Revised: 1 April 2021

DOI: 10.1002/jmv.27028

RESEARCH ARTICLE

JOURNAL OF WILEY

Alteration of type I interferon response is associated with subclinical atherosclerosis in virologically suppressed HIV-1-infected male patients

Letizia Santinelli ¹ 💿 📔 Gabriella De Girolamo ² 💿 📔 Cristian Borrazzo ² 💿 📔
Paolo Vassalini ² 💿 Claudia Pinacchio ² 💿 📔 Eugenio Nelson Cavallari ² 💿 📔
Maura Statzu ¹ 💿 Federica Frasca ¹ 💿 Mirko Scordio ¹ 💿 Camilla Bitossi ¹ 💿
Agnese Viscido ¹ 💿 Giancarlo Ceccarelli ² 💿 Massimo Mancone ³ 💿
Claudio Maria Mastroianni ² 💿 📔 Guido Antonelli ^{1,4} 💿 📔 Gabriella d'Ettorre ² 💿 📔
Carolina Scagnolari ¹ 💿

¹Department of Molecular Medicine, Laboratory of Virology, Affiliated to Istituto Pasteur Italia, Sapienza University of Rome, Viale di Porta Tiburtina, Rome, Italy

²Department of Public Health and Infectious Diseases, Sapienza University of Rome, Viale del Policlinico, Rome, Italy

³Department of Cardiovascular, Respiratory, Nephrology, Anaesthesiology and Geriatric Sciences, Sapienza University of Rome, Viale del Policlinico, Rome, Italy

⁴Microbiology and Virology Unit, Sapienza University Hospital "Policlinico Umberto I", Rome, Italy

Correspondence

Carolina Scagnolari, Department of Molecular Medicine, Laboratory of Virology, Affiliated to Institute Pasteur Italia, Sapienza University of Rome, Viale di Porta Tiburtina 28, 00185 Rome, Italy. Email: carolina.scagnolari@uniroma1.it

Funding information

Sapienza Università di Roma; Finanziamenti di ateneo per la Ricerca Scientifica, Grant/Award Number: 2019, RP11916B6EC60AF3; Ricerca Ateneo Sapienza, Grant/Award Numbers: Progetti Medi, 2018 DDA N. 801/2019; Ricerche Universitarie, Grant/Award Number: 2017 RM11715C586062AF; Gilead Sciences Gilead fellowship Programme 2016 (Title: Analysis of immunogenetic patterns, viral burden and

Abstract

Given human immunodeficiency virus-1 (HIV-1)-infected patients have alterations in the type I interferon (IFN-I) pathway and are also at elevated risk of atherosclerosis, we evaluated IFN-I response and subclinical cardiovascular disease (CVD) association in HIV-1-infected patients. Transcript levels of IFN- α/β and IFN-stimulated gene 56 (ISG56) were evaluated by RT/real-time PCR in peripheral blood mononuclear cells collected from asymptomatic HIV-1-positive male patients at high risk of developing CVD (n = 34) and healthy subjects (n = 21). Stenosis degree (\geq or <50%), calcium volume score, calcium Agatston score, and myocardial extracellular volume were examined by coronary computerized tomography scan. Carotid intimamedia thickness (cIMT), Framingham risk score, atherosclerotic cardiovascular disease (ASCVD) score, and risk score developed by data collection on adverse effects of anti-HIV drugs (D:A:D) were also measured. Increased IFN- α , IFN- β , and ISG56 levels were observed in all HIV-1-infected males compared to healthy controls (p < .001 for all genes analyzed). HIV-1-infected patients with a stenosis degree \geq 50% showed a higher Framingham risk score (p = .019), which was correlated with IFN- β and ISG56 levels. HIV-1-infected males with enhanced IFN-I levels and stenosis displayed a higher ASCVD calculated risk (p = .011) and D:A:D score (p = .004). Also, there was a trend toward higher IFN- α and ISG56 mRNA levels in HIV-1positive patients with an increased cIMT (p > .05). Dysregulation of IFN-I response might participate in the pathogenesis of HIV-1-associated CVD.

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

 $\ensuremath{\mathbb{C}}$ 2021 The Authors. Journal of Medical Virology published by Wiley Periodicals LLC

cardiovascular diseases: a new integrative model for the management of HIV-1 patients suffering from cardiovascular damage)

KEYWORDS

atherosclerosis, cardiovascular disease, human immunodeficiency virus, IFN-stimulated genes, innate immunity, interferon

1 | INTRODUCTION

Since the introduction of combined antiretroviral therapy (cART), the life quality and survival of people living with human immunodeficiency virus-1 (HIV-1) have considerably enhanced.¹ Nevertheless, as a result of the harmful effects of HIV-1, exposure to prolonged cART and accelerated aging, cardiovascular disease (CVD) increasingly afflicts people living with HIV-1.^{2,3} The variety of CVD in HIV-1 is not only restricted to enhanced atherosclerosis, but includes pulmonary hypertension, ventricular dysfunction related to myocarditis, complex cerebrovascular disease, pericardial pathology, endocarditis, and cardiac tumors.^{4,5} The pathogenesis behind the cardiovascular HIV-1-associated complications is multifactorial, and involves distinct CVD risk parameters, but also viral and host factors associated with immunological and metabolic dysfunction⁶ observed during HIV-1 infection. As a result of these observations, great interest has been directed toward the role of type I interferon (IFN-I) in atherosclerosis.⁷⁻¹³

As HIV-1-infected patients have an enhanced activation of IFN-I signalling¹⁴⁻¹⁷ and are at elevated risk of developing atherosclerosis,^{18,19} we hypothesized that persistent upregulation of IFN-I and/or IFN-related pathways might be involved in the development and progression of atherosclerosis during HIV-1 infection. Therefore, we evaluated the gene expression level of IFN- α , IFN- β , and IFN-stimulated gene 56 (ISG56),²⁰ a well-established marker of IFN-I activation, in peripheral blood mononuclear cells (PBMCs) collected from HIV-1-infected male patients at high risk for developing CVD, examining whether alterations in IFN-I/ISG56 mRNA levels might be associated with progression of atherosclerosis.

