



Review

The biological pathways of Alzheimer disease: a review

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Abstract: Alzheimer disease is a progressive neurodegenerative disorder, mainly affecting older people, which severely impairs patients' quality of life. In the recent years, the number of affected individuals has seen a rapid increase. It is estimated that up to 107 million subjects will be affected by 2050 worldwide. Research in this area has revealed a lot about the biological and environmental underpinnings of Alzheimer, especially its correlation with β -Amyloid and Tau related mechanics; however, the precise molecular events and biological pathways behind the disease are yet to be discovered. In this review, we focus our attention on the biological mechanics that may lie behind Alzheimer development. In particular, we briefly describe the genetic elements and discuss about specific biological processes potentially associated with the disease.

Keywords: Alzheimer disease; biological pathways; immune system; oxidative stress

1. Alzheimer's disease

Dementia is a broad category of neurodegenerative pathologies, whose main symptom is a decline in cognitive ability severe enough to interfere with activities of daily living. Among them, Alzheimer Disease (AD) is the most common type, accounting for 60% and up to 80% of the total Dementia cases.

According to the Global Burden of Disease Study, AD is one of the fastest rising diseases among the leading causes of death [1,2]. About 10% of people of age 65 and older suffer from AD [3,4]. Epidemiologic data in the last decade highlighted a sharp increase of AD incidence: According to the estimations, the number of AD subjects will increase from the 26.6 million worldwide in 2006 up to 107 million by 2050, with 16.5 in Europe [3,4]. To note, 68% of the increase would be localized in the low- and middle-income countries [5].

AD has a progressive nature, and eventually leads to a severe cognitive decline. It mainly affects older people (>65 years old) even though earlier onset forms are also described in literature (10% of AD cases) [6]. From a clinical point of view, AD is staged in four different phases: preclinical, mild, moderate and late-stage (see Box 1). This classification is mainly based on cognitive decline.

Box 1. AD Stages [6,7]

The clinical classification of AD is mainly based on the severity of cognitive decline and the histopathological alterations. Four stages are usually described:

Preclinical: This phase is often overlooked since no severe symptoms are present. It is usually classified as mild cognitive impairment. In this phase the earliest pathological changes begin, and hit entorhinal cortex (first) and hippocampus (later). From a symptomatologic point of view, subjects at this stage present mild memory loss with relative sparing of long-term memories. No significant impairment in their daily activities.

Mild Alzheimer Disease: In this phase cognitive symptoms start manifesting. The pathological alterations reach the cerebral cortex during this phase. From a symptomatologic point of view, along with memory loss, there is an inability to remember new information, forgetting things and appointments, followed by impairment in problem-solving, judgment and executive functioning. Also subjects manifest personality changes, mood swings and loss of spontaneity. Further, states of confusion and disorientation are commonly seen.

Moderate Alzheimer Disease: In this phase the symptoms severity increase further. The pathological damage further spreads to the areas responsible for language, reasoning and sensory processing (cerebral cortex). Other than an increased severity of symptoms from the previous phases, behavioral problems and social withdrawal tendencies begin to appear. This is followed by language disorder and impairment of visuo-spatial skills. Of note, subjects at this stage have trouble recognizing their own dears.

Severe Alzheimer Disease: In this phase, subjects completely lose their independency for daily activities. The pathological damage in this stage is believed to cover all of the cortex areas. From a symptomatologic point of view, the affected subjects cognitive abilities reach its lowest state, further systemic symptoms starts to appear, including difficulty performing learned motor tasks (dyspraxia), olfactory dysfunction, sleep disturbances, extrapyramidal motor signs like dystonia, akathisia, and parkinsonian symptoms.

To note, other staging systems were developed for AD, including the one introduced by Braak and Braak [7,8]. This model is based on topographical staging of neurofibrillary tangles and divides AD progression into 6 stages. This Braak staging is an integral part of the National Institute on Aging and Reagan Institute neuropathological criteria for the diagnosis of AD [7].

The severity and the prevalence of AD promoted a flourishing research in order to develop strategies to confront this pathology.

In the 2002–2012 decade, over 240 drugs for AD were tested in publicly and privately funded clinical trials registered on the National Institutes of Health registry (clinicaltrials.gov) [9]. However, none of the pharmacologic treatments developed so far is able to cure or halt AD progression [10–16]. The two categories of approved drugs, Cholinesterase Inhibitors (tacrine, donepezil, rivastigmine, and galantamine) and partial N-methyl D-aspartate (NMDA) antagonists (memantine), only provide a temporary amelioration of symptoms [17–20]. For a brief summary, please see supplementary Box 1. Furthermore, these drugs have a variable efficacy among the subjects [21].

The absence of a definitive treatment is related to the lack of knowledge about the precise molecular mechanics and the events behind the disease [5].

Although the grade and type of symptoms may vary greatly from person to person [22], post mortem observations on AD subjects' Central Nervous System (CNS) evidenced some common features: (1) synaptic loss, (2) accumulation of abnormal neuritic plaques and (3) presence of neurofibrillary tangles [23–25]. The first lesions are believed to start accumulating 10–15 years before the onset of cognitive symptoms [26].

2. Physiopathological manifestations

Neuritic plaques are spheroid-like microscopic lesions characterized by a core of extracellular amyloid β peptide ($A\beta$) and surrounded by abnormal axonal endings. $A\beta$ is derived from a large protein called amyloid precursor protein (APP). APP can be cleaved by the action of enzymes named α -, β -, and γ -secretase. In normal individuals, APP is firstly cleaved by α secretase and then γ secretase [27,28]. The action of α secretase eliminates the risk of formation of an $A\beta$ peptide.

In the neurons of the subjects suffering from AD, it is the β -secretase enzyme that acts to cleave the APP molecule instead of α secretase, and the resulting sAPP β is released from the cell [29]. The sequential cleavage by β and then γ -secretase results in $A\beta_{40}$, $A\beta_{42}$ (β -amyloid 40 and β -amyloid 42) and C99 amino acid peptides [30]. The increased concentration of $A\beta_{42}$ (from thereon $A\beta$) favors the formation of oligomers, which have neurotoxic properties [31]. These oligomers tend to cluster around meningeal and cerebral vessels and gray matter in AD. These deposits coalesce to form the miliary structures that are known as plaques [7]. Interestingly, recent data suggest the importance of cholesterol for the γ -secretase cleavage of amyloid precursor protein, the last step of $A\beta$ formation [32].

Regarding neurofibrillary tangles, they are fibrillary intracytoplasmatic structures in neurons formed by a protein called Tau. Physiologically, the primary function of Tau protein is to stabilize axonal microtubules. Usually, Tau is bound to microtubules and presents a certain number of phosphate molecules attached to it. The phosphorylation mechanics of Tau are altered in AD for reasons not entirely understood. This alteration leads to an abnormal increase of the phosphorylation, which in turn causes the detachment of Tau molecules from microtubules. Detached hyperphosphorylated Tau proteins tend to assemble to form filamentous structures known as paired helical filaments, which in turn aggregate in the insoluble neurofibrillary tangles [33,34]. Post mortem analyses on AD subjects demonstrated that few of the many different types of neurons in the brain develop abnormal Tau aggregates [35,36]. They are all projection neurons with high concentrations of neurofilaments whose axons are excessively long and thin compared to the size of their soma [36]. Analyses on AD subjects on various stages evidenced that neurofibrillary tangles start to form in the

transentorhinal cortex, spread to the hippocampus, and then progress to cover the cerebral cortex at later phases [37–39].

Another typical feature of AD is the granulo-vacuolar degeneration observed in the hippocampal pyramidal cells. This degeneration is likely associated to the cognitive decline: alterations of cognition processes are correlated to the decrease in density of presynaptic buttons from pyramidal neurons in laminae III and IV [7]. The decrease of synaptic buttons is likely associated to the vascular degeneration. Indeed, the risk of dementia is increased fourfold with subcortical infarcts; also the presence of a cerebrovascular disease exacerbates the degree of dementia and its rate of progression. However, the mechanisms behind are not fully determined [7].

3. Genetics

Despite a large part of its biological background is not characterized, AD has a strong genetic correlation with 3 genes: APP and the genes for the presenilin 1 (PSEN1) and presenilin 2 (PSEN2) proteins. Alterations within these genes are directly correlated with plaques formation. Literature data demonstrated that subjects inheriting mutations within APP or PSEN1 genes are guaranteed to develop AD, while those inheriting mutations within PSEN2 gene have a 95 percent chance of developing the disease [40]. The cases of AD caused by alterations within the three genes are labeled as autosomal dominant familial AD, because of its inheritance model. It generally develops within 60 years of age, sometimes as early as 30 years of age [5]. For this reason, it is also often indicated as early onset AD (EOAD). This form presents a clear molecular background and it's easily recognized since it runs in families. Up to 5% of all AD cases are of this type [41].

The largest part of AD cases (more than 95% AD) presents a sporadic manifestation and usually manifests between the 60 and 65 years of age (Late onset AD or LOAD). This form has a more cryptic genetic background [41]:

Both EOAD and LOAD may occur in people with a positive family history of AD. About 60% of EOAD subjects have relatives affected by AD [42]. Around 13% of these subjects present an autosomal dominant inheritance with at least 3 generations affected [42]. EOAD cases may also occur in LOAD families [43].

Regarding the risk factors, contrary to EOAD single-gene inheritance, virtually all LOAD cases likely involve multiple susceptibility genes and environmental factors [2,43,44]. Although none of them can be labeled as causative, they are associated with an increased risk of developing the disease. Excluding the known three causative genes of EAOD form, only one other gene evidenced a strong association with AD risk: the Apolipoprotein E gene (APOe).

APOe is the gene encoding for the Apolipoprotein E, whose function is to bind lipids and sterols and transport them through the lymphatic and circulatory systems. In particular, APOe is in charge of cholesterol transport in the brain [45,46]. Several studies associated the isoform e4 of this protein to an increased risk of developing AD [47–50]. At least one APOe4 allele is found in about 15–25% of the general population and can reach 50% in AD population [50,51]. The fine molecular mechanisms behind the risk increase operated by APOe4 are not completely characterized, but seem to be related to the formation of neurofibrillary tangles [52,53] and amyloid clearance processes [54,55]. Also, this isoform alters multiple processes [56], in particular lipid metabolism [57,58]. The e4 isoform is structurally unstable compared to other isoforms and thus it is not able to perform its functions normally. Further, the loss of cysteine residues decreases its antioxidant potential [59] and

may lead to an increased production of oxidant molecules like malondialdehyde and hydroxynonenal (products of lipid peroxidation), as observed in APOe4 allele carriers [60,61]. Data obtained from cell cultures evidenced how APOe4 promotes oxidative stress and the generation of neurotoxic fragments which impairs mitochondrial activity [62–64]. Furthermore, a reduction in cerebral glucose utilization was observed in brain regions typically affected by AD [65,66].

Following the discovery of APOe4 association with LOAD, a number of studies investigated other genes involved in the same molecular cascades of APOE. More than 500 candidates were proposed in the last two decades (see Table 1 and [67,68] and references therein). They showed correlations with Tau phosphorylation, vacuolar sorting proteins, metallo-proteins, glucose and insulin metabolism, nitrous oxide synthesis, oxidative stress, growth factors, neurodevelopment, pruning-, inflammation- and lipid-related pathways. The blooming of association studies on new genes led to the creation of a genetic database (www.alzgene.org) by Bertram and colleagues [69]. The site collects numerous publications regarding genetic data of AD and several genes appear to affect AD risk. Table 1 reports the top ranked genes involved in the risk of AD (from Alzgene.org). It has to be noted that these genes are believed to have a limited effect on the overall prevalence of AD because they are rare or only slightly affect (in total a ~20% increase or decrease) AD risk [69].

Table 1. Genes significantly associated with Alzheimer’s Disease (Retrieved from alzgene.org).

Gene Name	Extended Name	Brief summary (exert from genecards.org)	References
Genes associated with Early Onset Alzheimer’s Disease			
APP	Amyloid β Precursor Protein	This gene encodes for a cell surface receptor and trans membrane precursor protein that is usually cleaved by α , β and γ secretases to form a number of peptides. The peptides are overall involved in functions relevant to neurite growth, neuronal adhesion and axonogenesis. In particular, some of them promote transcriptional activation, while others seem to be the basis for amyloid plaques formation. In addition, two of the peptides are antimicrobial peptides, having been shown to have bactericidal and antifungal activities. Mutations in this gene have been implicated in autosomal dominant Alzheimer.	[27,70–73]
PSEN1	Presenilin 1	This gene encodes for the catalytic subunit of the γ -secretase complex, which is involved in the cleavage of integral membrane proteins, including APP. It plays a role in Notch and Wnt signaling cascades and regulation of downstream processes via its role in processing key regulatory proteins. It stimulates cell-cell adhesion and is required for normal embryonic brain and skeleton development. Alterations of this subunit were correlated with autosomal dominant Alzheimer.	[74]
PSEN2	Presenilin 2	This gene encodes for the catalytic subunit of the γ -secretase complex, which is involved in the cleavage of integral membrane proteins, including APP. Two alternatively spliced transcript variants encoding different isoforms of PSEN2 have been identified. It requires the other members of the γ -secretase complex to have a protease activity. It may play a role in intracellular signaling and gene expression or in linking chromatin to the nuclear membrane. It may function in the cytoplasmic partitioning of proteins.	[75]

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Gene Name	Extended Name	Brief summary (exert from genecards.org)	References
Genes related to Late Onset Alzheimer's Disease			
ABCA7	ATP Binding Cassette Subfamily A Member 7	This gene encodes for a member of the ABC1 subfamily. Members of the ABC1 subfamily comprise the only major ABC subfamily found exclusively in multicellular eukaryotes. This transporter is often found in myelo-lymphatic tissues. The function of this protein is not entirely understood, but it is believed that it may have a role in lipid homeostasis in cells of the immune system, and in phagocytosis mechanics.	[76,77]
ADAM10	ADAM Metallopeptidase Domain 10	This gene encodes an ADAM family member whose role is related to its cleaving properties. Interact with several proteins, including TNF- α and E-cadherin. It is also involved with constitutive and regulated α -secretase cleavage of APP. Finally, contributes to the normal cleavage of the cellular prion protein.	[78]
APOE	Apolipoprotein E	The protein encoded by this gene is a major apoprotein of the chylomicron and is essential for the normal catabolism of triglyceride-rich lipoprotein constituents. This protein Mediates the binding, internalization, and catabolism of lipoprotein particles and can serve as a ligand for the LDL (Apo B/E) receptor. Mutations in this gene result in an impaired clearance of chylomicron and VLDL remnants. Further, several evidences in literature associate specific APOE isoforms with an increased risk of Alzheimer disease.	[79,80]
BACE1	β -Secretase 1	This gene encodes a member of the peptidase A1 family of aspartic proteases. This trans membrane protease catalyzes acts on APP. In particular, it leads to the generation and extracellular release of β -cleaved soluble APP, and a corresponding cell-associated C-terminal fragment which is later released by γ -secretase.	[81,82]
BIN1	Bridging integrator 1	This gene encodes several isoforms of a nucleocytoplasmic adaptor protein which are expressed in several different tissues. Of note, isoforms expressed in the central nervous system may be involved in synaptic vesicle endocytosis. This gene has been identified as one of the most relevant risk locus for late onset Alzheimer's disease (LOAD).	[83–86]
C9ORF72	Chromosome 9 Open Reading Frame 72	The protein encoded by this gene plays an important role in the regulation of endosomes trafficking. It may regulate autophagy-related mechanics. Also, it regulates actin dynamics in motor neurons by inhibiting the GTP-binding activity of ARF6, leading to ARF6 inactivation. Positively regulates axon extension and axon growth cone size in spinal motor neurons. It plays a role within the hematopoietic system in restricting inflammation and the development of autoimmunity.	[87]

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Gene Name	Extended Name	Brief summary (exert from genecards.org)	References
Genes related to Late Onset Alzheimer's Disease			
CD33	CD33 Molecule (Sialic Acid-Binding Ig-Like Lectin 3)	This gene encodes for a protein associated with Hematopoietic Stem Cell Differentiation Pathways and Lineage-specific Markers and Innate Immune System. In the immune response, it may act as an inhibitory receptor upon ligand induced tyrosine phosphorylation by recruiting cytoplasmic phosphatase(s) via their SH2 domain(s) that block signal transduction through dephosphorylation of signaling molecules.	[88–90]
CLU	Clusterin	The protein encoded by this gene is a secreted chaperone seemingly involved in several basic biological events including cell death, tumor progression, and neurodegenerative disorders. Its action inhibits formation of amyloid fibrils by APP, APOC2, B2M, CALCA, CSN3, SNCA and aggregation-prone LYZ variants.	[48,91–93]
CR1	Complement Receptor 1	The gene encodes a monomeric single-pass type I membrane glycoprotein. The protein mediates cellular binding to particles and immune complexes that have activated complement.	[94,95]
FUS	FUS RNA Binding Protein	This gene encodes a multifunctional protein component of the heterogeneous nuclear ribonucleoprotein (hnRNP) complex which is involved in pre-mRNA splicing and the export of fully processed mRNA to the cytoplasm. Its action influence gene expression, maintenance of genomic integrity and mRNA/microRNA processing.	[96]
GRN	Progranulin	This gene encodes for the 88 kDa precursor protein, progranulin. Cleavage of the signal peptide produces mature granulin which can be further cleaved into a variety of active, 6 kDa peptides. Granulin family members are important in normal development, wound healing, and tumorigenesis. Granulins have possible cytokine-like activity. They may play a role in inflammation, wound repair, and tissue remodeling.	[97]
LRRK2	Leucine-rich repeat kinase 2	The protein is present largely in the cytoplasm but also associates with the mitochondrial outer membrane. It is mainly involved in autophagy related mechanics. Of note, this protein seems to regulate neuronal process morphology in the central nervous system. Also, it plays a role in synaptic vesicle trafficking.	[98,99]
PICALM	Phosphatidylinositol Binding Clathrin Assembly Protein α -synuclein	This gene encodes a clathrin assembly protein, which recruits clathrin and adaptor protein complex 2 (AP2) to cell membranes at sites of coated-pit formation and clathrin-vesicle assembly. Alterations within this gene are associated with Alzheimer.	[100–102]
SNCA	Synuclein A	This gene may serve to integrate presynaptic signaling and membrane trafficking, in particular in the regulation of dopamine release and transport. It reduces neuronal responsiveness to various apoptotic stimuli. It has been found that SNCA peptides are a major component of amyloid plaques in the brains of patients with Alzheimer's disease. Also, it seems involved in the fibrillization of microtubule-associated protein Tau.	[103,104]

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Gene Name	Extended Name	Brief summary (exert from genecards.org)	References
Genes related to Late Onset Alzheimer's Disease			
SORL1	Sortilin Related Receptor 1	This gene encodes protein that belongs to at least two families: the vacuolar protein sorting 10 (VPS10) domain-containing receptor family, and the low density lipoprotein receptor (LDLR) family. Its role is likely related to endocytosis mechanics. SORL1 seems to be also associated with Alzheimer endophenotypes such as abstract thought, verbal memory, total brain volume, and white matter hyperintensities among people AD free. Regarding its mechanics SORL1 is believed to act through the regulation of the APP-containing endocytic vesicles trafficking.	[86,105–108]
MAPT	Microtubule Associated Protein Tau	This gene encodes the microtubule-associated protein Tau (MAPT). MAPT transcripts are differentially expressed in the nervous system, depending on stage of neuronal maturation and neuron type. It promotes microtubule assembly and stability, and might be involved in the establishment and maintenance of neuronal polarity. MAPT gene mutations have been associated with several neurodegenerative disorders such as Alzheimer's disease.	[109–111]
TARDBP	TAR DNA Binding Protein	This protein is involved in the regulation of CFTR splicing. It seems to be related to microRNA biogenesis, apoptosis and cell division. Hyperphosphorylation of this protein has recently been described in association with cognitive impairment, especially in the context of Alzheimer's disease pathology.	[112,113]
TREM2	Triggering Receptor Expressed On Myeloid Cells 2	This gene encodes a membrane protein that forms a receptor signaling complex with the TYRO protein tyrosine kinase binding protein. It is involved in immune response and in chronic inflammation. Defects in this gene are a cause of polycystic lipomembranous osteodysplasia with sclerosing leukoencephalopathy (PLOS). Inactivating mutations of TREM2 have been associated with an autosomal recessive form of early-onset dementia in literature.	[114,115]

Several other candidates and loci have been identified so far but replication analyses encountered some difficulties. Also, their contribute towards AD risk is deemed inferior to the genes of Table 1. Of them we want to report HLA-DRB5/DRB1, PTK2B, SLC24A4, RIN3, INPP5D, MEF2C, NME8, ZCWPW1, CELF1, FERMT2, CASS4, EPHA1, CD2AP, MS4A, CUGBP2, ATP5PD, MTHFD1L, GAB2, MEOX2 and PCDH11X [68,77,83,89,116–124].

