

Invited Mini Review

Emerging role of bystander T cell activation in autoimmune diseases

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Autoimmune disease is known to be caused by unregulated self-antigen-specific T cells, causing tissue damage. Although antigen specificity is an important mechanism of the adaptive immune system, antigen non-related T cells have been found in the inflamed tissues in various conditions. Bystander T cell activation refers to the activation of T cells without antigen recognition. During an immune response to a pathogen, bystander activation of self-reactive T cells via inflammatory mediators such as cytokines can trigger autoimmune diseases. Other antigen-specific T cells can also be bystander-activated to induce innate immune response resulting in autoimmune disease pathogenesis along with self-antigen-specific T cells. In this review, we summarize previous studies investigating bystander activation of various T cell types (NKT, $\gamma\delta$ T cells, MAIT cells, conventional CD4⁺, and CD8⁺ T cells) and discuss the role of innate-like T cell response in autoimmune diseases. In addition, we also review previous findings of bystander T cell function in infection and cancer. A better understanding of bystander-activated T cells versus antigen-stimulated T cells provides a novel insight to control autoimmune disease pathogenesis. [BMB Reports 2022; 55(2): 57-64]

INTRODUCTION

Autoimmune disease is known to be caused by the dysregulated immune response mediated via self-antigen-specific T cells. Genetic and environmental factors such as infection have been proposed to explain the significant and sensitive immune response to self-antigens. However, it is not still clear how a few clones of self-reactive T cells can trigger significant tissue damage in autoimmune diseases. Recently, antigen non-related T cells

have been shown to play a potential role in the pathogenesis of autoimmune diseases, as well as infection and cancer (1-3).

The antigen specificity of T cells is derived from T cell receptors (TCRs), which interact with antigen-derived peptide presented by the components of major histocompatibility complex (MHC) expressed on the surface of the antigen-presenting cells (APCs). Activated T cells express antigen-specific immunity via clonal expansion and differentiate into memory cells resulting in long-term immunity. However, antigen non-specific T cells are also involved in immune response because they can be activated independent of antigen (4-6). Such T cell activation is called "bystander activation". During pathogen infection and inflammatory response, cytokines, toll-like receptors (TLRs), ligands and other immune mediators activate self-reactive T cells, which activate the autoimmune response (Fig. 1A). Besides, initial self-antigen-mediated inflammatory response including activation of self-antigen-specific T cells can also lead to bystander activation of other antigen-specific memory T cells, which secrete pathogenic inflammatory cytokines contributing to autoimmune diseases (Fig. 1B). Therefore, in this review, we summarize previous studies investigating bystander activation of T cells in various conditions and emphasize their potential role in autoimmune disease as well as infection and cancer.

HISTORY OF BYSTANDER T CELL STUDIES

The phenomenon of T cell activation without TCR signaling has been reported since the 1980s. In 1989, Yang *et al.* reported the proliferation and activation of LCMV-specific cytotoxic T lymphocytes (CTLs) following infection of lymphocytic choriomeningitis virus (LCMV)-hyperimmunized mice with various viruses such as mouse cytomegalovirus (MCMV) and vaccinia virus (V) (7). They reported that LCMV-specific CTLs, which are not specific to the challenged virus, are bystander-activated by cytokines produced by virus infection. In 1994, when resting CD4⁺ T cells were stimulated with IL-2, IL-6, and TNF- α *in vitro*, the expression of CD69 on both CD45RO⁺ and CD45RA⁺ CD4 T cells was upregulated and CD45RO⁺ cells secreted IFN- γ and IL-4 (8). In 1996, Tough *et al.* established bystander T cell activation of CD8⁺ CD44^{high} T cells *in vivo* suggesting that bystander activation is mediated by type I interferon rather

