reconstruction. *J Macromol Sci Part B Phys* 2018; 57(2): 100–109.
- Ahn S, Yoon H, Kim G, et al. Designed three-dimensional collagen scaffolds for skin tissue regeneration. *Tissue Eng Part C Methods* 2010; 16(5): 813–820.
- Kim J, Lee KM, Han SH, et al. Development of stabilized dual growth factor-loaded hyaluronate collagen dressing matrix. *J Tissue Eng* 2021; 12: 2041731421999750.
- Wang Y, Wang Z and Dong Y. Collagen-based biomaterials for tissue engineering. ACS Biomater Sci Eng 2023; 9: 1132–1150.

- Shahverdi M, Seifi S, Akbari A, et al. Melt electrowriting of PLA, PCL, and composite PLA/PCL scaffolds for tissue engineering application. *Sci Rep* 2022; 12(1): 19935.
- Woodruff MA and Hutmacher DW. The return of a forgotten polymer—polycaprolactone in the 21st century. *Prog Polym Sci* 2010; 35: 1217–1256.
- Stowers RS. Advances in extracellular matrix-mimetic hydrogels to guide stem cell fate. *Cells Tissues Organs* 2022; 211: 36–53.
- Li X, Sun Q, Li Q, et al. Functional hydrogels with tunable structures and properties for tissue engineering applications. *Front Chem* 2018; 6: 499.
- Uppuluri VNVA, Thukani Sathanantham S, Bhimavarapu SK, et al. Polymeric hydrogel scaffolds: skin tissue engineering and regeneration. *Adv Pharm Bull* 2022; 12(3): 437–448.
- Kohli N, Sharma V, Brown SJ, et al. Synthetic polymers for skin biomaterials. In: García-Gareta E (ed) *Biomaterials for skin repair and regeneration*. Amsterdam: Elsevier, 2019, pp.125–149.
- Dudziak M, Topolniak I, Silbernagl D, et al. Long-time behavior of surface properties of microstructures fabricated by multiphoton lithography. *Nanomater* 2021; 11(12): 3285.
- Jonušauskas L, Gailevičius D, Mikoliūnaitė L, et al. Optically clear and resilient free-form μ-optics 3D-printed via ultrafast laser lithography. *Materials* 2017; 10(1): 10.
- Mačiulaitis J, Deveikytė M, Rekštytė S, et al. Preclinical study of SZ2080 material 3D microstructured scaffolds for cartilage tissue engineering made by femtosecond direct laser writing lithography. *Biofabrication* 2015; 7(1): 015015.
- Costa BNL, Adão RMR, Maibohm C, et al. Cellular interaction of bone marrow mesenchymal stem cells with polymer and hydrogel 3D microscaffold templates. ACS Appl Mater Interfaces 2022; 14(11): 13013–13024.
- Sun M, Sun X, Wang Z, et al. Synthesis and properties of gelatin methacryloyl (GelMA) hydrogels and their recent applications in load-bearing tissue. *Polymers* 2018; 10: 1290.
- Choi JR, Yong KW, Choi JY, et al. Recent advances in photo-crosslinkable hydrogels for biomedical applications. *Biotechniques* 2019; 66: 40–53.
- Stillman Z, Jarai BM, Raman N, et al. Degradation profiles of poly(ethylene glycol)diacrylate (PEGDA)-based hydrogel nanoparticles. *Polym Chem* 2020; 11(2): 568–580.
- Chaudhari AA, Vig K, Baganizi DR, et al. Future prospects for scaffolding methods and biomaterials in skin tissue engineering: A review. *Int J Mol Sci* 2016; 17: 1974.
- Hosseini M and Shafiee A. Engineering bioactive scaffolds for skin regeneration. *Small* 2021; 17(41): 2101384.
- Kennedy KM, Bhaw-Luximon A and Jhurry D. Skin tissue engineering: biological performance of electrospun polymer scaffolds and translational challenges. *Regen Eng Transl Med* 2017; 3: 201–214.
- Qin J, Chen F, Wu P, et al. Recent advances in bioengineered scaffolds for cutaneous wound healing. *Front Bioeng Biotechnol* 2022; 10: 841583.
- Tarassoli SP, Jessop ZM, Al-Sabah A, et al. Skin tissue engineering using 3D bioprinting: an evolving research field. *J Plast Reconstr Aesthet Surg* 2018; 71: 615–623.