Finally, Environmental factors like female gender [125], older age [126], low education level [127], smoking habit [128], obesity [129] and diabetes mellitus [129] increase the risk of AD. It is noteworthy to cite metals' levels as risk factors of AD. It was pointed out how metal homeostasis may be one of the processes which lead to the conformational changes of amyloidogenic proteins in AD [130]. Furthermore, while the variation of metals' concentrations is mainly environmental, alterations of protein transporters in the Blood Brain Barrier such as DMT1 or Transferrin may also alter metal homeostasis in the brain [131].

4. Molecular pathway contribution to AD risk

The number of genetic factors described are important contributors to AD, but even those cannot fully explain the totality of AD cases. Rather than single genes, a better approach would be

investigating AD as an event related to alterations affecting entire biological pathways. A plethora of mechanisms, including neuroinflammation [132], oxidative stress [133,134], defects in mitochondrial dynamics and function [135], cholesterol and fatty acid metabolism as well as glucose energetic pathways impairments in the brain [136,137], autophagy failure [138] and other less studied mechanisms have been proposed to contribute to AD. In this section we will report a brief panoramic discussion on these mechanisms. It has to be noted that while the various processes are discussed separately, they are strictly linked with each other and often work in a synergic way in the CNS.

5. Chronic inflammation and immune system

Among the processes thought to be involved with AD, inflammation is one of the main biological mechanisms considered. Inflammation in the brain, is a well-established (acute) process which defend the body against infection, toxins and injury. When the delicate equilibrium between pro- and anti-inflammatory signaling is disrupted, it results in chronic (neuro)inflammation [139–141].

Multiple literature data pointed out that neuro-inflammation (and pruning, as discussed below) has a role in the neurodegeneration processes commonly observed in AD [142–146].

Although, neuroinflammation is not typically associated to AD onset on its own, it plays a key role in increasing the severity of the disease by exacerbating A β and Tau pathologies [147].

A prolonged neuroinflammation state increases the concentration levels of proinflammatory cytokines in the microenvironment. The increase of cytokines triggers several potentially harmful effects: it induces mitochondrial stress in neurons, either directly or indirectly, including via A β signaling. It also increases Oxidative stress [148–150] and Blood-Brain Barrier (BBB) permeability which likely influence AD progression [151,152]. Furthermore, increased cytokines levels can influence other processes potentially related to AD. For instance, IL-18 increases the levels of Cdk5 and GSK-3 β , which are involved in Tau hyperphosphorylation [153]. Also, the activation of Cdk5 pathway causes Golgi fragmentation, neuronal and mitochondrial fragmentation. Interestingly, the inhibition of Cdk5 was proven to improve AD subjects' conditions [154–156].

Several processes and natural compounds can trigger immune responses and deviate this process from its physiological behavior. In particular, there has been a lot of focus recently on the complement system.

The complement system facilitates the immune system' response. It's excessive activation exacerbates AD symptoms [157], since it influence A β , Tau, and APOE4 interaction in AD [158,159] (for more details see [160]). Cholesterol also contributes to AD pathogenesis by inducing interleukin 1 β production through cytoplasmic sensor NLRP3 [161]. Strictly related to the immune function, viral infections, in particular Herpes Viruses (HSV1, HSV2), are also associated with AD. In particular, this type of infection is believed to potentially be an important risk factor for AD onset. Indeed, HSV1 (oral herpes) and HSV2 (genital herpes) can trigger A β aggregation, and their DNA is common in A β plaques. HSV1 reactivation is associated with Tau hyperphosphorylation and possibly Tau propagation. Recurrent reactivation may produce neuronal damage and AD pathology. This data open for a potential viral hypothesis of AD, where an increased risk may follow neurotropic infection [162,163]. The main players behind neuroinflammation and immune processes, are the non-neuronal cells that populate the brain (Neuroglia) [144,164]. These once "supporting" cells were indeed demonstrated to have an important role in the regulation of Brain processes,

including protection, neurotransmission and microenvironment homeostasis [165,166]. Physiologically, these cells have the ability to phagocytize A β proteins, thus acting against AD-related A β accumulation [167]. However, this protective function can be inhibited by increased levels of inflammatory cytokines such as IL-1 β , TNF- α and IFN- γ [140,168]. Neuroglia is composed by different cell types. Box 2 briefly reports their potential role in association with AD.

Box 2. Non-Neuronal Cells

The continuous research on AD and brain in general, demonstrated how the physiological functions of SNC are not linked to neurons functions only. Rather, it was demonstrated that the once “supporting” cells types present in SNC have an important role in the regulation of Brain processes, including neurotransmission and microenvironment homeostasis [165,166]. Alterations of their function may profoundly impact the physiological functions of the brain, and a large amount of literature data linked these alteration to several pathological states, including AD.

Microglia: Microglia are the resident phagocytes and innate immune cells of the brain, protecting from SNC from infections and also regulate the homeostasis. Over the past few decades, there has been an increased interest in microglia since the discovery of its involvement in synaptic plasticity dysfunctions. In particular, an alteration of the physiological functions of the cells has potentially harmful consequences for neuronal function, eventually leading to cognitive loss and behavioral deficits [169]. Also, synaptic plasticity has been demonstrated to be a key mechanics which is intimately linked with A β formation and with the long-term potentiation (LTP) pathways in AD [170,171]. Recent data support a possible role of microglia in AD risk, and it is now widely accepted that clustered populations of reactive microglia are hallmarks of AD. Microglia cells in the brain are usually in an inactivated state. In this state they are still able to perform some control functions [172–174], but do not seem to influence AD-related processes. Their activation, though, can trigger several molecular processes that correlate with an increased risk of AD. In particular, when activated microglia may trigger pro-inflammatory (M1 state) or anti-inflammatory (M2 state) processes [175]. M2 state seems to play a role in neuroprotective-related processes. The action of microglial cells in M1 state instead, are linked to AD development since it promotes neuroinflammation through the secretion of pro-inflammatory cytokines. As described before, neuroinflammation is detrimental for neurons and may lead to neurodegenerative processes. Also, cells in M1 state may release ROS, thus increasing the OS in the micro-environment [176]. As such, M1 microglia cells may also act as a bridge of different processes both related with AD. Several studies have also focused on the existent relationship of microglia with A β plaques formation and Tau hyper-phosphorylation, evidencing an interesting correlation with the main events of AD development [169]. However, little is known about microglial activities in early stage AD and in particular, on its role in synaptic dysfunction [169].

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Box 2. Non-Neuronal Cells

Astrocytes: like microglia cells, Astrocytes are one of the subtypes composing neuroglia. Once believed to carry out only a support function to neurons, it is now known that Astrocytes can influence neuronal and vascular function. Accordingly, the functional consequences of astrocyte dysfunction in AD can be severe.

It was evidenced in literature that astrocyte action is important in the onset and development of neurodegenerative events, including the ones related to AD pathology [177]. It has to be noted, though, that their role is likely secondary to other alterations, since it seems that astrocytes have a neuroprotective function in the brain and are activated (reactive astrogliosis) in critical cases in order to limit neural damage in the brain [178]. However, chronic activation ultimately leads to imbalances “exacerbating cellular pathology and behavioral outcomes” [177]. In particular, literature data demonstrated that several events related to AD, like A β accumulation, synaptic dysfunction, neuronal energy metabolism and neuroinflammation are strictly linked to astrocyte action [179]. From this point of view, astrocyte may be considered as another bridge that links together the several processes related to AD development. Furthermore, it was evidenced that the role of astrocytes in controlling neurovascular interactions may be correlated with brain hypoperfusion and/or impaired functional hyperemia, and both events are commonly observed in AD subjects [177]. This suggests an association between AD alterations and neurovascular functions. Consistently, it is interesting to note that A β accumulation seems to be mainly localized in brain areas with reduced cerebral blood flow [180–182]. Still, it has to be noted that the reduced vascularization may be a consequence of A β accumulation rather than a cause.

Neutrophils: Recently, some interesting literature data demonstrated neuroglial cells are not the only cell types that can be correlated with AD risk. Recent studies observed that the blood circulating immune cells, the neutrophils, seem to promote AD pathogenesis by triggering neuroinflammation-related AD events [183]. These cells are key regulators of the immune system because they communicate and interact with adaptive immune system cells during infections, chronic inflammation and autoimmune diseases [184,185]. The mechanics behind the neutrophil-dependent damage in AD are not entirely understood, especially because they should not be able to pass BBB. According to this, their effect should be indirect, through the increase of pro-inflammatory cytokines, or subordinated to BBB damage. The recent discovery of Neutrophil extracellular traps (neutrophil mechanics of defense against infection) [183] in AD models, added a new potential way for the cells to influence AD-related mechanics (we recommend to consult [183] for further details).

6. Oxidative stress

Oxidative stress (OS) has been widely recognized as a prodromal factor associated to AD [186]. Cell control on OS is particularly important to maintain the balanced microenvironment needed for multiple biological processes, from bioenergetics to other essential functions such as vesicle transport or post-transcriptional modulations [187]. Neurons in particular are very sensible to OS as their normal antioxidant content is low and their membranes are rich of polyunsaturated fatty acids [188].

Multiple causes, mainly related to mitochondrial dysfunctions and energy metabolism impairments (as it will be described later in the text) can increase cellular OS. Even the natural process of aging is believed to physiologically increase OS with time [189]. At neuronal level, oxidative damage related to age [190,191] strongly impairs the synaptic components involved in neuronal plasticity [189,192], cytoskeletal dynamics [193] and cellular communication [194], all processes known to be impaired in AD.

According to the current knowledge regarding OS it is not clear if OS increase could trigger AD onset. However, increased OS is often observed in the brain of early-stage AD subjects [195]. Further, it seems to have a key role in AD severity and propagation. Indeed, it has been demonstrated that A β enhances OS, and represents a source of radical oxygen and nitrogen species (ROS, RNS) [186]. This process is mainly linked with the function of important proteic mediators of OS, including NOX, TGF- β , NF- κ B and Nrf2 [196]. The increased OS state in turn triggers several molecular events that are strictly linked with AD symptomatology development [197].

First of all, elevated ROS and RNS concentrations promote Tau phosphorylation, which in turn promotes the destabilization of microtubules. In neurons, microtubules are of primary importance since they maintain these cells polarization and intracellular trafficking [189]. Their destabilization often translates in a decreased function of synapses [198].

ROS can propagate toward the membrane, and then oxidize proteins and nucleic acids, as confirmed by post-mortem AD patients' investigations. In particular, nucleic acid oxidization can cause lethal damage to cell (please refer to Mitochondria section) but can also promote protein aggregation. Indeed, recent observations indicated that Pre-translational mRNA oxidation synthesize peptides that are prone to aggregate [199]. mRNA oxidation seems to be a cytoanatomically selective event that hit mRNAs translated by ribosomes that reside tethered to the mitochondrial outer membrane [32]. The aggregates heavily impair the physiological cell functions. In neurons this could have severe consequences for the normal brain function.

Further, OS exert its effect on the choline recycling from the synapse processes, leading to ACh deficiency [200]. In late stage AD, levels of presynaptic high-affinity choline transporter 1 (CHT1) were observed to be decreased in synaptosomes in the hippocampus and neocortex of humans [201]. Further, high OS inactivates nAChRs, thus inducing long-term depression of cholinergic transmission [202].

An increased OS state leads to an imbalance between pro- and anti- apoptotic processes [153,203], leading to apoptosis and then neurodegeneration [204]. Interestingly, particularly low ROS concentrations may trigger proliferative signals in neurons, which highly reduce these cells functionality [204].

ROS can also regulate BBB function through enhancing the expression of several metallo-proteinases, and in particular the 9 isoform (MMP9). MMP9 expression is important in brain micro-

vascular environment since its alterations are associated with an increased BBB permeability that promotes AD progression through the extravasation of inflammatory factors and ROS in the brain [205,206].

Another effect of ROS is to influence the energy metabolism of the brain: it indirectly regulates neuronal cells permeability to glucose, decreasing GLUT-3 expression in AD neurons [207,208] and GLUT-1 in BBB [207]. These molecular events lead to a glucose hypometabolism state in the brain, which has relevant consequences in cells like neurons, whose physiological roles require high levels of energy [186].

Finally, excessive ROS inevitably lead to lipid peroxidation [209]. Polyunsaturated fatty acids are rich in the brain [210]. They could be degraded into malondialdehyde, which causes DNA damage and toxic stress in cells [211].

7. Energy metabolism

Numerous evidences implicate Energy metabolism with AD development [212]. In particular, glucose hypometabolism has been detected in the frontal, parietal, temporal, and posterior cingulate cortices of AD patients, and interests especially the basal forebrain cholinergic neurons degeneration [213]. AD intracellular lesions usually develop in neurons presenting long and thin axons compared to the soma [214] and present a reduced or absent myelin sheath. It is interesting to note that these type of neurons are the ones with highest energy requirements [215]. From a molecular point of view, glucose hypometabolism is likely linked with OS. For instance, it was observed that the expression and function of key enzymes in the glycolytic cascade is reduced with high ROS levels [32,216,217]. Energy alterations were also linked to a decreased functionality of enzymes related to TCA cycle, which lead neurons to a hypometabolic state [200,218–222]. High Cerebrospinal fluid pyruvate and lactate has been widely reported in AD patients compared with healthy elderly controls [223].

Furthermore, chronic treatment with pyruvate could alleviate short and long-term memory deficits via other pathogenic pathways without reducing amyloid- and Tau-dependent pathology in preclinical AD models [224]. Insulin signaling has been the focus of multiple studies for its association with AD [225–228]; A β oligomers can bind insulin receptors causing their internalization. The sheer increase of these oligomers in AD subjects triggers an inhibition of the insulin signaling pathways [229]. In sum, the alteration of insulin signaling (or an increased resistance to insulin) ultimately triggers neuroinflammation and neurodegenerative processes through: (1) an increase of A β concentrations; and (2) a GSK3 β -dependent (Glycogen Synthase Kinase 3 β) hyperphosphorylation of Tau protein [229–231]. The potential role of insulin signaling in AD is supported by the observation that insulin-related pathologies, like Diabetes Mellitus, increase AD risk [232].

8. Vascular related mechanics

Cerebrovascular abnormalities are common comorbidity in patients with Alzheimer's disease (AD) [233]. They may concur to the onset of cognitive impairment and dementia.

Vascular dysfunctions cause altered brain blood flow and pressure at the level of the brain [234]. These events are detrimental for the normal cerebral function that would result in disturbed homeostasis, but also in blood-brain barrier (BBB) damage and micro-fractures in cerebral vases [235],

with increased risk of neuroinflammation [236]. Ultimately, they lead to an increased neuronal death rate and eventually cause the onset and the progression of cognitive impairments [137].

Vascular dysfunctions also set in motion a chain of metabolic events strictly related to AD [237–242], including OS. Studies on animal models linked ROS production to the increased concentration levels of the Advanced Glycation Endproducts (AGE) proteins and their receptors (RAGE) in the vascular system. These events are linked to the formation of A β plaques [243–246]. Interestingly, RAGE is also correlated with inflammatory function and innate immunity, since it has the ability to recognize specific structural motifs (RAGE is also classified as a pattern recognition receptor). A further link with AD events is that brain levels of AGE and RAGE expression have increased in AD subjects [244,247]. Their expression also tends to increase naturally with age, offering a possible explanation for the increased risk to AD with Age [243,244,247].

A chronic hypoperfusion state also promotes the formation of A β , through the activation of the adaptive response to hypoxia and reduced clearance via perivascular draining [248,249]. Consistently, it is interesting to note that A β accumulation seems to be mainly localized in brain areas with reduced cerebral blood flow [180–182]. Also, the use of angiotensin receptor blockers or angiotensin converting enzyme inhibitors and diuretics has a protective effect against AD [250–252]. Nevertheless, the use of these compounds in AD treatment is not indicated, since reduced blood pressure may also trigger detrimental effects including memory impairments and an overall increased risk of AD [253–255]. According to our knowledge, in AD the cerebral vascularization undergoes cellular, morphological and structural alterations, which influence the blood flow [235]. However, accumulating evidences suggest that dysfunction of the cerebral vasculature may precede AD neuropathology, implicating a causal role in the etiology of AD, at least in some cases. Indeed, for long time it was believed that some cases of AD may have a vascular origin with neurodegenerative consequences [248]. Whether vascular dysfunctions are causative of AD have yet to be determined. However it is know that the cardiovascular system strictly interacts with AD neuropathology in multiple ways and that the interaction is bidirectional. Making vascular dysfunction an integral part of AD pathophysiology.

9. Neurodevelopment and neurotransmission associated processes

Neurodevelopmental and Neurotransmission related pathways are likely associated with AD development and in particular with its cognitive symptoms [256]. Physiologically, these processes consists in the proliferation, differentiation and maturation of neural stem cells (NSC) and the modulation of their interactions through synapse- and neurotransmission- related processes.

Several reports have indicated a significant reduction of 5-HT [257], DA [258] and NE [259] levels as well as their receptors in AD brain. In AD, loss of 5-HT results in depression, anxiety and agitation [260], dysregulation of DA release leads to reward-mediated memory formation deficits [261], and low level of NE impairs spatial memory function [262]. Additionally, the cholinergic system, which regulates memory function and behavior via the release of the neurotransmitter acetylcholine (ACh) [263], was found to be altered in AD. Accumulation of intraneuronal A β in AD degenerates basal forebrain cholinergic neurons and reduces ACh levels [264], which in turn leads to memory deficits [265]. Further, mitochondrial dysfunctions are associated with monoaminergic inactivity through various mechanisms. Higher MAO expression, in particular in association with the APOE4, also contributes to a reduced production of monoamines [200].

Further, literature evidences pointed out how pruning pathway becomes aberrantly up-regulated in early stages of AD, mediating synaptic loss [266]. Synaptic pruning is regulated by several signals, including cytokine TGF β [267], an inflammation-related factor whose levels have been found to be increased in AD [268–270].

These processes can be influenced by multiple factors, including OS and inflammation, which were already discussed, and novel processes like epigenetic control [271].

Recently it was observed that some new molecular cascades were triggered by A β . A β may act through PANX1 expression increase. PANX1 is a protein involved in the modulation of neurotransmission, neurogenesis and synaptic plasticity [189,272]. An increase of this protein under inflammatory conditions contributes to neuronal death [273]. A β can also bind to several postsynaptic partners, including NMDAR and type 1 metabotropic glutamate receptor 5 (mGluR5) [189]. The binding of such receptors cause NMDAR-dependent synaptic depression and spine elimination [274]. Recently, the link between A β , AD and these processes has been also associated to ryanodine receptors (RyR) function [189].

