


Cancer Biomarkers and Precision Oncology: A Review of Recent Trends and Innovations

Maha AlDoughaim¹, Nada AlSuhebany¹, Mohammed AlZahrani¹, Tariq AlQahtani¹, Sahar AlGhamdi¹, Hisham Badreldin¹ and Hana Al Alshaykh²

¹College of Pharmacy, King Saud Bin Abdul Aziz University for Health Sciences (KSAU-HS), King Abdullah International Medical Research Center (KAIMRC), Riyadh, Saudi Arabia. ²Pharmaceutical Care Division, King Faisal Specialist Hospital and Research Center (KFSHRC), Riyadh, Saudi Arabia.

Clinical Medicine Insights: Oncology
Volume 18: 1–15
© The Author(s) 2024
Article reuse guidelines:
sagepub.com/journals-permissions
DOI: 10.1177/11795549241298541



ABSTRACT: The discovery of cancer-specific biomarkers has resulted in major advancements in the field of cancer diagnostics and therapeutics, therefore significantly lowering cancer-related morbidity and mortality. Cancer biomarkers can be generally classified as prognostic biomarkers that predict specific disease outcomes and predictive biomarkers that predict disease response to targeted therapeutic interventions. As research in the area of predictive biomarkers continues to grow, precision medicine becomes far more integrated in cancer treatment. This article presents a general overview on the most recent advancements in the area of cancer biomarkers, immunotherapy, artificial intelligence, and pharmacogenomics of the Middle East.

KEYWORDS: Cancer biomarkers, precision medicine, targeted therapy, artificial intelligence, pharmacogenomics

RECEIVED: July 8, 2024. **ACCEPTED:** October 22, 2024.

TYPE: Review

FUNDING: The author(s) received no financial support for the research, authorship, and/or publication of this article.

DECLARATION OF CONFLICTING INTERESTS: The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

CORRESPONDING AUTHOR: Maha AlDoughaim, College of Pharmacy, King Saud Bin Abdul Aziz University for Health Sciences (KSAU-HS), King Abdullah International Medical Research Center (KAIMRC), Prince Mutib Ibn Abdullah Ibn Abdulaziz Rd, Ar Rimayah, Riyadh 11481, Saudi Arabia. Email: doughaimm@ksau-hs.edu.sa

Introduction

Cancer biomarkers have become integral in cancer research and drug development, playing a crucial role in guiding decision-making and ensuring the success of novel antitumor therapeutics.¹ There are 2 main types of biomarkers: disease-related and drug-related biomarkers. Disease-related biomarkers aid in risk stratification, identifying patients with cancer within the general population, staging cancer, and predicting disease outcomes.² Conversely, drug-related biomarkers provide valuable information about prognosis and survival following clinical intervention.^{2,3} These predictive factors assist in identifying patients who are most likely to benefit from specific treatments and assessing treatment effectiveness and potential side effects.^{2,3} Despite some progress in biomarker discovery, only a few biomarkers have been translated into routine clinical use. An ideal biomarker should be easily and reliably measured with high analytical specificity and sensitivity in a noninvasive or minimally invasive manner and at a low cost.⁴ However, the translation of biomarkers from the laboratory to clinical practice faces challenges such as small sample sizes; mismatched case and control specimens; and errors in sample collection, handling, storage, preparation, and analysis.⁴

The discovery of biomarkers is significant in the era of targeted therapy, where personalized medicine aims to predict the response to medical treatment and tailor interventions based on individual biological information.^{4,5} Cancer mortality rate continued to decline from 2019 to 2020 by 1.5%, and this

decline contributed to a 33% overall reduction in cancer deaths, preventing an estimated 3.8 million fatalities.⁶ This progress is largely attributed to advancements in cancer treatment, especially noticeable in the decrease in mortality rates for cancer diseases such as leukemia, melanoma, and kidney cancer, with an approximately 2% annual decline from 2016 to 2020.⁶

Recent trends and innovations in cancer biomarkers have revolutionized cancer treatment. This comprehensive review article aims to provide an in-depth overview of these biomarkers while also discussing the successes and obstacles encountered with targeted therapies. In addition, we provide insight into utilizing artificial intelligence (AI) in the field of cancer biomarkers. Finally, we discuss the current state and trends of cancer biomarkers in underrepresented Middle Eastern populations.

Precision Medicine and Targeted Therapies

Precision medicine represents a shift from the one-size-fits-all approach of traditional cancer treatments to more personalized, targeted interventions, which has shown promise in improving treatment outcomes and reducing the side effects associated with cancer therapies.⁷ Targeted cancer therapies are generally divided into 2 groups: small molecule drugs and monoclonal antibodies (mAbs).⁸ Small molecules (<900 Da) exert their effects by penetrating the cell and binding to specific cellular proteins to alter downstream signaling pathways. Monoclonal antibodies are larger proteins produced through recombinant technologies, and mAbs exert their therapeutic action by interacting with extracellular targets.^{9–11} Antibody drug conjugates (ADCs) are mAbs linked with a cytotoxic payload. This novel concept, which was introduced in the year 2000, has led to precise cancer cell

Hana Al Alshaykh is also affiliated to College of Pharmacy, AlFaisal University, Riyadh, Kingdom of Saudi Arabia.



Creative Commons Non Commercial CC BY-NC: This article is distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 License (<https://creativecommons.org/licenses/by-nc/4.0/>) which permits non-commercial use, reproduction and distribution of the work without further permission provided the original work is attributed as specified on the SAGE and Open Access pages (<https://us.sagepub.com/en-us/nam/open-access-at-sage>).

Cancer biomarkers: Major Targets for Cancer Therapies

Cancer Biomarkers :

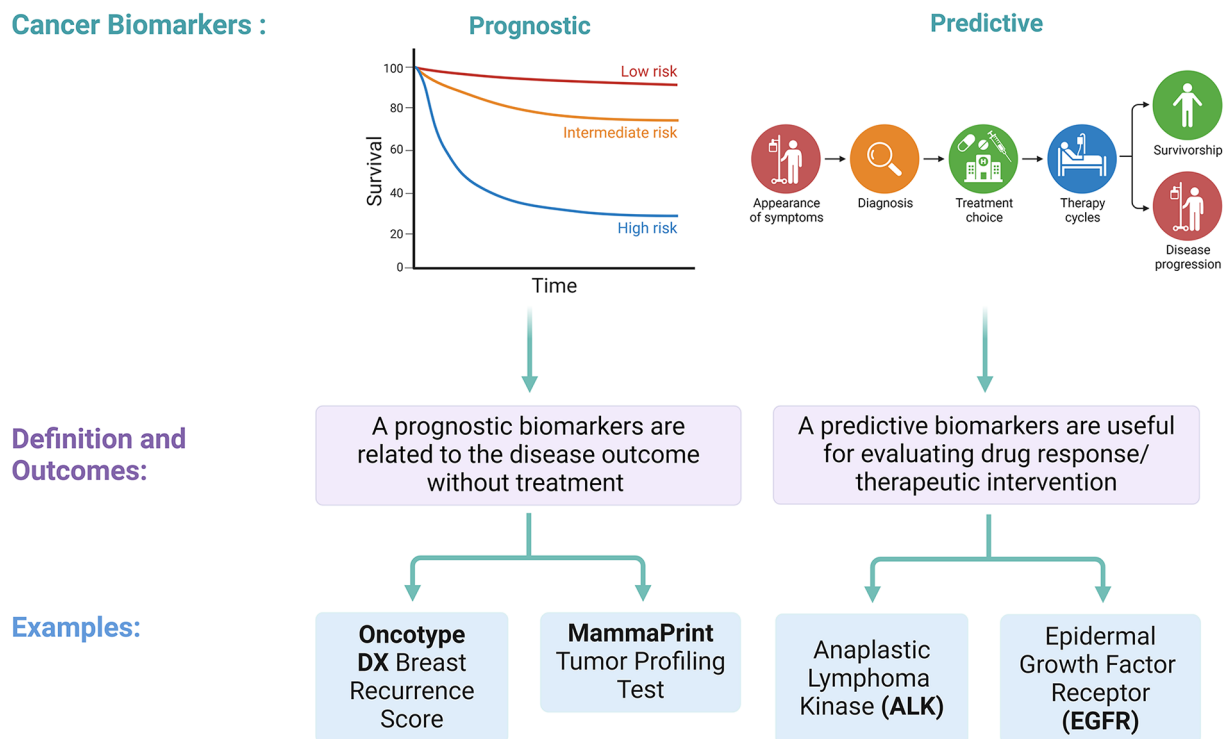


Figure 1. Biomarker classification based on treatment response. Prognostic biomarkers may be detected by standardized tests (eg, Oncotype Dx, MammaPrint). Predictive biomarkers that predict the response to targeted therapies including ALK and EGFR.

targeting with less cytotoxic chemotherapy-associated toxicity than conventional chemotherapy.^{12,13} In contrast to chemotherapy, targeted therapies are designed to specifically target some molecular structures within cells, often resulting in fewer and less severe side effects.⁸

In the realm of precision medicine, patients with cancer who receive treatment based on biomarker-guided approaches have shown improved outcomes compared with those receiving traditional treatment methods.¹⁴ The hallmarks of cancer, as described by Hanahan and Weinberg, provide a foundational framework for understanding cancer biology and have significantly influenced the development of rational drug designs. Among these hallmarks, selective growth and proliferative advantage are critical.¹⁴ An improved understanding of the hallmarks of cancer has led to a shift from nonselective, traditional therapies to more effective targeted treatments, which will be discussed in this section.

Biomarker classification

Cancer biomarkers are divided into prognostic and predictive biomarkers. Prognostic biomarkers provide insights into disease outcomes, while predictive biomarkers are useful for evaluating drug responses^{15,16} (Figure 1). Biomarkers may also be categorized based on patient genetics as either germline or somatic. Furthermore, a more complex framework exists for categorizing

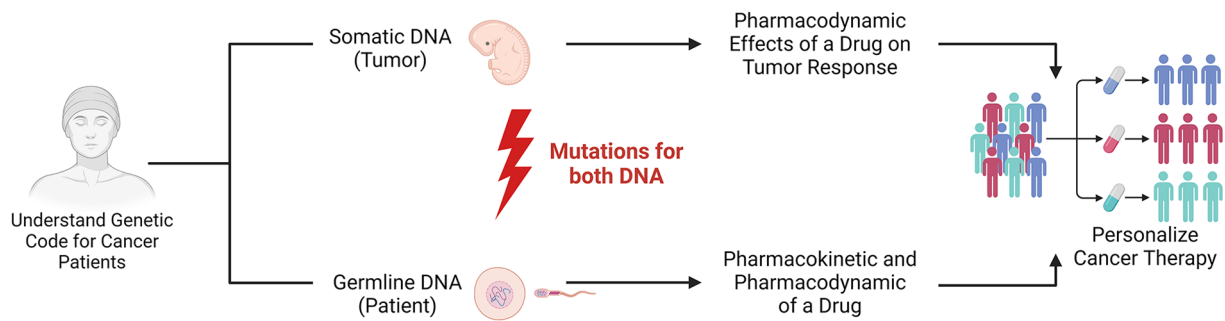
the significance and pathogenicity of various biomarkers^{16,17} (Figure 2).

Biomarkers in solid cancers

The fast-growing field of targeted therapies for cancer can be categorized according to their effect on one or more of the hallmarks of cancer.¹⁴ This section will discuss the role of cancer biomarkers in the top 3 reported solid cancers worldwide, which are breast, lung, and colorectal cancers (CRCs).⁶ In Table 1, we summarize the most common treatment-driven biomarkers and their targeted cancer therapies.

Lung cancer. Lung cancer, which includes small and non-small cell cancer types, is the leading cause of cancer deaths globally, with 1.8 million deaths/year.¹⁶ Despite advances in targeted therapies improving the prognosis of patients with advanced lung cancer, late-stage patients still face poor outcomes, highlighting the importance of early detection and aggressive treatment.¹⁵ In lung cancer treatment, personalized therapy based on molecular profiling has become crucial and used in multiple treatment algorithms, such as in the National Comprehensive Cancer Network (NCCN) and European Society for Medical Oncology (ESMO) guidelines.^{18,19}

DNA Analysis in Cancer Pharmacogenomics



Tier Based Classification of Somatic Variants

①	②	③	④
Tier I	Tier II	Tier III	Tier IV
Variants with Strong Clinical Significance	Variants with Potential Clinical Significance	Variants with Unknown Clinical Significance	Variants that are Benign or Likely Benign

Figure 2. Biomarker classification based on genomics and clinical utility. Somatic DNA, found in tumors, is useful for assessing the pharmacodynamic effects of a drug on the tumor response. Germline or inherited DNA variations predict the pharmacokinetic effect of a drug and its response. Multiple frameworks exist for categorizing the pathogenicity of somatic variants. One classification system is categorized as follows: Tier I for variants with strong clinical significance, tier II for variants with potential clinical significance, tier III for variants with unknown clinical significance, and tier IV for variants that are benign or likely benign.