2 | MATERIALS AND METHODS

2.1 | Study population

Thirty-four HIV-1-infected male patients successfully treated with cART, attending the Division of Infectious Diseases, at Department of Public Health and Infectious Diseases, "Sapienza" University of Rome (Italy) were enrolled from 2018 to 2019. Ageand gender-matched healthy subjects with no history of previous severe CVD and metabolic syndrome were included as a control group (n = 21). For each HIV-1-infected patient, medical and family history, body mass index and smoke status, time from HIV-1 diagnosis, length of cART treatment, and CD4 + T cell count were collected. To evaluate the risk of CVD, the following

cardiovascular parameters were measured: Framingham 10 years risk score, atherosclerotic cardiovascular disease (ASCVD) 10 years score, data collection on adverse effects of anti-HIV Drugs (D:A:D) 10 years estimated risk score, extracellular volume (ECV) and carotid artery intima-media thickness (cIMT). Inclusion criteria were as follows: (1) men at least 18 years old, (2) being in cART with HIV-1 RNA < 37 copies/ml, (3) CD4 + T cell counts >400 cells/mm³, (4) absence of metabolic syndrome. The study was approved by the institutional review board (Department of Public Health and Infectious Diseases, Sapienza, University of Rome) and the Ethics Committee (Sapienza, University of Rome), and all study participants signed written informed consent.

2.2 | Multi-detector computed tomography (MDTC) protocol and coronary angiography (CA)

All HIV-1-infected patients were tested for the severity of CVD using a low-dose prospectively ECG-triggered computerized tomography (CT) coronary angiography protocol, with a 64-slice multidetector MDCT scanner (Somatom Definition Siemens medical Solution, Forchheimen), as previously described.²¹ Calcium score was quantified using the Agatston Calcium Score method²² from 3 mm nonoverlapping sections by using a semiautomated software (calcium scoring CT; Siemens Medical Solutions). The luminal stenosis degree was classified as mild (Grade I: 30%-49%), moderate (grade II: 50%-69%), severe (Grade III: 70%-99%), or coronary occlusion (Grade IV: 100%). A threshold of 50% luminal narrowing in any coronary segment greater than 1.5 mm in diameter was adopted to define clinically significant coronary stenosis.²³ All HIV-1-infected patients with coronary lesions ≥ grade II at MDCT analysis were considered for coronary angiography. This evaluation was performed using the standard technique with angiograms that were examined by an experienced blinded operator to the MDTC results. The American Heart Association (AHA) segmentation model²⁴ was applied and the degree of stenosis was quantified (quantitative coronary angiography, QCA) (Allura Xper FD 10; Philips Medical Systems). Significant stenosis was defined as a reduction in diameter ≥50%.

2.3 | CT image analysis

Pre- and post-contrast Hounsfield units (HU) were measured on a Picture Archiving and Communication System. The regions of interest (ROIs) were drawn first on the equilibrium phase image EY-MEDICAL VIROLOGY

SANTINELLI ET AL.

at the myocardial septum and within the left ventricular chamber; the mean area was 3 cm^2 (range, $1.5-5 \text{ cm}^2$). Mean attenuation at the ROI was recorded in HU and Myocardial ECV fraction was calculated using the following equation:

$$\mathsf{ECV}(\%) = \lambda \cdot (1 - H) \cdot 100\%,$$

where the contrast agent partition coefficient (λ) represents the ratio of the change in the blood and myocardial attenuation (Δ HU) and *H* is the haematocrit level. The change in attenuation (Δ HU) was determined with the following equation: Δ HU = HU_{post} – HU_{pre}, where HU_{post} and HU_{pre} are attenuations after and before administration of iodinated contrast material, respectively.

2.4 | Carotid intima-media thickness measurement (c-IMT)

c-IMT measurement was performed using a B-mode ultrasound recording with a 7- to 14-MZ array probe (ESAOTE-technology). c-IMT was measured at a distance of at least 5 mm below the distal end of the common carotid artery. The mean value (expressed as mm) of 3 measurements was calculated for each HIV-1-positive patient and used as the final measurement of internal c-IMT. Normal c-IMT was defined as IMT < 0.9 mm and pathological c-IMT was considered as IMT \ge 0.9 mm. To avoid inter-operator differences, all the measurements of c-IMT were performed by a single operator.

2.5 | Peripheral blood mononuclear cells isolation

Fresh peripheral blood samples (20 ml) were collected by venepuncture in Vacutainer tubes containing EDTA (BD Biosciences) from HIV-1-infected men and healthy individuals, and processed by Ficoll-Hypaque density gradient centrifugation (Lympholyte, Cedarlane Labs) to obtain PBMCs. PBMCs were washed twice in phosphate-buffered saline and stored at -80° C as dried pellets for RNA extraction.