- 92. Heinrich MA, Liu W, Jimenez A, et al. 3D bioprinting: from benches to translational applications. *Small* 2019; 15: e1805510.
- Gupta S, Bissoyi A and Bit A. A review on 3D printable techniques for tissue engineering. *BioNanoScience* 2018; 8(3): 868–883.
- Guzzi EA, Bischof R, Dranseikiene D, et al. Hierarchical biomaterials via photopatterning-enhanced direct ink writing. *Biofabrication* 2021; 13(4): 044105.
- Chen J, Fan Y, Dong G, et al. Designing biomimetic scaffolds for skin tissue engineering. *Biomater Sci* 2023; 11: 3051–3076.
- Soman P, Lee JW, Phadke A, et al. Spatial tuning of negative and positive Poisson's ratio in a multi-layer scaffold. *Acta Biomater* 2012; 8(7): 2587–2594.
- Pourchet LJ, Thepot A, Albouy M, et al. Human Skin 3D bioprinting using scaffold-free approach. *Adv Healthc Mater* 2017; 6(4). Epub ahead of print 2016 Dec 15. DOI: 10.1002/adhm.201601101.
- Gomes S, Querido D, Ferreira JL, et al. Using water to control electrospun polycaprolactone fibre morphology for soft tissue engineering. *J Polym Res* 2019; 26(9): 26.
- Smith JA and Mele E. Electrospinning and additive manufacturing: adding three-dimensionality to electrospun scaffolds for tissue engineering. *Front Bioeng Biotechnol* 2021;
   DOI: 10.3389/fbioe.2021.674738
- 100. Larrondo L and John Manley RS. Electrostatic fiber spinning from polymer melts. I. Experimental observations on fiber formation and properties. *J Polym Sci Polym Phys Ed* 1981; 19(6): 909–920.
- 101. Larrondo L and St. John Manley R. Electrostatic fiber spinning from polymer melts. II. Examination of the flow field in an electrically driven jet. *J Polym Sci Polym Phys Ed* 1981; 19(6): 921–932.
- 102. Larrondo L and St. John Manley R. Electrostatic fiber spinning from polymer melts. III. Electrostatic deformation of a pendant drop of polymer melt. *J Polym Sci Polym Phys Ed* 1981; 19(6): 933–940.
- 103. Sun B. The mechanics of fibrillar collagen extracellular matrix. *Cell Rep Phys Sci* 2021; 2: 100515.
- 104. Wong SC, Baji A and Leng S. Effect of fiber diameter on tensile properties of electrospun poly(ε-caprolactone). *Polymer* 2008; 49(21): 4713–4722.
- 105. Brown TD, Dalton PD and Hutmacher DW. Direct writing by way of melt electrospinning. *Adv Mater* 2011; 23(47): 5651–5657.
- 106. Hrynevich A, Elçi BŞ, Haigh JN, et al. Dimension-based design of melt electrowritten scaffolds. *Small* 2018; 14(22): e1800232.
- 107. Paxton NC, Lanaro M, Bo A, et al. Design tools for patient specific and highly controlled melt electrowritten scaffolds. *J Mech Behav Biomed Mater* 2020; 105: 103695.
- 108. Brown TD, Edin F, Detta N, et al. Melt electrospinning of poly(ε-caprolactone) scaffolds: phenomenological observations associated with collection and direct writing. *Mater Sci Eng C* 2015; 45: 698–708.
- 109. Dokuchaeva AA, Timchenko TP, Karpova EV, et al. Effects of electrospinning parameter adjustment on the mechanical behavior of poly-ε-caprolactone vascular scaffolds. *Polymers* 2022; 14(2): 14.
- 110. Stachewicz U, Dijksman JF, Soudani C, et al. Surface free energy analysis of electrospun fibers based on

Rayleigh-Plateau/Weber instabilities. *Eur Polym J* 2017; 91: 368–375.