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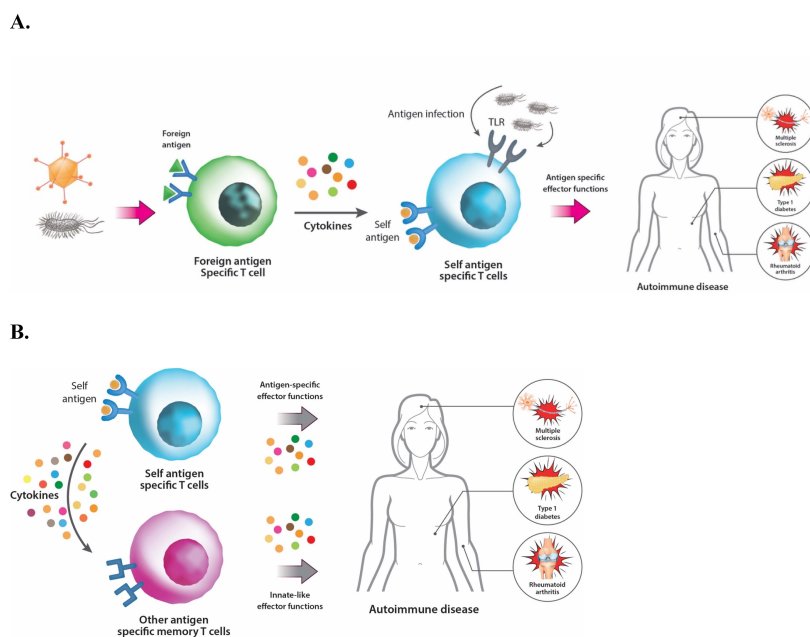


Fig. 1. Model of bystander activation of T cells. (A) Pathogen infection triggers inflammation which in turn bystander-activates self-antigen specific T cells. (B) During self-reactive T cell mediated inflammation, other antigen-specific T cells are bystander activated to produce inflammatory cytokines.

than TCR signaling (4). Another group showed that memory T cells proliferate upon NKT cell activation via IL-12 and IFN- γ -dependent pathway in addition to type I interferon. In the late 1990s, although substantial evidence suggested that T cells can be activated via TCR-independent pathway, the role of bystander-activated T cells in immune response was disputed. Zarozinski *et al.* disputed that polyclonal expansion of CTL response to LCMV infection is virus-specific and bystander activation of non-virus-specific CTL did not play a significant role (9). Additionally, Kaja *et al.* used virus-peptide tetramer staining to show that when adult mice were infected with LCMV, 50%-70% of activated CD8⁺ T cells were LCMV-specific in both primary and secondary immune response (10). However, it is clear that bystander T cell activation occurs during pathogen invasion and contributes to disease pathogenesis. In 2005, it was shown that parasitic infection may have a significant impact on the dynamics of CD4⁺ T cell populations regardless of antigen specificity (11). OVA-specific (DO11) T cells were transferred to naive recipient mice and the recipient was immunized to generate memory DO11 cells. After 6 weeks, the mouse was exposed to *Leishmania donovani* inducing expansion of both naive and memory DO11 cells. Guo *et al.* suggested that rested Th1, Th2, and Th17 cells can be bystander-activated by the IL-1 family and STAT-activating cytokines to secrete effector cytokines (12). In addition, *Nippostrongylus brasiliensis*-induced Th2 cells responded to a challenge with papain or house dust mite extract (HDM) by releasing IL-13 without TCR signaling (13). In 2018, it was reported that during acute hepatitis A (AHA), non-HAV-specific memory CD8⁺ T cells were bystander-activated via IL-15 to aggravate liver injury independent of TCR (1). We previously reported that memory Th17 cells can

be stimulated by IL-1 β and IL-23 without antigen recognition and may exacerbate experimental autoimmune encephalomyelitis (2). Likewise, intensive studies reported that bystander T cell activation and effector function can affect tissue damage.

BYSTANDER ACTIVATION OF VARIOUS T CELL TYPES

T cells are composed of different populations, each of which expresses unique surface markers and transcription factors. In the thymus, conventional T cells develop into CD4⁺ or CD8⁺ T cells with TCRs consisting of α and β chains responsible for antigen specificity. However, unconventional T cells such as $\gamma\delta$ T cells carry TCRs with γ and δ chains instead of α and β chains. Natural killer T (NKT) cells and mucosal-associated invariant T (MAIT) cells are unconventional T cells with a limited diversity or specificity to the restricted group of antigens. They are usually localized in tissues such as liver and mucosa for tissue immunity but also present in the circulation.

