Neuropsychiatric Disease and Treatment

Open Access Full Text Article

ORIGINAL RESEARCH

Resveratrol ameliorates chronic unpredictable mild stress-induced depression-like behavior: involvement of the HPA axis, inflammatory markers, BDNF, and Wnt/ β -catenin pathway in rats

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Abstract: Classic antidepressant drugs are modestly effective across the population and most are associated with intolerable side effects. Recently, numerous lines of evidence suggest that resveratrol (RES), a natural polyphenol, possesses beneficial therapeutic activity for depression. The aim of the present study was to explore whether RES exhibits an antidepressant-like effect in a depression model and to explore the possible mechanism. A depression model was established via chronic unpredictable mild stress (CUMS), after which the model rats in the RES and fluoxetine groups received a daily injection of RES or fluoxetine, respectively. The sucrose preference test, open field test, and forced swimming test were used to explore the antidepressantlike effects of RES. The activity of the hypothalamic-pituitary-adrenal (HPA) axis was evaluated by detecting the plasma corticosterone concentration and hypothalamic mRNA expression of corticotrophin-releasing hormone. The plasma interleukin-6 (IL-6), C-reactive protein (CRP), and tumor necrosis factor- α (TNF- α) concentrations were measured by enzyme-linked immunosorbent assay. Hippocampal protein expression of brain-derived neurotrophic factor (BDNF) and the Wnt/ β -catenin pathway were analyzed by western blot. The results showed that RES relieved depression-like behavior of CUMS rats, as indicated by the increased sucrose preference and the decreased immobile time. Rats that received RES treatment exhibited reduced plasma corticosterone levels and corticotrophin-releasing hormone mRNA expression in the hypothalamus, suggesting that the hyperactivity of the HPA axis in CUMS rats was reversed by RES. Moreover, after RES treatment, the rats exhibited increased plasma IL-6, CRP, and TNF- α concentrations. Furthermore, RES treatment upregulated the hippocampal protein levels of BDNF and the relative ratio of p- β -catenin/ β -catenin while downregulating the relative ratio of p-GSK-3β/GSK-3β. Our findings suggest that RES improved depressive behavior in CUMS rats by downregulating HPA axis hyperactivity, increasing BDNF expression and plasma IL-6, CRP, and TNF- α concentrations, and regulating the hippocampal Wnt/ β -catenin pathway. Keywords: brain-derived neurotrophic factor, chronic unpredictable mild stress, depression, hypothalamus-pituitary-adrenal axis, resveratrol, Wnt/ β -catenin pathway, interleukin-6,

Introduction

Depression is one of the leading causes of disability, affecting up to 120 million people worldwide.¹ According to the World Health Organization Global Burden of Disease, depression is anticipated to become the leading cause of long-term disability by the year 2030.⁵⁷ Based on the monoamine neurotransmitter hypothesis of depression,²

Neuropsychiatric Disease and Treatment 2017:13 2727-2736

C-reactive protein, tumor necrosis factor-a

2727

© 2017 Yang et al. This work is published and licensed by Dove Medical Press Limited. The full terms of this license are available at https://www.dovepress.com/terms.php hereby accept the Terms. Non-commercial uses of the work are permitted without any further permission from Dove Medical Press Limited, provided the work is properly attributed. For permission for commercial use of this work, large accept and the commercial uses of the work are permitted without any further permission from Dove Medical Press Limited, provided the work is properly attributed. For permission for commercial use of this work, large see paragraphs 4.2 and 5 of our Terms (http://www.dovepress.com/terms.php). raising monoamine levels in synaptic clefts has been the main strategy for treating depressive patients over the past 60 years. However, not all patients with depressive disorders respond to classic antidepressants, and months of treatment are usually required, indicating that monoamine depletion might not be the unique pathogenesis of depression.³ In addition to this, there is also growing evidence regarding adverse reactions to the clinical use of antidepressants, including dizziness, tremor, cognitive impairments, urinary retention, and sexual dysfunction.⁴ Therefore, there is an urgent need to explore the pathogenesis of depression and improve the efficacy and safety of depression treatment with better drugs.

