

OPEN ACCESS

EDITED BY
Brij Bhan Singh,
The University of Texas Health Science
Center at San Antonio, United States

Maurizio Molinari, Institute for Research in Biomedicine (IRB), Switzerland Cara B Gonzales, The University of Texas Health Science

Center at San Antonio, United States

*CORRESPONDENCE Richard Zimmermann, richard.zimmermann@uks.eu

SPECIALTY SECTION

This article was submitted to Membrane Physiology and Membrane Biophysics, a section of the journal Frontiers in Physiology

RECEIVED 08 August 2022 ACCEPTED 16 September 2022 PUBLISHED 03 October 2022

CITATION

Zimmermann JSM, Linxweiler J, Radosa JC, Linxweiler M and Zimmermann R (2022), The endoplasmic reticulum membrane protein Sec62 as potential therapeutic target in SEC62 overexpressing tumors. *Front. Physiol.* 13:1014271. doi: 10.3389/fphys.2022.1014271

COPYRIGHT © 2022 Zimmermann, Linxweiler,

This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Radosa, Linxweiler and Zimmermann.

The endoplasmic reticulum membrane protein Sec62 as potential therapeutic target in *SEC62* overexpressing tumors

Julia S. M. Zimmermann¹, Johannes Linxweiler², Julia C. Radosa¹, Maximilian Linxweiler³ and Richard Zimmermann⁴*

¹Department of Gynecology, Obstetrics and Reproductive Medicine, Saarland University, Homburg, Germany, ²Department of Urology and Pediatric Urology, Saarland University, Homburg, Germany, ³Department of Otorhinolaryngology, Head and Neck Surgery, Saarland University, Homburg, Germany, ⁴Competence Center for Molecular Medicine, Saarland University, Homburg, Germany

The human SEC62 gene is located on chromosome 3q, was characterized as a tumor driver gene and is found to be overexpressed in an ever-growing number of tumors, particularly those with 3q26 amplification. Where analyzed, SEC62 overexpression was associated with poor prognosis. Sec62 protein is a membrane protein of the endoplasmic reticulum (ER) and has functions in endoplasmic reticulum protein import, endoplasmic reticulum-phagy and -in cooperation with the cytosolic protein calmodulin- the maintenance of cellular calcium homeostasis. Various human tumors show SEC62 overexpression in immunohistochemistry and corresponding cell lines confirm this phenomenon in western blots and immunofluorescence. Furthermore, these tumor cells are characterized by increased stress tolerance and migratory as well as invasive potential, three hallmarks of cancer cells. Strikingly, plasmid-driven overexpression of SEC62 in non-SEC62 overexpressing cells introduces the same three hallmarks of cancer into the transfected cells. Depletion of Sec62 from either type of SEC62 overexpressing tumor cells by treatment with SEC62-targeting siRNAs leads to reduced stress tolerance and reduced migratory as well as invasive potential. Where tested, treatment of SEC62 overexpressing tumor cells with the small molecule/calmodulin antagonist trifluoperazine (TFP) phenocopied the effect of SEC62-targeting siRNAs. Recently, first phase II clinical trials with the prodrug mipsagargin/G202, which targets cellular calcium homeostasis in prostate cells as well as neovascular tissue in various tumors were started. According to experiments with tumor cell lines, however, SEC62 overexpressing tumor cells may be less responsive or resistant against such treatment. Therefore, murine tumor models for tumor growth or metastasis were evaluated with respect to their responsiveness to treatment with a mipsagargin analog (thapsigargin), or trifluoperazine, which had previously been in clinical use for the treatment of schizophrenia, or with the combination of both drugs. So far, no additive effect of the two drugs was observed but trifluoperazine had an inhibitory effect on tumor growth and metastatic potential in the models. Here, we review the state of affairs.

KEYWORDS

cellular calcium homeostasis, endoplasmic reticulum, endoplasmic reticulum-phagy, hallmarks of cancer, prodrug mipsagargin/G202, protein biogenesis, Sec62 protein, tumor driver gene SEC62

Introduction

The endoplasmic reticulum (ER) of nucleated cells forms a vast membrane network, which extends from the nuclear envelope to the cell periphery (Figure 1, left) (reviewed by Lang et al., 2017; Lang et al., 2019; Molinari, 2020; Sicking et al., 2021; Walter and Ron, 2011). Briefly summarizing these reviews, the mammalian ER has major functions in gene expression, cellular calcium homeostasis and signal transduction (Figure 2). About thirty percent of the cellular proteome is synthesized by ER-bound ribosomes, which amounts to approximately ten thousand different proteins (Figure 1, center and right). Most of these proteins are either integrated into the ER membrane or imported into the ER lumen, as mediated by signal peptides or signal peptide-equivalent transmembrane helices in the precursor polypeptides. In addition, a complex transport machinery comprising about

one hundred proteins in the cytosol, the ER membrane and the ER lumen is involved in these biosynthetic processes (Figure 2, upper left section). In the ER, the proteins are folded and assembled and non-ER proteins, such as plasma membrane or secretory proteins, are transported to the cell surface by vesicular transport. In the case of misfolding or misassembly of a certain protein due to mutation or of various proteins due to environmental conditions (such as low energy), ER-associated protein degradation (ERAD) kicks in or entire sections of the ER are destroyed by autophagy (ER-phagy) (Figure 2, upper right section). When these destructive mechanism fail in resolving the situation, signal transduction mechanisms are activated, which increase the capacity for folding and the destructive mechanisms in the affected cells, termed unfolded protein response (UPR), or initiate programmed cell death/apoptosis in order to save the organ or organism (Figure 2, lower right section). Besides these stress-related signal

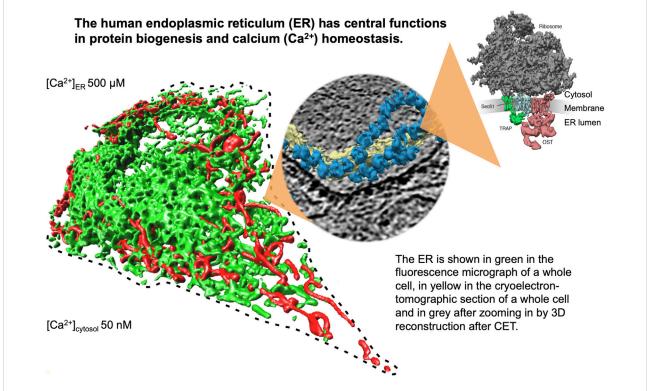


FIGURE 1

The endoplasmic reticulum (ER) of nucleated human cells has major functions in cellular calcium homeostasis, signal transduction and protein biogenesis. Going from left to right, it is shown here after fluorescence microscopy following expression of a GFP-targeted ER protein and a RFP-targeted mitochondrial protein, after cryoelectron tomography of a HeLa cell, and after 3D reconstruction of the human Sec61 complex together with its interaction partners ribosome, TRAP and oligosaccharyltransferase (Pfeffer et al., 2015). The Figure and Figure legend were adapted from Sicking et al. (2021).

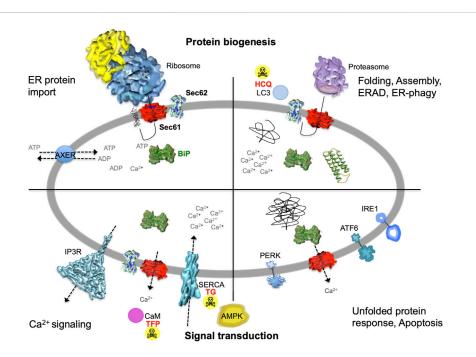


FIGURE 2
The heterotrimeric Sec61 complex, Sec62, IP3R and SERCA in the ER membrane are major players in protein biogenesis and Ca²⁺ -dependent signal transduction at the ER. These proteins are shown here in a schematic cross section of the human ER together with additional proteins and the various functions of the human ER. The non-annotated structures refer to a not yet-folded polypeptide, a natively folded protein, and an aggregate of non-native polypeptides, respectively. Although connected to the UPR, another level of complexity that is related to energy homeostasis of the ER lumen, particularly the ATP/ADP exchanger in the ER membrane (AXER/SLC35B1) and the ER to cytosol low energy signal transduction pathway, which involves cytosolic AMPK as well as Ca²⁺ efflux from the ER via the Sec61 channel, is not covered in this manuscript for the sake of clarity (Klein et al., 2018; Yong et al., 2019; Zimmermann and Lang, 2020; Schwarzbaum et al., 2022). AMPK, AMP-activated protein kinase; CaM, calmodulin; HCQ, hydroxychloroquine (autophagy inhibitor at the level of lysosomes); IP3R, IP3-receptor; SERCA, sarcoplasmic/endoplasmic reticulum Ca²⁺

ATPase; TFP, trifluoperazine (CaM antagonist); TG, thapsigargin (SERCA inhibitor). The Figure and Figure legend were adapted from Melnyk et al. (2022).

transduction processes, the ER plays a major role in Ca²⁺mediated signal transduction that is closely related to cellular calcium homeostasis (Figure 2, lower left section). In a resting cell, there is an almost one thousand fold Ca2+ gradient between ER and the cytosol (Figure 1). However, upon stimulation of the cell by e.g. a hormone on the cell surface, the second messenger IP3 is produced near the plasma membrane and diffuses through the cytosol to the ER, where it binds to the IP3-receptor (IP3R). Activation of the IP3R causes massive efflux of Ca2+ from the ER and activates various signaling pathways, including increased expression of certain genes. When the cell stimulation subsides, the cell returns to the resting state and a Ca2+ pump in the ER membrane (termed sarcoplasmic/ endoplasmic reticulum calcium ATPase/SERCA) regenerates the steep Ca2+ gradient between ER and cytosol. In addition, SERCA is constantly busy with pumping back Ca²⁺ into the ER lumen that had passively leaked from the ER. Recent years characterized the major ER protein import channel, the Sec61 channel, as one of the Ca2+ leak channels of the ER membrane in all nucleated cells. The Sec61 channel is associated with the ER membrane protein Sec62, which has functions in ER protein import, ER-phagy and keeping passive ER Ca^{2+} efflux via the Sec61 channel at bay (Figure 2).

