



Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.



# Nanosponges: An overlooked promising strategy to combat SARS-CoV-2

Ebrahim Mostafavi<sup>a,b,\*</sup>, Siavash Iravani<sup>c,\*</sup>,  
Rajender S. Varma<sup>d</sup>

<sup>a</sup>Stanford Cardiovascular Institute, Stanford University School of Medicine, Stanford, CA 94305, USA

<sup>b</sup>Department of Medicine, Stanford University School of Medicine, Stanford, CA 94305, USA

<sup>c</sup>Faculty of Pharmacy and Pharmaceutical Sciences, Isfahan University of Medical Sciences, Isfahan, Iran

<sup>d</sup>Regional Centre of Advanced Technologies and Materials, Czech Advanced Technology and Research Institute, Palacky University in Olomouc, Slechtitelu 27, 783 71 Olomouc, Czech Republic

Among explored nanomaterials, nanosponge-based systems have exhibited inhibitory effects for the biological neutralization of, and antiviral delivery against, severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2). More studies could pave the path for clarification of their biological neutralization mechanisms as well as the assessment of their long-term biocompatibility and biosafety issues before clinical translational studies. In this review, we discuss recent advances pertaining to antiviral delivery and inhibitory effects of nanosponges against SARS-CoV-2, focusing on important challenges and opportunities. Finally, as promising approaches for recapitulating the complex structure of different organs/tissues of the body, we discuss the use of 3D *in vitro* models to investigate the mechanism of SARS-CoV-2 infection and to find therapeutic targets to better manage and eradicate coronavirus 2019 (COVID-19).

**Keywords:** SARS-CoV-2; COVID-19; Nanosponges; Biological neutralization; Antiviral delivery; Viral inactivation



Ebrahim Mostafavi is currently at Stanford Cardiovascular Institute, Stanford School of Medicine. His research interests revolve around the use of cardiac iPSC cells to design and develop *in vitro* models (organoids, 3D bioprinted constructs, vascular grafts) for cardiovascular disease modeling and drug screening, as well as CRISPR/Cas gene-editing for cardiovascular diseases. During the course of his PhD, he received training at both Northeastern University and Harvard Medical School on engineering and development of (nano)biomaterials, nanocarriers, and 3D scaffolds (hydrogels, 3D bioprinted constructs, and nanoporous scaffolds) to create biologically complex systems for tissue engineering, regenerative/translational medicine, cancer therapy, and infectious diseases. Thus far, he has received several prestigious awards. Dr. Mostafavi is serving as Associate Editor-in-Chief of several prestigious journals within Elsevier, Springer, Dove Medical Press, T&F, Frontiers, etc. He is also an Editorial Board Member of 25+ prestigious biomedical and materials science journals. So far, he contributed to writing more than 30 book chapters and edited several books. His scholarly work consists of >110 publications with an H-index of 25 (I10-index = 50).



Dr. Iravani (Pharm.D., Ph.D.) has worked on several academic research projects at the Isfahan University of Medical Sciences (Faculty of Pharmacy and Pharmaceutical Sciences), including green and eco-friendly synthesis of nanomaterials, plant-derived nanostructures, phytochemical analysis, MXenes and their derivatives, carbon-based nanocomposites, drug/gene delivery nanosystems, biomedical engineering, and drug nanoparticles. His previous experience, of more than twelve years, centers on drug development and industrial pharmacy in various capacities including research and development, formulation, and quality control. Dr. Iravani has authored over 105 peer-reviewed scientific publications including 18 book chapters and two scientific books.



Prof. Rajender Varma (H-Index 120, Highly Cited Res. 2016, 18, 19, 20, 21) born in India (Ph.D., Delhi University 1976) is a senior scientist at U.S. EPA with a visiting position at RCPMT, Palacky University, Olomouc, Czech Republic. He has over 48 years of multidisciplinary research experience ranging from eco-friendly synthetic methods using microwaves, ultrasound, etc. to greener assembly of nanomaterials and sustainable appliances of magnetically retrievable nanocatalysts in benign media. He is a member of the editorial advisory board of several international journals, published over 820 papers, and awarded 17 U.S. Patents, 9 books, 29 book chapters, and 3 encyclopedia contributions with 55,200 citations.

\* Corresponding authors at: Stanford Cardiovascular Institute, Stanford University School of Medicine, Stanford, CA 94305, USA (E. Mostafavi), Faculty of Pharmacy and Pharmaceutical Sciences, Isfahan University of Medical Sciences, 81746-73461 Isfahan, Iran (S. Iravani). Mostafavi, E. ([ebimsv@stanford.edu](mailto:ebimsv@stanford.edu); [ebi.mostafavi@gmail.com](mailto:ebi.mostafavi@gmail.com)), Iravani, S. ([siavashira@gmail.com](mailto:siavashira@gmail.com)).

## Introduction

Various antivirals and immune modulatory agents as well as potential inhibitors, either organic or inorganic materials, have been proposed against SARS-CoV-2.<sup>1–3</sup> Recent advances in nanoscience and nanotechnology have revolutionized various fields of research, particularly medicine.<sup>4–8</sup> The application of nanotechnology in medicine has led to the emergence of a new realm of research, known as nanomedicine, opening new horizons and applications.<sup>9</sup> For instance, nanomedicine has shown significant achievements in, and potential for, the management of SARS-CoV-2, which causes COVID-19, including, but not limited to, the design of innovative drug and delivery systems with high efficiency and efficacy, the design of smart nanobiosensors for early detection of the virus, production of high-efficiency personal protective equipment, and so on.<sup>10,11</sup> However, comprehensive clinical trials and clinical translational studies are needed to evaluate the efficacy of these approaches. Innovative pharmacological formulations are under development, and elaborative works have focused on the preparation of drugs and inhibitory agents for combating viral infections and modulating efficient antiviral immunity<sup>12,13</sup>; pharmacological agents have been studied for homing in on the different stages of the life cycle of SARS-CoV-2. One of the main challenges to developing drugs and potential inhibitory agents with completed preclinical/clinical assessments is the time required for exhaustive clinical trials.<sup>14</sup> In addition, the impact of viral mutations can have a significant effect on both the resistance and susceptibility of the virus to inhibitory agents as well as their pathogenicity; the ensuing complications ought to be considered when designing advanced nanoscale materials against SARS-CoV-2.<sup>15,16</sup> Thus, there is an urgent need to fast-track efficient inhibitory agents, such as nanodevices and nanoformulations, with high safety to control and eliminate this virus.<sup>10,17–20</sup> Some of the targets comprise the RNA-dependent polymerases, hemagglutinin esterase, spike (S) and envelop (E) proteins, and viral proteases (e.g., 3-chymotrypsin-like protease), which have been explored as inhibitory agents, whereas other candidates are aimed at targeting the angiotensin-converting enzyme 2 (ACE2) receptor to treat COVID-19.<sup>21–23</sup> In addition, various

nanovaccines, including mRNA–lipid nanoparticle-based vaccines, have been developed and are in clinical trials for further evaluations against a variety of viruses, including Zika, influenza, and rabies.<sup>24</sup> However, the higher costs associated with maintaining, manufacturing, and transporting thermolabile molecules (such as RNA, or lipids) might be prohibitive to massive vaccination of the developing world.<sup>25</sup>

Limitations to the efficacy and delivery of drugs, and their associated adverse effects, have led researchers to move toward nanocarriers with unique properties, optimal effectiveness, specificity, and fewer adverse effects; an effective solution is to deploy nanostructures to effectively transport antiviral drugs or other formulations.<sup>26,27</sup> In this context, nanosponges have shown several advantages, such as biocompatibility, porosity, biomimetic features, sustained release behavior, and therapeutic activity (e.g., antimicrobial action against pathogenic bacteria), which make them suitable candidates for improving the bioavailability, stability, and solubility of therapeutic agents or drugs to provide the desired pharmacokinetics (PK) effects<sup>28–30</sup>; nanosponges can generate a variety of complexes with hydrophilic or lipophilic molecules, improving their transferring and protecting them from degradability.<sup>31–33</sup> For instance,  $\beta$ -cyclodextrin nanosponge-based delivery systems<sup>34</sup> were designed to formulate lipophilic drugs (e.g., dexibuprofen), offering an alternative strategy for enhancing the solubility of these drugs and improving their oral administration.<sup>35</sup> Versatile applications of nanosponges in different fields of biomedical, pharmaceuticals, environmental, catalysis, and sensors, along with their advantages in each field, are discussed in detail elsewhere<sup>36–39</sup>; in this review, we focus mainly on the delivery of drugs and antiviral agents using nanosponges.

