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## **Manipulating Oxidative Stress Following Ionizing Radiation**

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It is now well accepted that the ionizing radiation-generated reactive oxygen species (ROS), that constitute ~2/3 of the effects of external beam radiation, do not only produce direct tumor cell death, but also affect the surrounding microenvironment. Moreover, this indirect effect of radiation may result in systemic effects, specifically the initiation of an inflammatory response.

We have previously shown the existence of an endothelial-stem cell linked pathway that is activated by single-high-dose radiotherapy (SDRT) (>8 Gy/fraction). This endothelial-stem cell linkage pathway mediates normal tissue injury after SDRT of the intestines, lung and salivary glands [1–3], suggesting that this represents a generic response mechanism for mammalian tissue damage injury by large single-dose irradiation, i.e. ablative radiation therapy. Prior studies reported by Moeller et al. [4] provided strong evidence that bursts of ROS are generated by waves of hypoxia/reoxygenation that occur after each radiation exposure in response to conventional fractionated radiotherapy in tumors. Recently, Mizrachi et al. [3] demonstrated that SDRT-induced salivary glands (SG) hypofunction was to a large extent mediated by microvascular dysfunction involving ceramide and ROS generation.

ROS generation is important in the maintenance of homeostasis between pro-apoptotic and pro-survival signals [5–7]. However, enhanced accumulation of ROS generates chronic pathological conditions. While a large number of studies on oxidative stress focused on the mitochondria-generated ROS, Wortel et al. [8] in their recent study confirmed that the endothelium is one of the major sources of ROS and identified the different types of ROS generated by the SDRT-induced oxidative stress, both within the plasma membrane and in the cytosol of endothelial cells (Figure 1). Similar findings were previously reported by our

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collaborators in the same cells in response to Fas ligand, tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), endostatin, and homocysteine [9–15].

Ours as well as others previous studies have shown that the endothelial cells within the different tissues are the most sensitive cells to the effects of ionizing radiation (IR), since they are 20-fold enriched in secretory acid sphingomyelinase (ASMase), as compared to any other cell type in the body [16]. We also showed that ceramide is a sphingolipid messenger capable of initiating apoptotic cascades in response to various stressful stimuli, including IR [17,18]. IR induced alterations in the plasma membrane hydrolyzes sphingomyelin to generate ceramide via sphingomyelinases activation [19,20]. The sphingomyelinases are expressed preferentially in the vascular endothelium [21], suggesting that these mechanisms may be of particular relevance for vascular structure and function. Furthermore, data derived from acid sphingomyelinase-knock-out mice, showed that they have a radioresistant vasculature and are partially protected from end-organ radiation injury [2,22] emphasizing the biological significance of this phenomenon.

Advances in cancer diagnosis and treatment have led to increases in life expectancy, bringing forward the issue of long-term treatment-related morbidity and mortality in these patients. Numerous reports have concluded that these patients should be regarded as long-term cancer survivors rather than as healthy individuals due to their long-term risk of developing treatment-related adverse events, and in particular cardiovascular events. With this clinical background in mind, several groups have embarked in the search for the mechanism(s) by which these effects are inflicted upon the cardiovascular system with special attention to the vascular wall and the vascular endothelium [23–26].

Radiation induces microvascular dysfunction via activation of the acid sphingomyelinase (ASMase)/ceramide pathway. Microvascular dysfunction is crucial for tumor response to radiation. ASMase activation triggers the generation of ceramide-rich platforms (CRPs), NADPH oxidase (NOX) activation and subsequent production of ROS, resulting in microvascular endothelial dysfunction (Figure 1) [27]. Elevated ROS formation in the vascular wall is a key feature of all cardiovascular diseases and a likely contributor to endothelial cell dysfunction, vascular inflammation and plaque formation. The NOX family of enzymes comprises seven members (NOX1-5, DUOX1-2), with each one of them displaying distinct patterns of expression, intracellular compartmentalization, regulation, and biological function. NOX-derived ROS control multiple aspects of cell physiology via redox-activated signaling pathways. Nevertheless, NOX over-activity, a condition that is typically associated with significant up-regulation of its expression, has been increasingly reported in a variety of cardiovascular diseases [28]. Numerous studies have demonstrated that expression and activity of at least two isoforms of NADPH oxidase - NOX1 and NOX2 - is increased in animal models of hypertension, diabetes and atherosclerosis. Several studies in transgenic mice support the role for NOX1- and/or NOX2-containing oxidases as sources of excessive vascular ROS production and as triggers of endothelial cell dysfunction in hypertension, atherosclerosis and diabetes [29].

