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REVIEW ARTICLE

Role of MicroRNA, LncRNA, and Exosomes in the Progression of Osteoarthritis: A Review of Recent Literature

Fang Xie, MM^{1*} ^(D), Yong-li Liu, MM^{1*}, Xiu-yuan Chen, MM¹, Qian Li, MM¹, Jia Zhong, MM¹, Bin-yu Dai, MM¹, Xian-fang Shao, MD¹, Guan-bao Wu, MD²

¹Affiliated Changde Hospital, Hunan University of Traditional Chinese Medicine, Changde and ²Department of Orthopaedics, Affiliated Hospital of Hunan Academy of Traditional Chinese Medicine, Changsha, China

Osteoarthritis (OA) is a common clinical degenerative disease characterized by the destruction of articular cartilage, which has an increasing impact on people's lives and social economy. The pathogenesis of OA is complex and unclear, and there is no effective way to block its progress. The study of the pathogenesis of OA is the prerequisite for the early diagnosis and effective treatment of OA. To define the pathogenesis of OA, this review considers the pathological mechanism of OA that involves microRNA, lncRNA, and exosomes. More and more evidence shows that microRNA, lncRNA, and exosomes are closely related to OA. MicroRNA inhibits the target gene by binding to the 3'-untranslated region of the targets. LncRNA usually competes with microRNA to regulate the expression level of down-stream genes, while exosomes, as a carrier of intercellular information transfer, transmit the biological information of mother cells to target cells, and the effect of exosomes secreted by different cells on OA are different. In this review, we emphasized that different microRNA, lncRNA, and exosomes have different regulatory effects on chondrocyte proliferation and apoptosis, extracellular matrix degradation and inflammation. Besides, we classified and analyzed these molecules according to their effects on the progress of OA. Based on the analysis of the reported literature, this review reveals some pathogenesis of OA, and emphasizes that microRNA, lncRNA, and exosomes have great potential to assist early diagnosis and effective treatment of OA.

Key words: Exosomes; LncRNA; MicroRNA; Osteoarthritis

Introduction

O steoarthritis (OA) is a common degenerative disease related to age, obesity, gender, weight, and trauma¹. It is characterized by synovial hyperplasia, osteophyte formation, subchondral osteosclerosis, progressive articular cartilage destruction, and cartilage loss caused by the imbalance of extracellular matrix synthesis and catabolism². According to statistics, OA has an impact on the lives of 250 mn people around the world, bringing an annual economic burden of more than US\$89.1bn³. At present, the treatment strategy for early and middle stage OA is to relieve joint pain and intraarticular injection, and the treatment strategy for late stage OA is joint replacement surgery. However, although these treatments can alleviate the symptoms of OA and improve the quality of life of patients to a certain extent, they have little effect on blocking the progressive development of OA. The specific pathogenesis of OA is still not clear. The existing evidence shows that the pathogenesis of OA is related to inflammatory factors, abnormal apoptosis of chondrocytes, and degradation of extracellular matrix. During

Address for correspondence Xian-fang Shao, MD, Affiliated Changde Hospital, Hunan University of Traditional Chinese Medicine, No.588 of Binhu Road, Changde, China 415000; Tel:+86-0736-7893888; Email: 13973657615@139.com, Guan-bao Wu, MD, Department of Orthopaedics, Affiliated Hospital of Hunan Academy of Traditional Chinese Medicine, No.58 of Lushan Road, Yuelu District, Changsha, China 410006; Tel:+86-0731-88854064; Email:yhywgb@126.com

*These authors contributed equally to this study.

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the development of OA, TNF-a, IL-1, IL-6, and other inflammatory factors were abnormally expressed, which led to the increase of chondrocyte apoptosis and the degradation of extracellular matrix^{1,4}. At present, there is still a lack of effective means for the early diagnosis and treatment of OA.

MicroRNA (miRNA) are a kind of multifunctional non-coding RNA molecule with 22-25 bases encoded by endogenous genes. MiRNA regulate the stability and translation of mRNA, inhibit splicing and translation, inhibit target gene expression, and regulate downstream pathway by fully complementary binding with the 3'- untranslated region (3'-UTR) of target mRNA⁵⁻⁷. Long non-coding RNA (LncRNA) is a kind of non-coding RNA with a length of more than 200 bases. It does not encode proteins. It is a regulatory molecule. According to their positions relative to the protein coding genes, it can be divided into five categories: (i) antisense lncRNA; (ii) enhancer lncRNA; (iii) large International non-coding RNA; (iv) bidirectional lncRNA; and (v) intronic script lncRNA (intron lncRNA)^{8,9}. LncRNA binds and isolates microRNA away from the sites that act on mRNA, thereby reducing the effect of microRNA on mRNA expression¹⁰. Figure 1 illustrates the interaction mechanism of LncRNA, microRNA, and mRNA. Exosomes (Exo) is a kind of vesicle with double plasmalemma structure, which is secreted by cells. It can express CD63, CD9, and other marker proteins on its surface. The membrane contains protein molecules, mRNA, miRNA, lncRNA, and other signal substances, carries the specific cytokines of mother cells, and targets the proximal cells through autocrine and paracrine, or the distal cells through the circulatory system tissue, information exchange between cells^{11,12}.

In recent years, with the development of molecular biology technology, the important role of miRNA, lncRNA, and Exo in disease progression has been gradually discovered by researchers. MicroRNA, lncRNA, and Exo are also expected to be an important way to explain the pathogenesis, early diagnosis, and treatment of OA. The purpose of this review is to summarize the key role of microRNA, lncRNA, and Exo in the development of OA. All the information is extracted from the high-quality literature retrieved in the PubMed database.

Methods

With the help of the library platform of Hunan University of Traditional Chinese Medicine, PubMed database was searched. The literature from 2010 to 2019 were searched with the keywords "osteoarthritis," "microRNA," "lncRNA," "exosomes," and the language was English. Figure 2 illustrates the flow chart of searched results.

Eight hundred and thirty-four related citations were obtained by literature search. Three hundred and sixty-six related citations were obtained by taking experimental studies related to the pathological process of OA as the selection criteria and deleting the duplicate part. The two authors independently screened the titles and abstracts of each article, excluding the comprehensive research, clinical research, meeting report, and case report. The screening process was completed by the Rayyan QCRI software, and the dispute part was solved by the third author independently. A total of 158 documents have been full-text reviewed. Through the full-text review, there are 100 documents in line with our selection criteria, including 35 documents related to







microRNA and OA, 54 documents related to lncRNA and OA, and 11 documents related to exosomes and OA. A descriptive analysis was performed on included studies.

