

Indole and p-cresol in feces of healthy subjects: concentration, kinetics, and correlation with microbiome

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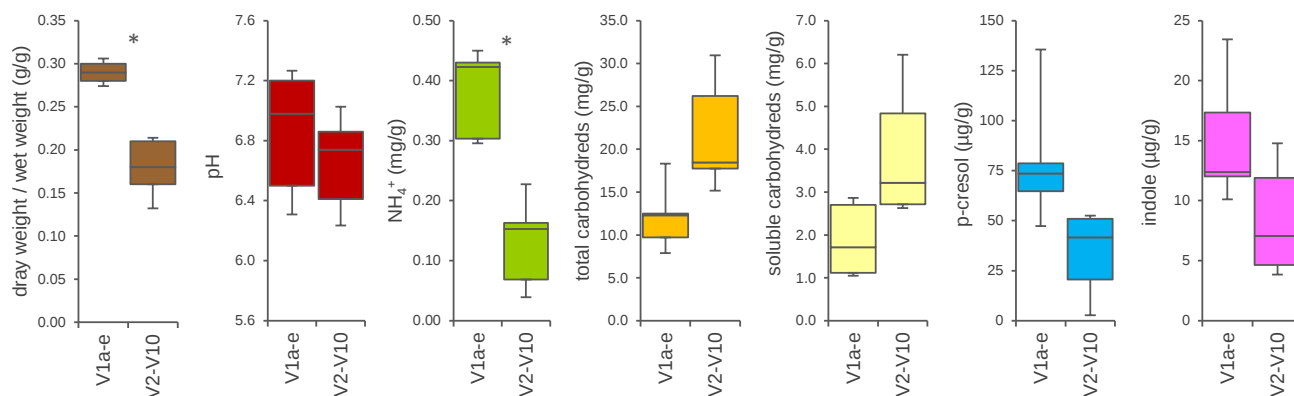
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Supplementary Material

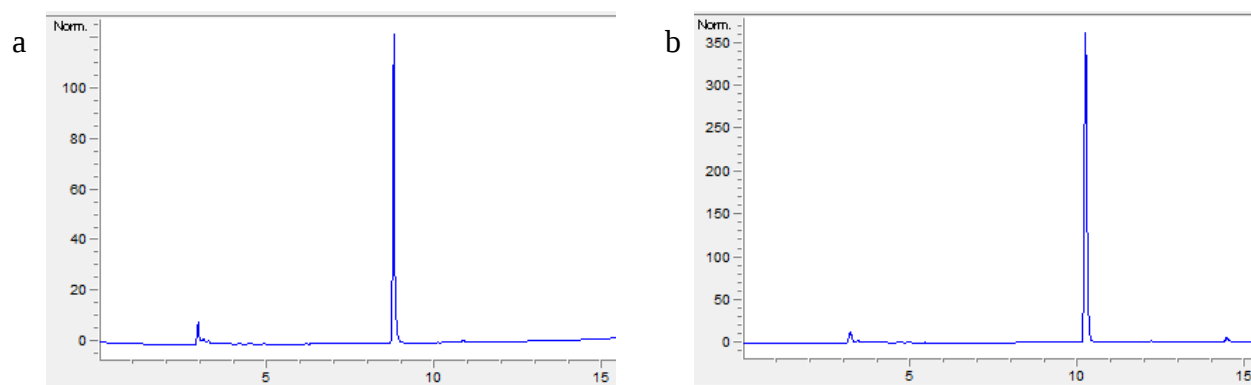
Supplementary Table 1. Biotransformation of p-cresol and indole with resting cells of 33 *Bifidobacterium* strains belonging to 8 different species or subspecies and 26 Lactobacillaceae ascribed to 15 taxa. The percentage of removal of the two molecules from the supernatants were reported as means $\pm$ standard deviation of three independent experiments.

