#### **RESEARCH ARTICLE**



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# Genetic diversity of Gymnocypris chilianensis (Cypriniformes, Cyprinidae) unveiled by the mitochondrial DNA D-loop region

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#### ABSTRACT

In order to analyze the genetic diversity and genetic differentiation of Gymnocypris chilianensis, D-loop region of the mitochondrial DNA was sequenced in 50 individuals of G. chilianensis obtained from 2 geographic locations (Heihe River and Shule River) and 25 individuals of G. przewalskii (Qinghai Lake). Twenty-five homologous sequences of another G. eckloni (Yellow River) downloaded from GenBank were analyzed together. The sequences were analyzed by using the MEGA (version 7.0) and DnaSP (version 6.0) software. The results revealed that 82 haplotypes were detected among 100 individuals. The haplotype diversity (Hd) and nucleotide diversity (Pi) of G. chilianensis of the Shule River were  $0.963 \pm 0.029$  and  $0.00414 \pm 0.00069$ , which were lower than those of 3 other populations. The genetic distance of G. chilianensis in both Heihe River and Shule River was 0.0013. The genetic distances between the 2 G. chilianensis populations and the G. eckloni were 0.0148 and 0.0141, respectively. Population differentiation values (Fst) and gene flow (Nm) showed that 4 population had occurred obvious genetic differentiation (Fst: 0.20811  $\sim$  0.98863. p < 0.01; Nm < 1). Compared with G. przewalskii and G. eckloni, the differentiation degree was more significant between Heihe River G. chilianensis and Shule River G. chilianensis (Fst = 0.98863, p < 0.01; Nm = 0.00287). Maximum Likelihood (ML) phylogenetic tree showed that G. chilianensis had further genetic distance with G. eckloni and G. przewalskii. In conclution, G. chilianensis (HH&SL) had lower genetic diversity and further genetic distance than G. przewalskii (QH) and G. eckloni (YL). We suggest strengthen the protection of genetic resources of G. chilianensis.

#### Introduction

Gymnocypris chilianensis is a species of the genus Gymnocypris which belongs to the subfamily Schizothoracinae of the family Cyprinidae. It is mainly distributed in the Shiyang River, Heihe River and Shule Rivers, inland river basin of Hexi Corridor in Gansu province of China (Li and Chang 1974; Wu and Wu 1992). The Hexi Corridor is located at the border of the Tibetan Plateau, the Loess Plateau and the Inner Mongolian Plateau. The three inland rivers in the Hexi Corridor originate from the Qilian Mountains. Water resources for supply of three rivers mainly depend on snow and rain water from the Qilian Mountains (Zhang et al. 2015). Gymnocypris chilianensis has delicious meat, rich nutritional value, and high economic value. In recent years, the distribution area and the wild resources of G. chilianensis are decreasing dramatically, which is mainly influenced by natural and artificial factors. As a result, its genetic diversity is diminishing progressively. The adaptability of species is positively correlated with genetic diversity. Lack of genetic diversity poses a huge threat to species living in anthropogenically disturbed habitats (Xiao et al. 2013). To protect and utilize the resources of G. chilianensis, it is necessary to investigate the genetic diversity and population structure of G. chilianensis.

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At present, there are few studies on the genetic diversity and population structure of G. chilianensis. Zhao et al. (2011) used mitochondrial DNA sequence data (a control region and the cytochrome b gene; 1894 bp) to assess the phylogeographic structure of G. chilianensis in the inland river watersheds of the Hexi Corridor in Gansu Province, China. Moreover, we have studied the genetic diversity and taxonomic status of G. chilianensis based on the mitochondrial DNA cytochrome b gene (Zhang et al. 2015). The results showed that both the haplotype diversity and nucleotide diversity were lower than the other populations. Analysis of sequence differences indicates that G. chilianensis is sufficiently diverged from G. przewalskii and G. eckloni to the extent that it has reached species level, it provides a reference basis for G. chilianensis to be regarded as an independent species of Gymnocypris. As a non-coding fragment of mtDNA, the D-loop region is not subject to the pressure of coding selection, and has long been considered to have the fastest mutation rate in the mitochondrial genome. Compared with cytochrome b gene of mtDNA, it is more suitable for the genetic diversity, population genetic structure and phylogeographic analysis (Cann et al. 1984).

