Revolutionising oral organoids with artificial intelligence

Jiawei Yang¹, Nicholas G. Fischer², Zhou Ye^{1,*}

Key Words:

artificial intelligence; bioprinting; dental stem cells; machine learning; oral disease; oral organoids

From the Contents

Introduction	372
Overview of Oral Organoids	374
Overview of Artificial Intelligence	379
Challenges and Perspectives	383

ABSTRACT

The convergence of organoid technology and artificial intelligence (AI) is poised to revolutionise oral healthcare. Organoids - three-dimensional structures derived from human tissues - offer invaluable insights into the complex biology of diseases, allowing researchers to effectively study disease mechanisms and test therapeutic interventions in environments that closely mimic in vivo conditions. In this review, we first present the historical development of organoids and delve into the current types of oral organoids, focusing on their use in disease models, regeneration and microbiome intervention. We then compare single-source and multi-lineage oral organoids and assess the latest progress in bioprinted, vascularised and neural-integrated organoids. In the next part of the review, we highlight significant advancements in AI, emphasising how AI algorithms may potentially promote organoid development for early disease detection and diagnosis, personalised treatment, disease prediction and drug screening. However, our main finding is the identification of remaining challenges, such as data integration and the critical need for rigorous validation of AI algorithms to ensure their clinical reliability. Our main viewpoint is that current AI-enabled oral organoids are still limited in applications but, as we look to the future, we offer insights into the potential transformation of AI-integrated oral organoids in oral disease diagnosis, oral microbial interactions and drug discoveries. By synthesising these components, this review aims to provide a comprehensive perspective on the current state and future implications of AI-enabled oral organoids, emphasising their role in advancing oral healthcare and improving patient outcomes.

*Corresponding author:

Zhou Ye, zhouye22@hku.hk.

http://doi.org/10.12336/ biomatertransl.2024.04.004

How to cite this article: Yang, J.; Fischer, NG.; Ye, Z. Revolutionising oral organoids with artificial intelligence. *Biomater Transl.* **2024**, *5*(4), 372-389.



Introduction

Organoids represent а groundbreaking advancement in biological research, offering simplified three-dimensional (3D) models of human organs grown from stem cells. These miniature structures replicate key aspects of tissue architecture and function, yielding invaluable insights into organ development, disease mechanisms and potential therapeutic interventions.^{1, 2} The evolution of organoids has transitioned from basic cell culture to sophisticated models that mimic the complexity of human tissues, driven by advancements in stem cell biology, tissue engineering and regenerative medicine. Recent studies have demonstrated

the potential of organoids to maintain critical physiological and pathological characteristics of their parental tissues to model various diseases and facilitate drug discovery.^{3, 4}

Oral diseases present a unique set of complexities due to the diverse cell types involved. Conditions like periodontitis, oral cancer and dental caries involve multifaceted pathological processes that are challenging to study with traditional twodimensional cell cultures or animal models. Given the high prevalence and significant impact of these diseases on overall health, there is an urgent need for advanced models to elucidate underlying mechanisms and develop effective treatments.⁵ Oral organoids have emerged as

particularly significant models, simulating the architecture and functionality of oral tissues such as the gingiva, periodontal ligament and dental pulp.⁶ These models pave the way for personalised medicine and novel therapeutic approaches.^{7,8}

The integration of artificial intelligence (AI) into oral organoid research marks a significant advancement, enhancing our ability to analyse complex biological systems and improve treatment strategies for oral diseases. AI technologies, particularly machine learning (ML) algorithms, have been employed to analyse high-dimensional data generated from organoid cultures, facilitating the identification of cellular heterogeneity and drug responses.⁹ For instance, the application of deep learning (DL) techniques have enabled researchers to classify organoid differentiation states and predict treatment outcomes based on transcriptomic data, streamlining drug discovery processes.¹⁰ AI also plays a vital role in optimising organoid bioprinting, ensuring the precision and reproducibility. Researchers can create more reliable *in vitro* models that closely mimic the physiological conditions of the oral cavity by employing AI to monitor organoid development quality.¹¹

Moreover, AI's potential extends to personalised medicine, where ML algorithms can analyse patient-specific data to tailor treatments based on individual responses.¹² This personalised approach enhances therapeutic efficacy and minimises adverse effects, improving patient outcomes in managing oral diseases. As the field evolves, the synergy between AI and organoid technology is expected to yield novel insights into the pathophysiology of oral conditions and drive innovative therapeutic strategies.

Hence, in this narrative review, we examine how oral organoids are revolutionising our understanding of oral health and disease while examining how AI can amplify these advancements. Integrating AI with organoid technology will enable researchers to overcome traditional limitations, enhance experimental precision and accelerate clinical translation, ultimately pushing the frontiers of oral health research (**Figure 1**).

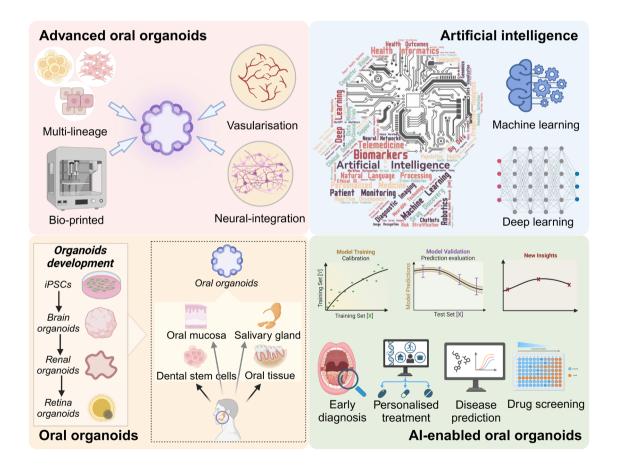


Figure 1. Schematic illustration of the development of oral organoids and perspectives of AI-enabled oral organoids. Created with BioRender.com. AI: artificial intelligence.

¹ Applied Oral Sciences and Community Dental Care, Faculty of Dentistry, The University of Hong Kong, Hong Kong Special Administrative Region, China; 2 MDRCBB, Minnesota Dental Research Center for Biomaterials and Biomechanics, University of Minnesota, Minneapolis, MN, USA

Overview of Oral Organoids History of organoids

Research over the last half century has shown cells cultured in two-dimensional environments, such as tissue culture polystyrene, do not represent in vivo biology.^{13, 14} This discrepancy contributes to the high failure rate of new drugs as they translate from benchtop to clinical trials, coining the term "valley of death" to describe this critical bottleneck in drug development.¹⁵ With rising ethical concerns around animal testing, there's an urgent need for alternative models such as 3D culture platforms.¹⁶ Organoids, derived from induced pluripotent stem cells (iPSCs), embryonic stem cells (ESCs), or adult stem cells (ASCs), recreate the structure and function of organs. A critical aspect of organoids is the ability of the construct to self-organise or spatially organise and form clusters. Therefore, here we use a general definition for organoids taken from Sakalem et al.¹⁷ as "refer[ring] to 3D cultures that present multiple cell lineages in co-culture [or more] that are able to spatially organise and form clusters".

Cells are typically supported by a complex mixture of factors in the culture media to create organoids.¹⁸ IPSC and ESC organoids normally involve differentiation protocols with multiple different culture media to mimic developmental cues.¹⁹ Organoids are typically formed by suspending or supporting cell in myriad scaffold materials (like Matrigel[®])²⁰ or scaffold-free techniques including droplets hanging from plates²¹ and air-liquid-interface models.²²

IPSC/ESC-derived organoids generally exhibit more diverse and organ-like functions and can expand more readily than those from ASCs.²³ Organoids derived from ASCs tend to exhibit spheroid shapes while iPSC/ESC tend to show more complex geometries associated with brain, lung and kidney.²⁴ While iPSCs are technically more complex, they can differentiate into nearly any lineage and better model organ development compared to ASCs, such as mesenchymal stem cells, which are widely studied for their immune-regulating properties.^{25, 26}

Oral organoids

Organoids have emerged as a significant tool in dental, oral and craniofacial medicine, addressing critical healthcare challenges in these fields. Organoids facilitate the investigation of complex biological interactions, particularly between ectoderm and mesoderm during tooth development.²⁷ Here we spotlight key advances in oral organoids for oral mucosa, salivary glands, teeth and other oral tissues.

Oral mucosa organoids

The oral mucosa acts as a barrier against the microbial and physical challenges in the oral environment. Collected surgical oral resection tissue can be cultured with a basement membrane extract and Rho-associated kinase inhibitor to create human oral mucosal organoids, which express basal cell markers, proliferation markers and keratin and overall resemble natural mucosa allowing expansion for up to 6 months.²⁸ Similarly, lingual mucosa organoids have been developed to exhibit stratified squamous epithelial layers by

culturing with epidermal growth factor, the Wnt signallingactivator R-Spondin1 and the transforming growth factor β inhibitor noggin (**Figure 2A**).²⁹

Co-cultures are often employed to investigate paracrine signalling and organoid interactions. For instance, oral squamous cell carcinoma organoids co-cultured with fibroblasts demonstrated enhanced organoid formation due to lactate and other secreted factors.³⁰ Organoid culture conditions vary significantly across different disease states, as evidenced by comparisons between normal oesophageal tissue, gastroesophageal junction, oesophageal squamous cell carcinoma and Barrett's oesophagus.³¹

Salivary gland organoids

Salivary glands are crucial for daily functions like speaking and eating and their dysfunction - especially after radiation therapy - can significantly affect oral health. These glands feature branching duct systems composed mainly of acinar, ductal and myoepithelial cells, with their structure resulting from epithelial-mesenchymal crosstalk.³² Salivary gland organoids can be established using fragments or ESCs. For instance, Tanaka et al.³³ demonstrated the potential of ESCs to create organoids that showed mature salivary gland function when transplanted into a defective mouse parotid gland model (**Figure 2B**).

Research has revealed that salivary gland organoids grown in hyaluronic acid hydrogels express key functional markers like tight junction proteins and α -amylase³⁴ and are responsive to neurotransmitters when implanted in resected parotid glands of immunocompromised rats.³⁵ Rat tail collagen has served as a scaffold for self-renewing organoids derived from submandibular gland cells, which differentiate into acinar and ductal cells.³⁶

Branching morphogenesis, essential for salivary gland development, is relevant to other organs as well. Coculturing embryonic submandibular gland epithelial cells with bone marrow-derived mesenchymal stem cells in Matrigel[®] has produced self-assembling organoids with branching morphology influenced by cell ratios.³⁷ Factors like laminin 111 and fibroblast growth factor 2 promoted the differentiation of epithelial progenitor cells into terminal buds.³⁸ These advancements contribute to our understanding of salivary gland branching morphogenesis and potential regenerative strategies.

Tooth and dental stem cell organoids

Engineering of teeth has been a long-held goal given myriad dental diseases and tooth loss. A key challenge in constructing tooth organoids is recreating the spatiotemporal crosstalk between dental epithelium and mesenchyme.⁶ An early study combined dissociated dental epithelial and dental mesenchymal cells from tooth germs at the cap stage in a collagen gel, successfully generating tooth-like structure upon implantation.³⁹ This approach has been adapted for canine⁴⁰ and porcine⁴¹ models.

Most tooth-derived stem cells are obtained from human third molars, complicating the creation of anterior tooth organoids. This has prompted the use of pluripotent stem

cells alongside dental mesenchymal stem cells as feeder cells to form tooth organoids that develop tooth-like structures postimplantation.⁴² An emerging stem cell for epithelial cells is the cell rests of Malassez,⁴³ which can differentiate into ameloblasts crucial for dental hard tissue when cultured with dental pulp stem cells (DPSCs).⁴⁴

Other innovative approaches for developing tooth organoids involve culturing adult dental epithelial stem cells in Matrigel[®], relying on various signalling pathways to produce highly elongated hydroxyapatite (**Figure 2C**).⁴⁵ Additionally, Bektas et al.⁴⁶ created methacrylated gelatine microparticles to culture DPSCs and dental epithelial cells, resulting in differentiated dental cells. Calabrese et al.⁴⁷ combined dental pulp stem/ progenitor cells and periodontal ligament stem/progenitor cells to mimic spatial organisation and mineral patterns of tooth roots, including rudimentary periodontal ligament tissue. These advancements represent promising steps toward effective tooth regeneration.