RyR is Ca²⁺ channel present in three different isoforms (with different responses to Ca²⁺) in the brain. It acts as redox sensors modulating different processes such as neuronal development, apoptosis, gene transcription, synaptic transmission and neuronal plasticity [275]. In particular, Hippocampal RyR has a critical role in many forms of synaptic plasticity [276]. Anomalous RyR channel function occurs in AD pathology [277,278]. In particular, it was observed that both A β and OS would promote aberrant activation of RyR, generating continuous cytoplasmic Ca²⁺ signals [189]. These anomalous Ca²⁺ signals lead to mitochondrial and NOX2-mediated ROS generation [279] and glial activation [280].

Glial activation is an important event in AD. Indeed, recent data demonstrated how the physiological functions of SNC are not linked to neurons functions only. Rather, it was demonstrated that the once “supporting” cells types present in SNC have an important role in the regulation of Brain processes, including neurotransmission and microenvironment homeostasis [165,166]. As briefly described in Box 2, Microglia modulate synaptogenesis, synapse tagging and elimination or synaptic pruning, help repair damage from injury and modulate synaptic transmission through cytokines [281] and in particular through complement proteins release [271,282].

These elements act as signals activating circulating macrophages that express complement receptors (C1qR and CR3) and microglial cells in the brain [267,283,284]. Microglia activation triggers synapses removal through phagocytosis or by releasing soluble synaptotoxic factors [285–287]. Several complement components are normally down-regulated in CNS [266,283]; their overexpression mediates synapse elimination [266,267,283,288]. Interestingly, studies in human and mouse brains observed an age-related up-regulation of C1q that deposits in synapses, particularly in the hippocampus, one of the most vulnerable regions to synapse loss in AD [289]. C3 has also been shown to contribute to synapse loss and dysfunction in the mouse hippocampus during normal aging [290]. Also, injection of A β oligomers in wild-type mice leads to up-regulation of C3 levels, which promote microglial removal of synaptic connections [256]. Further, down-regulating C3 in animal models of AD showed a decreased frequency of neurodegenerative phenomena [291,292].

Mechanistically, A β oligomers increase the expression of C3 in microglia and astrocytes. Elevated C3 levels promote microglia recruitment and mark synapses for elimination. It has to be noted that synaptic loss may be triggered by different mechanisms; some of them might not necessarily follow the trajectory of amyloid buildup [291,293].

Microglia function could be altered by several processes, including ROS generation and inflammation [279,280]. In particular, it was observed that glial cells actively release glutamate and ATP in presence of A β . This process trigger pro-apoptotic cascades in neurons [294].

10. Autophagy impairments

Autophagy is emerging as an important process in the regulation of neuronal and glial cells health in AD. Physiologically, Autophagy is an adaptive process induced under different forms of stress, including nutrient deprivation, hypoxia, and infection [295].

It is a complex process that consists of several sequential steps, sequestration, degradation, and amino acid/peptide generation, mediated by a unique organelle called the autophagosome, a vesicle that contains cellular material targeted to be degraded (macromolecules and organelles) by an intracellular degradation system [296].

Although all cell types have the ability to turn on autophagy, a growing body of studies suggest the importance of this process, specifically, in neurons [297].

Inhibition of autophagy events is causally linked to neurodegeneration, indicating the relevance of autophagy in the neuronal homeostasis regulation [298]. Of note, there is substantial evidence that deregulation of autophagy occurs in AD patients and AD animal models [186]. In this regard, both genetic and environmental factors connect autophagy with AD. For example, Apolipoprotein E4 (apoE4) overexpression potentially causes an A β accumulation in lysosomes, leading to neuronal death in the hippocampus [299,300]. Further, emerging evidence indicates that high ROS levels inhibit the fusion between autophagosomes and lysosomes [186].

Studies on animal models of AD reported that restoring the physiological autophagosomes clearance prevents the manifestation of cognitive symptoms [301].

In more detail, autophagy could be further classified Macroautophagy and Mitophagy: both are somewhat implicated in AD.

Macroautophagy, is tightly regulated degradative pathway that targets cellular macromolecules and eliminates them through proteolytic enzymes in autophagolysosomes [302–304]. Some data evidenced how this process is impaired in AD brains [303], where a pathological accumulation of autophagosomes can be observed [305]. In details, AD pathogenesis has been linked to through an impaired merging processes between the autophagosomal-lysosomal system and the consequent reduction/block of protein aggregates turnover [306,307]. Autophagic vacuoles, isolated from a variety of tissues, were shown to be enriched in APP, gamma-secretase components, PSEN1 and nicastrin, which are required to generate A β [308,309]. Interestingly, studies in animal models evidenced how abnormalities in this process occur early in AD pathogenesis, way before the appearance of neurofibrillary tangles or senile plaques. This indicates that induction of macroautophagy is not a consequence of amyloid deposition [306]. According to the autophagic hypothesis, the block of this process and the consequent accumulation of autophagosomes trigger neuronal degeneration [310] and leads to the release of these vesicles in the extracellular space where they form the characteristic AD plaques [306,307].

Mitophagy, is the selective autophagic removal of mitochondria. This process is particularly important in neurons, where these organelles play a crucial role in cell survival [311]. In physiological conditions, dysfunctional mitochondria are removed from cytoplasm through mitophagy, an event that in AD disease is suppressed by excessive levels of ROS and A β [186].

Generally, initiation of the mitophagy pathway occurs via the relocation of cardiolipin, a diphosphatidylglycerol lipid, from the inner mitochondrial membrane to outer mitochondrial membrane [312]. Evidence has indicated that two proteins PTEN-induced putative kinase1 (PINK1) and parkin can also initiate the mitophagy pathway leading to autophagosome-mediated mitochondrial degradation [313]. In AD, high levels of A β and Tau inhibit the expression of PINK1 and parkin, thereby reducing the number of autophagosomes leading to increased dysfunctional lysosomes and the severe disease pathology [312,314].

11. Protein misfolding and misfolded protein clearance

AD research is ever increasing, and everyday new suggestive hypotheses are postulated to tentatively explain that “over 90% of cases” (LOAD) with a cryptic cause. Other than the above cited processes, it has been suggested that AD pathological alterations may be attributed to an incorrect folding of proteins [315]. It has been suggested that pathologically misfolded proteins can influence properly folded proteins to alter their conformation, thus resulting in the propagation of disease [316]. Further, it has been observed that misfolded proteins could also trigger chronic inflammation through different activation pathways [317].

The possibility of propagation of the misfolded proteins between cells is not impossible: there are several processes developed for cell-cell communication, including synaptic transmission, direct communication through gap junction and paracrine signaling [318] as well as vesicle transporting (see exosomes section), through which a cell to cell protein exchange takes place. From a cytoanatomical point of view protein misfolding is a process that mainly take place in Endoplasmatic reticulum (ER). Physiologically, misfolded proteins in this compartment are eliminated through a fine molecular cascade (ER-associated degradation or ERAD) that recognizes, ubiquitinates, and retrotranslocates misfolded proteins to the cytosolic 26S proteasome [319]. Some studies indicate cross talk between ERAD pathways and OS in AD, where the inhibition of some key enzymes correlates with APP accumulation and A β production [186]. Interestingly, Tau accumulation can trigger a cycle where the presence of Tau inhibits ERAD (by binding HRD1): in turn ERAD inhibition leads to an increased Tau aggregation (together with other misfolded proteins) [186]. Misfolded protein continue accumulation ultimately lead to the activation of the Unfolded Protein Response, which trigger apoptosis and, consequently, neurodegeneration [320,321]. For a more in deep discussion on ERAD cascade please refer to [186].

12. Metal hypothesis

Metal ions have a significant role in the brain, since they are required to regulate the neuronal activity in the synapses and many other biological functions [186]. There is increasing evidence suggesting that metal balance impairments, either excess or deficiency of metal ions, are involved in a series of processes that can result in neurodegeneration and cell death [322]. These processes are the hallmarks of various neurodegenerative diseases, including AD [130]. Metals are important elements that effect protein conformation, when their homeostasis is disrupted, several protein misfolding events may appear inside the cell [323]. Regarding AD, it was observed that APP play a crucial role in metal homeostasis and increases metal concentrations when its function is altered [324]. Indeed, their concentrations were significantly increased in AD brain samples [325].

The presence of metal ions (especially for zinc, iron and copper) was detected in A β aggregates. In particular, it was observed that the amyloidogenic protein and its precursors have the ability to bind metals [130]. At the same time, bound metals can force proteins to alter their conformations [326], promoting metal-A β complexes generation and AD progression [32]. A β oligomerization reduces the availability of free metal ions. As Cu and Fe are elements needed for the electron transport, their chelation could inhibit oxidative phosphorylation. (which equals to impaired energy production and ROS production) [204]. Other elements, like Al and Fe, are also important determinants of protein conformation. In particular, Al is noteworthy since is a rather strong inducer of A β oligomerization and Al-aggregated A β exhibit tight binding to the surface of neurons and a high predisposition to form fibrillar deposits in cellular models [327]. Supporting its role in AD, Al is considered to be an environmental risk factor for the disease [327]. Notably, increased Fe levels accompany initial aggregation and accumulation of A β in specific brain regions like hippocampus [328]. Moreover, Fe also promotes Tau aggregation [189,329].

The role of metals in the brain is not only limited to protein folding processes. Indeed, they also play important roles in SNC physiologic mechanics, including neurotransmitter synthesis, neural information processing, and neuronal myelination [330]. It was demonstrated that deficiency of these metals produces severe adverse effects on central nervous system functions, especially learning and memory, functions intimately linked with AD development. Metal levels increase promote several pro-OS reactions and lead to (1) ROS generation through a Fenton-like mechanism [32,189,331,332] (2) increased ROS lifetime [333] and (3) oxidative phosphorylation decrease in mitochondria [32].

Finally, it was observed that Zn²⁺ and Cu²⁺ ions can be secreted into synaptic clefts [334–340] and an excess of these metals triggers a cascade of events that lead to hypoperfusion, thus increasing the risk of neuronal death [130]. While this is not one of the principal biologic processes investigated in AD, the role of metals may be more profound than expected. For a more in depth review regarding the role of metals in AD, please refer to [130,322].

13. A role for exosomes?

The detection of A β and Tau in AD subjects' cerebrospinal fluid, evidenced the existence of a mechanism to transport these proteins into the extracellular fluid [341]. While it is possible for cell to reverse their content into the extracellular environment (exocytosis), it has been demonstrated that Tau proteins released into extracellular fluid are cut in their mid-region. This cut prevents Tau aggregation which instead is commonly observed in AD [341]; thus, other transport mechanisms should exist.

Exosomes, are small vesicles on the scale of nanometers, which are widely present in extracellular fluids [341]. Recent data revealed that these vesicles are involved in cell-cell information transfer [341,342], are released by numerous types of cells [343] and carry genetic material, various bioactive proteins and lipids [342]. In particular, recent studies evidenced that exosomes take part into neuron-glia interaction network and their role likely affect important processes [342]. Given, their function, exosomes may contribute to A β and Tau propagation in AD brains [344–348].

Supporting this possibility, there are some observations reporting that in the early stage of AD, some neurons show morphological changes in their endosome compartment, which seems to increase in size; roughly at the same time A β levels begin to rise [349,350]. It was postulated that the

intracellular A β accumulates in endosomes [351]. Noteworthy, these endosomes also contain endocytic multivesicular bodies, whose vesicles are precursors of exosomes [352,353]. Further, A β is also released to the extracellular environment in association with exosomes in cellular models of AD [342,344]. In humans, A β plaques contains traces of molecular markers of exosomes (flotillin-1 and Alix proteins) [342] and A β rich exosomes can be found in the blood of AD subjects [354]. Interestingly, injecting exosomes from AD in the cerebrospinal fluid of animal models can induce AD symptomatology [341]. Further, exosomes from A β -treated astrocytes contain the pro-apoptotic PAR-4 protein and these vesicles cause PAR-4 associated apoptosis in naive cultures [355].

Finally, some data have demonstrated that mRNA and miRNA species are present in exosomes. It is still not known if miRNA levels variation is a cause or consequence of the neurodegenerative process. However, several reports evidenced that specific miRNAs, including miR-9, miR-29a/b, miR-107, miR-124, miR-128, miR-132, miR-134, miR-137 and miR-212 levels are altered in AD and are associated with neurodevelopmental processes [342]. Exosomes, by transporting these transcripts, could propagate AD-related damage to other cells.

As an additional effort to shed some light on Alzheimer background we also enriched our literature research with a pathway analysis based on recent genetic findings (see Table 1 and supplementary Table 1) in order to evidence functionally organized networks and related biological processes. Multiple clusters (a total of 6) were evidenced from the initial selection of genes. Each one is linked to different biological processes (Gene Ontology: Biological Processes database—<http://geneontology.org/>) [356]. The results are reported as supplementary material.

14. Mitochondria

Concluding the discussion on the biological processes, we want to focus on a central organelle which may very well occupy a central position in LOAD pathophysiology. Mitochondria are the cellular central point for energy-related processes.

The correlation of Mitochondria with AD is known in literature, supported by both genetic and molecular observations and is believed to occupy a relevant position in the pathogenesis of the disease [204]. A trend of inheritance do exist in AD cases: children of female AD subjects are at higher risk for developing AD than children of male AD subjects [357]. Maternally inherited AD may account for over 20% of all LOAD cases [357], supporting a possible involvement of mtDNA genes [358–360]. A number of studies have demonstrated that mitochondrial integrity declines with age, affecting multiple brain functions such as memory, learning and sensory processes [307,361], which are commonly impaired in AD.

The mitochondrial cascade hypothesis was one of the potential theories advanced to explain AD pathogenesis [200,204]. According to this hypothesis, polymorphisms within mtDNA and nuclear genes encoding for Electron Transport Chain (ETC) determine the efficiency of mitochondrial energy production as well as the accumulation rate of ROS byproducts. ROS production rate is strictly related to mtDNA alterations.

In particular, ROS accumulation rate is a good indicator of mitochondrial life and his energy production efficacy, since higher is ROS rate, higher is the accumulation of mtDNA damage. The decrease of energy production efficacy is mainly due to alterations of the complexes (I–IV), which are one of the most documented changes in AD [200,362]. As observed in the previous section, OS also triggers lipid peroxidation and protein oxidation.

Interestingly, some recent data pointed on the regulation of nuclear components through ROS signaling [363]. In details, mitochondrial ROS can upregulate the expression of assembly factors of I, II, III, and IV complexes [364], evidencing the bidirectional talk between mitochondria and nucleus [186]. Additionally, miRNAs seem to be implicated in the regulation of ROS signaling [189].

With the increase of ROS three different responses are triggered: (1) A β production from APP, which further increase ROS generation; (2) pro-apoptotic cascade by releasing factor such as cytochrome C; (3) hypoxic signaling cascade, which provides a signal for re-entry into mitotic cycling (destined to fail in neurons) and with resultant aneuploidy, Tau phosphorylation, and neurofibrillary tangle formation [204]. Phosphorylated Tau increases dynamin-related protein 1 (DRP1) levels, which ultimately cause excessive mitochondrial fragmentation [200].

The hypothesis etiologically divides LOAD from EOAD. Focusing on A β production, in LOAD this may represent a compensatory event occurring in response to the primary mitochondrial pathology, while in EOAD A β production is strictly a toxic phenomenon.

This fact was included in a recent redefinition of the mitochondrial hypothesis which was distinguished into primary and secondary mitochondrial cascade [200], stressing the distinction between mitochondrial dysfunction-induced (primary) and A β induced (secondary) AD onset. In the latter the dysfunction is merely an intermediated step.

Either way, A β production have harmful effects on mitochondria. According to literature data, A β contributes to OS [365,366]. In addition, A β promotes mitochondria fragmentation through S-nitrosylation of Drp1 and consequent augmented GTPase activity [307]. It also inhibits transmembrane translocase on mitochondria through cyclophylin D interactions, increases mitochondrial fission and permeability and downregulates mitochondrial fusion [200].

Mitochondria are essential organelles in every human cell type since play a key role in cell survival or death, regulating both energy metabolism and apoptotic pathways. However, their role in neurons is more than that, since these cells have high energy demands, in particular for neuronal synaptic activity [367] and neurotransmission processes in general [200]. In high OS conditions, mitochondria has a limited serotonergic efficiency caused by membrane permeabilization and altered serotonergic metabolism. Monoaminergic activity in general is reduced by mitochondria dysfunction [200]. Finally, some recent literature data indicated that mitochondria actively interact with ER through localized subdomain named mitochondria-associated ER membranes. The activity within these areas is increased in AD cells [368]. Notably, this data may infer to a possible bridge between mitochondria functions and cells protein production, including potential protein misfolding-related processes.

15. The impact of the different biological contributors toward the risk

With the exception of around 5% of autosomal dominant AD mutations (EOAD), a number of other biological processes contribute to AD cases world-wide. As introduced in the previous sections it is unlikely that a single factor (or mechanics) is responsible for synaptic dysfunctions and the other pathological changes that occur in AD subjects' brain. In this paragraph, we want to briefly illustrates how the several pathways discussed, may be linked through transduction signaling cascades. While the latter may be not directly causative of AD, they undoubtedly have a key role in AD propagation and symptoms manifestation.

As discussed in the Mitochondria section, whether A β production is the causative event (EOAD) or caused by other processes, including mitochondria dysfunction, its cytotoxicity is primarily exerted on mitochondria. The two main consequences are an increased OS and a reduced (aerobic) energy production. ROS acts on lipids promoting their peroxidation and perturbing their homeostasis. Physiologically, the latter are used not only for energy production but also as secondary messengers [369]. Several genes involved in lipid homeostasis, including APOE, CLU, SORL1 and ABCA7 were correlated with AD [370]. Polyunsaturated fatty acids could participate in myriad signal transduction within the brain directly or after enzymatic conversion to a series of mediators. For example, Activated sphingolipids are signaling molecules that serve as intracellular second messengers [371]. Their shortage has a potentiation effect on A β secretion [342], thus causing its accumulation (and plaques formation). Literature data indicates that this shortage is often observed in AD brains, especially in the inferior temporal and middle frontal gyri [369]. Further, Cholesterol and its esters low concentration correlate with increased of A β production [370].

Cellular Ca²⁺ signals regulate several important processes in neurons [372,373]. Literature evidences indicate that its dyshomeostasis may play a key role in the pathogenesis of AD [374,375]. Interestingly, there is ample some data reports that Ca²⁺ dysregulation may even precede the formation of A β plaques and neurofibrillary tangles in AD brain, suggesting disruption in cytoplasmic Ca²⁺ is an earlier event [372,376].

Intracellular Ca²⁺ is usually stored in the ER. Its release in the cytosol is controlled by RyRs and inositol 1,4,5-trisphosphate receptors (InsP₃R) [377]. Ca²⁺ intake from extracellular compartment is controlled by store-operated Ca²⁺ entry (SOCE) pathway and, by voltage-gated Ca²⁺ channels (VGCC) in neurons. Also, it has been shown Ca²⁺ influx through the CALHM1 channel or NMDAR stimulates α -secretase processing of APP [375], thus protecting from A β accumulation. It was observed that high OS state and A β aggregates can interfere with Ca²⁺ homeostasis as they can trigger Ca²⁺ release from ER stores through the InsP₃R and RyR [375,378]. Increased Ca²⁺ levels in the cells interferes with the physiological function of VGCCs, thus impairing neurotransmission [373]. Imbalanced cellular Ca²⁺ also contributes to pathophysiological conditions such as accumulation of A β plaques and neurofibrillary tangles, protein misfolding, necrosis, apoptosis, autophagy deficits, and degeneration [375,379–381]. Finally, excess cytosolic Ca²⁺ exacerbates mitochondria dysfunction and dysregulates KIF5-Miro-Trak-mediated mitochondrial transport to synapses [200]. For a more detailed discussion on Ca²⁺ please refer to [375,381].