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Dai et al. (2010) have studied the genetic diversity of Wujiang River population of *Schizothorax kozlovi* by using mtDNA *D-loop* region. Its genetic diversity was found to very low and the protection for the population is necessary. Zhang et al. (2013) used the *D-loop* region to study the four populations of *Gymnocypris przewalskii* (Lake Qinghai, Lake Keluke, Ganzi River and Cao Dalian). It was concluded that *Gymnocypris przewalskii* has high genetic diversity, and there is a certain degree of genetic differentiation among populations, especially the Keruke Lake population has been highly differentiated, but the level of genetic diversity is very low, it should be priority to protect.

In this study, we analyzed the genetic diversity and population structure by focusing on *G. chilianensis* and using the mtDNA D-loop gene as a marker, with the aim of providing a theoretical basis for resource conservation and utilization of *G. chilianensis*.

# **Materials and methods**

# Study area

A total of 50 individuals of *G. chilianensis* were collected, among which 25 individuals each were obtained from Heihe (HH)(N100°30'; E38°57') and Shule (SL) (N96°13'; E42°21') rivers. Besides, 25 individuals of *G. przewalskii* were collected from Qinghai Lake (QH) (N99°38'; E36°32').

# Sample collection

All individuals were identified based on morphological characteristics (Li and Chang 1974; Zhao 2005). A part of caudal fin was collected from each individual, preserved in 95% ethanol and stored at -20 °C in the Fisheries Laboratory of Gansu Agricultural University. Genomic DNA was extracted by proteinase K digestion followed by a standard phenolchloroform method (Green and Sambrook 2012). 25 *D-loop* region sequences of *G. eckloni* (YL) which were derived from the mainstream of the Yellow River in Qinghai Province of China, were downloaded from the GenBank database (GenBank accession No. FJ601074-FJ601098)

# Amplification, cloning and sequencing

*D-loop* sequences were amplified with the new specific primers: F5'-GGGATATGTCATCCTTTATGG-3', R5'-GGGTTTGACAAGAATAA CAGG-3'. The PCR reaction was performed in a total volume of 50  $\mu$ L containing 50 ng of template DNA, 100  $\mu$ M of dNTPs (Takara, Dalian, China), 2.5 mM of 1 × PCR reaction buffer, 1 U of Taq DNA polymerase (Takara), 0.4  $\mu$ M of each primer and 19.3  $\mu$ L of ddH<sub>2</sub>O. PCR was carried out under the following conditions: an initial denaturation at 95 °C for 3 min, followed by 30 cycles of denaturation at 72 °C for 90 s, followed by a final extension at 72 °C for 10 min.

Subsequently, PCR products were analyzed using 1% agarose gel and purified using the Universal DNA Purification Kit (TianGen, Beijing, China). Purified PCR products were subsequently cloned into pMD-19T Simple Vector (TaKaRa) and the ligation products transformed into DH5 $\alpha$  Chemically Competent Cell *Escherichia coli* (TianGen) according to the manufacturer's instructions. Colonies of *E. coli* harboring recombinant clones were screened for the presence of inserts of the expected size by LacZ blue-white selection and identified by double digestion with *Eco*RI and *Pst*I. The positive recombinant clones were sequenced from both directions (Sangon Biotech, Shanghai, China).

# Data analysis

The *D-loop* sequences were separately aligned and trimmed to equal lengths using the BioEdit program (http://www.mbio. ncsu.edu/bioedit/bioedit.html) and ClustalX 2.1 program (http:// www.clustal.org/). The nucleotide composition and genetic distance among four populations (the average genetic distances based on Kimura's two-parameter model) were computed using MEGA 7.0 software (http://www.megasoftware.net/), with the standard errors estimated by bootstrapping using 1000 replicates (Kimura 1980). Genetic diversity parameters were estimated using DnaSP 6 software (http://www.ub.edu/dnasp/). For phylogenetic analysis, Gymnodiptychus pachycheilus (GenBank accession No. of D-loop: FJ601172) and Ptychobarbus dipogon (GenBank accession No. of D-loop: FJ601178) were taken as the out-group, Maximum Likelihood (ML) phylogenetic tree based on *D-loop* haplotypes was constructed using MEGA 7.0, and statistical support was estimated using 1000 bootstrap replicates. Haplotype network (median-joining) was generated using Network 5 software (http://www.fluxus-engineering.com/) (Liao et al. 2016; Gao et al. 2017; Nisar et al. 2019).

