Significance of genetic variants in *DLC1* and their association with hepatocellular carcinoma

CHENG-RONG XIE^{*}, HONG-GUANG SUN^{*}, YU SUN, WEN-XIU ZHAO, SHENG ZHANG, XIAO-MIN WANG and ZHEN-YU YIN

Department of Hepatobiliary Surgery, Zhongshan Hospital, Xiamen University, Fujian Provincial Key Laboratory of Chronic Liver Disease and Hepatocellular Carcinoma, Xiamen, Fujian 361004, P.R. China

Received July 7, 2014; Accepted April 20, 2015

DOI: 10.3892/mmr.2015.3970

Abstract. DLC1 has been shown to be downregulated or absent in hepatocellular carcinoma (HCC) and is associated with tumorigenesis and development. However, only a small number of studies have focused on genetic variations of DLC1. The present study performed exon sequencing for the DLC1 gene in HCC tissue samples from 105 patients to identify functional genetic variation of DLC1 and its association with HCC susceptibility, clinicopathological features and prognosis. A novel missense mutation and four non-synonymous single nucleotide polymorphisms (SNPs; rs3816748, rs11203495, rs3816747 and rs532841) were identified. A significant correlation of rs3816747 polymorphisms with HCC susceptibility was identified. Compared to individuals with the GG genotype of rs3816747, those with the GA (odds ratio (OR)=0.486; P=0.037) or GA+AA genotype (OR=0.51; P=0.039) were associated with a significantly decreased HCC risk. Furthermore, patients with the GC+CC genotype of rs3816748, the TC+CC genotype of rs11203495 or the GA+AA genotype of rs3816747 had small-sized tumors compared with those carrying the wild-type genotype. No significant association of DLC1 SNPs with the patients' prognosis was found. These results indicated that genetic variations in the DLC1 gene may confer a risk for HCC.

Introduction

Hepatocellular carcinoma (HCC) is one of the most common cancers worldwide, with >600,000 mortalities per year, 55%

E-mail: yinzy@xmu.edu.cn

of which are in China (1). Risk factors of HCC include hepatitis B (HBV) or -C infection, high alcohol intake, smoking and nonalcoholic fatty liver disease (2). HCC is increasingly becoming a serious health problem in China. Recent studies have focused on finding somatic mutations in HCC by whole-exome sequencing. The results of these studies have shown that HCC is associated with somatic mutations in several genes, including *TP53*, *ARID1A*, *IRF2*, *MLL4* and *CTNNB1* (3-5). However, these studies did not fully elucidate the underlying mechanisms of the carcinogenesis and occurrence of HCC.

Deleted in liver cancer 1 (DLC1), which is located on chromosome 8p22, was first mapped in 1998 by Yuan et al (6). DLC1 consists of 18 exons. As a tumor suppressor gene, the protein encoded by DLC1 can regulate the structure of the actin cytoskeleton and inhibit cell proliferation, migration, invasion and angiogenesis (7-9). DLC1 has been shown to be downregulated in several cancer types, including HCC, breast cancer, nasopharyngeal cancer and cervical cancer (10). Promoter hypermethylation, low-level acetylated H3 and H4 and enriched H2K27me3 are responsible for silencing DLC1 expression (11-13). Besides the epigenetic silencing mechanisms of DLC1 in HCC, little is known about the association of DLC1 variants with HCC. Single nucleotide polymorphisms (SNPs) are the most frequent genetic variations in the human genome. Epidemiological studies have demonstrated that genetic variants are involved in various phases of carcinogenesis, which may determine susceptibility to the development of HCC (14,15). The synonymous SNP rs621554 of DLC1 was reported to be significantly associated with HBV-associated HCC (16). However, the non-synonymous SNPs of DLC1 that typically alter the gene product by changing the amino acid sequence of the protein have not yet been investigated in association with HCC. The present study aimed to determine the association of non-synonymous SNPs of DLC1 with HCC susceptibility, clinicopathological features and prognosis in a Chinese population.

Materials and methods

Ethics statement. The present study was approved by the ethics committee of Xiamen Zhongshan Hospital (Xiamen, China). All subjects provided written informed consent.