Results

MicroRNA

In the pathogenesis of OA, miRNA has the biological functions of regulating chondrocyte apoptosis and proliferation, extracellular matrix metabolism, inflammatory response, and so on¹³⁻¹⁶. The keywords of miRNA and osteoarthritis were searched in the PubMed database, 39 of which were research literature on molecular mechanism. It was found that there were 16 kinds of miRNA inhibiting OA process and 14 kinds of miRNA promoting OA process (Table 1).

TABLE 1 classification of miRNA in OA process		
Inhibiting OA process (16 kinds)	Promoting OA process(14 kinds)	
$\begin{array}{c} {\rm miR}\text{-}132^{17.18}, {\rm miR}\text{-}107^{19}, \\ {\rm miR}\text{-}149\text{-}5p^{20}, {\rm miR}\text{-}93\text{-}5p^{21}, \\ {\rm miR}\text{-}335\text{-}5p^{22}, {\rm miR}\text{-}4784^{23}, \\ {\rm miR}\text{-}106a\text{-}5p^{24}, {\rm miR}\text{-}145^{25}, \\ {\rm miR}\text{-}140^{26,27}, {\rm miR}\text{-}221^{28}, \\ {\rm miR}\text{-}381^{29}, {\rm miR}\text{-}105^{30}, \\ {\rm miR}\text{-}210^{31}, {\rm miR}\text{-}29a^{27}, \\ {\rm miR}\text{-}488^{32}, {\rm miR}\text{-}125b^{33}, \\ {\rm miR}\text{-}101^{34} \end{array}$	$\begin{array}{l} {\rm miR-146b}^{35.36}, {\rm miR-34a}^{37.38}, \\ {\rm miR-181a}^{37.39,40}, {\rm miR-582-5p}^{41}, \\ {\rm miR-324-5p}^{42}, {\rm miR-21-5p}^{43}, \\ {\rm miR-483-5p}^{44.45}, {\rm miR-384-5p}^{46}, \\ {\rm miR-155}^{39}, {\rm miR-98}^{47}, \\ {\rm miR-127-5p}^{48}, {\rm miR-16-5p}^{49}, \\ {\rm miR-101}^{50}, {\rm miR-146a}^{51} \end{array}$	

Fig. 2 The flow chart of literature search and screening.

Apoptosis is a form of programmed cell death, which involves a series of gene activation, expression, and regulation⁵². Out of control apoptosis can cause cancer, autoimmune diseases, and degenerative diseases. Over apoptosis of chondrocytes is the main pathological manifestation of OA⁵³. MiRNA promote or inhibit chondrocyte apoptosis by regulating the expression of key molecules involved in apoptosis signaling pathways. MiR-34a and miR-108a have a synergistic relationship in chondrocytes, which can jointly promote the activity of p50 NF- κ B, reduce the expression of Bcl-2, and promote chondrocyte apoptosis³⁷; in addition, miR-98 can also inhibit the transcription of Bcl-2 and the apoptosis of chondrocytes⁴⁷. According to the research of Zhao *et al.*⁴⁰, miR-181a can inhibit the expression of Glycerol-3-Phosphate Dehydrogenase 1-Like Protein (GPD1L) and accelerate the apoptosis of chondrocytes by binding with the 3'- UTR end of GPD1L. In addition, miR-181a may have a cooperative relationship with miR-155³⁹. There are other targets in miR-34a, and Yang et al.³⁸ have proved that miR-34a targets cysteine-rich angiogenic inductor 61 (CYR61), and inhibiting the expression of miR-34a can promote the proliferation of chondrocytes. MiR-146b can inhibit the expression of alpha-2-macroglobulin (A2M), improve the activity of proteolytic enzyme, promote cell apoptosis, and accelerate the development of OA35. MiR-384-5p inhibit cell proliferation and induce apoptosis by targeting Sox946. MiR-127-5p targeting combined with OPN inhibit chondrocyte proliferation⁴⁸. Li et al.³¹ showed that miR-210 inhibit the expression of HIF-3 α and promoted the proliferation of

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chondrocytes. MiR-146a target Smad4 to promote chondrocyte apoptosis⁵¹. MiR-29a can upregulate the expression of type II collagen, reduce the expression of MMP13, resist the apoptosis of chondrocytes, and delay the development of OA²⁷.

The extracellular matrix (ECM) of articular cartilage is mainly composed of proteoglycan and type II collagen, including a small number of chondrocytes. ECM contains a large number of signal molecules, which actively participate in the control of cell growth, polarity, shape, migration, and metabolism. The disruption of ECM catabolic balance will lead to the occurrence of OA⁵⁴. Wang et al.⁴⁵ found that miR-483-5p directly target Matn3 and TIMP2 to promote the degradation of ECM, chondrocyte hypertrophy, and cartilage angiogenesis, and accelerate the process of OA. Zheng et al.²⁸ confirmed that miR-221 was down regulate in OA, and miR-221 could inhibit the degradation of cartilage extracellular matrix through SDF1 / CXCR4 signaling pathway. MiR-145 inhibit the phosphorylation of MMK4 and alleviate the degradation of cartilage extracellular matrix caused by TNF - α stimulation²⁵. Liu *et al.*²³ found that the expression of miR-4784 is low in the early stage of OA, overexpression of miR-4784 can increase the expression of type II collagen in cartilage extracellular matrix, decrease the expression of MMP3, and inhibit the degradation of extracellular matrix. Li et al.⁴⁹ found that the expression of miR-16-5p in OA tissue is higher than that in normal tissue and further experiments confirm that miR-16-5p can target Smad3 to promote the degradation of cartilage extracellular matrix and promote the development of OA. In the experiments of Dai et al.⁵⁰, they found that silencing miR-101 can increase the expression of Sox9 and type II collagen and proteoglycan, thus inhibiting the degradation of cartilage extracellular matrix. However, Gao et al.³⁴ found that the expression of miR-101 increases while that of Sox9 and Runx2 decreases.

Long Non-Coding RNA

The pathogenesis of OA is still unclear, but a large amount of evidence shows that the interaction between lncRNA and miRNA plays an important role in the development of OA. LncRNA can competitively bind miRNA, act as competitive endogenous RNA (CeRNA), reduce the combination of miRNA and downstream genes, and increase the transcription and expression of downstream genes^{55,56}. LncRNA has become an early diagnosis and effective treatment target of OA. In the PubMed database, 45 relevant pieces of literature were searched with the keywords of lncRNA and osteoarthritis, and 34 kinds of lncRNA were involved in these studies, including 17 kinds of lncRNA molecules inhibiting the development of OA and 15 kinds of lncRNA molecules promoting the development of OA (Table 2).