2.6 | Real-time RT-PCR assays for IFN- α , IFN- β , and ISG56 mRNA expression

Quantitative real-time PCR for the analysis of IFN- α , IFN- β , and ISG56 mRNAs levels was carried out with the LightCycler480 instrument (Roche), as previously described.^{14,25} Briefly, total RNA was extracted from PBMCs collected from HIV-1-infected patients and healthy individuals using a commercial RNA purification assay (Norgen Biotek Corporation) and reverse-transcribed using the High-Capacity cDNA Reverse Transcription Kit (Applied Biosystems), according to the manufacturer's protocol. All primers and probes were added to the Probes Master Mix (Roche, Basel, Switzerland) at 500 and 250 nm respectively, in a final volume of 20 μ l. The β -glucuronidase housekeeping gene was considered as an internal control. Gene expression values were calculated by the comparative C_t method. The primers and probe sequences

used for IFN- α , (Hs. PT.58.24294810.g) and IFN- β (Hs. PT.58.39481063.g) were purchased from Integrated DNA Technologies (IDT). The primers and probe sequences used for ISG56 were the following: forward, 5'- TGAGAAGCTCTAGCCAACAACATGTC-3'; reverse, 5'-GAGCTTTATCCACAGAGCCTTTTC-3'; probe 5'-(6FAM) TATG TCTTTCGATATGCAGCCA-AGTTTTACCG (TAM)-3'.

2.7 | HIV-1 RNA measurement and CD4 + T lymphocyte count

HIV-1 viral load was determined by a Versant HIV-1 RNA kPCR assay (Siemens Healthineers) which has a detection limit of 37 copies/ml. Absolute CD4+T lymphocyte count was performed by FACScalibur flow cytometer (Becton Dickinson).

2.8 | Statistical analysis

Patients data were expressed as median/IQR (interguartile range) (age, CD4 + T cell count at enrolment, CD4 + T cell count at Nadir, years from HIV-1 diagnosis, years on ART, Framingham risk score, ASCVD score, D:A:D score, calcium score, calcium score Agatston, ECV, luminal stenosis) or as frequency (percentage) (class of anti-HIV-1 drug, smokers, opportunistic infections, diabetes, family history of CVD, previous CVD). The demographic and clinical characteristics of HIV-1-infected patients and healthy controls were compared using Student's t and χ^2 tests. Differences in the clinical characteristics (Framingham risk score, ASCVD risk score, D:A:D score, Calcium volume/cm³ score, Calcium score Agatston, Luminal stenosis, ECV range) between HIV-1-positive patients with a cIMT ≥ 0.9 mm and <0.9 mm were evaluated using the Mann-Whitney U test. Differences in IFN-α, IFN-β, and ISG56 mRNA levels between HIV-1-infected patients and healthy controls were analyzed using the Mann-Whitney U test. The same test was applied to evaluate any differences in the transcript level of IFN-I genes between HIV-1-infected patients with coronary stenosis ≥50% and <50%, and between HIV-1-infected patients with cIMT \ge 0.9 mm and <0.9 mm. Spearman's rho coefficient was calculated to assess the correlation between IFN- α , IFN- β , ISG56, and the Framingham score in HIV-1-infected patients. Statistical analyses were performed with SPSS v.25.0 for Windows: a p value less than .05 was considered statistically significant.

3 | RESULTS

3.1 | Patients characteristics

Overall, the study population included 34 cART-treated HIV-1-infected males at high risk of developing CVD and 21 healthy subjects. The demographic and clinical features of the HIV-1-positive men and healthy controls are reported in Table 1. In particular, HIV-1-infected males, with

MEDICAL VIROLOGY -WILEY

TABLE 1 Demographic and clinical characteristics of HIV-1-positive males and healthy controls

Item ^a	Healthy male controls, <i>n</i> = 21	HIV-1-infected men, <i>n</i> = 34	Luminal stenosis ≥50%, n = 11 (A)	<50%, n = 23 (B)	A versus B p value
Age (years)	55 (42.5-60.5)	47.5 (40.2–51.5)	48 (42-54.2)	45 (40-48)	.082
HIV-1 RNA (copies/ml) ^b	NA ^c	<37	<37	<37	1.000
CD4 + T at enrolment (cells/mm ³)	NA	448 (346-532.2)	490 (326.5-655)	446 (376-487.5)	.236
CD4 + T nadir (cells/mm ³)	NA	185.5 (108.2-292.5)	140 (53–229.2)	247 (155-391)	.023
Years from HIV-1 diagnosis	NA	9.5 (5-10)	10 (5-10.2)	7(3–10)	.245
Years on cART	NA	7.5 (5-10)	8 (5-10)	6 (0-10)	.283
Anti-HIV-1 drug class, n (%)					
NNRTI	NA	15 (44.1)	3 (27.2)	12 (52.1)	.177
PI	NA	22 (64.7)	10 (90.9)	12 (52.1)	.033
ABC	NA	15 (44.1)	8 (72.7)	7 (30.4)	.022
Opportunistic infections, n (%)	NA	14 (41.1)	5 (45.5)	9 (39)	.021
Diabetes, n (%)	0 (0)	0 (0)	0 (0)	0 (0)	1.000
Family history of CVD, n (%)	3 (15)	2 (13)	2 (13)	3 (15)	.340
Previous CVD, n (%)	O (O)	0 (0)	0 (0)	0 (0)	1.000
Framingham risk score (%) ^d	NA	3 (1-6)	4 (1-9)	3 (1-3)	.019
ASCVD risk score (%) ^d	NA	4 (2.8–5.9)	5.2 (3.8-8)	3 (2.8–3.9)	.011
D:A:D risk score (%) ^d	NA	12.2 (7.4–20.9)	18 (8.9–26.7)	8 (4-12)	.004
Calcium volume score (cm ³)	NA	35.4 (10-146.95)	35.4 (10-74.3)	6.4 (0-13.6)	<.001
Calcium score Agatston (HU)	NA	49.2 (11-245.3)	49.2 (11-102.3)	7.0 (0-15)	<.001
ECV (%)	NA	30.5 (28.4-34.2)	32.3 (28.4-36.2)	29.4 (28.4-31.5)	.518

Abbreviations: ABC, abacavir; ASCVD, atherosclerotic cardiovascular disease; cART, combined antiretroviral therapy; CVD, cardiovascular disease; ECV, extracellular volume; HIV-1, human immunodeficiency virus-1; NNRTI, non-nucleoside reverse transcriptase inhibitor; PI, protease inhibitor.

