

A new strategy to target regulatory T cells in solid tumors

Audrey Thomas-Schoemann,^{1,2,3,*} Frédéric Batteux² and Jérôme Alexandre^{2,4}

¹Université Paris Descartes; Unité de Formation et de Recherche des Sciences Pharmaceutiques et Biologiques; Paris, France; ²Université Paris Descartes; Sorbonne Paris Cité; Faculté de Médecine; Paris, France; ³Unité Fonctionnelle de Pharmacocinétique et Pharmacochimie; Assistance Publique-Hôpitaux de Paris; Hôpital Cochin; Paris, France; ⁴Service d'Oncologie; Assistance Publique-Hôpitaux de Paris; Groupement des Hôpitaux Paris Centre; Paris, France

Keywords: regulatory T cell, arsenic trioxide, colon cancer, oxidative stress, nitrosative stress, breast cancer

The depletion of regulatory T cells (Tregs) is a promising therapeutic strategy to enhance antitumor immune responses. Our recent findings indicate that low doses of arsenic trioxide can delay tumor growth in murine models of colon and breast cancer by depleting Tregs through oxidative and nitrosative bursts.

An increased number of regulatory T cells (Tregs) has been observed in the blood and tumor tissues of a high proportion of cancer patients, a phenomenon that promotes tumor progression and adversely affects prognosis.¹ Tregs express a series of markers including CD4, CD25 and the transcription factor FOXP3, which enables cells to suppress immune responses. The adverse effects of Tregs on the immune response are underscored by the observation that Treg depletion can enhance the efficacy of immunotherapeutic strategies.² Cyclophosphamide (CPM), a DNA alkylating agent, is used in numerous chemotherapy regimens. Clinical data reveal that although high-dose CPM exert potent immunosuppressive effects, low-dose CPM has an immunostimulatory activity. In this context, it has been shown that low-dose CPM decreases the number of Tregs and hence can enhance the antitumor activity of adoptively transferred T cells as well as of antitumor vaccines.³

Several strategies have been or are being developed to target Tregs.² Two different human anti-CD25 antibodies (basiliximab and daclizumab) and the recombinant interleukin (IL)-2-diphtheria toxin conjugate known as denileukin diftotox are currently under development. Ipilimumab, a monoclonal antibody targeting CTLA-4 (which is required for

Treg function), has provided impressive results in advanced melanoma patients.² Finally, as outlined in a review by Galluzzi et al., several others targeted agents stimulate tumor-specific immune responses, and some of them have been associated with decreased levels of circulating or tumor-infiltrating Tregs.⁴ More specifically, a recent study has revealed that targeted agents that block the vascular endothelial growth factor A (VEGFA)/ VEGF receptor 2 (VEGFR2) axis (e.g., bevacizumab and sunitinib) inhibit Treg proliferation triggered by VEGFA.⁵

Arsenic trioxide (As₂O₃) has been associated with substantial clinical efficacy in the treatment of promyelocytic leukemia patients. In addition, preclinical studies have shown that other hematological cancer and solid tumors are susceptible to As₂O₃. Although the exact mechanisms underlying the antitumor effects of this agent remain unclear, As₂O₃ has been recognized as a powerful inducer of oxidative stress in tumor cells.⁶

We recently demonstrated that low-dose As₂O₃ increases antitumor immune response in colon tumor-bearing mice by modulating Treg abundance.⁷ We first observed that tumor-bearing mice display an increased proportion of Tregs among the splenic CD4⁺ cell population, contributing to immune escape. We showed

that As₂O₃ induces the selective depletion of Tregs both in vitro and in vivo. Indeed, As₂O₃ depleted Tregs in both the spleen and the tumor tissues of mice bearing murine colon carcinoma CT26 cells. Low-dose As₂O₃ was found to exert antitumor effects that are closely related to Treg depletion in both colon (Ct26 cells) and breast (4T1 cells) carcinoma murine models. As₂O₃ turned out to be able to restore the activity of immune cells adoptively transferred from donor mice, thus enhance their antitumor potential. Our results confirmed a previous study that had revealed that As₂O₃ can exacerbate immune responses against breast cancer cells.⁸ In this study, As₂O₃ was shown to increase the cytotoxicity of lymphokine-activated killer (LAK) cells. Tregs were not specifically explored, but they are well known to affect the antitumor activity of multiple effector cells, including LAK cells.