The combination of lncRNA and miRNA interferes with the inhibition of miRNAs on the expression of downstream target genes. That is to say, the expression of lncRNA is negatively correlated with the corresponding miRNAs and positively correlated with the expression of downstream

target genes. Competitive binding of mRNA to miRNA is the main way for lncRNA to regulate biological functions. In the known research, it has been confirmed that some lncRNA have multi-target and multi-level regulatory effect. The study of Wang *et al.*⁵⁷ confirmed that lncRNA FOXD2-AS1 was low expression in OA patients, further experiments show that lncRNA FOXD2-AS1 promote chondrocyte proliferation and inhibit the development of OA through miR-27a / TLR4 axis. Cao et al.58 confirm that lncRNA FOXD2-AS1 inhibit the expression of miR-206 and upregulate the expression of CCND1, through miR-206 / CCND1 axis, and it promote the survival and development of chondrocytes and hinder the process of OA, besides, lncRNA DANCR promote the proliferation of OA chondrocytes through miR-216a / JAK2 axis 76 and miR-577 / Sphk273 axis. LncRNA CIR can promote apoptosis through miR-130a / Bim axis⁸⁹, and inhibit the expression of miR- $27b^{91}$ to promote the degradation of extracellular matrix. LncRNA PVT1 target regulation of miR-14992 and miR-488⁹³ expression promotes apoptosis, extracellular matrix degradation, and inflammatory response. LncRNA HOTAIR inhibit miR-12496 expression or promote inflammatory response and apoptosis through miR-17-5p / FUT2 /βcatenin axis⁹⁶.

Jiang et al.⁶⁵ found that the expression of lncRNA PACER is low in OA and inhibited cell apoptosis. The negative correlation between the expression of PACER and lncRNA HOTAIR suggested that there is also a mechanism of mutual regulation between lncRNA. There are also some lncRNA that have been proved to be related to OA, but the specific regulatory mechanism is not clear. Li et al.⁵⁹ found that lncRNA ANCR can promote chondrocyte proliferation in OA patients, and its expression was negatively correlated with TGF - β 1. Huang *et al.*⁶⁰ alleviate the inflammatory response in OA by inhibiting the expression of IL-6. Yang et al.⁸¹ found that when the expression of lncRNA LOC101928134 is downregulated, the expression of IFN1 increase and activate the downstream JAK / STAT signal pathway, inhibiting the proliferation of synovium and cartilage destruction, but the specific regulatory mechanism of LOC101928134 is not clear. In addition, lncRNA ZFAS1 can inhibit the activation of Wnt3a pathway and promote the proliferation and migration of chondrocytes⁷³. The expression of GACAT3 is negatively correlated with IL-6, inhibits the activity of IL-6 / STAT pathway, and promotes chondrocyte proliferation⁷⁴. The study of Park et al.⁸⁴ found that the overexpression of lncRNA Nespas can inhibit the expression of miR-291a-3p, miR-196a-5p, miR-23a-3p, miR-24-3p, and Let-7a-5p. After analysis, these miRNAs target ACSL6, but the transcriptional binding of these miRNAs with lncRNA Nespas and ACSL6 needs further experimental verification.

Interestingly, Zhang *et al.*⁷⁶ found that the expression of lncRNA DANCR increases in OA patients, DANCR promotes the proliferation and inflammatory response of OA chondrocytes, inhibits apoptosis, and promotes the progression of OA. Fan *et al.*⁷⁷ also found that the expression of

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TABLE 2 classification of IncRNA in OA process			
	LncRNA	Targets	Cell process
Inhibiting OA process	FOXD2-AS157,58	MiR-27a/TLR4 ⁵⁷	Chondrocyte proliferation (+)
		MiR-206/CCND1 ⁵⁸	Chondrocyte proliferation (+)
	ANCR ⁵⁹		Chondrocyte proliferation (+)
	DILC ⁶⁰		Inflammation(-)
	MIR4435-2HG ⁶¹		Chondrocyte proliferation (+)
	SNHG1 ⁶²	MiR-16-5P	Inflammation (-)
	SNHG5 ⁶³	MiR-26a/Sox2	Chondrocyte proliferation (+)
	HULC ⁶⁴	MiR-101	Inflammation (-)
	PACER ⁶⁵		Chondrocyte apoptosis (-)
	MEG3 ^{66,67}	MiR-93/TGFBR2 ⁶⁶	Degradation of extracellular matrix (-)
		MiR-16/SAMD7 ⁶⁷	Chondrocyte proliferation (+)
	LINC00341 ⁶⁸	MiR-141/YAF2	Chondrocyte apoptosis (-)
	ATB ⁶⁹	MiR-223	Inflammation (-)
	PMS2L2 ⁷⁰	MiR-203	Inflammation (-)
	MALAT1 ⁷¹	MiR-150-5p/AKT3	Chondrocyte proliferation (+)
	ROR ⁷²		Chondrocyte apoptosis (-)
	ZFAS1 ⁷³		Chondrocyte proliferation (+)
	GACAT3 ⁷⁴		Chondrocyte proliferation (+)
	UFC1 ⁷⁵	MiR-34a	Chondrocyte apoptosis (-)
Promoting OA process	MIAI ¹⁷	MiR-132	Chondrocyte proliferation (-)
	DANCR ^{76,77}	MiR-216a/JAK2 ⁷⁶	OA Chondrocyte proliferation and inflammation (+
	2	MiR-577/Sphk2 ⁷⁷	OA Chondrocyte proliferation (+)
	TM1P3 ⁷⁸	MiR-22/MMP13	Degradation of extracellular matrix (+)
	CTD-2574D22.4 ⁷⁹		Chondrocyte apoptosis and inflammation (+)
	TNFSF10 ⁸⁰	MiR-376/FGFR1	OA Chondrocyte proliferation and inflammation (+)
	L0C101928134 ⁸¹		Chondrocyte apoptosis (+)
	CASA2 ⁸²	TL-17	Chondrocyte apoptosis (+)
	CHRF ⁸³	MiR-146a/JAK1/STAT3	Inflammation (+)
	Nespas ⁸⁴		Chondrocyte apoptosis (+)
	H19 ⁸⁵	MiR-130a	Chondrocyte apoptosis (+)
	THRIL ⁸⁶	MiR-125b	Inflammation (+)
	TUG ⁸⁷	MiR-195/MMP-13	
	P21 ⁸⁸	MIR-195/MMP-13 MIR-130b/PTEN/AKT	Degradation of extracellular matrix (+)
	CIR ^{89–91}	MiR-1300/PTEN/AKT MiR-130a/Bim ⁸⁹	Chondrocyte apoptosis (+)
	CIR	MIR-130a/BIM-5	Chondrocyte apoptosis $(+)$
		N/D 071 91	Chondrocyte autophagy (+) ⁹⁰
	PVT1 ^{92,93}	MiR-27b ⁹¹	Degradation of extracellular matrix (+)
	PVI1	MiR-149 ⁹²	Degradation of extracellular matrix (+)
	NICT94	MiR-488 ⁹³	Chondrocyte apoptosis (+)
	XIST ⁹⁴	MiR-211/CXR4/MAPK	Chondrocyte apoptosis (+)
	MBNL1-AS1 ⁹⁵		Chondrocyte apoptosis (+)
	HOTAIR ⁹⁶	MiR-124 ⁹⁶	Inflammation (+)
	67	MiR-17-5p/FUT2/β-catenin ⁹⁶	Degradation of extracellular matrix (+)
	FAS-AS1 ⁹⁷		Degradation of extracellular matrix (+)
	MSR ⁹⁸	MiR-152	Degradation of extracellular matrix (+)
	PCGEM1 ⁹⁹	MiR-770	Chondrocyte apoptosis (+)