Other oral tissue organoids

Dental, oral and craniofacial structures are highly diverse, leading to the development of various organoids for their study. For instance, taste bud organoids can be cultured by combing circumvallate epithelium and foliate papillae cells within Matrigel[®], which helps localise stem cells and taste receptors along with their innervation (**Figure 2D**).⁴⁸ This concept has been extended to engineering artificial tongues using a decellularised extracellular matrix.⁴⁹

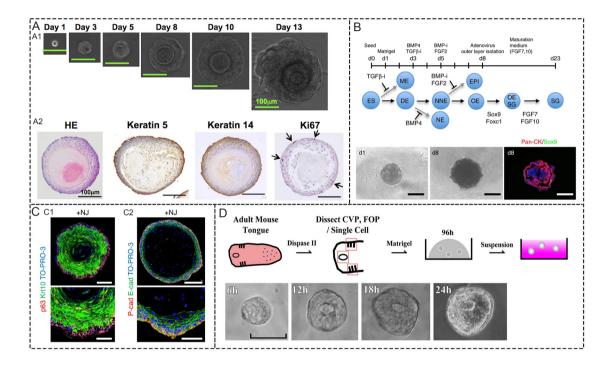


Figure 2. Various oral organoid applications. (A) Lingual mucosa organoid growth and histological analysis of the organoids. Staining for Ki67 showed that some cells actively proliferated in the outer periphery (arrows). Scale bars: 100 μm. Reprinted from Hisha et al.²⁹ (B) Generation of a salivary gland organoid and immunofluorescence analysis of pan-cytokeratin (Pan-CK, red) and Sox9 (green). Scale bars: 300 μm. Reprinted from Tanaka et al.³³ (C) Confocal images of a 3D culture system for murine dental epithelial organoids. Scale bars: 100 μm (top), 50 μm (bottom). Reprinted from Kim et al.⁴⁵ (D) Schematic of suspension-culture method for taste bud organoids and the time-lapse imaging of suspension-cultured organoids under a bright-field microscope. Red squares indicate CVP and FOP. Scale bar: 50 μm. Reprinted from Adpaikar et al.⁴⁸ Copyright 2024, Springer Nature. BMP4: bone morphogenetic protein inhibitor; CVP: adult mice circumvallate papilla; DE: definitive ectoderm; E-cad: E-cadherin; EPI: epidermis; ES: embryonic stem cells; FGF: fibroblast growth factor; FOP: Foliate papillae; Foxc1: forkhead box C1; HE: haematoxylin eosin staining; Ki67: Kiel 67; Krt10: Keratin 10; ME: mesendoderm; NE: neural ectoderm; NJ: Noggin and Jagged1; NNE: non-neural ectoderm; OE: oral ectoderm; OE-SG: salivary gland placode; p63: tumour protein 63; P-cad: P-cadherin; Sox9: SRY-box transcription factor 9; TGFβ-i: transforming growth factor β inhibitor; TO-PRO-3: Thiazole red.

Cartilage exists throughout the head and neck, with hyaline cartilage found in the nose and fibrocartilage in the temporomandibular joint.⁵⁰ Crispim and Ito⁵¹ developed hyaline cartilage organoids from chondrocytes in a notochordal cell-derived matrix. These organoids expressed prototypical hyaline cartilage features like type II collagen, type VI collagen, glycosaminoglycans, with SRYbox transcription factor 9-positive cells. Future research may expand on these models by incorporating concepts from other cartilage organoids, such as those related to osteoarthritis and larger joints in the human body.⁵²

Applications of oral organoids Disease model

Oral organoids have become a crucial tool for developing disease models, especially oral cancers and salivary gland disorders. Organoids derived from human salivary glands have been shown to maintain key functional characteristics, making them suitable for modelling salivary gland diseases and testing potential therapeutic interventions (**Figure 3A**).^{53, 54} Salivary gland organoids were also utilised to create a swelling

model, induced by cholinergic stimulation, to simulate the physiological conditions of saliva secretion, revealing the underlying mechanism of salivary gland disorders and innovative treatments.^{53, 54} Patient-derived oral carcinogenesis organoids can accurately reflect tumour characteristics and responses to therapies,^{55, 56} making it possible to explore the genetic alterations and tumour microenvironment for mastering cancer biology and developing targeted treatments (**Figure 3B**).³⁰

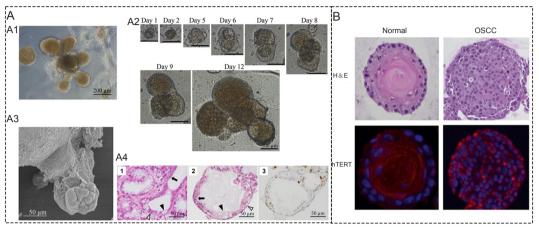


Figure 3. Applications of oral organoids as disease models. (A) Morphological and functional analyses of the human salivary-gland-derived organoids. The organoid has two layers of cells, an inner lining of epithelial (arrow in A4-2) and mucous cells (arrowhead in A4-2) and an outer lining of cells (open arrowhead in A4-2). These inner and outer layers are reminiscent of the luminal inner epithelial (arrow in A4-1) and mucous (arrowhead in A4-1) cells and outer myoepithelial cells (open arrowhead in A4-1), respectively, in the region of the intercalated duct connecting to the secretory end piece of the normal salivary gland. Scale bar: $200 \,\mu$ m (A1), $100 \,\mu$ m (A2), $50 \,\mu$ m (A3, A4). Reprinted from Yoshimoto et al.⁵³ (B) H&E analysis and hTERT expression in normal oral and OSCC organoids. Reprinted from Yoon et al.⁵⁵ hTERT: human telomerase reverse transcriptase; H&E: haematoxylin eosin staining; OSCC: oral squamous cell carcinoma.

The application of organoids extends beyond cancer modelling. Gao et al.⁶ discussed how oral organoids can be used to study various diseases, including maxillofacial tumours and tooth dysplasia, thereby contributing to a better understanding of oral health and disease mechanisms. Furthermore, the progressive clustered regularly interspaced palindromic repeats clustered regularly interspaced short palindromic repeats (CRISPR) and CRISPR-associated protein 9 (CRISPR/Cas9) for gene editing has enabled oral organoids to model specific genetic alterations associated with oral diseases, enhancing the relevance of these models for translational research.⁵⁶

Regeneration

Advances in bioprinting technology have enabled the creation of salivary gland organoids for drug discovery and regenerative therapies, minimising the reliance on animal models.^{57, 58} Studies have found that bioengineered salivary glands, developed using embryonic organ-inductive potential stem cells, can regenerate fully functional salivary glands *in vitro*, highlighting the potential for clinical applications in treating conditions like xerostomia.^{57, 59, 60} Oral organoids derived from dental stem cells present promising opportunities for tooth regeneration. These stem cells can differentiate into various cell types necessary for the regeneration of dental tissues, including dentine and periodontal ligaments.^{61, 62}

376

Creating organoids from these cells opens up possibilities for investigating odontogenic mechanisms and developing biomimetic artificial teeth, potentially transforming dental restorative practices (**Figure 4**).^{60, 63, 64}

Oral organoids are also valuable for studying the mechanisms of tissue regeneration and repair. The application of intermittent compressive forces has been proven to enhance cell cycling and reduce apoptosis in embryoid bodies derived from iPSCs, suggesting that mechanical stimuli can positively influence the regenerative capacity of these cells.⁶⁵ Furthermore, oral organoids help in addressing diseases that affect oral health.

Microbiome interaction

Oral organoids' 3D structures assist in investigating the oral microbiome and host-tissue interactions in a controlled condition. Traditional methods often miss the dynamic nature of the oral cavity. Dysbiosis, or an imbalance in microbial communities, can lead to oral diseases, including periodontitis and dental caries.^{66,67} Oral organoids can simulate the oral environment and examine how specific microbial species interact with epithelial cells, potentially leading to inflammation and tissue damage. This approach allows for a more nuanced understanding of the mechanisms underlying oral diseases and the role of microbial communities in their progression.

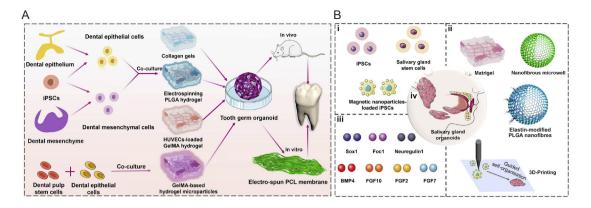


Figure 4. Oral organoid applications for tooth and salivary gland formation. (A) Scheme of engineered tooth germ organoids fabrication for tooth regeneration. Reprinted from Wang and Sun.⁶⁰ (B) Scheme of generation of engineered salivary gland organoids. Reprinted from Wang and Sun.⁶⁰ 3D: three-dimensional; BMP4: bone morphogenetic protein 4; FGF: fibroblast growth factor; Foc1: *Fusarium oxysporum* f. sp. cubense race 1; GelMA: gelatine methacrylate; HUVECs: human umbilical vein endothelial cells; iPSCs: induced pluripotent stem cells; PCL: polycaprolactone; PLGA: poly(lactic-co-glycolic acid); Sox1: SRY-box transcription factor 1.

Moreover, oral organoids can be employed to investigate how microbes transmit between the oral cavity and other body sites, like the gastrointestinal tract. Salivary gland organoids are being employed to investigate the regeneration mechanisms in the context of inflammatory bowel disease, where salivary gland impairment occurs due to systemic inflammation.⁶⁸ This research could lead to novel therapeutic strategies for managing oral symptoms associated with such diseases. By using organoids derived from oral and intestinal tissues, researchers can study how oral microbes influence gut microbiome composition and function, which is relevant for systemic diseases linked to oral health, such as cardiovascular disease and diabetes.^{69,70}

Another exciting area of research involves modelling hostpathogen interactions using oral organoids. Forbester et al.⁷¹ have demonstrated their utility in studying how pathogenic bacteria interact with epithelial cells, assisting in developing targeted therapies. Organoids are valuable for evaluating various treatments on microbial communities are provide insights into restoring a healthy microbiome balance.⁷²

Advanced oral organoids

Single-source vs. multi-lineage oral organoids

Single-source oral organoids, derived from a single progenitor cell type, replicate the architecture and function of specific oral tissues, enabling controlled experiments to study cellular behaviours. Hemeryck et al.⁷³ created an organoid model from human dental follicle tissue, showing a tooth epithelial stemness phenotype similar to the epithelial cell rests of Malassez, confirmed through single-cell transcriptomics. Song et al.⁷⁴ demonstrated that transforming growth factor β 1 promotes differentiation in organoids derived from human gingival mesenchymal stem cells without compromising viability. However, the limited cellular diversity of singlesource organoids restricts their ability to model complex biological processes.⁷⁵ In contrast, multi-lineage oral organoids, developed from various progenitor cell types, offer a more accurate representation of oral tissues. This closely mimics the physiological environment and facilitates the study of interactions between different cell types.⁷⁶ Recent advancements in growth factors and small molecule cocktails that simulate organ stem cell niches enhance their capacity for differentiation and self-renewal into complex structures.⁶

Furthermore, advanced imaging and sequencing technologies enable real-time monitoring and gene expression analysis, enriching our understanding of organoid biology.³ Multilineage oral organoids are particularly promising for drug testing and personalised medicine, allowing for the assessment of individual responses to therapies using patient-derived cells. This approach is especially relevant for oral diseases where patient-specific factors significantly influence treatment outcomes, making multi-lineage organoids a powerful platform to explore cellular dynamics and processes like tissue regeneration, inflammation and cancer progression.^{77, 78}

Bioprinted oral organoids

3D bioprinting technology is revolutionising oral organoid manufacturing by overcoming limitations of traditional 3D culture methods, such as size constraints and lack of reproducibility, as well as insufficient vascular and immune cell interactions.⁷⁹ By layering bio-inks, bioprinting allows precise control over organoid composition and structure, facilitating the creation of larger, more complex tissue-like models. This is particularly beneficial for studying oral cancers, as chimeric organoids combining normal and cancerous cells can be developed to explore tumour microenvironments and cellular interactions.^{56, 80, 81} Enhanced reproducibility and structural fidelity enable organoids to be engineered with specific traits essential for accurate disease modelling, which is especially important in oral cancer research for recreating tumour heterogeneity.^{56, 81, 82}

Bio-inks derived from natural materials have further improved organoid viability and functionality, creating a more physiologically relevant environment.⁸³ Integrating microfluidic systems with bioprinted organoids enables real-time analysis of dynamic cellular behaviour and drug response,⁸⁴ simulating physiological conditions like nutrient flow.⁸⁵ Magnetic 3D bioprinting has also been used to create salivary gland organoids, replicating functional epithelial compartments responsive to neurotransmitters (**Figure 5A**).^{86, 87} These neuroepithelial organoids hold promise for drug discovery and cytotoxicity screening, particularly for conditions like dry mouth syndrome.