G protein-coupled receptors (GPCRs) regulate cell responses to external molecules, like neurotransmitters [382,383] and it was demonstrated that their action is involved in both long- and short-term memory biological processes [384], known to be impaired in AD.

GPCRs can control genetic expression through their interaction with the extracellular signal-regulated kinases (ERKs or ERK1/2). ERKs is involved in a wide array of signals and signal transduction pathways, including the ones involved in the formation of memory traces [385,386] and the ones involved in the regulation of neuronal plasticity (both protein synthesis and protein phosphorylation in dendrites) [387,388]. Interestingly, G-GPCRs produce the secondary messenger inositol 1,4,5-triphosphate (IP₃) which binds to IP₃R on ER and efflux the Ca²⁺ into cytoplasm [381].

Dysfunction of the mammalian target of rapamycin (mTOR) pathway has been implicated in both neurodevelopmental and neurodegenerative diseases, including AD [389–392]. Numerous studies have demonstrated mTOR hyperactivation in AD brain [393]. Physiologically, mTOR is a regulator of survival, differentiation, and development of neurons [394]. It was observed that A β

accumulation activates mTOR pathway through phosphorylation of mTOR inhibitor PRAS40 [390]. Further, hypoenergetic states seems also to trigger this pathway [395]. The aberrant activation of the mTOR pathway leads to an alteration of protein synthesis and autophagy's mechanisms which in turn are related to the accumulation of inclusion bodies, a common feature of AD ([391,392,395] and references therein). During neural development, mTOR regulates neuronal growth, differentiation and interconnectivity [396–398]. It regulates homeostasis by directly influencing gene transcription, protein and lipid synthesis and organelle biogenesis and maintenance [391]. This pathway appears to be influenced by ERKs and GPCRs action [399]. In turn mTOR pathway inhibits PI3K/Akt signaling, thus triggering GSK-3 β activation [394]. Accumulating evidence also supports a correlation between alterations in mTOR signaling and aging [400]. Finally, cell cultures from animal models of AD interestingly display a reduced mTOR signaling [401]. Alterations of this pathway may be linked to the excessive burden of misfolded proteins [399].

The balance of pro- and anti-apoptotic Bcl-2 family proteins (i.e. Bcl-2, Bad, Bim and Bax) has been known to have a role in neuronal cell death [402]. The levels of these proteins are altered in AD brains neurons [402]. Although the exact molecular details are not completely described, it is believed that A β is able to increase Bim and decrease Bcl-2 levels [402]. The pro-apoptotic protein (Bim) and anti-apoptotic protein (Bcl-2) are known to exert their activity through heterodimerization with Bax protein [403]. Bax is usually localized in the cytosol or associated with the membranes of mitochondria and/or ER [404]. Upon activation, Bax changes conformation and dwells into mitochondrial membranes where it promotes the release of cytochrome C and triggers apoptosis [405,406]. With the presence A β , Bim overexpression leads to the activation of Bax which triggers neuronal cell death [402]. The activation of this cascade is strictly correlated with the neurodegenerative events associated with AD.

The increase of A β production negatively impact survivability of neurons. In physiological conditions, it has been observed that the predominant APP-cleaved fragment (sAPP α) interacts with IGF1 and insulin receptor. This interaction together with different neurotrophins (NGF, BDNF) mediates the activation of the PI3K/Akt pathway. This action promotes neuronal survival, possibly through phosphorylation and inhibition of glycogen synthase kinase-3 β (GSK-3 β) [395]. The accumulation of A β directly compete with the effects mediated by the different neurotrophins, downregulating PI3K/Akt pathway and ultimately overactivating GSK-3 β [395]. The activation of GSK-3 β accounts for several features of AD such as increased A β production and plaque accumulation [407], reduced memory performance, neurogenesis and synaptogenesis (astroglial-related) [395,408], as well as increased Tau phosphorylation, Inflammation (microglia) and neurodegeneration (mitochondrial intrinsic apoptotic and the death receptor-mediated extrinsic apoptotic pathways) [409]. Additionally, A β direct competition with insulin for its receptor, increases the latter levels in the brain microenvironment. High insulin levels promote inflammatory responses in the brain, based on increased TNF α , interleukin 1 β and 6 (IL1 β and IL6) [395].

Following the progression of AD, brain neurons find themselves in a status of reduced energy production and the microenvironment has elevated pro-inflammatory cytokines (see previous sections). In these conditions Notch signaling is upregulated [410]. Physiologically, Notch signaling has an essential role in vascular development and (neuro) angiogenesis through the modulation of VEGFR2 [411]. It is also associated with synaptic plasticity, and glial cell activation [412,413]. Recently, some data hypothesized that Notch signaling may be implicated AD as Notch1 is often found in A β plaques [410,414].

Despite its physiological role, it has been observed that chronic activation of Notch1 negatively impact the brain microenvironment, in particular the delicate connection of the brain with cardiovascular system. Indeed, Notch signaling, in association with VEGF, has been demonstrated to cause impaired blood flow, further reducing the nutrients intake by neurons (worsening the already weak energetic state). Notch also induce BBB leakages which has severe impact on the brain and may accelerate A β accumulation [410]. Finally, Notch is involved in regulating APP proteolysis [410]. Despite the still lacking data, Notch signaling, is a potential candidate that links various AD related processes with vascular impairments and may explain how they interact.

A β accumulation in AD is strictly linked with the upregulation of DKK1 gene [415,416]. The translated protein is an inhibitor of Wnt (canonical) pathway. Physiologically, Wnt signaling has important functions linked to plasticity and cognition. Among these, it promotes the endocytosis, the recycle of vesicles and modulate neurotransmitter release [417] and receptor transcription [418]. It also facilitates the induction of LTP [419].

Wnt signaling has three major cascades: the canonical Wnt/ β -catenin, planar cell polarity (PCP), and Wnt/Ca²⁺ pathways. Interestingly, Wnt/ β -catenin and Wnt/PCP modulate synapse maturation; specifically, Wnt/ β -catenin promotes synapses stability, while Wnt/PCP promotes synapse retraction [420]. With the over expression of DKK1 by A β , the canonical pathway is suppressed skewing the balance toward the non-canonical PCP pathway. This event accelerates synaptic damage [420] and participate to the progression of cognitive symptoms of AD. Further, canonical Wnt/ β -catenin pathway promotes the phosphorylation (and inactivation) of GSK-3 β [419]. Thus, its inhibition produces similar effects to the ones described in PI3K/Akt signaling. The diagram in Figure 1 reports a simplified view of the pathways connections discussed.

16. Methods

The main aim of literature research was to extrapolate biological pathways potentially involved with the etiopathogenesis of Alzheimer disease. Phase (1) A preliminary research was done selecting reviews (from 2013 to 2018) on PubMed using the keywords: Alzheimer AND (Genetics OR Biological cascades) in order to prepare a background for the paper. At the time of the research, 1427 elements were obtained. Data were screened for genes association and molecular cascades. Phase (2) Genes with multiple references of association with AD plus the ones extracted from Alzgene.org dataset were grouped and clustered (using Gene Ontology Database for gene-gene correlations). Cluster data from phase 2 was also used to perform pathway analyses (see exploratory data for details, and Supplementary Figure 1 for a summary). The data obtained was used as criteria for pathway selection. Phase (3) Data from phase 1 and phase 2 were used to a more detailed research: each pathway was used as keyword (Alzheimer AND Pathway name) for a refined literature research on PubMed and Google scholar. A total of 2204 (Immune System), 3836 (Inflammation), 4653 (Oxidative Stress), 2792 (Energy Metabolism), 746 (Autophagy), 130 (Exosomes-mechanics), 469 (Vascular-Related Mechanics), 4150 (Signal Transduction) and 169 (Neurodevelopment and Neurotransmission) works, partially overlapping, were obtained. Starting from the most recent ones, literature findings were filtered and screened for information.

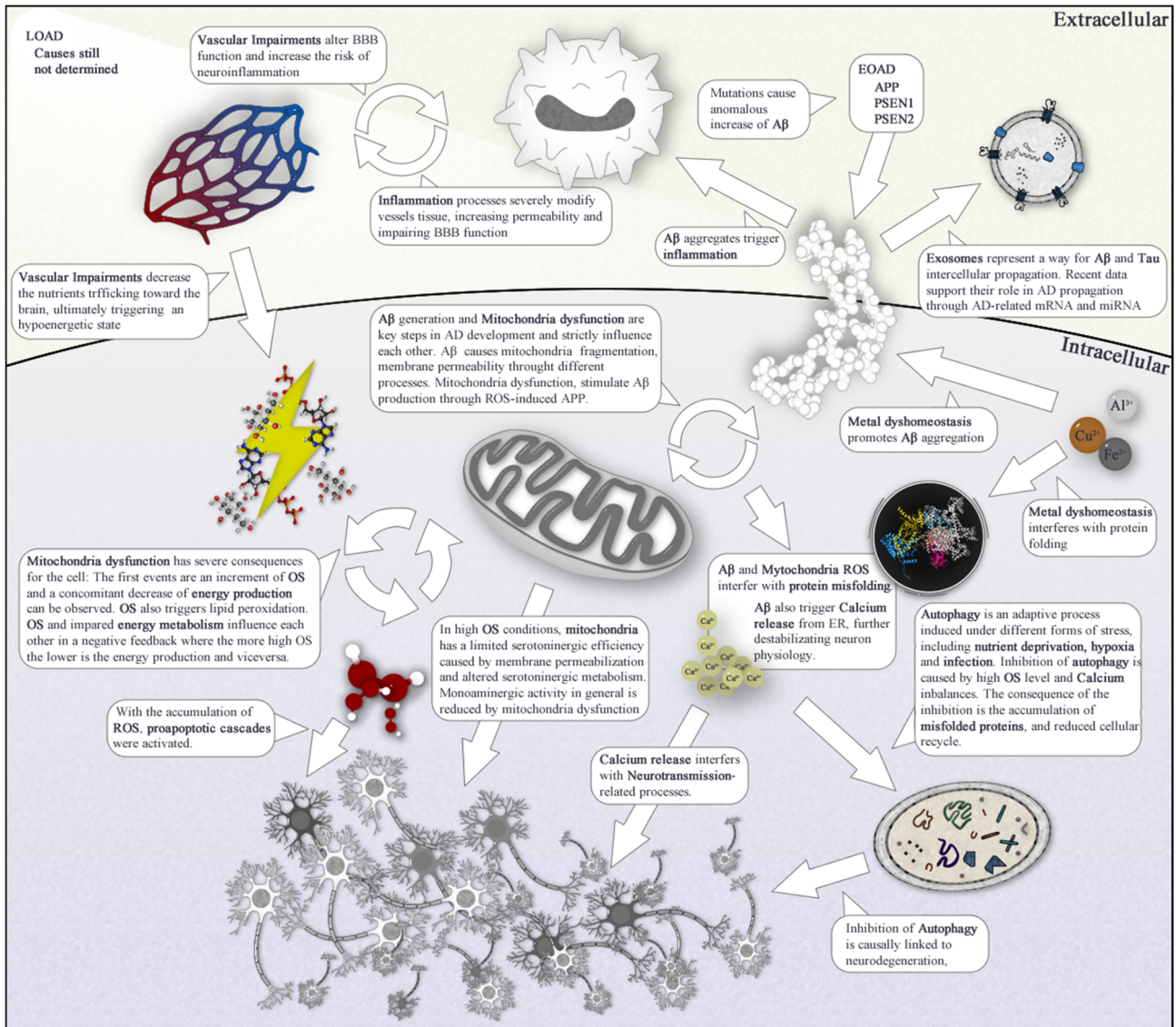


Figure 1. Graphical view of pathways connections.

17. Conclusive remarks

AD is one of the principal causes of disability and decreased quality of life among the elderly. Despite the active research on the field, many fundamental questions remain regarding the molecular background of this disease.

The old theories linking AD to the genes directly related to A β formation and Tau hyperphosphorylation cannot explain the complexity of AD and neither give a definite target for treatment. Indeed, the conventional treatments based on the A β hypothesis have failed to treat or prevent AD. Around 99.6% of drug candidates targeting amyloid pathways, including β secretase inhibitors, γ secretase inhibitors, and A β itself, do not have a significant therapeutic effect on AD subjects [9].

Recent studies and the new hypotheses for AD have indicated many new potential targets [421–424] even though we are still far from a definitive solution. The evidences derived from the study of the disease and its treatment, together with the current concept of AD as “multifactorial pathology”,

clearly indicates the necessity to consider wider approaches which include the complex interactions between different pathways. Part of these biological processes was introduced and described in this review.

Other than the familial form-associated genes (PSEN1, PSEN2, APP) and the high risk variants of APOe, the AD background is obscure. Its complexity is not to be underestimated and it is likely that multiple factor may trigger AD pathological changes in the brain. From what it was discussed, whether the etiological cause, AD progression hit Mitochondria at some point. The central role of this organelle is plausible, considering its importance in neurons. It's destabilization could arise following A β accumulation (EOAD main etiological cause), reduced intake of nutrients (which makes mitochondria sensible to vascular impairments). Even aging itself (accumulation of damage on mtDNA) slowly decrease mitochondria efficiency. However, mitochondria dysfunction alone is not enough to explain why the damage is mainly restricted on neurons. Additionally, all the propagation mechanics remain elusive and the molecular cascades behind the propagation path (see Box 1) is not completely understood.

The key to further deepen the studies of AD understands how these processes interact with each other. Thereby, a focus on multiple pathways or functional cascades may be useful to pinpoint novel potential target for AD treatment.

Conflicts of interest

The authors declare there are no conflicts of interest.

References

1. GBD 2015 Mortality and Causes of Death Collaborators (2016) Global, regional, and national life expectancy, all-cause mortality, and cause-specific mortality for 249 causes of death, 1980–2015: a systematic analysis for the Global Burden of Disease Study 2015. *Lancet* 388: 1459–1544.
2. APA (2020) 2020 Alzheimer's disease facts and figures. *Alzheimers Dement*, in press.
3. Brookmeyer R, Johnson E, Ziegler-Graham K, et al. (2007) Forecasting the global burden of Alzheimer's disease. *Alzheimers Dement* 3: 186–191.
4. Prince M, Bryce R, Albanese E, et al. (2013) The global prevalence of dementia: a systematic review and metaanalysis. *Alzheimers Dement* 9: 63–75. e2.
5. Alzheimer's Association (2018) 2018 Alzheimer's disease facts and figures. *Alzheimers Dement* 14: 367–429.
6. Perl DP (2010) Neuropathology of Alzheimer's disease. *Mt Sinai J Med* 77: 32–42.
7. Kumar A, Tsao JW (2018) *Alzheimer Disease*. StatPearls. Treasure Island (FL).
8. Braak H, Braak E (1996) Development of Alzheimer-related neurofibrillary changes in the neocortex inversely recapitulates cortical myelogenesis. *Acta Neuropathol* 92: 197–201.
9. Cummings JL, Morstorf T, Zhong K (2014) Alzheimer's disease drug-development pipeline: few candidates, frequent failures. *Alzheimers Res Ther* 6: 37.
10. Hussein W, Saglik BN, Levent S, et al. (2018) Synthesis and biological evaluation of new cholinesterase inhibitors for Alzheimer's disease. *Molecules* 23: 2033.

11. Leblhuber F, Steiner K, Schuetz B, et al. (2018) Probiotic supplementation in patients with Alzheimer's dementia—An explorative intervention study. *Curr Alzheimer Res* 15: 1106–1113.
12. Farlow MR, Salloway S, Tariot PN, et al. (2010) Effectiveness and tolerability of high-dose (23 mg/d) versus standard-dose (10 mg/d) donepezil in moderate to severe Alzheimer's disease: A 24-week, randomized, double-blind study. *Clin Ther* 32: 1234–1251.
13. Homma A, Atarashi H, Kubota N, et al. (2016) Efficacy and safety of sustained release donepezil high dose versus immediate release donepezil standard dose in Japanese patients with severe Alzheimer's disease: a randomized, double-blind trial. *J Alzheimers Dis* 52: 345–357.
14. Winblad B, Kilander L, Eriksson S, et al. (2006) Donepezil in patients with severe Alzheimer's disease: double-blind, parallel-group, placebo-controlled study. *Lancet* 367: 1057–1065.
15. Feldman H, Gauthier S, Hecker J, et al. (2005) Efficacy and safety of donepezil in patients with more severe Alzheimer's disease: a subgroup analysis from a randomized, placebo-controlled trial. *Int J Geriatr Psychiatry* 20: 559–569.
16. Black SE, Doody R, Li H, et al. (2007) Donepezil preserves cognition and global function in patients with severe Alzheimer disease. *Neurology* 69: 459–469.
17. Howard R, McShane R, Lindesay J, et al. (2015) Nursing home placement in the donepezil and memantine in moderate to severe Alzheimer's disease (DOMINO-AD) trial: secondary and post-hoc analyses. *Lancet Neurol* 14: 1171–1181.
18. Bond M, Rogers G, Peters J, et al. (2012) The effectiveness and cost-effectiveness of donepezil, galantamine, rivastigmine and memantine for the treatment of Alzheimer's disease (review of Technology Appraisal No. 111): a systematic review and economic model. *Health Technol Assess* 16: 1–470.
19. Zhang N, Wei C, Du H, et al. (2015) The effect of memantine on cognitive function and behavioral and psychological symptoms in mild-to-moderate Alzheimer's disease patients. *Dement Geriatr Cogn Disord* 40: 85–93.
20. Molinuevo JL, Berthier ML, Rami L (2011) Donepezil provides greater benefits in mild compared to moderate Alzheimer's disease: implications for early diagnosis and treatment. *Arch Gerontol Geriatr* 52: 18–22.
21. Takeda A, Loveman E, Clegg A, et al. (2006) A systematic review of the clinical effectiveness of donepezil, rivastigmine and galantamine on cognition, quality of life and adverse events in Alzheimer's disease. *Int J Geriatr Psychiatry* 21: 17–28.
22. Lam B, Masellis M, Freedman M, et al. (2013) Clinical, imaging, and pathological heterogeneity of the Alzheimer's disease syndrome. *Alzheimers Res Ther* 5: 1–14.
23. Jack CR Jr, Albert MS, Knopman DS, et al. (2011) Introduction to the recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimers Dement* 7: 257–262.
24. Albert MS, DeKosky ST, Dickson D, et al. (2011) The diagnosis of mild cognitive impairment due to Alzheimer's disease: recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimers Dement* 7: 270–279.
25. McKhann GM, Knopman DS, Chertkow H, et al. (2011) The diagnosis of dementia due to Alzheimer's disease: recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimers Dement* 7: 263–269.