DANCR increases in OA patients and promotes the proliferation of OA chondrocytes through miR-577 / Sphk2 axis. The study of Huang *et al.*⁸⁰ confirmed that lncRNA TNFSF10 promotes OA chondrocyte proliferation, inhibits apoptosis, and promotes inflammatory response through miR-376 / FGFR1 axis. These experiments prove that OA chondrocytes are relate to inflammatory response, and promoting the proliferation of OA chondrocytes can accelerate the development of OA. In other words, the proliferation and inflammatory response of degenerated chondrocytes can accelerate the degeneration of cartilage¹⁰⁰. OA chondrocytes have different biological functions from normal chondrocytes.

Exosomes

The biological characteristics of Exo, which is not secreted by cells, are different, and its effect on OA is also different. Domenis *et al.*¹⁰¹ extracted and identified synovial-fluidderived exosomes of patients with osteoarthritis, and found that SF-Exo can promote inflammatory response. The Exo secreted by IL-1 β stimulate synovial fibroblasts can increase the expression of MMP-13 and ADAMTS5, decrease the expression of COL2A1 and ACAN, promote the degradation of cartilage extracellular matrix, and accelerate the development of OA¹⁰². Because Exo can carry the biological information of mother cells, more research regards Exo as a strategy of treatment. The study of Qi *et al.*¹⁰³ confirms that



through chondrocyte proliferation, chondrocyte apoptosis, extracellulsr matrix degradation and inflammation. Exo secrete by mesenchymal stem cells can promote Akt phosphorylation by inhibiting p38 and ERK phosphorylation, and inhibit chondrocyte apoptosis cause by mitochondrial dysfunction. Bone-marrow-mesenchymal-stem-

phosphorylation by inhibiting p38 and ERK phosphorylation, and inhibit chondrocyte apoptosis cause by mitochondrial dysfunction. Bone-marrow-mesenchymal-stemcells Exo¹⁰⁴, embryonic-mesenchymal-stem-cells Exo¹⁰⁵, and adipose-mesenchymal-stem-cells Exo¹⁰⁶ can promote the expression of type II collagen and proteoglycan, inhibit the expression of MMP-13, ADAMTS5 and proinflammatory factors, and maintain the balance of cartilage extracellular matrix. Exosomes can also regulate the biological functions of target cells by carrying miRNA and lncRNA. Wu *et al.*¹⁰⁷ found that miR-100-5p was abundant in the Exo of subpatellar fat pad mesenchymal stem cells, and the activity of mTOP autophagy pathway in chondrocytes is inhibited by miR-100-5p to adjust the gait of OA in a rat model. Sun et al.¹⁰⁸ confirmed that Exo can promote chondrocyte proliferation and inhibit MMP-13 expression through miR-302c. The study of Mao et al.¹⁰⁹ confirmed that the expression of miR-92a in Exo of OA chondrocytes was lower than that of normal chondrocytes and further experiments show that human mesenchymal stem cells inhibit the expression of Wnt5a through miR-92a and alleviate the degeneration of articular cartilage. The Exo secrete by miR-140-5p overexpress synovial stem cells can promote the proliferation and migration of chondrocytes¹¹⁰. Liu *et al.*¹¹¹ confirmed that human mesenchymal stem cells could inhibit the apoptosis of chondrocytes induced by IL-1 β and promote cartilage repair through lncRNA KLF3-AS1.

Conclusions

MicroRNA, lncRNA, and Exo have strong regulatory effects on the pathological process of OA, so they are expected to be the targets of early diagnosis and treatment of OA. Figure 3 illustrates the mechanism of the study. The interaction mechanism of these three factors may be as follows: Exo secreted by other cells contains different lncRNA, micro-RNA, or mRNA; when Exo contacts the cell membrane, these substances are transferred to the target cell; in the cell, microRNA can bind mRNA to inhibit its expression. However, lncRNA can competitively bind microRNA to alleviate the inhibition of microRNA on the expression of downstream genes. All in all, their regulatory role in cells is ultiachieved by influencing mRNA expression mately downstream. From the current literature reports, the pathological research of OA involves microRNA, lncRNA, and Exo, which are basically in vitro experiments or animal model experiments. With the progress of molecular biotechnology in the later research, it is hoped that more clinical experimental reports will be published. In addition, due to the diversity of biological functions of microRNA and lncRNA and the characteristics of Exo information transmission media, research into the fusion of these three factors Orthopaedic Surgery Volume 12 • Number 3 • June, 2020 DEGENERATIVE MECHANISM OF OSTEOARTHRITIS

may effectively promote the research process of the pathological mechanism of OA.

Authorship Declaration

All authors listed meet the authorship criteria according to the latest guidelines of the International Committee of Medical Journal Editors, and all authors are in agreement with the manuscript.

1. Wang Y, Fan X, Xing L, Tian F. Wnt signaling: a promising target for

osteoarthritis therapy. Cell Commun Signal, 2019, 17: 97. 2. Chen D, Shen J, Zhao W, et al. Osteoarthritis: toward a comprehensive

understanding of pathological mechanism. Bone Res, 2017, 5: 16044.

3. GBD 2016 Disease and Injury Incidence and Prevalence Collaborators. Global, regional, and national incidence, prevalence, and years lived with disability for 328 diseases and injuries for 195 countries, 1990–2016: a systematic analysis for the Global Burden of Disease Study 2016. Lancet, 390: 1211–1259.