Vascularised oral organoids

Vascularisation is essential for organoid survival and functionality, providing nutrients and oxygen while removing waste. Xu et al.⁶⁴ created dental pulp organoids with endothelial and DPSCs, highlighting the role of vascularisation in organoid development, especially in studying dentin and pulp interactions in teeth.⁶² IPSCs can differentiate into endothelial cells crucial for vascular structures. Recently, 3D human iPSC-derived blood vessel organoids resembling native vessels were created.⁸⁸ Vascularised liver organoids derived from iPSCs further showcase the potential of stem cells in forming vascularisation (Figure 5B).89 Additionally, some specific growth factors, such as Wnt family member 2B, have been reported to significantly enhance vascular development in organoids by increasing endothelial cell number.⁹⁰ Hence, with advancements in stem cells and specific growth factors, vascularised oral organoids with complex structures and adequate functions will be designed for promising applications. Recent efforts to enhance vascularisation have been drawn from various organ systems. Holkom et al.⁹¹ designed oral organoids to examine how the tumour microenvironment regulates angiogenesis in oral squamous cell carcinoma, identifying nicotinamide N-methyltransferase as a potential target for antiangiogenic therapy (Figure 5C). Kidney organoids cultured on kidney-decellularised extracellular matrices exhibited strong vascularisation, suggesting similar methods could benefit oral organoids.92

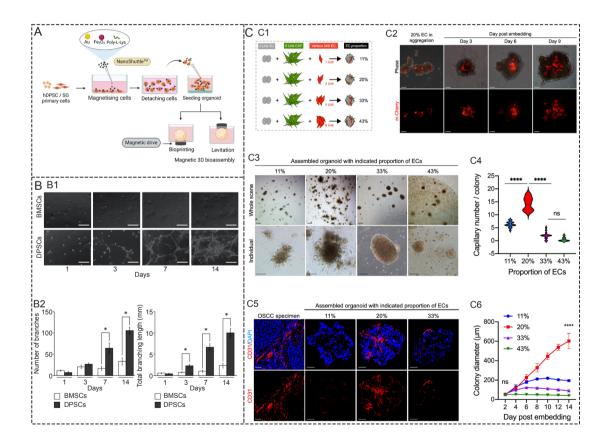


Figure 5. Applications of advanced oral organoids. (A) SG organoid biofabrication workflow utilising two different magnetic 3D bioassembly platforms. Reprinted from Klangprapan et al.⁸⁷ (B) Comparison of the sprouting ability of BMSCs and DPSCs, showcasing the potential of stem cells in forming vascularised organoids. *P < 0.05. Scale bars: 200 µm. Reprinted from Li et al.⁸⁹ (C) Generation of assembled organoid comprising ECs, fibroblasts and cancer cells. ****P < 0.0001. Scale bars: 100 µm. Reprinted from Holkom et al.⁹¹ Copyright 2024, with permission from Wiley. 3D: three-dimensional; Au: gold; BMSCs: bone marrow-derived mesenchymal stem cells; CAF: cancer-associated fibroblast; CD31: cluster of differentiation 31; DAPI: blue-fluorescent DNA stain; DPSCs: dental pulp stem cells; EC: endothelial cells; FAO: fibroblast-attached organoid; Fe₂O₃: ferric oxide; hDPSC: human dental pulp stem cells; ns: not significant; OSCC: oral squamous cell carcinoma; Poly-L-Lys: polylysine; SG: salivary gland; TC: tumour cell.

Neural-integrated oral organoids

The development of neural-integrated oral organoids is a cutting-edge approach for creating more physiologically relevant models for studying oral health and disease by combining neural and oral epithelial tissues. 3D bioprinting technology enables the precise spatial arrangement of various cell types, including neural and epithelial cells, within oral organoids. Li et al.⁹³ have demonstrated the potential of bioprinting to create brain-like co-culture constructs, which could be adapted for oral organoid fabrication. Such oral organoids could provide insights into the interactions between neural and epithelial cells in the oral cavity, which are crucial for understanding conditions like pain, inflammation and cancer.

Integrating neural stem cells into oral organoids enhances their functionality by mimicking *in vivo* neural-epithelial interactions. Research has emphasised the importance of incorporating both vascular and neural populations to improve physiological relevance and enable longitudinal analyses of functional development.⁹⁴ Additionally, the development of an integrated transcriptomic cell atlas of human neural organoids has provided insights into cellular composition and differentiation pathways.⁹⁵ This methodology can be applied to oral organoids to explore how neural integration affects gene expression and cellular interactions, ultimately improving organoid functionality.

Overview of Artificial Intelligence

With its ability to mimic human intelligence and perform tasks that typically require human cognition, AI is reshaping the landscape of healthcare, finance, education and more. In healthcare, AI is being utilised to predict disease outbreaks, assist in surgical procedures and optimise treatment protocols, paving the way for a future where AI technology plays a crucial role in improving human health and well-being.⁹⁶ Here we generally introduce the evolution of AI through several significant stages, the concept of integrating AI into organoids and provide specific examples of their use in various applications.

Evolution of artificial intelligence

During the 1950s and 1960s, AI research focused on symbolic reasoning and rule-based systems, which laid the groundwork for early AI applications. Pioneers like Alan Turing and John McCarthy were instrumental in shaping foundational ML and AI concepts. Alan Turing introduced the Turing Test in 1950 that set a benchmark for evaluating machine intelligence, while John McCarthy officially created the term "artificial intelligence" at the 1956 Dartmouth Conference.⁹⁷ Following that, people were optimistic about AI research, as the established machines could solve complex problems, making computers intelligent.⁹⁸

In the 1980s, AI research shifted towards symbolic AI and the development of "expert systems", which aimed to emulate human decision-making in specific domains like medicine and finance.^{99, 100} Expert systems relied on a knowledge base and an inference engine that processed and combined these symbols to make decisions. Despite their initial success and the potential to handle complex tasks, expert systems were limited by their inability to learn and adapt over time, relying on static representations of knowledge and rules.⁹⁸

AI evolution met a pivotal turning point as ML emerged in the 1990s. ML introduced algorithms that could learn from data and improve performance without explicit programming. This led to supervised learning models¹⁰¹ and unsupervised learning models,¹⁰² expanding AI applications in areas such as image recognition,¹⁰³ natural language processing¹⁰⁴ and recommendation systems.¹⁰⁵ Inspired by biological neural networks, these advancements allowed machines to learn from data rather than relying solely on pre-defined rules.

Today, AI has evolved into advanced systems incorporating reinforcement learning, generative models and autonomous systems. These technologies enable the creation of intelligent agents that can tackle complex real-world challenges. Integrating ML with emerging technologies highlights AI's transformative influence across sectors, such as healthcare, finance and transportation, significantly shaping modern society.¹⁰⁶

How is artificial intelligence assisting organoids development?

Initially, the development of organoids relied on traditional biological methods, focusing on isolating and culturing stem cells to create 3D structures.⁶ Organoid design and applications are exploring new avenues as AI is booming. As an essential component of AI, ML makes computers effectively process and analyse vast amounts of data (Figure 6).^{107, 108} ML algorithms have been employed to analyse single-cell transcriptomics data¹⁰⁹ and identify organoid phenotypes by automating the analysis of biological images.¹¹⁰ Indeed, in oral medicine in general, machine learning approaches for clinical radiographic image analysis is booming with some countries granting approval for clinical use.¹¹¹ In organoid research, ML helps optimise growth conditions by analysing data from experiments to identify the best combinations of growth factors and extracellular matrix components.¹¹² This data-driven approach improves the reproducibility and functionality of organoids.

DL is a subset of ML that significantly advances data analysis through multi-layer neural networks to enhance feature extraction and transformation.¹¹³ Unlike traditional ML methods, DL processes raw data directly via neural networks, allowing DL to efficiently train models end-to-end and perform exceptionally well with large datasets.¹¹⁴ Techniques like convolutional neural networks excel in image classification, including organoid detection.¹¹⁵⁻¹¹⁷ The development of OrgaQuant, a deep convolutional neural network, automates the detection of human intestinal organoids in bright-field images, significantly increasing research efficiency.¹¹⁸ Image analysis may be particularly powerful in oral organoids for systems looking at the interactions between cell populations during tooth formation, like ectoderm and mesoderm cells during tooth development.²⁷

Additionally, AI integration into bioprinting technologies optimises 3D printing parameters, facilitating the creation of complex and functional organoids that better mimic native tissue architecture.¹¹⁹ This has wide potential application for printing of the multi-tissue periodontal complex.

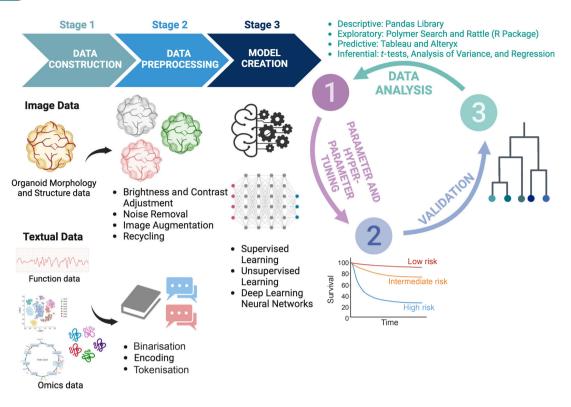


Figure 6. Integration of AI organoid system in three steps: data construction, data preprocessing and model creation. Reprinted from Maramraju et al.¹⁰⁸ AI: artificial intelligence.

Artificial intelligence-enabled organoids Early detection and diagnosis

AI-enabled organoids are proving to be valuable tools for early detection and diagnosis, offering a unique platform for studying tumour biology and drug responses. By using organoids derived from patient biopsies, researchers can closely examine the biological characteristics of tumours and identify novel biomarkers that inform clinical decision-making.¹²⁰ This approach is especially important in cancers like gastrointestinal stromal tumours, where patient-derived organoids have been instrumental in evaluating drug sensitivity and predicting treatment outcomes.^{120, 121} Moreover, the integration of AI into the analysis of organoid data significantly enhances the accuracy of these assessments. AI algorithms can detect subtle changes that may signal disease progression, thereby improving our understanding of tumour dynamics and enabling more tailored treatment strategies.⁴

In addition, AI-enabled organoids facilitate more precise and efficient analysis of complex biological systems, deepening our understanding of disease mechanisms and refining diagnostic capabilities. As highlighted by Maramraju et al.¹⁰⁸ AI-driven analyses yield results that surpass traditional human assessment, a critical factor for the accuracy needed in preclinical trials and diagnostics. This sentiment is echoed by Lampart et al.¹²² who developed analytical tools that enable high-throughput and high-content screenings, further demonstrating how ML enriches organoid research. The ability to sift through extensive datasets generated by organoid studies is critical for detecting subtle phenotypic changes that may signal disease progression or

therapeutic responses. Advanced imaging techniques exemplify AI's transformative role in this field. Gritti et al.¹²³ have introduced ML-based Organoids Analysis (MOrgAna) software for quantitatively analysing organoids, enabling researchers to conduct detailed morphological assessments (**Figure 7A**). This capability allows for the observation and analysis of dynamic changes in organoid structures over time, offering critical insights into developmental processes and disease states.