The stress response and the resulting hypothalamic– pituitary–adrenal (HPA) axial hyperactivity are considered to be one of the most powerful triggers for depressive disorders.⁵ Preclinical and clinical studies have indicated that the stress response can result in depression-like behavior accompanied by hyperactivity of the HPA axis,^{6,7} as indicated by increased serum glucocorticoid concentrations and corticotrophinreleasing hormone (CRH) expression in the hypothalamus.⁸ Chronic mild unpredictable stress (CUMS) has been used in animal depression research because it can simulate the representative symptoms of depression, including anhedonia and despairing behavior.⁹ Given the roles of the HPA axis in the pathophysiology of depression, plasma corticosterone concentrations and hypothalamic CRH expression were evaluated in CUMS rats in this study.

Increasing evidence supports the view that depression is accompanied by activation of the inflammatory response system, and overproduction of pro-inflammatory cytokines may play a role in the pathophysiology of depressive disorders.^{10,11} It has been reported that the most robust evidence-based inflammatory markers associated with depressive disorder include interleukin-6 (IL-6), C-reactive protein (CRP), and tumor necrosis factor- α (TNF- α).¹² Moreover, numerous studies have demonstrated that increased levels of plasma IL-6 and CRP are related to an increased risk of depressive episodes^{13,14} and can even be used to predict subsequent depressive symptoms.15 In animals, it has been shown that depression-like behaviors are induced by cytokines¹⁶ and cytokine inducers, including lipopolysaccharide administration¹⁷ and chronic mild stress.¹⁸ Therefore, IL-6, CRP, and TNF- α levels were evaluated in CUMS rats with and without antidepressant administration in this study.

Recent evidence has indicated that decreased levels of brain-derived neurotrophic factor (BDNF) in the hippocampus can cause reduced proliferation of hippocampal neurons and result in depressive disorders.¹⁹ Moreover, depression-like effects in rats induced by chronic stress or glucocorticoid administration have been reported to be related to reduced hippocampal BDNF levels.^{20,21} Furthermore, some antidepressants are reported to achieve their efficacy by increasing hippocampal BDNF expression.^{22,23} Although the underlying mechanisms remain unknown, it has been reported that BDNF appears to be a direct target of Wnt signaling.²⁴ Together with the fact that the canonical Wnt signaling pathway has been reported to be implicated in mood disorders such as bipolar disorder and major depression,^{25,26} the protein expression levels of BDNF and the Wnt/ β -catenin pathway were further investigated in CUMS rats in the present study.

Resveratrol (trans-3,5,4'-trihydroxy-trans-stilbene, RES), a phytoalexin with antioxidant properties, is found in a wide range of foods, especially in grapes and red wine. In the last decade, RES has been shown to possess multiple biological and pharmacological activities such as antiinflammatory, antioxidant, anticarcinogenic, antiaging, neuroprotective, and cardioprotective effects.²⁷ Recently, RES was reported to exert antidepressant-like effects, alleviating depression-like symptoms/behaviors in rodent animal models.^{28,29} Additionally, it has been shown that RES treatment can alter the monoaminergic system and the molecular markers related to depression.^{30,31} However, it remains unknown whether RES can improve depression-like symptoms in CUMS rats, which involve a complicated balance of the HPA axis, the inflammatory reaction, BDNF, and the Wnt/ β -catenin pathway.

In the present study, a rat depression model was established using CUMS. Behavior tests, including the open field test (OFT), sucrose preference test (SPT), and forced-swimming test (FST), were used to probe the effects of RES.

Experimental procedures Drugs

RES was provided by Sigma Chemical Co. (St Louis, MO, USA). Fluoxetine hydrochloride (Prozac) was provided by Eli Lilly Pharmaceuticals. RES and fluoxetine were dissolved to become a mixed suspension in an aqueous solution of 0.5% sodium carboxymethyl cellulose.