 Woodell-May JE, Sommerfeld SD. Role of inflammation and the immune system in the progression of osteoarthritis. J Orthop Res, 2020, 38: 253–257.
 Bartel DP. MicroRNAs: target recognition and regulatory functions. Cell, 2009, 136: 215–233.

6. Mo YY. MicroRNA regulatory networks and human disease. Cell Mol Life Sci, 2012. 69: 3529–3531.

7. Gebert L, MacRae IJ. Regulation of microRNA function in animals. Nat Rev Mol Cell Biol, 2019, 20: 21–37.

8. Quinn JJ, Chang HY. Unique features of long non-coding RNA biogenesis and function. Nat Rev Genet, 2016, 17: 47–62.

9. Huynh NP, Anderson BA, Guilak F, McAlinden A. Emerging roles for long noncoding RNAs in skeletal biology and disease. Connect Tissue Res, 2017, 58: 116–141.

10. Ballantyne MD, McDonald RA, Baker AH. IncRNA/MicroRNA interactions in the vasculature. Clin Pharmacol Ther, 2016, 99: 494–501.

11. Cheng X, Zhang G, Zhang L, et al. Mesenchymal stem cells deliver

exogenous miR-21 via exosomes to inhibit nucleus pulposus cell apoptosis and reduce intervertebral disc degeneration. J Cell Mol Med, 2018, 22: 261–276. **12.** Colombo M, Raposo G, Théry C. Biogenesis, secretion, and intercellular

interactions of exosomes and other extracellular vesicles. Annu Rev Cell Dev Biol, 2014, 30: 255–289.

13. Malemud CJ. MicroRNAs and osteoarthritis. Cell, 2018, 7: 92.

14. Al-Modawi RN, Brinchmann JE, Karlsen TA. Multi-pathway protective effects of MicroRNAs on human chondrocytes in an in vitro model of osteoarthritis. Mol Ther Nucleic Acids, 2019, 17: 776–790.

15. Le LT, Swingler TE, Clark IM. Review: the role of microRNAs in osteoarthritis and chondrogenesis. Arthritis Rheum, 2013, 65: 1963–1974.

16. Swingler TE, Niu L, Smith P, *et al*. The function of microRNAs in cartilage and osteoarthritis. Clin Exp Rheumatol, 2019, 37: 40–47.

17. Li C, Pan S, Song Y, Li Y, Qu J. Silence of IncRNA MIAT protects ATDC5 cells against lipopolysaccharides challenge via up-regulating miR-132. Artif Cells Nanomed Biotechnol, 2019, 47: 2521–2527.

 Zhou X, Luo D, Sun H, et al. MiR-132-3p regulates ADAMTS-5 expression and promotes chondrogenic differentiation of rat mesenchymal stem cells. J Cell Biochem, 2018, 119: 2579–2587.

19. Lin SS, Yuan LJ, Niu CC, Tu YK, Yang CY, Ueng S. Hyperbaric oxygen inhibits the HMGB1/RAGE signaling pathway by upregulating Mir-107 expression in human osteoarthritic chondrocytes. Osteoarthritis Cartilage, 2019, 27: 1372–1381.

20. Çelik E, Bayram C, Denkbaş EB. Chondrogenesis of human mesenchymal stem cells by microRNA loaded triple polysaccharide nanoparticle system. Korean J Couns Psychother, 2019, 102: 756–763.

21. Xue H, Tu Y, Ma T, et *al.* miR-93-5p attenuates IL-1 β -induced chondrocyte apoptosis and cartilage degradation in osteoarthritis partially by targeting TCF4. Bone, 2019, 123: 129–136.

22. Zhong G, Long H, Ma S, Shunhan Y, Li J, Yao J. miRNA-335-5p relieves chondrocyte inflammation by activating autophagy in osteoarthritis. Life Sci, 2019, 226: 164–172.

23. Liu J, Yu Q, Ye Y, Yan Y, Chen X. Abnormal expression of miR-4784 in chondrocytes of osteoarthritis and associations with chondrocyte hyperplasia. Exp Ther Med, 2018, 16: 4690–4694.

24. Ji Q, Qi D, Xu X, et al. Cryptotanshinone protects cartilage against developing osteoarthritis through the miR-106a-5p/GLIS3 Axis. Mol Ther Nucleic Acids, 2018, 11: 170–179.

25. Hu G, Zhao X, Wang C, *et al.* MicroRNA-145 attenuates TNF-α-driven cartilage matrix degradation in osteoarthritis via direct suppression of MKK4. Cell Death Dis, 2017, 8: e3140.

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References

26. Si HB, Zeng Y, Liu SY, et al. Intra-articular injection of microRNA-140 (miRNA-140) alleviates osteoarthritis (OA) progression by modulating extracellular matrix (ECM) homeostasis in rats. Osteoarthritis Cartilage, 2017, 25: 1698–1707.
27. Li X, Zhen Z, Tang G, Zheng C, Yang G. MiR-29a and MiR-140 protect chondrocytes against the anti-proliferation and cell matrix signaling changes by IL-1β. Mol Cells, 2016, 39: 103–110.

28. Zheng X, Zhao FC, Pang Y, et al. Downregulation of miR-221-3p contributes to IL-1β-induced cartilage degradation by directly targeting the SDF1/CXCR4 signaling pathway. J Mol Med (Berl), 2017, 95: 615–627.

29. Chen W, Sheng P, Huang Z, *et al.* MicroRNA-381 regulates chondrocyte hypertrophy by inhibiting histone Deacetylase 4 expression. Int J Mol Sci, 2016, 17: 1377.

30. Ji Q, Xu X, Xu Y, et al. miR-105/Runx2 axis mediates FGF2-induced ADAMTS expression in osteoarthritis cartilage. J Mol Med (Berl), 2016, 94: 681–694.
31. Li Z, Meng D, Li G, Xu J, Tian K, Li Y. Overexpression of microRNA-210 promotes chondrocyte proliferation and extracellular matrix deposition by

targeting HIF-3 α in osteoarthritis. Mol Med Rep, 2016, 13: 2769–2776. **32.** Song J, Kim D, Lee CH, Lee MS, Chun CH, Jin EJ. MicroRNA-488 regulates in target of categories of acteoarthritis. L Biamod

zinc transporter SLC39A8/ZIP8 during pathogenesis of osteoarthritis. J Biomed Sci, 2013, 20: 31.

33. Matsukawa T, Sakai T, Yonezawa T, *et al.* MicroRNA-125b regulates the expression of aggrecanase-1 (ADAMTS-4) in human osteoarthritic chondrocytes. Arthritis Res Ther, 2013, 15: R28.