We observed immunostimulatory effects at low As₂O₃ concentrations (0.5–1 μM) in vitro and with a single 1 mg/Kg dose in vivo. In previous studies, As₂O₃ doses ranging from 2 to 6.5 mg/Kg/day for one to six weeks were required to obtain therapeutic effects against solid tumors.⁹ The immunogenic effects of low-dose As₂O₃ appear to be related to the high sensitivity of Tregs to this agent. It has previously been shown

*Correspondence to: Audrey Thomas-Schoemann; Email: schoemann.audrey@gmail.com

Submitted: 12/17/12; Accepted: 12/19/12

Citation: Thomas-Schoemann A, Batteux F, Alexandre J. A new strategy to target regulatory T cells in solid tumors. *Oncoimmunology* 2013; 2:e23338; <http://dx.doi.org/10.4161/onci.23338>

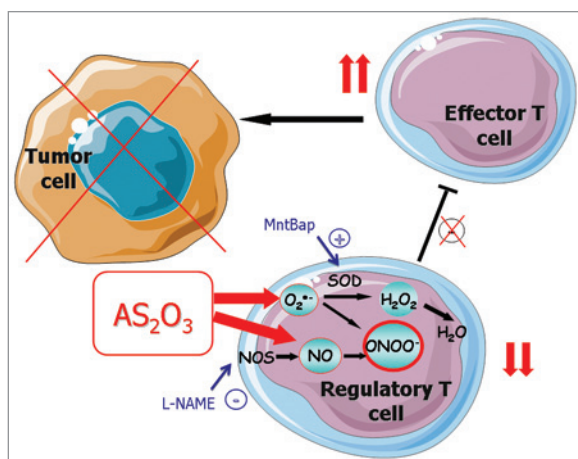


Figure 1. Arsenic trioxide can deplete regulatory T cells through the production of ONOO⁻, thus improving the antitumor activity of effector T cells. Previous studies have shown that arsenic trioxide (As₂O₃) is able to induce the intracellular accumulation of superoxide anion (O₂⁻) and nitrite oxide (NO). Superoxide dismutases (SODs) catalyze the dismutation of O₂⁻ in hydrogen peroxide (H₂O₂), which is converted in H₂O by catalase and glutathione peroxidase. O₂⁻ may react with NO to form peroxynitrite (ONOO⁻). Our data suggest that the depletion of regulatory T cells (Tregs) as induced by As₂O₃ is related to O₂⁻ and NO production, resulting in ONOO⁻ accumulation, as it is limited by the NO synthase (NOS) inhibitor NG-nitro-L-arginine methyl ester (L-NAME) and by the SOD mimic manganese [III] tetrakis-(5,10,15,20)-benzoic acid porphyrin (MnTBAP).

that low-dose CPM similarly decreases the number of Tregs. However, CPM may exert a toxic effect on other lymphocyte populations. Indeed, we observed that CPM induces a splenocyte depletion that could not be observed with As₂O₃. These data suggest that the specificity of As₂O₃ for Tregs may be higher than that of CPM.

Finally, we demonstrated that the depletion of Tregs as induced by As₂O₃ is mediated by the generation of reactive oxygen and nitrogen species. We showed that As₂O₃ promotes the accumulation of hydrogen peroxide (H₂O₂) and/or peroxynitrite (ONOO⁻) in Tregs, as demonstrated via the fluorescent probe H₂DCFDA (2',7'-dichlorodihydrofluorescein diacetate). Our data strongly suggest that Treg depletion as induced by As₂O₃ is related to ONOO⁻ production, since it was inhibited by the nitric oxide (NO) synthase inhibitor NG-nitro-L-arginine methyl ester (L-NAME) and by the superoxide dismutase mimic manganese [III] tetrakis-(5,10,15,20)-benzoic acid porphyrin (MnTBAP) (Fig. 1). Thus, Treg depletion as induced by As₂O₃ is related to superoxide and NO production resulting in the accumulation of ONOO⁻. In contrast, As₂O₃ failed to induce ONOO⁻ accumulation,

in non-Treg CD4⁺ cells. Such differential effects of As₂O₃ on Treg and non-Treg CD4⁺ cells could be related to differences in the redox status of these cells. The threshold for toxicity might be more easily reached in Tregs upon As₂O₃ exposure because of higher basal levels of NO and ONOO⁻ in these cells, resulting in the overwhelming of antioxidant defenses. This redox-based differential effect of As₂O₃ has previously been observed in acute promyelocytic leukemia cells,¹⁰ which would be more susceptible to As₂O₃ cytotoxicity because of their relatively high basal NADPH oxidase activity.