strain	p-cresol	indole
<i>Bifidobacterium animalis</i> subsp. <i>lactis</i> WC 0432	5.3 $\pm$ 1.3	35.2 $\pm$ 8.9
<i>Bifidobacterium bifidum</i> MB 254	0.7 $\pm$ 0.2	39.9 $\pm$ 5.4
<i>Bifidobacterium bifidum</i> WC 0417	0.9 $\pm$ 0.3	29.6 $\pm$ 4.0
<i>Bifidobacterium bifidum</i> WC 0418	0.0 $\pm$ 0.0	38.5 $\pm$ 11.8
<i>Bifidobacterium breve</i> B 2429	0.0 $\pm$ 0.0	25.2 $\pm$ 7.9
<i>Bifidobacterium breve</i> WC 0420	0.0 $\pm$ 0.0	28.2 $\pm$ 0.6
<i>Bifidobacterium breve</i> WC 0421	3.0 $\pm$ 0.7	26.4 $\pm$ 1.8
<i>Bifidobacterium breve</i> WC 0422	0.0 $\pm$ 0.0	31.5 $\pm$ 0.5
<i>Bifidobacterium breve</i> WC 0423	13.0 $\pm$ 1.9	24.0 $\pm$ 4.3
<i>Bifidobacterium catenulatum</i> WC 0458	0.7 $\pm$ 0.2	36.4 $\pm$ 2.7
<i>Bifidobacterium catenulatum</i> WC 0469	0.0 $\pm$ 0.0	39.9 $\pm$ 5.6
<i>Bifidobacterium lactis</i> WC 0413	0.9 $\pm$ 0.3	29.0 $\pm$ 5.0
<i>Bifidobacterium lactis</i> WC 0414	0.0 $\pm$ 0.0	28.7 $\pm$ 0.4
<i>Bifidobacterium lactis</i> WC 0455	1.9 $\pm$ 0.2	28.9 $\pm$ 12.0
<i>Bifidobacterium lactis</i> WC 0459	13.6 $\pm$ 2.3	37.8 $\pm$ 3.2
<i>Bifidobacterium longum</i> MB 201	1.3 $\pm$ 0.3	26.9 $\pm$ 0.1
<i>Bifidobacterium longum</i> MB 219	0.3 $\pm$ 0.2	40.0 $\pm$ 2.8
<i>Bifidobacterium longum</i> subsp. <i>infantis</i> MB 208	0.0 $\pm$ 0.0	41.7 $\pm$ 6.0
<i>Bifidobacterium longum</i> subsp. <i>infantis</i> MB 256	3.5 $\pm$ 0.7	39.0 $\pm$ 3.3
<i>Bifidobacterium longum</i> subsp. <i>infantis</i> WC 0434	0.0 $\pm$ 0.0	30.5 $\pm$ 4.6
<i>Bifidobacterium longum</i> subsp. <i>longum</i> WC 0436	13.6 $\pm$ 2.4	38.0 $\pm$ 5.0
<i>Bifidobacterium longum</i> subsp. <i>longum</i> WC 0438	0.0 $\pm$ 0.0	39.0 $\pm$ 4.2
<i>Bifidobacterium longum</i> subsp. <i>longum</i> WC 0439	13.2 $\pm$ 1.5	40.2 $\pm$ 3.3
<i>Bifidobacterium longum</i> subsp. <i>longum</i> WC 0440	13.1 $\pm$ 0.4	25.3 $\pm$ 3.1
<i>Bifidobacterium longum</i> subsp. <i>longum</i> WC 0443	0.0 $\pm$ 0.0	39.7 $\pm$ 4.8
<i>Bifidobacterium pseudocatenulatum</i> MB 114	18.2 $\pm$ 4.5	29.0 $\pm$ 3.0
<i>Bifidobacterium pseudocatenulatum</i> MB 116	5.4 $\pm$ 1.2	24.5 $\pm$ 1.4
<i>Bifidobacterium pseudocatenulatum</i> MB 243	0.0 $\pm$ 0.0	31.2 $\pm$ 3.4
<i>Bifidobacterium pseudocatenulatum</i> WC 0400	13.8 $\pm$ 2.1	28.8 $\pm$ 1.4
<i>Bifidobacterium pseudocatenulatum</i> WC 0401	0.0 $\pm$ 0.0	30.0 $\pm$ 0.3
<i>Bifidobacterium pseudocatenulatum</i> WC 0403	0.0 $\pm$ 0.0	26.2 $\pm$ 0.9
<i>Bifidobacterium pseudocatenulatum</i> WC 0407	12.9 $\pm$ 1.5	26.8 $\pm$ 0.7
<i>Bifidobacterium pseudocatenulatum</i> WC 0408	2.5 $\pm$ 0.5	31.7 $\pm$ 5.2
<i>Lactocaseibacillus paracasei</i> WC 0227	13.1 $\pm$ 1.8	4.3 $\pm$ 0.7
<i>Lactocaseibacillus rhamnosus</i> WC 0232	15.7 $\pm$ 2.0	2.4 $\pm$ 1.1
<i>Lactocaseibacillus rhamnosus</i> WC 0295	20.8 $\pm$ 1.9	11.2 $\pm$ 2.4
<i>Lactocaseibacillus rhamnosus</i> WC0216	11.0 $\pm$ 1.6	0.8 $\pm$ 1.1
<i>Lactiplantibacillus pentosus</i> WC 0308	15.4 $\pm$ 2.2	4.9 $\pm$ 2.3
<i>Lactiplantibacillus plantarum</i> WC 0230	19.9 $\pm$ 2.0	4.5 $\pm$ 2.1
<i>Lactiplantibacillus plantarum</i> WC 0214	16.1 $\pm$ 2.1	7.1 $\pm$ 3.3
<i>Lactiplantibacillus plantarum</i> WC 0292	21.4 $\pm$ 1.9	14.9 $\pm$ 2.1
<i>Lactobacillus acidophilus</i> WC 0203	14.4 $\pm$ 1.9	9.1 $\pm$ 2.3
<i>Lactobacillus acidophilus</i> WC 0281	20.7 $\pm$ 1.8	1.3 $\pm$ 0.1
<i>Lactobacillus delbrueckii</i> subsp. <i>delbrueckii</i> WC 0286	14.0 $\pm$ 2.0	0.9 $\pm$ 1.2
<i>Lactobacillus delbrueckii</i> subsp. <i>lactis</i> WC 0290	14.9 $\pm$ 2.1	16.3 $\pm$ 2.9
<i>Lactobacillus delbrueckii</i> subsp. <i>lactis</i> WC 0291	19.6 $\pm$ 1.7	9.3 $\pm$ 1.7
<i>Lactobacillus gasseri</i> WC 0213	16.9 $\pm$ 2.4	13.9 $\pm$ 2.6
<i>Latilactobacillus curvatus</i> WC 0301	16.3 $\pm$ 1.2	10.9 $\pm$ 2.3
<i>Latilactobacillus sakei</i> WC 0313	15.8 $\pm$ 2.2	10.5 $\pm$ 1.9
<i>Latilactobacillus sakei</i> WC 0314	19.8 $\pm$ 1.7	6.8 $\pm$ 1.3
<i>Levilactobacillus brevis</i> WC 0282	20.3 $\pm$ 1.9	8.5 $\pm$ 1.4
<i>Ligilactobacillus ruminis</i> WC 0225	21.3 $\pm$ 2.0	6.9 $\pm$ 0.7
<i>Limosilactobacillus fermentum</i> WC 0212	15.4 $\pm$ 1.2	8.4 $\pm$ 2.1
<i>Limosilactobacillus mucosae</i> WC 0229	18.6 $\pm$ 2.0	3.8 $\pm$ 2.2
<i>Limosilactobacillus reuteri</i> WC 0215	14.7 $\pm$ 1.1	5.1 $\pm$ 1.7
<i>Limosilactobacillus reuteri</i> WC 0224	20.7 $\pm$ 1.9	2.3 $\pm$ 0.3
<i>Limosilactobacillus reuteri</i> WC 0231	19.0 $\pm$ 2.2	8.2 $\pm$ 2.3
<i>Limosilactobacillus reuteri</i> WC 0293	21.4 $\pm$ 2.0	14.3 $\pm$ 2.5
<i>Limosilactobacillus reuteri</i> WC 0294	20.3 $\pm$ 1.9	8.8 $\pm$ 1.3