^aData are expressed as median (IQR) or percentages. No statistically significant differences were recorded in demographic characteristics and family history of CVD between HIV-1-infected patients and heathy controls, p > .05 using Student's t and χ^2 tests.

^bHIV-1 viral load was determined by versant HIV-1 RNA kPCR assay (Siemens Healthcare Diagnostic), which has a detection limit of 37 copies/ml. ^cNA, not applicable.

^dFramingham 10 years risk score, ASCVD 10 years risk score and D:A:D 10 years risk score are expressed as median (IQR) of percentage values.



FIGURE 1 IFN- α , IFN- β , and ISG56 gene expression levels in HIV-1-infected patients and healthy controls. The mRNA levels of (A) IFN- α , (B) IFN- β , and (C) ISG56 were evaluated in PBMCs of 34 HIV-1-infected patients and 21 healthy controls; *p < .001 for all genes analyzed, determined by Mann–Whitney *U* test. IFN- α , interferon- α ; ISG56, IFN-stimulated gene 56; HIV-1, human immunodeficiency virus-1

EY-MEDICAL VIROLOGY

a median age of 47.5 years (IQR, 40.2–51.5 years), had a median CD4 + T cell count of 448 cells/mm³ (IQR, 346–532.2 cells/mm³) and they had received cART for a median of 7.5 years (IQR, 5–10 years). Eleven out of 34 HIV-1-infected patients (32.4%) showed a luminal stenosis \geq 50% while 67.6% (*n* = 23) had a luminal stenosis <50% (Table 1). HIV-1-infected patients with luminal stenosis \geq 50% had a higher ASCVD calculated risk score, calcium volume and calcium Agatston scores than those with stenosis degree <50% (Table 1, *p* < .001 for all cardiovascular parameters). Opportunistic infections were detected in 45.5% (5 of 11) of HIV-1-infected patients with a stenosis level \geq 50% compared to 39.1% (9 of 23) in those with a lower cardiovascular risk (Table 1, *p* = .002). The analysis of patients' smoking history showed that no one had never smoked.

3.2 | Luminal stenosis, CV risk, and IFN-I response in HIV-1-infected patients

As IFN-I played a complex and controversial role in HIV-1 immunopathogenesis, 16,26,27 we measured the gene expression levels of IFN-α, IFN-β, and ISG56 in PBMCs of HIV-1-infected patients and healthy subjects. Results indicated that HIV-1-infected men have an increased amount of IFN-α, IFN-β, and ISG56 mRNA in PBMCs compared to healthy controls (Figure 1A-C; p < .001 for all genes analyzed). In particular, an average of 80-fold increase in IFN-α, IFN-β, and ISG56 levels were observed in HIV-1-infected males (Figure 1A-C). As an elevated IFN-I response has been related to vascular damage and CVD development,²⁸ we evaluated whether the alteration in IFN-I response might be associated with the presence of a subclinical atherosclerosis in HIV-1-infected men. We found that ISG56 mRNA levels were higher in HIV-1-infected patients with a luminal stenosis ≥50% than in those with a luminal stenosis <50% (Figure 2C, p = .017). A trend toward an increase in the expression of IFN-α-mRNA was also observed in those patients with a luminal stenosis ≥50%, however no statistically significant differences were recorded for both IFN- α and IFN- β (Figure 2A,B, p > .05). Of note, HIV-1-infected patients with a stenosis ≥50% showed higher **TABLE 2** Correlation between IFN- α , IFN- β , ISG56 and the Framingham 10 years risk score in HIV-1-infected men, stratified according to the grade of their luminal stenosis

	Framingham 10 years risk Luminal stenosis ≥50%		score Luminal stenosis <50%	
IFN-α mRNA	r = .587	p = .126	r =238	p = .31- 3
IFN-β mRNA	r = .707	p = .048	<i>r</i> = .110	p = .62- 0
ISG56 mRNA	r = .731	p = .040	r =264	p = .23- 6

Note: Spearman's rho coefficient was used to assess the correlation between IFN- α , IFN- β , ISG56, and the Framingham 10 years risk score. Significant correlations are highlighted in bold.

Abbreviations: HIV-1, human immunodeficiency virus-1; IFN-α, interferon-α; ISG56, IFN-stimulated gene 56.

Framingham risk score (Table 1, p = .019). Levels of this cardiological parameter were positively correlated with those of IFN- β and ISG56 but not of IFN- α (Table 2).

As the Framingham risk score has been associated with an underestimation of the CVD risk in HIV-1-infected subjects with a longer duration of cART,²⁹ we compared the IFN-I and ISG56 levels in HIV-1-infected patients with a luminal stenosis <50% and those with stenosis \geq 50% considering D:A:D equation. Results indicated that HIV-1-positive patients with a stenosis level lower than <50% and decreased IFN- α , IFN- β , and ISG56 mRNA levels (Figure 2A-C) had a lower risk of CVD compared to those with a higher stenosis degree and IFN-I response (Table 1, *p* = .004; Figure 2A-C).