26. Beason-Held LL, Goh JO, An Y, et al. (2013) Changes in brain function occur years before the onset of cognitive impairment. *J Neurosci* 33: 18008–18014.
27. O'Brien RJ, Wong PC (2011) Amyloid precursor protein processing and Alzheimer's disease. *Annu Rev Neurosci* 34: 185–204.
28. Wilkins HM, Swerdlow RH (2017) Amyloid precursor protein processing and bioenergetics. *Brain Res Bull* 133: 71–79.
29. Vassar R, Kovacs DM, Yan R, et al. (2009) The beta-secretase enzyme BACE in health and Alzheimer's disease: regulation, cell biology, function, and therapeutic potential. *J Neurosci* 29: 12787–12794.
30. De Strooper B, Vassar R, Golde T (2010) The secretases: enzymes with therapeutic potential in Alzheimer disease. *Nat Rev Neurol* 6: 99–107.
31. Walsh DM, Klyubin I, Fadeeva JV, et al. (2002) Naturally secreted oligomers of amyloid beta protein potently inhibit hippocampal long-term potentiation in vivo. *Nature* 416: 535–539.
32. Chen YY, Wang MC, Wang YN, et al. (2020) Redox signaling and Alzheimer's disease: from pathomechanism insights to biomarker discovery and therapy strategy. *Biomark Res* 8: 42.
33. Alonso AC, Zaidi T, Grundke-Iqbal I, et al. (1994) Role of abnormally phosphorylated tau in the breakdown of microtubules in Alzheimer disease. *Proc Natl Acad Sci U S A* 91: 5562–5566.
34. Alonso AC, Grundke-Iqbal I, Iqbal K (1996) Alzheimer's disease hyperphosphorylated tau sequesters normal tau into tangles of filaments and disassembles microtubules. *Nat Med* 2: 783–787.
35. Braak H, Alafuzoff I, Arzberger T, et al. (2006) Staging of Alzheimer disease-associated neurofibrillary pathology using paraffin sections and immunocytochemistry. *Acta Neuropathol* 112: 389–404.
36. Morrison BM, Hof PR, Morrison JH (1998) Determinants of neuronal vulnerability in neurodegenerative diseases. *Ann Neurol* 44: S32–44.
37. Braak H, Braak E (1985) On areas of transition between entorhinal allocortex and temporal isocortex in the human brain. Normal morphology and lamina-specific pathology in Alzheimer's disease. *Acta Neuropathol* 68: 325–332.
38. Mesulam MM (1999) Neuroplasticity failure in Alzheimer's disease: bridging the gap between plaques and tangles. *Neuron* 24: 521–529.
39. Serrano-Pozo A, Frosch MP, Masliah E, et al. (2011) Neuropathological alterations in Alzheimer disease. *Cold Spring Harb Perspect Med* 1: a006189.
40. Goldman JS, Hahn SE, Catania JW, et al. (2011) Genetic counseling and testing for Alzheimer disease: joint practice guidelines of the American College of Medical Genetics and the National Society of Genetic Counselors. *Genet Med* 13: 597–605.
41. Bekris LM, Yu CE, Bird TD, et al. (2010) Genetics of Alzheimer disease. *J Geriatr Psychiatry Neurol* 23: 213–227.
42. Brickell KL, Steinbart EJ, Rumbaugh M, et al. (2006) Early-onset Alzheimer disease in families with late-onset Alzheimer disease: a potential important subtype of familial Alzheimer disease. *Arch Neurol* 63: 1307–1311.
43. Bird TD (2008) Genetic aspects of Alzheimer disease. *Genet Med* 10: 231–239.
44. Serretti A, Artioli P, Quartesan R, et al. (2005) Genes involved in Alzheimer's disease, a survey of possible candidates. *J Alzheimers Dis* 7: 331–353.

45. Mahley RW (1988) Apolipoprotein E: cholesterol transport protein with expanding role in cell biology. *Science* 240: 622–630.
46. Puglielli L, Tanzi RE, Kovacs DM (2003) Alzheimer's disease: the cholesterol connection. *Nat Neurosci* 6: 345–351.
47. Harold D, Abraham R, Hollingworth P, et al. (2009) Genome-wide association study identifies variants at CLU and PICALM associated with Alzheimer's disease. *Nat Genet* 41: 1088–1093.
48. Lambert JC, Heath S, Even G, et al. (2009) Genome-wide association study identifies variants at CLU and CR1 associated with Alzheimer's disease. *Nat Genet* 41: 1094–1099.
49. Saunders AM, Strittmatter WJ, Schmechel D, et al. (1993) Association of apolipoprotein E allele epsilon 4 with late-onset familial and sporadic Alzheimer's disease. *Neurology* 43: 1467–1472.
50. Farrer LA, Cupples LA, Haines JL, et al. (1997) Effects of age, sex, and ethnicity on the association between apolipoprotein E genotype and Alzheimer disease. A meta-analysis. APOE and Alzheimer Disease Meta Analysis Consortium. *JAMA* 278: 1349–1356.
51. Zlokovic BV (2013) Cerebrovascular effects of apolipoprotein E: implications for Alzheimer disease. *JAMA Neurol* 70: 440–444.
52. Ghebremedhin E, Schultz C, Braak E, et al. (1998) High frequency of apolipoprotein E epsilon4 allele in young individuals with very mild Alzheimer's disease-related neurofibrillary changes. *Exp Neurol* 153: 152–155.
53. Shi Y, Yamada K, Liddelow SA, et al. (2017) ApoE4 markedly exacerbates tau-mediated neurodegeneration in a mouse model of tauopathy. *Nature* 549: 523–527.
54. Ulrich JD, Ulland TK, Mahan TE, et al. (2018) ApoE facilitates the microglial response to amyloid plaque pathology. *J Exp Med* 215: 1047–1058.
55. Mahley RW (2016) Central nervous system lipoproteins: ApoE and regulation of cholesterol metabolism. *Arterioscler Thromb Vasc Biol* 36: 1305–1315.
56. Yamazaki Y, Zhao N, Caulfield TR, et al. (2019) Apolipoprotein E and Alzheimer disease: pathobiology and targeting strategies. *Nat Rev Neurol* 15: 501–518.
57. Verghese PB, Castellano JM, Garai K, et al. (2013) ApoE influences amyloid-beta (A β) clearance despite minimal apoE/A β association in physiological conditions. *Proc Natl Acad Sci U S A* 110: E1807–1816.
58. Huang Y, Mahley RW (2014) Apolipoprotein E: structure and function in lipid metabolism, neurobiology, and Alzheimer's diseases. *Neurobiol Dis* 72: 3–12.
59. Miyata M, Smith JD (1996) Apolipoprotein E allele-specific antioxidant activity and effects on cytotoxicity by oxidative insults and beta-amyloid peptides. *Nat Genet* 14: 55–61.
60. Montine KS, Olson SJ, Amarnath V, et al. (1997) Immunohistochemical detection of 4-hydroxy-2-nonenal adducts in Alzheimer's disease is associated with inheritance of APOE4. *Am J Pathol* 150: 437–443.
61. Ramassamy C, Averill D, Beffert U, et al. (1999) Oxidative damage and protection by antioxidants in the frontal cortex of Alzheimer's disease is related to the apolipoprotein E genotype. *Free Radic Biol Med* 27: 544–553.
62. Chen Y, Zhang J, Zhang B, et al. (2016) Targeting insulin signaling for the treatment of Alzheimer's disease. *Curr Top Med Chem* 16: 485–492.
63. Mahley RW, Huang Y (2012) Apolipoprotein e sets the stage: response to injury triggers neuropathology. *Neuron* 76: 871–885.

64. Zhong N, Weisgraber KH (2009) Understanding the association of apolipoprotein E4 with Alzheimer disease: clues from its structure. *J Biol Chem* 284: 6027–6031.
65. Reiman EM, Caselli RJ, Chen K, et al. (2001) Declining brain activity in cognitively normal apolipoprotein E epsilon 4 heterozygotes: a foundation for using positron emission tomography to efficiently test treatments to prevent Alzheimer's disease. *Proc Natl Acad Sci U S A* 98: 3334–3339.
66. Reiman EM, Chen K, Alexander GE, et al. (2004) Functional brain abnormalities in young adults at genetic risk for late-onset Alzheimer's dementia. *Proc Natl Acad Sci U S A* 101: 284–289.
67. Skotte N (2010) Genome-wide association studies identify new interesting loci for late-onset Alzheimer's disease. *Clin Genet* 77: 330–332.
68. Chouraki V, Seshadri S (2014) Genetics of Alzheimer's disease. *Adv Genet* 87: 245–294.
69. Bertram L, McQueen MB, Mullin K, et al. (2007) Systematic meta-analyses of Alzheimer disease genetic association studies: the AlzGene database. *Nat Genet* 39: 17–23.
70. Goldgaber D, Lerman MI, McBride OW, et al. (1987) Characterization and chromosomal localization of a cDNA encoding brain amyloid of Alzheimer's disease. *Science* 235: 877–880.
71. Kang J, Lemaire HG, Unterbeck A, et al. (1987) The precursor of Alzheimer's disease amyloid A4 protein resembles a cell-surface receptor. *Nature* 325: 733–736.
72. Robakis NK, Wisniewski HM, Jenkins EC, et al. (1987) Chromosome 21q21 sublocalisation of gene encoding beta-amyloid peptide in cerebral vessels and neuritic (senile) plaques of people with Alzheimer disease and Down syndrome. *Lancet* 1: 384–385.
73. Tanzi RE, Gusella JF, Watkins PC, et al. (1987) Amyloid beta protein gene: cDNA, mRNA distribution, and genetic linkage near the Alzheimer locus. *Science* 235: 880–884.
74. Sherrington R, Rogaev EI, Liang Y, et al. (1995) Cloning of a gene bearing missense mutations in early-onset familial Alzheimer's disease. *Nature* 375: 754–760.
75. Levy-Lahad E, Wasco W, Poorkaj P, et al. (1995) Candidate gene for the chromosome 1 familial Alzheimer's disease locus. *Science* 269: 973–977.
76. Zhao QF, Yu JT, Tan MS, et al. (2015) ABCA7 in Alzheimer's Disease. *Mol Neurobiol* 51: 1008–1016.
77. Hollingworth P, Harold D, Sims R, et al. (2011) Common variants at ABCA7, MS4A6A/MS4A4E, EPHA1, CD33 and CD2AP are associated with Alzheimer's disease. *Nat Genet* 43: 429–435.
78. Marcello E, Saraceno C, Musardo S, et al. (2013) Endocytosis of synaptic ADAM10 in neuronal plasticity and Alzheimer's disease. *J Clin Invest* 123: 2523–2538.
79. Corder EH, Saunders AM, Risch NJ, et al. (1994) Protective effect of apolipoprotein E type 2 allele for late onset Alzheimer disease. *Nat Genet* 7: 180–184.
80. Royston MC, Mann D, Pickering-Brown S, et al. (1994) Apolipoprotein E epsilon 2 allele promotes longevity and protects patients with Down's syndrome from dementia. *Neuroreport* 5: 2583–2585.
81. Coimbra JRM, Marques DFF, Baptista SJ, et al. (2018) Highlights in BACE1 inhibitors for Alzheimer's disease treatment. *Front Chem* 6: 178.
82. Holsinger RM, Goense N, Bohorquez J, et al. (2013) Splice variants of the Alzheimer's disease beta-secretase, BACE1. *Neurogenetics* 14: 1–9.

83. Tan MS, Yu JT, Tan L (2013) Bridging integrator 1 (BIN1): form, function, and Alzheimer's disease. *Trends Mol Med* 19: 594–603.
84. Seshadri S, Fitzpatrick AL, Ikram MA, et al. (2010) Genome-wide analysis of genetic loci associated with Alzheimer disease. *JAMA* 303: 1832–1840.
85. Liu G, Zhang S, Cai Z, et al. (2013) BIN1 gene rs744373 polymorphism contributes to Alzheimer's disease in East Asian population. *Neurosci Lett* 544: 47–51.
86. Miyashita A, Koike A, Jun G, et al. (2013) SORL1 is genetically associated with late-onset Alzheimer's disease in Japanese, Koreans and Caucasians. *PLoS One* 8: e58618.
87. Harms M, Benitez BA, Cairns N, et al. (2013) C9orf72 hexanucleotide repeat expansions in clinical Alzheimer disease. *JAMA Neurol* 70: 736–741.
88. Jiang T, Yu JT, Hu N, et al. (2014) CD33 in Alzheimer's disease. *Mol Neurobiol* 49: 529–535.
89. Deng YL, Liu LH, Wang Y, et al. (2012) The prevalence of CD33 and MS4A6A variant in Chinese Han population with Alzheimer's disease. *Hum Genet* 131: 1245–1249.
90. Tan L, Yu JT, Zhang W, et al. (2013) Association of GWAS-linked loci with late-onset Alzheimer's disease in a northern Han Chinese population. *Alzheimers Dement* 9: 546–553.
91. Liu G, Wang H, Liu J, et al. (2014) The CLU gene rs11136000 variant is significantly associated with Alzheimer's disease in Caucasian and Asian populations. *Neuromolecular Med* 16: 52–60.
92. Komatsu M, Shibata N, Kuerban B, et al. (2011) Genetic association between clusterin polymorphisms and Alzheimer's disease in a Japanese population. *Psychogeriatrics* 11: 14–18.
93. Yu JT, Ma XY, Wang YL, et al. (2013) Genetic variation in clusterin gene and Alzheimer's disease risk in Han Chinese. *Neurobiol Aging* 34: 1921. e17–23.
94. Jin C, Li W, Yuan J, et al. (2012) Association of the CR1 polymorphism with late-onset Alzheimer's disease in Chinese Han populations: a meta-analysis. *Neurosci Lett* 527: 46–49.
95. Crehan H, Holton P, Wray S, et al. (2012) Complement receptor 1 (CR1) and Alzheimer's disease. *Immunobiology* 217: 244–250.
96. Nacmias B, Piaceri I, Bagnoli S, et al. (2014) Genetics of Alzheimer's disease and frontotemporal dementia. *Curr Mol Med* 14: 993–1000.
97. Perry DC, Lehmann M, Yokoyama JS, et al. (2013) Progranulin mutations as risk factors for Alzheimer disease. *JAMA Neurol* 70: 774–778.
98. Li HL, Lu SJ, Sun YM, et al. (2013) The LRRK2 R1628P variant plays a protective role in Han Chinese population with Alzheimer's disease. *CNS Neurosci Ther* 19: 207–215.
99. Zhao Y, Ho P, Yih Y, et al. (2011) LRRK2 variant associated with Alzheimer's disease. *Neurobiol Aging* 32: 1990–1993.
100. Parikh I, Fardo DW, Estus S (2014) Genetics of PICALM expression and Alzheimer's disease. *PLoS One* 9: e91242.
101. Chung SJ, Lee JH, Kim SY, et al. (2013) Association of GWAS top hits with late-onset Alzheimer disease in Korean population. *Alzheimer Dis Assoc Disord* 27: 250–257.
102. Liu G, Zhang S, Cai Z, et al. (2013) PICALM gene rs3851179 polymorphism contributes to Alzheimer's disease in an Asian population. *Neuromolecular Med* 15: 384–388.
103. Wang Q, Tian Q, Song X, et al. (2016) SNCA gene polymorphism may contribute to an increased risk of Alzheimer's disease. *J Clin Lab Anal* 30: 1092–1099.
104. Linnertz C, Lutz MW, Ervin JF, et al. (2014) The genetic contributions of SNCA and LRRK2 genes to Lewy body pathology in Alzheimer's disease. *Hum Mol Genet* 23: 4814–4821.

105. Vardarajan BN, Zhang Y, Lee JH, et al. (2015) Coding mutations in SORL1 and Alzheimer disease. *Ann Neurol* 77: 215–227.
106. Yin RH, Yu JT, Tan L (2015) The role of SORL1 in Alzheimer's disease. *Mol Neurobiol* 51: 909–918.
107. Rogaeva E, Meng Y, Lee JH, et al. (2007) The neuronal sortilin-related receptor SORL1 is genetically associated with Alzheimer disease. *Nat Genet* 39: 168–177.
108. Lambert JC, Ibrahim-Verbaas CA, Harold D, et al. (2013) Meta-analysis of 74,046 individuals identifies 11 new susceptibility loci for Alzheimer's disease. *Nat Genet* 45: 1452–1458.
109. Allen M, Kachadorian M, Quicksall Z, et al. (2014) Association of MAPT haplotypes with Alzheimer's disease risk and MAPT brain gene expression levels. *Alzheimers Res Ther* 6: 39.
110. Zhang N, Yu JT, Yang Y, et al. (2011) Association analysis of GSK3B and MAPT polymorphisms with Alzheimer's disease in Han Chinese. *Brain Res* 1391: 147–153.
111. Bloom GS (2014) Amyloid-beta and tau: the trigger and bullet in Alzheimer disease pathogenesis. *JAMA Neurol* 71: 505–508.
112. James BD, Wilson RS, Boyle PA, et al. (2016) TDP-43 stage, mixed pathologies, and clinical Alzheimer's-type dementia. *Brain* 139: 2983–2993.
113. Brouwers N, Bettens K, Gijselink I, et al. (2010) Contribution of TARDBP to Alzheimer's disease genetic etiology. *J Alzheimers Dis* 21: 423–430.
114. Guerreiro R, Wojtas A, Bras J, et al. (2013) TREM2 variants in Alzheimer's disease. *N Engl J Med* 368: 117–127.
115. Lu Y, Liu W, Wang X (2015) TREM2 variants and risk of Alzheimer's disease: a meta-analysis. *Neurol Sci* 36: 1881–1888.
116. Naj AC, Jun G, Beecham GW, et al. (2011) Common variants at MS4A4/MS4A6E, CD2AP, CD33 and EPHA1 are associated with late-onset Alzheimer's disease. *Nat Genet* 43: 436–441.
117. Wijsman EM, Pankratz ND, Choi Y, et al. (2011) Genome-wide association of familial late-onset Alzheimer's disease replicates BIN1 and CLU and nominates CUGBP2 in interaction with APOE. *PLoS Genet* 7: e1001308.
118. Boada M, Antunez C, Ramirez-Lorca R, et al. (2014) ATP5H/KCTD2 locus is associated with Alzheimer's disease risk. *Mol Psychiatry* 19: 682–687.
119. Beecham GW, Naj AC, Gilbert JR, et al. (2010) PCDH11X variation is not associated with late-onset Alzheimer disease susceptibility. *Psychiatr Genet* 20: 321–324.
120. Miar A, Alvarez V, Corao AI, et al. (2011) Lack of association between protocadherin 11-X/Y (PCDH11X and PCDH11Y) polymorphisms and late onset Alzheimer's disease. *Brain Res* 1383: 252–256.
121. Jiang T, Yu JT, Wang YL, et al. (2014) The genetic variation of ARRB2 is associated with late-onset Alzheimer's disease in Han Chinese. *Curr Alzheimer Res* 11: 408–412.
122. Gorski DH, Walsh K (2003) Control of vascular cell differentiation by homeobox transcription factors. *Trends Cardiovasc Med* 13: 213–220.
123. Rovelet-Lecrux A, Legallic S, Wallon D, et al. (2012) A genome-wide study reveals rare CNVs exclusive to extreme phenotypes of Alzheimer disease. *Eur J Hum Genet* 20: 613–617.
124. Wu Z, Guo H, Chow N, et al. (2005) Role of the MEOX2 homeobox gene in neurovascular dysfunction in Alzheimer disease. *Nat Med* 11: 959–965.
125. Mielke MM, Vemuri P, Rocca WA. (2014) Clinical epidemiology of Alzheimer's disease: assessing sex and gender differences. *Clin Epidemiol* 6: 37–48.