Personalized medicine for lung cancer involves screening all advanced-stage patients for key mutations, such as those in epidermal growth factor receptor (EGFR), anaplastic lymphoma kinase (ALK), and ROS1, to tailor treatment plans.^{8,18} The EGFR is overexpressed in 10% to 50% of patients with non-small cell lung cancer (NSCLC), making it a great molecular target. The presence of EGFR exon 19 deletions or EGFR exon 21 L858R mutations is predictive of treatment benefit from EGFR-TKI therapy, such as osimertinib. The 2 predominant mutations observed in the EGFR gene are deletions in exons 19 and 21. On the contrary, approximately 5% of patients with NSCLC exhibit rearrangements in the ALK gene, and ALK inhibitors, such as alectinib, brigatinib, ceritinib, crizotinib, and lorlatinib, all of which have received Food and Drug Administration (FDA) approval, can be used in the metastatic setting.^{8,18} However, drug resistance remains a challenge in lung cancer treatment.¹⁸⁻²⁰ Common resistance mechanisms include mutations such as T790M in EGFR and amplification of MET or HER2. Combining drugs targeting different mechanisms is a potential strategy to combat this resistance.¹⁸

Breast cancer. Breast cancer is the most frequently diagnosed cancer worldwide, with approximately 2.3 million new cases

each year.⁶ This disease is highly heterogeneous, involving various genetic and environmental factors, making its treatment complex.⁸ Genetic factors such as mutations in the BRCA1 and BRCA2 genes significantly increase breast cancer risk. The prognosis and response to treatment vary greatly depending on tumor size, type, histological score, metastasis status, and receptor status, necessitating individualized treatment approaches.²¹

In breast cancer, targeted therapies have become increasingly important, particularly for estrogen receptor (ER)-positive cancers, which constitute approximately 75% of all breast cancer cases.²¹ Tamoxifen, an FDA-approved endocrine therapy, is widely used for these cancers.²² Fulvestrant, another ER antagonist, has shown efficacy comparable to that of anastrozole in clinical trials.²³ For human epidermal growth factor receptor 2 (HER2)-positive breast cancer, trastuzumab, a mAb, has been shown to be effective at improving survival rates.²¹ Pertuzumab, another antibody targeting HER2:HER3 dimerization, has shown significant results in metastatic breast cancer. Lapatinib, a tyrosine kinase inhibitor (TKI) targeting HER2 and EGFR, is effective when used in combination with trastuzumab.²⁴ On the contrary, triple-negative breast cancer (TNBC) cells, particularly those with BRCA1/2 dysfunction, respond well to poly-ADP ribose polymerase inhibitors (PARPi), such as olaparib and talazoparib.^{22,25}

Table 1. Selected treatment-driven cancer biomarkers in the settings of advanced solid cancer diseases.

CANCER TYPE	BIOMARKERS	FDA-APPROVED THERAPIES	TARGETED THERAPY CLASSIFICATION
Lung cancer	EGFR exon 19 and 21	Afatinib, erlotinib, dacomitinib, gefitinib, osimertinib	Small molecules
	EGFR Exon 20 Insertion Mutation	Amivantamab-vmjw	Monoclonal antibody (bispecific)
	ALK Rearrangement	Alectinib, brigatinib, ceritinib, crizotinib, lorlatinib	Small molecules
	ROS1 Rearrangement	Ceritinib, crizotinib, entrectinib, repotrectinib	Small molecules
	BRAF V600E Mutation	Dabrafenib/trametinib, Encorafenib/binimetinib, Dabrafenib, Vemurafenib	Small molecules
	NTRK1/2/3 Gene Fusion	Larotrectinib, entrectinib	Small molecules
	MET Skipping Mutation	Capmatinib, crizotinib, tepotinib	Small molecules
	RET Rearrangement	Selpercatinib, pralsetinib, cabozantinib	Small molecules
Breast cancer	Germline or somatic BRCA1/2 mutation	PARPi (olaparib, talazoparib)	Small molecules
	HER2-positive	Trastuzumab, pertuzumab	Monoclonal antibody combination
		Neratinib	Small molecules
	HER2-Negative and ER/PR-positive	CDK 4/6 inhibitors (palbociclib, abemaciclib, ribociclib)	Small molecules
	ESR1 mutation	Elacestrant	Small molecule
	PIK3CA mutation	Alpelisib + fulvestrant	Small molecules
	AKT1/PTEN-activating mutation	Capivasertib + fulvestrant	Small molecules
Colorectal cancer	(KRAS/NRAS/BRAF WT and left-sided tumors only)	EGFR monoclonal antibodies (panitumumab, or cetuximab)	Monoclonal antibodies
	VEGF	Bevacizumab, aflibercept and ramucirumab	Monoclonal antibodies
		regorafenib and fruquintinib.	Small molecules
	KRAS G12C mutation	Sotorasib, adagrasib	Small molecules
	HER2-positive and RAS and BRAF wildtype	Trastuzumab/lapatinib, trastuzumab/tucatinib	Monoclonal antibody/ small molecule combinations
		Trastuzumab/pertuzumab	Monoclonal antibody combination
		fam-trastuzumab deruxtecan-nxki (T-DXd)	Monoclonal antibody (ADC)
BRAF V600E mutation	Encorafenib/panitumumab, encorafenib/cetuximab	Small molecule/monoclonal antibody combination	
	dabrafenib/trametinib	Small molecules	

Sacituzumab govitecan is an ADC that targets the Trop-2 surface antigen expressed in breast cancer and was initially approved for relapsed/refractory TNBC. In the recent Tropics-02 trial, sacituzumab govitecan demonstrated longer overall survival (OS) in treatment-resistant HR-positive breast cancer, indicating a greater role for ADC in addressing treatment resistance in breast cancer.²⁶ Furthermore, low-HER2 breast cancer classification has gained increased attention since

the approval of trastuzumab deruxtecan (T-DXd) for patients with advanced breast cancer with low HER2 expression. In the DESTINY-Breast04 trial, T-DXd was associated with favorable progression-free survival (PFS) and OS compared with standard chemotherapy.²⁷

Emerging targeted therapies for breast cancer include the inhibition of intracellular molecular pathways. For example, mTOR pathway inhibitors have shown improved outcomes in

resistant endocrine breast cancers.²⁸ CDK 4/6 inhibitors, such as palbociclib, abemaciclib, and ribociclib, in combination with aromatase inhibitors, have been shown to be effective in treating hormone receptor-positive breast cancer, significantly improving disease-free survival (DFS) rates.²⁹ These advances have led to the FDA approval of several new drugs, such as elacestrant for ESR1 mutations, PI3K inhibitors, and AKT inhibitors, in a short period of time.

Despite these advances, drug resistance remains a significant challenge in breast cancer treatment. Resistance mechanisms include increased drug efflux, enhanced metabolic enzyme production, and alterations in drug targets.³⁰ To address resistance to single agents, combination therapy has been widely applied to treat breast cancer.

The use of personalized medicine for the treatment of breast cancer, driven by molecular stratification and multi-gene microarray analysis, is evolving rapidly. This approach enables tailored treatment choices and specific target selection, reducing breast cancer-related mortality and changing the standard of care.²¹ The use of molecular assays and multi-gene microarrays, such as Oncotype DX and MammaPrint, allows for more individualized treatment decisions among patients with breast cancer. With ongoing advancements in targeted therapy, informed by gene expression analysis and high-throughput screening, the field is moving toward more effective, personalized treatments.

Colorectal cancer. Colorectal cancer ranks as the third most common and second deadliest cancer worldwide.⁶ Notably, 25% of patients with CRC are diagnosed at a late stage, rendering surgery ineffective.³¹ This highlights an urgent need for new therapeutic strategies to improve survival rates and mitigate the severity of CRC. Advancements in chemotherapy and targeted therapies have markedly improved outcomes for patients with metastatic CRC (mCRC), extending the average survival to nearly 40 months.⁸

The introduction of mAbs targeting EGFR, such as cetuximab and panitumumab, has been pivotal.³² These treatments have shown significant promise in improving PFS and OS, especially when combined with chemotherapy regimens such as FOLFOX.^{31,33} However, the efficacy of these anti-EGFR drugs as second-line treatments remains limited. In addition, antiangiogenic agents targeting vascular endothelial growth factor (VEGF), such as bevacizumab, aflibercept, regorafenib, and ramucirumab, have been approved for mCRC treatment following positive clinical trial results. However, these agents do not require testing for VEGF in patients with cancer since VEGF proteins are upregulated in CRC tissue and are one of the hallmarks of cancer.^{14,31} Recently, the FDA granted approval to fruquintinib, a novel oral kinase inhibitor that acts on VEGFR 1, 2, and 3.³⁴ Its effectiveness and safety were assessed in 2 randomized, double-blind, phase 3 clinical trials named FRESCO and FRESCO-2.^{35,36}

Other recent targeted therapies for mCRC include 2 KRAS G12C inhibitors, sotorasib and adagrasib.³¹ KRAS G12C inhibitors are recommended for the treatment of previously treated patients with mCRC harboring this mutation. Sotorasib or adagrasib should be given in combination with cetuximab or panitumumab or may be considered as a single agent. Other targeted therapies used for mCRC include BRAF V600E mutation, for which treatment combinations such as cetuximab or panitumumab combined with encorafenib are being used. In patients with mCRC with HER2 amplification, 4 different treatment regimens can be used: fam-trastuzumab deruxtecan-nxki (T-DXd) as a single agent or trastuzumab combined with pertuzumab, lapatinib, or tucatinib.³¹

Despite advances in molecular medicine and targeted drug development, drug resistance in mCRC remains a significant challenge. Resistance to anti-EGFR therapy, linked to mutations in genes such as BRAF, RAS, and PIK3CA, limits the effectiveness of these treatments to approximately 40% of patients with mCRC.³⁷ Addressing resistance mechanisms and developing new biomarkers and therapeutic pathways are crucial for enhancing treatment efficacy. Personalized medicine has gained importance in CRC treatment, with multiple targeted therapies showing promise. The molecular diversity of tumors necessitates individualized treatment plans, guided by biomarkers such as KRAS mutations, to tailor therapies such as cetuximab and panitumumab to each patient's specific needs.³⁷ However, there is a growing need for new indicators to predict responses to both existing and experimental treatments.

Selected targeted cancer therapies in hematologic cancers

The swift advancement of targeted therapies for blood cancers is transforming how we approach treatment, focusing on key aspects of cancer development. This section explores how cancer biomarkers play a crucial role in the most common global blood cancers. By pinpointing treatment biomarkers, treatment can be customized for better effectiveness and fewer side effects. In Table 2, we present a summary of significant treatment-driven biomarkers and the corresponding targeted therapies for cancer. This individualized method offers great potential for improving patient results, reducing negative impacts, and changing the scope of hematologic malignancy treatment.

Acute myeloid leukemia. Acute myeloid leukemia (AML) accounts for 80% of all acute leukemia in adults worldwide.³⁹ Despite the availability of multiple prognostic factors for AML, the mortality rate is still high.⁴⁰ FMS-like tyrosine kinase 3 (FLT3) is considered one of the crucial biomarkers tested in AML patients and can classify patients as high versus low risk.⁴¹ The role of FLT3 in hematopoietic cells includes controlling cell proliferation, differentiation, and survival.⁴² Patients with AML who tested positive for FLT3 were

Table 2. Selected treatment-driven cancer biomarkers for hematologic malignancies.