The apolipoprotein-E-deficient (ApoE-/-) mouse is the most widely studied animal model of hypercholesterolemia and atherosclerosis [30,31]. It has been reported that ApoE-/-/

p47<sup>phox</sup>\_/– double KO are protected, compared to the ApoE\_/– single KO mice, from the development of aortic atherosclerotic lesions [32–34]. Moreover, Drummond et al. [24] demonstrated that ApoE\_/–/NOX2\_/– double KO mice were also protected from endothelial cell dysfunction and development of atherosclerotic lesions, suggesting that at least some of the protective actions of p47<sup>phox</sup> deletion were likely due to inhibition of NOX2 activity. A major mechanism by which NOX-derived ROS contribute to vascular disease is via superoxide (O2\*-)-mediated inactivation of NO, resulting in loss of its vasoprotective actions, and the subsequent generation of the highly reactive ROS, peroxynitrite (OONO-) (Figure 1). Peroxynitrite is a powerful oxidant that causes irreversible damage to macromolecules including proteins, lipids, and DNA, thereby disrupting crucial cell signaling pathways and promoting cell death.

Recent studies have indicated that NOX-derived O2<sup>•-</sup> production in the extracellular compartment may be markedly increased during vascular disease. Atherogenic stimuli such as tumor necrosis factor-α (TNF-α), endostatin, cholesterol, and homocysteine increase endothelial cell expression of NOX1 and NOX2, cause CRP generation, resulting in NOX2 activation in the plasma membrane (PM) [12,15,35,36]. The translocation of p47<sup>phox</sup> to the cytosolic face of these CRP-containing NOX2 aggregates is likely to result in 'hot spots' of activity in the endothelial cells PM. This, combined with the accumulation of macrophages (which normally express NOX2 oxidase in the PM) in the vessel wall, will result in markedly higher amounts of O2<sup>•</sup> being generated in the extracellular space, thereby increasing the likelihood of NO breakdown and production of OONO-. This may explain, at least in part, why the majority of oxidative damage detected in atherosclerotic lesions occurs in the extracellular matrix space [37]. All cardiovascular pathologies have in common excessive NOX-dependent ROS formation associated with up-regulation of the various NOX subtypes. In this study, Wortel et al., [8] were able to demonstrate for the first time that IR induces excessive NOX activation and ROS generation. This study and other studies published recently by their collaborators [38], demonstrate that ROS generation is an indispensable mediator of SDRT-induced ischemia/reperfusion pathobiology in tumors. They also demonstrated the transient and immediate O2<sup>•-</sup> generation and the subsequent accumulation of peroxynitrite (OONO<sup>-</sup>) after SDRT, which resulted in impaired endothelial function.

Recently, the first class of the dual NOX1 and NOX4 pharmacological inhibitors, GKT137831, received the approval for phase II clinical trial for the treatment of diabetic nephropathy. According to a recent press release of Genkyotex, the leading pharmaceutical company that develops NOX inhibitors, treatment of patients with diabetic nephropathy with GKT137831, significantly reduced liver enzymes and markers of inflammation. Moreover, the beneficial effects of GKT137831 were reported in several experimental models of disease, including atherosclerosis, hypertension, and diabetes [39–41], emphasizing the major role of the endothelial dysfunction in these pathological conditions and the involvement of NOX-mediated ROS.

Obtaining profound knowledge on the pathogenesis of IR-induced vascular injury could help stratify patients who might be at risk for cardiovascular events and potentially identify specific biomarkers that would warrant close surveillance and indicate specific diagnostic

interventions. We postulate that acute IR-induced microvascular dysfunction of normal tissue vasculature may contribute to and increase the risk for long-term cardiovascular morbidity and other vascular related diseases.

Wortel's [8] finding that sildenafil protected endothelial cells from RT-induced oxidative stress in the endothelial cells via reduction of NOX-mediated ROS formation and, thus inhibited the pro-apoptotic ASMase/ceramide pathway, is of paramount importance. The endothelium plays a role in the initiation of pulmonary oxidative injury induced by Ischemia/Reperfusion (I/R), and endothelial cells could be either a source or a target of oxidants. Oxidative injury to endothelium has been found to set the stage for secondary pulmonary injury by leukocytes [42,43].