Personalised treatment and health monitoring

AI-enabled organoids are valuable in the realm of personalised medicine, offering the ability to create patient-derived organoids that closely replicate the characteristics of individual tumours or tissues. This innovation allows for a precise assessment of how specific therapeutic agents will affect individual patients. Studies have demonstrated that pancreatic cancer organoids can be used for drug screening directly on patient cells, facilitating treatment planning tailored to the unique genetic makeup of the tumour.4, 124 A potential use for AI and salivary glands is identification of early markers of saliva gland dysfunction from cultured organoids. The incorporation of microfluidic systems in organoid culture further enhances this process by enabling real-time monitoring of drug responses, which is critical for developing personalised treatment plans. Natarajan et al.¹²⁵ have showcased a microfluidic co-culture system that allows for live monitoring of T cell interactions with liver organoids, providing insights into immune responses and therapeutic strategies against viral infections. Takahashi126 emphasised the role of organoids in drug discovery and personalised medicine,

Biomaterials Translational

noting their ability to resemble various organs and respond to treatments in ways that reflect patient-specific responses. By integrating AI into the analysis of organoid data, researchers can enhance the efficiency of drug screening processes, leading to the identification of effective therapies tailored to individual patients. Park et al.¹²⁷ investigated the use of AI in predicting the differentiation of kidney organoids derived from human iPSCs. By utilising AI algorithms to assess the maturity of these organoids, researchers can select the most suitable models for clinical applications, thereby enhancing the reliability of drug testing and improving treatment outcomes for kidney-related diseases.

In terms of health monitoring, AI-enabled organoids serve as a dynamic platform for assessing disease progression and treatment efficacy. By integrating AI algorithms, researchers can analyse complex data generated from organoid experiments, gaining a comprehensive understanding of how individual organoids respond to treatments over time. The organoid brightfield identification-based therapy screening (OrBITS) platform developed by Deben et al.¹²⁸ enables labelfree, time-lapse monitoring of patient-derived organoids, significantly enhancing the ability to track organoid health and therapy responses (**Figure 7B**). This capability is essential for making timely adjustments to treatment strategies based on real-time data. Furthermore, applying AI to analyse organoid growth kinetics addresses the intra-tumoural heterogeneity commonly observed in cancers. By utilising ML techniques to assess individual organoid responses rather than relying on bulk cultures, researchers can uncover the variability in treatment responses among different tumour cells.¹²⁹

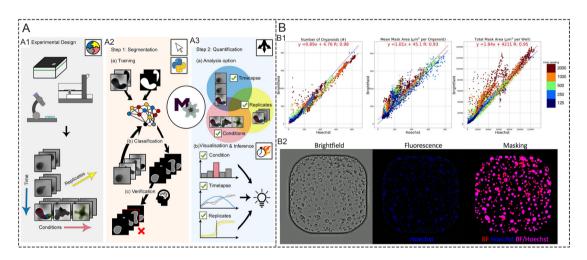


Figure 7. AI-enabled organoids for early diagnosis and health monitoring. (A) MOrgAna workflow schematic: experimental design, segmentation and quantification. Reprinted from Gritti et al.¹²³ (B) Brightfield *versus* Hoechst analysis of organoids. Reprinted from Deben et al.¹²⁸ AI: artificial intelligence; BF: brightfield; MOrgAna: machine-learning based organoids analysis; OrBITS: organoid brightfield identification-based therapy screening.

Disease prediction

AI-enabled organoids are revolutionising human disease modelling by closely mimicking tissue physiological and pathological characteristics. Cai et al.¹³⁰ have explored the potential of brain organoids, combined with AI, to create predictive models that simulate disease progression and response to therapies. In the realm of retinal diseases, Kegeles et al.¹³¹ have demonstrated that convolutional neural networks could accurately predict retinal differentiation in retinal organoids (**Figure 8A**). This predictive capability is vital for early diagnosis and intervention, enabling the identification of individuals at risk of developing vision-threatening conditions. The ability to forecast disease progression based on organoid behaviour enhances the potential for timely therapeutic strategies.

AI-enabled organoids are also valuable in studying neurodegenerative diseases, such as Alzheimer's and Parkinson's diseases. Esmail and Danter¹³² have developed

iPSC-derived brain organoids to model the genetics of Alzheimer's progression, yielding insights into the disease's pathophysiology. Similarly, Monzel et al.¹³³ employed ML techniques to predict neurotoxicity in human midbrain organoids, demonstrating the potential for AI to assess the impact of environmental toxins on neuronal health. By utilising AI to analyse organoid responses to various stimuli, researchers can predict disease progression and identify potential therapeutic interventions. In oral medicine, this will likely be most immediately useful for oral squamous cell carcinoma cancer organoids, which are more advanced and ready for AI-integration than other oral organoid systems.

Drug screening

As discussed above, organoids derived from patient-specific cells replicate an individual's unique genetic and phenotypic characteristics, providing a platform for drug testing that closely mimics the patient's tissues. The work of Phan et

al.¹³⁴ demonstrated that high-throughput screening using patient-derived tumour organoids can identify actionable drug sensitivities, creating a preclinical platform for precision medicine.

AI-enabled organoids further enhance the establishment of high-throughput drug screening platforms that can efficiently evaluate the efficacy of various compounds. Takahashi et al. conducted large-scale drug cytotoxicity screenings using human intestinal organoids, demonstrating the feasibility of screening extensive libraries of pharmacologically active compounds.¹³⁵ By utilising dispersed intestinal epithelial cells from organoids, they achieved a homogeneous cell population, which is crucial for minimising variability in drug response assessments. This method exemplifies how organoid technology can be adapted for high-throughput applications, making it a valuable tool in drug discovery.

Integrating ML algorithms into organoid drug screening

enhances data analysis and interpretation. Branciforti et al.¹²⁹ have highlighted the use of DL-based pipelines to analyse drug screening results from cancer organoids, combining imaging techniques with gene expression and protein interaction data. This comprehensive analysis leads to a more nuanced understanding of drug efficacy and mechanisms of action, ultimately improving the predictive power of drug screening assays. For example, the oral squamous cell carcinoma organoid system developed by Zhang et al.¹³⁶ for drug screening could be enhanced with AI. Furthermore, the automated microfluidic platforms developed by Schuster et al.137 enable dynamic and combinatorial drug screening of tumour organoids, allowing for the simultaneous testing of multiple drug combinations (Figure 8B). This approach not only accelerates the drug discovery process but also provides insights into the interactions between different therapeutic agents, which is essential for developing effective combination therapies.

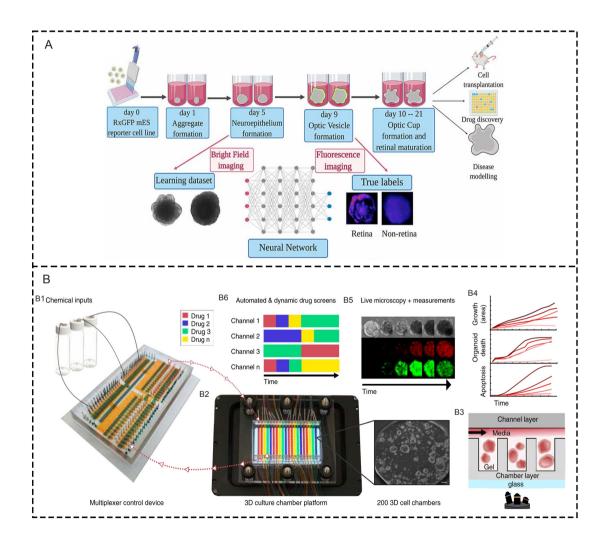


Figure 8. AI-enabled organoids for disease prediction and drug screening. (A) Schematic flow of retinal differentiation experiments using a neural network to predict retinal differentiation. Reprinted from Kegeles et al.¹³¹ (B) Automated microfluidic 3D cellular and organoid culture platform for dynamical drug perturbations. Reprinted from Schuster et al.¹³⁷ 3D: three-dimensional; AI: artificial intelligence; mES: mouse embryonic stem cell; RxGFP: mES reporter cell line.

Challenges and Perspectives

Challenges in developing oral organoids

Oral organoids, miniature models of oral tissues, have greatly expanded knowledge of oral biology and disease, yet several challenges remain in their development and applications. Tissues like the gingiva, periodontal ligament and dental pulp exhibit complex cellular interactions that are difficult to replicate *in vitro*. Achieving functional organoids requires precise control over the microenvironment and signalling pathways to accurately mirror the native tissue complexity.¹³⁸ Stringent quality control and standardisation are vital to ensure reproducibility and reliability in research outcomes. Ahn et al.¹³⁹ highlighted that replicating the *in vivo* environment poses significant challenges, necessitating integration of oral organoids into regulatory frameworks that prioritise quality and safety.

Without established manufacturing standards, variability in oral organoid quality can lead to inconsistent findings, complicating the translation of results into clinical applications.¹⁴⁰ Differences in the quality and type of stem cells, whether derived from different individuals or tissues, further complicate the development of consistent oral organoids. For instance, oral organoids derived from DPSCs and periodontal ligament stem cells may show differing differentiation potentials based on their origin.⁵⁶ Addressing these challenges is crucial for advancing oral organoid technology in research and clinical settings.

Moreover, ensuring equitable access to organoid technology is vital for its broader adoption in clinical practice, especially in resource-limited settings where specialised equipment and trained personnel may be lacking.¹⁴¹ The high costs associated with organoid production and analysis may be prohibitive for some patients, hindering their access to personalised treatments.

Challenges in artificial intelligence-driven oral organoids

Integrating AI into the development of oral organoids offers promising opportunities, yet critical considerations need to be addressed to ensure ethical and reliable application of AI. The process involves analysing sensitive health data from patientderived tissues, raising concerns about data privacy and algorithmic bias that could reinforce existing inequalities.^{142, 143} Mahmood et al.¹⁴⁴ underscored the necessity of context-specific quality assurance measures, which include acceptance testing prior to clinical use and continuous quality control monitoring. By addressing biases in the training data and ensuring that the data used for AI algorithms is representative and of high quality, researchers can improve the reliability of predictions made by AI systems in organoid studies. Furthermore, as AI technologies progress, regulatory frameworks need to adapt to ensure the safety and efficacy of AI-driven organoid applications. Ranjbar et al.¹⁴⁵ emphasised the importance of establishing new management systems and quality assurance mechanisms in healthcare organisations to accommodate the unique challenges posed by AI systems.

Rigorous validation of AI-driven models against experimental

results is crucial due to the complexity of biological systems, which may limit predictive accuracy.¹⁴⁶ Variability in organoid culture conditions, such as differences in media composition and environmental factors, can lead to inconsistent organoid characteristics, which may affect the outcomes of experiments.¹⁰⁸ The extensive data generated from organoid studies often surpasses the capabilities of traditional analysis methods, increasing the risk of data overload and model overfitting.^{147, 148} Thus, developing AI models capable of accurately capturing complex biological behaviours remains a significant challenge.

Ensuring the quality and consistency of oral organoids is critical for reliable research outcomes. While AI can assist in monitoring and analysing organoid characteristics, it also necessitates new quality control protocols, as traditional methods may be inadequate. Without robust quality assurance, the reproducibility of AI-driven results may be compromised, hindering further clinical translation. In addition, generating AI-enabled oral organoids requires seamless interoperability among various platforms and technologies to effectively analyse and provide actionable insights.

Perspectives of artificial intelligence-enabled oral organoids

While AI-driven oral organoids are still relatively rare, advancements in other types of organoids inspire their designs and applications. Integrating AI with oral organoids can significantly enhance diagnostic and treatment capabilities, analyse the interactions between oral microbiota and host cells and facilitate novel oral drug development.

For diagnostics, AI algorithms can analyse high-resolution imaging data from organoids, identifying disease markers and treatment responses for quicker and more accurate diagnoses. The landscape of AI models for predicting oral diseases has evolved, leveraging ML and DL techniques to enhance diagnostic accuracy and improve patient management. In treatment planning, AI optimises the testing of therapeutic drugs on organoids, allowing for patient-specific therapy adjustments.^{149,150} By simulating disease dynamics and analysing drug efficacy, AI-driven models can identify resistance mechanisms and early intervention biomarkers.^{151,152}

Moreover, ML techniques can mine multi-omics data to uncover patterns related to disease states and microbial interactions. AI can analyse the interplay between oral microbiota and host cells in organoid systems, identifying microbial signatures associated with disease and facilitating the development of personalised preventive strategies.^{153, 154} Understanding the roles of specific bacterial communities in periodontitis can inform the design of probiotics or other therapeutic interventions aimed at restoring a healthy oral microbiome. AI-driven organoids can model these interactions, providing insights into how microbial communities influence tissue health and disease progression. By analysing data from organoid studies, AI can identify patterns of microbial colonisation and their effects on host responses, potentially leading to the discovery of novel therapeutic targets. This research is vital for developing strategies to manipulate the

oral microbiome in favour of health, thereby preventing or mitigating diseases such as periodontitis.