Animals

After 7 days of adaptive breeding, 40 male adult Sprague Dawley (SD) rats were randomly allocated to four groups, including a control group (control +0.5% sodium carboxymethyl cellulose), a chronic unpredictable mild stress group (CUMS +0.5% sodium carboxymethyl cellulose), RES treatment group (15 mg/kg/day RES + CUMS), and a fluoxetine-treated CUMS group (2 mg/kg/day fluoxetine hydrochloride + CUMS). Rats were maintained at 21°C–25°C with 50%–60% relative humidity, under a 12/12 h light–dark cycle (lights on at 07:30 h). Rats in the control group were housed five per cage (43 cm length ×31 cm width ×19 cm height) with free access to water and food. Rats in the other three groups were housed five per cage and stressed according to the CUMS procedure described below. The experiments were performed after approval of the protocol by the Animal Care and Use Committee at Anhui Medical University and complied with the National Institutes of Health Guide for the Care and Use of Laboratory Animals (NIH publication No 85-23, revised 1985).

Chronic unpredictable mild stress procedure

Stressors were administered once daily between 08:40 and 10:40 for 3 weeks, with the exception of the 24 h duration stressors. The stressors were used to induce a depressive state, including 1) social crowding (10 rats per cage, 24 h); 2) swimming at 30°C (20 min); 3) in a cage tilted at 30° from the horizontal (24 h); 4) swimming at 8°C–10°C (5 min); 5) in a wet cage (24 h); 6) tail pinch (2 min); and 7) food and water deprivation (24 h). The different stressors were

randomly distributed, with an interval of 7 days between repetitions. All stressors were administered three times within 21 days. The schedule of the experimental design is shown in Figure 1.

Behavioral tests

Behavioral tests were executed in a noise-proof room. All behavioral tests were performed between 08:00 and 14:00, with matching between the groups.

Sucrose preference test

All the rats were singly housed in a cage (28 cm length \times 17 cm width \times 14.5 cm height) after a 12-h period of food and water deprivation. Then, all rats were given two bottles containing water or 2% sucrose solution. Sucrose and water bottles were placed in randomly assigned sides of the cage. Six hours later, the weights of water and sucrose consumed were measured. The rats' sensitivity to reward was calculated as the percentage of total liquid intake attributed to the 2% sucrose solution, according to the following equation: Sucrose solution(g)/(Sucrose solution [g] + water [g]) \times 100%.



Figure I Schedule of the experimental design.

Abbreviations: BDNF, brain-derived neurotrophic factor; CRH, corticotrophin-releasing hormone; CRP, C-reactive protein; CUMS, chronic unpredictable mild stress; D, day; ELISA, enzyme-linked immunosorbent assay; FST, forced swimming test; IL-6, interleukin-6; OFT, open field test; PCR, polymerase chain reaction; RES, resveratrol; SPT, sucrose preference test; TNF-α, tumor necrosis factor-α.

Open field test

The open field arena, 100×100 cm in size, was divided into 16 square areas. Rats were placed randomly at one corner of the apparatus. During a 5-min observation period, all rat movements were recorded by a camera interfaced to a computer running the ANY maze video imaging software (Stoelting Co, Wood Dale, IL, USA). After testing each individual rat, the open field arena surface was cleaned with 75% ethyl alcohol. The total distance was recorded to measure locomotion. The distance, frequency, and duration in the center, and frequencies of grooming and defecation were recorded to measure anxiety. The frequency of rearing was recorded to measure exploratory behavior.

Forced swimming test

The FST was conducted by placing the rat in a cylinder (60 cm in height $\times 25$ cm in diameter) containing 30 cm of water at 24°C–25°C. The FST paradigm includes two sections: an initial 15-min pretest followed by a 5-min test 24 h later. A rat was considered to be immobile when it remained floating in the water in an upright position, making only very small movements to keep its head above the water. A rat was considered to be struggling when it made active movements with its forepaws in and out of the water along the side of the swim chamber. A rat was considered to be swimming or circular movements.