3.3 | cIMT level and IFN-I expression relationship in the development of atherosclerosis

(C) (B) (A) relative to B-glucuronidase 50 80 30 relative to **B**-glucu 40 60 20 30 40 20 IFN-a mRNA expression-10 d 20 10 ISG56 mRNA (IFN-B mRNA C 0 0 Luminal Stenosis Luminal Stenosis Luminal Stenosis Luminal Stenosis Luminal Stenosis Luminal Stenosis >50% <50% >50% <50%

FIGURE 2 IFN- α , IFN- β , and ISG56 gene expression levels in HIV-1-infected patients with a stenosis degree \geq 50% and <50%. The mRNA levels of (A) IFN- α , (B) IFN- β , and (C) ISG56 were evaluated in PBMCs of HIV-1-infected patients with a stenosis degree \geq 50% (*n* = 23) and <50% (*n* = 11); **p* < .05, determined by Mann–Whitney *U* test. IFN- α , interferon- α ; ISG56, IFN-stimulated gene 56; HIV-1, human immunodeficiency virus-1; mRNA, messenger RNA; PBMC, peripheral blood mononuclear cell

cIMT is considered a surrogate measure of subclinical atherosclerosis that can predict CVD events among the general population. $^{\rm 30}$ Thus, we

JOURNAL O WILEY MEDICAL VIROLOGY

4935

TABLE 3 Comparison of CV risk
factors in HIV-1-infected males stratified
according to the cIMT \geq 0.9 mm
or <0.9 mm

I

Item ^a	cIMT ≥ 0.9 mm, <i>n</i> = 19	cIMT <0.9 mm, <i>n</i> = 15	p value ^b
Framingham risk score (%) ^c	5 (2-9)	1 (1-3)	<.001
ASCVD risk score (%) ^c	5.1 (3.6-6.4)	2.9 (2.3-3.5)	.003
D:A:D risk score (%) ^c	18 (10-22)	4 (3-12)	.001
Calcium volume score (cm ³)	3.3 (0-35.4)	0	<.001
Calcium score agatston (HU)	4.9 (0-49.2)	0	<.001
Luminal stenosis (%)	50 (40-58)	25 (20-30)	<.001
ECV (%)	33.2 (28.67-35.7)	28.66 (26.8-30.4)	.008

Abbreviations: ASCVD, atherosclerotic cardiovascular disease; cIMT, carotid intima-media thickness; CV, cardiovascular; ECV, extracellular volume.

^aData are expressed as median (IQR) or percentage.

^bDifferences in the clinical characteristics between HIV-1-positive patients with a cIMT ≥ 0.9 mm and <0.9 mm were evaluated using the Mann-Whitney U test.

^cFramingham 10 years risk score, ASCVD 10 years risk score and D:A:D 10 years risk score are expressed as median (IQR) of percentage values.

examined whether the enhanced levels of IFN-I/ISG56 and luminal stenosis found in HIV-1-infected men might be linked to a different cIMT degree. Nineteen (55.9%) HIV-1-infected males showed a cIMT \geq 0.9 mm, a value directly associated to CVD development ³¹ (Table 3). This group of HIV-1-infected patients had higher calcium volume score, calcium Agatston score, and ECV compared to those with a cIMT <0.09 mm. (Table 3, p < .001). Furthermore, HIV-1-infected males with an increased cIMT showed a trend toward an upregulation of ISG56 and a deregulation of IFN- α mRNA expression (Figure 3A,C; p > .05), compared to those with a cIMT <0.9 mm.

DISCUSSION 4

CVD represent one of the main causes of morbidity in people living with HIV-1. Here, we evaluated whether IFN-I response can be associated to subclinical atherosclerosis during HIV-1 infection. As expected, IFN- α , IFN- β , and ISG56 transcripts were increased in PBMCs of HIV-1-infected patients compared to healthy controls.^{14,16} To understand whether an upregulation in the IFN-I signature might promote the development of cardiovascular damage, we examined the relationship between levels of IFN- α/β , ISG56 and those of different cardiovascular parameters (e.g., coronary luminal stenosis, Framingham risk score, ASCVD risk score, calcium Agatston score, D:A:D risk score, cIMT and ECV). Despite the low Framingham risk score, we found a deep coronary obstruction in about 32% of HIV-1infected patients, using a cut-off value of 50% for defining significant stenosis, $^{32-34}$ which was correlated to an increased IFN- β and ISG56 expression in PBMCs. Interestingly, it has been reported that HIV-1infected patients with a low Framingham risk score are at high risk of developing subclinical carotid atherosclerosis.^{21,35} The presence of opportunistic infections have been also associated with an increased risk of developing CVD in patients with luminal stenosis ≥50%.³⁶ Beyond its status as an independent risk factor, smoking increases



FIGURE 3 IFN- α . IFN- β , and ISG56 gene expression levels in HIV-1-infected patients with a cIMT \geq 0.9 mm and <0.9 mm. The mRNA levels of (A) IFN- α , (B) IFN- β , and (C) ISG56 were evaluated in PBMCs of HIV-1-infected patients with a cIMT \geq 0.9 mm (*n* = 19) and <0.9 mm (n = 15); *p < .05, determined by Mann-Whitney U test. cIMT, carotid intima-media thickness; IFN-α, interferon-α; ISG56, IFN-stimulated gene 56; HIV-1, human immunodeficiency virus-1; PBMC, peripheral blood mononuclear cell

EY-MEDICAL VIROLOGY

the risk of CVD,⁶ but the analysis of smoking history showed that noone of HIV-1-infected patients had ever smoked.