# Results

#### **Base composition**

A total of 100 homologous sequences of 747 bp were used for the base composition analysis. The average base composition was T = 32.1%, C = 21.9%, A = 31.9%, and G = 15.1%. The base compositions of *G. chilianensis*, *G. eckloni* and *G. przewalskii* were very similar in the *D-loop* region, indicating that the base composition did not change with the variation. Average Homoplasy of 4 population compared with each other was 98.1%.

# Gene mutation

A total of 100 homologous sequences of 747 bp were used for the genetic analysis of gene mutations, and a total of 124 mutation sites were detected, accounting for 16.4% of the total analyzed sites, including 119 transitions and 9 transversions. The transition/transversion ratio was 13.2 and significantly greater than 2.0. Therefore, the mutagenesis was not needed saturated and weighted analysis in the phylogenetic analysis (Knight and Mindell 1993).

Table 1. Genetic diversity indices of the four Gymnocypris populations studied.

Population	Sample size	Нр	V (%)	S	Hd (SD)	Pi (SD)
НН	25	20	80	36	0.973 (0.022)	0.00626 (0.00072)
SL	25	20	80	35	0.963 (0.029)	0.00414 (0.00069)
QH	25	24	96	55	0.997 (0.012)	0.00831 (0.00132)
YL	25	18	72	22	0.943 (0.037)	0.00745 (0.00050)

Note: HH = Heihe River *G. chilianensis*; SL = Shule River *G. chilianensis*; YL = Yellow River *G. eckloni* and QH = Qinghai Lake *G. przewalskii.* S: number of polymorphic (segregating) sites; Hd: haplotype (gene) diversity; Pi: nucleotide diversity; SD: standard deviation.

#### **Genetic diversity**

A total of 82 haplotypes were recovered from 100 aligned sequences and the haplotype diversity and nucleotide diversity indices were shown in Table 1. The haplotype diversity of the Qinghai Lake *G. przewalskii* population was the highest (Hd = 0.997), compared with the lowest haplotype diversity of the Yellow River *G. eckloni* (Hd = 0.943). The nucleotide diversity of the Qinghai Lake *G. przewalskii* was the highest (Pi = 0.00831), compared with the lowest nucleotide diversity of the Shule River *G. chilianensis* (Pi = 0.0014).

#### **Genetic differentiation**

Genetic distance within and among populations was estimated using the Kimura's two-parameter model, with values ranging from 0.0042 to 0.0084 and from 0.0013 to 0.0148, respectively (Table 2), suggesting that the genetic distances among populations were higher than within populations. The genetic distance between the Heihe River *G. chilianensis* population and the *G. eckloni* population were found to be the highest (0.0148), followed by the genetic distance between Heihe River *G. chilianensis* population and the *G. przewalskii* population (0.0143), while the genetic distance between the Heihe River *G. chilianensis* population and the Shule River *G. chilianensis* population were the lowest (0.0013).

DNASP 6 software was used to calculate the *Fst* and *Nm* among the 4 populations, and the results were shown in Table 3. *Fst* (*Fst* > 0.25) and *Nm* (*Nm* < 1) of the four populations reached the level of high differentiation, with the highest *Fst* and the lowest *Nm* between Heihe River *G. chilianensis* and Shule River *G. chilianensis*, the lowest *Fst* and the highest *Nm* between Qinghai Lake *G. przewalskii* and Yellow River *G. eckloni*.

#### Matrilineal genealogical analysis

The matrilineal genealogical analysis showed that 82 haplotypes diverged into two main branches. One main branch contains the Heihe River *G. chilianensis*. Another branch contains Shule River *G. chilianensis*, *G. przewalskii* and *G. eckloni* which the Shule River *G. chilianensis* from a haplotype clade, bootstrap values were 84%, *G. przewalskii* and *G. eckloni* from another haplotype clade, bootstrap values were 73% (Figure 1).

# Haplotype network analysis

To understand the relationships of haplotypes, the median joining network was constructed for the identified haplotypes. All the haplotypes were clustered into two big groups

**Table 2.** Genetic distance among the four *Gymnocypris* populations (below diagonal) and genetic distance within populations (in bold, above diagonal).