Correspondence to: Dr Zhen-Yu Yin, Department of Hepatobiliary Surgery, Zhongshan Hospital, Xiamen University, Fujian Provincial Key Laboratory of Chronic Liver Disease and Hepatocellular Carcinoma, 209 South Hubin Road, Xiamen, Fujian 361004, P.R. China

^{*}Contributed equally

Key words: deleted in liver cancer 1, single nucleotide polymorphism, hepatocellular carcinoma, mutation

Study subjects. The present study analyzed 105 patients with HCC at Xiamen Zhongshan Hospital (Xiamen, China). Diagnosis was confirmed based on histological examination of the specimens. The specimens were transferred to a -80°C freezer immediately after surgery for long-term storage until analysis. Patient pathology information was obtained from pathology reports.

Measurement of α -fetoprotein (AFP). Serum samples were obtained from the above-mentioned patients. Serum AFP was measured by microchip capillary electrophoresis and a liquid-phase binding assay on a μ TAS Wako i30 Auto Analyzer (Wako Pure Chemical Industries, Ltd., Osaka, Japan). All processes were performed automatically and followed the manufacturer's instructions.

Sequencing exons of DLC1. To identify functional genetic variation of DLC1, the whole coding region was sequenced in all of the 105 HCC patients' samples. DNA was extracted from frozen tissue specimens using the TIANamp Genomic DNA kit (Tiangen Biotech, Beijing, China). The DLC1 exome sequencing for HCC samples was performed by BGI (Guangzhou, China).

Statistical analyses. All statistical analyses were performed using SPSS version 19.0 for Windows (International Business Machines, Armonk, NY, USA). χ^2 tests or Fisher's exact tests were used to determine the associations among the *DLC1* genotypes, HCC risk and clinicopathological characteristics. The strength of association between polymorphisms and HCC risk was assessed by calculating odds ratios (ORs) with the corresponding 95% confidence intervals (CIs). The Kaplan-Meier method with a log-rank test was used to establish whether the *DLC1* SNPs influence the prognosis of HCC patients. All tests were two-sided, and P-values less than 0.05 were considered to indicate statistically significant differences.

Results

Exome sequencing of DLC1. The present study identified genetic variants within the coding region of *DLC1* via exome sequencing. Previously reported mutations in *DLC1* were not identified in the subjects of the present study (17). However, in one patient, a novel heterozygous missense mutation $(T\rightarrow A)$ in exon 14 at nucleotide position 3,743 was identified (Fig. 1A). This mutation resulted in a change in the amino acid from Val to Ile at codon 1,248, which is in the Rho GAP domain. The 1,248th amino acid of the DLC1 protein is highly conserved among a variety of species (Fig. 1B).

For analysis, non-synonymous SNPs with the ability to affect encoded protein function were selected. A total of four non-synonymous SNPs were identified. The features of the non-synonymous SNPs of *DLC1* in HCC patients are shown in Table I.

Association of DLC1 SNPs with HCC susceptibility. A total of 197 records of healthy Chinese Han individuals from the 1,000 Genomes database were used as a control group (http://www.ncbi.nlm.nih.gov/variation/tools/1000genomes/). The association between each of the DLC1 genotypes and the risk of HCC is shown in Table II. A significant correlation of rs3816747

Table I. Characteristics of nonsynonymous SNPs of DLC1.

SNP ID	Allele	Exon	AA	AA position
rs3816748	G/C	2	Leu/Val	81
rs11203495	T/G	2	Gln/His	254
rs3816747	G/A	2	Thr/Ile	260
rs532841	C/T	9	Val/Met	791

SNP, single nucleotide polymorphism; AA, amino acid.

polymorphisms with HCC susceptibility was identified. Compared to individuals with the GG genotype of rs3816747, those carrying the GA (OR=0.486; 95%CI=0.245-0.962; P=0.037) or GA+AA (OR=0.51; 95%CI=0.267-0.974; P=0.039) genotype were associated with a significantly decreased risk of HCC. However, no significant differences were observed in the frequency distribution of rs3816748, rs11203495 and rs532841 between the case and control groups.