More recently, IFN-I have been shown to be a crucial modulators of atherosclerosis disease: the persistent expression of IFN-I can promote and sustain endothelial dysfunction, which in turn contribute to atherosclerosis development in patients suffering from SLE.³⁷ Specifically, IFN-I appears to be involved in atherosclerosis development through the following mechanisms: (i) increase in macrophage recruitment^{8,28}; (ii) foam cell formation⁹; (iii) disruption of vascular repair³⁸ and (iv) plaque progression.³⁷ The absence of IFN-I signalling can also counteract the formation of heart lesions and the recruitment of macrophages to arteries in a mouse model of atherosclerosis.³⁷ Hence, the enhanced levels of IFN-I and ISG56 observed in HIV-1-infected men might be caused by an excessive secretion of these cytokines by activated macrophages, given these cells are the major contributors of the atherosclerotic inflammatory response.^{39,40}

On the other hand, the upregulation of IFN-I response might favour macrophage attraction toward the atherosclerotic plaque.⁸ Indeed, blocking IFN-I signalling has been shown to decrease the macrophage accumulation at atherosclerotic plaque level.^{7,40} Accordingly, it is likely that HIV-1-associated systemic inflammation might favor monocytes migration across the vascular endothelium; moreover, HIV-1 infection can reduce reverse trans endothelial migration of macrophages present at the atherosclerotic level, promoting the persistence of macrophages within the plaques and increasing the risk of CVD expansion.⁴¹ In this complex scenario, IFN-I response has been shown to exhibit strong pro- and antiatherogenic properties.^{8–12}

Due to the increasing evidence of subclinical cardiovascular atherosclerosis events in young HIV-1-infected males compared to uninfected controls,⁴² our results also highlighted the importance of performing coronary CT scan in asymptomatic HIV-1-infected subjects for CVD. Although HIV-1-infected patients with luminal stenosis < 50% and cIMT < 0.9 mm might be considered as a control subgroup with no CVD, it could be more interesting to include a group of healthy donors with subclinical CVD to give more strength to the role played by IFN-I response as potential predictor of increased risk of CVD in asymptomatic HIV-1-positive individuals; nevertheless, we are highly confident that these observations remain an interesting preliminary source of data for better understanding the relationship between persistent IFN-I upregulation and subclinical atherosclerosis in HIV-1-infected patients.

In conclusion, overall, these results supported the existence of a relationship between a dysregulated expression of IFN- α , IFN- β , and ISG56 and the onset of atherosclerosis in long-termtreated HIV-1-infected men, suggesting that measurement of IFN-I pathways might help for identifying those individuals who are at higher risk for CVD. Additional studies on IFN-I signature in larger cohorts of HIV-1-infected patients with CVD risk factors are needed to assess any clinical correlations and to better define the importance of IFN-I pathways in the management of atherosclerosis.

ACKNOWLEDGMENTS

This study was supported by research grants to Carolina Scagnolari from Sapienza University of Rome (Ricerche Universitarie, 2017 RM11715C586062AF; Finanziamenti di ateneo per la Ricerca Scientifica-2019, RP11916B6EC60AF3), from Gilead Sciences (Gilead fellowship Programme 2016, Title: Analysis of immunogenetic patterns, viral burden and cardiovascular diseases: a new integrative model for the management of HIV-1 patients suffering from cardiovascular damage) and to Gabriella d'Ettorre from Sapienza University of Rome (Ricerca Ateneo Sapienza, Progetti Medi, 2018 DDA N. 801/2019).

CONFLICT OF INTERESTS

The authors declare that there are no conflict of interests.

AUTHOR CONTRIBUTIONS

Letizia Santinelli: investigation, data curation, formal analysis, writingoriginal draft; Gabriella De Girolamo: formal analysis, writing-original draft; Cristian Borrazzo: software, formal analysis, data curation; Paolo Vassalini: investigation, resources; Claudia Pinacchio: investigation, resources, data curation; Eugenio Nelson Cavallari: clinicical investigation, resources; Maura Statzu: investigation, resources, data curation; Federica Frasca: investigation, data curation; Mirko Scordio: investigation, data curation; Camilla Bitossi: investigation; Agnese Viscido: investigation; Giancarlo Ceccarelli: validation, visualization; Massimo Mancone: methodology, investigation; Claudio Maria Mastroianni: validation, visualization; Guido Antonelli: validation, visualization; Gabriella d'Ettorre: project administration, validation, visualization, funding acquisition; Carolina Scagnolari: conceptualization, validation, writing-original draft, editing and revision, visualization, supervision, project administration, funding acquisition. All authors approved the final manuscript.

PEER REVIEW

The peer review history for this article is available at https://publons. com/publon/10.1002/jmv.27028

DATA AVAILABILITY STATEMENT

The data are not publicly available due to them containing information that could compromise research participant privacy/ consent.

ORCID

Letizia Santinelli b http://orcid.org/0000-0002-5621-058X Gabriella De Girolamo b http://orcid.org/0000-0003-2667-0603 Cristian Borrazzo b http://orcid.org/0000-0002-2346-898X Paolo Vassalini b http://orcid.org/0000-0002-6336-0455 Claudia Pinacchio b http://orcid.org/0000-0003-4370-3366 Eugenio Nelson Cavallari b http://orcid.org/0000-0003-4370-3366 Eugenio Nelson Cavallari http://orcid.org/0000-0003-0070-2195 Federica Frasca b http://orcid.org/0000-0002-9086-4659 Mirko Scordio b http://orcid.org/0000-0003-4673-6681 Camilla Bitossi b http://orcid.org/0000-0003-0143-9413

JOURNAL OF MEDICAL VIROLOGY

Agnese Viscido b http://orcid.org/0000-0003-3724-9291 Giancarlo Ceccarelli http://orcid.org/0000-0001-5921-3180 Massimo Mancone http://orcid.org/0000-0002-9602-0684 Claudio Maria Mastroianni http://orcid.org/0000-0002-1286-467X Guido Antonelli http://orcid.org/0000-0002-2533-2939 Gabriella d'Ettorre http://orcid.org/0000-0002-3571-5677 Carolina Scagnolari http://orcid.org/0000-0003-1044-1478