YL	0.0148 (0.0062)	0.0141 (0.0061)	0.0013 (0.0007)	0.0075 (0.0019)
QH	0.0143 (0.0060)	0.0137 (0.0059)	0.0084 (0.0015)	
SL	0.0013 (0.0012)	0.0042 (0.0007)		
нн	0.0063 (0.0015)			
Group	HH (SD)	SL (SD)	QH (SD)	YL (SD)

Note: HH = Heihe River G. chilianensis; SL = Shule River G. chilianensis; YL = Yellow River G. eckloni and QH = Qinghai Lake G. przewalskii. SD: standard deviation.

**Table 3.** *Fst* (below diagonal) and *Nm* (above diagonal) among the 4 *Gymnocypris* populations.

Group	НН	SL	QH	YL
НН		0.00287	0.00369	0.00342
SL	0.98863 (p < 0.01)		0.07568	0.06762
QH	0.98545 ( <i>p</i> < 0.01)	0.76761 ( <i>p</i> < 0.01)		0.95128
YL	0.98648 ( <i>p</i> < 0.01)	0.78710 ( <i>p</i> < 0.01)	0.20811 (p < 0.01)	

Note: HH = Heihe River *G. chilianensis*; SL = Shule River *G. chilianensis*; YL = Yellow River *G. eckloni* and QH = Qinghai Lake *G. przewalskii*.

(haplogroups a and b). *G. chilianensis* (HH&SL) clustered on haplogroups a, *G. eckloni* (YL) and *G. przewalskii* (QH) clustered on haplogroups b and some haplotype cross distribution each other (Figure 2).

## Discussion

Haplotype diversity (Hd) and nucleotide diversity (Pi) are important indicators of population genetic variation. They are a valuable indicator for estimating the abundance or scarcity of the genetic diversity of a population (Li et al. 2011). In this study, the *D-loop* region sequence analysis showed that the haplotype diversity (Hd =0.9970) and nucleotide diversity (Pi = 0.00831) of G. przewalskii population were the highest among the four groups. The nucleotide diversity index of the Shule River G. chilianensis population was 0.00414, which was lower than the other populations analyzed in our study, but still well above the research results on the Shule River G. chilianensis (Pi = 0.0002) by Zhao et al. (2011), and much higher than the result of research on the Spinibarbus yunnanensis (Pi = 0.0014) by Li and Lu (1998). According to the range of Pi of genetic diversity proposed by Lan and Shi (1993), the genetic diversity of the Shule River G. chilianensis population was lower (Pi < 0.0047). Moreover, the combination of high haplotype diversity (Hd = 0.9630) with low nucleotide diversity (Pi = 0.00414) of the Shule River G. chilianensis population reveals possible mechanisms for the formation of this population, which multiply rapidly from a small effective fish population (Avise 2000). Our result was similar to the observed in Anabarilius grahami by Yang et al. (2008).

The genetic distance based on the *D-loop* region indicates that the genetic distances among populations were higher than within populations. Due to the lack of clear genetic distance criteria, which based on *D-loop* region, we cannot measure whether it reached the species differentiation level between individuals. In present study, the genetic distance based on the *D-loop* region sequence can only show the relationship between *G. chilianensis* and *G. eckloni* is distant, and the relationship between *G. chilianensis* and *G. przewalskii* is distant. The present result is accorded with the viewpoint of



Figure 1. Maximum Likelihood (ML) phylogenetic tree of the 78 D-loop haplotypes from the Gymnocypris populations. Note: HH = Heihe River G. chilianensis, SL = Shule River G. chilianensis, YL = Yellow River G. eckloni, QH = Qinghai Lake G. przewalskii, OG01 = Yellow River Gymnodiptychus pachycheilus, OG02 = Yellow River Ptychobarbus dipogon.