34. Gao F, Peng C, Zheng C, Zhang S, Wu M. miRNA-101 promotes chondrogenic differentiation in rat bone marrow mesenchymal stem cells. Exp Ther Med, 2019, 17: 175–180.

35. Liu X, Liu L, Zhang H, *et al*. MiR-146b accelerates osteoarthritis progression by targeting alpha-2-macroglobulin. Aging, 2019, 11: 6014–6028.

36. Budd E, de Andrés MC, Sanchez-Elsner T, Oreffo R. MiR-146b is down-regulated during the chondrogenic differentiation of human bone marrow derived skeletal stem cells and up-regulated in osteoarthritis. Sci Rep, 2017, 7: 46704.
37. Cheleschi S, Tenti S, Mondanelli N, *et al.* MicroRNA-34a and MicroRNA-181a mediate Visfatin-induced apoptosis and oxidative stress via NF-κB pathway in human osteoarthritic chondrocytes. Cell, 2019, 8: 874.

38. Yang B, Ni J, Long H, Huang J, Yang C, Huang X. IL-1 β -induced miR-34a upregulation inhibits Cyr61 to modulate osteoarthritis chondrocyte proliferation through ADAMTS-4. J Cell Biochem, 2018, 119: 7959–7970.

39. De Palma A, Cheleschi S, Pascarelli NA, Giannotti S, Galeazzi M, Fioravanti A. Hydrostatic pressure as epigenetic modulator in chondrocyte cultures: a study on miRNA-155, miRNA-181a and miRNA-223 expression levels. J Biomech, 2018, 66: 165–169.

40. Zhai X, Meng R, Li H, *et al.* miR-181a modulates chondrocyte apoptosis by targeting Glycerol-3-phosphate dehydrogenase 1-like protein (GPD1L) in osteoarthritis. Med Sci Monit. 2017. 23: 1224–1231.

41. Wang P, Dong R, Wang B, et al. Genome-wide microRNA screening reveals miR-582-5p as a mesenchymal stem cell-specific microRNA in subchondral bone of the human knee joint. J Cell Physiol, 2019, 234: 21877–21888.

42. Woods S, Barter MJ, Elliott HR, et *al.* miR-324-5p is up regulated in endstage osteoarthritis and regulates Indian hedgehog signalling by differing mechanisms in human and mouse. Matrix Biol, 2019, 77: 87–100.

43. Wang XB, Zhao FC, Yi LH, et al. MicroRNA-21-5p as a novel therapeutic target for osteoarthritis. Rheumatology (Oxford), 2019, 58: 1485–1497.

44. Wang H, Zhang H, Sun Q, *et al.* Chondrocyte mTORC1 activation stimulates miR-483-5p via HDAC4 in osteoarthritis progression. J Cell Physiol, 2019, 234: 2730–2740.

45. Wang H, Zhang H, Sun Q, *et al.* Intra-articular delivery of Antago-miR-483-5p inhibits osteoarthritis by modulating Matrilin 3 and tissue inhibitor of metalloproteinase 2. Mol Ther, 2017, 25: 715–727.

46. Zhang W, Cheng P, Hu W, *et al.* Inhibition of microRNA-384-5p alleviates osteoarthritis through its effects on inhibiting apoptosis of cartilage cells via the NFr B signaling pathway by targeting SOX9. Cancer Gene Ther, 2018, 25: 326–338. **47.** Wang J, Chen L, Jin S, *et al.* Altered expression of microRNA-98 in IL-1β-induced cartilage degradation and its role in chondrocyte apoptosis. Mol Med Rep, 2017, 16: 3208–3216.

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48. Tu M, Li Y, Zeng C, et al. MicroRNA-127-5p regulates osteopontin expression and osteopontin-mediated proliferation of human chondrocytes. Sci Rep, 2016, 6: 25032.

49. Li L, Jia J, Liu X, *et al.* MicroRNA-16-5p controls development of osteoarthritis by targeting SMAD3 in chondrocytes. Curr Pharm des, 2015, 21: 5160–5167.

50. Dai L, Zhang X, Hu X, Zhou Ć, Ao Y. Silencing of microRNA-101 prevents IL- 1β -induced extracellular matrix degradation in chondrocytes. Arthritis Res Ther, 2012, 14: R268.

51. Li J, Huang J, Dai L, *et al.* miR-146a, an IL-1 β responsive miRNA, induces vascular endothelial growth factor and chondrocyte apoptosis by targeting Smad4. Arthritis Res Ther, 2012, 14: R75.

52. Li D, Ni S, Miao KS, Zhuang C. PI3K/Akt and caspase pathways mediate oxidative stress-induced chondrocyte apoptosis. Cell Stress Chaperones, 2019, 24: 195–202.

53. Musumeci G, Castrogiovanni P, Trovato FM, *et al.* Biomarkers of chondrocyte apoptosis and autophagy in osteoarthritis. Int J Mol Sci, 2015, 16: 20560–20575.

54. Guilak F, Meyer BC, Ratcliffe A, Mow VC. The effects of matrix compression on proteoglycan metabolism in articular cartilage explants. Osteoarthritis Cartilage, 1994, 2: 91–101.

55. Sun H, Peng G, Ning X, Wang J, Yang H, Deng J. Emerging roles of long noncoding RNA in chondrogenesis, osteogenesis, and osteoarthritis. Am J Transl Res, 2019, 11: 16–30.

56. Chen WK, Yu XH, Yang W, et al. IncRNAs: novel players in intervertebral disc degeneration and osteoarthritis. Cell Prolif, 2017, 1: 50.

57. Wang Y, Cao L, Wang Q, Huang J, Xu S. LncRNA FOXD2-AS1 induces chondrocyte proliferation through sponging miR-27a-3p in osteoarthritis. Artif Cells Nanomed Biotechnol, 2019, 47: 1241–1247.

58. Cao L, Wang Y, Wang Q, Huang J. LncRNA FOXD2-AS1 regulates chondrocyte proliferation in osteoarthritis by acting AS a sponge of miR-206 to modulate CCND1 expression. Biomed Pharmacother, 2018, 106: 1220–1226.

59. Li Q, Zhang Z, Guo S, Tang G, Lu W, Qi X. LncRNA ANCR is positively correlated with transforming growth factor β1 in patients with osteoarthritis. J Cell Biochem, 2019, 120: 14226–14232.

60. Huang J, Liu L, Yang J, Ding J, Xu X. IncRNA DILC is downregulated in osteoarthritis and regulates IL-6 expression in chondrocytes. J Cell Biochem, 2019, 120: 16019–16024.

61. Xiao Y, Bao Y, Tang L, Wang L. LncRNA MIR4435-2HG is downregulated in osteoarthritis and regulates chondrocyte cell proliferation and apoptosis. J Orthop Surg Res, 2019, 14: 247.