CANCER TYPE	BIOMARKERS	FDA-APPROVED THERAPIES	TARGETED THERAPY CLASSIFICATION
AML	FLT3	Midostaurin, Gilteritinib, Quizartinib	Small molecules
	BCL-2	Venetoclax	Small molecule
	IDH1/IDH2	Ivosidenib, Olutasidenib, Enasidenib	Small molecules
CML	BCR-ABL	Imatinib, bosutinib, dasatinib, nilotinib, and ponatinib	Small molecules
Lymphoma	CD30	Brentuximab vedotin	Monoclonal antibody (ADC)
	ALK	Alectinib, Brigatinib, Ceritinib, Lorlatinib	Small molecules

Recent trends in cancer biomarkers underscore the continuous evolution of diagnostic and therapeutic strategies aimed at improving patient outcomes. Multiple trials are ongoing exploring the benefit of using targeted therapies and whether personalized medicine is more beneficial than standard-of-care treatment. The TAPUR Study, which stands for the Targeted Agent and Profiling Utilization Registry, is a clinical trial initiative in oncology.³⁸ It is designed to explore the effectiveness of targeted therapies outside of their approved indications. The TAPUR Study aims to investigate whether targeted therapies approved for one type of cancer may also be effective against other types of cancer with specific genetic mutations or biomarkers.

considered at high risk of relapse/recurrence compared with those missing this mutation.⁴² FLT3 mutations are classified as either internal tandem duplications, which are usually located within the juxta membrane, or tyrosine kinase domain, which are considered missense mutations.⁴¹ Luckily, multiple targeted agents, such as midostaurin, gilteritinib and quizartinib, have been approved for targeting FLT3.⁴³

B-cell lymphoma 2 inhibition (BCL-2) is another essential biomarker usually considered an adverse prognostic factor in AML.⁴⁴ The role of BCL-2 in hematopoietic cells is to control antiapoptotic proteins, which ultimately promote the progression of malignant cells. Venetoclax is an approved drug that targets BCL-2 in patients with AML.⁴⁵ Other prognostic biomarkers in AML include isocitrate dehydrogenase (IDH1 and IDH2), which are responsible for the conversion of alpha-ketoglutarate to 2-hydroxyglutarate, which disrupts epigenetic regulation and eventually blocks the differentiation of hematopoietic cells.⁴⁶ The prognostic potential of IDH in AML is not yet clear, with variations in prognosis based on IDH mutational isoforms.⁴⁷ There are 2 approved agents targeting IDH isoforms, namely, ivosidenib and olutasidenib for IDH1 mutation and enasidenib for IDH2 mutation.^{47,48} The identification and targeting of crucial biomarkers such as FLT3, BCL-2, and IDH have significantly improved the management of AML, offering hope for better outcomes in patients facing this challenging disease.

Chronic myeloid leukemia. The discovery of breakpoint cluster region and Abelson murine leukemia viral oncogene homolog (BCR-ABL) gene has played a tremendous role in understanding cancer development in patients with chronic myeloid leukemia (CML).⁴⁹ It was first described by Nowell et al⁵⁰ in 1960 and was then named after the city (Philadelphia chromosome) where it was first discovered. The fusion of BCR-ABL gene is responsible for the translocation of chromosomes 9 and 22, which enhances cancerous cell growth and proliferation.⁴⁹ Luckily, the BCR-ABL gene pathway is dependent on tyrosine

kinase activity, making it an optimal target for treating patients with CML. The first approved TKI-targeted agent in CML was imatinib, which was introduced to the market in early 2001 after its FDA approval, making it a transformational treatment for CML.⁵¹ However, despite the initial success of imatinib, resistance, and intolerance issues necessitated the exploration of alternative treatment strategies. This has prompted extensive research efforts aimed at developing novel agents capable of overcoming these challenges, such as developing second-generation TKIs targeting BCR-ABL, including dasatinib, bosutinib, nilotinib, and ponatinib.⁵¹ Despite initial successes, ongoing efforts to address resistance and intolerance issues have spurred the development of second-generation TKIs, revolutionizing the treatment landscape for patients with CML and offering renewed hope for improved outcomes.

Lymphoma. Malignant lymphoma is composed of a variety of types and is classified based on cell origin. This section summarizes some biomarkers that help in guiding disease management. One of the biomarkers commonly examined in lymphoma is CD30.⁵² CD30 is a membrane-bound ligand that is expressed in multiple forms of lymphoma, including diffuse large B-cell lymphoma (DLBCL).⁵² The signaling cascade of CD30 promotes cancer cell survival and proliferation and renders cancer cells resistant to self-destruction.⁵³ The US FDA approved the first ADC targeting CD30 in patients suffering from refractory Hodgkin lymphoma (HL) or anaplastic large cell lymphoma.^{54,55} Another biomarker rarely used in lymphomas is ALK.⁵⁶ The ALK fusion is usually observed in NSCLC, and it has been a target of interest in this population.⁵⁷ The ALK fusion has been reported to be expressed in DLBCL, T-cell lymphomas, and other types of lymphoma.³⁸ The presence of this gene alteration is attributed to worse prognosis in patients with lymphoma and is usually observed in patients who fail standard of care treatments.³⁸ Multiple ALK-targeted agents, such as alectinib, brigatinib, crizotinib, ceritinib, and lorlatinib, have been approved. The majority of those approved

agents were in the setting of NSCLC.⁵⁸ According to the NCCN guidelines, alectinib, brigatinib, ceritinib, and lorlatinib can be used for patients with lymphoma with ALK-expressing lymphoma if the first-line treatment fails.⁵⁹⁻⁶³ The identification of such biomarkers has significantly advanced the management of malignant lymphomas, offering targeted treatment options for patients with refractory disease.

Overcoming resistance to targeted therapy

Resistance to treatment is a problem that is often encountered with targeted cancer therapy, making it important to explore treatment strategies to overcome potential therapy failure. Resistance to targeted therapy may be adaptive, as cancer cells are able to overcome the signal blockade caused by TKIs through the use of different downstream cellular pathways.⁶⁴ Adaptive resistance is often observed in BRAF^{V600}-mutated melanoma if BRAF-directed therapy is used alone, reactivation of downstream signaling through the RAS-MAPK pathway occurs, resulting in treatment resistance. This highlights the importance of using combination therapy, such as the combination of dabrafenib and trametinib, to block multiple targets in the MAPK pathway to prevent the development of adaptive resistance.⁶⁵ On the contrary, acquired resistance occurs in patients who were previously responsive to treatment for a period of time; it can occur as a result of new mutations or gene amplification, leading to eventual disease progression.⁶⁴ Achieving a deep initial response is a strategy that aims to lower the chances of resistance development; this strategy is employed with hematological cancers such as CML.⁶⁴ The efficacy of chronic-phase CML treatment is assessed through molecular and cytogenetic milestones that correlate with a deep response, which is linked to decreased disease progression and improved survival.⁶⁶ Next-generation sequencing (NGS) serial tests are another strategy that has been successfully used to identify changes in tumor mutation expression once disease progression occurs, which can aid in tailoring new personalized treatment plans on the basis of molecular resistance patterns.^{67,68} As a result of this growing application, ESMO recently published a statement recommending performing NGS testing in various advanced cancer types, including cancers of unknown origin.⁶⁹

Predictive Biomarkers in Immunotherapy

The introduction of immune checkpoint inhibitors (ICPi) has dramatically changed the cancer treatment landscape.^{70,71} In 2011, ipilimumab, the first ICPi approved by the FDA, was used for the treatment of advanced-stage melanoma.⁷² Shortly thereafter, programmed death 1 (PD-1) inhibitors, including pembrolizumab and nivolumab, were introduced to the market, followed by the PD-1 ligand (PD-L1) inhibitors atezolizumab, durvalumab, and avelumab.⁷³⁻⁷⁸ Currently, PD-1/PD-L1 inhibitors are approved for the treatment of a wide range of solid tumors, including but not limited to melanoma, head and

neck, gastrointestinal, lung, breast, renal, and genitourinary cancers.⁷⁹ With the approval of pembrolizumab for the treatment of relapsed/refractory classical HL, indications have also expanded to include hematological malignancies after showing longer PFS in comparison with the standard of care.⁸⁰ Several biomarkers have been used to predict the response to ICPis; this section discusses the most recent advancements and challenges in this field.

Programmed death ligand 1

The PD-1/PD-L1 axis was first discovered in the early 1990s, and extensive research revealed its central role in immune tolerance.⁸¹ The PD-1 is a transmembrane protein expressed on activated immune white blood cells, including B and T lymphocytes, whereas PD-L1 is expressed on lymphocytes as well as various other body tissues and organs.⁸²⁻⁸⁵ The interaction between PD-1 and PD-L1 inactivates the immune response. Malignant cells overexpressing PD-L1 escape immune system surveillance, allowing them to continue to grow and proliferate.⁸⁶ Pembrolizumab was the first PD-1 inhibitor to gain accelerated FDA approval in 2014 for the treatment of unresectable melanoma.⁸⁷

The PD-L1 expression detection through immunohistochemistry (IHC) is the most commonly used biomarker detection method to predict response to immunotherapy.⁸⁸ The PD-L1 biomarker detection has now received approval for various types of cancers.⁸⁹ Despite its wide use, PD-L1 IHC detection has been shown to be less sensitive than other PD-L1 detection methods.⁹⁰ In addition, the absence of PD-L1 expression does not rule out a response to immunotherapy, as noted in both CheckMate238 and KEYNOTE-054, where a recurrence-free survival benefit was observed in all patients regardless of PD-L1 expression status.^{91,92} Moreover, IMPOWER-133 demonstrated an OS benefit in extensive-stage small-cell lung cancer patients with the addition of atezolizumab to etoposide and carboplatin chemotherapy backbone across all patients, including those with PD-L1 expression $\leq 1\%$.⁹³ These observations show variability in the thresholds for PD-L1 positivity as well as a great deal of heterogeneity in PD-L1 expression cutoffs between different trials and disease states, raising the need for additional research on alternative biomarkers to better predict immunotherapy responses.

Tumor mutational burden

Tumor mutational burden (TMB) refers to the extent of genetic derangements found in cancer cells.^{94,95} Genetic abnormalities may lead to the activation of oncogenes, deactivation of tumor suppressor genes, production of abnormal proteins, and expression of neoantigens on the cell surface.⁹⁴ The TMB is detected clinically using NGS technology through testing cancer tissues for the number of somatic mutations per

megabase (mut/Mb).⁹⁶ The US FDA approved 2 NGS panels for TMB testing in clinic: F1CDx and MSK-IMPACT.⁹⁶ The TMB was approved based on the results of the KEYNOTE-158 trial, where a subgroup of patients with TMB high (TMB-H) (defined as ≥ 10 mut/Mb) demonstrated an overall response and a complete response benefit.⁹⁷ Furthermore, a post hoc analysis of the TMB data from 12 trials using whole exome sequencing further confirmed a cutoff of 10 mut/Mb as an indicator of the response to ICPis.⁹⁸

The use of the TMB as a predictor of response to ICPi therapy is challenging in some tumors that exhibit high microsatellite instability (MSI-H) as MSI creates a greater mutational burden compared with that of MSI-stable tumors, making it difficult to set a pan-cancer threshold.^{99,100} Furthermore, TMB status failed to predict response in certain tumors, including but not limited to breast, prostate, and adrenocortical cancer.¹⁰¹ The aforementioned observations represent challenges with the utility of TMB as a treatment response biomarker.