Fig. 1 shows a schematic of the structure of cyclodextrin-based nanosponges. Nanosponges with high drug-loading or release behavior could improve the solubility of docetaxel in aqueous media, offering a promising strategy for the efficient and specific delivery of drugs, proteins/peptides, genetic materials, antineoplastic agents, and so on.<sup>40–42</sup> Lapatinib nanosponges were formulated to enhance solubility and bioavailability parameters and reduce the oral dose required for the anticancer drug lapa-

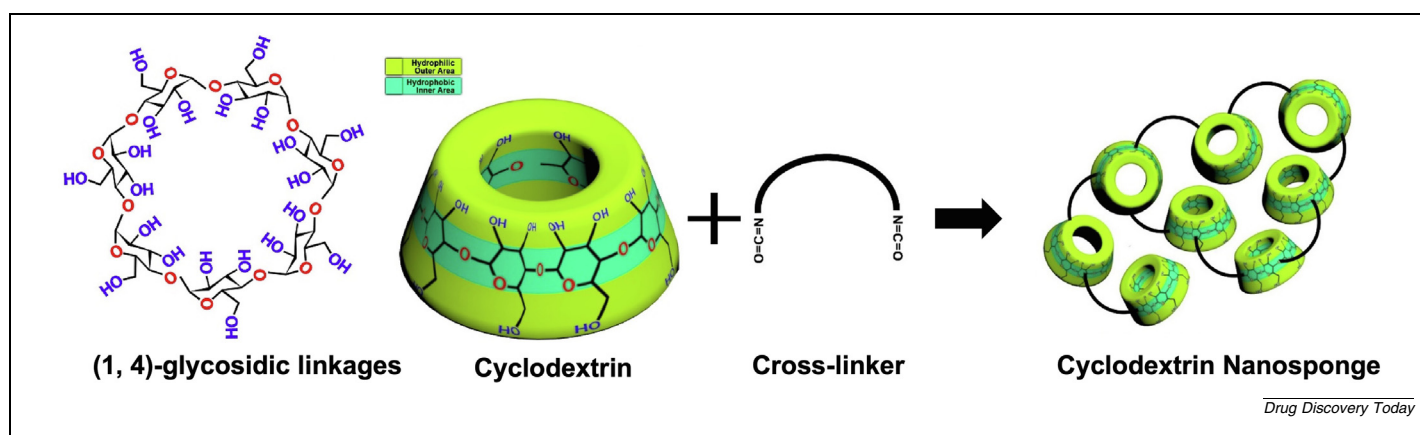


FIGURE 1

Schematic of the structure of cyclodextrin-based nanosponges. Adapted, with permission, from <sup>31</sup>.

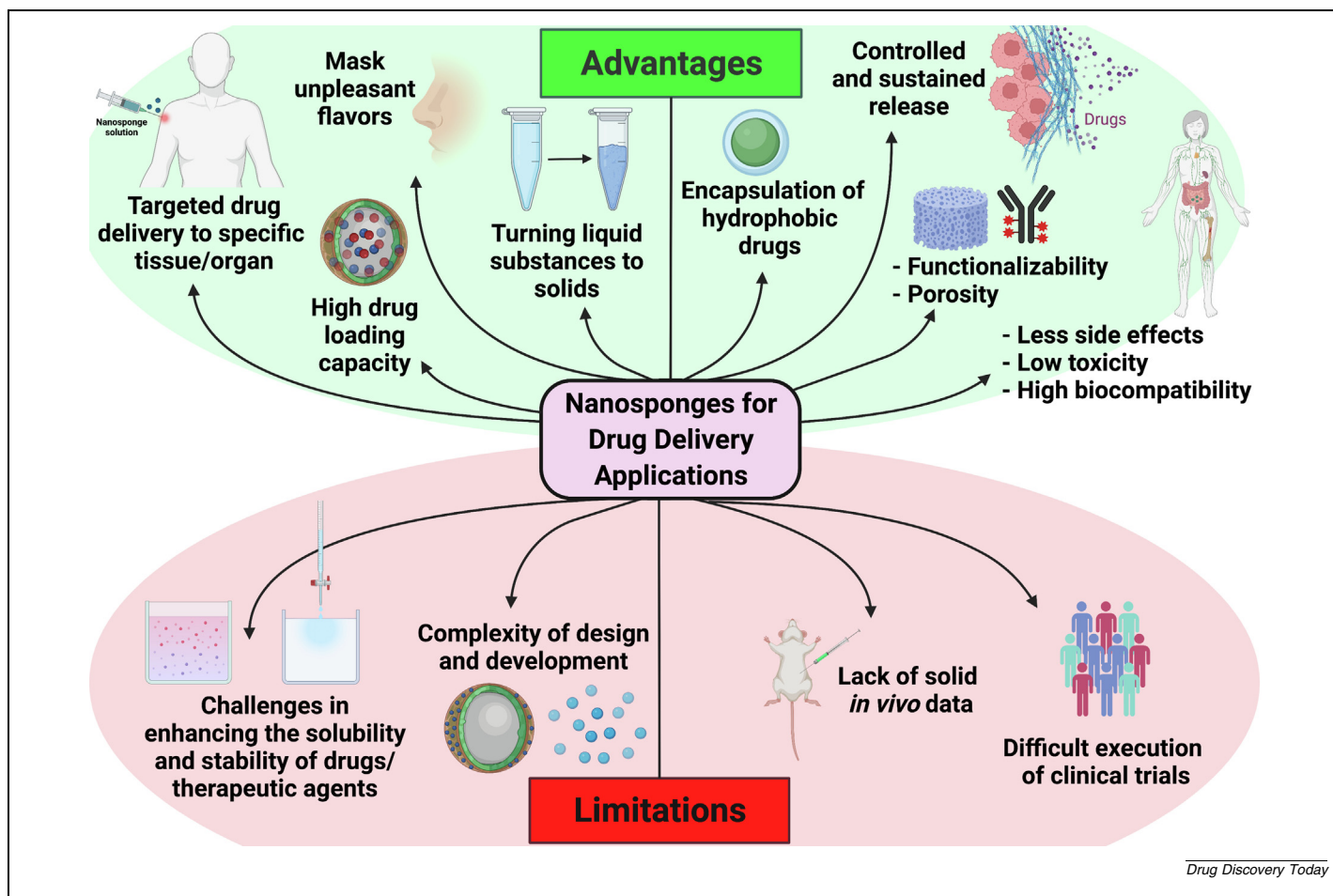
tinib.<sup>43</sup> Extensive research has been conducted to improve the selectivity, solubility, and targeting properties of anticancer drugs/agents. Palminteri *et al.*<sup>44</sup> reported an innovatively designed smart drug delivery system based on cyclodextrin nanosponges for glutathione-mediated delivery of resveratrol to the targeted cancer cells. In addition, the oral bioavailability of avanafil and dapoxetine could be improved by cyclodextrin nanosponges.<sup>45</sup> By contrast, recent advances in the preparation of bioinspired self-catabolic DNzyme nanosponges for programmable and controllable drug delivery and efficient gene-silencing activity provide promising opportunities for the development of smart gene therapeutic- and personalized nanomedicine-based strategies.<sup>46,47</sup>

One of the efficient ways to target viruses is via biological neutralization using biofunctionalized nanostructures to attach to harmful molecules/agents or pathogenic viruses to block their activities, thus preventing their replication. By developing efficient nano-scaled therapeutics with attractive advantages, such as low toxicity, targeted/sustained release behavior, good biosafety, and improved long-term biocompatibility, nanosponges can be suitable options for improving immunizations (cellular ad humoral immune responses) besides the specific/controlled delivery properties providing personalized therapeutics potentials.<sup>48–50</sup> Herein, recent advancements pertaining to the delivery

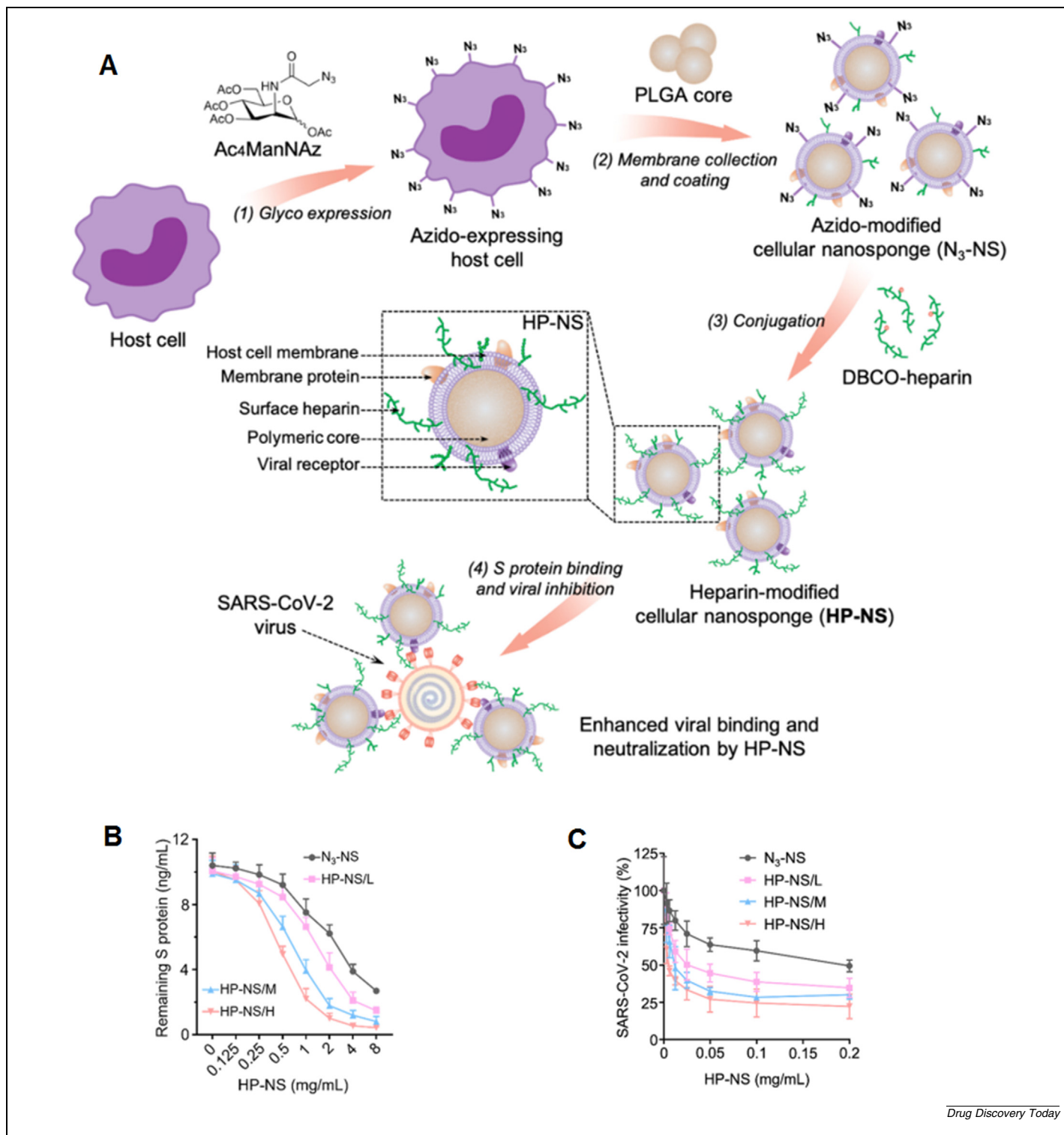
of antiviral agents as well as the inhibitory effects of nanosponge-based systems as an overlooked but promising nano-based strategy against SARS-CoV-2 are deliberated, focusing on important challenges and future perspectives.