Zhang et al. (2015), which means that the genetic relationship between G. chilianensis and G. przewalskii and G. eckloni is distant. Fst is an important indicator of genetic differentiation among populations. 0 < Fst < 0.05 indicates that there is no differentiation among its subgroups. 0.05< Fst < 0.15 indicates that there is moderate differentiation; 0.15< Fst < 0.25 indicates that there is high differentiation. In this study, we found that the Fst of the four populations reached  $0.20811 \sim 0.98863$  (p < 0.01), which indicated that obvious genetic differentiation had occurred in the four populations. Nm < 1 between populations indicates that the population may be differentiated due to genetic drift. Nm > 1 between populations indicates that the level of gene flow between populations is higher and the genetic differentiation between populations is smaller. When Nm > 4, the gene exchange between populations is more sufficient and the genetic differentiation is more smaller (Wright 1951, 1978). In this study, we found that the Nm values of the four populations were less than 1, indicating that geographic isolation has completely hindered the gene exchange of the four

populations. Compared with Qinghai Lake *G. przewalskii* and Yellow River *G. eckloni*, the genetic distance between Heihe River *G. chilianensis* and Shule River *G. chilianensis* was further and the differentiation degree was more significant.

The Maximum Likelihood (ML) phylogenetic trees showed that the populations of G. chilianensis (HH) clustered on one branch, G. chilianensis (SL) clustered on one branch, G. eckloni (YL) and G. przewalskii (QH) clustered on the other branch, and the bootstrap values of which were 83-99%. G. chilianensis had further genetic distance with G. eckloni and G. przewalskii. G. chilianensis should be recognized as an independent species, which is widely accepted (Wu et al. 1964; Li and Chang 1974; Zhang et al. 2013; Zhang et al. 2015), but Zhao (1991) suggested that G. chilianensis as a subspecies of G. eckloni. The results of this study provide a theoretical reference for their views. The results of this study also suggest the genetic distance among G. przewalskii and G. eckloni was closer (0.0100). The established Maximum Likelihood (ML) phylogenetic tree also showed that G. przewalskii and G. eckloni clustered into one branch and haplotype crossed each other.



**Figure 2.** Median joining network of the haplotypes reconstructed in present study for Gymnocypris fish. Note: A circle represents a haplotype, the blue stand for *G. eckloni* (YL), the green stand *G. przewalskii* (QH), orange and purple stand for *G. chilianensis* (HH&SL). HH = Heihe River *G. chilianensis*, SL = Shule River *G. chilianensis*, YL = Yellow River *G. eckloni* and QH = Qinghai Lake *G. przewalskii* 

Eighty-two haplotypes we observed were clustered into two clades and cross distribution each other (Figure 2). Clustering results in network haplotypes analysis were completely consistent with its clustering results in phylogenetic analysis, clustering results in haplotype network analysis can clearly indicate the relationship between four populations. Two populations of G. chilianensis (HH&SL) have developed genetic differentiation, but there was some cross distribution between haplotypes, indicating that the degree of genetic differentiation is not high. Other haplogroup contain G. eckloni (YL) and G. przewalskii (QH), and some haplotype cross distribution each other. In conclusion, G. przewalskii and G. eckloni might be subspecies of each other, however, further studies are required. The result of this study shows that the G. chilianensis (HH&SL) had lower genetic diversity than G. przewalskii (QH) and G. eckloni (YL). We suggest strengthen the protection of genetic resources of G. chilianensis. According to different groups, effective protection measures should be set up to actively protect the ecological environment in the basin and improve its habitat environment.

#### **Disclosure statement**

The authors declare no conflicts of interest.

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## Data availability statement

The data that support the findings of this study are available in figshare at https://doi.org/10.6084/m9.figshare.13122809.v1

#### References

- Avise JC. 2000. Phylogeography: the history and formation of species. 1st ed. Cambridge (MA): Harvard University Press.
- Cann RL, Brown WM, Wilson AC. 1984. Polymorphic sites and the mechanism of evolution in human mitochondrial DNA. Genetics. 106:479–499.
- Dai Y, Zou X, Xiao H. 2010. Genetic Diversity of the mtDNA *D-loop* in the Population of *Schizothorax kozlovi* from the Wujiang River. Sichuan J Zool. 29:505–509.
- Gao YS, Jia XX, Tang XJ, Fan YF, Lu JX, Huang SH, Tang MJ. 2017. The genetic diversity of chicken breeds from Jiangxi, assessed with BCDO2 and the complete mitochondrial DNA *D-loop* region. Plos One. 12:e0173192.
- Green MR, Sambrook J. 2012. Molecular cloning: a laboratory manual. 4th ed. New York (NY): Cold Spring Harbor Laboratory Press.
- Kimura M. 1980. A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. J Mol Evol. 16:111–120.
- Knight A, Mindell DP. 1993. Substitution bias, weighting of DNA sequence evolution, and the phylogenetic position of Fea's viper. Syst Biol. 42:18–31.
- Lan H, Shi L. 1993. The origin and genetic differentiation of native breeds of pigs in southwest China: an approach from mitochondrial DNA polymorphism. Biochem Genet. 31:51–60.
- Li S, Chang S. 1974. Two new species and one new subspecies of fishes from The Northern Part of Kansu Province, China. Acta Zoologica Sinica. 20:414–419.