62. Lei J, Fu Y, Zhuang Y, Zhang K, Lu D. LncRNA SNHG1 alleviates IL-1βinduced osteoarthritis by inhibiting miR-16-5p-mediated p38 MAPK and NF-κB signaling pathways. Biosci Rep, 2019, 39: BSR20191523.

63. Shen H, Wang Y, Shi W, Sun G, Hong L, Zhang Y. LncRNA SNHG5/miR-26a/SOX2 signal axis enhances proliferation of chondrocyte in osteoarthritis. Acta Biochim Biophys Sin, 2018, 50: 191–198.

64. Chu P, Wang Q, Wang Z, Gao C. Long non-coding RNA highly up-regulated in liver cancer protects tumor necrosis factor-alpha-induced inflammatory injury by down-regulation of microRNA-101 in ATDC5 cells. Int Immunopharmacol, 2019, 72: 148–158.

65. Jiang M, Liu J, Luo T, Chen Q, Lu M, Meng D. LncRNA PACER is downregulated in osteoarthritis and regulates chondrocyte apoptosis and IncRNA HOTAIR expression. Biosci Rep, 2019, 39: BSR20190404.

66. Chen K, Zhu H, Zheng MQ, Dong QR. LncRNA MEG3 inhibits the degradation of the extracellular matrix of chondrocytes in osteoarthritis via targeting miR-93/TGFBR2 Axis. Cartilage, 2019: 1947603519855759.

67. Xu J, Xu Y. The IncRNA MEG3 downregulation leads to osteoarthritis progression via miR-16/SMAD7 axis. Cell Biosci, 2017, 7: 69.

68. Yang Q, Li X, Zhou Y, Fu W, Wang J, Wei Q. A LINC00341-mediated regulatory pathway supports chondrocyte survival and may prevent osteoarthritis progression. J Cell Biochem, 2019, 120: 10812–10820.

69. Ying H, Wang Y, Gao Z, Zhang Q. Long non-coding RNA activated by transforming growth factor beta alleviates lipopolysaccharide-induced inflammatory injury via regulating microRNA-223 in ATDC5 cells. Int Immunopharmacol, 2019, 69: 313–320.

70. Li X, Yu M, Chen L, *et al*. LncRNA PMS2L2 protects ATDC5 chondrocytes against lipopolysaccharide-induced inflammatory injury by sponging miR-203. Life Sci, 2019, 217: 283–292.

71. Zhang Y, Wang F, Chen G, He R, Yang L. LncRNA MALAT1 promotes osteoarthritis by modulating miR-150-5p/AKT3 axis. Cell Biosci, 2019, 9: 54.
72. Yang Z, Tang Y, Lu H, et al. Long non-coding RNA reprogramming (IncRNA-ROR) regulates cell apoptosis and autophagy in chondrocytes. J Cell Biochem, 2018, 119: 8432–8440.

73. Ye D, Jian W, Feng J, Liao X. Role of long noncoding RNA ZFAS1 in proliferation, apoptosis and migration of chondrocytes in osteoarthritis. Biomed Pharmacother, 2018, 104: 825–831.

74. Li X, Ren W, Xiao ZY, Wu LF, Wang H, Guo PY. GACAT3 promoted proliferation of osteoarthritis synoviocytes by IL-6/STAT3 signaling pathway. Eur Rev Med Pharmacol Sci, 2018, 22: 5114–5120.

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75. Zhang G, Wu Y, Xu D, Yan X. Long Noncoding RNA UFC1 promotes proliferation of chondrocyte in osteoarthritis by acting as a sponge for miR-34a. DNA Cell Biol, 2016, 35: 691–695.

76. Zhang L, Zhang P, Sun X, Zhou L, Zhao J. Long non-coding RNA DANCR regulates proliferation and apoptosis of chondrocytes in osteoarthritis via miR-216a-5p-JAK2-STAT3 axis. Biosci Rep, 2018, 38: BSR20181228.

77. Fan X, Yuan J, Xie J, et *al.* Long non-protein coding RNA DANCR functions as a competing endogenous RNA to regulate osteoarthritis progression via miR-577/SphK2 axis. Biochem Biophys Res Commun, 2018, 500: 658–664.

78. Li Y, Li Z, Li C, Zeng Y, Liu Y. Long noncoding RNA TM1P3 is involved in osteoarthritis by mediating chondrocyte extracellular matrix degradation. J Cell Biochem, 2019, 120: 12702–12712.

79. Li L, Zhang L, Zhang Y, et al. Inhibition of Long non-coding RNA CTD-2574D22.4 alleviates LPS-induced apoptosis and inflammatory injury of chondrocytes. Curr Pharm des, 2019, 25: 2969–2974.

80. Huang B, Yu H, Li Y, Zhang W, Liu X. Upregulation of long noncoding TNFSF10 contributes to osteoarthritis progression through the miR-376-3p/FGFR1 axis. J Cell Biochem, 2019, 120: 19610–19620.

81. Yang DW, Zhang X, Qian GB, Jiang MJ, Wang P, Wang KZ. Downregulation of long noncoding RNA LOC101928134 inhibits the synovial hyperplasia and cartilage destruction of osteoarthritis rats through the activation of the Janus kinase/signal transducers and activators of transcription signaling pathway by upregulating IFNA1. J Cell Physiol, 2019, 234: 10523–10534.

82. Huang T, Wang J, Zhou Y, Zhao Y, Hang D, Cao Y. LncRNA CASC2 is upregulated in osteoarthritis and participates in the regulation of IL-17 expression and chondrocyte proliferation and apoptosis. Biosci Rep, 2019, 39: BSR20182454.

83. Yu C, Shi D, Li Z, Wan G, Shi X. Long noncoding RNA CHRF exacerbates IL-6-induced inflammatory damages by downregulating microRNA-146a in ATDC5 cells. J Cell Physiol, 2019, 234: 21851–21859.

84. Park S, Lee M, Chun CH, Jin EJ. The IncRNA, Nespas, is associated with osteoarthritis progression and serves as a potential new prognostic biomarker. Cartilage, 2019, 10: 148–156.

85. Hu Y, Li S, Zou Y. Knockdown of LncRNA H19 relieves LPS-induced damage by modulating miR-130a in osteoarthritis. Yonsei Med J, 2019, 60: 381–388.
86. Liu G, Wang Y, Zhang M, Zhang Q. Long non-coding RNA THRIL promotes LPS-induced inflammatory injury by down-regulating microRNA-125b in ATDC5 cells. Int Immunopharmacol, 2019, 66: 354–361.