#### Nanosponges against SARS-CoV-2

Nanosponges exist in both crystalline and paracrystalline forms, which are determined based mainly on the reaction/synthesis and processing conditions; crystallization of nanosponges can help in controlling and determining their drug-loading capacity.<sup>38</sup> Different approaches have been explored for synthesizing nanosponge-based systems, including interfacial phenomena, hot melting processes, hyper-crosslinked cyclodextrin, ultrasound-assisted synthesis, solvent condensation, microwave-assisted synthesis, interfacial condensation, mechanochemical synthesis, chain-growth polycondensation, and emulsion solvent evaporation<sup>33</sup>. A full description of each method can be found elsewhere.<sup>36,37</sup> In contrast to extensive research on the biological neutralization potential of cellular nanosponges, there are only limited investigations pertaining to their applicability as nanoplatforms for antiviral delivery (Fig. 2).<sup>26,51</sup> Cyclodextrin-based nanosponges (~400 nm) bearing carboxylic groups within their structures have been evaluated for the delivery of acyclovir and demonstrated high loading capacity



**FIGURE 2** Advantages and disadvantages of nanosponges in drug delivery applications.



Drug Discovery Today

**FIGURE 3**

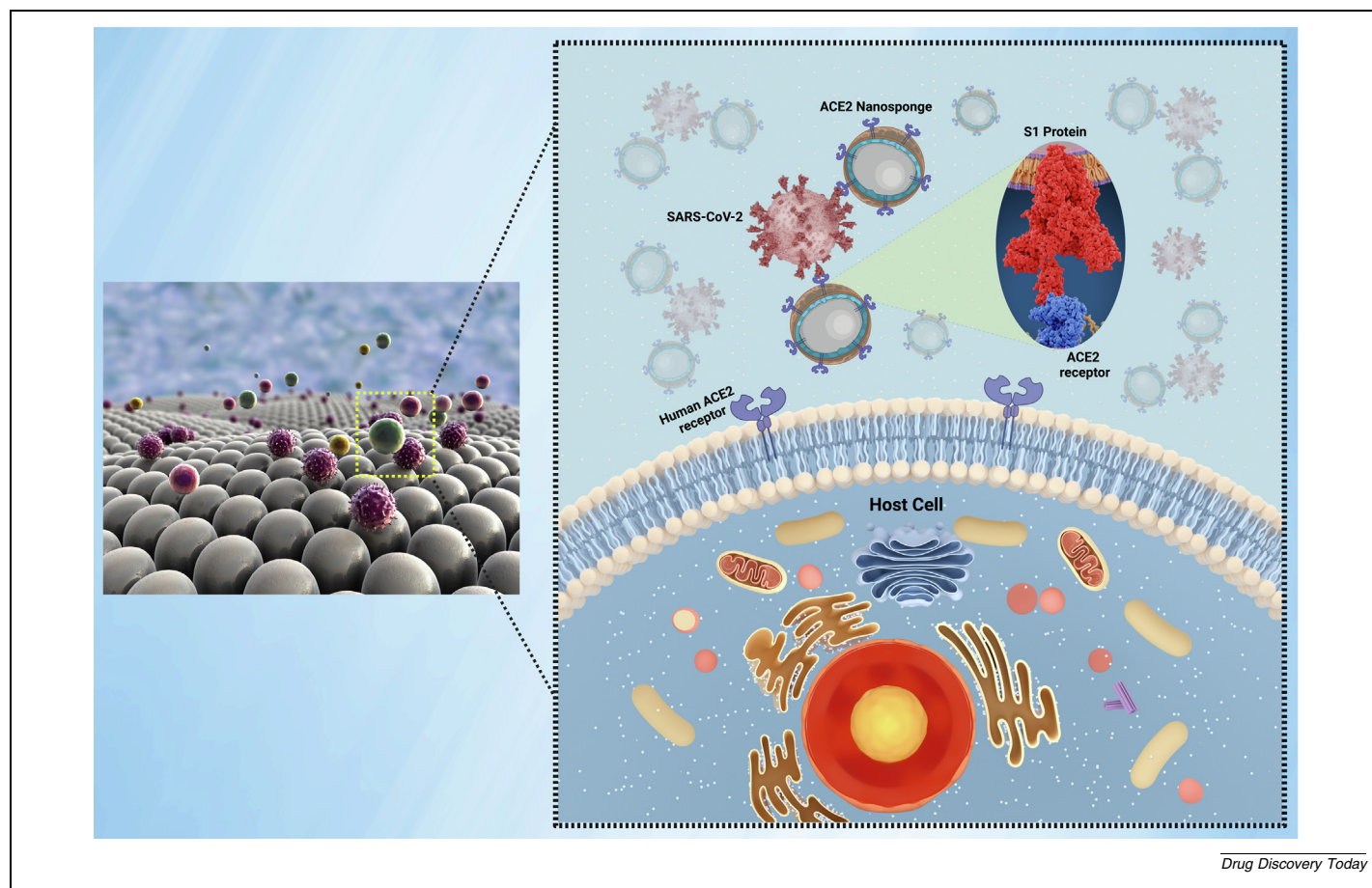
(a) Cellular nanosponges engineered for inhibitory effects against severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2). To express azido groups, the host cells were incubated with *N*-azidoacetylmannosamine-tetra-acylated (Ac4ManNAz). Then, the collected membranes were used to coat poly(lactic-co-glycolic acid) (PLGA) polymers to produce cellular nanosponges expressing azido groups (N<sub>3</sub>-NS). Functionalization with heparin was performed by applying dibenzocyclooctyne groups (DBCO-heparin), which conjugated to azido-NS via copper-free click chemistry. The resulting heparin-modified cellular nanosponges (HP-NS) were examined for SARS-CoV-2 viral infectivity applications. (b) Dose-dependent binding of different HP-NS preparations with SARS-CoV-2 S proteins. (c) Dose-dependent viral infectivity inhibition by N<sub>3</sub>-NS and various HP-NS formulations with low (L), medium (M), and high (H) heparin densities. Reproduced, with permission, from <sup>55</sup>.

for this antiviral drug (~70 % w/w) with prolonged-release performance<sup>52</sup>; further *in vivo* analysis is required in terms of comprehensive efficacy and biodistribution assessments of these nanosponges. Notably, the cytotoxicity and biosafety parameters are vital for future clinical and biomedical uses of nanosponge-based systems. Rao *et al.*<sup>53</sup> evaluated the cytotoxicity of  $\beta$ -cyclodextrin nanosponges constructed for encapsulation of a phytotherapeutic agent (Babchi oil) against HaCaT cell lines. The prepared nanosponge-based delivery system exhibited high biosafety, stability, and therapeutic effects with targeting properties, reducing the required dose/consumption of the drug/therapeutic agents and minimizing systemic adverse effects, with improved drug localization at the targeted sites. Babchi oil has been explored for its antibacterial, antifungal, antioxidant, anti-inflammatory, immunomodulatory, and antitumor effects; however, its antiviral effects should be further explored.<sup>53</sup>

Various innovatively designed nanomaterials have been widely explored as antivirals because of their potential to mimic the cellular attachment of pathogenic viruses. Given that these viruses bind to molecules on host cells using glycoproteins on their surfaces, nanosponges could be designed based on this premise after eliminating the cellular contents while retaining only the membranes. Subsequently, these membranes can be broken into thousands of tiny vesicles ~100-nm wide. Biocompatible

and biodegradable nanostructures built from polymers, such as poly(lactic-co-glycolic acid), could be coated with cellular membranes to form core-shell structures with high stability, functioning as a decoy of a human cell. The designed nanosponges with their binding points on the membranes can surround viruses, preventing them from entering host cells.<sup>54,55</sup> Rao *et al.*<sup>56</sup> reported a powerful two-step neutralization approach based on a decoy nanoparticle against COVID-19, including SARS-CoV-2 neutralization, followed by cytokine neutralization. These nanosponges can effectively protect host cells from infection by SARS-CoV-2 by competing with them for virus binding. By introducing these nanosponges, interactions between the complex of the SARS-CoV-2 S protein and human ACE2 decreased and, instead, viral receptors of the nanosponges showed high affinity for binding to ACE2.