In the year 2006, a first report associated the overexpression of the SEC62 gene, which is also termed TLOC1 and located at chromosome 3q26, with prostate cancer (Jung et al., 2006) (Figure 3). Subsequent studies identified SEC62 overexpression as a phenomenon associated with prostate cancer progression in patients and, in prostate cancer cells, as the reason for increased ER stress tolerance as well as increased migratory and invasive potential (Greiner et al., 2011a; Greiner et al., 2011b). Thus, the latter observations associated SEC62 overexpression with two hallmarks of cancer (Hanahan and Weinberg, 2000; Hanahan and Weinberg, 2011; Hanahan, 2022) (Figure 4). Therefore, the Sec62 protein was suggested as potential diagnostic marker as well as therapeutic target in prostate cancer (Greiner et al., 2011a). Subsequently, similar observations were made for non-small cell lung cancer (NSCLC) and thyroid cancer (Linxweiler et al., 2012; Linxweiler et al., 2014) and SEC62 overexpression was linked to poor prognosis in NSCLC (Linxweiler et al., 2013). Furthermore, the latter work

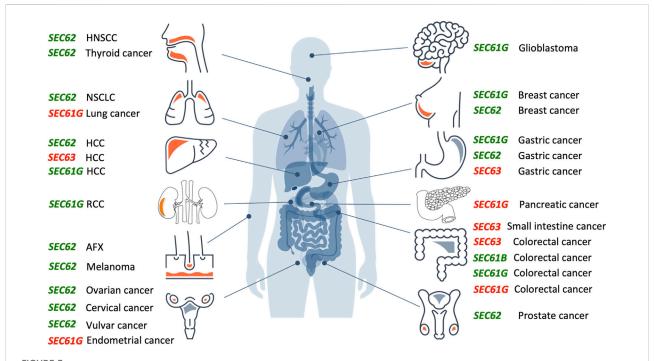


FIGURE 3

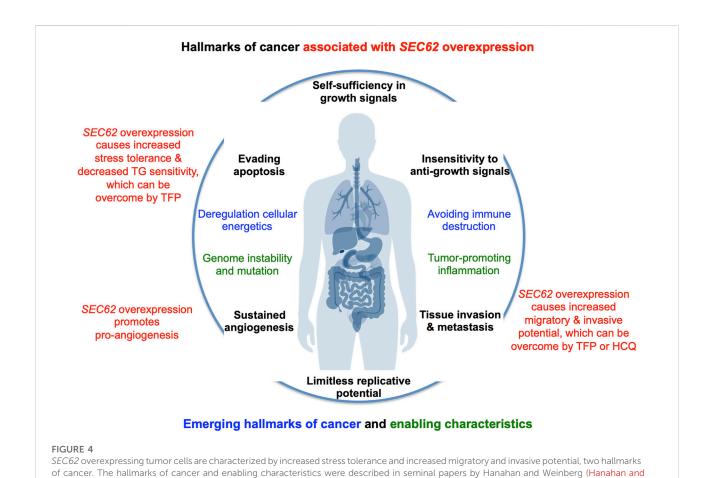
Overproduction or mutation of SEC61, SEC62 and SEC63 genes are associated with various human tumor diseases. Overview of genetic changes and altered expression of SEC61, SEC62, and SEC63 gene in human cancer entities segregated by the tissue of origin (from top left to bottom right: head and neck, lung, liver, kidneys, skin, female genital tract, brain, breast, stomach, pancreas, intestine, male genital tract). Green colored genes symbolize functional gain by overexpression and amplification, red colored genes symbolize functional loss by low expression, deletion, or mutation. AFX, atypical fibroxanthoma; HCC, hepatocellular carcinoma; HNSCC, head and neck squamous cell carcinoma; NSCLC, non-small cell lung cancer; RCC, renal cell carcinoma. The Figure and Figure legend were adapted from Sicking et al. (2021).

characterized a clinically tested small molecule and Calmodulinantagonist (trifluoperazine/TFP) as being able to suppress the effects of SEC62 overexpression on ER stress tolerance and migratory potential of tumor cells. Also in 2013, the SEC62/ TLOC1 gene was characterized as tumor driver gene (Hagerstrand et al., 2013). Next, head and neck squamous cell carcinomas (HNSCC) and cervical cancer of unknown primary (CUP) were added to the list of SEC62 overexpressing tumors (Wemmert et al., 2016; Bochen et al., 2017) as were dysplastic cervical lesions (Linxweiler et al., 2016), atypical fibroxanthoma and melanoma (Müller et al., 2019; Müller et al., 2021), mammary carcinoma/invasive ductal breast cancer, vulvar cancer/vulvar intraepithelial neoplasia, cervical cancer/cervical intraepithelial neoplasia (Takacs et al., 2019a; Takacs et al., 2019b; Takacs et al., 2019c) and gastric cancer (Su et al., 2022). Therefore, it appears that Sec62 may be a diagnostic and prognostic marker as well as a therapeutic target for various tumors, which warranted murine tumor models as the next logical step (Linxweiler et al., 2017). In light of the exciting developments in the field of targeted SERCA-inhibiting prodrugs, developed by John T. Isaacs, Sören B. Christensen and colleagues in the fight against prostate cancer as well as neovascular tissue in various tumors (Treiman et al., 1998; Denmeade et al., 2003, 2012; Doan et al., 2015; Mahalingam et al., 2016), we review the current state of the studies on *SEC62* overexpressing tumors and discuss where, in our opinion, the field should go from here.

Summary of results on mammalian Sec62 protein

Functions of the endoplasmic reticulum membrane protein Sec62

The mammalian Sec62 protein is in the center of a dynamic protein network (Figure 5). It is present in the ER membrane, contains two transmembrane helices, a short ER-lumenal loop plus two large cytosolic domains, forms a heterodimeric complex with the Sec63 protein and transiently associates with the heterotrimeric Sec61 complex (Daimon et al., 1997; Mayer et al., 2000; Tyedmers et al., 2000). These interactions involve a positively charged patch of amino acid residues in the N-terminal domain of Sec62 plus a negatively charged cluster at the C-terminus of Sec63 and the C-terminal domain of Sec62 (including two



Weinberg, 2000; Hanahan and Weinberg, 2011). The red texts refer to the hallmarks that are linked to SEC62 overexpressing cancer cells (Greiner

predicted EF hands) plus the N-terminus of Sec61a, respectively (Müller et al., 2010; Linxweiler et al., 2013). Notably, the Sec62/Sec63 interaction is enhanced by CK2mediated phosphorylation within the negatively charged patch of Sec63 (Ampofo et al., 2013) and the Sec62/ Sec61 interaction is sensitive to Ca2+, likely involving the EF hands in Sec62 (Linxweiler et al., 2013). Cytosolic interaction partners of Sec62 are Ca2+ (the two predicted EF hands), ribosomes and LC3 and involve at the level of Sec62 the putative EF hands, a ribosome binding site (RBS) and a LIR-motif within the more C-terminal EF hand, which are relevant for cellular calcium homeostasis, ER protein import and ER-phagy, respectively (Müller et al., 2010; Linxweiler et al., 2013; Fumagalli et al., 2016). The αsubunit of the Sec61 complex contains ten transmembrane helices, interacts with ribosomes via two RBS and contains an IQ-motif at its cytosolic N-terminus, which provides a binding site for the cytosolic EF hand protein calmodulin (CaM) (Kalies et al., 1994; Erdmann et al., 2011; Cheng et al., 2015). The Sec63 protein contains three transmembrane

helices and two domains in cytosol and ER lumen,

et al., 2011a; Greiner et al., 2011b; Linxweiler et al., 2013).

respectively, and comprises a binding site nucleoredoxin (NRX) near the cytosolic C-terminus and an ER-lumenal J-domain for the interactions with the chaperone BiP plus the EF hand protein calumenin (Tyedmers et al., 2005; Weitzmann et al., 2007; Dudek et al., 2009; Müller et al., 2011; Melnyk et al., 2022). The roles of the J-domain of Sec63 are to recruit BiP to the Sec62/ Sec63 complex, to stimulate the ATPase activity of BiP and, thereby, to allow activated BiP to bind a substrate, such as an oligopeptide within ER-lumenal loop seven of Sec61a (Weitzmann et al., 2007; Schäuble et al., 2012). Notably, BiP is present in the ER lumen at millimolar concentration, comprises a nucleotide-binding domain (NBD) plus a substrate binding domain (SBD) as well as multiple low affinity Ca2+ binding sites, and depends on ATP as well as a high ER-lumenal Ca²⁺ concentration for its activity (Melnyk et al., 2022). Sil1 is one of the nucleotide exchange factors for BiP, Grp170 is another one (Weitzmann et al., 2006). BiP also interacts with the ER-lumenal domains of ATF6, IRE1 and PERK and, thus, is connected to the UPR (Walter and Ron, 2011). In addition, IRE1a interacts with Sec61 and, thus,

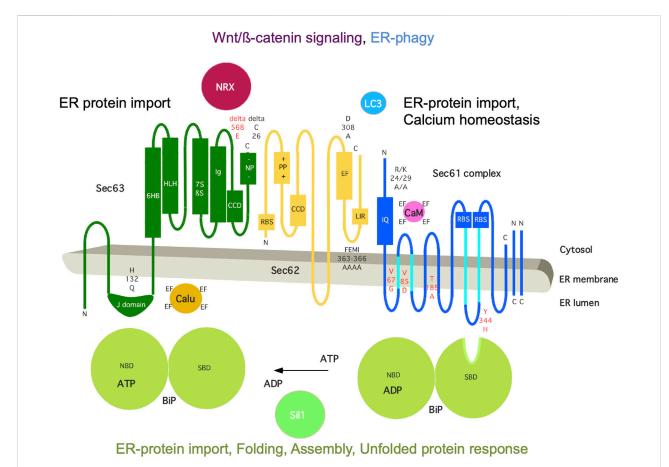


FIGURE 5

The human ER membrane protein Sec62 is part of a dynamic protein network. Topology and functionally relevant domains of the heterotrimeric Sec61 complex and its allosteric effectors BiP, Sec62 and Sec63 are shown (Mayer et al., 2000; Tyedmers et al., 2000). The cytosolic loops six and eight of the α-subunit of the Sec61 complex comprise a binding site for ribosomes (termed RBS) and ER-lumenal loop 7 a binding site for BiP (Schäuble et al., 2012; Cheng et al., 2015); the N-terminus of the same subunit contains an IQ-motif for binding of Ca²+-calmodulin (CaM) (Erdmann et al., 2011). The cofactors of BiP, Sec63 and Silt, are also shown, as are additional interaction partners of Sec62 (LC3) and Sec63 (nucleoredoxin/NRX, calumenin/Calu) (Tyedmers et al., 2005; Müller et al., 2011; Fumagalli et al., 2016). Disease-associated mutations of Sec61α and Sec63 are indicated in red (amino acid residues are given in single letter code) (reviewed by Sicking et al., 2021), as are rationally designed mutations in black, such as Sec61α^{R24A/K29A} (Erdmann et al., 2011), Sec62^{D308A} (Linxweiler et al., 2013), Sec62^{FEMIS63-366AAA} (Fumagalli et al., 2016), and Sec63^{H132O} as well as Sec63^{deltaC26} (Müller at al., 2010; Haßdenteufel et al., 2018; Schorr et al., 2020). C, C-terminus; CCD, coiled-coil domain for protein interaction; EF, EF hand; IQ, IQ-motif for CaM binding; LIR, LIR-motif for LC3 binding; N, N-terminus; NBD, nucleotide binding domain; -NP-, patch with negatively charged amino acid residues; +PP+, patch with positively charged amino acid residues; RBS, ribosome binding site; SBD, substrate binding site. The Figure and Figure legend were adapted from Sicking et al. (2021).