Natural killer T (NKT) cells

Invariant NKT (iNKT) cells recognize lipid antigens such as α -galactosylceramide (α -GalCer) presented by MHC class I-like molecule CD1d (14). NKT cells can be divided into type 1 and type 2 NKT cells. Type 1 NKT cells carry CD1d-restricted semi-variant $\alpha\beta$ TCR with limited β chain (V β 8, V β 7 or V β 2 in mice, and V β 11 in humans), while type 2 NKT cells are known to contain a broader TCR repertoire. Leite-de-Moraes reported that splenic CD4⁺ NKT cells in MHC class II-deficient mice secreted IFN- γ without further stimulation after a single injection of IL-12 plus IL-18 (15). In 2008, it was reported that IL-17RB⁺ CD4⁺ NKT cells secrete IL-13 and Th2 chemokines

upon stimulation with IL-25 *in vitro*, and depletion of these cells by IL-17RB-specific monoclonal antibodies or NKT cell-deficient mice failed to trigger IL-25-dependent airway hyper-sensitive reaction (AHR) in an animal model of asthma (16). According to Jean-Marc Doisne, NK1.1⁻ iNKT cells in peripheral lymph nodes stimulated by IL-1 and IL-23 secrete IL-17 and IL-22 to control early infections (17). In brief, various NKT subsets can exert effector functions in the TCR-independent pathway.

Gamma delta ($\gamma\delta$) T cells

Gamma delta ($\gamma\delta$) T cells are unique T cells carrying TCRs, each composed of γ and δ chains instead of α and β chains (14). They constitute about 4% of all T cells in the lymphoid tissue and are abundant in the skin and gut. Murine $\gamma\delta$ T cells can be roughly divided into two types: IFN- γ ⁺ and IL-17⁺ cells, depending on the type of cytokines they secrete (18). Haas *et al.* divided $\gamma\delta$ T cells into two subsets based on NK1.1 and CCR6 expression suggesting that stimulation with IL-12 and IL-18 induced IFN- γ production by NK1.1⁺ $\gamma\delta$ T cells, while IL-23 induced IL-17A synthesis by CCR6⁺ $\gamma\delta$ T cells (19). Bystander activation of human $\gamma\delta$ T cells remains to be studied more.

Mucosal-associated invariant T (MAIT) cells

Mucosal-associated invariant T cells (MAIT cells) express semi-invariant $\alpha\beta$ TCR and similar to other unconventional T cells have limited TCR diversity (V α 19-J α 33 and V β 8 or V β 6 in mice, V α 7.2-J α 33 and V β 2 or V β 13 in humans) (14). MAIT cells recognize microbial-derived vitamin B metabolites presented to β 2M-associated MHC-related 1 (MR1) (20). MAIT cells are CD161⁺ V α 7.2⁺ CD8⁺ CD4⁻ CD3⁺ T cells secreting effector cytokines independent of TCR. Compared with other T cell subsets, CD161^{high} CD8⁺ T cells in human PBMC express IFN- γ more sensitively in response to IL-12 and IL-18 (21). In addition, IFN- γ synthesis was TCR-independent and not inhibited by cyclosporin A, which inhibits calcineurin in TCR signaling. In 2016, Wilgenburg *et al.* found that MAIT cells are abundant and activated during human viral infections *in vivo* (22). They reported that this activation was TCR-independent but dependent on IL-18 with IL-12, IL-15, and/or IFN- α/β . Thus, MAIT cells might be activated in a TCR-independent manner and have effector functions in diseases.

CD4⁺ T cells

CD4 T cells, also known as T helper cells, can be divided into several types, such as Th1, Th2, Th17, and Treg. When they are activated, each effector CD4⁺ T cell type secretes specific cytokines following immune response. Interestingly, several studies show that CD4⁺ T cells can be activated even without TCR stimulation (23). In 1999, Gangappa *et al.* suggested that herpes simplex virus (HSV)-induced lesions may occur via bystander activation of CD4⁺ T cells because TCR-transgenic mice backcrossed to SCID mice recognizing on OVA peptide but not HSV proteins still develop ocular lesions upon HSV infection and CD4⁺ T cells are found in the lesions. Thus bystander