Overall, cellular binding and the entry of SARS-CoV-2 can be mediated by its S protein via attachment to the ACE2 receptor and glycosaminoglycans (e.g., heparin).<sup>55</sup> Accordingly, cellular nanosponges mimic host cells for attracting and neutralizing SARS-CoV-2 through natural cellular receptors, thus providing a broad-spectrum antiviral approach. Increasing the heparin density on the cellular nanosponge surface also improved the inhibitory effects of heparin against the virus. Consequently, azido was expressed on host cell membranes to prepare cellular nanos-



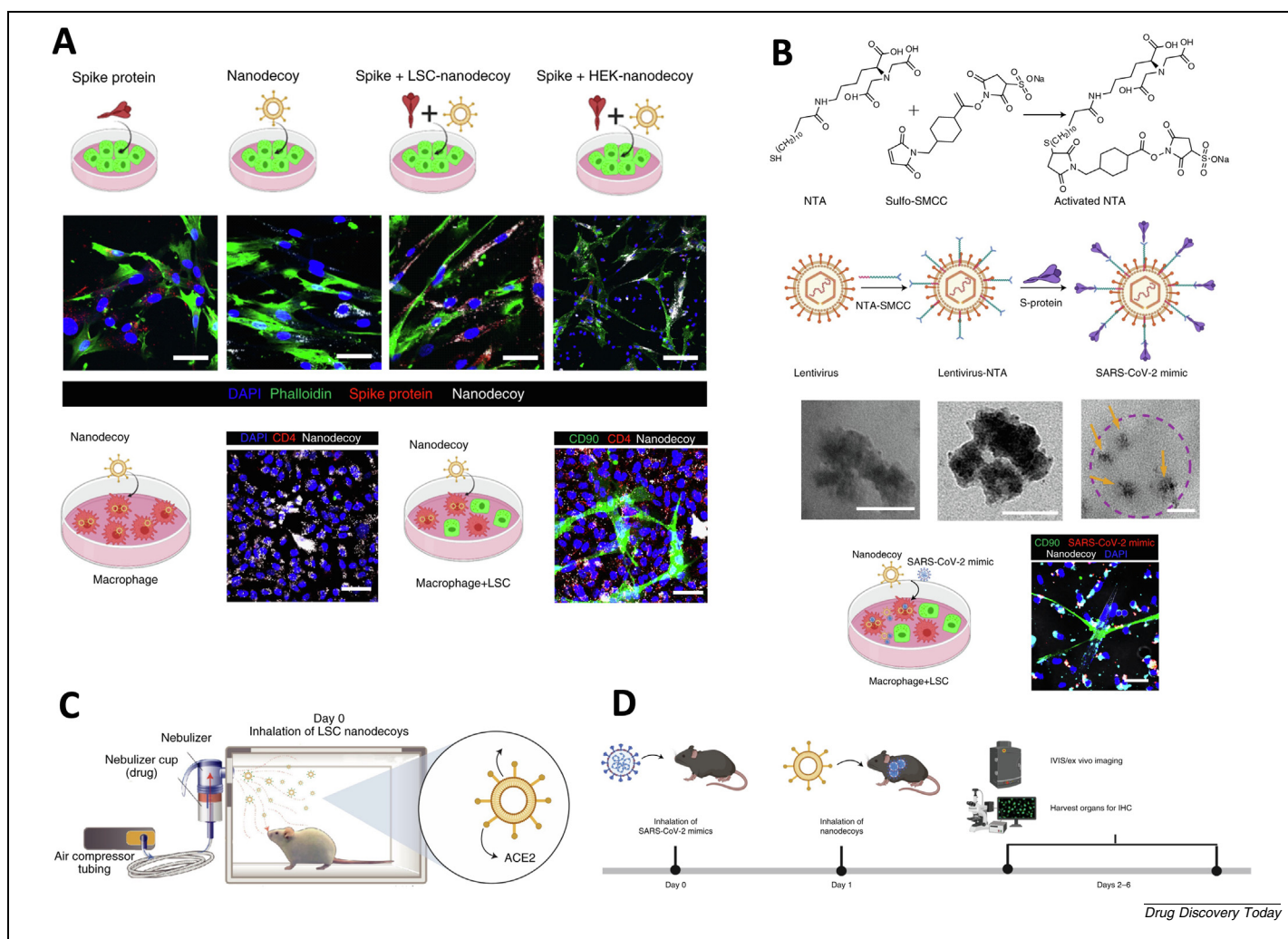
**FIGURE 4**

Membrane nanoparticles constructed from angiotensin-converting enzyme 2 (ACE2)-rich cells with inhibitory effects against severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2).

ponges with heparin adhered to their surfaces. These cellular nanosponges demonstrated significant binding potential with viral S glycoproteins, providing effectual inhibition against SARS-CoV-2 infectivity. Thus, the surface engineering of host-mimicking cellular nanosponges with glycosaminoglycans is a promising technique for enhancing inhibition of SARS-CoV-2, and could be extended to other glycan-dependent viruses (Fig. 3).<sup>55</sup> In addition, plasma membrane-derived nanosponges have been evaluated for *in vitro* neutralization of SARS-CoV-2; these plasma membranes originated from human lung epithelial type II cells or human macrophage-coated poly(lactic-co-glycolic acid) nanoparticles.<sup>54</sup> The designed nanosponges contained protein receptors essential for the cellular entry of SARS-CoV-2. Consequently, SARS-CoV-2 was neutralized, and its infectivity was reduced in a concentration-dependent manner; virus was unable to infect cells after the incubation with these engineered nanosponges. Such nanosponge-based platforms with promising inhi-

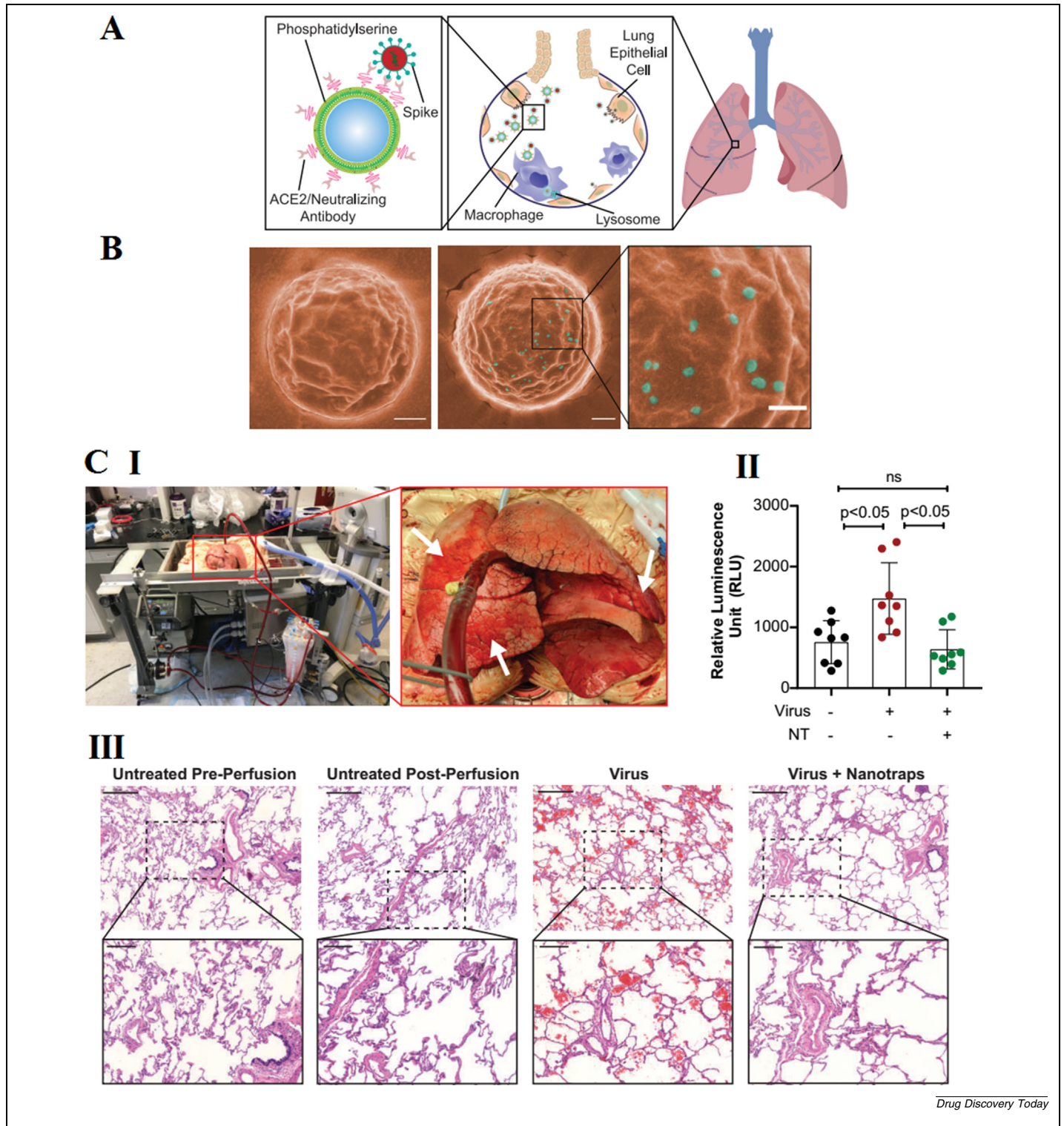
bitory effects should be further evaluated against SARS-CoV-2 mutants and other viral species. Notably, nanosponges shrouded with the cell membranes of macrophages offer the extra advantage of soaking up circulating inflammatory cytokine proteins generated as a result of the immune system response to viral infection.<sup>54</sup>