Recent *in vitro* research demonstrated that mutations occurring in a single-copy gene region or mutations with multiple copies were less likely to be eliminated from the cancer genome and are therefore denoted as persistent TMB (pTMB).¹⁰² Persistent mutations are associated with persistent mutation-associated neoantigen expression, which allows for durable recognition of the tumor by the immune system during immunotherapy. The utility of pTMB as a predictor of response to immunotherapy has been observed in a number of cancers, including melanoma, NSCLC, and head and neck cancers, thus indicating that the pTMB is promising for predicting response to immunotherapy.¹⁰²

Microsatellite instability

Gene microsatellites are short sequences of noncoding DNA that are typically composed of 1 to 6 base pairs that form during the DNA replication process.^{103,104} The cellular mismatch repair (MMR) system works to correct such errors that occur during replication and failure to correct replication errors results in the accumulation of short tandem repeats, which leads to MSI and ultimately tumor development.^{105,106} The MSI-high tumors are common in gastrointestinal (including colorectal) and endometrial cancers, making MSI a valuable biomarker for predicting the response to ICPi therapy.¹⁰⁶ Several methods are currently used to assess MSI/MMR status, including NGS, PCR, and IHC.¹⁰⁷ Interestingly, a new approach using machine learning (ML) models for magnetic resonance imaging (MRI) has been successfully used to predict MSI status in patients with rectal cancer, although further studies are needed; this approach represents a promising non-invasive diagnostic tool for patients with rectal cancer.¹⁰⁸ Moreover, PD-1 blockade with dostarlimab in patients with MMR-deficient locally advanced rectal cancer achieved a

100% complete response rate in patients who completed 1 year of dostarlimab treatment in a phase 1 study.¹⁰⁹ Currently, a phase 2 ongoing multicenter global trial is investigating the efficacy of dostarlimab on a larger scale in patients with MSI/MMR-deficient locally advanced rectal cancer and is expected to release preliminary results by 2026.¹¹⁰

AI Models in Response Prediction

In the field of clinical oncology and cancer research, there are at least 4 main types of biomarkers. First, susceptibility or risk biomarkers highlight an individual's likelihood of developing cancer. Second, diagnostic markers are used to determine whether cancer is present or absent. Third, prognostic biomarkers offer insights into the potential course of the disease, predicting its progression or recurrence. Finally, stratification markers help categorize cancer diagnoses into distinct molecular groups, facilitating tailored treatment plans and providing a clearer picture of possible disease trajectories.¹¹¹ Traditional histopathological methods, including IHC of solid tissues, remain the preferred approach for cancer diagnosis. However, the emerging field of liquid biopsy biomarkers holds significant potential for enhanced detection of early-stage, recurrent, or undetected cancers.¹¹¹ The rapid advancement of in-depth sequencing techniques and the analysis of genomes and transcriptomes from clinical samples form the foundation of precision medicine, a vision not yet fully achieved.¹⁵ Due to technological progress in DNA and RNA analysis, nucleic acid-derived biomarkers have taken the forefront. Biomarkers can manifest in various forms—molecular, histological, radiographic, or physiological. While many biomarkers are based on observable clinical symptoms, some are derived from physiological measurements or laboratory tests.^{15,111} Biomarkers that can be quantified depend heavily on established reference ranges to guide clinical decisions. Interestingly, most of the biomarkers used in current oncology are not quantifiable in a continuous manner. Genomic biomarkers, for instance, which are frequently used to forecast treatment outcomes or disease prognosis, typically present binary data. In addition, interpretations of histological, radiological, and MRI imaging biomarkers often involve subjective assessments by medical professionals.¹¹²

In the realm of oncology, emerging technologies such as AI and ML are being developed.¹¹³ Their strength lies in analyzing intricate nonlinear patterns in vast multidimensional datasets, a critical capability for contemporary clinical practices that require holistic analysis of real-world and multiomics data.¹¹⁴ The ML excels at fusing information about both the patient and the tumor, enhancing the precision of predictive biomarkers. This leads to more tailored treatments and helps identify patients who could most benefit from immuno-oncology.¹¹⁵ A number of ML models have been generated to classify NSCLC into its well-established subtypes and/or identify crucial NSCLC biomarkers that are differentially expressed across subtypes.^{116,117} If adequate information is provided, deep

learning models have proven to be superior to traditional ML algorithms, but the users of these models are still unable to recognize how different features contribute to the task at hand. The explainable AI (XAI) concept was recently introduced in an effort to fill the explainability gap in deep models.¹¹⁸ The XAI is a set of tools or techniques that aid developers in understanding the internal functions of a machine/deep learning model.¹¹⁸ The XAI identified a small number of clinically significant NSCLC biomarkers for possible use in targeted therapy. Fifty-two biomarkers (NSCLC-Biomarkers-Set) were identified using a deep learning framework that was created using the XAI algorithms GradientSHAP, IntegratedGradients, and DeepLIFT. Forty-five of the 52 biomarkers discovered using the suggested framework have already been identified in previous studies. The 7 genes identified by XAI include AP2M1, C9orf69, FLJ44635, ID2B, CEL, LOC442308, and LOC728758.¹¹⁸ These 7 genes have not yet been documented for subtyping NSCLC and could be the focus of additional studies by clinicians with the aim of developing targeted therapy for patients with NSCLC.¹¹⁸ The proposed framework's XAI-based feature selection method outperformed other feature selection methods in terms of classification accuracy. Moreover, 28 biomarkers with P values $\leq .05$ were found to be helpful for predicting survival outcomes, and 14 of the newly discovered biomarkers may be druggable.¹¹⁸

AI in digital pathology

The AI has been used in digital pathology to improve cancer diagnosis and prognostication. In one case study, a multimodal AI architecture was developed to analyze digital histopathology images and clinicopathologic data from patients with prostate cancer. This AI model was trained on data from multiple clinical trials and successfully predicted clinical outcomes such as biochemical recurrence and OS, showcasing its potential in precision oncology.^{119,120}

AI and DNA methylation analysis

Another example involves the use of AI in conjunction with DNA methylation analysis for lung cancer detection. Researchers applied AI algorithms, including support vector machines and deep learning, to analyze methylation changes in circulating cell-free tumor DNA. This approach identified potential biomarkers for lung cancer with high diagnostic accuracy, demonstrating AI's capability in enhancing early detection and understanding cancer pathogenesis.¹²¹

AI in breast cancer diagnosis

Machine learning and deep learning techniques have been extensively reviewed for their application in breast cancer diagnosis and classification through medical imaging. These AI methodologies have improved the accuracy and efficiency of

detecting and classifying breast cancer, providing valuable insights into disease progression and treatment planning.¹²²

Identification of novel biomarkers in precision oncology

A specific example of a biomarker identified through AI methodologies in precision oncology is the sphingomyelin (d18:0/22:0), which has been identified as a potential screening biomarker for lung cancer. This biomarker was discovered through a metabolomics study that analyzed metabolic perturbations related to lung cancer.¹²³ The study involved a nested case-control analysis within the Cancer Prevention Study cohorts, demonstrating the association of sphingomyelin (d18:0/22:0) with lung cancer risk. Another example is the identification of the biomarker VIPR1 in hepatocellular carcinoma (HCC) using ML algorithms. This biomarker was identified through the analysis of gene expression data and was found to have significant diagnostic value for HCC, showing potential as an independent prognostic factor.¹²⁴ These examples illustrate how AI and ML can be used to identify novel biomarkers that have significant implications for cancer diagnosis and treatment.

AI in multi-omics integration

The AI has been pivotal in integrating multiomics data, which include genomics, transcriptomics, proteomics, and metabolomics, to enhance cancer diagnosis and treatment. For instance, AI models have been used to analyze multimodal datasets, leading to improved cancer subtyping, risk stratification, and prognostication. A study demonstrated the use of AI to integrate multiomics data, which resulted in better diagnostic and therapeutic outcomes by identifying distinct cancer subtypes and predicting patient responses to treatments.¹²⁵

Multiomics and AI in Cancer

The aim of multiomics methods is to better understand the molecular and clinical characteristics of cancers by integrating numerous omics datasets collected from the same patients into unique frameworks. Currently, there are novel opportunities to further classify cancers into subtypes, enhance the prognosis and therapeutic outcome of these subtypes, and comprehend important pathophysiological processes through various molecular layers due to a wide range of emerging omics and multiview clustering algorithms. A multiomics analysis is a data-driven scientific study that aims to uncover the complexity of cells and their surroundings by analyzing a variety of high-dimensional datasets at several levels and sizes. The advent of high-throughput technologies in genomics and transcriptomics, coupled with increased efforts in large-scale research collaboration and computational algorithm innovation, has led to a paradigm change in cancer research toward multiomics approaches.¹²⁶⁻¹²⁸ In cancer, AI has proven to be

capable of analyzing complementary multimodal datasets. The advancement of cancer precision medicine has accelerated with the simultaneous development of AI algorithms and multiomics technology.¹²⁹ Artificial intelligence techniques are frequently used at different stages of clinical cancer treatment, as they have shown success in addressing the heterogeneity and complexity of multiomics data. Currently, AI algorithms are able to combine data from various platforms, such as radiomics, genomics, transcriptomics, proteomics, metabolomics, and pathomics, to enable more accurate identification of cancer subtypes and to suggest reliable tools for predicting cancer prognosis and response to treatment.^{130,131} This introduced the role of AI-based multiomics analysis to aid in cancer precision medicine, including cancer detection and screening, diagnosis, classification, and grading.¹²⁹

Cancer Biomarker Trends in Middle Eastern Populations

Validated biomarkers of genetic alterations are becoming key tools for disease prevention, disease prognosis, and therapy selection in oncology settings. These mutations are classified as either germline static or somatic dynamic mutations. Germline mutations are found in germ cells (detected in both normal and tumor cells) and have a role in identifying genetic predisposition and guiding screening and treatment. Somatic mutations are found only in nonreproductive cells and have a role in identifying predictive/prognostic mutations for disease prognosis and therapy selection.^{132,133} There are 53 FDA-recognized genes for predicting response to 87 cancer drugs and an additional 24 genes considered a standard of care and recognized in professional guidelines that show a predicted response to 45 FDA-approved drugs. The majority of these therapeutic biomarkers are somatic mutations, such as ALK rearrangement, BRAF, EGFR, RET rearrangements, and ROS-1¹³⁴ (Table 1).

The evidence investigating the prevalence of cancer biomarkers in the MENA region is limited.¹³⁵ In Saudi Arabia, a report showed the testing rate for the most common biomarkers assessed by cancer type¹³⁶ (Figure 3). In addition, Alharbi and colleagues revealed the prevalence of KRAS mutations among 248 Saudi patients diagnosed with CRC (49.6%), followed by NRAS mutations (2%) and BRAF mutations (0.4%). The survival rate is worse among carriers of KRAS mutations than among carriers of wild-type KRAS tumors.¹³⁷ This finding was consistent with more recent data from local and MENA region studies. Nasser and colleagues discovered several novel KRAS mutations in Saudi patients diagnosed with CRC.¹³⁸⁻¹⁴⁰ On the contrary, Rasool and colleagues reported novel mutations in the BRAF gene at c.1758delA, c.1826insT, c.1860insA, and c.1860insA/C in Saudi patients with CRC.¹⁴¹ Such mutation uniqueness could be attributed to environmental factors, among other factors, such as ethnic/racial variation, that reflect different gene methylation patterns and dissimilar molecular pathogenesis.^{140,142-144}

Investigational efforts explored the spectrum and frequency of germline variants through NGS-based multigene testing (180 genes associated with cancer predisposition) in Bahraini women with breast cancer. Their work detected 2 BRCA1 variants in 2 patients, namely, the missense variant c.287A > G (p.Asp96Gly) and the truncating variant c.1066C > T (p.Gln356Ter). Moreover, they yielded additional genomic information on low-penetrance genes, where 4 patients carried pathogenic/likely pathogenic or potential pathogenic variants together with distinct other variants of unknown significance.¹⁴⁵ Such efforts and findings can serve as supplementary knowledge to traditional genetic counseling.

Zooming into BRCA1 and BRCA 2 gene mutations, a double-strand DNA repair homologous mutation that showed a positive association in patients treated with PARP inhibitors.¹⁴⁵⁻¹⁴⁷ In addition, it has value in predicting response to platinum therapy.¹⁴⁸ An analysis of a large cohort from the Middle Eastern population revealed 2 novel founder mutations that account for 46.4% of all BRCA mutant cases and 1.6% of all breast cancer cases. Understanding the mutation distribution in the Middle Eastern population is exceptionally important for developing cost-effective genetic testing strategies in clinical practice.^{149,150} Regarding the clinical impact on patient outcome, in a small cohort of 61 Saudi female patients who were diagnosed with primary ovarian, fallopian tube, or peritoneal carcinoma, the results were consistent with those of previous studies showing a better response to chemotherapy owing to the deficiency of DNA repair mechanisms and increased DFS and OS associated with BRCA gene mutations, resulting in an 8-month increase in DFS in the BRCA mutant group, whose patients received either first- or second-line treatment.¹⁵¹

Embedding Cancer Biomarkers and Precision Oncology Within Health Systems

Integrating precision oncology into the standard of care seems to be at the top of the health care collective agenda. It has emerged as a priority not only because of its pivotal role in advancing patient care but also because of its potential to preserve health care resources while allocating and using cutting edge resources. Biomarker assays, often called companion diagnostics, are good examples of emerging clinical tests that are essential for identifying individuals who are expected to benefit from therapy. Currently, biomarker assays are needed to meet the needs of targeted therapy production and use in practice.¹⁵²

The Early Detection Research Network (EDRN) marked the threshold for the qualification of biomarkers during the development process to be inexpensive, robust, and translatable to guide risk assessment, detection, diagnosis, prognosis, and response to therapy. Approvals by regulatory agencies in conjunction with the drugs they are paired with are essential for the transition from research to practice.¹⁵³ Thus, a shift in the