In conclusion, we suggest for the first time that low doses of As₂O₃ may constitute a new strategy to deplete Tregs in colorectal tumors. Indeed, we have recently shown that As₂O₃ can deplete Tregs through oxidative and nitrosative bursts, thus improving antitumor immune responses. Our results offer a new opportunity to use low doses of As₂O₃ to enhance the antitumor activity of adoptive immunotherapy against human cancer.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

References

1. Curiel TJ, Coukos G, Zou L, Alvarez X, Cheng P, Mottram P, et al. Specific recruitment of regulatory T cells in ovarian carcinoma fosters immune privilege and predicts reduced survival. *Nat Med* 2004; 10:942-9; PMID:15322536; <http://dx.doi.org/10.1038/nm1093>.
2. Mouggiakakos D, Choudhury A, Lladser A, Kiessling R, Johansson CC. Regulatory T cells in cancer. *Adv Cancer Res* 2010; 107:57-117; PMID:20399961; [http://dx.doi.org/10.1016/S0065-230X\(10\)07003-X](http://dx.doi.org/10.1016/S0065-230X(10)07003-X).
3. Bracci L, Moschella F, Sestili P, La Sorsa V, Valentini M, Canini I, et al. Cyclophosphamide enhances the antitumor efficacy of adoptively transferred immune cells through the induction of cytokine expression, B-cell and T-cell homeostatic proliferation, and specific tumor infiltration. *Clin Cancer Res* 2007; 13:644-53; PMID:17252888; <http://dx.doi.org/10.1158/1078-0432.CCR-06-1209>.
4. Galluzzi L, Senovilla L, Zitvogel L, Kroemer G. The secret ally: immunostimulation by anticancer drugs. *Nat Rev Drug Discov* 2012; 11:215-33; PMID:22301798; <http://dx.doi.org/10.1038/nrd3626>.
5. Terme M, Pernet S, Marcheteau E, Sandoval F, Benhamouda N, Colussi O, et al. VEGFA-VEGF Receptor pathway blockade inhibits tumor-induced regulatory T cell proliferation in colorectal cancer. *Cancer Res* 2012; In Press; PMID:23108136; <http://dx.doi.org/10.1158/0008-5472.CAN-12-2325>.
6. Trachootham D, Alexandre J, Huang P. Targeting cancer cells by ROS-mediated mechanisms: a radical therapeutic approach? *Nat Rev Drug Discov* 2009; 8:579-91; PMID:19478820; <http://dx.doi.org/10.1038/nrd2803>.
7. Thomas-Schoemann A, Batteux F, Mongaret C, Nicco C, Chéreau C, Annereau M, et al. Arsenic Trioxide Exerts Antitumor Activity through Regulatory T Cell Depletion Mediated by Oxidative Stress in a Murine Model of Colon Cancer. *J Immunol* 2012; 189:5171-7; PMID:23105136; <http://dx.doi.org/10.4049/jimmunol.1103094>.
8. Baj G, Arnulfo A, Deaglio S, Mallone R, Vigone A, De Cesaris MG, et al. Arsenic trioxide and breast cancer: analysis of the apoptotic, differentiative and immunomodulatory effects. *Breast Cancer Res Treat* 2002; 73:61-73; PMID:12083632; <http://dx.doi.org/10.1023/A:1015272401822>.
9. Liu B, Pan S, Dong X, Qiao H, Jiang H, Krissansen GW, et al. Opposing effects of arsenic trioxide on hepatocellular carcinomas in mice. *Cancer Sci* 2006; 97:675-81; PMID:16827809; <http://dx.doi.org/10.1111/j.1349-7006.2006.00230.x>.
10. Wang J, Li L, Cang H, Shi G, Yi J. NADPH oxidase-derived reactive oxygen species are responsible for the high susceptibility to arsenic cytotoxicity in acute promyelocytic leukemia cells. *Leuk Res* 2008; 32:429-36; PMID:17804067; <http://dx.doi.org/10.1016/j.leukres.2007.06.006>.