Measurement of plasma corticosterone, IL-6, CRP, and TNF- α

Twenty-four hours after the FST, the rats were anesthetized by an intraperitoneal (i.p.) injection of chloral hydrate (300 mg/kg, Sinopharm Chemical Reagent Co., Ltd, Shanghai Shi, China). Then, blood was collected from the abdominal aorta in evacuated tubes with EDTA-K2. The plasma concentrations of corticosterone, IL-6, CRP, and TNF- α were measured using an enzyme-linked immunosorbent assay (ELISA; corticosterone: Enzo Life Sciences, Inc., Farmingdale, NY, USA; IL-6, CRP, and TNF- α : Yuanye Biotech. Co., LTD, Shanghai, China) according to the manufacturer's instructions.

RNA isolation and real-time PCR

After blood collection, rats were sacrificed by decapitation. Then, the hypothalamus was rapidly dissected on ice, frozen quickly in liquid nitrogen, and stored at -80°C. Total RNA was extracted using Trizol reagent (Invitrogen, Carlsbad, CA, USA) according to the manufacturer's instructions. Identical amounts of RNA (2 µg) were reverse transcribed into cDNA using a commercial real-time polymerase chain reaction (RT-PCR) kit (Promega, Fitchburg, WI, USA) according to the manufacturer's instructions. qPCR was performed using the SYBR Green PCR Kit (Applied Biosystems, Waltham, MA, USA) and an ABI Prism 7000 Sequence Detector system in a 25 µL volume for 40 cycles (15 s at 95°C; 60 s at 62°C) for rat β-actin and CRH. The primers used were β-actin Forward (5'-TTGCTGACAGGATGCAGAA-3'), Reverse (5'-ACCAATCCACACAGAGTACTT-3'); CRH Forward (5'-CAGAACAACAGTGCGGGCTCA-3'), Reverse (5'-AAGGCAGACAGGGCGACAGAG-3'). The relative amount of the target gene was calculated using the $2^{-\Delta\Delta Ct}$ method.

Western blot assays

Frozen tissue from the entire hippocampus was homogenized in ice-cold radioimmunoprecipitation assay (RIPA) buffer plus Complete Protease Inhibitor (Roche, Indianapolis, IN, USA). The dissolved proteins were collected after centrifugation at 10,000 g for 15 min at 4°C, and then, the supernatant was collected. Protein quantitation was conducted using a Lowry Protein Assay Kit (Meiji Biotech. Co., LTD., Shanghai, China). The same quantity (~40-50 µg) of protein from each rat was separated by 12% SDS-PAGE and then transferred onto a polyvinylidene difluoride membrane (Amersham Biosciences, Buckinghamshire, UK). The membrane was blocked with 5% skim milk for 3 h, probed with the primary antibodies targeting β -actin (1:800; Bioworld Technology, Inc., St Louis Park, MN, USA), BDNF (1:500; Santa Cruz Biotechnology, Inc., Dallas, TX, USA), GSK-3β, p-GSK-3β (Ser9), β-catenin, or p-β-catenin (1:1,000; Cell Signaling Technology, Danvers, MA, USA) at 4°C overnight, and then incubated with a horseradish peroxidase-conjugated secondary antibody (1:10,000) at 37°C for 2 h. The membranes were developed using an enhanced chemiluminescence detection system (Pierce Biotechnology, Rockford, IL, USA).

Statistical analysis

All tests were performed using the SPSS Graduate Pack 17.0 statistical software package (SPSS, Chicago, IL, USA). Data were expressed as the mean \pm SEM. The between-group effects on body weight were analyzed by a repeated measures one-way analysis of variance (ANOVA) with treatment group and weeks as the factors. Statistical analyses of behavioral performance, plasma corticosterone, IL-6, CRP, and TNF- α concentration, and expression of CRH, BDNF, p-GSK-3 β /GSK-3 β , or p- β -catenin/ β -catenin were carried out by

ANOVA followed by least significant difference (LSD) post hoc tests. A value of P < 0.05 was regarded as significant.

Results

RES administration improved depressionlike behavior caused by CUMS in rats

Figure 2 shows the effect of RES on the changes in body weight and behavior caused by CUMS. The repeated measures

ANOVA (four treatments ×3 weeks with repeated measures on weeks) showed that both the treatment and the time affected body weight (treatment effect: (F[3,36] =4.162, P=0.047); time effect: (F[3,108] =634.376, P<0.001); interaction: (F[9,108] =3.880, P=0.004)). The LSD test showed that the CUMS rats, with or without RES treatment, gained less body weight during the 3-week stress period compared with controls (F[3,36] =13.177, P=0.002).