activation of antigen-independent CD4⁺ T cells was detected in the virus-induced milieu. Therefore, cytokines are one of the factors inducing bystander activation of CD4⁺ T cells (24). According to the study by Unutmaz *et al.*, human naive (CD45RA⁺) CD4⁺ T cells and memory (CD45RO⁺) cells are activated upon stimulation with IL-2, TNF- α , and IL-6 to express activation markers, and initiate cell cycle and proliferation (2). Chakir *et al.* showed that activation of naive T cells by high doses of IL-2 respond to IL-12 and IL-18 even in the absence of TCR ligation and express signs of Th1 phenotype (25). Guo *et al.* suggested that Th2 secretes IL-13 and IL-5 by IL-33 and STAT5 activator in an antigen-independent manner (7). In addition, Th17 secretes IL-17 by IL-1 β and STAT3 activator (26, 27), and Th1 produces IFN- γ following stimulation by IL-18 and STAT4 inducer (27, 28). Jiang *et al.* revealed that memory CD4⁺ T cell cycling during HIV infection is related to the levels of lipopolysaccharide (LPS), plasma HIV RNA, and memory CD8⁺ T cell cycling (29). Therefore, exposure to microbial products, plasma viremia, or proinflammatory cytokines resulting from HIV infection rather than peptide stimulation increases memory CD4⁺ T cells in HIV disease. According to van Aalst *et al.* in 2017, injection of complete Freund's Adjuvant (CFA) leads to CD4⁺ T cell proliferation, activation, and intravenous transfer (30). In addition, this local bystander activation occurred when incomplete Freund's Adjuvant (IFA) was used as a booster after priming with CFA. Our previous study confirmed that CD4⁺ T cells primed with IL-1 β and IL-23 differentiated similar to pathogenic Th17 cells that express ROR γ t or GM-CSF (10). Using EAE, it was confirmed that memory-like Th17 cells which are not specific to antigen can infiltrate into the spinal cord, and secrete IL-17A, IFN- γ , and GM-CSF, suggesting that the actual antigen-nonspecific cells are bystander-activated and exhibit pathogenic effects.

CD8⁺ T cells

Zhang *et al.* first reported that memory-phenotype (CD44^{high}) CD8⁺ T cells can be activated by IL-15, which mimics the effects of type I IFN (31). They also suggest that IL-12, IL-18, and IFN- γ induce selective proliferation of CD44^{high} CD8⁺ T cells *in vivo*. In 2002, it was reported that IL-15 induced not only activation of effector CD8⁺ CTLs in an antigen-independent manner and maintained for up to 60 days but also proliferation and survival of memory-phenotype CD8⁺ T cells (32). In addition, cytokine immunotherapy in cancer can promote anti-tumor immunity mediated via bystander-activated memory CD8⁺ T cells activated by IL-2 or IL-12 and expressing NKG2D and granzyme B, while depletion of NKG2D in mice decreased antitumor effects after immunotherapy (33). Chu *et al.* confirmed that bystander-activated CTLs (BA-CTL) directly eliminate target cells in NKG2D induced with IL-12, IL-18, and IL-15 suggesting the role of memory CD8⁺ T cells in the early innate immune response (34). In 2014, Crosby *et al.* found that LCMV and *Listeria* immune mice exhibited a significant increase in the size of leishmanial lesions compared with mice infected with

Leishmania major alone (35). The bystander-activated LCMV specific memory CD8⁺ T cells play a role in an unrelated immune response and increase immunopathology via NKG2D-dependent mechanism because treatment with NKG2D-blocking antibodies diminished the increased number of lesions in LCMV immune mice. Besides, during HIV infection, IL-15 may induce substantial bystander expansion and cycling as well as granzyme B⁺ CD45RO⁺ CD8⁺ T cells related to high morbidity and mortality in untreated patients (36). Recently, during HAV infection, the role of bystander CD8⁺ T cells was established in acute hepatitis A (AHA). Kim et al. reported that HAV-unrelated CD8⁺ T cells were activated by IL-15 and developed intrinsic cytotoxicity via NKG2D and Nkp30 leading to liver injury (1). They also suggest that the migration of bystander-activated memory CD8⁺ T cells from the circulation to the liver is mediated by CCR5 (37).