REFERENCES

- Miners A, Phillips A, Kreif N, et al. Health-related quality-of-life of people with HIV in the era of combination antiretroviral treatment: a cross-sectional comparison with the general population. *Lancet HIV.* 2014;1(1):e32-e40. https://doi.org/10.1016/S2352-3018(14) 70018-9
- 2. Oursler KA, Sorkin JD. HIV and aging. Int J Infect Dis. 2016;53:59-60 . https://doi.org/10.1016/j.ijid.2016.11.414
- Barnes RP, Lacson CJA, Bahrami H. HIV infection and risk of cardiovascular diseases beyond coronary artery disease. *Curr Atheroscler Rep.* 2017;19(5):20. https://doi.org/10.1007/s11883-017-0652-3
- Grinspoon S, Carr A. Cardiovascular risk and body fat abnormalities in HIV-infected adults. N Engl J Med. 2005;352:48-62.
- 5. Ho JE, Hsue PY. Cardiovascular manifestations of HIV infection. *Heart*. 2009;95:1193-1202.
- Pinto DSM, da Silva MJLV. Cardiovascular disease in the setting of human immunodeficiency virus infection. *Curr Cardiol Rev.* 2018;14: 25-41. https://doi.org/10.2174/1573403X13666171129170046
- Levy Z. Low-dose interferon-alpha accelerates atherosclerosis in an LDL receptor-deficient mouse model. *Eur J Intern Med.* 2003;14: 479-483. https://doi.org/10.1016/j.ejim.2003.08.010
- Goossens P, Gijbels MJJ, Zernecke A, et al. Myeloid type I interferon signaling promotes atherosclerosis by stimulating macrophage recruitment to lesions. *Cell Metab.* 2010;12:142-153. https://doi.org/ 10.1016/j.cmet.2010.06.008
- Li J, Fu Q, Cui H, et al. Interferon-α priming promotes lipid uptake and macrophage-derived foam cell formation: a novel link between interferon-α and atherosclerosis in lupus. *Arthritis Rheum*. 2011;63: 492-502. https://doi.org/10.1002/art.30165
- Zhang LN, Velichko S, Vincelette J, et al. Interferon-beta attenuates angiotensin II-accelerated atherosclerosis and vascular remodeling in apolipoprotein E deficient mice. *Atherosclerosis.* 2008;197: 204-211. https://doi.org/10.1016/j.atherosclerosis.2007.03.019
- Daissormont ITMN, Christ A, Temmerman L, et al. Plasmacytoid dendritic cells protect against atherosclerosis by tuning T-cell proliferation and activity. *Circ Res.* 2011;109:1387-1395. https://doi. org/10.1161/CIRCRESAHA.111.256529
- 12. Koulis C, Chen YC, Hausding C, et al. Protective role for Toll-like receptor-9 in the development of atherosclerosis in apolipoprotein E-deficient mice. *Arterioscler Thromb Vasc Biol.* 2014;34:516-525. https://doi.org/10.1161/ATVBAHA.113.302407
- Kahlenberg JM, Kaplan MJ. Mechanisms of premature atherosclerosis in rheumatoid arthritis and lupus. *Annu Rev Med.* 2013;64: 249-263. https://doi.org/10.1146/annurev-med-060911-090007
- Pinacchio C, Scheri GC, Statzu M, et al. Type I/II interferon in HIV-1infected patients: expression in gut mucosa and in peripheral blood mononuclear cells and its modification upon probiotic supplementation. J Immunol Res. 2018;2018:1738676-1738677. https:// doi.org/10.1155/2018/1738676
- Doyle T, Goujon C, Malim MH. HIV-1 and interferons: who's interfering with whom? *Nat Rev Microbiol*. 2015;13:403-413. https://doi. org/10.1038/nrmicro3449
- Scagnolari C, Antonelli G. Type I interferon and HIV: subtle balance between antiviral activity, immunopathogenesis and the microbiome.

Cytokine Growth Factor Rev. 2018;40:19-31. https://doi.org/10.1016/j. cytogfr.2018.03.003