Association of DLC1 SNPs with clinicopathological characteristics of HCC. To further determine the clinicopathological significance of DLC1 SNPs, an univariate analysis was performed by using χ^2 tests or Fisher's exact tests to correlate the genotypes of these four non-synonymous SNPs with clinicopathological features (Table III). Of note, a significant association of rs3816748, rs11203495, rs3816747 and tumor size was revealed. Patients with a GC+CC genotype of rs3816748, TC+CC genotype of rs11203495 or genotype GA+AA of rs3816747 had small-sized tumors compared with those of wild-type genotype patients. However, these DLC1 SNPs had no significant association with gender, portal vein tumor thrombus, AFP and tumor differentiation level. Patients with a GC+CC genotype of rs3816748, TC+CC genotype of rs11203495 or GA+AA genotype of rs3816747 had small-sized tumors compared with those of wild-type genotype patients.

Association of DLC1 SNPs with HCC prognosis. The prognostic impact of DLC1 SNPs was assessed by using Kaplan-Meier survival curves and the log-rank test (Fig. 2). The patients were followed every three months subsequent to surgery until the follow-up deadline, January 2013, was reached. A total of 42.1% of the patients had succumbed to HCC at the end of the follow-up period. However, none of the DLC1 SNPs was associated with the overall survival rate. In addition, the association of tumor-free survival rates with the DLC1 SNPs was assessed. Similarly, no significant differences in tumor-free survival between different genotypes of DLC1 were observed (Fig. 3).

Discussion

The present study identified a novel missense mutation and four non-synonymous SNPs in *DLC1* in HCC samples via exome sequencing. A study by Park *et al* (17) reported that in 17 primary HCC tumor samples and 18 HCC cell lines, only one missense mutation (Val991IIe) in *DLC1* was detected, namely in the START domain in the Hep40 cell line. However, this mutation was not detected in the subjects of the present

Genotype	HCC, n (%)	1,000 GENOMES, n (%)	OR	95% CI	P-value
rs3816748					
GG	68 (65.38)	111 (56.35)	1 (reference)	-	-
GC	30 (28.85)	77 (39.09)	0.636	0.379-1.068	0.086
CC	6 (5.77)	9 (4.57)	1.088	0.371-3.192	0.878
GC+CC	36 (34.62)	86 (43.65)	0.683	0.418-1.118	0.129
rs11203495					
TT	21 (20.59)	38 (19.29)	1 (reference)	-	-
TC	47 (46.08)	101 (51.27)	0.842	0.446-1.590	0.596
CC	34 (33.33)	58 (29.44)	1.061	0.537-2.095	0.865
TC+CC	81 (79.41)	159 (80.71)	0.922	0.508-1.674	0.789
rs3816747					
GG	21 (20.59)	23 (11.68)	1 (reference)	-	-
GA	47 (46.08)	106 (53.81)	0.486	0.245-0.962	0.037
AA	34 (33.33)	68 (34.52)	0.548	0.266-1.126	0.1
GA+AA	81 (79.41)	174 (88.32)	0.51	0.267-0.974	0.039
rs532841					
CC	39 (37.14)	75 (38.07)	1 (reference)	-	_
СТ	47 (44.76)	89 (45.18)	1.016	0.601-1.715	0.954
TT	19 (18.10)	33 (16.75)	1.107	0.559-2.195	0.861
TC+TT	66 (62.86)	122 (61.93)	1.04	0.638-1.697	0.874

Table II. Genotypic analyses of *DLC1* single nucleotide polymorphisms and their association with HCC susceptibility in a Chinese population.

Bold values are statistically significant (P<0.05). OR, odds ratio; 95% CI, 95% confidence interval; HCC, hepatocellular carcinoma.

B	Mutation	Ν	L	A	Е	С	L	А
	Homo sapiens	Ν	L	Α	v	С	L	А
	Mus musculus	Ν	L	Α	۷	с	L	А
	Bos taurus	Ν	L	Α	v	С	L	А
	Rattus norvegicus	Ν	L	Α	v	с	L	А
	Canis lupus familiaris	Ν	L	Α	۷	с	L	А

Figure 1. (A) Sequence analysis shows nucleotide sequence with a novel heterozygous missense $T \rightarrow A$ mutation at 3,743 in *DLC1*. (B) The 1,248th amino acid of the DLC1 protein in different species.