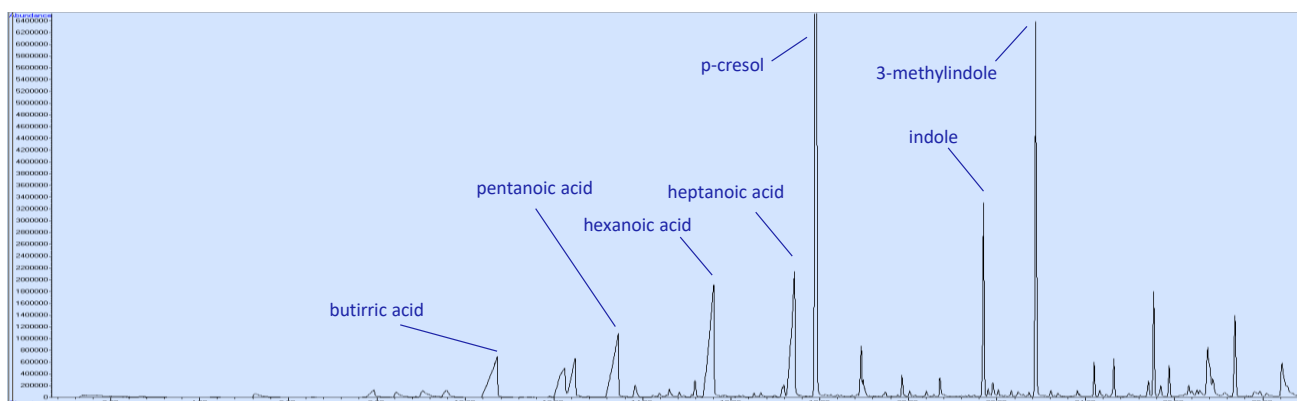
Supplementary Figure 1. Box-plot representation of the chemical parameters measured for the characterization of the fecal samples collected from 10 different volunteers (V1-10). Feces of volunteer 1 were collected five times (V1a-e). The mean values, 10th, 25th, 75th, 90th percentile and the significant differences are reported. * p<0.01