directly links ER protein import and Ca^{2+} leakage with UPR (Sundaram et al., 2017; Li et al., 2020). Interestingly, NRX interacts with protein Dishevelled 1 (Dvl1) and, thus, is linked to the Wnt signaling pathway and cadherin-controlled cell adhesion (Nelson and Nusse, 2004; Nusse and Clevers, 2017). It was proposed that oxidative stress simultaneously inhibits NRX/Dvl1 interaction and stimulates NRX/Sec63 interaction, thereby activating Wnt/ β -catenin signaling as well as affecting β -catenin/cadherin-controlled cell adhesion, which both are linked to cell migration and various cancers (Müller et al., 2011).

The Sec61 complex in the membrane of the ER provides the major entry point for precursor polypeptides with either N-terminal signal peptides or equivalent transmembrane helices into the ER lumen and membrane, respectively (Figures 1, 5) (reviewed by Lang et al., 2017; Lang et al., 2019). In the early phase of co- and posttranslational membrane translocation or integration, the signal peptides or transmembrane helices of precursor polypeptides are targeted to the Sec61 complex by various pathways. Next, binding of the precursor proteins to the closed Sec61 complex triggers opening of an aqueous polypeptide-conducting channel, which is formed by Sec61a (Pfeffer et al., 2015). This occurs spontaneously after binding of precursors with strong signal peptides to the channel or with additional support from auxiliary components, such as TRAP

(Figure 1, right) or Sec62/Sec63 plus BiP (Figure 5). Thus, while the Sec61-complex mediates import of most precursor polypeptides into the ER, the Sec61-associated Sec62/ Sec63 heterodimer supports ER protein import in a precursor-specific manner, typically in cooperation with the ER-lumenal Hsp70-type molecular chaperone BiP. Analyses of protein transport in human and murine cells showed a client-specific role of Sec62 in ER protein import, such as the posttranslational ER import of presecretory proteins with a content of less than 100 amino acid residues (Lakkaraju et al., 2012; Lang et al., 2012; Haßdenteufel et al., 2018; Haßdenteufel et al., 2019). However, these small precursor proteins do not represent the complete picture of Sec62 substrates, since the human Sec62/Sec63 complex was also found to be involved in cotranslational import of certain large precursor polypeptides, such as prion protein. Notably, the mammalian Sec62 protein is able to interact with the ribosome and to recruit, in collaboration with Sec63, BiP to the Sec61 channel. The latter interaction facilitates Sec61 channel opening for large precursors with weak signal peptides (Ziska et al., 2019; Schorr et al., 2020), the idea being that binding of BiP to loop seven of Sec61α provides binding energy for shifting the dynamic equilibrium of the Sec61 channel to the open state (Schäuble et al., 2012). The BiP binding site in Sec61a was characterized as a di-tyrosine motif-containing mini-helix in ER-lumenal loop 7 (Figure 5). Interestingly, homozygous mutation of tyrosine 344 to histidine in this loop seven is linked to Diabetes mellitus in mice and compromises co- and posttranslational ER import of Sec63-plus BiP-dependent precursor polypeptides when introduced into HeLa cells. Furthermore, BiP can bind to incoming precursor polypeptides and act on these as a molecular ratchet, which guarantees unidirectional transport (Tyedmers et al., 2003). Typical for an Hsp70, both BiP activities involve ATP, Ca²⁺, J-domain-proteins or Hsp40-type co-chaperones, such as Sec63, and nucleotide exchange factors, such as Sil1 (Melnyk et al., 2022). Following the same principles and interactions, BiP also plays a central role in folding and assembly of newly-imported polypeptides, such as immunoglobulins in the plasma cells of the immune system (reviewed by Melnyk et al., 2022), and supports efficient Sec61 channel closing to preserve Ca2+ homeostasis (Schäuble et al., 2012), which will be discussed next. In addition, BiP is a key player in various Ca2+-dependent and-independent signal transduction pathways, which report on ER energy- and protein-homeostasis (see below).

The mammalian ER also represents the major Ca²⁺ storage compartment in nucleated cells and allows the controlled release of Ca²⁺ from the ER. Interestingly, the open polypeptide-conducting Sec61 channel allows the passive passage of Ca²⁺ from the ER and, therefore, needs to be closely controlled (Wirth et al., 2003; Lang et al., 2011).

Sec61 channel closing either occurs spontaneously or is facilitated by allosteric effectors, such as the ER-lumenal BiP and/or the cytosolic Ca²⁺-calmodulin (Ca²⁺-CaM) (Erdmann et al., 2011; Schäuble et al., 2012). Originally, single-channel recordings from planar lipid bilayers characterized the Sec61 complex as a highly dynamic aqueous channel that is transiently opened by signal peptides and permeable to Ca2+ after completion of protein import (Wirth et al., 2003; Erdmann et al., 2011). The same biophysical approach showed that the Sec61 channel closure can be induced by binding of BiP or Ca²⁺-CaM (Erdmann et al., 2011). Next, the fact that BiP is involved in closing the Sec61 channel was confirmed at the cellular level by combination of siRNA-mediated gene silencing or pharmacological manipulation and live cell Ca2+ imaging (Schäuble et al., 2012). In addition, cytosolic Ca2+-CaM was shown under similar conditions to contribute to Sec61 channel closing via an unrelated mechanism after Ca2+ has started to leak from the ER (Erdmann et al., 2011). During the last 15 years, additional siRNA-mediated gene silencing and live cell Ca2+ imaging experiments identified the ER-lumenal J-domain proteins ERj3 and ERj6 as specific co-chaperones of BiP for channel closure and the putative EF hand- and Ca²⁺- binding protein Sec62 as a co-factor of CaM in Sec61 channel closing (Figures 2, 5) (Linxweiler et al., 2013; Schorr et al., 2015). The binding site of BiP was identified as the above-mentioned di-tyrosine motif-containing mini-helix within ER-lumenal loop seven of the Sec 61α and was shown to be relevant to the described mechanisms by mutagenesis studies (Schäuble et al., 2012). Again, the idea is that binding of BiP to loop seven of Sec61a provides binding energy for shifting the dynamic equilibrium of the Sec61 channel to the closed state. In case of inefficient channel closure in intact cells, Ca²⁺ starts to leak from the ER into the cytosol and binds calmodulin in the ER vicinity, and Ca²⁺-CaM is recruited to the IQ-motif in the Sec61 α-subunit (Erdmann et al., 2011) (Figures 2, 5). Once more, the involved binding energy favors channel closure. Apparently, binding of Ca2+-CaM is supported by Sec62, which may bind Ca2+ because of its putative EF hands within its cytosolic C-terminal domain and lets go of the Sec61 α-subunit (Linxweiler et al., 2013). After the Sec61 channel is closed, and Ca2+ leakage subsides, SERCA pumps Ca2+ back into the ER, CaM and Sec62 return to the respective Ca2+-free forms, and the next protein import cycle can be initiated. When these mechanisms fail, however, the passive Ca²⁺ efflux from the ER may lower BiP activity and, therefore, cause protein misfolding, which may be reported via the UPR, and, eventually lead to apoptosis.

Recently, a function of the mammalian Sec62 protein beyond its house-keeping functions in ER protein import and calcium homeostasis was found and, in contrast to the latter two, is activated by demand, possibly by

dephosphorylation of the C-terminal domain of Sec63 or by Ca²⁺ binding to the predicted EF hands of Sec62. Fumagalli et al. observed that monomeric Sec62 plays a crucial role in the recovery of human and murine cells from ER stress (Fumagalli et al., 2016). The term ER stress describes conditions of globally disturbed folding and assembly of proteins in the ER, which can be caused e.g. by reduced ER levels of ATP or Ca2+. Depending on the stress severity, the cell can either stimulate compensatory mechanisms such as the UPR or initiate apoptosis, which may involve Sec61 channel mediated Ca2+ efflux from the ER (Hara et al., 2014; Feliziani et al., 2020). During UPR, synthesis of the majority of proteins is inhibited while the production of ER chaperones such as BiP is drastically increased in order to facilitate the correct folding of ER-lumenal polypeptides. In the case that this rescue operation fails, misfolded proteins are degraded by the proteasome via ERAD or via ER-phagy (Pisoni and Molinari, 2016; Molinari, 2020). If the cell succeeds in coping with the ER stress, the expanded ER with its increased amount of ER-lumenal chaperones has to be brought back to a physiological size.

Therefore, small vesicles bearing ER-lumenal chaperones are pinched-off from the ER and engulfed by endolysosomes for degradation in an ER-phagy-related process, termed piecemeal micro-ER-phagy (Loi et al., 2019) (Figure 2). In this context, Sec62 was shown to bear a LIR-motif at its C-terminus functioning as a receptor for LC3 during recovery from ER stress e.g. as induced by the reversible SERCA inhibitor cyclopiazonic acid (Fumagalli et al., 2016). The interaction between the LIR-motif and LC3 is sensitive to mutation of the tetrapeptide 363FEMI366 to tetraalanine (Figure 5). Thus, Sec62 plays an important, Sec63-independent, role during compensation of ER stress, termed recovER-phagy. It remains to be seen if Sec62 also plays a general role in ERphagy that is involved in fighting ER stress and how it is activated for its role in these processes. It would not come as a surprise if the interaction of IRE1 α with Sec61, which links ER protein import and Ca2+ leakage with UPR, would play a role in one way or another (Sundaram et al., 2017; Li et al., 2020).