Hybrid membrane-coated biomimetic nanomaterials with attractive and unique biological properties have attracted increasing research attention for biological and biomedical applications, especially for drug delivery,<sup>57</sup> with an emphasis on immune therapy, tumor vaccines, phototherapy, and detoxification.<sup>58</sup> For example, membrane nanoparticles derived from ACE2-rich cells displayed efficient blocking effects against SARS-CoV-2 infectivity (Fig. 4) because they efficiently suppressed the entrance of SARS-CoV-2 S pseudo-virions into host cells by obstructing viral infectivity both *in vitro* and *in vivo*.<sup>59</sup> Biomimetic nanocarriers have been constructed for antiviral drug



**FIGURE 5**

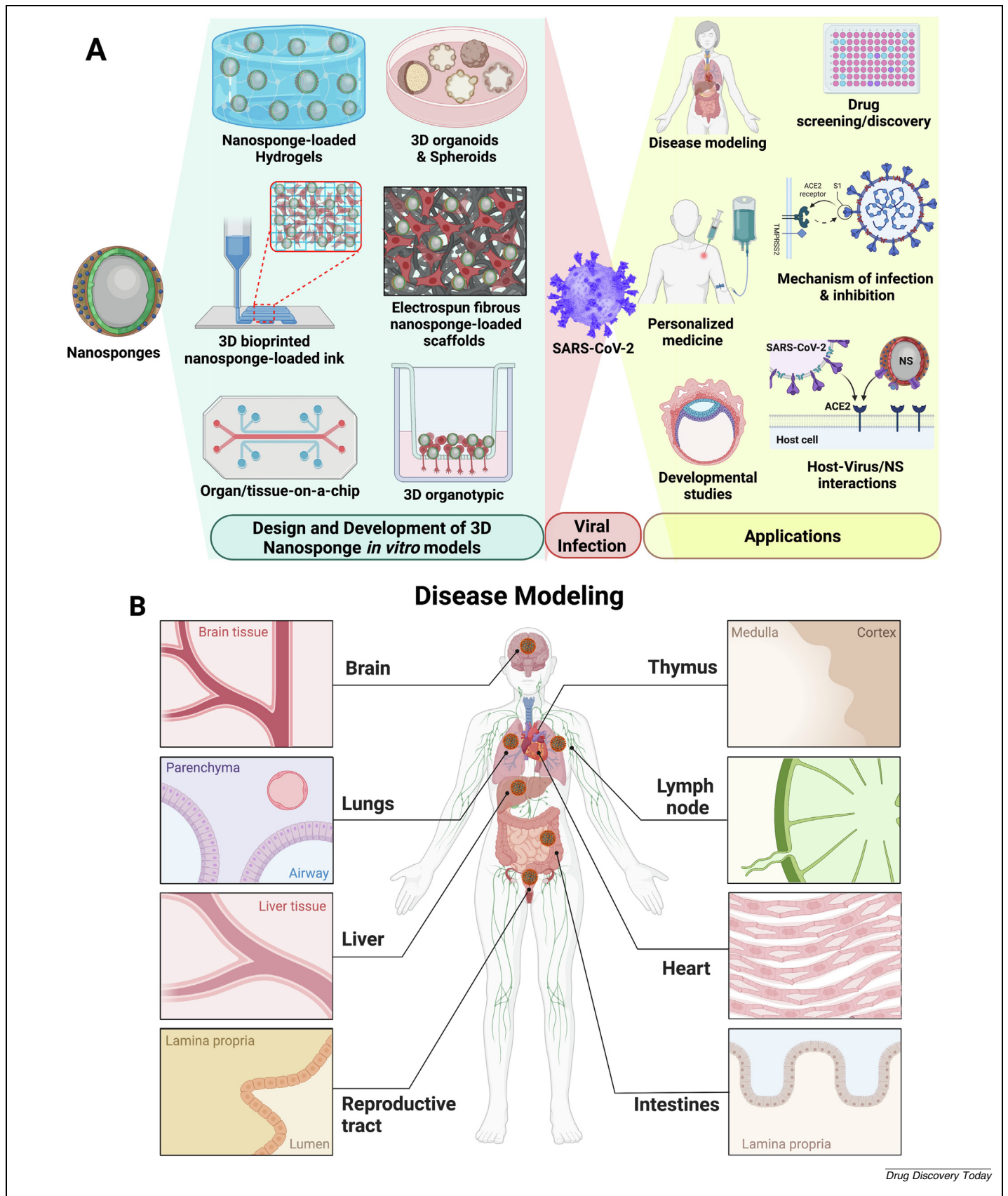
(a) Nanodecoys designed for neutralizing the S1 spike protein. S1 (red) and nanodecoys (white) interacted after their co-culture with lung cells (green). These nanodecoys were internalized by macrophages (confocal image; CD4, red). In addition, the nanodecoys were internalized by macrophages after co-culturing with lung cells (confocal image; CD90, green). Scale bars: 50  $\mu$ m. (b) SARS-CoV-2-mimicking viruses were neutralized using the designed nanodecoys. Scale bars: 100 nm. (c) Inhalation of nanodecoys in a mouse model: these nanodecoys directly accumulated in the lung, which is one of the main targets of SARS-CoV-2 replication/infection. (d) The inhalation of nanodecoys enhanced clearance of the SARS-CoV-2 mimic in a mouse model. Reproduced, with permission, from <sup>66</sup>.



**FIGURE 6**

(a) Functionalized liposomal-based nanotrappers utilizing anti-severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2)-neutralizing antibodies, phagocytosis-specific phosphatidylserines, or recombinant angiotensin-converting enzyme 2 (ACE2). The accumulation and trapping of SARS-CoV-2 virions was achieved by these nanotrappers in the lung, producing virus–nanotrap complexes. (b) Pseudo-colored scanning electron microscopy (SEM) images of nanotrap complexes with SARS-CoV-2 pseudovirus. (c) (i) Untreated (right upper lobe), virus alone (right middle lobe), and virus–nanotrap complexes (lingula) regions in a human *ex vivo* lung perfusion system. (ii) Luciferase expression quantification analyses, 8 h after infection. (iii) *Ex vivo* lung perfusion after hemoxilyn and eosin (H&E) staining, including untreated, virus alone, and virus–nanotrap complex samples. Reproduced, with permission, from <sup>69</sup>.





delivery applications against COVID-19 and concurrently exhibit anti-inflammatory effects.<sup>60</sup> Lopinavir-loaded polymeric nanoparticles were coated with macrophage membranes, neutralizing proinflammatory cytokines and suppressing macrophage/neutrophil activation. The increased potential to target inflammation as well as enhanced therapeutic effects resulting in the reduction of viral loads are the important advantages of these nanosystems.<sup>60</sup>

Zhou *et al.*<sup>61</sup> illustrated an innovative tool with prophylactic and therapeutic uses. Plasma membrane-ACE2-extracellular vesicles were deployed for neutralizing SARS-CoV-2 (pseudo-type and authentic) in human ACE2 transgenic mice, with efficient blocking effects reported against viral loads of authentic SARS-CoV-2, thus protecting the host from lung inflammation induced by SARS-CoV-2 infection.<sup>61</sup> Typically, antiviral agents target a singular viral site, thus suffer limitations of infectiveness against viruses because of their various mutations and escape strategies.<sup>62–64</sup> For instance, biomimetic proteolipid bilayer decoy receptor nanospheres constructed molecularly (~100-nm nanospheres) were explored against SARS-CoV-2, and demonstrated promising inhibitory effects. This nanosystem neutralized SARS-CoV-2 infections in animal/human cells and also trapped the viral particles. Intravenous (I)-injected nanosphere-based platforms can be suitable for multivalent capturing of SARS-CoV-2 in the body; however, preclinical and clinical evaluations are required to understand the underlying mechanisms and its efficacy against several mutated variants of SARS-CoV-2.<sup>62</sup> In particular, the emergence of new variants of concern (VOCs) of SARS-CoV-2 (such as Omicron and its different variants) caused health experts to re-evaluate the effectiveness of available strategies and think about more comprehensive and critical studies to better manage the ongoing pandemic as well as possible future outbreaks.<sup>64,65</sup> Li *et al.*<sup>66</sup> designed human lung spheroid cell-derived ACE2 nanodecoys for binding and neutralizing SARS-CoV-2, as well as protecting host lung cells from infection (Fig. 5); these nanodecoys with no noticeable toxicity could be transported through inhalation therapy and remained in the lungs for over 72 h after delivery, accelerating viral clearance and reducing lung injury. Such nanodecoys with unique properties and efficiency can be considered as potential inhibitory and therapeutic agents against COVID-19.<sup>66</sup> These nanosphere-like nanodecoys can be deployed to unravel crucial issues regarding drug development and nanotherapy, including the reduction of off-target effects (the improvement of targeting features) and unwanted biodistribution. Notably, complexes of nanosystems encompassing viruses can be used for the development of vaccine production technologies; cell membrane-mimicking nanodecoys with their ability to trap

and detain pathogenic viruses should be further evaluated for the systemic protection against, and prevention of, infectious diseases.<sup>67,68</sup>

Various nanotrappers have been designed to neutralize and capture SARS-CoV-2 and inhibit viral infections, as exemplified by functionalized nanospheres that perform as nanotrappers against SARS-CoV-2. Chen *et al.*<sup>69</sup> functionalized liposomal-based nanotrappers using anti-SARS-CoV-2-neutralizing antibodies, phagocytosis-specific phosphatidylserines, or recombinant ACE2 proteins to capture SARS-CoV-2 with complete blockage of viral infection; these nanotrappers illustrated high *in vitro* and *in vivo* biosafety and biocompatibility, representing an innovative nano-based strategy for inhibiting SARS-CoV-2 infection (Fig. 6).<sup>69</sup> It appears that these innovatively designed nanospheres- and other nanotrap-based nanoformulations have excellent potential for application as nasal sprays or inhalers<sup>70</sup> for simple and direct delivery/accumulation in the respiratory system. Moreover, additional formulations for specific targeting of these nanosystems to different sites of SARS-CoV-2 exposure show promise.