While it is known that sildenafil affects NO levels in the blood vessels, Wortel et al., [8] elucidated the cellular mechanism by which sildenafil, attenuates endothelial dysfunction and protects against radiation-induced erectile dysfunction (ED). This study showed that sildenafil inhibited RT-induced NOX generation of ROS in endothelial cells and protected them from apoptotic death via the acid sphingomyelinase (ASMase)/ceramide pathway. Specifically, by inhibiting ASMase and ceramide generation, sildenafil significantly inhibited O2\*- generation, and subsequently NO was not used to generate ONOO- [44]. ONOO can disrupt crucial cell signaling pathways and initiate cell death by causing damage to macromolecules, including proteins, lipids, and more critically DNA [3,44]. In addition, to reducing the toxicity of ONOO-, sildenafil maintained the bioavailability of NO in the endothelial cells, and therefore preserved its vasoprotective properties. Reduced NO bioavailability is considered one of the main characteristics of endothelial dysfunction, which is also linked to erectile dysfunction [45]. In addition, since endothelial cells are a major target for RT-induced pneumonitis, GI syndrome, kidney damage, and xerostomia and because it is known that sildenafil is well tolerated, repurposing this drug should be considered for the treatment and prevention of RT-induced side effects, including cardiotoxicity for which it was initially successfully introduced.

Many cancer survivors find themselves struggling with health issues related to prior cancer treatment many years after they are declared cancer-free. These problems include chronic pain, neuropathy, infertility, recurrent infections, memory problems, sexual health issues, cognitive impairments and more, including increased risk of secondary malignancy. For many cancer survivors, these health issues last a lifetime, and some might even be life threatening. Mitigating treatment side effects by protecting the vasculature, in particular the generation of ROS, may result in a significant improvement of patient's quality of life by reducing morbidity and mortality.

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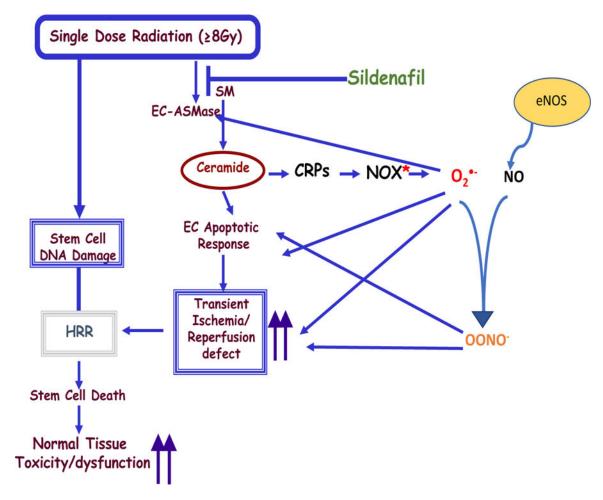


Figure 1: Sildenafil effect on Irradiated Stem/Progenitor cell-linked Endothelium.

Recent studies of single dose radiotherapy (SDRT) in mouse models provided a mechanism linking acid sphingomyelinase (ASMase)/ceramide-mediated microvascular injury with normal tissues stem cell demise or with the functional progenitors of the organ. SDRTinduced ceramide, having fusigenic properties, initiates the generation of CRPs. Subsequently, NOX subunits, such as gp91<sup>phox</sup> and p47<sup>phox</sup>, are aggregated, resulting in activated NOX via this process, and producing O2°-. O2°- may activate ASMase in a feedforward mechanism, enhancing CRPs clustering and forming positive amplifications of this process. O2<sup>•-</sup> coupling with NO generates another ROS, peroxynitrite (OONO<sup>-</sup>) but, most importantly depletes NO levels. Altogether, these processes constitute a redox signaling network or signalosome, resulting in endothelial dysfunction and impairment of endothelium-dependent vasodilation. Sildenafil significantly inhibited ROS production in general: the immediate O2\*- production and the subsequent OONO- generation, in endothelial cells, by stopping the feed-forward activation of ASMase, ceramide generation, and NOX activation and thus reduced endothelium injury and normal tissue toxicity. In the case of RT-induced gastrointestinal syndrome, the crypt stem cells use the homologous recombination repair (HRR) of DNA to recover. SDRT-induces transient Ischemia/ Reperfusion and thus inhibiting HRR, resulting in GI toxicity. Similarly, in other organs the damage to the vasculature results in normal tissue dysfunction.

ASMase: Acid Sphingomyelinase; BAECs: Bovine Aortic Endothelial Cells; CRPs: Ceramide-Rich Platforms; NOX: NADPH Oxidase; HRR: Homologous Recombination Repair