126. Fjell AM, McEvoy L, Holland D, et al. (2014) What is normal in normal aging? Effects of aging, amyloid and Alzheimer's disease on the cerebral cortex and the hippocampus. *Prog Neurobiol* 117: 20–40.
127. Lee JY, Chang SM, Jang HS, et al. (2008) Illiteracy and the incidence of Alzheimer's disease in the Yonchon County survey, Korea. *Int Psychogeriatr* 20: 976–985.
128. Durazzo TC, Mattsson N, Weiner MW, et al. (2014) Smoking and increased Alzheimer's disease risk: a review of potential mechanisms. *Alzheimers Dement* 10: S122–145.
129. Barnard ND, Bush AI, Ceccarelli A, et al. (2014) Dietary and lifestyle guidelines for the prevention of Alzheimer's disease. *Neurobiol Aging* 35: S74–78.
130. Kawahara M, Kato-Negishi M, Tanaka K (2017) Cross talk between neurometals and amyloidogenic proteins at the synapse and the pathogenesis of neurodegenerative diseases. *Metallomics* 9: 619–633.
131. Zheng W, Aschner M, Ghersi-Egea JF (2003) Brain barrier systems: a new frontier in metal neurotoxicological research. *Toxicol Appl Pharmacol* 192: 1–11.
132. Heneka MT, Carson MJ, El Khoury J, et al. (2015) Neuroinflammation in Alzheimer's disease. *Lancet Neurol* 14: 388–405.
133. Wang X, Wang W, Li L, et al. (2014) Oxidative stress and mitochondrial dysfunction in Alzheimer's disease. *Biochim Biophys Acta* 1842: 1240–1247.
134. Chen Z, Zhong C (2014) Oxidative stress in Alzheimer's disease. *Neurosci Bull* 30: 271–281.
135. DuBoff B, Feany M, Gotz J (2013) Why size matters-balancing mitochondrial dynamics in Alzheimer's disease. *Trends Neurosci* 36: 325–335.
136. Dik MG, Jonker C, Comijs HC, et al. (2007) Contribution of metabolic syndrome components to cognition in older individuals. *Diabetes Care* 30: 2655–2660.
137. Campos-Pena V, Toral-Rios D, Becerril-Perez F, et al. (2017) Metabolic syndrome as a risk factor for Alzheimer's disease: is abeta a crucial factor in both pathologies? *Antioxid Redox Signal* 26: 542–560.
138. Whyte LS, Lau AA, Hemsley KM, et al. (2017) Endo-lysosomal and autophagic dysfunction: a driving factor in Alzheimer's disease? *J Neurochem* 140: 703–717.
139. Ferreira ST, Clarke JR, Bomfim TR, et al. (2014) Inflammation, defective insulin signaling, and neuronal dysfunction in Alzheimer's disease. *Alzheimers Dement* 10: S76–83.
140. Kinney JW, Bemiller SM, Murtishaw AS, et al. (2018) Inflammation as a central mechanism in Alzheimer's disease. *Alzheimers Dement* 4: 575–590.
141. Rubio-Perez JM, Morillas-Ruiz JM (2012) A review: inflammatory process in Alzheimer's disease, role of cytokines. *Sci World J* 2012: 756357.
142. Mengel-From J, Christensen K, McGue M, et al. (2011) Genetic variations in the CLU and PICALM genes are associated with cognitive function in the oldest old. *Neurobiol Aging* 32: 554. e7–11.
143. Jonsson T, Stefansson H, Steinberg S, et al. (2013) Variant of TREM2 associated with the risk of Alzheimer's disease. *N Engl J Med* 368: 107–116.
144. Cunningham C (2013) Microglia and neurodegeneration: the role of systemic inflammation. *Glia* 61: 71–90.
145. Bodea LG, Wang Y, Linnartz-Gerlach B, et al. (2014) Neurodegeneration by activation of the microglial complement-phagosome pathway. *J Neurosci* 34: 8546–8556.

146. Mhatre SD, Tsai CA, Rubin AJ, et al. (2015) Microglial malfunction: the third rail in the development of Alzheimer's disease. *Trends Neurosci* 38: 621–636.
147. Zotova E, Nicoll JA, Kalara R, et al. (2010) Inflammation in Alzheimer's disease: relevance to pathogenesis and therapy. *Alzheimers Res Ther* 2: 1.
148. Musicco C, Capelli V, Pesce V, et al. (2009) Accumulation of overoxidized peroxiredoxin III in aged rat liver mitochondria. *Biochim Biophys Acta* 1787: 890–896.
149. Sutinen EM, Pirttila T, Anderson G, et al. (2012) Pro-inflammatory interleukin-18 increases Alzheimer's disease-associated amyloid-beta production in human neuron-like cells. *J Neuroinflammation* 9: 199.
150. Sutinen EM, Korolainen MA, Hayrinen J, et al. (2014) Interleukin-18 alters protein expressions of neurodegenerative diseases-linked proteins in human SH-SY5Y neuron-like cells. *Front Cell Neurosci* 8: 214.
151. Oakley R, Tharakan B (2014) Vascular hyperpermeability and aging. *Aging Dis* 5: 114–125.
152. McColl BW, Rose N, Robson FH, et al. (2010) Increased brain microvascular MMP-9 and incidence of haemorrhagic transformation in obese mice after experimental stroke. *J Cereb Blood Flow Metab* 30: 267–272.
153. Ojala JO, Sutinen EM, Salminen A, et al. (2008) Interleukin-18 increases expression of kinases involved in tau phosphorylation in SH-SY5Y neuroblastoma cells. *J Neuroimmunol* 205: 86–93.
154. Alvarez A, Toro R, Caceres A, et al. (1999) Inhibition of tau phosphorylating protein kinase cdk5 prevents beta-amyloid-induced neuronal death. *FEBS Lett* 459: 421–426.
155. Alvarez A, Munoz JP, Maccioni RB (2001) A Cdk5-p35 stable complex is involved in the beta-amyloid-induced deregulation of Cdk5 activity in hippocampal neurons. *Exp Cell Res* 264: 266–274.
156. Seo J, Kritskiy O, Watson LA, et al. (2017) Inhibition of p25/Cdk5 attenuates tauopathy in mouse and iPSC models of frontotemporal dementia. *J Neurosci* 37: 9917–9924.
157. Morgan BP (2018) Complement in the pathogenesis of Alzheimer's disease. *Semin Immunopathol* 40: 113–124.
158. Bonham LW, Desikan RS, Yokoyama JS, et al. (2016) The relationship between complement factor C3, APOE epsilon4, amyloid and tau in Alzheimer's disease. *Acta Neuropathol Commun* 4: 65.
159. Bonham LW, Geier EG, Fan CC, et al. (2016) Age-dependent effects of APOE epsilon4 in preclinical Alzheimer's disease. *Ann Clin Transl Neurol* 3: 668–677.
160. Fritzing DC, Benjamin DE (2016) The complement system in neuropathic and postoperative pain. *Open Pain J* 9: 26–37.
161. Sheedy FJ, Grebe A, Rayner KJ, et al. (2013) CD36 coordinates NLRP3 inflammasome activation by facilitating intracellular nucleation of soluble ligands into particulate ligands in sterile inflammation. *Nat Immunol* 14: 812–820.
162. Devanand DP (2018) Viral hypothesis and antiviral treatment in Alzheimer's disease. *Curr Neurol Neurosci Rep* 18: 55.
163. Duggan MR, Torkzaban B, Ahooyi TM, et al. (2020) Potential role for herpesviruses in Alzheimer's disease. *J Alzheimers Dis*, 1–15 (Preprint).
164. Wang JH, Cheng XR, Zhang XR, et al. (2016) Neuroendocrine immunomodulation network dysfunction in SAMP8 mice and PrP-hAbetaPPswe/PS1DeltaE9 mice: potential mechanism underlying cognitive impairment. *Oncotarget* 7: 22988–3005.

165. Lopategui Cabezas I, Herrera Batista A, Penton Rol G (2014) The role of glial cells in Alzheimer disease: potential therapeutic implications. *Neurologia* 29: 305–309.
166. Birch AM (2014) The contribution of astrocytes to Alzheimer's disease. *Biochem Soc Trans* 42: 1316–1320.
167. Takata K, Kitamura Y, Saeki M, et al. (2010) Galantamine-induced amyloid- β clearance mediated via stimulation of microglial nicotinic acetylcholine receptors. *J Biol Chem* 285: 40180–40191.
168. Koenigsnecht-Talboo J, Landreth GE (2005) Microglial phagocytosis induced by fibrillar beta-amyloid and IgGs are differentially regulated by proinflammatory cytokines. *J Neurosci* 25: 8240–8249.
169. Xie J, Wang H, Lin T, et al. (2017) Microglia-Synapse pathways: promising therapeutic strategy for Alzheimer's disease. *Biomed Res Int* 2017: 2986460.
170. Qi Y, Klyubin I, Harney SC, et al. (2014) Longitudinal testing of hippocampal plasticity reveals the onset and maintenance of endogenous human A β -induced synaptic dysfunction in individual freely behaving pre-plaque transgenic rats: rapid reversal by anti-A β agents. *Acta Neuropathol Commun* 2: 175.
171. Lo AC, Iscru E, Blum D, et al. (2013) Amyloid and tau neuropathology differentially affect prefrontal synaptic plasticity and cognitive performance in mouse models of Alzheimer's disease. *J Alzheimers Dis* 37: 109–125.
172. Giulian D, Baker TJ (1986) Characterization of amoeboid microglia isolated from developing mammalian brain. *J Neurosci* 6: 2163–2178.
173. Kreutzberg GW (1996) Microglia: a sensor for pathological events in the CNS. *Trends Neurosci* 19: 312–318.
174. Mecha M, Carrillo-Salinas FJ, Feliu A, et al. (2016) Microglia activation states and cannabinoid system: therapeutic implications. *Pharmacol Ther* 166: 40–55.
175. Henkel JS, Beers DR, Zhao W, et al. (2009) Microglia in ALS: the good, the bad, and the resting. *J Neuroimmune Pharmacol* 4: 389–398.
176. Stansley B, Post J, Hensley K (2012) A comparative review of cell culture systems for the study of microglial biology in Alzheimer's disease. *J Neuroinflammation* 9: 115.
177. Acosta C, Anderson HD, Anderson CM (2017) Astrocyte dysfunction in Alzheimer disease. *J Neurosci Res* 95: 2430–2447.
178. Sofroniew MV (2009) Molecular dissection of reactive astrogliosis and glial scar formation. *Trends Neurosci* 32: 638–647.
179. Ricci G, Volpi L, Pasquali L, et al. (2009) Astrocyte-neuron interactions in neurological disorders. *J Biol Phys* 35: 317–336.
180. Mattsson N, Tosun D, Insel PS, et al. (2014) Association of brain amyloid-beta with cerebral perfusion and structure in Alzheimer's disease and mild cognitive impairment. *Brain* 137: 1550–1561.
181. Huang KL, Lin KJ, Ho MY, et al. (2012) Amyloid deposition after cerebral hypoperfusion: evidenced on [(18)F]AV-45 positron emission tomography. *J Neurol Sci* 319: 124–129.
182. Okamoto Y, Yamamoto T, Kalaria RN, et al. (2012) Cerebral hypoperfusion accelerates cerebral amyloid angiopathy and promotes cortical microinfarcts. *Acta Neuropathol* 123: 381–394.
183. Pietronigro EC, Della Bianca V, Zenaro E, et al. (2017) NETosis in Alzheimer's disease. *Front Immunol* 8: 211.

184. Mantovani A, Cassatella MA, Costantini C, et al. (2011) Neutrophils in the activation and regulation of innate and adaptive immunity. *Nat Rev Immunol* 11: 519–531.
185. Tillack K, Breiden P, Martin R, et al. (2012) T lymphocyte priming by neutrophil extracellular traps links innate and adaptive immune responses. *J Immunol* 188: 3150–3159.
186. Llanos-Gonzalez E, Henares-Chavarino AA, Pedrero-Prieto CM, et al. (2019) Interplay between mitochondrial oxidative disorders and proteostasis in Alzheimer's disease. *Front Neurosci* 13: 1444.
187. Franco R, Vargas MR (2018) Redox biology in neurological function, dysfunction, and aging. *Antioxid Redox Signal* 28: 1583–1586.
188. Miller VM, Lawrence DA, Mondal TK, et al. (2009) Reduced glutathione is highly expressed in white matter and neurons in the unperturbed mouse brain—implications for oxidative stress associated with neurodegeneration. *Brain Res* 1276: 22–30.
189. Munoz P, Ardiles AO, Perez-Espinosa B, et al. (2020) Redox modifications in synaptic components as biomarkers of cognitive status, in brain aging and disease. *Mech Ageing Dev* 189: 111250.
190. Kumar A, Yegla B, Foster TC (2018) Redox signaling in neurotransmission and cognition during aging. *Antioxid Redox Signal* 28: 1724–1745.
191. Sbodio JI, Snyder SH, Paul BD (2019) Redox mechanisms in neurodegeneration: from disease outcomes to therapeutic opportunities. *Antioxid Redox Signal* 30: 1450–1499.
192. Arias-Cavieres A, Adasme T, Sanchez G, et al. (2017) Aging impairs hippocampal-dependent recognition memory and LTP and prevents the associated RyR Up-regulation. *Front Aging Neurosci* 9: 111.
193. Wilson C, Gonzalez-Billault C (2015) Regulation of cytoskeletal dynamics by redox signaling and oxidative stress: implications for neuronal development and trafficking. *Front Cell Neurosci* 9: 381.
194. Quintanilla RA, Orellana JA, von Bernhardi R (2012) Understanding risk factors for Alzheimer's disease: interplay of neuroinflammation, connexin-based communication and oxidative stress. *Arch Med Res* 43: 632–644.
195. Butterfield DA, Bader Lange ML, Sultana R (2010) Involvements of the lipid peroxidation product, HNE, in the pathogenesis and progression of Alzheimer's disease. *Biochim Biophys Acta* 1801: 924–929.
196. Chen YY, Yu XY, Chen L, et al. (2019) Redox signaling in aging kidney and opportunity for therapeutic intervention through natural products. *Free Radic Biol Med* 141: 141–149.
197. Bruce-Keller AJ, Gupta S, Knight AG, et al. (2011) Cognitive impairment in humanized APPxPS1 mice is linked to Abeta(1-42) and NOX activation. *Neurobiol Dis* 44: 317–326.
198. Kothari V, Luo Y, Tornabene T, et al. (2017) High fat diet induces brain insulin resistance and cognitive impairment in mice. *Biochim Biophys Acta* 1863: 499–508.
199. Wilkinson BL, Landreth GE (2006) The microglial NADPH oxidase complex as a source of oxidative stress in Alzheimer's disease. *J Neuroinflammation* 3: 30.
200. Wong KY, Roy J, Fung ML, et al. (2020) Relationships between mitochondrial dysfunction and neurotransmission failure in Alzheimer's disease. *Aging Dis* 11: 1291–1316.
201. Bissette G, Seidler FJ, Nemeroff CB, et al. (1996) High affinity choline transporter status in Alzheimer's disease tissue from rapid autopsy. *Ann N Y Acad Sci* 777: 197–204.

202. Campanucci VA, Krishnaswamy A, Cooper E (2008) Mitochondrial reactive oxygen species inactivate neuronal nicotinic acetylcholine receptors and induce long-term depression of fast nicotinic synaptic transmission. *J Neurosci* 28: 1733–1744.
203. Jomova K, Vondrakova D, Lawson M, et al. (2010) Metals, oxidative stress and neurodegenerative disorders. *Mol Cell Biochem* 345: 91–104.
204. Swerdlow RH, Khan SM (2004) A “mitochondrial cascade hypothesis” for sporadic Alzheimer’s disease. *Med Hypotheses* 63: 8–20.
205. Kim GW, Gasche Y, Grzeschik S, et al. (2003) Neurodegeneration in striatum induced by the mitochondrial toxin 3-nitropropionic acid: role of matrix metalloproteinase-9 in early blood-brain barrier disruption? *J Neurosci* 23: 8733–8742.
206. Ridnour LA, Dhanapal S, Hoos M, et al. (2012) Nitric oxide-mediated regulation of beta-amyloid clearance via alterations of MMP-9/TIMP-1. *J Neurochem* 123: 736–749.
207. Liu Y, Liu F, Iqbal K, et al. (2008) Decreased glucose transporters correlate to abnormal hyperphosphorylation of tau in Alzheimer disease. *FEBS Lett* 582: 359–364.
208. Nikinmaa M, Pursiheimo S, Soitamo AJ (2004) Redox state regulates HIF-1alpha and its DNA binding and phosphorylation in salmonid cells. *J Cell Sci* 117: 3201–3206.
209. Morris G, Walder K, Puri BK, et al. (2016) The deleterious effects of oxidative and nitrosative stress on palmitoylation, membrane lipid rafts and lipid-based cellular signalling: new drug targets in neuroimmune disorders. *Mol Neurobiol* 53: 4638–4658.
210. Zou Y, Watters A, Cheng N, et al. (2019) Polyunsaturated fatty acids from astrocytes activate PPARgamma signaling in cancer cells to promote brain metastasis. *Cancer Discov* 9: 1720–1735.
211. Marnett LJ (1999) Lipid peroxidation-DNA damage by malondialdehyde. *Mutat Res* 424: 83–95.
212. Lu Y, Ren J, Cui S, et al. (2016) Cerebral glucose metabolism assessment in rat models of Alzheimer’s disease: an 18F-FDG-PET study. *Am J Alzheimers Dis Other Demen* 31: 333–340.
213. Jeong DU, Oh JH, Lee JE, et al. (2016) Basal forebrain cholinergic deficits reduce glucose metabolism and function of cholinergic and GABAergic systems in the cingulate cortex. *Yonsei Med J* 57: 165–172.
214. Braak H, Del Tredici K (2015) The preclinical phase of the pathological process underlying sporadic Alzheimer’s disease. *Brain* 138: 2814–2833.
215. Morrison BM, Lee Y, Rothstein JD (2013) Oligodendroglia: metabolic supporters of axons. *Trends Cell Biol* 23: 644–651.
216. Grant CM, Quinn KA, Dawes IW (1999) Differential protein S-thiolation of glyceraldehyde-3-phosphate dehydrogenase isoenzymes influences sensitivity to oxidative stress. *Mol Cell Biol* 19: 2650–2656.
217. Avery SV (2011) Molecular targets of oxidative stress. *Biochem J* 434: 201–210.
218. Bubber P, Haroutunian V, Fisch G, et al. (2005) Mitochondrial abnormalities in Alzheimer brain: mechanistic implications. *Ann Neurol* 57: 695–703.
219. Gibson GE, Blass JP, Beal MF, et al. (2005) The alpha-ketoglutarate-dehydrogenase complex: a mediator between mitochondria and oxidative stress in neurodegeneration. *Mol Neurobiol* 31: 43–63.

220. Long J, Liu C, Sun L, et al. (2009) Neuronal mitochondrial toxicity of malondialdehyde: inhibitory effects on respiratory function and enzyme activities in rat brain mitochondria. *Neurochem Res* 34: 786–794.
221. Martin E, Rosenthal RE, Fiskum G (2005) Pyruvate dehydrogenase complex: metabolic link to ischemic brain injury and target of oxidative stress. *J Neurosci Res* 79: 240–247.
222. Salminen A, Haapasalo A, Kauppinen A, et al. (2015) Impaired mitochondrial energy metabolism in Alzheimer's disease: impact on pathogenesis via disturbed epigenetic regulation of chromatin landscape. *Prog Neurobiol* 131: 1–20.
223. Liguori C, Chiaravalloti A, Sancesario G, et al. (2016) Cerebrospinal fluid lactate levels and brain [18F]FDG PET hypometabolism within the default mode network in Alzheimer's disease. *Eur J Nucl Med Mol Imaging* 43: 2040–2049.
224. Isopi E, Granzotto A, Corona C, et al. (2015) Pyruvate prevents the development of age-dependent cognitive deficits in a mouse model of Alzheimer's disease without reducing amyloid and tau pathology. *Neurobiol Dis* 81: 214–224.
225. Talbot K, Wang HY, Kazi H, et al. (2012) Demonstrated brain insulin resistance in Alzheimer's disease patients is associated with IGF-1 resistance, IRS-1 dysregulation, and cognitive decline. *J Clin Invest* 122: 1316–1338.
226. Morales-Corraliza J, Wong H, Mazzella MJ, et al. (2016) Brain-Wide insulin resistance, Tau phosphorylation changes, and hippocampal neprilysin and Amyloid-beta alterations in a monkey model of Type 1 diabetes. *J Neurosci* 36: 4248–4258.
227. Yamamoto N, Ishikuro R, Tanida M, et al. (2018) Insulin-signaling pathway regulates the degradation of amyloid beta-protein via astrocytes. *Neuroscience* 385: 227–236.
228. Han X, Yang L, Du H, et al. (2016) Insulin attenuates beta-amyloid-associated Insulin/Akt/EAAT signaling perturbations in human astrocytes. *Cell Mol Neurobiol* 36: 851–864.
229. Ng RC, Chan KH (2017) Potential neuroprotective effects of adiponectin in Alzheimer's disease. *Int J Mol Sci* 18: 592.
230. Pei JJ, Khatoon S, An WL, et al. (2003) Role of protein kinase B in Alzheimer's neurofibrillary pathology. *Acta Neuropathol* 105: 381–392.
231. Anderson NJ, King MR, Delbruck L, et al. (2014) Role of insulin signaling impairment, adiponectin and dyslipidemia in peripheral and central neuropathy in mice. *Dis Model Mech* 7: 625–633.
232. Garcia-Casares N, Jorge RE, Garcia-Arnes JA, et al. (2014) Cognitive dysfunctions in middle-aged type 2 diabetic patients and neuroimaging correlations: a cross-sectional study. *J Alzheimers Dis* 42: 1337–1346.
233. Purnell C, Gao S, Callahan CM, et al. (2009) Cardiovascular risk factors and incident Alzheimer disease: a systematic review of the literature. *Alzheimer Dis Assoc Disord* 23: 1–10.
234. de la Torre JC (2012) Cardiovascular risk factors promote brain hypoperfusion leading to cognitive decline and dementia. *Cardiovasc Psychiatry Neurol* 2012: 367516.
235. Klohs J (2019) An integrated view on vascular dysfunction in Alzheimer's disease. *Neurodegener Dis* 19: 109–127.
236. Popovic M, Laumonier Y, Burysek L, et al. (2008) Thrombin-induced expression of endothelial CX3CL1 potentiates monocyte CCL2 production and transendothelial migration. *J Leukoc Biol* 84: 215–223.