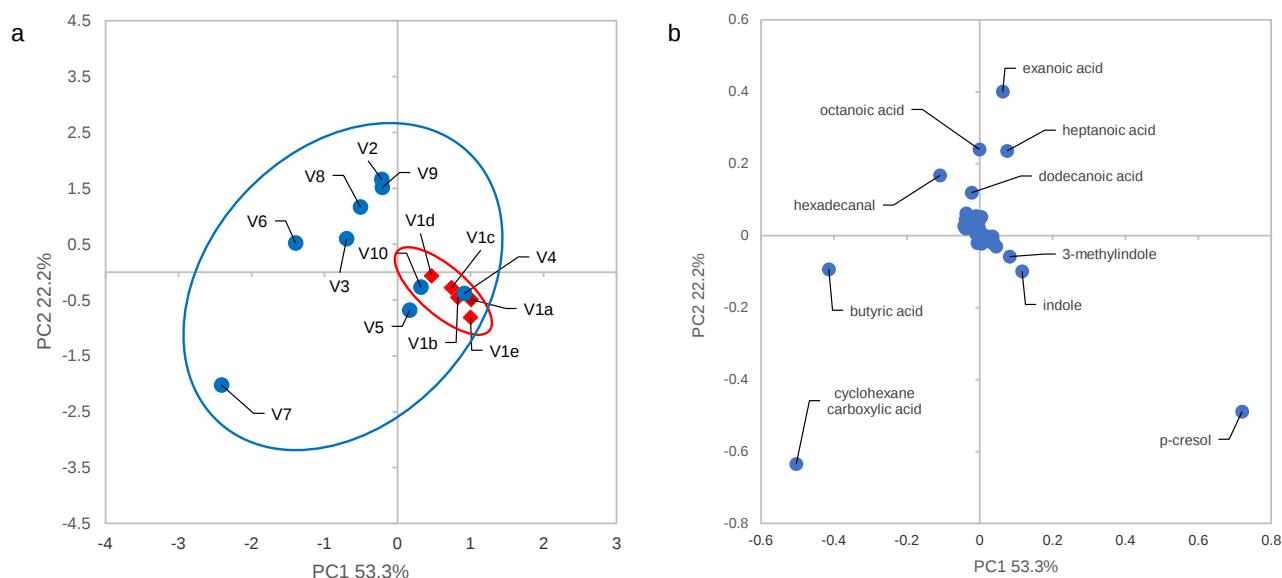
Supplementary Figure 2. Analysis of indole and p-cresol standard (1mM in H₂O) with HPLC, equipped with a diode array detector and a C18 column.