Figure 2 Effect of RES on the body weight and behavior of the CUMS rats.

Notes: The body weight (**A**), sucrose preference (**B**), and performance in the OFT (**C**, **D**) and FST (**E**) were observed. The data are presented as the mean \pm SEM, with n=10 for each group. **P*<0.05 and ***P*<0.01 compared to the control group. **P*<0.05 and ***P*<0.01 compared to the model group. **Abbreviations:** CUMS, chronic unpredictable mild stress; FST, forced swimming test; OFT, open field test; RES, resveratrol.



Figure 3 Effects of RES on the plasma corticosterone and the expression of CRH mRNA in the hypothalamus in CUMS rats. Notes: The plasma corticosterone (**A**), and the expression of CRH mRNA in the hypothalamus (**B**) are shown. The data are presented as the mean \pm SEM, with n=10 for each group. **P<0.01 compared to the control group. #P<0.05 and ##P<0.01 compared to the model group. **Abbreviations:** CRH, corticotrophin-releasing hormone; CUMS, chronic unpredictable mild stress; RES, resveratrol.

However, no significant differences between CUMS rats and CUMS rats treated with RES or fluoxetine were found (Figure 2A).

Compared to the control group, the sucrose preference of CUMS rats was significantly lower, which could be reversed by RES or fluoxetine treatment (F[3,36] = 4.289, P=0.044; Figure 2B).

In the OFT, no differences in performance, including the number of rearings (F[3,36] =1.312, P=0.336; Figure 2C), groomings (F[3,36] =0.222, P=0.878; Figure 2C), defecations (F[3,36] =2.778, P=0.110; Figure 2C), squares crossed (F[3,36] =0.462, P=0.717; Figure 2C), and distance (F[3,36] =1.178, P=0.377; Figure 2D), were observed among the four groups.

In the FST, no differences in the climbing (F[3,36]=0.820, P=0.519; Figure 2E) or swimming (F[3,36]=1.487, P=0.290; Figure 2E) time were observed among the four groups. However, the CUMS rats were immobile for a longer time (F[3,36]=7.272, P=0.011) than controls, which was reversed by RES and fluoxetine treatment (Figure 2E).

RES administration decreased hyperactivity of the HPA axis caused by CUMS

After three continuous weeks of CUMS, the plasma corticosterone concentrations of the CUMS rats were remarkably higher than that of the control rats (F[3,36]=9.747, P=0.002; Figure 3A). Treatment with RES or fluoxetine decreased the elevated corticosterone levels (Figure 3A). Consistently, hypothalamic CRH mRNA expression was inhibited by treatment with RES or fluoxetine (F[3,36]=6.223, P=0.017; Figure 3B).

RES administration decreased the elevated IL-6, CRP, and TNF- α concentrations induced by CUMS in rats

As shown in Figure 4, compared with the control group, the plasma IL-6 (F[3,36] =7.600, P=0.01; Figure 4A), CRP (F[3,36] =4.0554, P=0.049; Figure 4B), and TNF- α (F[3,36] =14.998, P=0.001; Figure 4C) concentrations of model rats were significantly higher than those of the controls, which was reversed by the RES or fluoxetine treatment.

RES administration increased BDNF protein expression and decreased activation of the canonical Wnt pathway in the hippocampus of CUMS rats

Figure 5 shows the protein expression levels of GSK-3 β , p-GSK-3 β (Ser9), β -catenin, p- β -catenin, and BDNF in the hippocampus. Compared to the control group, the hippocampal protein levels of BDNF were lower in the CUMS rats. However, these differences were ameliorated by RES or fluoxetine (F[3,36] =11.948, *P*=0.003; Figure 5A and B).

A lower relative ratio of p-GSK-3 β /GSK-3 β (F[3,36] =4.524, *P*=0.039; Figure 5C and D) and a higher relative ratio of p- β -catenin/ β -catenin (F[3,36] =14.943, *P*=0.001; Figure 5C and D) were observed in the hippocampus of the model rats compared with the control group, which were reversed by the RES or fluoxetine treatments.