study. *DLC1* mutation was rare and was only identified in one patient in the study. The novel mutation which was identified in the present study was located in exon 14, which encodes the Rho GAP domain. According to the pathological report of this patient, the tumor showed poor differentiation. This patient also had a shorter survival time (2 months) than the median survival time (61 months). Rho guanine triphosphatase (GTPase) activity is frequently deregulated in human cancers, and is involved in actin cytoskeleton remodeling, migration, metastasis, cell proliferation, transcription and tumorigenesis (18,19). The multiple function of DLC1 mainly depends on the Rho GAP domain, which catalyzes the conversion of an active GTP-bound Rho to the inactive guanine diphosphate-bound form (20). Therefore, DLC1 is able to inhibit tumorigenic and metastatic processes. Further functional analysis demonstrated that this mutant lost its tumor suppressive ability (data not shown).

A comprehensive genotyping analysis of a Chinese population provided evidence that a *DLC1* SNP was associated with susceptibility to HBV-associated HCC (16). This SNP was located in intron 19 of *DLC1*, which may influence the transcription of *DLC1* by changing the binding sites for a number of transcription factors or the target site of microRNA (16). However, the present study focused on non-synonymous *DLC1* SNPs in HCC patients. A significant association between rs3816747 polymorphisms and susceptibility to HCC was observed. The variant genotype GA+AA of rs3816747 conferred a 0.51-fold decreased susceptibility to HCC. It was also found that the variant genotypes GC+CC of rs3816748, TC+CC of rs11203495 and GA+AA of rs3816747 were significantly

		Gender		J.T.	ımor size			PVTT			AFP			Differenti	ation	
Genotypes	Male n (%)	Female n (%)	Ь	<5 cm n (%)	≥5 cm n (%)	Ь	No n (%)	Yes n (%)	Ь	<400 n (%)	≥400 n (%)	Ь	Low n (%)	Medium n (%)	High n (%)	P P
rs381674																
GG	57 (67.06)	11 (57.89)		12 (48.00)	51 (63.75)		23 (65.71)	37 (64.91)		40 (71.43)	19 (55.88)		6 (46.15)	50 (68.49)	5 (55.56)	
GC	23 (27.06)	7 (36.84)	0.824	11 (44.00)	15 (18.75)	0.083	10 (28.57)	16 (28.07)	1	13 (23.21)	14 (41.18)	0.099	7 (53.85)	18 (24.66)	3 (33.33)	0.226^{a}
CC	5 (5.88)	1 (5.26)		2 (8.00)	14 (17.50)		2 (5.71)	4 (7.02)		3 (5.36)	0 (00.0)		0 (00.0)	5 (6.85)	1 (11.11)	
GC+CC	28 (32.94)	8 (42.11)	0.448	13 (52.00)	29 (36.25)	0.024	12 (34.29)	20 (35.09)	0.937	16 (28.58)	14 (41.18)	0.182	7 (53.85)	23 (31.51)	4 (44.44)	0.280
rs11203495																
TT	14 (16.87)	6 (33.33)		2 (8.00)	19 (27.54)		5 (14.29)	14 (24.56)		15 (26.79)	4 (12.50)		1 (7.692)	17 (23.94)	1 (11.11)	
TC	41 (49.40)	6 (33.33)	0.304^{a}	13 (52.00)	28 (40.58)	0.133	18 (51.43)	24 (42.11)	0.466	27 (48.21)	15 (46.88)	0.169	4 (30.77)	31 (43.66)	6 (66.67)	0.159
CC	28 (33.73)	6 (33.33)		10 (40.00)	22 (31.88)		12 (34.29)	19 (33.33)		14 (25.00)	13 (40.63)		8 (61.54)	23 (32.39)	2 (22.22)	
TC+CC	69 (83.13)	12 (66.67)	0.196	23 (92.00)	50 (72.46)	0.045	30 (85.71)	43 (75.44)	0.237	41 (73.21)	28 (87.50)	0.117	12 (92.31)	54 (76.06)	8 (88.89)	0.368
rs3816747																
GG	14 (16.87)	6 (33.33)		2 (8.00)	19 (27.54)		5 (14.29)	14 (24.56)		15 (26.79)	4 (12.50)		1 (7.692)	17 (23.94)	1 (11.11)	
GA	41 (49.40)	6 (33.33)	0.304^{a}	13 (52.00)	28 (40.58)	0.133	18 (51.43)	24 (42.11)	0.466	27 (48.21)	15 (46.88)	0.169	4 (30.77)	31 (43.66)	6 (66.67)	0.159
AA	28 (33.73)	6 (33.33)		10 (40.00)	22 (31.88)		12 (34.29)	19 (33.33)		14 (25.00)	13 (40.63)		8 (61.54)	23 (32.39)	2 (22.22)	
GA+AA	69 (83.13)	12 (66.67)	0.196	23 (92.00)	50 (72.46)	0.045	30 (85.71)	43 (75.44)	0.237	41 (73.21)	28 (87.50)	0.117	12 (92.31)	54 (76.06)	8 (88.89)	0.368
rs532841																
CC	35 (40.70)	4 (21.05)		11 (44.00)	22 (30.99)		16 (43.24)	20 (35.09)		26 (46.43)	11 (32.35)		5 (38.46)	27 (36.49)	3 (33.33)	
CT	39 (45.35)	8 (42.11)	0.056	9 (36.00)	35 (49.30)	0.448^{a}	17 (45.95)	22 (38.60)	0.187	20 (35.71)	16 (47.06)	0.410	4 (30.77)	32 (43.24)	6 (66.67)	0.401
TT	12 (13.95)	7 (36.84)		5 (20.00)	14 (19.72)		4 (10.81)	15 (26.32)		10 (17.86)	7 (20.59)		4 (30.77)	15 (20.27)	0 (00.0)	
CT+TT	51 (59.30)	15 (78.95)	0.109	14 (56.00)	49 (69.01)	0.239	21 (56.76)	37 (64.91)	0.427	30 (53.57)	23 (67.65)	0.188	8 (61.54)	47 (63.51)	6 (66.67)	1
^a Fisher's exac with CC). AF	ot test. P<0.05 P, alpha-fetol	5 are given in protein; PVT	T, portal	-values indicat	e the followin ombus; P, P-v	ig: rs381 alue.	674, compare	d with GG; rs	1120349)5, compared	with TT; rs38	16747, со	ompared with	GG and rs53	(2841, comp	ared