237. Sole M, Esteban-Lopez M, Taltavull B, et al. (2019) Blood-brain barrier dysfunction underlying Alzheimer's disease is induced by an SSAO/VAP-1-dependent cerebrovascular activation with enhanced Aβ deposition. *Biochim Biophys Acta Mol Basis Dis* 1865: 2189–2202.
238. Zlokovic BV (2011) Neurovascular pathways to neurodegeneration in Alzheimer's disease and other disorders. *Nat Rev Neurosci* 12: 723–738.
239. Ramanathan A, Nelson AR, Sagare AP, et al. (2015) Impaired vascular-mediated clearance of brain amyloid β in Alzheimer's disease: the role, regulation and restoration of LRP1. *Front Aging Neurosci* 7: 136.
240. Montagne A, Pa J, Zlokovic BV (2015) Vascular plasticity and cognition during normal aging and dementia. *JAMA Neurol* 72: 495–496.
241. Montine TJ, Koroshetz WJ, Babcock D, et al. (2014) Recommendations of the Alzheimer's disease-related dementias conference. *Neurology* 83: 851–860.
242. Sweeney MD, Sagare AP, Zlokovic BV (2015) Cerebrospinal fluid biomarkers of neurovascular dysfunction in mild dementia and Alzheimer's disease. *J Cereb Blood Flow Metab* 35: 1055–1068.
243. Yan SD, Chen X, Fu J, et al. (1996) RAGE and amyloid-β peptide neurotoxicity in Alzheimer's disease. *Nature* 382: 685–691.
244. Miller MC, Tavares R, Johanson CE, et al. (2008) Hippocampal RAGE immunoreactivity in early and advanced Alzheimer's disease. *Brain Res* 1230: 273–280.
245. Lue LF, Walker DG, Brachova L, et al. (2001) Involvement of microglial receptor for advanced glycation endproducts (RAGE) in Alzheimer's disease: identification of a cellular activation mechanism. *Exp Neurol* 171: 29–45.
246. Carnevale D, Mascio G, D'Andrea I, et al. (2012) Hypertension induces brain β-amyloid accumulation, cognitive impairment, and memory deterioration through activation of receptor for advanced glycation end products in brain vasculature. *Hypertension* 60: 188–197.
247. Srikanth V, Maczurek A, Phan T, et al. (2011) Advanced glycation endproducts and their receptor RAGE in Alzheimer's disease. *Neurobiol Aging* 32: 763–777.
248. de la Torre J (2018) The vascular hypothesis of Alzheimer's disease: a key to preclinical prediction of dementia using neuroimaging. *J Alzheimers Dis* 63: 35–52.
249. Wierenga CE, Hays CC, Zlatar ZZ (2014) Cerebral blood flow measured by arterial spin labeling MRI as a preclinical marker of Alzheimer's disease. *J Alzheimers Dis* 42: S411–419.
250. Chuang YF, Breitner JCS, Chiu YL, et al. (2014) Use of diuretics is associated with reduced risk of Alzheimer's disease: the Cache County study. *Neurobiol Aging* 35: 2429–2435.
251. Ashby EL, Kehoe PG (2013) Current status of renin-aldosterone angiotensin system-targeting anti-hypertensive drugs as therapeutic options for Alzheimer's disease. *Expert Opin Investig Drugs* 22: 1229–1242.
252. Yasar S, Xia J, Yao W, et al. (2013) Antihypertensive drugs decrease risk of Alzheimer disease: ginkgo evaluation of memory study. *Neurology* 81: 896–903.
253. Thomas T, Miners S, Love S (2015) Post-mortem assessment of hypoperfusion of cerebral cortex in Alzheimer's disease and vascular dementia. *Brain* 138: 1059–1069.
254. Zhao Y, Gong CX (2015) From chronic cerebral hypoperfusion to Alzheimer-like brain pathology and neurodegeneration. *Cell Mol Neurobiol* 35: 101–110.
255. Glodzik L, Rusinek H, Pirraglia E, et al. (2014) Blood pressure decrease correlates with tau pathology and memory decline in hypertensive elderly. *Neurobiol Aging* 35: 64–71.

256. Hong S, Beja-Glasser VF, Nfonoyim BM, et al. (2016) Complement and microglia mediate early synapse loss in Alzheimer mouse models. *Science* 352: 712–716.
257. Gottfries CG, Bartfai T, Carlsson A, et al. (1986) Multiple biochemical deficits in both gray and white matter of Alzheimer brains. *Prog Neuropsychopharmacol Biol Psychiatry* 10: 405–413.
258. Storga D, Vrecko K, Birkmayer JG, et al. (1996) Monoaminergic neurotransmitters, their precursors and metabolites in brains of Alzheimer patients. *Neurosci Lett* 203: 29–32.
259. Arai H, Ichimiya Y, Kosaka K, et al. (1992) Neurotransmitter changes in early- and late-onset Alzheimer-type dementia. *Prog Neuropsychopharmacol Biol Psychiatry* 16: 883–890.
260. Lanari A, Amenta F, Silvestrelli G, et al. (2006) Neurotransmitter deficits in behavioural and psychological symptoms of Alzheimer's disease. *Mech Ageing Dev* 127: 158–165.
261. Nobili A, Latagliata EC, Viscomi MT, et al. (2017) Dopamine neuronal loss contributes to memory and reward dysfunction in a model of Alzheimer's disease. *Nat Commun* 8: 14727.
262. Chalermphanupap T, Kinkead B, Hu WT, et al. (2013) Targeting norepinephrine in mild cognitive impairment and Alzheimer's disease. *Alzheimers Res Ther* 5: 21.
263. Woolf NJ, Butcher LL (2011) Cholinergic systems mediate action from movement to higher consciousness. *Behav Brain Res* 221: 488–498.
264. Baker-Nigh A, Vahedi S, Davis EG, et al. (2015) Neuronal amyloid-beta accumulation within cholinergic basal forebrain in ageing and Alzheimer's disease. *Brain* 138: 1722–1737.
265. Bracco L, Bessi V, Padiglioni S, et al. (2014) Do cholinesterase inhibitors act primarily on attention deficit? A naturalistic study in Alzheimer's disease patients. *J Alzheimers Dis* 40: 737–742.
266. Stephan AH, Barres BA, Stevens B (2012) The complement system: an unexpected role in synaptic pruning during development and disease. *Annu Rev Neurosci* 35: 369–389.
267. Bialas AR, Stevens B (2013) TGF-beta signaling regulates neuronal C1q expression and developmental synaptic refinement. *Nat Neurosci* 16: 1773–1782.
268. Nakajima K, Tohyama Y, Maeda S, et al. (2007) Neuronal regulation by which microglia enhance the production of neurotrophic factors for GABAergic, catecholaminergic, and cholinergic neurons. *Neurochem Int* 50: 807–820.
269. Sierra A, Gottfried-Blackmore AC, McEwen BS, et al. (2007) Microglia derived from aging mice exhibit an altered inflammatory profile. *Glia* 55: 412–424.
270. Welser-Alves JV, Milner R (2013) Microglia are the major source of TNF-alpha and TGF-beta1 in postnatal glial cultures; regulation by cytokines, lipopolysaccharide, and vitronectin. *Neurochem Int* 63: 47–53.
271. Brucato FH, Benjamin DE (2020) Synaptic pruning in Alzheimer's disease: role of the complement system. *Glob J Med Res* 20.
272. Gajardo I, Salazar CS, Lopez-Espindola D, et al. (2018) Lack of pannexin 1 alters synaptic GluN2 subunit composition and spatial reversal learning in mice. *Front Mol Neurosci* 11: 114.
273. Flores-Munoz C, Gomez B, Mery E, et al. (2020) Acute pannexin 1 blockade mitigates early synaptic plasticity defects in a mouse model of Alzheimer's disease. *Front Cell Neurosci* 14: 46.
274. Liu J, Chang L, Song Y, et al. (2019) The role of NMDA receptors in Alzheimer's disease. *Front Neurosci* 13: 43.
275. Hidalgo C, Arias-Cavieres A (2016) Calcium, reactive oxygen species, and synaptic plasticity. *Physiology* 31: 201–215.

276. Lu YF, Hawkins RD (2002) Ryanodine receptors contribute to cGMP-induced late-phase LTP and CREB phosphorylation in the hippocampus. *J Neurophysiol* 88: 1270–1278.
277. Del Prete D, Checler F, Chami M (2014) Ryanodine receptors: physiological function and deregulation in Alzheimer disease. *Mol Neurodegener* 9: 21.
278. Oules B, Del Prete D, Greco B, et al. (2012) Ryanodine receptor blockade reduces amyloid-beta load and memory impairments in Tg2576 mouse model of Alzheimer disease. *J Neurosci* 32: 11820–11834.
279. SanMartin CD, Veloso P, Adasme T, et al. (2017) RyR2-Mediated Ca(2+) release and mitochondrial ROS generation partake in the synaptic dysfunction caused by amyloid beta peptide oligomers. *Front Mol Neurosci* 10: 115.
280. Munoz Y, Paula-Lima AC, Nunez MT (2018) Reactive oxygen species released from astrocytes treated with amyloid beta oligomers elicit neuronal calcium signals that decrease phospho-Ser727-STAT3 nuclear content. *Free Radic Biol Med* 117: 132–144.
281. Bitzer-Quintero OK, Gonzalez-Burgos I (2012) Immune system in the brain: a modulatory role on dendritic spine morphophysiology? *Neural Plast* 2012: 348642.
282. Fonseca MI, Chu SH, Hernandez MX, et al. (2017) Cell-specific deletion of C1qa identifies microglia as the dominant source of C1q in mouse brain. *J Neuroinflammation* 14: 48.
283. Schafer DP, Lehrman EK, Kautzman AG, et al. (2012) Microglia sculpt postnatal neural circuits in an activity and complement-dependent manner. *Neuron* 74: 691–705.
284. Gasque P (2004) Complement: a unique innate immune sensor for danger signals. *Mol Immunol* 41: 1089–1098.
285. Azevedo EP, Ledo JH, Barbosa G, et al. (2013) Activated microglia mediate synapse loss and short-term memory deficits in a mouse model of transthyretin-related oculoleptomeningeal amyloidosis. *Cell Death Dis* 4: e789.
286. Wang WY, Tan MS, Yu JT, et al. (2015) Role of pro-inflammatory cytokines released from microglia in Alzheimer's disease. *Ann Transl Med* 3: 136.
287. Liddel SA, Guttenplan KA, Clarke LE, et al. (2017) Neurotoxic reactive astrocytes are induced by activated microglia. *Nature* 541: 481–487.
288. Stevens B, Allen NJ, Vazquez LE, et al. (2007) The classical complement cascade mediates CNS synapse elimination. *Cell* 131: 1164–1178.
289. Stephan AH, Madison DV, Mateos JM, et al. (2013) A dramatic increase of C1q protein in the CNS during normal aging. *J Neurosci* 33: 13460–13474.
290. Shi Q, Colodner KJ, Matousek SB, et al. (2015) Complement C3-Deficient mice fail to display age-related hippocampal decline. *J Neurosci* 35: 13029–13042.
291. Shi Q, Chowdhury S, Ma R, et al. (2017) Complement C3 deficiency protects against neurodegeneration in aged plaque-rich APP/PS1 mice. *Sci Transl Med* 9: eaaf6295.
292. Reichwald J, Danner S, Wiederhold KH, et al. (2009) Expression of complement system components during aging and amyloid deposition in APP transgenic mice. *J Neuroinflammation* 6: 35.
293. Paolicelli RC, Jawaid A, Henstridge CM, et al. (2017) TDP-43 depletion in microglia promotes amyloid clearance but also induces synapse loss. *Neuron* 95: 297–308. e6.
294. Orellana JA, Shoji KF, Abudara V, et al. (2011) Amyloid beta-induced death in neurons involves glial and neuronal hemichannels. *J Neurosci* 31: 4962–4977.

295. Pena-Oyarzun D, Bravo-Sagua R, Diaz-Vega A, et al. (2018) Autophagy and oxidative stress in non-communicable diseases: a matter of the inflammatory state? *Free Radic Biol Med* 124: 61–78.
296. Mizushima N (2009) Physiological functions of autophagy. *Curr Top Microbiol Immunol* 335: 71–84.
297. Alirezaei M, Kiosses WB, Flynn CT, et al. (2008) Disruption of neuronal autophagy by infected microglia results in neurodegeneration. *PLoS One* 3: e2906.
298. Komatsu M, Waguri S, Chiba T, et al. (2006) Loss of autophagy in the central nervous system causes neurodegeneration in mice. *Nature* 441: 880–884.
299. Ji ZS, Mullendorff K, Cheng IH, et al. (2006) Reactivity of apolipoprotein E4 and amyloid beta peptide: lysosomal stability and neurodegeneration. *J Biol Chem* 281: 2683–2692.
300. Belinson H, Lev D, Masliah E, et al. (2008) Activation of the amyloid cascade in apolipoprotein E4 transgenic mice induces lysosomal activation and neurodegeneration resulting in marked cognitive deficits. *J Neurosci* 28: 4690–4701.
301. Friedman LG, Qureshi YH, Yu WH (2015) Promoting autophagic clearance: viable therapeutic targets in Alzheimer’s disease. *Neurotherapeutics* 12: 94–108.
302. Yu WH, Cuervo AM, Kumar A, et al. (2005) Macroautophagy—a novel Beta-amyloid peptide-generating pathway activated in Alzheimer’s disease. *J Cell Biol* 171: 87–98.
303. Nixon RA (2007) Autophagy, amyloidogenesis and Alzheimer disease. *J Cell Sci* 120: 4081–4091.
304. Klionsky DJ, Elazar Z, Seglen PO, et al. (2008) Does bafilomycin A1 block the fusion of autophagosomes with lysosomes? *Autophagy* 4: 849–850.
305. Nixon RA, Wegiel J, Kumar A, et al. (2005) Extensive involvement of autophagy in Alzheimer disease: an immuno-electron microscopy study. *J Neuropathol Exp Neurol* 64: 113–122.
306. Funderburk SF, Marcellino BK, Yue Z (2010) Cell “self-eating” (autophagy) mechanism in Alzheimer’s disease. *Mt Sinai J Med* 77: 59–68.
307. Silva DF, Esteves AR, Oliveira CR, et al. (2011) Mitochondria: the common upstream driver of amyloid-beta and tau pathology in Alzheimer’s disease. *Curr Alzheimer Res* 8: 563–572.
308. Yu WH, Kumar A, Peterhoff C, et al. (2004) Autophagic vacuoles are enriched in amyloid precursor protein-secretase activities: implications for beta-amyloid peptide over-production and localization in Alzheimer’s disease. *Int J Biochem Cell Biol* 36: 2531–2540.
309. Mizushima N (2005) A(beta) generation in autophagic vacuoles. *J Cell Biol* 171: 15–17.
310. Boland B, Kumar A, Lee S, et al. (2008) Autophagy induction and autophagosome clearance in neurons: relationship to autophagic pathology in Alzheimer’s disease. *J Neurosci* 28: 6926–6937.
311. Lautrup S, Lou G, Aman Y, et al. (2019) Microglial mitophagy mitigates neuroinflammation in Alzheimer’s disease. *Neurochem Int* 129: 104469.
312. Reddy PH, Oliver DM (2019) Amyloid Beta and phosphorylated Tau-Induced defective autophagy and mitophagy in Alzheimer’s disease. *Cells* 8: 488.
313. Kerr JS, Adriaanse BA, Greig NH, et al. (2017) Mitophagy and Alzheimer’s disease: cellular and molecular mechanisms. *Trends Neurosci* 40: 151–166.
314. Hu Y, Li XC, Wang ZH, et al. (2016) Tau accumulation impairs mitophagy via increasing mitochondrial membrane potential and reducing mitochondrial Parkin. *Oncotarget* 7: 17356–17368.

315. Ellisdon AM, Bottomley SP (2004) The role of protein misfolding in the pathogenesis of human diseases. *IUBMB Life* 56: 119–123.
316. Vingtdoux V, Sergeant N, Buee L (2012) Potential contribution of exosomes to the prion-like propagation of lesions in Alzheimer's disease. *Front Physiol* 3: 229.
317. Uddin MS, Tewari D, Sharma G, et al. (2020) Molecular mechanisms of ER stress and UPR in the pathogenesis of Alzheimer's disease. *Mol Neurobiol* 57: 2902–2919.
318. Schiera G, Di Liegro CM, Di Liegro I (2015) Extracellular membrane vesicles as vehicles for brain cell-to-cell interactions in physiological as well as pathological conditions. *Biomed Res Int* 2015: 152926.
319. Christianson JC, Ye Y (2014) Cleaning up in the endoplasmic reticulum: ubiquitin in charge. *Nat Struct Mol Biol* 21: 325–335.
320. Diehl JA, Fuchs SY, Koumenis C (2011) The cell biology of the unfolded protein response. *Gastroenterology* 141: 38–41. e2.
321. Dhakal S, Macreadie I (2020) Protein homeostasis networks and the use of yeast to guide interventions in Alzheimer's disease. *Int J Mol Sci* 21: 8014.
322. Kabir MT, Uddin MS, Zaman S, et al. (2020) Molecular mechanisms of metal toxicity in the pathogenesis of Alzheimer's disease. *Mol Neurobiol*, 1–20.
323. Cristovao JS, Santos R, Gomes CM (2016) Metals and neuronal metal binding proteins implicated in Alzheimer's disease. *Oxid Med Cell Longev* 2016: 9812178.
324. Gerber H, Wu F, Dimitrov M, et al. (2017) Zinc and copper differentially modulate amyloid precursor protein processing by gamma-secretase and amyloid-beta peptide production. *J Biol Chem* 292: 3751–3767.
325. Savellieff MG, Lee S, Liu Y, et al. (2013) Untangling amyloid-beta, tau, and metals in Alzheimer's disease. *ACS Chem Biol* 8: 856–865.
326. Dahms SO, Konnig I, Roeser D, et al. (2012) Metal binding dictates conformation and function of the amyloid precursor protein (APP) E2 domain. *J Mol Biol* 416: 438–452.
327. Flaten TP (2001) Aluminium as a risk factor in Alzheimer's disease, with emphasis on drinking water. *Brain Res Bull* 55: 187–196.
328. Ward RJ, Zucca FA, Duyn JH, et al. (2014) The role of iron in brain ageing and neurodegenerative disorders. *Lancet Neurol* 13: 1045–1060.
329. Lane DJR, Ayton S, Bush AI (2018) Iron and Alzheimer's disease: an update on emerging mechanisms. *J Alzheimers Dis* 64: S379–S395.
330. Tamano H, Takeda A (2011) Dynamic action of neurometals at the synapse. *Metallomics* 3: 656–661.
331. Ashraf A, Clark M, So PW (2018) The aging of iron man. *Front Aging Neurosci* 10: 65.
332. Ojala JO, Sutinen EM (2017) The role of interleukin-18, oxidative stress and metabolic syndrome in Alzheimer's disease. *J Clin Med* 6: 55.
333. Arrigoni F, Rizza F, Tisi R, et al. (2020) On the propagation of the OH radical produced by Cu-amyloid beta peptide model complexes. Insight from molecular modelling. *Metallomics* 12: 1765–1780.
334. Colvin RA, Jin Q, Lai B, et al. (2016) Visualizing metal content and intracellular distribution in primary hippocampal neurons with synchrotron X-ray fluorescence. *PLoS One* 11: e0159582.
335. Acevedo KM, Hung YH, Dalziel AH, et al. (2011) Copper promotes the trafficking of the amyloid precursor protein. *J Biol Chem* 286: 8252–8262.