Discussion

In the present study, we investigated the antidepressant effects of RES using the CUMS rat depression model. The results showed that RES can improve the depression-like behavior exhibited by rats subjected to CUMS, as indicated



Figure 4 Effects of RES on the plasma concentrations of IL-6, CRP, and TNF- α in CUMS rats.

Notes: The plasma concentrations of IL-6 (A), CRP (B), and TNF- α (C) are shown. The data are presented as the mean ± SEM, with n=10 for each group. *P<0.05 and **P<0.01 compared to the control group. *P<0.05 and ##P<0.01 compared to the model group.

Abbreviations: CRP, C-reactive protein; CUMS, chronic unpredictable mild stress; IL-6, interleukin-6; RES, resveratrol; TNF- α , tumor necrosis factor- α .



Figure 5 Effects of RES on the protein expressions of BDNF and the Wnt/ β -catenin pathway in the hippocampi of CUMS rats. Notes: Representative blots and the quantitative analysis of BDNF (**A**, **B**), GSK-3 β and p-GSK-3 β (**C**, **D**), β -catenin and p- β -catenin (**C**, **D**) are presented. The data are presented as the mean \pm SEM, with n=10 for each group. **P<0.01 compared to the control group. **P<0.05 and **P<0.01 compared to the model group. **Abbreviations:** BDNF, brain-derived neurotrophic factor; CUMS, chronic unpredictable mild stress; RES, resveratrol. by an increased sucrose preference index in the SPT and a decreased immobile time in the FST. Rats that received RES treatment had lower plasma corticosterone concentrations and hypothalamic CRH mRNA expression levels, indicating that RES treatment reversed the hyperactivity of the HPA axis observed in CUMS rats. RES treatment also increased the protein expression of BDNF and decreased the activation of the canonical Wnt pathway in the hippocampus of CUMS rats.

CUMS is often utilized in laboratory animals to mimic unpredictable life stressors that may contribute to the development of major depressive disorders in humans.³² In the present study, rats displayed typical anhedonia behavior and despairing behavior after 3 weeks of chronic stress, as indicated by their decreased sucrose preference in the SPT and their increased immobile time in the FST, which showed that the rat model of depression was established successfully. The OFT provides simultaneous analysis of locomotion, exploration, and anxiety.33 In the present study, no differences in performance in the OFT, including the amount of rearing, grooming, and defecation, the number of squares crossed and distance were observed among the four groups, indicating that RES and CUMS had no significant effect on the locomotor activity, exploration, and anxiety level of the rats. The SPT is generally used to evaluate anhedonia, defined as impairments in the ability to pursue, experience, and/or learn about pleasure, which is a typical symptom of depression in rodents.³⁴ Immobile time in the FST is taken as an index of despair behavior, which is another prominent symptom of depression.³⁵ In the present study, the CUMS rats showed a reduced sucrose preference index in the SPT and an increased immobile time in the FST, indicating depression-like symptoms in CUMS rats, which was consistent with the previous results.³⁶

It has been shown in clinical studies that RES is well tolerated with the dosages ranging from 200 to 1,000 mg^{37,38} daily. In terms of the formula for dose translation based on body surface area (BSA), the corresponding dose should range from 18 to 90 mg/kg in rats. With respect to preclinical studies, it has been reported that RES (15 mg/kg×16 days) can improve the depression-like behavior in subclinical hypothyroidism rats.³⁹ Consistently, another study also demonstrated that RES (10 and 40 mg/kg) exerts an antidepressant-like effect in an animal model of depression.²⁸ Therefore, a dose of 15 mg/kg/day was selected in the present study.

Hyperactivity of the HPA axis is one of the most powerful factors that trigger depressive disorders.⁵ It has been reported that RES treatment significantly reduces the increased serum

corticosterone level in a stressed animal model induced by repeated corticosterone treatment, which has been performed widely in adult rodents in various depression models.⁴⁰ Pang et al studied the effect of RES isomers on the HPA axis.⁴¹ The result showed that trans-RES may exert its antidepressantlike effect against post-stroke depression by modulating the HPA axis. Consistent with these studies, rats that received RES treatment had decreased plasma corticosterone concentrations and CRH mRNA expression in the hypothalamus in the present study, suggesting that the RES has a regulatory effect on the hyperactivity of the HPA axis in CUMS rats.