SEC61, SEC62 and SEC63 gene overexpression or mutation associated with human tumor diseases

Over the past 15 years, increasing evidence suggests a relevant role of *SEC61*, *SEC62* and *SEC63* genes in the development and tumor cell biology of human malignancies (Figure 3). For the *SEC61B* gene altered expression was reported for colorectal cancer (reviewed by Fan et al., 2011). Beginning in 2006, for the *SEC61G* gene

increased expression and gene amplification were reported for breast cancer, glioblastoma, gastric cancer, colorectal cancer, renal cancer and lung adenocarcinoma (Reis-Filho et al., 2006; Tsukamoto et al., 2008; Lu et al., 2009; Meng et al., 2021a; Chen et al., 2010; Xu et al., 2022). In glioblastoma multiforme, Liu et al. observed a significant correlation of high SEC61G expression with poor prognosis based on statistical analysis of data from the Cancer Genome Atlas cohort and the Chinese Glioma Genome Atlas cohort (Liu et al., 2019). Univariate and multivariate Cox proportional hazards regression verified SEC61G as an independent factor for prognosis and therapeutic outcome in these cohorts. Furthermore, mining of the COSMIC database identified in 2021 six mutations in the SEC61G gene that were associated with colorectal, endometrial, pancreatic and lung cancer, respectively, and linked to disturbed cellular calcium homeostasis due to altered Sec61 channel gating (Witham et al., 2021). In 2002, a first publication described frameshift mutations of SEC63 due to microsatellite instability in 38% of gastric cancers and 49% of colorectal cancers (Mori et al., 2002). Similar results were reported in 2005 and 2013 (Eschrich et al., 2005; Schulmann et al., 2005; Casper et al., 2013; Casper et al., 2021), where microsatellite instability associated SEC63 frameshift mutations were found in 56% of small-bowel cancers in patients with hereditary non-polyposis colorectal cancer (HNPCC) and in one case of hepatocellular carcinoma (HCC).

However, most extensive evidence for a causative role of a protein translocation component in the development and tumor cell biology of human cancer exists for the ER transmembrane protein Sec62. In 2006, a first study found SEC62 copy number gains in 7 of 13 prostate cancer samples as well as elevated Sec62 protein levels in three prostate cancer cell lines (Jung et al., 2006). In the following years, amplification and overexpression of the SEC62 gene were reported for various other cancer entities, including nonsmall cell lung cancer (NSCLC) (Greiner et al., 2011a; Linxweiler et al., 2012), thyroid cancer (Greiner et al., 2011a; Linxweiler et al., 2012), hepatocellular cancer (Weng et al., 2012; Du et al., 2019), ovarian cancer (Hagerstrand et al., 2013; Radosa et al., 2022), breast cancer (Hagerstrand et al., 2013; Takacs et al., 2019a), head and neck squamous cell carcinoma (HNSCC) (Wemmert et al., 2016; Bochen et al., 2017), cervical cancer (Takacs et al., 2019b), vulvar cancer (Takacs et al., 2019c), atypical fibroxanthoma (Müller et al., 2019), melanoma (Müller et al., 2021), gastric cancer (Su et al., 2022) and in larger prostate cancer patient cohorts (Greiner et al., 2011b). After screening the cBio portal for cancer genomics from over 72,000 cancer patients with 55 different tumor entities SEC62 gene alterations were reported for 2,595 patients and represented gene amplifications in the majority of cases (Cerami et al., 2012; Gao et al., 2013; Sicking et al., 2021). Up to now, SEC62 overexpression was linked to poor prognosis in prostate

cancer (Jung et al., 2006), hepatocellular cancer (Weng et al., 2012; Du et al., 2019), NSCLC (Linxweiler et al., 2013), HNSCC (Wemmert et al., 2016; Bochen et al., 2017), breast cancer (Takacs et al., 2019a), melanoma (Müller et al., 2021), colorectal cancer (Liu et al., 2021), ovarian cancer (Radosa et al., 2022) and gastric cancer (Su et al., 2022).

SEC62 overexpressing tumor cells have increased stress tolerance and migratory as well as invasive potential

From a functional point of view only few studies addressed the specific functional impact of altered SEC62 expression levels on cancer cell biology. A first step to uncover potential associations of SEC62 overexpression with tumor cell biology were correlative analyses with clinical data. Thereby, Greiner et al. found an association of high SEC62 expression level with higher Gleason Score in prostate cancer (Greiner et al., 2011b). In non-small cell lung cancer, high Sec62 levels correlated with the occurrence of lymph node metastases and tumor cell dedifferentiation (Linxweiler et al., 2012). Similarly, an association of SEC62 overexpression with lymphatic metastases was reported for head and neck squamous cell carcinoma (Bochen et al., 2017) as well as an association of SEC62 overexpression with distant metastases in breast cancer (Takacs et al., 2019a). These results indicated a potential role of Sec62 in cancer metastasis, which was strengthened by several functional studies. As a second step, a significant inhibition of cancer cell migration by SEC62 gene silencing was reported for prostate cancer cells (Greiner et al., 2011b), non-small cell lung cancer cells as well as thyroid carcinoma cells (Linxweiler et al., 2012), cervical cancer cells (Linxweiler et al., 2016), hepatocellular carcinoma cells (Du et al., 2019; Li et al., 2019) and head and neck squamous cell carcinoma cells (Bochen et al., 2017). Where tested, overproduction of a mutated Sec62, Sec62^{D308A} (Figure 5) that affected the more N-terminal EF hand had a similar effect on cell migration as SEC62 silencing, i.e. an even dominant negative effect over Sec62 (Linxweiler et al., 2013). Strikingly, *SEC62* overexpression stimulated the migratory potential and stress tolerance of otherwise poorly migrating and poorly stress tolerant cells such as HEK293, HeLa, Huh-7 and FaDu cells (Greiner et al., 2011a; Linxweiler et al., 2013; Bochen et al., 2017; Du et al., 2019). Furthermore, an influence of SEC62 expression level on stress tolerance of human cancer cells was suggested by several studies that reported a higher ER stress sensitivity induced by SEC62 silencing, which was phenocopied by overproduction of Sec62 $^{\hspace{-0.5cm} D308A}$ or administration of CaM antagonists, such as TFP (Greiner et al., 2011b; Linxweiler et al., 2012; Linxweiler et al., 2016).

These findings raised the question of how SEC62 overexpression causes increased stress tolerance and migratory and invasive potential of the respective cancer cells:

- 1) On first sight, the role of Sec62 in ER protein import may appear to be an unlikely candidate since the protein typically cooperates with Sec63 and simultaneous SEC63 overexpression was not observed in SEC62 overexpressing tumor cells (Greiner et al., 2011a). However, in our opinion it cannot entirely be dismissed since in some cases Sec62 was found to play a role in ER protein import without the involvement of Sec63 (Haßdenteufel et al., 2018; Lang et al., 2022). In this context it is tempting to speculate that the biogenesis of ADAM metalloproteinases, which are involved in proteolytic cleavage of extracellular matrix and cell adhesion proteins, as well as cadherins that play a direct role in cell adhesion, is limited by standard Sec62 levels. Therefore, it may be improved by increased levels of Sec62, thereby supporting cell migration. Indeed, the two αsecretases ADAM10 and ADAM17 were found to depend on Sec62 but not Sec63 in their ER import in HeLa cells (reviewed by Lang et al., 2022). As a caveat, however, loss of cadherin expression was found to promote tumorigenesis (Nelson and Nusse, 2004).
- 2) Therefore, the increased suppression of Ca2+ efflux via the Sec61 channel by Sec62 and the activated ER-phagy in SEC62 overexpressing cancer cells may appear as the more likely reasons for the improved migratory potential and stress tolerance (Erdmann et al., 2011; Fumagalli et al., 2016; Linxweiler et al., 2016; Bergmann et al., 2017). Increased suppression of Ca²⁺ efflux via the Sec61 channel by increased levels of Sec62 may prevent apoptosis, which involves an elevated cytosolic Ca²⁺ concentration, in combination with an increased capacity for ER-phagy due to the excess of free Sec62 may allow the cells to defend themselves instantly and more efficiently against ER stress and, thereby, also to prevent apoptosis. As another caveat, however, the mutations of the SEC61G gene that were associated with colorectal, endometrial, pancreatic and lung cancer were found to cause increased Ca2+ efflux from the ER due to altered Sec61 channel gating (Witham et al., 2021).

Overall, we favor the idea that all these Sec62 functions may be responsible for the increased stress tolerance and migratory potential of the SEC62 overexpressing cancer cells. Indeed, SEC62 overexpression but not overproduction of the EF hand mutant variant $Sec62^{D308A}$ allowed cells to tolerate higher levels of the irreversible SERCA inhibitor thapsigargin (TG) (Treiman et al., 1998; Lindner et al., 2020). Notably, however, cytosolic Ca^{2+} also plays a role in cell migration (Tsai et al., 2015), which has to be considered as a possible contributing factor, and the Sec63 interaction partner NXN may be relevant here by activating Wnt/ β -catenin signaling or affecting β -catenin/cadherin-controlled cell adhesion (Nelson and Nusse, 2004; Müller et al., 2011; Nusse and Clevers, 2017). Furthermore, the recently reported observation that SEC62 overexpression leads to overproduction of PAI-1 (coded by the SERPINE1

gene) and TNFRSF11B and the putative resulting hypoxia-induced tube formation as well as stimulated Wnt/ β -catenin signaling also have to be taken into future account (Meng et al., 2021b). Notably, it was suggested that it needs to be addressed if autophagy inhibitors such as bafilomycin A1, chloroquine (CQ) and hydroxychloroquine (HCQ) can cause decreased stress tolerance and/or decreased migratory as well as invasive potential (Bergmann et al., 2017). Alternatively, the overexpression of the recovER-phagy-deficient mutant variant Sec62^{FEMI363-366AAAA} should be tested as a potential cause for increased stress tolerance as well as increased migratory and invasive potential.

Depletion of Sec62 from tumor cells or treatment of the cells with the drug trifluoperazine reduce stress tolerance and migratory as well as invasive potential

Taken together, these data strongly indicate a role of SEC62 as a driver oncogene in various human cancers, which show a consistent association with poor prognosis, lymph node as well as distant metastasis and stress tolerance. This turns the Sec62 protein into an attractive target for anticancer therapy (Greiner et al., 2011a; Linxweiler et al., 2017). Since SEC62-targeting siRNAs are not a therapeutic option at present and Sec62 is not accessible to monoclonal antibodies due to its intracellular location, alternative strategies had to be developed to achieve at least a functional knock-down in SEC62 overexpressing tumors.