AI-enabled organoids are revolutionising drug discovery processes in oral healthcare. By utilising organoids that closely mimic human oral tissues, researchers can conduct highthroughput screening of potential therapeutic compounds.¹⁵⁵ AI algorithms can analyse the responses of these organoids to various drugs, identifying effective candidates for further development.¹⁵⁶ This approach not only accelerates the drug discovery timeline but also reduces the reliance on animal models, aligning with ethical considerations in research.¹⁵⁷ Moreover, AI can assist in predicting treatment outcomes based on organoid responses, thereby facilitating the development of more effective and targeted therapies for oral diseases.

Furthermore, AI can help to deal with the challenges in developing oral organoids mentioned above. AI has the potential to improve accessibility to oral organoids by streamlining production processes, ultimately reducing costs and resource demands. By leveraging AI algorithms, oral organoid culture conditions can be analysed and optimised to enhance yield and efficiency, making organoid technology more viable in diverse clinical settings.¹⁰⁸ AI can significantly contribute to quality control by utilising ML to monitor and evaluate oral organoid development in real-time, ensuring more consistent and reliable outcomes.¹¹⁸ This early detection of deviations allows for timely interventions to maintain quality standards. Scalability is a common challenge in oral organoid production¹⁵⁸ and AI can address this issue by automating aspects of the organoid culture process, such as monitoring environmental conditions and adjusting parameters in realtime to optimise growth.¹¹² Additionally, AI-driven predictive models can assist in prioritising organoid development based on their potential for successful drug responses, optimising resource allocation and enhancing efficiency.¹⁰⁸

Although the discussion of oral organoids and AI is evergrowing, due to the limited applications of AI-enabled oral organoids, we hereby encapsulate the progress made in the development of oral organoids and AI, offering our perspectives on the potential for AI to transform oral organoids in the future. Overall, the development of AI-driven oral organoid technology suggests a transformative shift in oral healthcare, emphasising personalised treatment, early disease detection and continuous patient monitoring. These organoids can streamline therapeutic interventions and improve outcomes by harnessing AI for predictive analytics and drug development.

Author contributions

JY and ZY conceptualised the review; JY and NGF drafted the manuscript; ZY revised the manuscript. All authors reviewed and approved the final version of the manuscript.

Financial support

This work was supported by the IADR Innovation in Oral Care Awards, the Innovation and Technology Fund (No. ITS/307/22) and the National Natural Science Foundation of China (NSFC) Young Scientist Fund (No. 82401187). Acknowledgement

Parts of the figures were created by using the tool BioRender (https://www.biorender.com/).

Conflicts of interest statement

The authors declare no conflicts of interest.

Open access statement

This is an open-access journal, and articles are distributed under the terms of the Creative Commons Attribution-Non-Commercial-Share Alike 4.0 License, which allows others to remix, tweak, and build upon the work noncommercially if appropriate credit is given. The new creations are licensed under identical terms.

- Xinaris, C.; Brizi, V.; Remuzzi, G. Organoid models and applications in biomedical research. *Nephron.* 2015, 130, 191-199.
- Liu, H.; Zhang, X.; Liu, J.; Qin, J. Vascularization of engineered organoids. *BMEMat.* 2023, *1*, e12031.
- Tang, X. Y.; Wu, S.; Wang, D.; Chu, C.; Hong, Y.; Tao, M.; Hu, H.; Xu, M.; Guo, X.; Liu, Y. Human organoids in basic research and clinical applications. *Signal Transduct Target Ther.* 2022, *7*, 168.
- Zhou, Z.; Cong, L.; Cong, X. Patient-derived organoids in precision medicine: drug screening, organoid-on-a-chip and living organoid biobank. *Front Oncol.* 2021, *11*, 762184.
- Lamont, R. J.; Koo, H.; Hajishengallis, G. The oral microbiota: dynamic communities and host interactions. *Nat Rev Microbiol.* 2018, *16*, 745-759.
- 6. Gao, X.; Wu, Y.; Liao, L.; Tian, W. Oral organoids: progress and challenges. *J Dent Res.* **2021**, *100*, 454-463.
- Almansoori, A. A.; Kim, B.; Lee, J. H.; Tran, S. D. Tissue engineering of oral mucosa and salivary gland: disease modeling and clinical applications. *Micromachines (Basel)*. 2020, 11, 1066.
- 8. Schutgens, F.; Clevers, H. Human organoids: tools for understanding biology and treating diseases. *Annu Rev Pathol.* **2020**, *15*, 211-234.
- Tebon, P. J.; Wang, B.; Markowitz, A. L.; Davarifar, A.; Tsai, B. L.; Krawczuk, P.; Gonzalez, A. E.; Sartini, S.; Murray, G. F.; Nguyen, H. T. L.; Tavanaie, N.; Nguyen, T. L.; Boutros, P. C.; Teitell, M. A.; Soragni, A. Drug screening at single-organoid resolution via bioprinting and interferometry. *Nat Commun.* 2023, *14*, 3168.
- Kong, J.; Lee, H.; Kim, D.; Han, S. K.; Ha, D.; Shin, K.; Kim, S. Network-based machine learning in colorectal and bladder organoid models predicts anti-cancer drug efficacy in patients. *Nat Commun.* 2020, *11*, 5485.
- 11. Lee, H. Engineering in vitro models: bioprinting of organoids with artificial intelligence. *Cyborg Bionic Syst.* **2023**, *4*, 0018.
- Moosa, Y.; Alizai, M. H. K.; Tahir, A.; Zia, S.; Sadia, S.; Fareed, M. T. Artificial intelligence in oral medicine. *Int J Health Sci (Qassim)*. 2023, 7, 1476-1488.
- Duval, K.; Grover, H.; Han, L. H.; Mou, Y.; Pegoraro, A. F.; Fredberg, J.; Chen, Z. Modeling physiological events in 2D vs. 3D cell culture. *Physiology (Bethesda)*. 2017, *32*, 266-277.
- 14. Discher, D. E.; Janmey, P.; Wang, Y. L. Tissue cells feel and respond to the stiffness of their substrate. *Science*. **2005**, *310*, 1139-1143.
- Seyhan, A. A. Lost in translation: the valley of death across preclinical and clinical divide – identification of problems and overcoming obstacles. *Transl Med Commun.* 2019, *4*, 18.
- Horejs, C. Organ chips, organoids and the animal testing conundrum. *Nat Rev Mater.* 2021, *6*, 372-373.
- Sakalem, M. E.; De Sibio, M. T.; da Costa, F.; de Oliveira, M. Historical evolution of spheroids and organoids, and possibilities of use in life sciences and medicine. *Biotechnol J.* 2021, *16*, e2000463.
- Geurts, M. H.; Clevers, H. CRISPR engineering in organoids for gene repair and disease modelling. *Nat Rev Bioeng.* 2023, *1*, 32-45.
- Corrò, C.; Novellasdemunt, L.; Li, V. S. W. A brief history of organoids. *Am J Physiol Cell Physiol.* 2020, 319, C151-C165.
- 20. Kim, W.; Gwon, Y.; Park, S.; Kim, H.; Kim, J. Therapeutic strategies of

three-dimensional stem cell spheroids and organoids for tissue repair and regeneration. Bioact Mater. 2023, 19, 50-74.

- 21. Panek, M.; Grabacka, M.; Pierzchalska, M. The formation of intestinal organoids in a hanging drop culture. Cytotechnology. 2018, 70, 1085-1095.
- 22. Lee, R. E.; Reidel, B.; Nelson, M. R.; Macdonald, J. K.; Kesimer, M.; Randell, S. H. Air-Liquid interface cultures to model drug delivery through the mucociliary epithelial barrier. Adv Drug Deliv Rev. 2023, 198, 114866.
- 23. Vandana, J. J.; Manrique, C.; Lacko, L. A.; Chen, S. Human pluripotentstem-cell-derived organoids for drug discovery and evaluation. Cell Stem Cell. 2023. 30. 571-591.
- 24. Novelli, G.; Spitalieri, P.; Murdocca, M.; Centanini, E.; Sangiuolo, F. Organoid factory: The recent role of the human induced pluripotent stem cells (hiPSCs) in precision medicine. Front Cell Dev Biol. 2022, 10, 1059579.
- 25. Shariati, L.; Esmaeili, Y.; Haghjooy Javanmard, S.; Bidram, E.; Amini, A. Organoid technology: current standing and future perspectives. Stem Cells. 2021, 39, 1625-1649.
- 26. Yang, Q.; Li, M.; Yang, X.; Xiao, Z.; Tong, X.; Tuerdi, A.; Li, S.; Lei, L. Flourishing tumor organoids: History, emerging technology, and application. Bioeng Transl Med. 2023, 8, e10559.
- 27. Thesleff, I.; Hurmerinta, K. Tissue interactions in tooth development. Differentiation. 1981, 18, 75-88.
- 28. Driehuis, E.; Kolders, S.; Spelier, S.; Lõhmussaar, K.; Willems, S. M.; Devriese, L. A.; de Bree, R.; de Ruiter, E. J.; Korving, J.; Begthel, H.; van Es, J. H.; Geurts, V.; He, G. W.; van Jaarsveld, R. H.; Oka, R.; Muraro, M. J.; Vivié, J.; Zandvliet, M.; Hendrickx, A. P. A.; Iakobachvili, N.; Sridevi, P.; Kranenburg, O.; van Boxtel, R.; Kops, G.; Tuveson, D. A.; Peters, P. J.; van Oudenaarden, A.; Clevers, H. Oral mucosal organoids as a potential platform for personalized cancer therapy. Cancer Discov. 2019, 9, 852-871.
- Hisha, H.; Tanaka, T.; Kanno, S.; Tokuyama, Y.; Komai, Y.; Ohe, 29. S.; Yanai, H.; Omachi, T.; Ueno, H. Establishment of a novel lingual organoid culture system: generation of organoids having mature keratinized epithelium from adult epithelial stem cells. Sci Rep. 2013, 3, 3224.
- 30. Zhao, H.; Jiang, E.; Shang, Z. 3D co-culture of cancer-associated fibroblast with oral cancer organoids. J Dent Res. 2021, 100, 201-208.
- 31. Jiang, Y.; Zhao, H.; Kong, S.; Zhou, D.; Dong, J.; Cheng, Y.; Zhang, S.; Wang, F.; Kalra, A.; Yang, N.; Wei, D. D.; Chen, J.; Zhang, Y. W.; Lin, D. C.; Meltzer, S. J.; Jiang, Y. Y. Establishing mouse and human oral esophageal organoids to investigate the tumor immune response. Dis Model Mech. 2024, 17, dmm050319.
- 32. Zhao, C.; Meng, C.; Cui, N.; Sha, J.; Sun, L.; Zhu, D. Organoid models for salivary gland biology and regenerative medicine. Stem Cells Int. 2021, 2021, 9922597.
- 33. Tanaka, J.; Ogawa, M.; Hojo, H.; Kawashima, Y.; Mabuchi, Y.; Hata, K.; Nakamura, S.; Yasuhara, R.; Takamatsu, K.; Irié, T.; Fukada, T.; Sakai, T.; Inoue, T.; Nishimura, R.; Ohara, O.; Saito, I.; Ohba, S.; Tsuji, T.; Mishima, K. Generation of orthotopically functional salivary gland from embryonic stem cells. Nat Commun. 2018, 9, 4216.
- 34. Pradhan-Bhatt, S.; Harrington, D. A.; Duncan, R. L.; Jia, X.; Witt, R. L.; Farach-Carson, M. C. Implantable three-dimensional salivary spheroid assemblies demonstrate fluid and protein secretory responses to neurotransmitters. Tissue Eng Part A. 2013, 19, 1610-1620.
- 35. Pradhan-Bhatt, S.; Harrington, D. A.; Duncan, R. L.; Farach-Carson, M. C.; Jia, X.; Witt, R. L. A novel in vivo model for evaluating functional

restoration of a tissue-engineered salivary gland. Laryngoscope. 2014, 124, 456-461.