Testing Rate of Common Biomarkers in Saudi Arabia

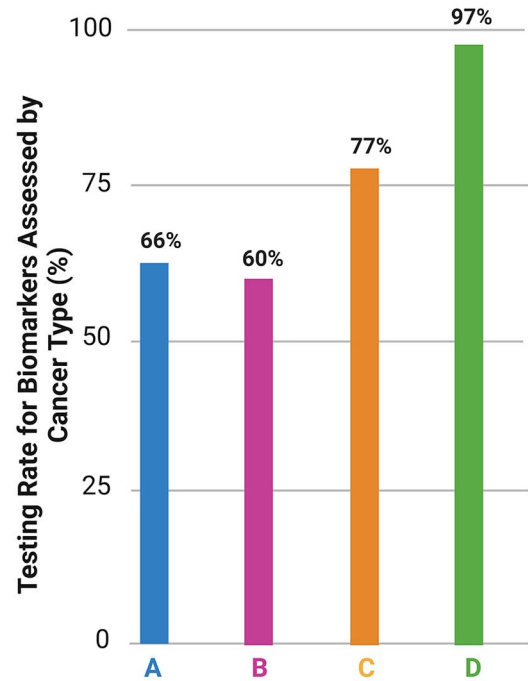
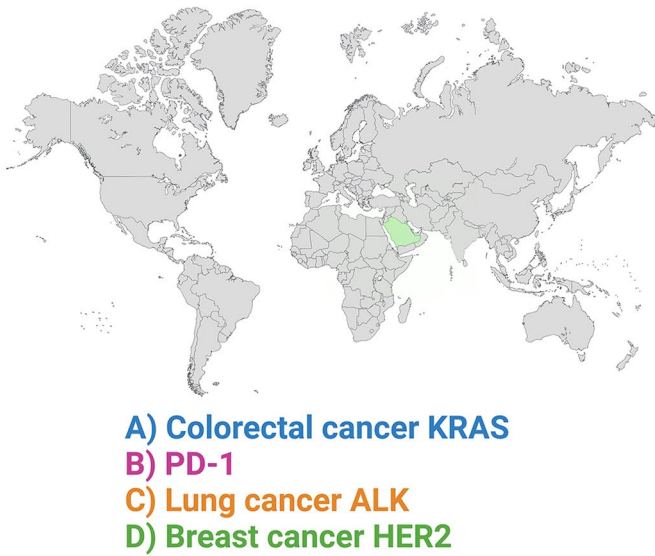


Figure 3. Common cancer biomarkers detected among Saudi patients. Testing rate for the most common biomarkers assessed by cancer type; breast cancer HER2 (97%), NSCLC ALK (77%), and PD1 (60%), and in colon cancer KRAS (~66%).

cognitive framework toward precision oncology in medicine and research demands redefining cancers on the basis of their molecular characteristics in addition to their organ of origin. Some enablers for this shift are the industrial development model, the development of sustainable reimbursement models that cover equal access high-throughput genomic sequencing of tumor genomes, and the engagement of stakeholders and regulatory agencies responsible for drug approvals to reinforce the adoption of genetic data from the early stages of drug approval all the way to the bedside.^{153,154} One of the success stories is the accelerated development of widely used targeted therapy due to the substantial results in early clinical trial phases. For instance, crizotinib, a TKI targeting ALK, was approved by the FDA in 2011 for the treatment of ALK-positive relapsed or refractory systemic anaplastic large-cell lymphoma in pediatric patients aged 1 year and older.⁸

An example of an initiative that paved the way for embedding cancer biomarker testing in practice was launching the Program for the Assessment of Clinical Cancer Tests (PACCT) by the National Cancer Institute (NCI) in 2000, which aimed to ensure the translation of new knowledge about cancer and new technologies into clinical practice and to develop more informative laboratory tools to help maximize the impact of cancer treatments.¹⁵⁵ Another program released in 2012 by the NCI was the Clinical Assay Development Program, which targeted the development of assays for therapy response prediction, mainly for use in clinical trials.¹⁵⁶

Such advances in molecular profiling have improved not only the genomic classification of a patient's tumor but also the

development, approval, and availability of precision therapies.¹⁵⁷ However, there are still unleashed possibilities for analyzing all cancer-associated genetic alterations in a single assay and germline and somatic mutations, such as with NGS.¹⁵⁷

On the contrary, there could be some challenges hindering the routine adoption of cancer biomarkers in practice. For instance, it is difficult to determine the associations among biological data, disease, and clinical translation to make meaningful medical decisions. The threshold for the lowest level of evidence acceptable for a variant allele to become a viable biomarker. Another major challenge is the translation of biomarkers from laboratory identification/isolation to clinical use and interpretation.^{8,158}

Challenges in Biomarker Development

Although the number of ongoing research efforts to establish clinically relevant biomarkers has increased significantly over the years, very few biomarkers have been used clinically. This can be attributed to several pitfalls during the identification and validation process. Studies with flaws in design and execution may fail to detect significant results. For example, collecting few samples or failing to match age and sex between healthy volunteers and patients could result in failure of biomarker identification.¹⁵⁹ Tumor heterogeneity represents a challenge in the area of biomarker discovery; cancer cells harbor genetic mutations as they replicate, making distinguishing driver mutations from passenger mutations difficult.^{4,160} The intertumor variability of biomarker cutoff values, such as TMB in ICPi therapy, makes standardizing cutoffs among various cancer

types difficult. Analyzing other biomarkers in the tumor micro-environment may aid in reaching a more accurate prediction of treatment response; however, this remains a growing area of research.¹⁵⁹ In addition, laboratory assays that fail to achieve adequate sensitivity and specificity will hinder the process of biomarker validation.¹⁵⁹ Problems encountered during sample collection and transportation could affect the quality of samples. Rigorous quality control measures must be followed during all stages from sample collection, testing, storage, and analysis. Guidelines and standards set by EDRN and PACCT help in streamlining the research and validation processes.¹⁵³

Future Directions

The rapidly developing field of cancer biomarkers holds a promising future, as many novel concepts have been introduced for the purpose of improving cancer detection and treatment. A novel development in the cancer treatment pipeline is the use of chimeric receptor antigen T-cell therapy (CAR-T) in solid tumors. The CAR-T cells were first approved in 2017 for hematological malignancies. Currently, multiple phase 1 and 2 CAR-T cell therapies targeting biomarkers including HER-2 and EGFR in solid tumors are in progress.¹⁶¹

The area of ADCs has experienced many advancements since the introduction of the first ADC, gemtuzumab ozogamicin in 2000. Several clinical trials are currently evaluating ADCs that target C-mesenchymal epithelial transition factor (c-Met). The c-Met is a tyrosine kinase receptor found on the surface of various cancer cells, making it a potential target for ADC agents.¹⁶² Like ADCs, immunotoxins (IT) are antibody conjugates that selectively deliver toxins to cancer cells.¹⁶³ Tagraxofusp is an IT used for blastic plasmacytoid dendritic cell neoplasm and is the only available IT product available on the market. Tagraxofusp has shown positive outcomes in clinical trials. A new recombinant bispecific IT that targets CCR4-IL2 for the treatment of cutaneous T-cell lymphoma is being tested in animal models, and it has demonstrated efficacy and an acceptable safety profile in rats and minipigs.¹⁶⁴

Conclusion

This comprehensive review covers the pivotal role of cancer biomarkers in revolutionizing oncology, particularly through the lens of precision medicine and targeted therapies. This review highlights the importance of categorizing biomarkers into prognostic and predictive biomarkers and their significant impact on managing solid and hematologic cancers. Innovations in biomarker research have led to the development of personalized treatment regimens, enhancing efficacy and reducing the adverse effects of cancer therapies. Moreover, the integration of AI and multiomics approaches can refine the prediction of treatment responses and advance the implementation of precision oncology.

Acknowledgements

Not Applicable

Author Contributions

Maha AlDoughaim: article concept, author coordination, writing sections (3, 3.1, 3.2, 3.3, 8, 9).

Nada AlSuhebany: writing sections (2, 2.1, 2.2, 2.2.1, 2.2.2, 2.2.3) and formulation of Table 1, second proof reading and revision.

Mohammed AlZahrani: writing sections (2.3, 2.3.1, 2.3.2, 2.3.3) and formulation of Table 2.

Tariq AlQahtani: writing sections 4 and 5.

Sahar AlGhamdi: writing introduction and drawing of Figures 1, 2, and 3.

Hisham Badreldin: writing article conclusion, proof reading, and revision.

Hana Al AlShaykh: writing sections 6 and 7.

Availability of Data and Materials

Not applicable to this article as no datasets were generated or analyzed for this article

Consent for Publication

Not Applicable

Ethics Approval and Consent to Participate

Not Applicable

ORCID iD

Maha AlDoughaim  <https://orcid.org/0009-0008-7229-9474>

REFERENCES

- Smith AD, Roda D, Yap TA. Strategies for modern biomarker and drug development in oncology. *J Hematol Oncol*. 2014;7:70. doi:10.1186/s13045-014-0070-8
- Mäbert K, Cojoc M, Peitzsch C, Kurth I, Souchelnyskiy S, Dubrovskaya A. Cancer biomarker discovery: current status and future perspectives. *Int J Radiat Biol*. 2014;90:659-677. doi:10.3109/09553002.2014.892229
- Normanno N, Apostolidis K, de Lorenzo F, et al. Cancer biomarkers in the era of precision oncology: addressing the needs of patients and health systems. *Semin Cancer Biol*. 2022;84:293-301. doi:10.1016/j.semcancer.2021.08.002
- Ileana Dumbrava E, Meric-Bernstam F, Yap TA. Challenges with biomarkers in cancer drug discovery and development. *Expert Opin Drug Discov*. 2018;13:685-690. doi:10.1080/17460441.2018.1479740
- Hyman DM, Taylor BS, Baselga J. Implementing genome-driven oncology. *Cell*. 2017;168:584-599. doi:10.1016/j.cell.2016.12.015
- Siegel RL, Miller KD, Wagle NS, Jemal A. Cancer statistics, 2023. *CA Cancer J Clin*. 2023;73:17-48. doi:10.3322/caac.21763
- Rulten SL, Grose RP, Gatz SA, Jones JL, Cameron AJM. The future of precision oncology. *Int J Mol Sci*. 2023;24:12613. doi:10.3390/ijms241612613
- Saeed RF, Awan UA, Saeed S, et al. Targeted therapy and personalized medicine. *Cancer Treat Res*. 2023;185:177-205. doi:10.1007/978-3-031-27156-4_10
- Joo WD, Visintin I, Mor G. Targeted cancer therapy—are the days of systemic chemotherapy numbered. *Maturitas*. 2013;76:308-314. doi:10.1016/j.maturitas.2013.09.008
- Nwibo DD, Levi CA, Nwibo MI. Small molecule drugs; down but not out: a future for medical research and therapeutics. *IOSR J Dent Med Sci*. 2015;14:70-77. doi: 10.9790/0853-14117077
- Lee YT, Tan YJ, Oon CE. Molecular targeted therapy: treating cancer with specificity. *Eur J Pharmacol*. 2018;834:188-196. doi:10.1016/j.ejphar.2018.07.034
- Sievers EL, Senter PD. Antibody-drug conjugates in cancer therapy. *Annu Rev Med*. 2013;64:15-29. doi:10.1146/annurev-med-050311-201823
- Fu Z, Li S, Han S, Shi C, Zhang Y. Antibody drug conjugate: the “biological missile” for targeted cancer therapy. *Sig Transduct Target Ther*. 2022;7:1-25. doi:10.1038/s41392-022-00947-7
- Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell*. 2011;144:646-674. doi:10.1016/j.cell.2011.02.013