Cell death triggered by unmitigated ER stress is seen as an important potential therapeutic strategy in cancer therapy (Denmeade et al., 2003; Lindner et al., 2020). Based on the role of Sec62 in the regulation of Ca²⁺ efflux through the Sec61 channel, CaM antagonists (e.g. TFP) were investigated as potential therapeutics at the level of cancer cell lines (Vandonselaar, et al., 1994; Linxweiler et al., 2013). Notably, TFP had previously been in clinical use for the treatment of schizophrenia (Marques et al., 2004). Interestingly, CaM antagonists such as TFP mimicked a functional Sec62 knockdown by stimulating Ca²⁺ efflux from the ER and inhibiting the migratory potential of cervix and prostate cancer cells, which was linked to inhibition of proliferation at lower doses of TG (Linxweiler et al., 2012; Linxweiler et al., 2016).

These findings on TFP have to be seen in light of the exciting developments in the field of SERCA-targeting prodrugs, developed by John T. Isaacs, Sören B. Christensen and colleagues, originally in the fight against prostate cancer as well as more recently directed at neovascular tissue in various tumors (Denmeade et al., 2003; Denmeade et al., 2012; Doan et al., 2015; Mahalingam et al., 2016). Briefly, prostate as well as vascular cells, present in a broad number of tumors, such as bladder-, breast-, hepatocellular-, renal-cancer and glioblastoma

multiforme, harbor the proteolytic enzyme that is termed prostate specific membrane antigen (PSMA) in their plasma membranes. Therefore, these cells can be specifically targeted by TG-conjugates, which comprise TG in combination with oligopeptides that are substrates for PSMA. Upon cleavage of the prodrug conjugates by PSMA, TG is selectively liberated in the vicinity of the target cells, inhibits SERCA in these cells and drives them into programmed cell death. According to experiments with tumor cell lines, however, SEC62 overexpressing tumor cells may be unresponsive or resistant against such treatment (Linxweiler et al., 2013). Therefore, murine tumor models for tumor growth or metastasis were evaluated with respect to their responsiveness to treatment with the mipsagargin/G202 analog TG in the presence of TFP, which had previously been in clinical use for the treatment of schizophrenia, and will be discussed next.

Trifluoperazine reduces tumor growth and metastatic potential in murine tumor models

Hagerstrand et al. were the first to demonstrate induction of subcutaneous tumor growth in C. Cg/AnNTac-Foxn1^{nunu} mice, inoculated with SEC62 overexpressing HMLE cells (Hagerstrand et al., 2013). Because of the promising in vitro results for TFP, in vivo studies in form of murine tumor models appeared to be warranted as the next logical step during recent years. One first in vivo study reported a significant inhibition of seeding and growth of a subcutaneously injected head and neck squamous cell carcinoma cell line (FaDu) in BALB/cAnNRj-Foxn1^{nu}/ Foxn1^{nu} mice by single and combined treatment with TFP and TG (Körbel et al., 2018). A second in vivo study focusing on lymphatic metastases addressed if the migration inhibition, which was found for various cancer cells in vitro, manifests as a clinically relevant phenotype in a living organism and observed a tendency of both drugs to suppress metastasis rate as well as to reduce metastasis size in an orthotopic xenograft mouse model of hypopharyngeal squamous cell carcinoma (Körner et al., 2022). Within the scope of such therapeutic concepts, the benefit of autophagy inhibitors was recently tested both at the level of gastric cancer cells and in a murine metastasis model for gastric cancer (Su et al., 2022). The striking observations were that SEC62 overexpression affects the levels of the matrix metalloproteinases MMP2 and MMP9 plus their inhibitor TIMP-1 and promotes the migratory and invasive potential of gastric cancer cells. In addition, it was observed that autophagy blockade by hydroxychloroquine (HCQ) indeed impairs the promotive effects of SEC62 overexpression on the migratory and invasive potential (Figure 4). Thus, it was concluded that the combination of reduction of the Sec62 level by lentiviral

shRNA and autophagy blockade exerts a synergistic antimetastatic effect and, therefore, represents a promising therapeutic strategy for metastases of gastric cancers (Su et al., 2022). However, the mechanism of TIMP-1 to MMP2 plus MMP9 balance change under conditions of SEC62 overexpression remains to be elucidated. Based on the observation that the ER import of ADAM metalloproteinases ADAM10 and ADAM17 and the protease inhibitor SERPINE2 depends on Sec62 (Lang et al., 2022), a direct role of SEC62 overexpression on ER import of TIMP-1 or MMP2 and MMP9 may also be considered. Furthermore, the possible influence of HCQ on ER stress tolerance remains to be analysed.

Discussion

Against the background of stimulation of tumor metastasis and the increased tolerance to ER stress and worsened prognosis in SEC62 overexpression, Sec62 presents itself as a potential target structure of a targeted antitumor therapy (Greiner et al., 2011a; Greiner et al., 2011b) (Figures 2, 4). Although, as previously described, several in vitro investigations observed that the migratory and metastatic potential of human tumor cells can significantly be reduced by a SEC62 siRNA transfection (Greiner et al., 2011a; Linxweiler et al., 2012, 2016; Bochen et al., 2017), transfer of this technology to humans does not appear possible at present. So far, it is due to significant systemic side effects and too low concentrations at the target site that small RNA molecules have not yet succeeded in tumor therapeutics in humans. However, the possibility of a targeted in vivo CRISPR/Cas9-based genome editing may represent a promising future approach, as may be true for microRNAs (He et al., 2019). In search of alternative mechanisms to inhibit Sec62 function, it was shown that treating tumor cells with the CaM antagonist TFP both in terms of migratory potential and cellular calcium homeostasis achieves the same phenotype as SEC62 silencing (Linxweiler et al., 2013) (Figure 4). However, the possible influence of TFP on pro-angiogenesis was not yet analysed. Since the CaM antagonist TFP was in clinical use for the treatment of psychiatric illnesses for many years, for this drug already exists extensive experience with a human application. Furthermore, the repurposing of this substance as a tumor therapeutic has already been developed by a few groups in recent years and has been investigated with promising results, in some cases in combination with bleomycin (Polischouk et al., 2007; Pulkoski-Gross et al., 2015; Sullivan et al., 2002; Sangodkar et al., 2012; Yeh et al., 2012; Zacharski et al., 1990; Zhelev et al., 2004). However, the possible influence of SEC62 overexpression on the TFP effect was not yet analysed in these studies. Based on the in vitro data, we expect that the TFP effect may be increased by an additional treatment with an inhibitor of the SERCA, in particular, since a dose-dependent proliferation inhibition was achieved with this combination in vitro. Currently, several groups already deal with the application of TG or of TG-related prodrugs (such as mipsgargin) as potential tumor therapeutics in advanced solid tumor diseases (Denmeade et al., 2003, 2012). A clinical phase I study (Dose-Escalation Phase 1 Study of G-202 in patients with advanced solid tumors, Clinical Trials Government Identifier NCT01056029) showed favorable pharmacokinetic properties with acceptable toxicity in the treated patients (Mahalingam et al., 2016). Furthermore, phase II clinical trials with the prodrug mipsagargin/ G202 are under way (Doan et al., 2015). Advanced investigations, and in particular clinical studies, will be required to evaluate clinical utility and response to therapy for SEC62 overexpressing tumors with TFP and TFP plus TGrelated prodrugs, which also affect angiogenesis because of the PSMA content of vascular cells (Doan et al., 2015). Along these same lines, based on the fascinating recent results on autophagy inhibitor HCQ in the metastases formation of SEC62 overexpressing gastric tumors (Su et al., 2022), combinatorial therapies of HCQ plus TFP and HCQ plus the targeted mipsagargin/G202, respectively, should be considered as potential future therapeutic options. Notably, CQ and HCQ are used in malaria prophylaxis and therapy and discussed in the therapy of various cancer entities (reviewed by Levy et al., 2017).

Author contributions

JZ and RZ wrote the original manuscript. All authors contributed to the manuscript and agreed to the final version of the manuscript.

Funding

JZ was supported by HOMFOR. Furthermore, the authors acknowledge support from the German Research Foundation (DFG) and Saarland University within the Open Access Publication Funding program.

Acknowledgments

The authors are grateful to their former colleague in Homburg Prof. Bernd Wullich (Department of Urology and Pediatric Urology, Erlangen University Medical Center, Erlangen, Germany) for drawing their attention to the overexpression of the *SEC62* gene in prostate cancer, to Lars Kästner and Peter Lipp (both Saarland University) for the fluorescence micrograph in Figure 1, and to Stefan Pfeffer and

Friedrich Förster (both formerly at the Max-Planck Institute of Biochemistry, Martinsried, Germany).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

References

Ampofo, E., Welker, S., Jung, M., Müller, L., Greiner, M., Zimmermann, R., et al. (2013). CK2 phosphorylation of human Sec63 regulates its interaction with Sec62. *Biochim. Biophys. Acta* 1830, 2938–2945. doi:10.1016/j.bbagen.2012.12.020

Bergmann, T. J., Fumagalli, F., Loi, M., and Molinari, M. (2017). Role of SEC62 in ER maintenance: A link with ER stress tolerance in SEC62-overexpressing tumors? *Mol. Cell. Oncol.* 4, e1264351. doi:10.1080/23723556.2016.1264351

Bochen, F., Adisurya, H., Wemmert, S., Lerner, C., Greiner, M., Zimmermann, R., et al. (2017). Effect of 3q oncogenes SEC62 and SOX2 on lymphatic metastasis and clinical outcome of head and neck squamous cell carcinomas. *Oncotarget* 8, 4922–4934. doi:10.18632/oncotarget.13986

Casper, M., Linxweiler, M., Linxweiler, J., Zimmermann, R., Glanemann, M., Lammert, F., et al. (2021). SEC62 and SEC63 expression in hepatocellular carcinoma (HCC) and tumor-surrounding liver tissue. Visc. Med. 37, 110–115. doi:10.1159/000513293

Casper, M., Weber, S. N., Kloor, M., Müllenbach, R., Grobholz, R., Lammert, F., et al. (2013). Hepatocellular carcinoma as extracolonic manifestation of Lynch syndrome indicates SEC63 as potential target gene in hepatocarcinogenesis. *Scand. J. Gastroenterol.* 48, 344–351. doi:10.3109/00365521.2012.752030