- 111. Karchin A, Simonovsky FI, Ratner BD, et al. Melt electrospinning of biodegradable polyurethane scaffolds. *Acta Biomater* 2011; 7(9): 3277–3284.
- 112. Hong JK and Madihally SV. Three-dimensional scaffold of electrosprayed fibers with large pore size for tissue regeneration. *Acta Biomater* 2010; 6(12): 4734–4742.
- 113. Pham QP, Sharma U and Mikos AG. Electrospun poly(epsilon-caprolactone) microfiber and multilayer nanofiber/microfiber scaffolds: characterization of scaffolds and measurement of cellular infiltration. *Biomacromolecules* 2006; 7(10): 2796–2805.
- 114. Bayati V, Abbaspour MR, Dehbashi FN, et al. A dermal equivalent developed from adipose-derived stem cells and electrospun polycaprolactone matrix: an in vitro and in vivo study. *Anat Sci Int* 2017; 92(4): 509–520.
- 115. Loewner S, Heene S, Baroth T, et al. Recent advances in melt electro writing for tissue engineering for 3D printing of microporous scaffolds for tissue engineering. *Front Bioeng Biotechnol* 2022; 10: 896719.
- 116. Bas O, De-Juan-Pardo EM, Meinert C, et al. Biofabricated soft network composites for cartilage tissue engineering. *Biofabrication* 2017; 9(2): 025014.
- 117. Saidy NT, Shabab T, Bas O, et al. Melt electrowriting of complex 3D anatomically relevant scaffolds. *Front Bioeng Biotechnol* 2020; 8: 793.
- 118. Wang Z, Wang L, Li T, et al. 3D bioprinting in cardiac tissue engineering. *Theranostics* 2021; 11: 7948–7969.
- Serpooshan V, Mahmoudi M, Hu DA, et al. Bioengineering cardiac constructs using 3D printing. *J 3D Print Med* 2017; 1(2): 123–139.
- 120. Gizaw M, Faglie A, Pieper M, et al. The role of electrospun fiber scaffolds in stem cell therapy for skin tissue regeneration. *Med One* 2019; 4: e190002.
- 121. Kataria K, Gupta A, Rath G, et al. In vivo wound healing performance of drug loaded electrospun composite nanofibers transdermal patch. *Int J Pharm* 2014; 469(1): 102–110.
- 122. Yu DG, Wang M and Ge R. Strategies for sustained drug release from electrospun multi-layer nanostructures. *Wiley Interdiscip Rev Nanomed Nanobiotechnol* 2022; 14(3): e1772.
- 123. Ding Y, Li W, Zhang F, et al. Electrospun fibrous architectures for drug delivery, tissue engineering and cancer therapy. Adv Funct Mater 2019. DOI: 10.1002/adfm.201802852
- 124. Liu H, Wang H, Lu X, et al. Electrospun structural nanohybrids combining three composites for fast helicide delivery. *Adv Compos Hybrid Mater* 2022; 5(2): 1017–1029.
- 125. Cerkez I, Sezer A and Bhullar SK. Fabrication and characterization of electrospun poly(e-caprolactone) fibrous membrane with antibacterial functionality. *R Soc Open Sci* 2017; 4(2): 160911.
- 126. Pezeshki-Modaress M, Mirzadeh H and Zandi M. Gelatin– GAG electrospun nanofibrous scaffold for skin tissue engineering: Fabrication and modeling of process parameters. *Mater Sci Eng C* 2015; 48: 704–712.
- 127. Lizarazo-Fonseca L, Muñoz Prieto E, Vera Graziano R, et al. Andamios eletrohilados de poli(ε-caprolactona) / colágeno con uso potencial en regeneración de tejido cutáneo. *Ciencia en Desarrollo* 2019; 10(2): 197–208.