15. Sarhadi VK, Armengol G. Molecular biomarkers in cancer. *Biomolecules*. 2022;12:1021. doi:10.3390/biom12081021
16. Patel JN, Ersek JL, Kim ES. Lung cancer biomarkers, targeted therapies and clinical assays. *Transl Lung Cancer Res*. 2015;4:503-514. doi:10.3978/j.issn.2218-6751.2015.06.02
17. Li MM, Datto M, Duncavage EJ, et al. Standards and guidelines for the interpretation and reporting of sequence variants in cancer: a Joint Consensus Recommendation of the Association for Molecular Pathology, American Society of Clinical Oncology, and College of American Pathologists. *J Mol Diagn*. 2017;19:4-23. doi:10.1016/j.jmoldx.2016.10.002
18. National Comprehensive Cancer Network (NCCN) Guidelines. Non-small cell lung cancer version 1. Published 2024. Accessed January 15, 2024. https://www.Nccn.Org/Professionals/Physician_gls/Pdf/Nscl.Pdf
19. Hendriks LE, Kerr KM, Menis J, et al. Oncogene-addicted metastatic non-small-cell lung cancer: ESMO clinical practice guideline for diagnosis, treatment and follow-up. *Ann Oncol*. 2023;34:339-357. doi:10.1016/j.annonc.2022.12.009
20. Yu HA, Arcila ME, Rekhtman N, et al. Analysis of tumor specimens at the time of acquired resistance to EGFR-TKI therapy in 155 patients with EGFR-mutant lung cancers. *Clin Cancer Res*. 2013;19:2240-2247. doi:10.1158/1078-0432.CCR-12-2246
21. National Comprehensive Cancer Network (NCCN) Guidelines. Breast cancer version 5. Published 2023. Accessed January 15, 2024. https://www.Nccn.Org/Professionals/Physician_gls/Pdf/Breast.Pdf
22. Hu R, Hilakivi-Clarke L, Clarke R. Molecular mechanisms of tamoxifen-associated endometrial cancer (review). *Oncol Lett*. 2015;9:1495-1501. doi:10.3892/ol.2015.2962
23. Boér K. Fulvestrant in advanced breast cancer: evidence to date and place in therapy. *Ther Adv Med Oncol*. 2017;9:465-479. doi:10.1177/1758834017711097
24. Yang X, Wu D, Yuan S. Tyrosine kinase inhibitors in the combination therapy of HER2 positive breast cancer. *Technol Cancer Res Treat*. 2020;19:1533033820962140.
25. Robson M, Im S-A, Senkus E, et al. Olaparib for metastatic breast cancer in patients with a germline BRCA mutation. *N Engl J Med*. 2017;377:523-533. doi:10.1056/NEJMoa1706450
26. Rugo HS, Bardia A, Marmé F, et al. Overall survival with sacituzumab govitecan in hormone receptor-positive and human epidermal growth factor receptor 2-negative metastatic breast cancer (TROPICS-02): a randomised, open-label, multicentre, phase 3 trial. *Lancet*. 2023;402:1423-1433. doi:10.1016/S0140-6736(23)01245-X
27. Modi S, Jacot W, Yamashita T, et al. Trastuzumab deruxtecan in previously treated HER2-low advanced breast cancer. *N Engl J Med*. 2022;387:9-20. doi:10.1056/NEJMoa2203690
28. Meisel JL, Venur VA, Gnani M, Carey L. Evolution of targeted therapy in breast cancer: where precision medicine began. *Am Soc Clin Oncol Educ Book*. 2018;38:78-86. doi:10.1200/EDBK_201037
29. Braal CL, Jongbloed EM, Wilting SM, Mathijssen RHJ, Koolen SLW, Jager A. Inhibiting CDK4/6 in breast cancer with palbociclib, ribociclib, and abemaciclib: similarities and differences. *Drugs*. 2021;81:317-331. doi:10.1007/s40265-020-01461-2
30. Li WJ, Zhong SL, Wu YJ, et al. Systematic expression analysis of genes related to multidrug-resistance in isogenic docetaxel- and adriamycin-resistant breast cancer cell lines. *Mol Biol Rep*. 2013;40:6143-6150. doi:10.1007/s11033-013-2725-x
31. National Comprehensive Cancer Network (NCCN) Guidelines. Colon cancer version 4. Published 2023. Accessed January 15, 2024. https://www.Nccn.Org/Professionals/Physician_gls/Pdf/Colon.Pdf
32. Cunningham D, Humblet Y, Siena S, et al. Cetuximab monotherapy and cetuximab plus irinotecan in irinotecan-refractory metastatic colorectal cancer. *N Engl J Med*. 2004;351:337-345. doi:10.1056/NEJMoa033025
33. Douillard J-Y, Siena S, Cassidy J, et al. Randomized, Phase III trial of panitumumab with infusional fluorouracil, leucovorin, and oxaliplatin (FOLFOX4) versus FOLFOX4 alone as first-line treatment in patients with previously untreated metastatic colorectal cancer: the PRIME study. *J Clin Oncol*. 2010;28:4697-4705. doi:10.1200/JCO.2009.27.4860
34. The United States Food and Drug Administration (FDA). FDA approves fruquintinib in refractory metastatic colorectal cancer. Accessed January 15, 2024. <https://www.Fda.Gov/Drugs/Resources-Information-Approved-Drugs/Fda-Approves-Fruquintinib-Refractory-Metastatic-Colorectal-Cancer>
35. Dasari A, Lonardi S, Garcia-Carbonero R, et al; and FRESKO-2 Study Investigators. Fruquintinib versus placebo in patients with refractory metastatic colorectal cancer (FRESKO-2): an international, multicentre, randomised, double-blind, phase 3 study. *Lancet*. 2023;402:41-53. doi:10.1016/S0140-6736(23)00772-9
36. Li J, Qin S, Xu R-H, et al. Effect of fruquintinib vs placebo on overall survival in patients with previously treated metastatic colorectal cancer: the FRESKO randomized clinical trial. *JAMA*. 2018;319:2486-2496. doi:10.1001/jama.2018.7855
37. Sforza V, Martinelli E, Ciardiello F, et al. Mechanisms of resistance to anti-epidermal growth factor receptor inhibitors in metastatic colorectal cancer. *World J Gastroenterol*. 2016;22:6345-6361. doi:10.3748/wjg.v22.i28.6345
38. Shreenivas A, Janku F, Gouda MA, et al. ALK fusions in the pan-cancer setting: another tumor-agnostic target? *NPJ Precis Oncol*. 2023;7:101. doi:10.1038/s41698-023-00449-x
39. Jani CT, Ahmed A, Singh H, et al. Burden of AML, 1990-2019: estimates from the global burden of disease study. *JCO Glob Oncol*. 2023;9:e2300229. doi:10.1200/GO.23.00229
40. Small S, Oh TS, Plataniak LC. Role of biomarkers in the management of acute myeloid leukemia. *Int J Mol Sci*. 2022;23:14543. doi:10.3390/ijms232314543
41. Kiyoi H, Kawashima N, Ishikawa Y. FLT3 mutations in acute myeloid leukemia: therapeutic paradigm beyond inhibitor development. *Cancer Sci*. 2020;111:312-322. doi:10.1111/cas.14274
42. Lyman SD, James L, Vanden Bos T, et al. Molecular cloning of a ligand for the Flt3/Flk-2 tyrosine kinase receptor: a proliferative factor for primitive hematopoietic cells. *Cell*. 1993;75:1157-1167. doi:10.1016/0092-8674(93)90325-k
43. Negotei C, Colita A, Mitu I, et al. A review of FLT3 kinase inhibitors in AML. *J Clin Med*. 2023;12:6429. doi:10.3390/jcm12206429
44. Kulsoom B, Shamsi TS, Afsar NA, Memon Z, Ahmed N, Hasnain SN. Bax, Bcl-2, and Bax/Bcl-2 as prognostic markers in acute myeloid leukemia: are we ready for Bcl-2-directed therapy? *Cancer Manag Res*. 2018;10:403-416. doi:10.2147/CMAR.S154608
45. Qian S, Wei Z, Yang W, Huang J, Yang Y, Wang J. The role of BCL-2 family proteins in regulating apoptosis and cancer therapy. *Front Oncol*. 2022;12:985363. doi:10.3389/fonc.2022.985363
46. Aiman W, Ali MA, Basit MA, et al. Efficacy and tolerability of isocitrate dehydrogenase inhibitors in patients with acute myeloid leukemia: a systematic review of clinical trials. *Leuk Res*. 2023;129:107077. doi:10.1016/j.leukres.2023.107077
47. Liu X, Gong Y. Isocitrate dehydrogenase inhibitors in acute myeloid leukemia. *Biomark Res*. 2019;7:22. doi:10.1186/s40364-019-0173-z
48. Issa GC, DiNardo CD. Acute myeloid leukemia with IDH1 and IDH2 mutations: 2021 treatment algorithm. *Blood Cancer J*. 2021;11:107. doi:10.1038/s41408-021-00497-1
49. Braun TP, Eide CA, Druker BJ. Response and resistance to BCR-ABL1-targeted therapies. *Cancer Cell*. 2020;37:530-542. doi:10.1016/j.ccell.2020.03.006
50. Nowell PC, Hungerford DA. Chromosome studies on normal and leukemic human leukocytes. *J Natl Cancer Inst*. 1960;25:85-109.
51. Jabbour E. Chronic myeloid leukemia: first-line drug of choice. *Am J Hematol*. 2016;91:59-66. doi:10.1002/ajh.24249
52. Nakashima M, Uchamaru K. CD30 expression and its functions during the disease progression of adult T-cell leukemia/lymphoma. *Int J Mol Sci*. 2023;24:8731. doi:10.3390/ijms24108731
53. Dumitru AV, Tăpoi DA, Halcu G, et al. The polyvalent role of CD30 for cancer diagnosis and treatment. *Cells*. 2023;12:1783. doi:10.3390/cells12131783
54. Richardson NC, Kasamon YL, Chen H, et al. FDA approval summary: brentuximab vedotin in first-line treatment of peripheral T-cell lymphoma. *Oncologist*. 2019;24:e180-e187. doi:10.1634/theoncologist.2019-0098
55. Crosswell HE, LaCasce AS, Bartlett NL, et al. Brentuximab vedotin with chemotherapy in adolescents and young adults with stage III or IV classical Hodgkin lymphoma in ECHELON-1. *Haematologica*. 2024;109:982-987. doi:10.3324/haematol.2023.283303
56. Sun R, Medeiros LJ, Young KH. Diagnostic and predictive biomarkers for lymphoma diagnosis and treatment in the era of precision medicine. *Mod Pathol*. 2016;29:1118-1142. doi:10.1038/modpathol.2016.92
57. Parvaresh H, Roozitalab G, Golandam F, Behzadi P, Jabbarzadeh Kaboli P. Unraveling the potential of ALK-targeted therapies in non-small cell lung cancer: comprehensive insights and future directions. *Biomedicines*. 2024;12:297. doi:10.3390/biomedicines12020297
58. Lin JJ, Riey GJ, Shaw AT. Targeting ALK: precision medicine takes on drug resistance. *Cancer Discov*. 2017;7:137-155. doi:10.1158/2159-8290.CD-16-1123
59. Fukano R, Mori T, Sekimizu M, et al. Alectinib for relapsed or refractory anaplastic lymphoma kinase-positive anaplastic large cell lymphoma: an open-label phase II trial. *Cancer Sci*. 2020;111:4540-4547. doi:10.1111/cas.14671
60. Chihara D, Wong S, Feldman T, et al. Outcome of patients with relapsed or refractory anaplastic large cell lymphoma who have failed brentuximab vedotin. *Hematol Oncol*. 2019;37:35-38. doi:10.1002/hon.2560
61. Richly H, Kim TM, Schuler M, et al. Ceritinib in patients with advanced anaplastic lymphoma kinase-rearranged anaplastic large-cell lymphoma. *Blood*. 2015;126:1257-1258. doi:10.1182/blood-2014-12-617779
62. Wirk B, Malysz J, Choi E, Songdej N. Lorlatinib induces rapid and durable response in refractory anaplastic lymphoma kinase-positive large B-cell lymphoma. *JCO Precis Oncol*. 2023;7:e2200536. doi:10.1200/PO.22.00536
63. NCCN. T-cell lymphomas (version 3). Published 2021. https://www.Nccn.Org/Professionals/Physician_gls/Pdf/t-Cell.Pdf
64. Vasan N, Baselga J, Hyman DM. A view on drug resistance in cancer. *Nature*. 2019;575:299-309. doi:10.1038/s41586-019-1730-1
65. Gumusay O, Vitiello PP, Wabl C, Corcoran RB, Bardelli A, Rugo HS. Strategic combinations to prevent and overcome resistance to targeted therapies in oncology. *Am Soc Clin Oncol Educ Book*. 2020;40:e292-e308. doi:10.1200/EDBK_280845
66. Hehlmann R, Lauseker M, Sauße S, et al. Assessment of imatinib as first-line treatment of chronic myeloid leukemia: 10-year survival results of the randomized CML study IV and impact of non-CML determinants. *Leukemia*. 2017;31:2398-2406. doi:10.1038/leu.2017.253