#### Engineered *in vitro* 3D-nanosponge models to combat SARS-CoV-2

The severity of COVID-19 and its exponential rate of spread (particularly newly emerged variants, such as Omicron) are clear. Therefore, the development of techniques and strategies, therapeutics targets, vaccines, and all other types of platform that can improve our understanding of the virus and its mechanism of infection in the body will help manage the spread of infection and eradicate the virus. However, because of a lack of appropriate and reliable *in vitro* models to recapitulate the complex structure of different organs of the body (from the early stages of development to a mature state), our understanding of the mechanism of SARS-CoV-2 infection in different tissues/organs is limited. Therefore, there is an unmet need for further well-developed models to be able to accurately investigate the ultrastructural, functional, molecular analysis, histochemical, and gene expression characteristics of different organs in the presence of SARS-CoV-2. Among various *in vitro* models, 3D *in vitro* models have been extensively used in viral and antiviral studies, with promising results,<sup>71</sup> given that such bioengineered 3D models provide a more realistic platform compared with simple 2D monolayer cell cultures. They can also properly and adequately recapitulate the physiological microenvironment of the tissue/organs of the body and, therefore, are able to successfully model SARS-CoV-2 virus infection and replication for the development of new drugs or vaccines to eventually manage COVID-19 (Fig. 7A). The most frequently used 3D *in vitro* models include hydrogels,<sup>72</sup> orga-

#### FIGURE 7

Development of nanosphere-laden 3D *in vitro* models to manage severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection and replication. (a) Once produced, nanospheres active against SARS-CoV-2 infection can be loaded into various 3D *in vitro* models, including hydrogels, organoids, spheroids, 3D bioprinted constructs, nanofibrous scaffolds, microfluidic-on-chips, and so on. The developed 3D model can be then used against SARS-CoV-2 infection for different applications, such as: discovery of novel and effective drugs to treat COVID-19; to study the development of different diseases in the presence of SARS-CoV-2, and the inhibitory roles of nanospheres in these developmental stages; to study the mechanism of infection and inhibition in the presence of nanospheres; to investigate host–virus complex interactions; and for disease modeling of (b) different tissues/organs that can be severely affected by SARS-CoV-2.

noids,<sup>71,73–75</sup> spheroids,<sup>76</sup> 3D bioprinted constructs,<sup>77,78</sup> nanofibrous scaffolds,<sup>79</sup> microfluidics-on-chips,<sup>73</sup> among others. For instance, microfluidics-on-a-chip or organoid models loaded with multiple cell lineages together with nanosponges are potential platforms for the early diagnosis of symptoms related to multiorgan infection. This is particularly relevant given that SARS-CoV-2 is capable of rapid spread to, and infection of, different organs of the body, including lung,<sup>80</sup> heart,<sup>81,82</sup> liver,<sup>74</sup> brain,<sup>75,83,84</sup> intestine,<sup>85</sup> reproductive system,<sup>86</sup> thymus,<sup>87</sup> and lymph nodes<sup>88</sup> (Fig. 7B).

### Concluding remarks, challenges, and prospects

Nanosponges have been deployed against SARS-CoV-2 with promising inhibitory effects for biological neutralization and antiviral drug delivery applications. However, more research is needed to clarify the precise mechanisms of action and their efficacy in animal disease models as well as their long-term biocompatibility and biosafety issues. Furthermore, some important challenging questions need to be addressed before clinical assessments, including the identification of suitable delivery pathways for these engineered nanosponges, given that they can be directly delivered to the lungs by an inhaler or intravenously transported for the treatment of cytokine storm-related complications. Future studies need to harness the additional beneficial

attributes of these membrane-cloaked nanosponges for vaccine and antiviral delivery as well as their use as broad-spectrum antiviral therapy. Additionally, such cellular nanosponges could be appropriate for the biological neutralization of chemical toxic agents, inflammatory cytokines, bacterial toxins, viral particles, and pathological antibodies. Remarkably, high-compatibility cell membrane-originating nanomaterials have opened the door to effective pharmacotherapy by circumventing the limitations of currently deployed methods, especially for immune modulation. 3D *in vitro* models can be used in viral and antiviral studies as platforms that can help understand better the SARS-CoV-2 and its mechanism of infection in the body and, thus, to manage the spread of infection, support the drug-screening process and discovery of therapeutic agents, eventually eradicating the virus.

### Conflict of Interest

The authors declare no competing interest.

### Acknowledgment

E.M. would like to acknowledge the support from the National Institute of Biomedical Imaging and Bioengineering (5T32EB009035).

### References

- 1 C. Wang, P.W. Horby, F.G. Hayden, G.F. Gao, A novel coronavirus outbreak of global health concern, *Lancet* 395 (2020) 470–473.
- 2 S. Irvani, R.S. Varma, Important roles of oligo- and polysaccharides against SARS-CoV-2: recent advances, *App Sci* 11 (2021) 3512.
- 3 T. Li, T. Huang, C. Guo, A. Wang, X. Shi, X. Mo, et al., Genomic variation, origin tracing, and vaccine development of SARS-CoV-2: a systematic review, *Innovation* 2 (2021) 100116.
- 4 V.P. Jain, S. Chaudhary, D. Sharma, N. Dabas, R.S.K. Lalji, B.K. Singh, et al., Advanced functionalized nanographene oxide as a biomedical agent for drug delivery and anti-cancerous therapy: a review, *Eur Polym J* 142 (2021) 110124.
- 5 G. Jamalipour Soufi, P. Irvani, A. Hekmatnia, E. Mostafavi, M. Khatami, S. Irvani, MXenes and MXene-based materials with cancer diagnostic applications: challenges and opportunities, *Comments on Inorg Chem* 42 (3) (2021) 174–207.
- 6 S.C. Jang, O.Y. Kim, C.M. Yoon, D.S. Choi, T.Y. Roh, J. Park, et al., Bioinspired exosome-mimetic nanovesicles for targeted delivery of chemotherapeutics to malignant tumors, *ACS Nano* 7 (2013) 7698–7710.
- 7 S. Irvani, R.S. Varma, Nanosponges for water treatment: progress and challenges, *Appl Sci* 12 (2022) 4182.
- 8 E. Mostafavi, H. Zare, Carbon-based nanomaterials in gene therapy, *OpenNano* (2022) 100062, <https://doi.org/10.1016/j.onano.2022.100062>.
- 9 E. Mostafavi, P. Soltantabar, T.J. Webster, Nanotechnology and picotechnology: a new arena for translational medicine, in: L. Yang, S.B. Bhaduri, T.J. Webster (Eds.), *Biomaterials in Translational Medicine*, Amsterdam; Elsevier, 2019, pp. 191–212.
- 10 M. Saravanan, E. Mostafavi, S. Vincent, H. Negash, R. Andavar, V. Perumal, et al., Nanotechnology-based approaches for emerging and re-emerging viruses: special emphasis on COVID-19, *Microb Pathog* 156 (2021) 104908.
- 11 S. Irvani, Nano- and biosensors for the detection of SARS-CoV-2: challenges and opportunities, *Mater Adv* 1 (2020) 3092–3103.
- 12 G. Farrugia, R.W. Plutowski, Innovation lessons from the COVID-19 pandemic, *Mayo Clin Proc* 95 (2020) 1574–1577.
- 13 M. Peplow, Nanotechnology offers alternative ways to fight COVID-19 pandemic with antivirals, *Nat Biotechnol* 39 (2021) 1172–1174.
- 14 Q. Peng, R. Peng, B. Yuan, M. Wang, J. Zhao, L. Fu, et al., Structural basis of SARS-CoV-2 polymerase inhibition by favipiravir, *Innovation* 2 (2021) 100080.
- 15 Q. Li, J. Wu, J. Nie, L. Zhang, H. Hao, S. Liu, et al., The impact of mutations in SARS-CoV-2 spike on viral infectivity and antigenicity, *Cell* 182 (2020) 1284–1294.
- 16 A.M. Seddon, Materials science in the time of coronavirus, *J Mater Sci* 55 (2020) 9145–9147.
- 17 Z. Tang, X. Zhang, Y. Shu, M. Guo, H. Zhang, W. Tao, Insights from nanotechnology in COVID-19 treatment, *Nanotoday* 36 (2021) 101019.
- 18 C. Zeng, X. Hou, M. Bohmer, Y. Dong, Advances of nanomaterials-based strategies for fighting against COVID-19, *View* 2 (2021) 20200180.
- 19 D. Desai, P. Shende, Nanoconjugates-based stem cell therapy for the management of COVID-19, *Stem Cell Rev* 17 (2021) 231–240.
- 20 N. Rabiee, S. Ahmadi, G.J. Soufi, A. Hekmatnia, M. Khatami, Y. Fatahi, et al., Quantum dots against SARS-CoV-2: diagnostic and therapeutic potentials, *J Chem Technol Biotechnol* 97 (2022) 1640–1654.
- 21 G.J. Soufi, A. Hekmatnia, M. Nasrollahzadeh, N. Shafiei, M. Sajjadi, P. Irvani, et al., SARS-CoV-2 (COVID-19): new discoveries and current challenges, *App Sci* 10 (2020) 3641.
- 22 G. Jamalipour Soufi, S. Irvani, Potential inhibitors of SARS-CoV-2: recent advances, *J Drug Target* 29 (2020) 349–364.
- 23 A.P.A. Carvalho, C.A. Conte-Junior, Recent advances on nanomaterials to COVID-19 management: a systematic review on antiviral/virucidal agents and mechanisms of SARS-CoV-2 inhibition/inactivation, *Global Challenges* 5 (2021) 2000115.
- 24 X.F. Li, Z. Cui, H. Fan, Q. Chen, L. Cao, H.Y. Qiu, et al., A highly immunogenic live-attenuated vaccine candidate prevents SARS-CoV-2 infection and transmission in hamsters, *Innovation* 3 (2022) 100221.
- 25 T. Kisby, A. Yilmazer, K. Kostarelos, Reasons for success and lessons learnt from nanoscale vaccines against COVID-19, *Nat Nanotechnol* 16 (2021) 843–850.
- 26 D. Lembo, F. Trotta, R. Cavalli, Cyclodextrin-based nanosponges as vehicles for antiviral drugs: challenges and perspectives, *Nanomedicine* 13 (2018) 477–480.
- 27 W. Chen, P. Xie, M. Pei, G. Li, Z. Wang, P. Liu, Facile construction of fluorescent traceable prodrug nanosponges for tumor intracellular pH/hypoxia dual-triggered drug delivery, *Colloid Interface Sci Commun* 46 (2022) 100576.
- 28 D. Desai, P. Shende, Drug-free cyclodextrin-based nanosponges for antimicrobial activity, *J Pharm Innovation* 16 (2021) 258–268.
- 29 S. Allahyari, F. Zahednezhad, M. Khatami, N. Hashemzadeh, P. Zakeri-Milani, F. Trotta, Cyclodextrin nanosponges as potential anticancer drug delivery systems to be introduced into the market, compared with liposomes, *J Drug Delivery Sci Technol* 67 (2022) 102931.