- 36. Lombaert, I. M.; Brunsting, J. F.; Wierenga, P. K.; Faber, H.; Stokman, M. A.; Kok, T.; Visser, W. H.; Kampinga, H. H.; de Haan, G.; Coppes, R. P. Rescue of salivary gland function after stem cell transplantation in irradiated glands. PLoS One. 2008, 3, e2063.
- Farahat, M.; Sathi, G. A.; Hara, E. S.; Taketa, H.; Kuboki, T.; 37. Matsumoto, T. MSCs feeder layers induce SMG self-organization and branching morphogenesis. PLoS One. 2017, 12, e0176453.
- Hosseini, Z. F.; Nelson, D. A.; Moskwa, N.; Sfakis, L. M.; Castracane, J.; 38. Larsen, M. FGF2-dependent mesenchyme and laminin-111 are niche factors in salivary gland organoids. J Cell Sci. 2018, 131, jcs208728.
- 39. Nakao, K.; Morita, R.; Saji, Y.; Ishida, K.; Tomita, Y.; Ogawa, M.; Saitoh, M.; Tomooka, Y.; Tsuji, T. The development of a bioengineered organ germ method. Nat Methods. 2007, 4, 227-230.
- Ono, M.; Oshima, M.; Ogawa, M.; Sonoyama, W.; Hara, E. S.; Oida, 40. Y.; Shinkawa, S.; Nakajima, R.; Mine, A.; Hayano, S.; Fukumoto, S.; Kasugai, S.; Yamaguchi, A.; Tsuji, T.; Kuboki, T. Practical wholetooth restoration utilizing autologous bioengineered tooth germ transplantation in a postnatal canine model. Sci Rep. 2017, 7, 44522.
- 41. Wang, F.; Wu, Z.; Fan, Z.; Wu, T.; Wang, J.; Zhang, C.; Wang, S. The cell re-association-based whole-tooth regeneration strategies in large animal, Sus scrofa. Cell Prolif. 2018, 51, e12479.
- 42. Kim, E. J.; Yoon, K. S.; Arakaki, M.; Otsu, K.; Fukumoto, S.; Harada, H.; Green, D. W.; Lee, J. M.; Jung, H. S. Effective differentiation of induced pluripotent stem cells into dental cells. Dev Dyn. 2019, 248, 129-139.
- Keinan, D.; Cohen, R. E. The significance of epithelial rests of Malassez 43. in the periodontal ligament. J Endod. 2013, 39, 582-587.
- Shinmura, Y.; Tsuchiya, S.; Hata, K.; Honda, M. J. Quiescent epithelial 44. cell rests of Malassez can differentiate into ameloblast-like cells. J Cell Physiol. 2008, 217, 728-738.
- 45. Kim, H. Y.; Cooley, V.; Kim, E. J.; Li, S.; Lee, J. M.; Sheyfer, D.; Liu, W.; Klein, O. D.; Joester, D.; Jung, H. S. Adult dental epithelial stem cell-derived organoids deposit hydroxylapatite biomineral. Int J Oral Sci. 2023. 15. 55.
- Kilic Bektas, C.; Zhang, W.; Mao, Y.; Wu, X.; Kohn, J.; Yelick, P. C. 46. Self-Assembled Hydrogel Microparticle-Based Tooth-Germ Organoids. Bioengineering (Basel). 2022, 9, 215.
- 47. Calabrese, T. C.; Rothermund, K.; Gabe, C. M.; Beniash, E.; Davidson, L. A.; Syed-Picard, F. N. Self-assembly of tooth root organoid from postnatal human dental stem cells. Tissue Eng Part A. 2024, 30, 404-414.
- Adpaikar, A. A.; Zhang, S.; Kim, H. Y.; Kim, K. W.; Moon, S. J.; Lee, J. 48. M.; Jung, H. S. Fine-tuning of epithelial taste bud organoid to promote functional recapitulation of taste reactivity. Cell Mol Life Sci. 2022, 79, 211.
- 49. Lee, J. S.; Cho, A. N.; Jin, Y.; Kim, J.; Kim, S.; Cho, S. W. Bio-artificial tongue with tongue extracellular matrix and primary taste cells. Biomaterials. 2018, 151, 24-37.
- Fan, Y.; Cui, C.; Li, P.; Bi, R.; Lyu, P.; Li, Y.; Zhu, S. Fibrocartilage stem 50. cells in the temporomandibular joint: insights from animal and human studies. Front Cell Dev Biol. 2021, 9, 665995.
- 51. Crispim, J. F.; Ito, K. De novo neo-hyaline-cartilage from bovine organoids in viscoelastic hydrogels. Acta Biomater. 2021, 128, 236-249.
- Dönges, L.; Damle, A.; Mainardi, A.; Bock, T.; Schönenberger, M.; 52. Martin, I.; Barbero, A. Engineered human osteoarthritic cartilage organoids. Biomaterials. 2024, 308, 122549.
- Yoshimoto, S.; Yoshizumi, J.; Anzai, H.; Morishita, K.; Okamura, K.; 53. Hiraki, A.; Hashimoto, S. Inhibition of Alk signaling promotes the

Biomaterials Translational

- 54. Tanaka, J.; Mishima, K. In vitro three-dimensional culture systems of salivary glands. *Pathol Int.* **2020**, *70*, 493-501.
- Yoon, A. J.; Santella, R. M.; Wang, S.; Kutler, D. I.; Carvajal, R. D.; Philipone, E.; Wang, T.; Peters, S. M.; Stewart, C. R.; Momen-Heravi, F.; Troob, S.; Levin, M.; AkhavanAghdam, Z.; Shackelford, A. J.; Canterbury, C. R.; Shimonosono, M.; Hernandez, B. Y.; McDowell, B. D.; Nakagawa, H. MicroRNA-based cancer mortality risk scoring system and hTERT expression in early-stage oral squamous cell carcinoma. *J Oncol.* 2021, 2021, 8292453.
- Chitturi Suryaprakash, R. T.; Kujan, O.; Shearston, K.; Farah,
 C. S. Three-dimensional cell culture models to investigate oral carcinogenesis: a scoping review. *Int J Mol Sci.* 2020, *21*, 9520.
- Shopova, D.; Yaneva, A.; Mihaylova, A.; Dinkova, A.; Bakova, D. Unlocking the future: bioprinting salivary glands-from possibility to reality. *J Funct Biomater.* 2024, *15*, 151.
- Shopova, D.; Mihaylova, A.; Yaneva, A.; Bakova, D. Advancing dentistry through bioprinting: personalization of oral tissues. *J Funct Biomater.* 2023, *14*, 530.
- 59. Chansaenroj, A.; Yodmuang, S.; Ferreira, J. N. Trends in salivary gland tissue engineering: from stem cells to secretome and organoid bioprinting. *Tissue Eng Part B Rev.* **2021**, *27*, 155-165.
- 60. Wang, Y.; Sun, Y. Engineered organoids in oral and maxillofacial regeneration. *iScience*. **2023**, *26*, 105757.
- 61. AR, Y. B.; Casasco, A.; Monti, M. Hypes and hopes of stem cell therapies in dentistry: a review. *Stem Cell Rev Rep.* **2022**, *18*, 1294-1308.
- Jeong, S. Y.; Lee, S.; Choi, W. H.; Jee, J. H.; Kim, H. R.; Yoo, J. Fabrication of dentin-pulp-like organoids using dental-pulp stem cells. *Cells.* 2020, 9, 642.
- Popowics, T.; Mulimani, P. Mammalian dental diversity: an evolutionary template for regenerative dentistry. *Front Dent Med.* 2023, 4, 1158482.
- Xu, X.; Li, Z.; Ai, X.; Tang, Y.; Yang, D.; Dou, L. Human threedimensional dental pulp organoid model for toxicity screening of dental materials on dental pulp cells and tissue. *Int Endod J.* 2022, *55*, 79-88.
- 65. Manokawinchoke, J.; Limraksasin, P.; Okawa, H.; Pavasant, P.; Egusa, H.; Osathanon, T. Intermittent compressive force induces cell cycling and reduces apoptosis in embryoid bodies of mouse induced pluripotent stem cells. *Int J Oral Sci.* 2022, *14*, 1.
- Liu, X.; Tong, X.; Zhu, J.; Tian, L.; Jie, Z.; Zou, Y.; Lin, X.; Liang, H.;
 Li, W.; Ju, Y.; Qin, Y.; Zou, L.; Lu, H.; Zhu, S.; Jin, X.; Xu, X.; Yang, H.;
 Wang, J.; Zong, Y.; Liu, W.; Hou, Y.; Jia, H.; Zhang, T. Metagenomegenome-wide association studies reveal human genetic impact on the oral microbiome. *Cell Discov.* 2021, *7*, 117.
- 67. Morrison, A. G.; Sarkar, S.; Umar, S.; Lee, S. T. M.; Thomas, S. M. The contribution of the human oral microbiome to oral disease: a review. *Microorganisms*. **2023**, *11*, 318.
- Monasterio, G.; Morales, R. A.; Bejarano, D. A.; Abalo, X. M.; Fransson, J.; Larsson, L.; Schlitzer, A.; Lundeberg, J.; Das, S.; Villablanca, E. J. A versatile tissue-rolling technique for spatial-omics analyses of the entire murine gastrointestinal tract. *Nat Protoc.* 2024, 19, 3085-3137.
- Plachokova, A. S.; Andreu-Sánchez, S.; Noz, M. P.; Fu, J.; Riksen, N. P. Oral microbiome in relation to periodontitis severity and systemic inflammation. *Int J Mol Sci.* 2021, *22*, 5876.
- 70. Herremans, K. M.; Riner, A. N.; Cameron, M. E.; McKinley, K. L.; Triplett, E. W.; Hughes, S. J.; Trevino, J. G. The oral microbiome,

pancreatic cancer and human diversity in the age of precision medicine. *Microbiome.* **2022**, *10*, 93.

- Forbester, J. L.; Goulding, D.; Vallier, L.; Hannan, N.; Hale, C.; Pickard, D.; Mukhopadhyay, S.; Dougan, G. Interaction of salmonella enterica serovar typhimurium with intestinal organoids derived from human induced pluripotent stem cells. *Infect Immun.* 2015, *83*, 2926-2934.
- Richiardone, E.; Van den Bossche, V.; Corbet, C. Metabolic studies in organoids: current applications, opportunities and challenges. *Organoids*. 2022, *1*, 85-105.
- 73. Hemeryck, L.; Hermans, F.; Chappell, J.; Kobayashi, H.; Lambrechts, D.; Lambrichts, I.; Bronckaers, A.; Vankelecom, H. Organoids from human tooth showing epithelial stemness phenotype and differentiation potential. *Cell Mol Life Sci.* 2022, *79*, 153.
- 74. Song, Y. M.; Na, K. H.; Lee, H. J.; Park, J. B. The effects of transforming growth factor-β1 on the differentiation of cell organoids composed of gingiva-derived stem cells. *Biomed Res Int.* 2022, 2022, 9818299.
- 75. Hof, L.; Moreth, T.; Koch, M.; Liebisch, T.; Kurtz, M.; Tarnick, J.; Lissek, S. M.; Verstegen, M. M. A.; van der Laan, L. J. W.; Huch, M.; Matthäus, F.; Stelzer, E. H. K.; Pampaloni, F. Long-term live imaging and multiscale analysis identify heterogeneity and core principles of epithelial organoid morphogenesis. *BMC Biol.* 2021, *19*, 37.
- 76. Rosenbluth, J. M.; Schackmann, R. C. J.; Gray, G. K.; Selfors, L. M.; Li, C. M.; Boedicker, M.; Kuiken, H. J.; Richardson, A.; Brock, J.; Garber, J.; Dillon, D.; Sachs, N.; Clevers, H.; Brugge, J. S. Organoid cultures from normal and cancer-prone human breast tissues preserve complex epithelial lineages. *Nat Commun.* **2020**, *11*, 1711.
- Bernal, P. N.; Bouwmeester, M.; Madrid-Wolff, J.; Falandt, M.; Florczak, S.; Rodriguez, N. G.; Li, Y.; Größbacher, G.; Samsom, R. A.; van Wolferen, M.; van der Laan, L. J. W.; Delrot, P.; Loterie, D.; Malda, J.; Moser, C.; Spee, B.; Levato, R. Volumetric bioprinting of organoids and optically tuned hydrogels to build liver-like metabolic biofactories. *Adv Mater.* **2022**, *34*, e2110054.
- Guan, Y.; Enejder, A.; Wang, M.; Fang, Z.; Cui, L.; Chen, S. Y.; Wang, J.; Tan, Y.; Wu, M.; Chen, X.; Johansson, P. K.; Osman, I.; Kunimoto, K.; Russo, P.; Heilshorn, S. C.; Peltz, G. A human multi-lineage hepatic organoid model for liver fibrosis. *Nat Commun.* 2021, *12*, 6138.
- Dey, M.; Ozbolat, I. T. 3D bioprinting of cells, tissues and organs. *Sci Rep.* 2020, *10*, 14023.
- Ren, Y.; Yang, X.; Ma, Z.; Sun, X.; Zhang, Y.; Li, W.; Yang, H.; Qiang, L.; Yang, Z.; Liu, Y.; Deng, C.; Zhou, L.; Wang, T.; Lin, J.; Li, T.; Wu, T.; Wang, J. Developments and opportunities for 3D bioprinted organoids. *Int J Bioprint.* 2021, *7*, 364.
- Reid, J. A.; Palmer, X. L.; Mollica, P. A.; Northam, N.; Sachs, P. C.; Bruno, R. D. A 3D bioprinter platform for mechanistic analysis of tumoroids and chimeric mammary organoids. *Sci Rep.* 2019, *9*, 7466.
- 82. Rossi, G.; Manfrin, A.; Lutolf, M. P. Progress and potential in organoid research. *Nat Rev Genet.* **2018**, *19*, 671-687.
- Molander, D.; Sbirkov, Y.; Bodurov, I.; Dikov, D.; Sarafian, V. Comparative analysis of bioinks in 3D bioprinted organoids of colorectal cancer. *Acta Morphol Anthropol.* 2022, *29*, 17-21.
- 84. Ma, J.; Wang, Y.; Liu, J. Bioprinting of 3D tissues/organs combined with microfluidics. *RSC Adv.* **2018**, *8*, 21712-21727.
- Maloney, E.; Clark, C.; Sivakumar, H.; Yoo, K.; Aleman, J.; Rajan, S. A. P.; Forsythe, S.; Mazzocchi, A.; Laxton, A. W.; Tatter, S. B.; Strowd, R. E.; Votanopoulos, K. I.; Skardal, A. Immersion bioprinting of tumor organoids in multi-well plates for increasing chemotherapy screening throughput. *Micromachines (Basel)*. 2020, *11*, 208.
- 86. Adine, C.; Ferreira, J. Bioprinting strategies to engineer functional

