

**REPLY: Apoptosis, A Double-Edge Sword!**



We appreciate the comments of Drs. Rossello and Yellon regarding our recent paper (1), and we agree that activation of pro-survival pathways in viable myocytes after brief periods of ischemia likely occurs alongside apoptotic myocyte death to provide protection against subsequent ischemic injury. Aside from the severe transmural ischemia associated with an acute transient occlusion that we used in our study, apoptosis-induced myocyte cell loss with compensatory myocyte cellular hypertrophy can arise in response to reversible subendocardial ischemia and can ultimately affect regional as well as global left ventricular function. Thus, modulating this dynamic interplay between myocyte survival and death may be most important with repetitive ischemia similar to angina in chronic coronary artery disease that can contribute to the development of ischemic cardiomyopathy. For example, we previously demonstrated that chronic repetitive subendocardial ischemia distal to a chronic coronary artery stenosis in swine leads to a progressive increase in apoptosis as the physiological significance of a coronary stenosis increases (2,3). This is followed by regional myocyte loss, compensatory cellular hypertrophy, and physiological features consistent with hibernating myocardium. Interestingly, proteomic profiling has demonstrated that this is accompanied by an up-regulation of a variety of pro-survival proteins as well as a down-regulation in mitochondrial metabolism (4,5). This is functionally significant, as hibernating myocardium exhibits depressed mitochondrial respiration and a reduced rate of ATP depletion during simulated zero flow ischemia *in vitro* (6). Others have also demonstrated activation of pro-survival pathways during repetitive non-transmural ischemia (7). These cardioprotective adaptations are reversed by alleviating demand-induced ischemia with coronary revascularization (8). Collectively, these findings support the notion that hibernating myocardium becomes ischemia-tolerant to prevent a supply-demand imbalance and reduce mitochondrial oxidative stress as a counter-regulatory process to attenuate myocyte loss from apoptosis in a fashion similar to that proposed by Rossello and Yellon for ischemic pre-conditioning and infarction. We agree that further work is needed to understand whether favorable manipulation of the balance between pro- and anti-apoptotic signaling pathways can have a long-term effect on myocyte

loss after ischemia and prevent the development of ischemic cardiomyopathy.

Brian R. Weil, PhD

Rebeccah F. Young, MA

Xiaomeng Shen, BS

Gen Suzuki, MD, PhD

Jun Qu, PhD

Saurabh Malhotra, MD, MPH

\*John M. Carty, Jr., MD

\*Division of Cardiovascular Medicine

Jacobs School of Medicine and Biomedical Sciences

University at Buffalo

Clinical Translational Research Center, Suite 7030

875 Ellicott Street

Buffalo, New York 14203

E-mail: [carty@buffalo.edu](mailto:carty@buffalo.edu)

<http://dx.doi.org/10.1016/j.jacbs.2017.07.009>

Please note: This work has received funding from the National Heart, Lung, and Blood Institute (HL-055324, HL-061610, and F32HL-114335), the National Center for Advancing Translational Sciences (UL1TR001412), the Department of Veterans Affairs (IIOIBX002659), the George E. Becker Fund for Heart Research, and the Albert and Elizabeth Rekate Fund in Cardiovascular Medicine. The authors have reported that they have no relationships relevant to the contents of this paper to disclose.

Published by Elsevier on behalf of the American College of Cardiology Foundation. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>)

**REFERENCES**

1. Weil BR, Young RF, Shen X, et al. Brief myocardial ischemia produces cardiac troponin I release and focal myocyte apoptosis in the absence of pathological infarction in swine. *J Am Coll Cardiol Basic Trans Science* 2017; 2:105-14.
2. Lim H, Fallavollita JA, Hard R, Kerr CW, Carty JM Jr. Profound apoptosis-mediated regional myocyte loss and compensatory hypertrophy in pigs with hibernating myocardium. *Circulation* 1999;100:2380-6.
3. Fallavollita JA, Lim H, Carty JM. Myocyte apoptosis and reduced SR gene expression precede the transition from chronically stunned to hibernating myocardium. *J Mol Cell Cardiol* 2001;33:1937-44.
4. Page B, Young R, Iyer V, et al. Persistent regional downregulation in mitochondrial enzymes and upregulation of stress proteins in swine with chronic hibernating myocardium. *Circ Res* 2008;102:103-12.
5. Duan XT, Young R, Straubinger RM, et al. A straightforward and highly efficient precipitation/on-pellet digestion procedure coupled with a long gradient nano-LC separation and Orbitrap mass spectrometry for label-free expression profiling of the swine heart mitochondrial proteome. *J Proteome Res* 2009;8:2838-50.
6. Hu Q, Suzuki G, Young RF, Page BJ, Fallavollita JA, Carty JM Jr. Reductions in mitochondrial O<sub>2</sub> consumption and preservation of high-energy phosphate levels after simulated ischemia in chronic hibernating myocardium. *Am J Physiol Heart Circ Physiol* 2009;297:H223-32.
7. Depre C, Tomlinson JE, Kudej RK, et al. Gene program for cardiac cell survival induced by transient ischemia in conscious pigs. *Proc Natl Acad Sci U S A* 2001;98:9336-41.
8. Page BJ, Banas MD, Suzuki G, et al. Revascularization of chronic hibernating myocardium stimulates myocyte proliferation and partially reverses chronic adaptations to ischemia. *J Am Coll Cardiol* 2015;65:684-97.