Cerami, E., Gao, J., Dogrusoz, U., Gross, B. E., Sumer, S. O., Aksoy, B. A., et al. (2012). The cBio cancer genomics portal: An open platform for exploring multidimensional cancer genomics data. *Cancer Discov.* 2, 401–404. doi:10.1158/2159-8290.CD-12-0095

Chen, J. S., Chen, K.-T., Fan, C.-W., Han, C.-L., Chen, y.-J., Yu, J.-S., et al. (2010). Comparison of membrane fraction proteomic profiles of normal and cancerous human colorectal tissues with gel-assisted digestion and iTRAQ labeling mass spectrometry. *FEBS J.* 277, 3028–3038. doi:10.1111/j.1742-4658. 2010.07712.x

Cheng, Z., Jiang, Y., Mandon, E. C., and Gilmore, R. (2015). Identification of cytoplasmic residues of Sec61p involved in ribosome binding and cotranslational translocation. *J. Cell Biol.* 168, 67–77. doi:10.1083/jcb.200408188

Daimon, M., Susa, S., Suzuki, K., Kato, T., Yamatani, K., and Sasaki, H. (1997). Identification of a human cDNA homologue to the Drosophila translocation protein 1 (Dtrp1). *Biochem. Biophys. Res. Commun.* 230, 100–104. doi:10.1006/bbrc.1996.5892

Denmeade, S. R., Jakobsen, C. M., Janssen, S., Khan, S. R., Garrett, E. S., Lilja, H., et al. (2003). Prostate-specific antigen-activated thapsigargin prodrug as targeted therapy for prostate cancer. *J. Natl. Cancer Inst.* 95, 990–1000. doi:10.1093/jnci/95. 13.990

Denmeade, S. R., Mhaka, A. M., Rosen, D. M., Brenne, W. N., Dalrymple, S., Dach, I., et al. (2012). Engineering a prostate-specific membrane antigen-activated tumor endothelial cell prodrug for cancer therapy. *Sci. Transl. Med.* 4, 140ra86. doi:10.1126/scitranslmed.3003886

Doan, N. T. Q., Sandholt Paulsen, E., Sehgal, P., Moller, J. V., Nissen, P., Denmeade, S. R., et al. (2015). Targeting thapsigargin towards tumors. *Steroids* 97, 2–7. doi:10.1016/j.steroids.2014.07.009

Du, J., Zhao, Z., Zhao, H., Liu, D., Chen, J., Cheng, B., et al. (2019). Sec62 promotes early recurrence of hepatocellular carcinoma through activating integrina/CAV1 signalling. *Oncogenesis* 8, 74. doi:10.1038/s41389-019-0183-6

Dudek, J., Benedix, J., Cappel, S., Greiner, M., Jalal, C., Müller, L., et al. (2009). Functions and pathologies of BiP and its interaction partners. *Cell. Mol. Life Sci.* 65, 1556–1569. doi:10.1007/s00018-009-8745-y

Erdmann, F., Schäuble, N., Lang, S., Jung, M., Honigmann, A., Ahmad, M., et al. (2011). Interaction of calmodulin with Sec61a limits Ca²⁺ leakage from the endoplasmic reticulum. *EMBO J.* 30, 17–31. doi:10.1038/emboj.2010.284

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Eschrich, S., Yang, I., Bloom, G., Kwong, K. Y., Boulware, D., Cantor, A., et al. (2005). Molecular staging for survival prediction of colorectal cancer patients. J. Clin. Oncol. 23, 3526–3535. doi:10.1200/JCO.2005.00.695

Fan, C.-W., Chan, C.-C., Chen, K.-T., Twu, J., Huang, Y.-S., Han, C.-L., et al. (2011). Identification of $SEC61\beta$ and its autoantibody as biomarkers for colorectal cancer. *Clin. Chim. Acta.* 412, 887–893. doi:10.1016/j.cca.2011.01.012

Feliziani, C., Quasollo, G., Holstein, D., Fernandez, M., Paton, J. C., Paton, A. W., et al. (2020). Endoplasmic reticulum stress induces Ca2+ signaling system initiated by Ca2+ microdomains. *bioRxiv Prepr.* doi:10.1101/2020.10.07. 328849

Fumagalli, F., Noack, J., Bergmann, T., Cebollero, E., Pisoni, G. B., Fasana, E., et al. (2016). Translocon component Sec62 acts in endoplasmic reticulum turnover during stress recovery. *Nat. Cell Biol.* 18, 1173–1184. doi:10.1038/ncb3423

Gao, J., Aksoy, B. A., Dogrusoz, U., Dresdner, G., Gross, B., Sumer, S. O., et al. (2013). Integrative analysis of complex cancer genomics and clinical profiles using the cBioPortal. *Sci. Signal.* 6, pl1. doi:10.1126/scisignal.2004088

Greiner, M., Kreutzer, B., Jung, V., Grobholz, R., Hasenfus, A., Stöhr, R., et al. (2011a). Silencing of the *SEC62* gene inhibits migratory and invasive potential of various tumor cells. *Int. J. Cancer* 128, 2284–2295. doi:10.1002/jic.25580

Greiner, M., Kreutzer, B., Lang, S., Jung, V., Cavalié, A., Unteregger, G., et al. (2011b). Sec62 protein level is crucial for the ER stress tolerance of prostate cancer. *Prostate* 71, 1074–1083. doi:10.1002/pros.21324

Hagerstrand, D., Tong, A., Schumacher, S. E., Ilic, N., Shen, R. R., Cheung, H. W., et al. (2013). Systematic interrogation of 3q26 identifies TLOC1 and SKIL as cancer drivers. *Cancer Discov.* 3, 1044–1057. doi:10.1158/2159-8290.CD-12-0502

Hanahan, D. (2022). Hallmarks of cancer: New dimensions. *Cancer Discov.* 12, 31–46. doi:10.1158/2159-8290.CD-21-1059

Hanahan, D., and Weinberg, R. A. (2011). Hallmarks of cancer: The next generation. *Cell* 144, 646–674. doi:10.1016/j.cell.2011.02.013

Hanahan, D., and Weinberg, R. A. (2000). The hallmarks of cancer. *Cell* 100, 57–70. doi:10.1016/s0092-8674(00)81683-9

Hara, T., Mahadevan, J., Kanekura, K., Hara, M., Lu, S., and Urano, F. (2014). Calcium efflux from the endoplasmic reticulum leads to ß cell death. *Endo* 155, 758. doi:10.1210/en.2013-1519

Haßdenteufel, S., Johnson, N., Paton, A. W., Paton, J. C., High, S., and Zimmermann, R. (2018). Chaperone-mediated Sec61 channel gating during ER import of small precursor proteins overcomes Sec61 inhibitor-reinforced energy barrier. *Cell Rep.* 23, 1373–1386. doi:10.1016/j.celrep.2018.03.122

Haßdenteufel, S., Nguyen, D., Helms, V., Lang, S., and Zimmermann, R. (2019). ER import of small human presecretory proteins: Components and mechanisms. *FEBS Lett.* 593, 2506–2524. doi:10.1002/1873-3468.13542

He, H., Wu, W., Sun, Z., and Chai, L. (2019). MiR-4429 prevented gastric cancer progression through targeting METTL3 to inhibit m6A-caused stabilization of SEC62. *Biochem. Biophys. Res. Commun.* 517, 581–587. doi:10.1016/j.bbrc.2019.

Jung, V., Kamradt, J., Kindich, R., Jung, M., Mueller, M., Schulz, W. A., et al. (2006). Genomic and expression analysis of the 3q25-q26 amplification unit reveals TLOC1/SEC62 as a probable target gene in prostate cancer. *Mol. Cancer Res.* 4, 169–176. doi:10.1158/1541-7786.MCR-05-0165

Kalies, K. U., Görlich, D., and Rapoport, T. A. (1994). Binding of ribosomes to the rough endoplasmic reticulum mediated by the Sec61p-complex. *J. Cell Biol.* 126, 925–934. doi:10.1083/jcb.126.4.925