salivary gland organoids. In *Organ tissue engineering*, Eberli, D.; Lee, S. J.; Traweger, A., eds.; Springer International Publishing: Cham, 2021; pp 173-194.

- 87. Klangprapan, J.; Souza, G. R.; Ferreira, J. N. Bioprinting salivary gland models and their regenerative applications. *BDJ Open.* **2024**, *10*, 39.
- Meijer, E. M.; Koch, S. E.; van Dijk, C. G. M.; Maas, R. G. C.; Chrifi, I.; Szymczyk, W.; Besseling, P. J.; Pomp, L.; Koomen, V.; Buikema, J. W.; Bouten, C. V. C.; Verhaar, M. C.; Smits, A.; Cheng, C. 3D human iPSC blood vessel organoids as a source of flow-adaptive vascular cells for creating a human-relevant 3D-scaffold based macrovessel model. *Adv Biol (Weinh).* 2023, *7*, e2200137.
- Li, A.; Sasaki, J. I.; Abe, G. L.; Katata, C.; Sakai, H.; Imazato, S. Vascularization of a bone organoid using dental pulp stem cells. *Stem Cells Int.* 2023, 2023, 5367887.
- Zhao, X.; Zhang, Z.; Luo, Y.; Ye, Q.; Shi, S.; He, X.; Zhu, J.; Zhu, Q.; Zhang, D.; Xia, W.; Zhang, Y.; Jiang, L.; Cui, L.; Ye, Y.; Xiang, Y.; Hu, J.; Zhang, J.; Lin, C. P. Trophoblast and blood vessel organoid cultures recapitulate the role of WNT2B in promoting intravillous vascularization in human intrauterine and ectopic pregnancy. *bioRxiv.* 2022. doi: 10.1101/2022.04.18.488605.
- Holkom, M.; Yang, X.; Li, R.; Chen, Y.; Zhao, H.; Shang, Z. Fibroblast regulates angiogenesis in assembled oral cancer organoid: A possible role of NNMT. *Oral Dis.* 2024. doi: 10.1111/odi.14945.
- 92. Kim, J. W.; Nam, S. A.; Yi, J.; Kim, J. Y.; Lee, J. Y.; Park, S. Y.; Sen, T.; Choi, Y. M.; Lee, J. Y.; Kim, H. L.; Kim, H. W.; Park, J.; Cho, D. W.; Kim, Y. K. Kidney decellularized extracellular matrix enhanced the vascularization and maturation of human kidney organoids. *Adv Sci* (*Weinh*). 2022, 9, e2103526.
- 93. Li, Y. E.; Jodat, Y. A.; Samanipour, R.; Zorzi, G.; Zhu, K.; Hirano, M.; Chang, K.; Arnaout, A.; Hassan, S.; Matharu, N.; Khademhosseini, A.; Hoorfar, M.; Shin, S. R. Toward a neurospheroid niche model: optimizing embedded 3D bioprinting for fabrication of neurospheroid brain-like co-culture constructs. *Biofabrication*. 2020, *13*, 015014.
- Del Dosso, A.; Urenda, J. P.; Nguyen, T.; Quadrato, G. Upgrading the physiological relevance of human brain organoids. *Neuron*. 2020, 107, 1014-1028.
- He, Z.; Dony, L.; Fleck, J. S.; Szałata, A.; Li, K. X.; Slišković,
 I.; Lin, H. C.; Santel, M.; Atamian, A.; Quadrato, G.; Sun, J.;
 Paşca, S. P.; Camp, J. G.; Theis, F.; Treutlein, B. An integrated
 transcriptomic cell atlas of human neural organoids. *bioRxiv*. 2023. doi: 10.1101/2023.10.05.561097.
- Jiang, F.; Jiang, Y.; Zhi, H.; Dong, Y.; Li, H.; Ma, S.; Wang, Y.; Dong, Q.; Shen, H.; Wang, Y. Artificial intelligence in healthcare: past, present and future. *Stroke Vasc Neurol.* 2017, *2*, 230-243.
- 97. Miller, G. A. The cognitive revolution: a historical perspective. *Trends Cogn Sci.* **2003**, *7*, 141-144.
- Joint Research Centre. *Historical evolution of artificial intelligence*. Luxembourg: Publications Office of the European Union: 2020.
- 99. Tripathi, K. P. A review on knowledge-based expert system: concept and architecture. *Artif Intell Tech.* **2021**, *AIT*, 21-25.
- 100. Bahrammirzaee, A. A comparative survey of artificial intelligence applications in finance: artificial neural networks, expert system and hybrid intelligent systems. *Neural Comput Applic.* **2010**, *19*, 1165-1195.
- 101. Zou, J.; Huss, M.; Abid, A.; Mohammadi, P.; Torkamani, A.; Telenti, A. A primer on deep learning in genomics. *Nat Genet.* **2019**, *51*, 12-18.
- 102. Alkhayrat, M.; Aljnidi, M.; Aljoumaa, K. A comparative dimensionality reduction study in telecom customer segmentation using deep learning and PCA. *J Big Data*. **2020**, *7*, 9.

- 103. Saito, Y.; Oikawa, M.; Nakazawa, H.; Niide, T.; Kameda, T.; Tsuda, K.; Umetsu, M. Machine-learning-guided mutagenesis for directed evolution of fluorescent proteins. ACS synthetic biology. 2018, 7, 2014-2022.
- 104. Nagarhalli, T. P.; Vaze, V.; Rana, N. K. In *Impact of Machine Learning in Natural Language Processing: A Review*, 2021 Third International Conference on Intelligent Communication Technologies and Virtual Mobile Networks (ICICV), 4-6 Feb. 2021; 2021; pp 1529-1534.
- 105. Furtado, F.; Singh, A. Movie recommendation system using machine learning. *Int J Res Ind.* **2020**, *9*, 84-98.
- Sharma, M.; Savage, C.; Nair, M.; Larsson, I.; Svedberg, P.; Nygren, J.
 M. Artificial Intelligence Applications in Health Care Practice: Scoping Review. J Med Internet Res. 2022, 24, e40238.
- 107. Pushpanathan, K.; Hanafi, M.; Mashohor, S.; Fazlil Ilahi, W. F.
 Machine learning in medicinal plants recognition: a review. *Artif Intell Rev.* 2021, *54*, 305-327.
- Maramraju, S.; Kowalczewski, A.; Kaza, A.; Liu, X.; Singaraju, J. P.; Albert, M. V.; Ma, Z.; Yang, H. AI-organoid integrated systems for biomedical studies and applications. *Bioeng Transl Med.* 2024, *9*, e10641.
- 109. Chen, K. Y.; Srinivasan, T.; Lin, C.; Tung, K. L.; Gao, Z.; Hsu, D. S.; Lipkin, S. M.; Shen, X. Single-cell transcriptomics reveals heterogeneity and drug response of human colorectal cancer organoids. *Annu Int Conf IEEE Eng Med Biol Soc.* 2018, 2018, 2378-2381.
- 110. Jordan, M. I.; Mitchell, T. M. Machine learning: Trends, perspectives, and prospects. *Science*. **2015**, *349*, 255-260.
- 111. Feher, B.; Tussie, C.; Giannobile, W. V. Applied artificial intelligence in dentistry: emerging data modalities and modeling approaches. *Front Artif Intell.* 2024, *7*, 1427517.
- 112. Mukherjee, S.; Yadav, G.; Kumar, R. Recent trends in stem cell-based therapies and applications of artificial intelligence in regenerative medicine. *World J Stem Cells.* **2021**, *13*, 521-541.
- 113. LeCun, Y.; Bengio, Y.; Hinton, G. Deep learning. *Nature*. **2015**, *521*, 436-444.
- 114. van der Laak, J.; Litjens, G.; Ciompi, F. Deep learning in histopathology: the path to the clinic. *Nat Med.* 2021, *27*, 775-784.
- 115. Purwono, P.; Ma; #039; arif, A.; Rahmaniar, W.; Fathurrahman, H. I. K.; Frisky, A. Z. K.; Haq, Q. M. U. Understanding of convolutional neural network (CNN): a review. *Int J Robot Control Syst.* **2023**, *2*, 739-748.
- 116. Park, C.; Took, C. C.; Seong, J. K. Machine learning in biomedical engineering. *Biomed Eng Lett.* **2018**, *8*, 1-3.
- 117. Al-Askar, H.; Radi, N.; MacDermott, Á. Chapter 7 Recurrent neural networks in medical data analysis and classifications. In *Applied computing in medicine and health*, Al-Jumeily, D.; Hussain, A.; Mallucci, C.; Oliver, C., eds.; Morgan Kaufmann: Boston, 2016; pp 147-165.
- 118. Kassis, T.; Hernandez-Gordillo, V.; Langer, R.; Griffith, L. G. OrgaQuant: human intestinal organoid localization and quantification using deep convolutional neural networks. *Sci Rep.* 2019, *9*, 12479.
- 119. Shin, J.; Lee, Y.; Li, Z.; Hu, J.; Park, S. S.; Kim, K. Optimized 3D bioprinting technology based on machine learning: a review of recent trends and advances. *Micromachines (Basel)*. 2022, *13*, 363.
- 120. Cao, Y.; Zhang, X.; Chen, Q.; Rao, X.; Qiu, E.; Wu, G.; Lin, Y.; Zeng, Z.; Zheng, B.; Li, Z.; Cai, Z.; Wang, H.; Han, S. Patient-derived organoid facilitating personalized medicine in gastrointestinal stromal tumor with liver metastasis: a case report. *Front Oncol.* 2022, *12*, 920762.
- 121. Wang, H. M.; Zhang, C. Y.; Peng, K. C.; Chen, Z. X.; Su, J. W.; Li, Y. F.; Li, W. F.; Gao, Q. Y.; Zhang, S. L.; Chen, Y. Q.; Zhou, Q.; Xu, C.; Xu, C. R.; Wang, Z.; Su, J.; Yan, H. H.; Zhang, X. C.; Chen, H. J.; Wu,

Y. L.; Yang, J. J. Using patient-derived organoids to predict locally advanced or metastatic lung cancer tumor response: a real-world study. *Cell Rep Med.* **2023**, *4*, 100911.