67. Park JJ, Chu A, Li J, et al. Repeat next-generation sequencing testing on progression in men with metastatic prostate cancer can identify new actionable alterations. *JCO Precis Oncol*. 2024;8:e2300567. doi:10.1200/PO.23.00567
68. Colomer R, Miranda J, Romero-Laorden N, et al. Usefulness and real-world outcomes of next generation sequencing testing in patients with cancer: an observational study on the impact of selection based on clinical judgement. *EClinicalMedicine*. 2023;60:102029. doi:10.1016/j.eclinm.2023.102029
69. Mosele MF, Westphalen CB, Stenzinger A, et al. Recommendations for the use of next-generation sequencing (NGS) for patients with advanced cancer in 2024: a report from the ESMO Precision Medicine Working Group. *Ann Oncol*. 2024;35:588-606. doi:10.1016/j.annonc.2024.04.005
70. Robert C. A decade of immune-checkpoint inhibitors in cancer therapy. *Nat Commun*. 2020;11:3801. doi:10.1038/s41467-020-17670-y
71. Xin Yu J, Hubbard-Lucey VM, Tang J. Immuno-oncology drug development goes global. *Nat Rev Drug Discov*. 2019;18:899-900. doi:10.1038/d41573-019-00167-9
72. Hodi FS, O'Day SJ, McDermott DF, et al. Improved survival with ipilimumab in patients with metastatic melanoma. *N Engl J Med*. 2010;363:711-723. doi:10.1056/NEJMoa1003466
73. Davis AA, Patel VG. The role of PD-L1 expression as a predictive biomarker: an analysis of all US Food and Drug Administration (FDA) approvals of immune checkpoint inhibitors. *J Immunother Cancer*. 2019;7:278. doi:10.1186/s40425-019-0768-9
74. Garon EB, Hellmann MD, Rizvi NA, et al. Five-year overall survival for patients with advanced non-small-cell lung cancer treated with pembrolizumab: results from the phase I KEYNOTE-001 study. *J Clin Oncol*. 2019;37:2518-2527. doi:10.1200/JCO.19.00934
75. Raedler LA. Opdivo (Nivolumab): second PD-1 inhibitor receives FDA approval for unresectable or metastatic melanoma. *Am Health Drug Benefits*. 2015;8:180-183.
76. Felip E, Altorki N, Zhou C, et al; and IMPower010 Investigators. Adjuvant atezolizumab after adjuvant chemotherapy in resected stage IB-IIIa non-small-cell lung cancer (IMPower010): a randomised, multicentre, open-label, phase 3 trial. *Lancet*. 2021;398:1344-1357. doi:10.1016/S0140-6736(21)02098-5
77. Antonia Scott J, Augusto V, Davey D, et al. Durvalumab after chemoradiotherapy in stage III non-small-cell lung cancer. *N Engl J Med*. 2017;377:1919-1929. doi:10.1056/NEJMoa1709937
78. Kaufman HL, Russell JS, Hamid O, et al. Updated efficacy of avelumab in patients with previously treated metastatic merkel cell carcinoma after ≥ 1 year of follow-up: JAVELIN Merkel 200, a phase 2 clinical trial. *J Immunother Cancer*. 2018;6:7. doi:10.1186/s40425-017-0310-x
79. Vaddepally RK, Kharel P, Pandey R, Garje R, Chandra AB. Review of indications of FDA-approved immune checkpoint inhibitors per NCCN guidelines with the level of evidence. *Cancers*. 2020;12:738. doi:10.3390/cancers12030738
80. Kuruvilla J, Ramchandren R, Santoro A, et al; and KEYNOTE-204 Investigators. Pembrolizumab versus brentuximab vedotin in relapsed or refractory classical hodgkin lymphoma (KEYNOTE-204): an interim analysis of a multicentre, randomised, open-label, phase 3 study. *Lancet Oncol*. 2021;22:512-524. doi:10.1016/S1470-2045(21)00005-X
81. Bardhan K, Anagnostou T, Boussiotis VA. The PD1:PD-L1/2 pathway from discovery to clinical implementation. *Front Immunol*. 2016;7:550. doi:10.3389/fimmu.2016.00550
82. Jiang X, Wang J, Deng X, et al. Role of the tumor microenvironment in PD-L1/PD-1-mediated tumor immune escape. *Mol Cancer*. 2019;18:10. doi:10.1186/s12943-018-0928-4
83. Dong H, Zhu G, Tamada K, Chen L. B7-H1, a third member of the B7 family, co-stimulates T-cell proliferation and interleukin-10 secretion. *Nat Med*. 1999;5:1365-1369. doi:10.1038/70932
84. Keir ME, Butte MJ, Freeman GJ, Sharpe AH. PD-1 and its ligands in tolerance and immunity. *Annu Rev Immunol*. 2008;26:677-704. doi:10.1146/annurev.immunol.26.021607.090331
85. Yamazaki T, Akiba H, Iwai H, et al. Expression of programmed death 1 ligands by murine T cells and APC. *J Immunol*. 2002;169:5538-5545. doi:10.4049/jimmunol.169.10.5538
86. Iwai Y, Ishida M, Tanaka Y, Okazaki T, Honjo T, Minato N. Involvement of PD-L1 on tumor cells in the escape from host immune system and tumor immunotherapy by PD-L1 blockade. *Proc Natl Acad Sci USA*. 2002;99:12293-12297. doi:10.1073/pnas.192461099
87. KEYTRUDA® (Pembrolizumab) for injection prescribing information. Merck Sharp & Dohme Corp. Published September 2014. www.Accessdata.Fda.Gov/Drugsatfda_docs/Label/2014/125514lbl.pdf
88. Anagnostou V, Landon BV, Medina JE, Forde P, Velculescu VE. Translating the evolving molecular landscape of tumors to biomarkers of response for cancer immunotherapy. *Sci Transl Med*. 2022;14:eabo3958. doi:10.1126/scitranslmed.abo3958
89. Center for Devices and Radiological Health. List of cleared or approved companion diagnostic devices. U.S. Food and Drug Administration. Accessed March 3, 2024. https://www.Fda.Gov/Medical-Devices/in-Vitro-Diagnostics/List-Cleared-or-Approved-Companion-Diagnostic-Devices-in-Vitro-and-Imaging-Tools
90. Lu S, Stein JE, Rimm DL, et al. Comparison of biomarker modalities for predicting response to PD-1/PD-L1 checkpoint blockade: a systematic review and meta-analysis. *JAMA Oncol*. 2019;5:1195-1204. doi:10.1001/jamaoncol.2019.1549
91. Eggermont AMM, Kicinski M, Blank CU, et al. Five-year analysis of adjuvant pembrolizumab or placebo in stage III melanoma. *NEJM Evid*. 2022;1:EVI-Doa2200214. doi:10.1056/EVIDo2200214
92. Larkin J, Del Vecchio M, Mandalá M, et al. Adjuvant nivolumab versus ipilimumab in resected stage III/IV melanoma: 5-year efficacy and biomarker results from CheckMate 238. *Clin Cancer Res*. 2023;29:3352-3361. doi:10.1158/1078-0432.CCR-22-3145
93. Liu SV, Reck M, Mansfield AS, et al. Updated overall survival and PD-L1 subgroup analysis of patients with extensive-stage small-cell lung cancer treated with atezolizumab, carboplatin, and etoposide (IMpower133). *J Clin Oncol*. 2021;39:619-630. doi:10.1200/JCO.20.01055
94. Chan TA, Yarchoan M, Jaffee E, et al. Development of tumor mutation burden as an immunotherapy biomarker: utility for the oncology clinic. *Ann Oncol*. 2019;30:44-56. doi:10.1093/annonc/mdy495
95. Zehir A, Benayed R, Shah RH, et al. Mutational landscape of metastatic cancer revealed from prospective clinical sequencing of 10,000 patients. *Nat Med*. 2017;23:703-713. doi:10.1038/nm.4333
96. Makrooni MA, O'Sullivan B, Seoighe C. Bias and inconsistency in the estimation of tumour mutation burden. *BMC Cancer*. 2022;22:840. doi:10.1186/s12885-022-09897-3
97. Maio M, Ascierto PA, Manzyuk L, et al. Pembrolizumab in microsatellite instability high or mismatch repair deficient cancers: updated analysis from the phase II KEYNOTE-158 study. *Ann Oncol*. 2022;33:929-938. doi:10.1016/j.annonc.2022.05.519
98. Cristescu R, Aurora-Garg D, Albright A, et al. Tumor mutational burden predicts the efficacy of pembrolizumab monotherapy: a pan-tumor retrospective analysis of participants with advanced solid tumors. *J Immunother Cancer*. 2022;10:e003091. doi:10.1136/jitc.2021-003091
99. Mo SF, Cai ZZ, Kuai WH, Li X, Chen YT. Universal cutoff for tumor mutational burden in predicting the efficacy of anti-PD-(L)1 therapy for advanced cancers. *Front Cell Dev Biol*. 2023;11:1209243. doi:10.3389/fcell.2023.1209243
100. McGrail DJ, Pilié PG, Rashid NU, et al. High tumor mutation burden fails to predict immune checkpoint blockade response across all cancer types. *Ann Oncol*. 2021;32:661-672. doi:10.1016/j.annonc.2021.02.006
101. Xu F, Guan Y, Zhang P, et al. Tumor mutational burden presents limiting effects on predicting the efficacy of immune checkpoint inhibitors and prognostic assessment in adrenocortical carcinoma. *BMC Endocr Disord*. 2022;22:130. doi:10.1186/s12902-022-01017-3
102. Niknafs N, Balan A, Cherry C, et al. Persistent mutation burden drives sustained anti-tumor immune responses. *Nat Med*. 2023;29:440-449. doi:10.1038/s41591-022-02163-w
103. Garrido-Ramos MA. Satellite DNA: an evolving topic. *Genes*. 2017;8:230. doi:10.3390/genes8090230
104. Li K, Luo H, Huang L, Luo H, Zhu X. Microsatellite instability: a review of what the oncologist should know. *Cancer Cell Int*. 2020;20:16. doi:10.1186/s12935-019-1091-8
105. Lower SS, McGurk MP, Clark AG, Barbash DA. Satellite DNA evolution: old ideas, new approaches. *Curr Opin Genet Dev*. 2018;49:70-78. doi:10.1016/j.gde.2018.03.003
106. Cortes-Ciriano I, Lee S, Park W-Y, Kim T-M, Park PJ. A molecular portrait of microsatellite instability across multiple cancers. *Nat Commun*. 2017;8:15180. doi:10.1038/ncomms15180
107. Ratovomanana T, Cohen R, Svrcek M, et al. Performance of next-generation sequencing for the detection of microsatellite instability in colorectal cancer with deficient DNA mismatch repair. *Gastroenterology*. 2021;161:814-826. doi:10.1053/j.gastro.2021.05.007
108. Zhang W, Yin H, Huang Z, et al. Development and validation of MRI-based deep learning models for prediction of microsatellite instability in rectal cancer. *Cancer Med*. 2021;10:4164-4173. doi:10.1002/cam4.3957
109. Cercek A, Lumish M, Sinopoli J, et al. PD-1 blockade in mismatch repair-deficient, locally advanced rectal cancer. *N Engl J Med*. 2022;386:2363-2376. doi:10.1056/NEJMoa2201445
110. Cercek A, Bachet J-B, Capdevila J, et al. Phase II, single-arm, open-label study of dostarlimab monotherapy in previously untreated patients with stage II/III dMMR/MSI-H locally advanced rectal cancer. *J Clin Oncol*. 2023;41:TPS3639. doi:10.1200/JCO.2023.41.16_suppl.TPS3639
111. Califf RM. Biomarker definitions and their applications. *Exp Biol Med (Maywood)*. 2018;243:213-221. doi:10.1177/1535370217750088
112. FDA-NIH Biomarker Working Group. *BEST (Biomarkers, Endpoints, and Other Tools)*. Resource Food and Drug Administration (US); 2016.
113. Davenport T, Kalakota R. The potential for artificial intelligence in healthcare. *Future Healthc J*. 2019;6:94-98. doi:10.7861/futurehosp.6-2-94
114. Yap J, Yolland W, Tschandl P. Multimodal skin lesion classification using deep learning. *Exp Dermatol*. 2018;27:1261-1267. doi:10.1111/exd.13777