- Klein, M.-C., Zimmermann, K., Schorr, S., Landini, M., Klemens, P., Altensell, J., et al. (2018). AXER is an ATP/ADP exchanger in the membrane of the endoplasmic reticulum. *Nat. Commun.* 9, 3489. doi:10.1038/s41467-018-06003-9
- Körbel, C., Linxweiler, M., Wemmert, S., Bochen, F., Schick, B., Meyer, M., et al. (2018). Treatment of SEC62 over-expressing tumors by thapsigargin and trifluoperazine. *Biomol. Concepts* 9, 53–63. doi:10.1515/bmc-2018-0006
- Körner, S., Bochen, F., Wemmert, S., Körbel, C., Menger, M., Kühn, J. P., et al. (2022). Antagonizing Sec62 function in intracellular Ca^{2+} homeostasis represents a novel therapeutic strategy for head and neck cancer. *Front. Physiol.* 13, 880004. doi:10.3389/fphys.2022.880004
- Lakkaraju, A. K. K., Thankappan, R., Mary, C., Garrison, J. L., Taunton, J., and Strub, K. (2012). Efficient secretion of small proteins in mammalian cells relies on Sec62-dependent posttranslational translocation. *Mol. Biol. Cell* 23, 2712–2722. doi:10.1091/mbc.E12-03-0228
- Lang, S., Benedix, J., Fedeles, S. V., Schorr, S., Schirra, C., Schäuble, N., et al. (2012). Different effects of Sec61α, Sec62 and Sec63 depletion on transport of polypeptides into the endoplasmic reticulum of mammalian cells. *J. Cell Sci.* 125, 1958–1969. doi:10.1242/jcs.096727
- Lang, S., Erdmann, F., Jung, M., Wagner, R., Cavalié, A., and Zimmermann, R. (2011). Sec61 complexes form ubiquitous ER Ca $^{2+}$ leak channels. *Channels* 5, 228–235. doi:10.4161/chan.5.3.15314
- Lang, S., Nguyen, D., Bhadra, P., Jung, M., Helms, V., and Zimmermann, R. (2022). Signal peptide features determining the substrate specificities of targeting and translocation components in human ER protein import. *Front. Physiol.* 13, 833540. doi:10.3389/fphys.2022.833540
- Lang, S., Nguyen, D., Pfeffer, S., Förster, F., Helms, V., and Zimmermann, R. (2019). "Functions and mechanisms of the human ribosome-translocon complex," in *Macromolecular protein complexes II: Structure and function subcellular Biochemistry*. Editors J. R. Harris and J. Marles-Wrigth (New York, United States: Springer international publishing), 93, 83–141. doi:10.1007/978-3-030-28151-9 4
- Lang, S., Pfeffer, S., Lee, P.-H., Cavalié, A., Helms, V., Förster, F., et al. (2017). An update on Sec61 channel functions, mechanisms, and related diseases. *Front. Physiol.* 8, 887. doi:10.3389/fphys.2017.00887
- Levy, J. M., Towers, C. G., and Thorburn, A. (2017). Targeting autophagy in cancer. Nat. Rev. Cancer 17, 528–542. doi:10.1038/nrc.2017.53
- Li, W. T., Zou, A. E., Honda, C. O., Zheng, H., Wang, X. Q., Kisseleva, T., et al. (2019). Etiology-specific analysis of hepatocellular carcinoma transcriptome reveals genetic dysregulation in pathways implicated in immunotherapy efficacy. *Cancers* 11, 1273. doi:10.3390/cancers11091273
- Li, X., Sun, S., Appathurai, S., Sundaram, A., Plumb, R., and Mariappan, M. (2020). A molecular mechanism for turning off IRE1a signaling during endoplasmic reticulum stress. *Cell Rep.* 33, 108563. doi:10.1016/j.celrep.2020. 108563
- Lindner, P., Christensen, S. B., Nissen, P., Moller, J. V., and Engedal, N. (2020). Cell death induced by the ER stressor thapsigargin involves death receptor 5, a non-autophagic function of MAP1LC3B, and distinct contributions from unfolded protein response components. *Cell Commun. Signal.* 18, 12. doi:10.1186/s. 12964-019-0499-z
- Linxweiler, J., Kollipara, L., Zahedi, R., Lampel, P., Zimmermann, R., and Greiner, M. (2014). Proteomic insights into non-small cell lung cancer: New ideas for cancer diagnosis and therapy from a functional viewpoint. *EuPA Open Proteom.* 4, 25–39. doi:10.1016/j.euprot.2014.05.004
- Linxweiler, M., Bochen, F., Schick, B., Wemmert, S., Al Kadah, B., Greiner, M., et al. (2016). Identification of SEC62 as a potential marker for 3q amplification and cellular migration in dysplastic cervical lesions. *BMC Cancer* 16, 676. doi:10.1186/s12885-016-2739-6
- Linxweiler, M., Linxweiler, J., Barth, M., Benedix, J., Jung, V., Kim, Y.-J., et al. (2012). Sec62 bridges the gap from 3q amplification to molecular cell biology in Non-Small Cell Lung Cancer. *Am. J. Pathol.* 180, 473–483. doi:10.1016/j.ajpath. 2011.10.039
- Linxweiler, M., Schick, B., and Zimmermann, R. (2017). Let's talk about Secs: Sec61, Sec62 and Sec63 in signal transduction, oncology and personalized medicine. Signal Transduct. Target. Ther. 2, e17002. doi:10.1038/sigtrans. 2017.2
- Linxweiler, M., Schorr, S., Jung, M., Schäuble, N., Linxweiler, J., Langer, F., et al. (2013). Targeting cell migration and the endoplasmic reticulum stress response with calmodulin antagonists: A clinically tested small molecule phenocopy of SEC62 gene silencing in human tumor cells. *BMC Cancer* 13, 574. doi:10.1186/1471-2407-13-574
- Liu, B., Liu, J., Liao, Y., Jin, C., Zhang, Z., Zhao, J., et al. (2019). Identification of SEC61G as a novel prognostic marker for predicting survival and response to

- therapies in patients with glioblastoma. *Med. Sci. Monit.* 25, 3624–3635. doi:10.12659/MSM.916648
- Liu, X., Su, K., Sun, X., Jiang, Y., Wang, L., Hu, C., et al. (2021). Sec62 promotes stemness and chemoresistance of human colorectal cancer through activating Wnt/ β -catenin pathway. *J. Exp. Clin. Cancer Res.* 40, 132. doi:10.1186/s13046-021-01934-6
- Loi, M., Raimondi, A., Morone, D., and Molinari, M. (2019). ESCRT-III-driven piecemeal micro-ER-phagy remodels the ER during recovery from ER stress. *Nat. Commun.* 10, 5058. doi:10.1038/s41467-019-12991-z
- Lu, Z., Zhou, L., Killela, P., Rasheed, A. B., Di, C., Poe, W. E., et al. (2009). Glioblastoma proto-oncogene SEC61gamma is required for tumor cell survival and response to endoplasmic reticulum stress. *Cancer Res.* 69, 9105–9111. doi:10.1158/0008-5472.CAN-09-2775
- Mahalingam, D., Wilding, G., Denmeade, S., Saratopoulas, J., Cosgrove, D., Cetnar, J., et al. (2016). Mipsagargin, a novel thapsigargin-based PSMA-activated prodrug: Results of a first-in-man phase I clinical trial in patients with refractory, advanced or metastatic solid tumours. *Br. J. Cancer* 114, 986–994. doi:10.1038/bjc.2016.72
- Marques, L. O., Lima, M. S., and Soares, B. G. (2004). Trifluoperazine for schizophrenia. *Cochrane Database Syst. Rev.* 2004, CD003545. doi:10.1002/14651858.CD003545.pub2
- Mayer, H.-A., Grau, H., Kraft, R., Prehn, S., Kalies, K.-U., Hartmann, E., et al. (2000). Mammalian Sec61 is associated with Sec62 and Sec63. *J. Biol. Chem.* 275, 14550–14557. doi:10.1074/jbc.275.19.14550
- Melnyk, A., Lang, S., Sicking, M., Zimmermann, R., and Jung, M. (2022). "Co-chaperones of the human endoplasmic reticulum: An update," in *The networking of chaperones by Co-chaperones subcellular Biochemistry*. Editors A. Edkins and G. L. Blatch (New York, United States: Springer international publishing), 101.
- Meng, H., Jiang, X., Wang, J., Sang, Z., Guo, L., Yin, G., et al. (2021a). SEC61G is upregulated and required for tumor progression in human kidney cancer. *Mol. Med. Rep.* 23, 427. doi:10.3892/mmr.2021.12066
- Meng, Y., Zhao, H., Zhao, Z., Yin, Z., and Du, J. (2021b). Sec62 promotes proangiogenesis of hepatocellular carcinoma cells under hypoxia. *Cell biochem. Biophys.* 79, 747–755. doi:10.1007/s12013-021-01008-6
- Molinari, M. (2020). ER-phagy: Eating the factory. $Mol.\ Cell\ 78, 811-813.\ doi:10.\ 1016/J.molcel.2020.05.002$
- Mori, Y., Sato, F., Selaru, F. M., Olaru, A., Perry, K., Kimos, M. C., et al. (2002). Instabilotyping reveals unique mutational spectra in microsatellite-unstable gastric cancers. *Cancer Res.* 62, 3641–3645.
- Müller, C. S. L., Kreie, L., Bochen, F., Pfuhl, T., Smola, S., Gräber, S., et al. (2019). Expression of 3q oncogene SEC62 in atypical fibroxanthoma immunohistochemical analysis of 41 cases and correlation with clinical, viral and histopathologic features. *Oncol. Lett.* 17, 1768–1776. doi:10.3892/ol.2018.9767
- Müller, C. S. L., Pföhler, C., Wahl, M., Bochen, F., Körner, S., Kühn, J. P., et al. (2021). Expression of SEC62 oncogene in benign, malignant and borderline melacytic tumors: Unmasking the wolf in sheep's clothing. *Cancers* 13, 1645. doi:10.3390/cancers13071645
- Müller, L., Diaz de Escauriaza, M., Lajoie, P., Theis, M., Jung, M., Müller, A., et al. (2010). Evolutionary gain of function for the ER membrane protein Sec62 from yeast to humans. *Mol. Biol. Cell* 21, 691–703. doi:10.1091/mbc.e09-08-0730
- Müller, L., Funato, Y., Miki, H., and Zimmermann, R. (2011). An interaction between human Sec63 with nucleoredoxin may provide the missing link between the SEC63 gene and polycystic liver disease. FEBS Lett. 585, 596–600. doi:10.1016/j. febslet.2011.01.024
- Nelson, W. J., and Nusse, R. (2004). Convergence of Wnt, beta-catenin, and cadherin pathways. *Science* 303, 1483–1487. doi:10.1126/science.1094291
- Nusse, R., and Clevers, H. (2017). Wnt/ β -Catenin signaling, disease, and emerging therapeutic modalities. *Cell* 169, 985–999. doi:10.1016/j.cell.2017.05.016
- Pfeffer, S., Burbaum, L., Unverdorben, P., Pech, M., Chen, Y., Zimmermann, R., et al. (2015). Structure of the native Sec61 protein-conducting channel. *Nat. Commun.* 6, 8403. doi:10.1038/ncomms9403
- Pisoni, G., and Molinari, M. (2016). Five questions (with their answers) on ER-associated degradation. *Traffic* 17, 341–350. doi:10.1111/tra.12373
- Polischouk, A. G., Holgersson, A., Zong, D., Stenerlöw, B., Karlsson, H. L., Möller, L., et al. (2007). The antipsychotic drug trifluoperazine inhibits DNA repair and sensitizes non small cell lung carcinoma cells to DNA doublestrand break induced cell death. *Mol. Cancer Ther.* 6, 2303–2309. doi:10.1158/1535-7163.MCT-06-0402
- Pulkoski-Gross, A., Li, J., Zheng, C., Li, Y., Ouyang, N., Rigas, B., et al. (2015). Repurposing the antipsychotic trifluoperazine as an antimetastasis agent. *Mol. Pharmacol.* 87, 501–512. doi:10.1124/mol.114.096941

Radosa, J. C., Kasoha, M., Schilz, A.-C., Linxweiler, M., Linxweiler, B., Takacs, Z. F., et al. (2022). PKC inhibits Sec61 translocon-mediated sarcoplasmic reticulum Ca²⁺ leak in smooth muscle cells. *Front. Physiol.* 13, 925023. doi:10.3389/fphys. 2022.925023

Reis-Filho, J. S., Pinheiro, C., Lambros, M. B. K., Milanezi, F., Carvalho, S., Savage, K., et al. (2006). EGFR amplification and lack of activating mutations in metaplastic breast carcinomas. *J. Pathol.* 209, 445–453. doi:10.1002/path.2004