- 122. Lampart, F. L.; Iber, D.; Doumpas, N. Organoids in high-throughput and high-content screenings. *Front Chem Eng.* **2023**, *5*, 1120348.
- 123. Gritti, N.; Lim, J. L.; Anlaş, K.; Pandya, M.; Aalderink, G.; Martínez-Ara, G.; Trivedi, V. MOrgAna: accessible quantitative analysis of organoids with machine learning. *Development*. 2021, 148, dev199611.
- 124. Sharick, J. T.; Walsh, C. M.; Sprackling, C. M.; Pasch, C. A.; Pham,
 D. L.; Esbona, K.; Choudhary, A.; Garcia-Valera, R.; Burkard, M.
 E.; McGregor, S. M.; Matkowskyj, K. A.; Parikh, A. A.; Meszoely,
 I. M.; Kelley, M. C.; Tsai, S.; Deming, D. A.; Skala, M. C. Metabolic heterogeneity in patient tumor-derived organoids by primary site and drug treatment. *Front Oncol.* 2020, *10*, 553.
- 125. Natarajan, V.; Simoneau, C. R.; Erickson, A. L.; Meyers, N. L.; Baron, J. L.; Cooper, S.; McDevitt, T. C.; Ott, M. Modelling T-cell immunity against hepatitis C virus with liver organoids in a microfluidic coculture system. *Open Biol.* **2022**, *12*, 210320.
- 126. Takahashi, T. Organoids for drug discovery and personalized medicine. *Annu Rev Pharmacol Toxicol.* **2019**, *59*, 447-462.
- 127. Park, K.; Lee, J. Y.; Lee, S. Y.; Jeong, I.; Park, S. Y.; Kim, J. W.; Nam, S. A.; Kim, H. W.; Kim, Y. K.; Lee, S. Deep learning predicts the differentiation of kidney organoids derived from human induced pluripotent stem cells. *Kidney Res Clin Pract.* **2023**, *42*, 75-85.
- 128. Deben, C.; De La Hoz, E. C.; Compte, M. L.; Van Schil, P.; Hendriks, J. M. H.; Lauwers, P.; Yogeswaran, S. K.; Lardon, F.; Pauwels, P.; Van Laere, S.; Bogaerts, A.; Smits, E.; Vanlanduit, S.; Lin, A. OrBITS: label-free and time-lapse monitoring of patient derived organoids for advanced drug screening. *Cell Oncol (Dordr).* **2023**, *46*, 299-314.
- 129. Branciforti, F.; Salvi, M.; D'Agostino, F.; Marzola, F.; Cornacchia, S.; De Titta, M. O.; Mastronuzzi, G.; Meloni, I.; Moschetta, M.; Porciani, N.; Sciscenti, F.; Spertini, A.; Spilla, A.; Zagaria, I.; Deloria, A. J.; Deng, S.; Haindl, R.; Szakacs, G.; Csiszar, A.; Liu, M.; Drexler, W.; Molinari, F.; Meiburger, K. M. Segmentation and multi-timepoint tracking of 3D cancer organoids from optical coherence tomography images using deep neural networks. *Diagnostics (Basel).* **2024**, *14*, 1217.
- 130. Cai, H.; Ao, Z.; Tian, C.; Wu, Z.; Liu, H.; Tchieu, J.; Gu, M.; Mackie, K.; Guo, F. Brain organoid computing for artificial intelligence. *bioRxiv*. 2023. doi: 10.1101/2023.02.28.530502.
- 131. Kegeles, E.; Naumov, A.; Karpulevich, E. A.; Volchkov, P.; Baranov,
 P. Convolutional neural networks can predict retinal differentiation in retinal organoids. *Front Cell Neurosci.* 2020, *14*, 171.
- 132. Esmail, S.; Danter, W. R. NEUBOrg: artificially induced pluripotent stem cell-derived brain organoid to model and study genetics of alzheimer's disease progression. *Front Aging Neurosci.* **2021**, *13*, 643889.
- 133. Monzel, A. S.; Hemmer, K.; Kaoma, T.; Smits, L. M.; Bolognin, S.; Lucarelli, P.; Rosety, I.; Zagare, A.; Antony, P.; Nickels, S. L.; Krueger, R.; Azuaje, F.; Schwamborn, J. C. Machine learning-assisted neurotoxicity prediction in human midbrain organoids. *Parkinsonism Relat Disord.* 2020, *75*, 105-109.
- 134. Phan, N.; Hong, J. J.; Tofig, B.; Mapua, M.; Elashoff, D.; Moatamed, N. A.; Huang, J.; Memarzadeh, S.; Damoiseaux, R.; Soragni, A. A simple high-throughput approach identifies actionable drug sensitivities in patient-derived tumor organoids. *Commun Biol.* 2019, 2, 78.
- 135. Takahashi, Y.; Inoue, Y.; Sato, S.; Okabe, T.; Kojima, H.; Kiyono, H.; Shimizu, M.; Yamauchi, Y.; Sato, R. Drug cytotoxicity screening using human intestinal organoids propagated with extensive cost-reduction strategies. *Sci Rep.* 2023, *13*, 5407.

- Zhang, X. Y.; Sui, Y.; Shan, X. F.; Wang, L. M.; Zhang, L.; Xie, S.; Cai,
 Z. G. Construction of oral squamous cell carcinoma organoids in vitro
 3D-culture for drug screening. *Oral Dis.* 2024. doi: 10.1111/odi.15044.
- 137. Schuster, B.; Junkin, M.; Kashaf, S. S.; Romero-Calvo, I.; Kirby, K.; Matthews, J.; Weber, C. R.; Rzhetsky, A.; White, K. P.; Tay, S. Automated microfluidic platform for dynamic and combinatorial drug screening of tumor organoids. *Nat Commun.* **2020**, *11*, 5271.
- 138. Trisno, S. L.; Philo, K. E. D.; McCracken, K. W.; Catá, E. M.; Ruiz-Torres, S.; Rankin, S. A.; Han, L.; Nasr, T.; Chaturvedi, P.; Rothenberg, M. E.; Mandegar, M. A.; Wells, S. I.; Zorn, A. M.; Wells, J. M. Esophageal organoids from human pluripotent stem cells delineate Sox2 functions during esophageal specification. *Cell Stem Cell*. **2018**, *23*, 501-515.e7.
- 139. Ahn, S. J.; Lee, S.; Kwon, D.; Oh, S.; Park, C.; Jeon, S.; Lee, J. H.; Kim, T. S.; Oh, I. U. Essential guidelines for manufacturing and application of organoids. *Int J Stem Cells*. **2024**, *17*, 102-112.
- 140. Abdul, N. S.; Shivakumar, G. C.; Sangappa, S. B.; Di Blasio, M.; Crimi, S.; Cicciù, M.; Minervini, G. Applications of artificial intelligence in the field of oral and maxillofacial pathology: a systematic review and metaanalysis. *BMC Oral Health.* **2024**, *24*, 122.
- 141. Ravn, T.; Sørensen, M. P.; Capulli, E.; Kavouras, P.; Pegoraro, R.; Picozzi, M.; Saugstrup, L. I.; Spyrakou, E.; Stavridi, V. Public perceptions and expectations: disentangling the hope and hype of organoid research. *Stem Cell Reports.* **2023**, *18*, 841-852.
- 142. Farhud, D. D.; Zokaei, S. Ethical issues of artificial intelligence in medicine and healthcare. *Iran J Public Health*. **2021**, *50*, i-v.
- 143. Kluge, E. W. Artificial intelligence in healthcare: ethical considerations. *Healthc Manage Forum.* **2020**, *33*, 47-49.
- 144. Mahmood, U.; Shukla-Dave, A.; Chan, H. P.; Drukker, K.; Samala, R. K.; Chen, Q.; Vergara, D.; Greenspan, H.; Petrick, N.; Sahiner, B.; Huo, Z.; Summers, R. M.; Cha, K. H.; Tourassi, G.; Deserno, T. M.; Grizzard, K. T.; Näppi, J. J.; Yoshida, H.; Regge, D.; Mazurchuk, R.; Suzuki, K.; Morra, L.; Huisman, H.; Armato, S. G., 3rd; Hadjiiski, L. Artificial intelligence in medicine: mitigating risks and maximizing benefits via quality assurance, quality control, and acceptance testing. *BJR Artif Intell.* 2024, *1*, ubae003.
- 145. Ranjbar, A.; Mork, E. W.; Ravn, J.; Brøgger, H.; Myrseth, P.; Østrem, H. P.; Hallock, H. Managing risk and quality of ai in healthcare: are hospitals ready for implementation? *Risk Manag Healthc Policy*. 2024, *17*, 877-882.
- 146. Aldoihi, S.; Alblalaihid, K.; Alzemaia, F.; Almoajel, A. In *The Role of System Modeling on Artificial Intelligence: A Review of Emerging Trends*, 2023 IEEE International Conference on Metrology for eXtended Reality, Artificial Intelligence and Neural Engineering (MetroXRAINE), 25-27 Oct. 2023; 2023; pp 161-165.
- 147. Altukroni, A.; Alsaeedi, A.; Gonzalez-Losada, C.; Lee, J. H.; Alabudh, M.; Mirah, M.; El-Amri, S.; Ezz El-Deen, O. Detection of the pathological exposure of pulp using an artificial intelligence tool: a multicentric study over periapical radiographs. *BMC Oral Health.* 2023, 23, 553.
- 148. Olczak, J.; Pavlopoulos, J.; Prijs, J.; Ijpma, F. F. A.; Doornberg, J. N.; Lundström, C.; Hedlund, J.; Gordon, M. Presenting artificial intelligence, deep learning, and machine learning studies to clinicians and healthcare stakeholders: an introductory reference with a guideline and a Clinical AI Research (CAIR) checklist proposal. *Acta Orthop.* 2021, 92, 513-525.
- Sikri, A.; Sikri, J.; Gupta, R. Artificial intelligence in prosthodontics and oral implantology – a narrative review. *lob Acad J Dent Oral Health.* 2023,

5, 13-19.

- 150. García-Pola, M.; Pons-Fuster, E.; Suárez-Fernández, C.; Seoane-Romero, J.; Romero-Méndez, A.; López-Jornet, P. Role of artificial intelligence in the early diagnosis of oral cancer. A scoping review. *Cancers (Basel).* 2021, 13, 4600.
- 151. Chakraborty, D.; Ghosh, D.; Kumar, S.; Jenkins, D.; Chandrasekaran, N.; Mukherjee, A. Nano-diagnostics as an emerging platform for oral cancer detection: current and emerging trends. Wiley Interdiscip Rev Nanomed Nanobiotechnol. 2023, 15, e1830.
- Patil, S.; Albogami, S.; Hosmani, J.; Mujoo, S.; Kamil, M. A.; Mansour, M. A.; Abdul, H. N.; Bhandi, S.; Ahmed, S. Artificial intelligence in the diagnosis of oral diseases: applications and pitfalls. *Diagnostics (Basel)*.
 2022, 12, 1029.
- 153. Li, C.; Zhou, H.; Gou, H.; Fan, Z.; Zhang, Y.; Tang, P.; Huang, J.; Xu, Y.; Li, L. Autoinducer-2 produced by oral microbial flora and alveolar bone loss in periodontitis. *J Periodontal Res.* 2024, *59*, 576-588.
- 154. Lee, Y. H. Application of artificial intelligence for the management of oral diseases. *J Oral Med Pain*. **2022**, *47*, 107-108.

Biomaterials Translational

- 155. Li, F.; Zhang, P.; Wu, S.; Yuan, L.; Liu, Z. Advance in human epithelialderived organoids research. *Mol Pharm.* **2021**, *18*, 3931-3950.
- 156. Fatima, I.; Grover, V.; Raza Khan, I.; Ahmad, N.; Yadav, A. Artificial intelligence in medical field. *EAI Endorsed Trans Pervasive Health Technol.* 2023, 9, 4713.
- 157. Weerarathna, I. N.; Kamble, A. R.; Luharia, A. Artificial intelligence applications for biomedical cancer research: a review. *Cureus.* 2023, 15, e48307.
- 158. Mun, S. J.; Hong, Y. H.; Shin, Y.; Lee, J.; Cho, H. S.; Kim, D. S.; Chung, K. S.; Son, M. J. Efficient and reproducible generation of human induced pluripotent stem cell-derived expandable liver organoids for disease modeling. *Sci Rep.* 2023, *13*, 22935.

Received: September 28, 2024 Revised: October 20, 2024 Accepted: November 1, 2024 Available online: November 15, 2024