- 30 J. Deng, Q.J. Chen, W. Li, Z. Zuberi, J.X. Feng, Q.L. Lin, et al., Toward improvements for carrying capacity of the cyclodextrin-based nanosponges: recent progress from a material and drug delivery, *J Mater Sci* 56 (2021) 5995–6015.
- 31 A.P. Sherje, B.R. Dravyakar, D. Kadam, M. Jadhav, Cyclodextrin-based nanosponges: a critical review, *Carbohydr Polym* 173 (2017) 37–49.
- 32 Y. Khazaei Monfared, M. Mahmoudian, C. Cecone, F. Caldera, P. Zakeri-Milani, A. Matencio, et al., Stabilization and anticancer enhancing activity of the peptide nisin by cyclodextrin-based nanosponges against colon and breast cancer cells, *Polymers* 14 (2022) 594.
- 33 S. Pawar, P. Shende, A comprehensive patent review on  $\beta$ -cyclodextrin cross-linked nanosponges for multiple applications, *Recent Pat Nanotechnol* 14 (2020) 75–89.
- 34 S. Allahyari, F. Zahednezhad, M. Khatami, N. Hashemzadeh, P. Zakeri-Milani, F. Trotta, Cyclodextrin nanosponges as potential anticancer drug delivery systems to be introduced into the market, compared with liposomes, *J Drug Delivery Sci Technol* 67 (2021) 102931.
- 35 Q. Khalid, M. Ahmad, M.U. Minhas, F. Batool, N.S. Malik, M. Rehman, Novel  $\beta$ -cyclodextrin nanosponges by chain growth condensation for solubility enhancement of dexibuprofen: characterization and acute oral toxicity studies, *J Drug Delivery Sci Technol* 61 (2021) 102089.
- 36 G. Utzeri, P.M. Matias, D. Murtinho, A.J. Valente, Cyclodextrin-based nanosponges: overview and opportunities, *Front Chem* 10 (2022) 859406.
- 37 K. Tiwari, S. Bhattacharya, The ascension of nanosponges as a drug delivery carrier: preparation, characterization, and applications, *J Mater Sci: Mater Med* 33 (3) (2022) 1–21.
- 38 A. Girigoswami, K. Girigoswami, Versatile applications of nanosponges in biomedical field: a glimpse on SARS-CoV-2 management Published online June 20, *Bionanosci* (2022), <https://doi.org/10.1007/s12668-022-01000-1>.
- 39 R.K. Jani, N. Patel, Z. Patel, P. Dave, V. Upadhyay, Nanosponges as a biocatalyst carrier—an innovative drug delivery technology for enzymes, proteins, vaccines, and antibodies, *Biocatal Agric Biotechnol* (2022) 102329.
- 40 S.S.B. Rizvi, N. Akhtar, M.U. Minhas, A. Mahmood, K.U. Khan, Synthesis and characterization of carboxymethyl chitosan nanosponges with cyclodextrin blends for drug solubility improvement, *Gels* 8 (2022) 55.
- 41 I. Krabicová, S.L. Appleton, M. Tannous, G. Hoti, F. Caldera, A. Rubin Pedrazzo, et al., History of cyclodextrin nanosponges, *Polymers* 12 (2020) 1122.
- 42 F. Caldera, M. Tannous, R. Cavalli, M. Zanetti, F. Trotta, Evolution of cyclodextrin nanosponges, *Int J Pharm* 531 (2017) 470–479.
- 43 S. Allahyari, N. Esmailnezhad, H. Valizadeh, M. Ghorbani, M. Jelvehgari, F. Ghazi, et al., In-vitro characterization and cytotoxicity study of flutamide loaded cyclodextrin nanosponges, *J Drug Delivery Sci Technol* 61 (2021) 102275.
- 44 M. Palminteri, N.K. Dhakar, A. Ferraresi, F. Caldera, C. Vidoni, F. Trotta, et al., Cyclodextrin nanosponge for the GSH-mediated delivery of resveratrol in human cancer cells, *Nanotheranostics* 5 (2021) 197–212.
- 45 A.R. Gardouh, S. Elhusseiny, S. Gad, Mixed avanafil and dapoxetine hydrochloride cyclodextrin nano-sponges: preparation, in-vitro characterization, and bioavailability determination, *J Drug Delivery Sci Technol* 68 (2022) 103100.
- 46 J. Wang, S. Yu, Q. Wu, X. Gong, S. He, J. Shang, et al., A self-catabolic multifunctional DNzyme nanosponge for programmable drug delivery and efficient gene silencing, *Angew Chem* 133 (2021) 10861–10869.
- 47 Y. Jin, L. Liang, X. Sun, G. Yu, S. Chen, S. Shi, et al., Deoxyribozyme-nanosponges for improved photothermal therapy by overcoming thermoresistance, *NPG Asia Mater* 10 (2018) 373–384.
- 48 S.P. Varahachalam, B. Lahooti, M. Chamaneh, S. Bagchi, T. Chhibber, et al., Nanomedicine for the SARS-CoV-2: state-of-the-art and future prospects, *Int J Nanomed* 16 (2021) 539–560.
- 49 M. Nasrollahzadeh, M. Sajjadi, G. Jamalipour Soufi, S. Iravani, R.S. Varma, Nanomaterials and nanotechnology-associated innovations against viral infections with a focus on coronaviruses, *Nanomaterials* 10 (2020) 1072.
- 50 Y. Duan, S. Wang, Q. Zhang, W. Gao, L. Zhang, Nanoparticle approaches against SARS-CoV-2 infection, *Curr Opin Solid State Mater Sci* 25 (2021) 100964.
- 51 S.V. Chilajwar, P.P. Pednekar, K.R. Jadhav, G.J.C. Gupta, V.J. Kadam, Cyclodextrin-based nanosponges: a propitious platform for enhancing drug delivery, *Expert Opin Drug Delivery* 11 (2014) 111–120.
- 52 D. Lembo, S. Swaminathan, M. Donalio, A. Civra, L. Pastoro, D. Aquilano, et al., Encapsulation of acyclovir in new carboxylated cyclodextrin-based nanosponges improves the agent's antiviral efficacy, *Int J Pharm* 443 (2013) 262–272.
- 53 S. Kumar, Pooja, F. Trotta, R. Rao, Encapsulation of Babchi oil in cyclodextrin-based nanosponges: physicochemical characterization, photodegradation, and in vitro cytotoxicity studies, *Pharmaceutics* 10 (2018) 169.
- 54 Q. Zhang, A. Honko, J. Zhou, H. Gong, S.N. Downs, J.H. Vasquez, et al., Cellular nanosponges inhibit SARS-CoV-2 infectivity, *Nano Lett* 20 (2020) 5570–5574.
- 55 X. Ai, D. Wang, A. Honko, Y. Duan, I. Gavrih, R.H. Fang, et al., Surface glycan modification of cellular nanosponges to promote SARS-CoV-2 inhibition, *J Am Chem Soc* 143 (2021) 17615–17621.
- 56 L. Rao, S. Xia, W. Xu, R. Tian, G. Yu, C. Gu, et al., Decoy nanoparticles protect against COVID-19 by concurrently adsorbing viruses and inflammatory cytokines, *Proc Natl Acad Sci USA* 117 (44) (2020) 27141–27147.
- 57 Y. Yang, K. Wang, Y. Pan, L. Rao, G. Luo, Engineered cell membrane-derived nanoparticles in immune modulation, *Adv Sci* 21 (2021) 2102330.
- 58 Y. Zhao, A. Li, L. Jiang, Y. Gu, J. Liu, Hybrid membrane-coated biomimetic nanoparticles (HM@BNPs): a multifunctional nanomaterial for biomedical applications, *Biomacromolecules* 22 (2021) 3149–3167.
- 59 C. Wang, S. Wang, Y. Chen, J. Zhao, S. Han, G. Zhao, et al., Membrane nanoparticles derived from ACE2-rich cells block SARS-CoV-2 infection, *ACS Nano* 15 (2021) 6340–6351.
- 60 Q. Tan, L. He, X. Meng, W. Wang, H. Pan, W. Yin, et al., Macrophage biomimetic nanocarriers for anti-inflammation and targeted antiviral treatment in COVID-19, *J Nanobiotechnol* 19 (2021) 173.
- 61 F. Xie, P. Su, T. Pan, X. Zhou, H. Li, H. Huang, et al., Engineering extracellular vesicles enriched with palmitoylated ACE2 as COVID-19 therapy, *Adv Mater* 33 (2021) 2103471.
- 62 B. Esmaeli-Azad, I. Federico, F. Rojas, J. Zapf, ViruClear: molecularly designed biomimetic nanosponges for prevention and treatment of SARS-CoV-2 infections in COVID-19 patients, *FASEB J* 35 (2021).
- 63 S. Wang, D. Wang, Y. Duan, Z. Zhou, W. Gao, L. Zhang, Cellular nanosponges for biological neutralization, *Adv Mater* 34 (2022) 2107719.
- 64 E. Mostafavi, A.K. Dubey, L. Teodori, S. Ramakrishna, A. Kaushik, SARS-CoV-2 Omicron variant: a next phase of the COVID-19 pandemic and a call to arms for system sciences and precision medicine, *MedComm* 3 (1) (2022) e119.
- 65 S. Tiwari, S. Juneja, A. Ghosal, N. Bandara, R. Khan, S.L. Wallen, et al., Antibacterial and antiviral high-performance nano-systems to mitigate new SARS-CoV-2 variants of concerns, *Curr Opin Biomed Eng* 21 (2021) 100363.
- 66 Z. Li, Z. Wang, P.C. Dinh, D. Zhu, K.D. Popowski, H. Lutz, et al., Cell-mimicking nanodecoys neutralize SARS-CoV-2 and mitigate lung injury in a non-human primate model of COVID-19, *Nat Nanotechnol* 16 (2021) 942–951.
- 67 L. Rao, R. Tian, X. Chen, Cell-membrane-mimicking nanodecoys against infectious diseases, *ACS Nano* 14 (2020) 2569–2574.
- 68 K.-C. Yang, J.-C. Lin, H.-H. Tsai, C.-Y. Hsu, V. Shih, C.-M.-J. Hu, Nanotechnology advances in pathogen- and host-targeted antiviral delivery: multipronged therapeutic intervention for pandemic control, *Drug Delivery Transl Res* 11 (2021) 1420–1437.
- 69 M. Chen, J. Rosenberg, X. Cai, A.C.H. Lee, J. Shi, M. Nguyen, et al., Nanotraps for the containment and clearance of SARS-CoV-2, *Matter* 4 (2021) 2059–2082.
- 70 R. Kumar, P. Mehta, K.R. Shankar, M.A.K. Rajora, Y.K. Mishra, E. Mostafavi, et al., Nanotechnology-assisted metered-dose inhalers (MDIs) for high-performance pulmonary drug delivery applications, *Pharm Res* (2022), <https://doi.org/10.1007/s11095-022-03286-y>. Published online May 12.
- 71 V. Monteil, H. Kwon, P. Prado, A. Hagelkrüys, R.A. Wimmer, M. Stahl, et al., Inhibition of SARS-CoV-2 infections in engineered human tissues using clinical-grade soluble human ACE2, *Cell* 181 (4) (2020) 905–913.
- 72 J. Chen, B. Wang, J.S. Caserto, K. Shariati, P. Cao, Y. Pan, et al., Sustained delivery of SARS-CoV-2 RBD subunit vaccine using a high affinity injectable hydrogel scaffold, *Adv Healthcare Mater* 11 (2) (2022) 2101714.
- 73 R. Ramezankhani, R. Solhi, Y.C. Chai, M. Vosough, C. Verfaillie, Organoid and microfluidics-based platforms for drug screening in COVID-19, *Drug Discovery Today* 27 (2022) 1062–1076.
- 74 Y. Wang, S. Liu, H. Liu, W. Li, F. Lin, L. Jiang, et al., SARS-CoV-2 infection of the liver directly contributes to hepatic impairment in patients with COVID-19, *J Hepatol* 73 (2020) 807–816.
- 75 B.Z. Zhang, H. Chu, S. Han, H. Shuai, J. Deng, Y.F. Hu, et al., SARS-CoV-2 infects human neural progenitor cells and brain organoids, *Cell Res* 30 (2020) 928–931.
- 76 D. Omer, O. Pleniceanu, Y. Gnatek, M. Namestnikov, O. Cohen-Zontag, S. Goldberg, et al., Human kidney spheroids and monolayers provide insights into SARS-CoV-2 renal interactions, *J Am Soc Nephrol* 32 (2021) 2242–2254.
- 77 W.-K. Hsiao, B. Lorber, A. Paudel, Can 3D printing of oral drugs help fight the current COVID-19 pandemic (and similar crisis in the future)?, *Expert Opin Drug Delivery* 17 (7) (2020) 899–902.
- 78 F. Akter, Y. Araf, I.B. Naser, S.K. Promon, Prospect of 3D bioprinting over cardiac cell therapy and conventional tissue engineering in the treatment of COVID-19 patients with myocardial injury, *Regener Ther* 18 (2021) 447–456.