Sangodkar, J., Dhawan, N. S., Melville, H., Sigh, V. J., Yuan, E., Rana, H., et al. (2012). Targeting the FOXO1/KLF6 axis regulates EGFR signaling and treatment response. *J. Clin. Invest.* 122, 2637–2651. doi:10.1172/JCI62058

Schäuble, N., Lang, S., Jung, M., Cappel, S., Schorr, S., Ulucan, Ö., et al. (2012). BiP-mediated closing of the Sec61 channel limits Ca²⁺ leakage from the ER. *EMBO J.* 31, 3282–3296. doi:10.1038/emboj.2012.189

Schorr, S., Klein, M.-C., Gamayun, I., Melnyk, A., Jung, M., Schäuble, N., et al. (2015). Co-chaperone specificity in gating of the polypeptide conducting channel in the membrane of the human endoplasmic reticulum. *J. Biol. Chem.* 290, 18621–18635. doi:10.1074/jbc.M115.636639

Schorr, S., Nguyen, D., Haßdenteufel, S., Nagaraj, N., Cavalié, A., Greiner, M., et al. (2020). Identification of signal peptide features for substrate specificity in human Sec62/Sec63-dependent ER protein import. *FEBS J.* 287, 4612–4640. doi:10. 1111/febs.15274

Schulmann, K., Brasch, F. E., Kunstmann, E., Engel, C., Pagenstecher, C., Vogelsang, H., et al. (2005). HNPCC-Associated small bowel cancer: Clinical and molecular characteristics. *Gastroenterology* 128, 590–599. doi:10.1053/j.gastro.2004.12.051

Schwarzbaum, P. J., Schachter, J., and Bredeston, L. M. (2022). The broad range di- and tri-nucleotide exchanger SLC35B1 displays asymmetrical affinities for ATP transport across the ER membrane. *J. Biol. Chem.* 298, 101537. doi:10.1016/j.jbc. 2021.101537

Sicking, M., Lang, S., Bochen, F., Drenth, J. P. H., Zacharia, M., Zimmermann, R., et al. (2021). Complexity and specificity of Sec61 channelopathies: Human diseases affecting gating of the Sec61 complex. *Cells* 10, 1036. doi:10.3390/cells10051036

Su, S., Shi, Y.-T., Chu, Y., Jiang, M.-Z., Wu, N., Xu, B., et al. (2022). Sec62 promotes gastric cancer metastasis through mediating UPR-induced autophagy activation. *Cell. Mol. Life Sci.* 79, 133. doi:10.1007/s00018-022-04143-2

Sullivan, G. F., Garcia-Welch, A., White, E., Lutzker, S., and Hait, W. N. (2002). Augmentation of apoptosis by the combination of bleomycin with trifluoperazine in the presence of mutant p53. *J. Exp. Ther. Oncol.* 2, 19–26. doi:10.1046/j.1359-4117. 2002.01002.x

Sundaram, A., Plumb, R., Appathurai, S., and Mariappan, M. (2017). The Sec61 translocon limits IRE1a signaling during the unfolded protein response. *eLife* 6, e27187. doi:10.7554/eLife.27187

Takacs, F. Z., Radosa, J. C., Bochen, F., Juhasz-Böss, I., Solomayer, E.-F., Bohle, R. M., et al. (2019c). Sec62/Ki67 and p16/ki67 dual-staining immunocytochemistry in vulvar cytology for the identification of vulvar intraepithelial neoplasia and vulvar cancer: A pilot study. *Arch. Gynecol. Obstet.* 299, 825–833. doi:10.1007/s00404-018-5021-0

Takacs, F. Z., Radosa, J. C., Bohle, R. M., Bochen, F., Juhasz-Böss, I., Solomayer, E.-F., et al. (2019b). Sec62/Ki67 dual staining in cervical cytology specimens: A new marker for high-grade dysplasia. *Arch. Gynecol. Obstet.* 299, 481–488. doi:10.1007/s00404-018-4981-4

Takacs, F. Z., Radosa, J. C., Linxweiler, M., Kasohah, M., Bohle, R. M., Bochen, F., et al. (2019a). Identification of 3q oncogene SEC62 as a marker for distant metastasis and poor clinical outcome in invasive ductal breast cancer. *Arch. Gynecol. Obstet.* 299, 1405–1413. doi:10.1007/s00404-019-05081-4

Treiman, M., Caspersen, C., and Christensen, S. B. (1998). A tool coming of age: Thapsigargin as an inhibitor of sarco-endoplasmic reticulum Ca(2+)-ATPases. *Trends Pharmacol. Sci.* 19, 131–135. doi:10.1016/s0165-6147(98)01184-5

Tsai, F.-C., Kuo, G.-H., Chang, S.-W., and Tsai, P.-J. (2015). Ca2+ signaling in cytoskeletal reorganization, cell migration, and cancer metastasis. *Biomed. Res. Int.* 2015, 409245. doi:10.1155/2015.409245

Tsukamoto, Y., Uchida, T., Karnan, S., Noguchi, T., Nguyen, L. T., Tanigawa, M., et al. (2008). Genome-wide analysis of DNA copy number alterations and

gene expression in gastric cancer. J. Pathol. 216, 471–482. doi:10.1002/path.

Tyedmers, J., Lerner, M., Bies, C., Dudek, J., Skowronek, M. H., Haas, I. G., et al. (2000). Homologs of the yeast Sec complex subunits Sec62p and Sec63p are abundant proteins in dog pancreas microsomes. *Proc. Natl. Acad. Sci. U. S. A.* 97, 7214–7219. doi:10.1073/pnas.97.13.7214

Tyedmers, J., Lerner, M., Nastainczyk, W., and Zimmermann, R. (2005). Calumenin and reticulocalbin are associated with the protein translocase of the mammalian endoplasmic reticulum. *J. Biol. Sci.* 5, 70–75. doi:10.3923/jbs.2005. 70.75

Tyedmers, J., Lerner, M., Wiedmann, M., Volkmer, J., and Zimmermann, R. (2003). Polypeptide-binding proteins mediate completion of co-translational protein translocation into the mammalian endoplasmic reticulum. *EMBO Rep.* 4, 505–510. doi:10.1038/sj.embor.embor826

Vandonselaar, M., Hickie, R. A., Quail, J. W., and Delbaere, L. T. (1994). Trifluoperazine-induced conformational change in Ca(2+)-calmodulin. *Nat. Struct. Biol.* 1, 795–801. doi:10.1038/nsb1194-795

Walter, P., and Ron, D. (2011). The unfolded protein response: From stress pathway to homeostatic regulation. *Science* 334, 1081–1086. doi:10.1126/science.

Weitzmann, A., Baldes, C., Dudek, J., and Zimmermann, R. (2007). The hsp70-molecular chaperone network in the pancreatic endoplasmic reticulum: A quantitative approach. *FEBS J.* 274, 5175–5187. doi:10.1111/j.1742-4658-2007-06020

Weitzmann, A., Volkmer, J., and Zimmermann, R. (2006). The nucleotide exchange factor activity of Grp 170 may explain the non-lethal phenotype of loss of Sil1 function in man and mouse. *FEBS Lett.* 580, 5237–5240. doi:10.1016/j. febslet.2006.08.055

Wemmert, S., Lindner, Y., Linxweiler, J., Wagenpfeil, S., Bohle, R., Niewald, M., et al. (2016). Initial evidence for Sec62 as a prognostic marker in advanced head and neck squamous cell carcinoma. *Oncol. Lett.* 11, 1661–1670. doi:10.3892/ol.2016. 4135

Weng, L., Du, J., Zhou, Q., Cheng, B., Li, J., Zhang, D., et al. (2012). Identification of cyclin B1 and Sec62 as biomarkers for recurrence in patients with HBV-related hepatocellular carcinoma after surgical resection. *Mol. Cancer* 11, 39. doi:10.1186/1476-4598-11-39

Wirth, A., Jung, M., Bies, C., Frien, M., Tyedmers, J., Zimmermann, R., et al. (2003). The Sec61p complex is a dynamic precursor activated channel. *Mol. Cell* 12, 261–268. doi:10.1016/s1097-2765(03)00283-1

Witham, C. M., Paxman, A. L., Baklous, L., Steuart, R. F. L., Schulz, B. L., and Mousley, C. J. (2021). Cancer associated mutations in Sec61 γ alter the permeability of the ER translocase. *PLoS Genet.* 17, e1009780. doi:10.1371/journal.pgen.1009780

Xu, S., Li, X., Geng, J., Cao, Y., Yu, Y., and Qi, L. (2022). Sec 61γ is a vital protein in the endoplasmic reticulum membrane promoting tumor metastasis and invasion in lung adenocarcinoma. *Brit. J. Cancer.* in Press.

Yeh, C. T., Wu, A. T., Chang, P. M., Chen, K.-Y., Yang, C.-N., Yang, S.-C., et al. (2012). Trifluoperazine, an antipsychotic agent, inhibits cancer stem cell growth and overcomes drug resistance of lung cancer. *Am. J. Respir. Crit. Care Med.* 186, 1180–1188. doi:10.1164/rccm.201207-1180OC

Yong, J., Bischof, H., Burgstaller, S., Siirin, M., Murphy, A., Malli, R., et al. (2019). Mitochondria supply ATP to the ER through a mechanism antagonized by cytosolic Ca^{2+} . eLife 8, e49682. doi:10.7554/eLife.49682

Zacharski, L. R., Moritz, T. E., Haakenson, C. M., O'Doonell, J. F., Ballard, H. S., Johnson, G. J., et al. (1990). Chronic calcium antagonist use in carcinoma of the lung and colon: A retrospective cohort observational study. *Cancer Invest.* 8, 451–458. doi:10.3109/07357909009012067

Zimmermann, R., and Lang, S. (2020). A little AXER ABC: ATP, BiP, and calcium form a triumvirate orchestrating energy homeostasis of the endoplasmic reticulum. *Contact* 3, 251525642092679. doi:10.1177/2515256420926795

Ziska, A., Tatzelt, J., Dudek, J., Paton, A. W., Paton, J. C., Zimmermann, R., et al. (2019). The signal peptide plus a cluster of positive charges in prion protein dictate chaperone-mediated Sec61-channel gating. *Biol. Open* 8, bio040691. doi:10.1242/bio.040691