- Scagnolari C, Monteleone K, Selvaggi C, et al. ISG15 expression correlates with HIV-1 viral load and with factors regulating T cell response. *Immunobiology*. 2016;221(2):282-290. https://doi.org/10. 1016/j.imbio.2015.10.007
- Triant VA, Lee H, Hadigan C, Grinspoon SK. Increased acute myocardial infarction rates and cardiovascular risk factors among patients with human immunodeficiency virus disease. J Clin Endocrinol Metab. 2007;92:2506-2512. https://doi.org/10.1210/jc.2006-2190
- Obel N, Thomsen HF, Kronborg G, et al. Ischemic heart disease in HIV-infected and HIV-uninfected individuals: a population-based cohort study. *Clin Infect Dis.* 2007;44:1625-1631. https://doi.org/10. 1086/518285
- Li Y, Li C, Xue P, et al. ISG56 is a negative-feedback regulator of virus-triggered signaling and cellular antiviral response. Proc Natl Acad Sci U S A. 2009;106(19):7945-7950. https://doi.org/10.1073/ pnas.0900818106
- d' Ettorre G, Francone M, Mancone M, et al. Significant coronary stenosis detected by coronary computed angiography in asymptomatic HIV infected subjects. J Infect. 2012;64(1):82-88. https://doi. org/10.1016/j.jinf.2011.09.007
- Agatston AS, Janowitz WR, Hildner FJ, Zusmer NR, Viamonte M Jr, Detrano R. Quantification of coronary calcium using ultrafast computed tomography. J Am Coll Cardiol. 1990;15:827e32-832e32. https://doi.org/10.1016/0735-1097(90)90282-t
- Miller JM, Dewey M, Vavere AL, et al. Coronary CT angiography using 64 detector rows: methods and design of the multi-centre trial CORE-64. Eur Radiol. 2009;19:816e28-828e28. https://doi.org/10. 1007/s00330-008-1203-7
- King SB 3rd, Smith SC Jr, Hirshfeld JW Jr, et al. Focused update of the ACC/AHA/SCAI 2005 guideline update for percutaneous coronary intervention: a report of the American College of Cardiology/ American Heart Association task force on practice guidelines: 2007 Writing group to review new evidence and update the ACC/AHA/ SCAI 2005 guideline update for percutaneous coronary intervention, writing on behalf of the 2005 writing committee. *Circulation*. 2008;117:261e95. https://doi.org/10.1161/CIRCULATIONAHA. 107.188208
- Santinelli L, Statzu M, Pierangeli A, et al. Increased expression of IL-32 correlates with IFN-γ, Th1 and Tc1 in virologically suppressed HIV-1-infected patients. *Cytokine*. 2019;120:273-281. https://doi. org/10.1016/j.cyto.2019.01.012
- Hughes R, Towers G, Noursadeghi M. Innate immune interferon responses to human immunodeficiency virus-1 infection. *Rev Med Virol.* 2012;22:257-266. https://doi.org/10.1002/rmv.1708
- Trinchieri G. Type I interferon: friend or foe. J Exp Med. 2010;207: 2053-2063. https://doi.org/10.1084/jem.20101664
- Somers EC, Zhao W, Lewis EE, et al. Type I interferons are associated with subclinical markers of cardiovascular disease in a cohort of systemic lupus erythematosus patients. *PLOS One*. 2012;7(5): e37000. https://doi.org/10.1371/journal.pone.0037000
- Law M, Friis-Moller N, El-Sadr W, et al. The use of the framingham equation to predict myocardial infarctions in HIV-infected patients: comparison with observed events in the D:A:D study. *HIV Med.* 2006;7(4):218-230. https://doi.org/10.1111/j.1468-1293.2006. 00362.x
- Hodis HN. The role of carotid arterial intima-media thickness in predicting clinical coronary events. Ann Intern Med. 1998;128: 262-269. https://doi.org/10.7326/0003-4819-128-4-199802150-00002
- 31. Simova I. Intima-media thickness: appropriate evaluation and proper measurement. *E-J ESC Council Cardiol Pract Eur Soc Cardiol*. 2015.
- 32. Schuijf JD, Bax JJ, Shaw LJ, et al. Meta-analysis of comparative diagnostic performance of magnetic resonance imaging and

ILEY-MEDICAL VIROLOGY

multislice computed tomography for noninvasive coronary angiography. *Am Heart J.* 2006;151:404e11-411e11. https://doi.org/10. 1016/j.ahj.2005.03.022

- Watkins MW, Hesse B, Green CE, et al. Detection of coronary artery stenosis using 40-channel computed tomography with multisegment reconstruction. Am J Cardiol. 2007;99:175e81-181e81. https://doi.org/10.1016/j.amjcard.2006.07.081
- Carrascosa P, Capuñay C, Deviggiano A, et al. Accuracy of low-dose prospectively gated axial coronary CT angiography for the assessment of coronary artery stenosis in patients with stable heart rate. *J Cardiovasc Comput Tomogr.* 2010;4:197e205-205e205. https://doi. org/10.1016/j.jcct.2010.04.001
- León R, Reus S, López N, et al. Subclinical atherosclerosis in low framingham risk HIV patients. Eur J Clin Invest. 2017;47(8): 591-599. https://doi.org/10.1111/eci.12780
- Osibogun O, Ogunmoroti O, Michos ED, et al. HIV/HCV coinfection and the risk of cardiovascular disease: a meta-analysis. *J Viral Hepat*. 2017;24(11):998-1004.
- Thacker SG, Zhao W, Smith CK, et al. Type I interferons modulate vascular function, repair, thrombosis, and plaque progression in murine models of lupus and atherosclerosis. Arthritis Rheum. 2012;64:2975-2985. https://doi.org/10.1002/art.34504
- Denny MF, Thacker S, Mehta H, et al. Interferon-alpha promotes abnormal vasculogenesis in lupus: a potential pathway for premature atherosclerosis. *Blood*. 2007;110(8):2907-2915. https:// doi.org/10.1182/blood-2007-05-089086

- Singh RB, Mengi SA, Xu YJ, Arneja AS, Dhalla NS. Pathogenesis of atherosclerosis: a multifactorial process. *Exp Clin Cardiol*. 2002;7(1):40-53.
- Boshuizen MCS, de Winther MPJ. Interferons as essential modulators of atherosclerosis. Arterioscler Thromb Vasc Biol. 2015;35:1579-1588. https://doi.org/10.1161/ATVBAHA.115. 305464
- Crowe SM, Westhorpe CLV, Mukhamedova N, Jaworowski A, Sviridov D, Bukrinsky M. The macrophage: the intersection between HIV infection and atherosclerosis. J Leukoc Biol. 2010; 87(4):589-598. https://doi.org/10.1189/jlb.0809580
- Lo J, Abbara S, Rocha-Filho JA, Shturman L, Wei J, Grinspoon SK. Increased prevalence of subclinical coronary atherosclerosis detected by coronary computed tomography angiography in HIV-infected men. AIDS. 2010;24:243e53-253e53. https://doi. org/10.1097/QAD.0b013e328333ea9e

How to cite this article: Santinelli L, De Girolamo G, Borrazzo C, et al. Alteration of type I interferon response is associated with subclinical atherosclerosis in virologically suppressed HIV-1-infected male patients. *J Med Virol*. 2021;93:4930-4938. https://doi.org/10.1002/jmv.27028