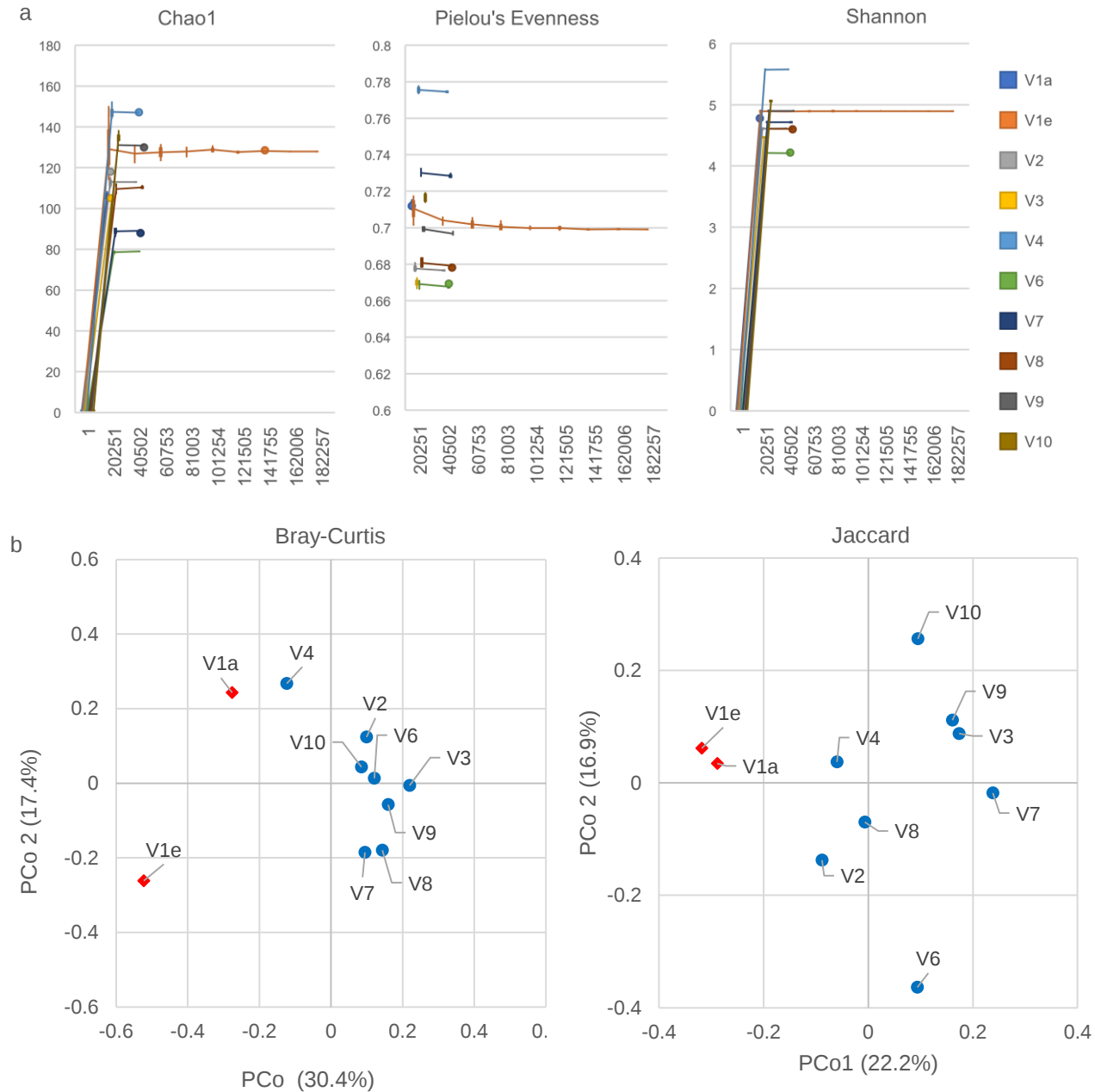
Supplementary Figure 3. Representative chromatogram of the SPME-GC-MS analysis of fecal samples. The main VOCs, occurring most frequently and abundantly are indicated.