- Abrigo M, McArthur SL and Kingshott P. Electrospun nanofibers as dressings for chronic wound care: advances, challenges, and future prospects. *Macromol Biosci* 2014; 14: 772–792.
- 129. Hewitt E, Mros S, Mcconnell M, et al. Melt-electrowriting with novel milk protein/PCL biomaterials for skin regeneration. *Biomed Mater* 2019; 14(5):055013. DOI: 10.1088/1748-605X/ab3344.
- Melchels FPW, Feijen J and Grijpma DW. A review on stereolithography and its applications in biomedical engineering. *Biomaterials* 2010; 31: 6121–6130.
- 131. Lu Y, Mapili G, Suhali G, et al. A digital micro-mirror device-based system for the microfabrication of complex, spatially patterned tissue engineering scaffolds. *J Biomed Mater Res A* 2006; 77(2): 396–405.
- 132. Chaudhary R, Fabbri P, Leoni E, et al. Additive manufacturing by digital light processing: a review. *Prog Addit Manuf* 2023; 8: 331–351.
- 133. Bifano M. Digital light processing: A review on the printing resolution and the materials options. *Appl Comput Eng* 2022; 1: 17–25.
- 134. Lin FS, Lee JJ, Lee AKX, et al. Calcium silicate-activated gelatin methacrylate hydrogel for accelerating human dermal fibroblast proliferation and differentiation. *Polymers* 2021; 13(1): 1–14.
- 135. Mo X, Ouyang L, Xiong Z, et al. Advances in digital light processing of hydrogels. *Biomed Mater* 2022; 17: 042002.
- 136. Varghese G, Moral M, Castro-García M, et al. Fabrication and characterisation of ceramics via low-cost DLP 3D printing. *Bol Soc Esp Cerám Vidr* 2018; 57(1): 9–18.
- 137. Choi JW, Kim GJ, Hong S, et al. Sequential process optimization for a digital light processing system to minimize trial and error. *Sci Rep* 2022; 12(1): 13553.
- 138. Zhao Z, Tian X and Song X. Engineering materials with light: Recent progress in digital light processing based 3D printing. *J Mater Chem C* 2020; 8: 13896–13917.
- 139. Li H, Dai J, Wang Z, et al. Digital light processing (DLP)based (bio)printing strategies for tissue modeling and regeneration. *Aggregate* 2022; 4(2): e270.
- 140. Gong J, Qian Y, Lu K, et al. Digital Light Processing (DLP) in tissue engineering: from promise to reality, and perspectives. *Biomed Mater* 2022; 17(6). DOI: 10.1088/1748-605X/ac96ba.
- 141. Zhang J, Hu Q, Wang S, et al. Digital light processing based three-dimensional printing for medical applications. *Int J Bioprinting* 2020; 6(1): 242.
- 142. Zhou F, Hong Y, Liang R, et al. Rapid printing of bioinspired 3D tissue constructs for skin regeneration. *Biomaterials* 2020; 258: 120287.
- 143. Olejnik A, Semba JA, Kulpa A, et al. 3D bioprinting in skin related research: recent achievements and application perspectives. ACS Synth Biol 2022; 11: 26–38.
- 144. Lee V, Singh G, Trasatti JP, et al. Design and fabrication of human skin by three-dimensional bioprinting. *Tissue Eng Part C Methods* 2014; 20(6): 473–484.
- 145. He P, Zhao J, Zhang J, et al. Bioprinting of skin constructs for wound healing. *Burns Trauma* 2018; 6: 5.
- 146. Pontiggia L, Van Hengel IA, Klar A, et al. Bioprinting and plastic compression of large pigmented and vascularized human dermo-epidermal skin substitutes by means of a new robotic platform. *JTissue Eng* 2022; 13: 20417314221088513.
- 147. Zhao H, Xu J, Yuan H, et al. 3D printing of artificial skin patches with bioactive and optically active polymer

materials for anti-infection and augmenting wound repair. *Mater Horiz* 2022; 9(1): 342–349.

- 148. Javaid M and Haleem A. 3D bioprinting applications for the printing of skin: A brief study. *Sens Int* 2021; 2: 100123.
- 149. Weng T, Zhang W, Xia Y, et al. 3D bioprinting for skin tissue engineering: current status and perspectives. *J Tissue Eng* 2021; 12: 20417314211028574.
- 150. Zhang M, Zhang C, Li Z, et al. Advances in 3D skin bioprinting for wound healing and disease modeling. *Regen Biomater* 2023; 10: rbac105.
- 151. Klak M, Bryniarski T, Kowalska P, et al. Novel strategies in artificial organ development: What is the future of medicine? *Micromachines* 2020; 11(7): 646.
- 152. Li J, Chen M, Fan X, et al. Recent advances in bioprinting techniques: approaches, applications and future prospects. J *Transl Med* 2016; 14(1): 271.
- 153. Yu J, Park SA, Kim WD, et al. Current advances in 3D bioprinting technology and its applications for tissue engineering. *Polymers* 2020; 12: 1–30.

- 154. Bishop ES, Mostafa S, Pakvasa M, et al. 3-D bioprinting technologies in tissue engineering and regenerative medicine: Current and future trends. *Genes Dis* 2017; 4: 185–195.
- 155. Zhang X and Zhang Y. Tissue engineering applications of three-dimensional bioprinting. *Cell Biochem Biophys* 2015; 72(3): 777–782.
- 156. Seol YJ, Lee H, Copus JS, et al. 3D bioprinted biomask for facial skin reconstruction. *Bioprinting* 2018; 10: 10.
- 157. Liu P, Shen H, Zhi Y, et al. 3D bioprinting and in vitro study of bilayered membranous construct with human cellsladen alginate/gelatin composite hydrogels. *Colloids Surf B Biointerfaces* 2019; 181: 1026–1034.
- 158. Niu C, Wang L, Ji D, et al. Fabrication of SA/Gel/C scaffold with 3D bioprinting to generate micro-nano porosity structure for skin wound healing: a detailed animal in vivo study. *Cell Regen* 2022; 11(1): 10.
- 159. de León EHP, Valle-Pérez AU, Khan ZN, et al. Intelligent and smart biomaterials for sustainable 3D printing applications. *Curr Opin Biomed Eng* 2023; 26: 100450.