87. Tang LP, Ding JB, Liu ZH, Zhou GJ. LncRNA TUG1 promotes osteoarthritisinduced degradation of chondrocyte extracellular matrix via miR-195/MMP-13 axis. Eur Rev Med Pharmacol Sci, 2018, 22: 8574–8581.

88. Han W, Liu J. LncRNA-p21 inhibited the proliferation of osteosarcoma cells via the miR-130b/PTEN/AKT signaling pathway. Biomed Pharmacother, 2018, 97: 911–918.

89. Lu Z, Luo M, Huang Y. IncRNA-CIR regulates cell apoptosis of chondrocytes in osteoarthritis. J Cell Biochem, 2018, 120: 5.
90. Wang CL, Peng JP, Chen XD. LncRNA-CIR promotes articular cartilage

90. Wang CL, Peng JP, Chen XD. LncRNA-CIR promotes articular cartilage degeneration in osteoarthritis by regulating autophagy. Biochem Biophys Res Commun, 2018, 505: 692–698.

91. Li YF, Li SH, Liu Y, Luo YT. Long Noncoding RNA CIR promotes chondrocyte extracellular matrix degradation in osteoarthritis by acting as a sponge for Mir-27b. Cell Physiol Biochem, 2017, 43: 602–610.

92. Zhao Y, Zhao J, Guo X, She J, Liu Y. Long non-coding RNA PVT1, a molecular sponge for miR-149, contributes aberrant metabolic dysfunction and inflammation in IL-1 β -simulated osteoarthritic chondrocytes. Biosci Rep, 2018, 38: BSR20180576.

93. Li Y, Li S, Luo Y, Liu Y, Yu N. LncRNA PVT1 regulates chondrocyte apoptosis in osteoarthritis by acting as a sponge for miR-488-3p. DNA Cell Biol, 2017, 36: 571–580.

94. Li L, Lv G, Wang B, Kuang L. The role of IncRNA XIST/miR-211 axis in modulating the proliferation and apoptosis of osteoarthritis chondrocytes through CXCR4 and MAPK signaling. Biochem Biophys Res Commun, 2018, 503: 2555–2562.

95. Li XF, Wang ZQ, Li LY, Zhao GQ, Yu SN. Downregulation of the long noncoding RNA MBNL1-AS1 protects sevoflurane-pretreated mice against ischemia-reperfusion injury by targeting KCNMA1. Exp Mol Med, 2018, 50: 115.

96. Hu J, Wang Z, Shan Y, Pan Y, Ma J, Jia L. Long non-coding RNA HOTAIR promotes osteoarthritis progression via miR-17-5p/FUT2/ β -catenin axis. Cell Death Dis, 2018, 9: 711.

97. Zhu JK, He TD, Wei ZX, Wang YM. LncRNA FAS-AS1 promotes the degradation of extracellular matrix of cartilage in osteoarthritis. Eur Rev Med Pharmacol Sci, 2018, 22: 2966–2972.

98. Liu Q, Hu X, Zhang X, *et al.* The TMSB4 Pseudogene LncRNA functions as a competing endogenous RNA to promote cartilage degradation in human osteoarthritis. Mol Ther, 2016, 24: 1726–1733.

99. Kang Y, Song J, Kim D, *et al.* PCGEM1 stimulates proliferation of osteoarthritic synoviocytes by acting as a sponge for miR-770. J Orthop Res, 2016, 34: 412–418.

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100. Goldring MB. Chondrogenesis, chondrocyte differentiation, and articular cartilage metabolism in health and osteoarthritis. Ther Adv Musculoskeletal Dis, 2012, 4: 269–285.

101. Domenis R, Zanutel R, Caponnetto F, *et al.* Characterization of the Proinflammatory profile of synovial fluid-derived Exosomes of patients with osteoarthritis. Mediators Inflamm, 2017, 2017: 4814987.

102. Kato T, Miyaki S, Ishitobi H, *et al.* Exosomes from IL-1 β stimulated synovial fibroblasts induce osteoarthritic changes in articular chondrocytes. Arthritis Res Ther, 2014, 16: R163.

103. Qi H, Liu DP, Xiao DW, Tian DC, Su YW, Jin SF. Exosomes derived from mesenchymal stem cells inhibit mitochondrial dysfunction-induced apoptosis of chondrocytes via p38, ERK, and Akt pathways. In Vitro Cell Dev Biol Anim, 2019, 55: 203–210.

104. Cosenza S, Ruiz M, Toupet K, Jorgensen C, Noël D. Mesenchymal stem cells derived exosomes and microparticles protect cartilage and bone from degradation in osteoarthritis. Sci Rep, 2017, 7: 16214.

105. Wang Y, Yu D, Liu Z, *et al.* Exosomes from embryonic mesenchymal stem cells alleviate osteoarthritis through balancing synthesis and degradation of cartilage extracellular matrix. Stem Cell Res Ther, 2017, 8: 189.

106. Tofiño-Vian M, Guillén MI, Pérez Del Caz MD, Castejón MA, Alcaraz MJ. Extracellular vesicles from adipose-derived Mesenchymal stem cells Downregulate senescence features in osteoarthritic osteoblasts. Oxid Med Cell Longev, 2017, 2017: 7197598.

107. Wu J, Kuang L, Chen C, et al. miR-100-5p-abundant exosomes derived from infrapatellar fat pad MSCs protect articular cartilage and ameliorate gait abnormalities via inhibition of mTOR in osteoarthritis. Biomaterials, 2019, 206: 87–100.

108. Sun H, Hu S, Zhang Z, Lun J, Liao W, Zhang Z. Expression of exosomal microRNAs during chondrogenic differentiation of human bone mesenchymal stem cells. J Cell Biochem, 2019, 120: 171–181.

109. Mao G, Zhang Z, Hu S, *et al.* Exosomes derived from miR-92a-3p-overexpressing human mesenchymal stem cells enhance chondrogenesis and suppress cartilage degradation via targeting WNT5A. Stem Cell Res Ther, 2018, 9: 247.

110. Tao SC, Yuan T, Zhang YL, Yin WJ, Guo SC, Zhang CQ. Exosomes derived from miR-140-5p-overexpressing human synovial mesenchymal stem cells enhance cartilage tissue regeneration and prevent osteoarthritis of the knee in a rat model. Theranostics, 2017, 7: 180–195.

111. Liu Y, Zou R, Wang Z, Wen C, Zhang F, Lin F. Exosomal KLF3-AS1 from hMSCs promoted cartilage repair and chondrocyte proliferation in osteoarthritis. Biochem J, 2018, 475: 3629–3638.