Given that inflammation may play a role in depression, antiinflammatory agents may be a useful antidepressant therapy or adjuvant to traditional therapies. Research involving RES treatment in rodent models of inflammatory diseases have demonstrated downregulation of inflammationinduced biomarkers, including proinflammatory mediators (IL-1 β , IL-6, TNF- α , and TNF- β),⁴² and upregulation of inflammation-reduced biomarkers, including antiinflammatory protein (IL-10).⁴³ Consistently, the plasma IL-6, CRP, and TNF- α concentrations of model rats increased significantly, which was reversed by the RES or fluoxetine treatment in the present study. Thus, as one of the initial pharmacological activities, the antiinflammatory activity of RES might be involved in its antidepressant-like effects.

GSK-3ß regulates phosphorylation and subsequent degradation of β -catenin, thereby preventing aberrant activation of the canonical Wnt pathway.44 Clinical studies have assessed the levels or activity of total GSK-3 β protein in the prefrontal cortex in patients with mood disorders, including depression.⁴⁵ Evidence also suggests that inhibition of GSK-3β might contribute to an antidepressant effect.⁴⁶ Consistent with the result that the enzymatic activity of GSK-3 β was increased in depressed suicide victims,45 lower ratios of p-GSK-3 β /GSK-3 β in the hippocampus were found in the CUMS rats, which could underlie depression-like behavior. The decreased ratio of p-GSK-3β/GSK-3β was ameliorated by RES treatment. L803-mts, a known GSK-3β inhibitor,⁴⁷ was reported to induce an antiimmobility effect in the FST. Moreover, a lower GSK-3ß expression level in heterozygotic GSK-3 $\beta^{+/-}$ mice is associated with a reduced immobile time in the FST.⁴⁸ These studies, together with the result of this study, suggest a role for GSK-3 β in the antidepressant effects of RES and highlight GSK-3 β as a potential target in the treatment of human depression.

 β -catenin, a substrate of GSK-3 β , is involved in cerebral development, cognitive function, and dendritic growth.^{49,50} Increasing evidence has indicated that β -catenin levels in

the hippocampus can serve as a marker for antidepressant effects.⁵¹ Moreover, β -catenin protein levels in the postmortem prefrontal cortices of depressed patients were lower compared to healthy controls.⁵² Consistently, the ratio of p- β -catenin/ β -catenin was increased in the hippocampus of the model rats, which was reversed by the RES treatment. The findings from this study, together with the reports mentioned above, reinforce the observation that RES might involve regulation of the canonical Wnt pathway in its antidepressant-like effect.

BDNF, a neurotrophin, is involved in neuronal proliferation, development, differentiation, and maturation.⁵³ There are several reports on the functional interaction between the BDNF and Wnt signaling cascades. Recent studies have demonstrated that BDNF expression is regulated by the Wnt signaling pathway.⁵⁴ Another study reported that BDNF and Wnt signaling cooperatively regulate dendritic spine formation.55 Moreover, BDNF also mobilizes synaptic vesicles, enhances synapse formation, regulates axonal morphogenesis, and promotes neurite growth from neonatal cochlear ganglion explants by disrupting β -catenin interactions.⁵⁶ In the present study, our results showed that RES treatment could upregulate the reduced hippocampal protein levels of BDNF in CUMS rats. Given the relationship between BDNF and the canonical Wnt pathway, it is logical to hypothesize that the upregulation of BDNF expression caused by RES might be due to its effect on the canonical Wnt pathway.

Conclusion

In conclusion, our results demonstrate that RES ameliorated depression-like behavior in CUMS rats. This effect may be due, at least in part, to regulation of the HPA axis, inflammatory markers, BDNF, and the Wnt/ β -catenin pathway in the hippocampus.

Disclosure

The authors report no conflicts of interest in this work.

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