Table III. Correlation between DLC1 single nucleotide polymorphism genotypes and clinicopathological characteristics in study subjects.



Figure 2. Survival analysis of *DLC1* single nucleotide polymorphisms in hepatocellular carcinoma based on genotypes. (A) Kaplan-Meier curves of overall survival for rs3816748 (GG, 50 individuals; GC, 20; and CC, 4), (B) rs11203495 (TT, 16; TG, 36; and TT, 20), (C) rs3816747 (GG, 16; GA, 36; and AA, 20) and (D) rs532841 (CC, 24; CT, 38; and TT, 13) genotypes in *DLC1*.



Figure 3. Survival analysis of *DLC1* single nucleotide polymorphisms in hepatocellular carcinoma based on genotypes. (A) Kaplan-Meier curves of tumor-free survival for rs3816748 (GG, 67 individuals; GC, 30; and CC, 6), (B) rs11203495 (TT, 21; TG, 46; and TT, 34), (C) rs3816747 (GG, 21; GA, 46; and AA, 34) and (D) rs532841 (CC, 39; CT, 46; and TT, 19) genotypes in *DLC1*.

associated with decreased tumor size. It was therefore hypothesized that the non-synonymous SNPs that change the amino acid sequence of the protein may alter the function of the DLC1 protein. The DLC1 gene encodes four isoforms, which occur due to a heterogeneity in promoters and alternative splicing at the 3'-end (20). The rs3816748, rs11203495, and rs3816747 polymorphisms are all located in exon 2 of DLC1. Exon 2 encodes the N-terminal region before the SAM domain, which only exists in isoform 1 and isoform 3 (21). However, the function of this region has remained to be fully elucidated. It has been reported that DLC1 isoform 1 is less efficient in exerting its tumor-suppressive activity compared with DLC1 isoform 2, which lacks the N-terminal region (22). In the present study, it was therefore hypothesized that this region may alter the tumor growth suppressive effect in an auto-inhibitory manner through interaction with other proteins, or by changing its localization. These non-synonymous SNPs in exon 2 change the amino acid sequence of the N-terminal region, which may decrease the auto-inhibitory effect. Further functional study of the N-terminal region and genetic variants will be necessary to reveal the molecular mechanisms underlying the association between non-synonymous SNPs of DLC1 and HCC.