- 79 S.A. Mohamad, E.M. Zahran, M.R.A. Fadeel, A. Albohy, M.A. Safwat, New acaciin-loaded self-assembled nanofibers as MPro inhibitors against BCV as a surrogate model for SARS-CoV-2, *Int J Nanomed* 16 (2021) 1789.
- 80 S. Damiani, M. Fiorentino, A. De Palma, M.P. Foschini, T. Lazzarotto, L. Gabrielli, et al., Pathological post-mortem findings in lungs infected with SARS-CoV-2, *J Pathol* 253 (2021) 31–40.
- 81 L. Chen, X. Li, M. Chen, Y. Feng, C. Xiong, The ACE2 expression in human heart indicates new potential mechanism of heart injury among patients infected with SARS-CoV-2, *Cardiovasc Res* 116 (6) (2020) 1097–1100.
- 82 J.W. Li, T.W. Han, M. Woodward, C.S. Anderson, H. Zhou, Y.D. Chen, et al., The impact of 2019 novel coronavirus on heart injury: a systematic review and meta-analysis, *Prog Cardiovasc Dis* 63 (4) (2020) 518–524.
- 83 A.S. Zubair, L.S. McAlpine, T. Gardin, S. Farhadian, D.E. Kuruvilla, S. Spudich, Neuropathogenesis and neurologic manifestations of the coronaviruses in the age of coronavirus disease 2019: a review, *JAMA Neurol* 77 (8) (2020) 1018–1027.
- 84 M. Kujawska, E. Mostafavi, A. Kaushik, SARS-CoV-2 getting into the brain; neurological phenotype of COVID-19, and management by nano-biotechnology, *Neural Regener Res* 2022 (18) (2023) 1.
- 85 F. Xiao, M. Tang, X. Zheng, Y. Liu, X. Li, H. Shan, Evidence for gastrointestinal infection of SARS-CoV-2, *Gastroenterology* 158 (6) (2020) 1831–1833.
- 86 N. Holtmann, P. Edimiris, M. Andree, C. Doehmen, D. Baston-Buest, O. Adams, et al., Assessment of SARS-CoV-2 in human semen—a cohort study, *Fertil Steril* 114 (2) (2020) 233–238.
- 87 M.P. Lins, S. Smaniotto, Potential impact of SARS-CoV-2 infection on the thymus, *Can J Microbiol* 67 (1) (2021) 23–28.
- 88 M.P. Steinbuck, L.M. Seenappa, A. Jakubowski, L.K. McNeil, C.M. Haqq, P.C. DeMuth, A lymph node-targeted amphiphile vaccine induces potent cellular and humoral immunity to SARS-CoV-2, *Sci Adv* 7 (6) (2021) eabe5819.