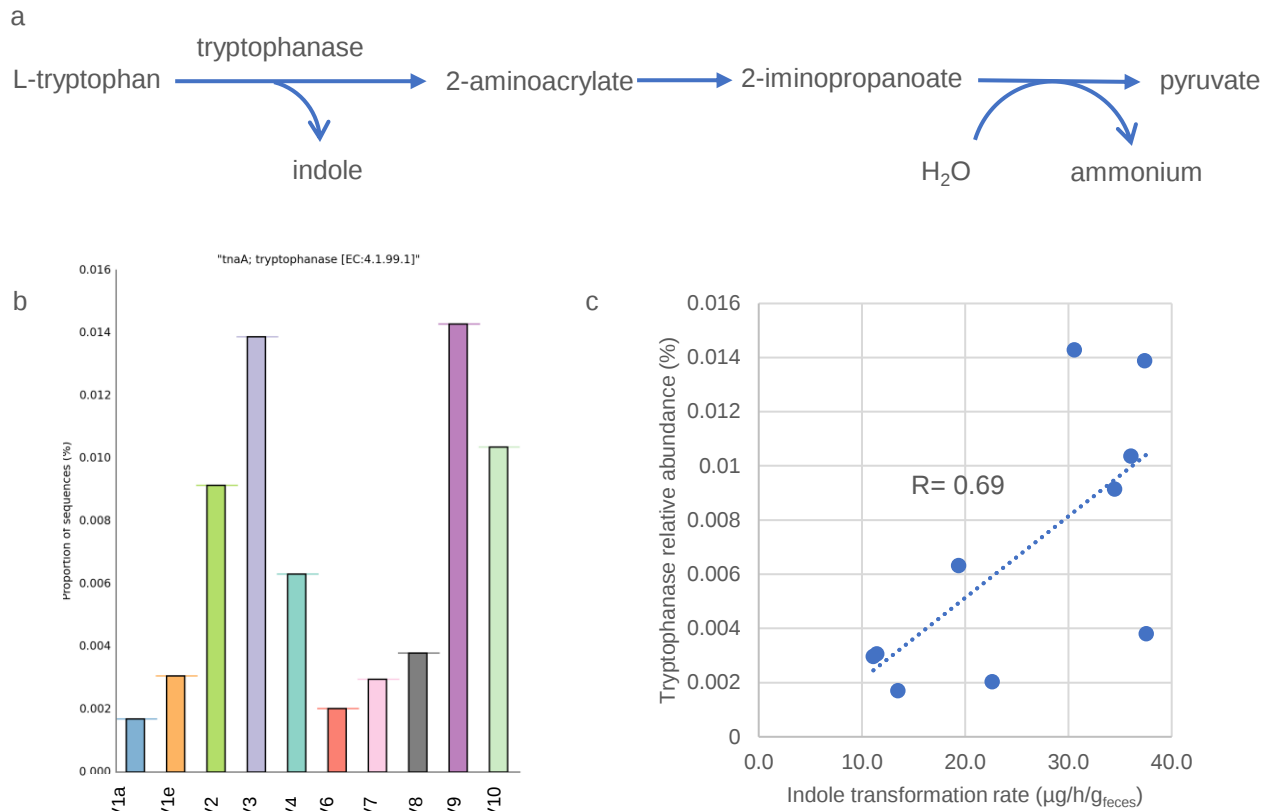
Supplementary Figure 4. PCA analysis of VOCs detected by SPME-GC-MS analyses in the fecal samples. **a** PC1 vs PC2 plot. Red diamond, samples V1a-e; blue circle, samples V2-V10. Ellipses represent the 95% of variability of the observed parameters ascribed to the corresponding set of samples. **b** Score plot showing the VOCs mainly contributing to PC1 and PC2.



Supplementary Figure 5. Analysis of microbiota composition of the fecal samples obtained by 16S rRNA gene profiling. **a** Alpha rarefaction plots of the main alpha diversity indexes. **b** Beta-diversity analyses: PCoA based on Bray-Curtis (quantitative) and Jaccard (qualitative) distance matrices.



Supplementary Figure 6: a Metabolic pathway of tryptophan degradation into indole. **b** Relative abundance of tryptophanase (*tnaA*) in the metagenomes predicted by PICRUST2. **c** Pearson's correlation between indole accumulation rate in TT slurries and tryptophanase relative abundance.



Supplementary Figure 7: Heatmap of Spearman’s correlation between microbial species, VOCs identified by HS-SPME-GC-MS analyses and coprometry parameters. Blue cladogram on the right represent features clusters obtained with UPGMA algorithm using PAST software according to their correlation with VOCs.