Previous studies have reported associations between SNPs in certain genes and HCC prognosis, including rs2640908 in PER, rs1741981 in HDAC1 and rs2547547 in HDAC3 (23,24). However, in the present study, no significant association of DLC1 SNPs with prognosis was found. Similarly, Ko et al (22) reported that DLC1 expression was not associated with prognosis. Although restoration of DLC1 resulted in the inhibition of colony formation, cell migration and invasion in vitro as well as reduction of the development and metastasis of tumors in vivo and in vitro (25), DLC1 did not influence the prognosis in HCC patients. Recently, Roessler et al (26) identified survival-associated driver genes by conducting an integrative genomic profiling analysis of 76 patients with HBV-associated HCC. Six tumor suppressor genes, including *DLC1* on chromosome 8p, were found to be deleted in patients with poor prognosis (26). These studies suggested that DLC1 SNPs on their own do not have any important role in HCC prognosis.

In conclusion, mutations of *DLC1* in the HCC patients examined in the present study were rare. The rs3816747 polymorphism in *DLC1* had a predictive value for HCC susceptibility in a Chinese population. Three SNPs (rs3816748, rs11203495 and rs3816747) in exon 2 were associated with tumor size. However, these SNPs were not associated with prognosis and are therefore not suitable as prognosis markers. To the best of our knowledge, the present study was the first to investigate the association of all non-synonymous SNPs of *DLC1* found in HCC patient samples with HCC susceptibility, clinicopathological characteristics and prognosis in Chinese subjects. However, a larger population and further exploration of the molecular mechanisms of *DLC1* SNPs may be required for further confirmation of this association.

Acknowledgements

This study was supported by grants from the National Natural Science Foundation of China (nos. 81172286 and 81372618),

the '973' Program (no. 2013CB910803) and the National Key Sci-Tech Special Project of China (no. 2012ZX10002-011-005). The support by Fujian Provincial Key Laboratory of Chronic Liver Disease and Hepatocellular Carcinoma administration at Zhongshan Hospital (Xiamen, China) is highly appreciated.