115. Klauschen F, Müller KR, Binder A, et al. Scoring of tumor-infiltrating lymphocytes: from visual estimation to machine learning. *Semin Cancer Biol.* 2018;52:151-157. doi:10.1016/j.semcancer.2018.07.001
116. Coudray N, Ocampo PS, Sakellaropoulos T, et al. Classification and mutation prediction from non-small cell lung cancer histopathology images using deep learning. *Nat Med.* 2018;24:1559-1567. doi:10.1038/s41591-018-0177-5
117. Huang Z, Johnson TS, Han Z, et al. Deep learning-based cancer survival prognosis from RNA-Seq data: approaches and evaluations. *BMC Med Genomics.* 2020;13:41. doi:10.1186/s12920-020-0686-1
118. Dwivedi K, Rajpal A, Rajpal S, Agarwal M, Kumar V, Kumar N. An explainable AI-driven biomarker discovery framework for non-small cell lung cancer classification. *Comput Biol Med.* 2023;153:106544. doi:10.1016/j.combiomed.2023.106544
119. Esteve A, Feng J, Huang S-C, et al. Development and validation of a prognostic AI biomarker using multi-modal deep learning with digital histopathology in localized prostate cancer on NRG oncology phase III clinical trials. *J Clin Oncol.* 2022;40:222. doi:10.1200/JCO.2022.40.6_suppl.222
120. Sandler HM, Tran PT, Rodgers JP, et al. Artificial intelligence predictive model for hormone therapy use in prostate cancer. *Res Sq.* Published online April 21, 2023. doi:10.21203/rs.3.rs-2790858/v1
121. Bahado-Singh R, Vlachos KT, Aydas B, Gordevicis J, Radhakrishna U, Vishweswaraiah S. Precision oncology: artificial intelligence and DNA methylation analysis of circulating cell-free DNA for lung cancer detection. *Front Oncol.* 2022;12:790645. doi:10.3389/fonc.2022.790645
122. Radak M, Lafta HY, Fallahi H. Machine learning and deep learning techniques for breast cancer diagnosis and classification: a comprehensive review of medical imaging studies. *J Cancer Res Clin Oncol.* 2023;149:10473-10491. doi:10.1007/s00432-023-04956-z
123. Tang Z, Liang D, Deubler EL, et al. Abstract 3025: a pooled analysis of lung cancer metabolomics in the cancer prevention studies revealing sphingomyelin (D18:0/22:0) as a potential screening biomarker. *Cancer Res.* 2023;83:3025. doi:10.1158/1538-7445.AM2023-3025
124. Ge S, Xu CR, Li YM, et al. Identification of the diagnostic biomarker VIPR1 in hepatocellular carcinoma based on machine learning algorithm. *J Oncol.* 2022;2022:2469592. doi:10.1155/2022/2469592
125. Wei L, Niraula D, Gates EDH, et al. Artificial intelligence (AI) and machine learning (ML) in precision oncology: a review on enhancing discoverability through multiomics integration. *Br J Radiol.* 2023;96:20230211. doi:10.1259/bjr.20230211
126. Basu A, Bodycombe NE, Cheah JH, et al. An interactive resource to identify cancer genetic and lineage dependencies targeted by small molecules. *Cell.* 2013;154:1151-1161. doi:10.1016/j.cell.2013.08.003
127. Berns K, Bernards R. Understanding resistance to targeted cancer drugs through loss of function genetic screens. *Drug Resist Updat.* 2012;15:268-275. doi:10.1016/j.drug.2012.10.002
128. Koboldt DC, Zhang Q, Larson DE, et al. VarScan 2: somatic mutation and copy number alteration discovery in cancer by exome sequencing. *Genome Res.* 2012;22:568-576. doi:10.1101/gr.129684.111
129. He X, Liu X, Zuo F, Shi H, Jing J. Artificial intelligence-based multi-omics analysis fuels cancer precision medicine. *Semin Cancer Biol.* 2023;88:187-200. doi:10.1016/j.semcancer.2022.12.009
130. Kann BH, Hosny A, Aerts HJWL. Artificial intelligence for clinical oncology. *Cancer Cell.* 2021;39:916-927. doi:10.1016/j.ccell.2021.04.002
131. Bera K, Schalper KA, Rimm DL, Velcheti V, Madabhushi A. Artificial intelligence in digital pathology—new tools for diagnosis and precision oncology. *Nat Rev Clin Oncol.* 2019;16:703-715. doi:10.1038/s41571-019-0252-y
132. Howlader N, Cronin KA, Kurian AW, Andridge R. Differences in breast cancer survival by molecular subtypes in the United States. *Cancer Epidemiol Biomarkers Prev.* 2018;27:619-626. doi:10.1158/1055-9965.EPI-17-0627
133. Wilske B, Schierz G, Preac-Mursic V, Weber K, Pfister HW, Einhäupl K. Serological diagnosis of erythema migrans disease and related disorders. *Infection.* 1984;12:331-337. doi:10.1007/BF01651147
134. OncoKBTM—Msk's precision oncology knowledge base. Accessed December 26, 2024. <https://www.oncokb.org>
135. Kourie HR, Ibnshamsah F, Zoucin J, et al. The first Middle East and North Africa expert consensus recommendations for the management of advanced colorectal cancer. *Future Oncol.* 2022;18:2733-2744. doi:10.2217/fon-2022-0282
136. Mukherjee B. *Oncology Market Trends in GCC Countries.* IQYIA; 2021.
137. Alharbi A, Bin Dokhi H, Almuhaimeen G, Alomran F, Masuadi E, Alomran N. Prevalence of colorectal cancer biomarkers and their impact on clinical outcomes in Riyadh, Saudi Arabia. *PLoS ONE.* 2021;16:e0249590. doi:10.1371/journal.pone.0249590
138. Benmokhtar S, Laraqui A, Hilali F, et al. RAS/RAF/MEK/ERK/MapK pathway mutations as predictive biomarkers in Middle Eastern colorectal cancer: a systematic review. *Clin Med Insights Oncol.* 2024;18:11795549241255651. doi:10.1177/11795549241255651
139. Saharti S. KRAS/NRAS/BRAF mutation rate in Saudi academic hospital patients with colorectal cancer. *Cureus.* 2022;14:e24392. doi:10.7759/cureus.24392
140. Naser WM, Shawarby MA, Al-Tamimi DM, et al. Novel KRAS gene mutations in sporadic colorectal cancer. *PLoS ONE.* 2014;9:e113350. doi:10.1371/journal.pone.0113350
141. Rasool M, Natesan Pushparaj P, Buhmeida A, Karim S. Mutational spectrum of BRAF gene in colorectal cancer patients in Saudi Arabia. *Saudi J Biol Sci.* 2021;28:5906-5912. doi:10.1016/j.sjbs.2021.06.048
142. Chan AO, Soliman AS, Zhang Q, et al. Differing DNA methylation patterns and gene mutation frequencies in colorectal carcinomas from Middle Eastern countries. *Clin Cancer Res.* 2005;11:8281-8287. doi:10.1158/1078-0432.CCR-05-1000
143. Jafari M, Laraqui A, Baba W, et al. Prevalence and patterns of mutations in RAS/RAF/MEK/ERK/MapK signaling pathway in colorectal cancer in North Africa. *BMC Cancer.* 2022;22:1142. doi:10.1186/s12885-022-10235-w
144. Hughes LAE, Simons CCJM, van den Brandt PA, van Engeland M, Weijenberg MP. Lifestyle, diet, and colorectal cancer risk according to (epi)genetic instability: current evidence and future directions of molecular pathological epidemiology. *Curr Colorectal Cancer Rep.* 2017;13:455-469. doi:10.1007/s11888-017-0395-0
145. Al-Kafaji G, Jassim G, AlHajeri A, et al. Investigation of germline variants in Bahraini women with breast cancer using next-generation sequencing based-multigene panel. *PLoS ONE.* 2023;18:e0291015. doi:10.1371/journal.pone.0291015
146. Blair AB, Groot VP, Gemenetzis G, et al. BRCA1/BRCA2 germline mutation carriers and sporadic pancreatic ductal adenocarcinoma. *J Am Coll Surg.* 2018;226:630-637. doi:10.1016/j.jamcollsurg.2017.12.021
147. Mirza MR, Monk BJ, Herrstedt J, et al; and ENGOT-OV16/NOVA Investigators. Niraparib maintenance therapy in platinum-sensitive, recurrent ovarian cancer. *N Engl J Med.* 2016;375:2154-2164. doi:10.1056/NEJMoa1611310
148. Coleman RL, Oza AM, Lorusso D, et al; and ARIEL3 Investigators. Rucaparib maintenance treatment for recurrent ovarian carcinoma after response to platinum therapy (ARIEL3): a randomised, double-blind, placebo-controlled, phase 3 trial. *Lancet.* 2017;390:1949-1961. doi:10.1016/S0140-6736(17)32440-6
149. Bu R, Siraj AK, Al-Obaisi KAS, et al. Identification of novel BRCA founder mutations in Middle Eastern breast cancer patients using capture and Sanger sequencing analysis. *Int J Cancer.* 2016;139:1091-1097. doi:10.1002/ijc.30143
150. Farra C, Dagher C, Hamadeh L, El Saghir N, Mukherji D. BRCA mutations in a cohort of Iraqi patients presenting to a Tertiary Referral Center. *BMC Med Genet.* 2019;20:154. doi:10.1186/s12881-019-0885-9
151. Agha N, Alshamsan B, Al-Farsi S, et al. Assessing frequency and clinical outcomes of BRCA mutated ovarian cancer in Saudi women. *BMC Cancer.* 2022;22:18. doi:10.1186/s12885-021-09123-6
152. Moore DC, Guinigundo AS. Revolutionizing cancer treatment: harnessing the power of biomarkers to improve patient outcomes. *J Adv Pract Oncol.* 2023;14:4-8. doi:10.6004/jadpro.2023.14.3.15
153. Purkayastha K, Dhar R, Pethusamy K, et al. The issues and challenges with cancer biomarkers. *J Cancer Res Ther.* 2023;19:S20-S35. doi:10.4103/jcrt.jcrt_384_22
154. Rassy E, Heard JM, Andre F. The paradigm shift to precision oncology between political will and cultural acceptance. *ESMO Open.* 2023;8:101622. doi:10.1016/j.esmoop.2023.101622
155. Taube SE, Abrams JS. Program for the assessment of clinical cancer tests (PACCT): assisting the development of tailored cancer therapy. *Per Med.* 2005;2:363-369. doi:10.2217/17410541.2.4.363
156. Williams PM, Lively TG, Jessup JM, Conley BA. Bridging the gap: moving predictive and prognostic assays from research to clinical use. *Clin Cancer Res.* 2012;18:1531-1539. doi:10.1158/1078-0432.CCR-11-2203
157. Stein MK, Oluoha O, Patel K, VanderWalde A. Precision medicine in oncology: a review of multi-tumor actionable molecular targets with an emphasis on non-small cell lung cancer. *J Pers Med.* 2021;11:518. doi:10.3390/jpm11060518
158. Schwartzberg L, Kim ES, Liu D, Schrag D. Precision oncology: who, how, what, when, and when not? *Am Soc Clin Oncol Educ Book.* 2017;37:160-169. doi:10.1200/EDBK_174176
159. Drucker E, Krapfenbauer K. Pitfalls and limitations in translation from biomarker discovery to clinical utility in predictive and personalised medicine. *EPMA J.* 2013;4:7. doi:10.1186/1878-5085-4-7
160. Lawrence MS, Stojanov P, Mermel CH, et al. Discovery and saturation analysis of cancer genes across 21 tumour types. *Nature.* 2014;505:495-501. doi:10.1038/nature12912
161. Amorós-Pérez B, Rivas-Pardo B, Gómez Del Moral M, Subiza JL, Martínez-Naves E. State of the art in CAR-T cell therapy for solid tumors: is there a sweeter future? *Cells.* 2024;13:725. doi:10.3390/cells13090725
162. Mer AH, Mirzaei Y, Misamogoo F, et al. Progress of antibody-drug conjugates (ADCs) targeting c-Met in cancer therapy; insights from clinical and pre-clinical studies. *Drug Deliv Transl Res.* 2024;14:2963-2988. doi:10.1007/s13346-024-01564-3
163. Dosio F, Brusca P, Cattel L. Immunotoxins and anticancer drug conjugate assemblies: the role of the linkage between components. *Toxins (Basel).* 2011;3:848-883. doi:10.3390/toxins3070848
164. Wang Z, Ma J, Zhang H, et al. CCR4-IL2 bispecific immunotoxin is more effective than brentuximab for targeted therapy of cutaneous T-cell lymphoma in a mouse CTCL model. *FEBS Open Bio.* 2023;13:1309-1319. doi:10.1002/2211-5463.13625