336. Hickey JL, James JL, Henderson CA, et al. (2015) Intracellular distribution of fluorescent copper and zinc bis(thiosemicarbazonato) complexes measured with fluorescence lifetime spectroscopy. *Inorg Chem* 54: 9556–9567.
337. Alaverdashvili M, Hackett MJ, Pickering IJ, et al. (2014) Laminar-specific distribution of zinc: evidence for presence of layer IV in forelimb motor cortex in the rat. *Neuroimage* 103: 502–510.
338. Ciccotosto GD, James SA, Altissimo M, et al. (2014) Quantitation and localization of intracellular redox active metals by X-ray fluorescence microscopy in cortical neurons derived from APP and APLP2 knockout tissue. *Metallomics* 6: 1894–1904.
339. Craddock TJ, Tuszynski JA, Chopra D, et al. (2012) The zinc dyshomeostasis hypothesis of Alzheimer's disease. *PLoS One* 7: e33552.
340. Christensen MK, Geneser FA (1995) Distribution of neurons of origin of zinc-containing projections in the amygdala of the rat. *Anat Embryol* 191: 227–237.
341. Jiang L, Dong H, Cao H, et al. (2019) Exosomes in pathogenesis, diagnosis, and treatment of Alzheimer's disease. *Med Sci Monit* 25: 3329–3335.
342. Song Z, Xu Y, Deng W, et al. (2020) Brain derived exosomes are a double-edged sword in Alzheimer's disease. *Front Mol Neurosci* 13:79.
343. van Niel G, Porto-Carreiro I, Simoes S, et al. (2006) Exosomes: a common pathway for a specialized function. *J Biochem* 140: 13–21.
344. Eitan E, Suire C, Zhang S, et al. (2016) Impact of lysosome status on extracellular vesicle content and release. *Ageing Res Rev* 32: 65–74.
345. Asai H, Ikezu S, Tsunoda S, et al. (2015) Depletion of microglia and inhibition of exosome synthesis halt tau propagation. *Nat Neurosci* 18: 1584–1593.
346. Dinkins MB, Enasko J, Hernandez C, et al. (2016) Neutral sphingomyelinase-2 deficiency ameliorates Alzheimer's disease pathology and improves cognition in the 5XFAD mouse. *J Neurosci* 36: 8653–8667.
347. Xiao T, Zhang W, Jiao B, et al. (2017) The role of exosomes in the pathogenesis of Alzheimer's disease. *Transl Neurodegener* 6: 3.
348. Yuyama K, Igarashi Y (2017) Exosomes as carriers of Alzheimer's amyloid-ss. *Front Neurosci* 11: 229.
349. Cataldo AM, Barnett JL, Pieroni C, et al. (1997) Increased neuronal endocytosis and protease delivery to early endosomes in sporadic Alzheimer's disease: neuropathologic evidence for a mechanism of increased beta-amyloidogenesis. *J Neurosci* 17: 6142–6151.
350. Cataldo A, Rebeck GW, Ghetti B, et al. (2001) Endocytic disturbances distinguish among subtypes of Alzheimer's disease and related disorders. *Ann Neurol* 50: 661–665.
351. Cataldo AM, Petanceska S, Terio NB, et al. (2004) Abeta localization in abnormal endosomes: association with earliest Abeta elevations in AD and Down syndrome. *Neurobiol Aging* 25: 1263–1272.
352. Takahashi RH, Milner TA, Li F, et al. (2002) Intraneuronal Alzheimer abeta42 accumulates in multivesicular bodies and is associated with synaptic pathology. *Am J Pathol* 161: 1869–1879.
353. Langui D, Girardot N, El Hachimi KH, et al. (2004) Subcellular topography of neuronal Abeta peptide in APPxPS1 transgenic mice. *Am J Pathol* 165: 1465–1477.
354. Fiandaca MS, Kapogiannis D, Mapstone M, et al. (2015) Identification of preclinical Alzheimer's disease by a profile of pathogenic proteins in neurally derived blood exosomes: a case-control study. *Alzheimers Dement* 11: 600–607. e1.

355. Wang G, Dinkins M, He Q, et al. (2012) Astrocytes secrete exosomes enriched with proapoptotic ceramide and prostate apoptosis response 4 (PAR-4): potential mechanism of apoptosis induction in Alzheimer disease (AD). *J Biol Chem* 287: 21384–21395.
356. Bindea G, Mlecnik B, Hackl H, et al. (2009) ClueGO: a Cytoscape plug-in to decipher functionally grouped gene ontology and pathway annotation networks. *Bioinformatics* 25: 1091–1093.
357. Mosconi L, Berti V, Swerdlow RH, et al. (2010) Maternal transmission of Alzheimer's disease: prodromal metabolic phenotype and the search for genes. *Hum Genomics* 4: 170–193.
358. Birky CW Jr (1995) Uniparental inheritance of mitochondrial and chloroplast genes: mechanisms and evolution. *Proc Natl Acad Sci U S A* 92: 11331–11338.
359. Luo SM, Ge ZJ, Wang ZW, et al. (2013) Unique insights into maternal mitochondrial inheritance in mice. *Proc Natl Acad Sci U S A* 110: 13038–13043.
360. Yu Z, O'Farrell PH, Yakubovich N, et al. (2017) The mitochondrial DNA polymerase promotes elimination of paternal mitochondrial genomes. *Curr Biol* 27: 1033–1039.
361. Boumezbeur F, Mason GF, de Graaf RA, et al. (2010) Altered brain mitochondrial metabolism in healthy aging as assessed by in vivo magnetic resonance spectroscopy. *J Cereb Blood Flow Metab* 30: 211–221.
362. Rhein V, Song X, Wiesner A, et al. (2009) Amyloid-beta and tau synergistically impair the oxidative phosphorylation system in triple transgenic Alzheimer's disease mice. *Proc Natl Acad Sci U S A* 106: 20057–20062.
363. Larosa V, Remacle C (2018) Insights into the respiratory chain and oxidative stress. *Biosci Rep* 38.
364. Carden T, Singh B, Mooga V, et al. (2017) Epigenetic modification of miR-663 controls mitochondria-to-nucleus retrograde signaling and tumor progression. *J Biol Chem* 292: 20694–20706.
365. Spuch C, Ortolano S, Navarro C (2012) New insights in the amyloid-Beta interaction with mitochondria. *J Aging Res* 2012: 324968.
366. Yan SD, Stern DM (2005) Mitochondrial dysfunction and Alzheimer's disease: role of amyloid-beta peptide alcohol dehydrogenase (ABAD). *Int J Exp Pathol* 86: 161–171.
367. Cho DH, Nakamura T, Fang J, et al. (2009) S-nitrosylation of Drp1 mediates beta-amyloid-related mitochondrial fission and neuronal injury. *Science* 324: 102–105.
368. Area-Gomez E, Schon EA (2017) On the pathogenesis of Alzheimer's disease: the MAM Hypothesis. *FASEB J* 31: 864–867.
369. Snowden SG, Ebshiana AA, Hye A, et al. (2017) Association between fatty acid metabolism in the brain and Alzheimer disease neuropathology and cognitive performance: a nontargeted metabolomic study. *PLoS Med* 14: e1002266.
370. Kao YC, Ho PC, Tu YK, et al. (2020) Lipids and Alzheimer's disease. *Int J Mol Sci* 21: 1505.
371. Czubowicz K, Jesko H, Wencel P, et al. (2019) The role of ceramide and Sphingosine-1-Phosphate in Alzheimer's disease and other neurodegenerative disorders. *Mol Neurobiol* 56: 5436–5455.
372. Popugaeva E, Pchitskaya E, Bezprozvanny I (2018) Dysregulation of intracellular calcium signaling in Alzheimer's disease. *Antioxid Redox Signal* 29: 1176–1188.
373. Ruiz A, Matute C, Alberdi E (2009) Endoplasmic reticulum Ca(2+) release through ryanodine and IP(3) receptors contributes to neuronal excitotoxicity. *Cell Calcium* 46: 273–281.

374. Bezprozvanny I (2009) Calcium signaling and neurodegenerative diseases. *Trends Mol Med* 15: 89–100.
375. Tong BC, Wu AJ, Li M, et al. (2018) Calcium signaling in Alzheimer's disease & therapies. *Biochim Biophys Acta Mol Cell Res* 1865: 1745–1760.
376. Etcheberrigaray R, Hirashima N, Nee L, et al. (1998) Calcium responses in fibroblasts from asymptomatic members of Alzheimer's disease families. *Neurobiol Dis* 5: 37–45.
377. Berridge MJ (2009) Inositol trisphosphate and calcium signalling mechanisms. *Biochim Biophys Acta* 1793: 933–940.
378. Huang WJ, Zhang X, Chen WW (2016) Role of oxidative stress in Alzheimer's disease. *Biomed Rep* 4: 519–522.
379. Mauvezin C, Neufeld TP (2015) Bafilomycin A1 disrupts autophagic flux by inhibiting both V-ATPase-dependent acidification and Ca-P60A/SERCA-dependent autophagosome-lysosome fusion. *Autophagy* 11: 1437–1438.
380. Medina DL, Di Paola S, Peluso I, et al. (2015) Lysosomal calcium signalling regulates autophagy through calcineurin and TFEB. *Nat Cell Biol* 17: 288–299.
381. Mondal AC (2019) Role of GPCR signaling and calcium dysregulation in Alzheimer's disease. *Mol Cell Neurosci* 101: 103414.
382. Fernandez-Fernandez D, Rosenbrock H, Kroker KS (2015) Inhibition of PDE2A, but not PDE9A, modulates presynaptic short-term plasticity measured by paired-pulse facilitation in the CA1 region of the hippocampus. *Synapse* 69: 484–496.
383. Zhang G, Stackman RW Jr (2015) The role of serotonin 5-HT_{2A} receptors in memory and cognition. *Front Pharmacol* 6: 225.
384. Raote I, Bhattacharya A, Panicker MM (2007) Serotonin 2A (5-HT_{2A}) receptor function: ligand-dependent mechanisms and pathways. In: Chattopadhyay A, editor. *Serotonin Receptors in Neurobiology*. Boca Raton (FL): (Frontiers in Neuroscience), 105–132.
385. Chang L, Karin M (2001) Mammalian MAP kinase signalling cascades. *Nature* 410: 37–40.
386. Hullinger R, O'Riordan K, Burger C (2015) Environmental enrichment improves learning and memory and long-term potentiation in young adult rats through a mechanism requiring mGluR5 signaling and sustained activation of p70s6k. *Neurobiol Learn Mem* 125: 126–134.
387. Allen KD, Gourov AV, Harte C, et al. (2014) Nucleolar integrity is required for the maintenance of long-term synaptic plasticity. *PLoS One* 9: e104364.
388. Borroto-Escuela DO, Tarakanov AO, Guidolin D, et al. (2011) Moonlighting characteristics of G protein-coupled receptors: focus on receptor heteromers and relevance for neurodegeneration. *IUBMB Life* 63: 463–472.
389. Spilman P, Podlutskaya N, Hart MJ, et al. (2010) Inhibition of mTOR by rapamycin abolishes cognitive deficits and reduces amyloid-beta levels in a mouse model of Alzheimer's disease. *PLoS One* 5: e9979.
390. Caccamo A, Maldonado MA, Majumder S, et al. (2011) Naturally secreted amyloid-beta increases mammalian target of rapamycin (mTOR) activity via a PRAS40-mediated mechanism. *J Biol Chem* 286: 8924–8932.
391. Lipton JO, Sahin M (2014) The neurology of mTOR. *Neuron* 84: 275–291.
392. Oddo S (2012) The role of mTOR signaling in Alzheimer disease. *Front Biosci* 4: 941–952.

393. Caccamo A, De Pinto V, Messina A, et al. (2014) Genetic reduction of mammalian target of rapamycin ameliorates Alzheimer's disease-like cognitive and pathological deficits by restoring hippocampal gene expression signature. *J Neurosci* 34: 7988–7998.
394. Hodges SL, Reynolds CD, Smith GD, et al. (2018) Molecular interplay between hyperactive mammalian target of rapamycin signaling and Alzheimer's disease neuropathology in the NS-Pten knockout mouse model. *Neuroreport* 29: 1109–1113.
395. Gabbouj S, Ryhanen S, Marttinen M, et al. (2019) Altered insulin signaling in Alzheimer's disease brain—special emphasis on PI3K-Akt Pathway. *Front Neurosci* 13: 629.
396. Magri L, Cambiaghi M, Cominelli M, et al. (2011) Sustained activation of mTOR pathway in embryonic neural stem cells leads to development of tuberous sclerosis complex-associated lesions. *Cell Stem Cell* 9: 447–462.
397. Li YH, Werner H, Puschel AW (2008) Rheb and mTOR regulate neuronal polarity through Rap1B. *J Biol Chem* 283: 33784–33792.
398. Urbanska M, Gozdz A, Swiech LJ, et al. (2012) Mammalian target of rapamycin complex 1 (mTORC1) and 2 (mTORC2) control the dendritic arbor morphology of hippocampal neurons. *J Biol Chem* 287: 30240–30256.
399. Franco R, Martinez-Pinilla E, Navarro G, et al. (2017) Potential of GPCRs to modulate MAPK and mTOR pathways in Alzheimer's disease. *Prog Neurobiol* 149: 21–38.
400. Perluigi M, Di Domenico F, Butterfield DA (2015) mTOR signaling in aging and neurodegeneration: at the crossroad between metabolism dysfunction and impairment of autophagy. *Neurobiol Dis* 84: 39–49.
401. Ma T, Hoeffler CA, Capetillo-Zarate E, et al. (2010) Dysregulation of the mTOR pathway mediates impairment of synaptic plasticity in a mouse model of Alzheimer's disease. *PLoS One* 5: e12845.
402. Kudo W, Lee HP, Smith MA, et al. (2012) Inhibition of Bax protects neuronal cells from oligomeric Aβ neurotoxicity. *Cell Death Dis* 3: e309.
403. Tait SW, Green DR (2010) Mitochondria and cell death: outer membrane permeabilization and beyond. *Nat Rev Mol Cell Biol* 11: 621–632.
404. Gross A, McDonnell JM, Korsmeyer SJ (1999) BCL-2 family members and the mitochondria in apoptosis. *Genes Dev* 13: 1899–1911.
405. Putcha GV, Deshmukh M, Johnson EM Jr. (1999) BAX translocation is a critical event in neuronal apoptosis: regulation by neuroprotectants, BCL-2, and caspases. *J Neurosci* 19: 7476–7485.
406. Edlich F, Banerjee S, Suzuki M, et al. (2011) Bcl-x(L) retrotranslocates Bax from the mitochondria into the cytosol. *Cell* 145: 104–116.
407. Su Y, Ryder J, Li B, et al. (2004) Lithium, a common drug for bipolar disorder treatment, regulates amyloid-beta precursor protein processing. *Biochemistry* 43: 6899–6908.
408. Xu X, Zhang A, Zhu Y, et al. (2018) MFG-E8 reverses microglial-induced neurotoxic astrocyte (A1) via NF-κB and PI3K-Akt pathways. *J Cell Physiol* 234: 904–914.
409. Jimenez S, Torres M, Vizuete M, et al. (2011) Age-dependent accumulation of soluble amyloid beta (Aβ) oligomers reverses the neuroprotective effect of soluble amyloid precursor protein-alpha (sAPP(α)) by modulating phosphatidylinositol 3-kinase (PI3K)/Akt-GSK-3β pathway in Alzheimer mouse model. *J Biol Chem* 286: 18414–18425.

410. Cho SJ, Yun SM, Jo C, et al. (2019) Altered expression of Notch1 in Alzheimer's disease. *PLoS One* 14: e0224941.
411. Taylor KL, Henderson AM, Hughes CC (2002) Notch activation during endothelial cell network formation in vitro targets the basic HLH transcription factor HESR-1 and downregulates VEGFR-2/KDR expression. *Microvasc Res* 64: 372–383.
412. Yoon KJ, Lee HR, Jo YS, et al. (2012) Mind bomb-1 is an essential modulator of long-term memory and synaptic plasticity via the Notch signaling pathway. *Mol Brain* 5: 40.
413. Basak O, Giachino C, Fiorini E, et al. (2012) Neurogenic subventricular zone stem/progenitor cells are Notch1-dependent in their active but not quiescent state. *J Neurosci* 32: 5654–5666.
414. Brai E, Alina Raio N, Alberi L (2016) Notch1 hallmarks fibrillary depositions in sporadic Alzheimer's disease. *Acta Neuropathol Commun* 4: 64.
415. Caricasole A, Copani A, Caraci F, et al. (2004) Induction of Dickkopf-1, a negative modulator of the Wnt pathway, is associated with neuronal degeneration in Alzheimer's brain. *J Neurosci* 24: 6021–6027.
416. Rosi MC, Luccarini I, Grossi C, et al. (2010) Increased Dickkopf-1 expression in transgenic mouse models of neurodegenerative disease. *J Neurochem* 112: 1539–1551.
417. Cerpa W, Godoy JA, Alfaro I, et al. (2008) Wnt-7a modulates the synaptic vesicle cycle and synaptic transmission in hippocampal neurons. *J Biol Chem* 283: 5918–5927.
418. Farias GG, Valles AS, Colombres M, et al. (2007) Wnt-7a induces presynaptic colocalization of alpha 7-nicotinic acetylcholine receptors and adenomatous polyposis coli in hippocampal neurons. *J Neurosci* 27: 5313–5325.
419. Tapia-Rojas C, Inestrosa NC (2018) Loss of canonical Wnt signaling is involved in the pathogenesis of Alzheimer's disease. *Neural Regen Res* 13: 1705–1710.
420. Elliott C, Rojo AI, Ribe E, et al. (2018) A role for APP in Wnt signalling links synapse loss with beta-amyloid production. *Transl Psychiatry* 8: 179.
421. Aso E, Ferrer I (2014) Cannabinoids for treatment of Alzheimer's disease: moving toward the clinic. *Front Pharmacol* 5: 37.
422. Solas M, Francis PT, Franco R, et al. (2013) CB2 receptor and amyloid pathology in frontal cortex of Alzheimer's disease patients. *Neurobiol Aging* 34: 805–808.
423. Barnado A, Crofford LJ, Oates JC (2016) At the Bedside: Neutrophil extracellular traps (NETs) as targets for biomarkers and therapies in autoimmune diseases. *J Leukoc Biol* 99: 265–278.
424. Hamilton A, Esseltine JL, DeVries RA, et al. (2014) Metabotropic glutamate receptor 5 knockout reduces cognitive impairment and pathogenesis in a mouse model of Alzheimer's disease. *Mol Brain* 7: 40.



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