References

- 1. Jemal A, Bray F, Center MM, Ferlay J, Ward E and Forman D: Global cancer statistics. CA Cancer J Clin 61: 69-90, 2011.
- Llovet JM, Bustamante J, Castells A, *et al*: Natural history of untreated nonsurgical hepatocellular carcinoma: rationale for the design and evaluation of therapeutic trials. Hepatology 29: 62-67, 1999.
- 3. Cleary SP, Jeck WR, Zhao X, *et al*: Identification of driver genes in hepatocellular carcinoma by exome sequencing. Hepatology 58: 1693-1702, 2013.
- 4. Huang J, Deng Q, Wang Q, *et al*: Exome sequencing of hepatitis B virus-associated hepatocellular carcinoma. Nat Genet 44: 1117-1121, 2012.
- Guichard C, Amaddeo G, Imbeaud S, *et al*: Integrated analysis of somatic mutations and focal copy-number changes identifies key genes and pathways in hepatocellular carcinoma. Nat Genet 44: 694-698, 2012.
- Yuan BZ, Miller MJ, Keck CL, Zimonjic DB, Thorgeirsson SS and Popescu NC: Cloning, characterization and chromosomal localization of a gene frequently deleted in human liver cancer (DLC-1) homologous to rat RhoGAP. Cancer Res 58: 2196-2199, 1998.
- Yamaga M, Kawai K, Kiyota M, Homma Y and Yagisawa H: Recruitment and activation of phospholipase C (PLC)-delta1 in lipid rafts by muscarinic stimulation of PC12 cells: contribution of p122RhoGAP/DLC1, a tumor-suppressing PLCdelta1 binding protein. Adv Enzyme Regul 48: 41-54, 2008.
- Zhou X, Thorgeirsson SS and Popescu NC: Restoration of DLC-1 gene expression induces apoptosis and inhibits both cell growth and tumorigenicity in human hepatocellular carcinoma cells. Oncogene 23: 1308-1313, 2004.
- Shih YP, Liao YC, Lin Y and Lo SH: DLC1 negatively regulates angiogenesis in a paracrine fashion. Cancer Res 70: 8270-8275, 2010.
- Ullmannova V and Popescu NC: Expression profile of the tumor suppressor genes DLC-1 and DLC-2 in solid tumors. Int J Oncol 29: 1127-1132, 2006.
- Yuan BZ, Durkin ME and Popescu NC: Promoter hypermethylation of DLC-1, a candidate tumor suppressor gene, in several common human cancers. Cancer Genet Cytogenet 140: 113-117, 2003.
- Kim TY, Jong HS, Song SH, *et al*: Transcriptional silencing of the DLC-1 tumor suppressor gene by epigenetic mechanism in gastric cancer cells. Oncogene 22: 3943-3951, 2003.
- Au SL, Wong CC, Lee JM, Wong CM and Ng IO: EZH2-mediated H3K27me3 is involved in epigenetic repression of deleted in liver cancer 1 in human cancers. PLoS One 8: e68226, 2013.
- Jung SW, Park NH, Shin JW, et al: Prognostic impact of telomere maintenance gene polymorphisms in hepatocellular carcinoma patients with chronic hepatitis B. Hepatology 59: 1912-1920, 2014.
- Dong QZ, Zhang XF, Zhao Y, et al: Osteopontin promoter polymorphisms at locus-443 significantly affect the metastasis and prognosis of human hepatocellular carcinoma. Hepatology 57: 1024-1034, 2013.
- 16. Dong X, Zhou G, Zhai Y, *et al*: Association of DLC1 gene polymorphism with susceptibility to hepatocellular carcinoma in Chinese hepatitis B virus carriers. Cancer Epidemiol 33: 265-270, 2009.
- Park SW, Durkin ME, Thorgeirsson SS and Popescu NC: DNA variants of DLC-1, a candidate tumor suppressor gene in human hepatocellular carcinoma. Int J Oncol 23: 133-137, 2003.
- Buongiorno P and Bapat B: Rho GTPases and cancer. Prog Mol Subcell Biol 40: 29-53, 2005.
- Gómez del Pulgar T, Benitah SA, Valerón PF, Espina C and Lacal JC: Rho GTPase expression in tumourigenesis: evidence for a significant link. Bioessays 27: 602-613, 2005.
- 20. Zhou X, Zimonjic DB, Park SW, Yang XY, Durkin ME and Popescu NC: DLC1 suppresses distant dissemination of human hepatocellular carcinoma cells in nude mice through reduction of RhoA GTPase activity, actin cytoskeletal disruption and down-regulation of genes involved in metastasis. Int J Oncol 32: 1285-1291, 2008.

- 21. Low JS, Tao Q, Ng KM, *et al*: A novel isoform of the 8p22 tumor suppressor gene DLC1 suppresses tumor growth and is frequently silenced in multiple common tumors. Oncogene 30: 1923-1935, 2011.
- 22. Ko FC, Yeung YS, Wong CM, *et al*: Deleted in liver cancer 1 isoforms are distinctly expressed in human tissues, functionally different and under differential transcriptional regulation in hepatocellular carcinoma. Liver Int 30: 139-148, 2010.
- 23. Zhao B, Lu J, Yin J, *et al*: A functional polymorphism in PER3 gene is associated with prognosis in hepatocellular carcinoma. Liver Int 32: 1451-1459, 2012.
- 24. Yang Z, Zhou L, Wu LM, Xie HY, Zhang F and Zheng SS: Combination of polymorphisms within the HDAC1 and HDAC3 gene predict tumor recurrence in hepatocellular carcinoma patients that have undergone transplant therapy. Clin Chem Lab Med 48: 1785-1791, 2010.
- 25. Wong CM, Yam JW, Ching YP, *et al*: Rho GTPase-activating protein deleted in liver cancer suppresses cell proliferation and invasion in hepatocellular carcinoma. Cancer Res 65: 8861-8868, 2005.
- Roessler S, Long EL, Budhu A, *et al*: Integrative genomic identification of genes on 8p associated with hepatocellular carcinoma progression and patient survival. Gastroenterology 142: 957-966, e12, 2012.