- Li S, L G. 1998. Diversity of mitochondrial DNA in the populations of silver carp, bighead carp, grass carp and black carp in the middle- and lower reaches of the Yangtze River. Acta Zoologica Sinica. 44:82–93.
- Li H, Zhu W, Yang N, Song W, Wang J, Xu W, et al. 2011. The genetic variation of the mtDNA *D-loop* region in domestic ducks, Mei ducks and wild ducks. Chin J Anim Vet Sci. 42:1213–1219.
- Liao Y, Mo G, Sun J, Wei F, Liao DJ. 2016. Genetic diversity of Guangxi chicken breeds assessed with microsatellites and the mitochondrial DNA *D-loop* region. Mol Biol Rep. 43:415–425.
- Nisar A, Waheed A, Khan S, Feng X, Shah AH. 2019. Population structure, genetic diversity and phylogenetic analysis of different rural and commercial chickens of Pakistan using complete sequence of mtDNA *D*-*loop*. Mitochondrial DNA A DNA Mapp Seq Anal. 30:273–280.
- Wright S. 1978. Evolution and the genetics of populations. Vol 4. Variability within and among natural populations. Chicago (IL): University of Chicago Press; p. 179–182.
- Wright S. 1951. The genetical structure of populations. Ann Eugen. 15: 323-354.
- Wu X, Cao W, Yi B, Yang G, Luo Y. 1964. Ichthyography of cyprinidae in china. 1st ed. Shanghai (China): Shanghai Science and Technology Press.
- Wu Y, Wu C. 1992. The fishes of the Qinghai-Xizang Plateau. 1st ed. Chengdu (China): Sichuan Science and Technology Press.
- Xiao T, Lu C, Chao L, Zhang M, Gu Y, Sun X. 2013. Genetic diversity and phylogenetic analysis of eight Cyprinoid aquaculture breeds based on mitochondrial Cyt *b* gene. J Fish China. 37:344–350.

- Yang B, Chen XY, Yang JX. 2008. Structure of the mitochondrial DNA control region and population genetic diversity analysis of *Anabarilius* grahami (Regan). Zool Res. 29:379–385.
- Zhang Y, Du Y, Lou Z, Wang T. 2013. Molecular phylogeny of schizothoracinae fishes in Gansu Province based on mitochondrial cytochrome b gene sequences. J Northwest Normal Univ Nat Sci. 49: 91–102.
- Zhang R, Li G, Tang Y, Zhang C, Zhao K. 2013. Genetic diversity and differentiation of mitochondrial DNA *D-loop* region in *Gymnocypris przewalskii*. Chin Agric Sci Bull. 29:71–76.
- Zhang J, Liu Z, Zhang B, Yin X, Wang L, Shi H, Kang Y. 2015. Genetic diversity and taxonomic status of *Gymnocypris chilianensis* based on the mitochondrial DNA cytochrome *b* gene. Genet Mol Res. 14: 9253–9260.
- Zhao K. 2005. Molecular phylogenetics and phylogeography of Schizothoracinae (Teleostei:Cyprinidae) in Lake Qinghai and adjacent drainages. Northwest A&F University.
- Zhao T. 1991. Fish-fauna and zoogeographical division of Hexi-Alashan Region, the Northwest China. Acta Zoologica Sinica. 37:153–167.
- Zhao K, Duan Z, Peng Z, Gan X, Zhang R, He S, Zhao X. 2011. Phylogeography of the endemic *Gymnocypris chilianensis* (Cyprinidae): sequential westward colonization followed by allopatric evolution in response to cyclical Pleistocene glaciations on the Tibetan Plateau. Mol Phylogenet Evol. 59:303–310.