BYSTANDER-ACTIVATED T CELL AND AUTOIMMUNITY

Multiple sclerosis

Multiple sclerosis (MS) is a chronic autoimmune disease mainly induced by uncontrolled infiltration of T and B cells into the central nervous system (CNS) resulting in inflammation and demyelination in the CNS. In experimental autoimmune encephalomyelitis (EAE), one of the murine models of MS, the activation or adoptive transfer of myelin-recognizing T cells such as myelin oligodendrocyte glycoprotein (MOG)-specific T cells is known to play a key role in demyelination. Although, MOG-specific T cells are required for tissue damage in CNS during EAE progression, surprisingly, most CNS-infiltrated T cells are unrelated to MOG antigen specificity (38-40). Non-myelin-specific T cells such as ovalbumin (OVA)-specific T cells increased the susceptibility to EAE by enhancing the number and function of APC in the CNS following injection of a synthetic peptide of myelin basic protein (MBP) (41). Toll-like receptors (TLRs) play an important role in innate immune activation; however, the loss of TLR2 and TLR4 in CD4⁺ T cells reduced disease symptoms in the EAE model, which regulates the production of IL-17, IL-21, ROR γ t, and IFN- γ (42, 43). In addition, TLR2, TLR4, and TLR9 are highly expressed on CD4⁺ and CD8⁺ T cells of patients with MS compared with healthy controls, and the proportion of cytokine-secreting TLR⁺ T cells are correlated with the degree of brain lesion (44). In addition, injection of LPS induces bystander activation of MBP-specific T cells contributing to EAE in MBP TCR-transgenic mice, which requires physical contact with antigen-presenting cells via CD86 co-stimulation (45). Recently, our studies revealed that IL-1 β and IL-23 contributed to bystander activation of OVA-specific Th17 cells, which turn pathogenic by producing IL-17A, IFN- γ , and GM-CSF. As a result, the augmented antigen-specific T cell response led to severe EAE symptoms (2). Collectively, multiple reports demonstrated the presence of nonmyelin-specific T cells in the CNS with significant effector functions contributing to autoimmune encephalomyelitis.

Rheumatoid arthritis

Rheumatoid arthritis (RA) is an autoimmune disease caused by the destruction of joint cartilage and bone. Autoreactive CD4⁺ T cells secrete cytokines resulting in the activation of osteoclasts and destruction of joint elements. Several studies investigating the factors triggering RA proposed bystander activation of T cells as one of the causes. In 2000, Tan et al. found many Epstein-Barr virus (EBV)-specific CD8⁺ T cells in synovial fluid (46). These virus-specific T cells in joints express activation markers such as CD69 and CD38 and secrete pro-inflammatory cytokines, suggesting bystander activation of CD8⁺ T cells during arthritic inflammation. In 2004, Kobayashi et al. revealed the proliferation of autoantigen-reactive T cells induced by bystander activation via different antigenic epitopes, leading to severe lesions in aged mice with Sjogren's syndrome (47). Also, Sobek et al. demonstrated T cells can be activated by TLR2 signaling initiating RA by IFN- γ secretion in the absence of cognate antigen recognition (48). Brennan et al. found that cytokine-activated T cells (Tck cells) partially resemble the phenotypes of T cells in RA synovial tissue such as CD25^{high}, CD69^{high}, HLA-DR^{high}, CD49d^{high}, and the effector function suggested the role of cytokine-activated effector memory T cells in the synovium (49).

Type 1 diabetes

Type 1 diabetes (T1D) is an autoimmune disease caused by the selective destruction of insulin-producing β cells in the pancreas of islets of Langerhans. In 1998, Marc S. Horwitz et al. revealed that resting autoreactive T cells were re-stimulated by coxsackie virus infection and induced T1D (50). Infection with rotavirus, enterovirus, and influenza A virus can also induce T1D (51). Therefore, pathogen infection triggers bystander activation of self-reactive T cells resulting in induction of autoimmune diseases such as T1D. During the development of T1D in non-obese diabetic (NOD) mice, innate-like T cells such as iNKT17 cells infiltrate the pancreas and secrete IL-17, which exacerbates T1D (52). Likewise, in NOD mice, $\gamma\delta$ T cells infiltrate the islets of pancreas resulting in severe T1D when $\gamma\delta$ T cells are adoptively transferred to NOD mice (53). In 2017, Rouxel et al. proposed that MAIT cells play a role in direct killing of β cells in human T1D and NOD mice (54). Therefore, innate-like T cells can contribute to T1D pathogenesis via independent TCR activation.

ROLE OF BYSTANDER T CELLS IN OTHER DISEASES

Infection

Bystander T cell activation has been observed in studies of infection. Tough et al. first reported CD8⁺ CD44^{high} T cell proliferation in response to type I interferon induced by polyinosinic-polycytidylic acid (poly(I:C)) and LPS injection (4, 5). During *Burkholderia pseudomallei* infection, CD8⁺ CD44^{high} T cells are a major source of rapid synthesis of IFN- γ suggesting bystander activation of CD8 T cells by IL-12 and IL-18 induced

by the infection (55). In the early 2000s, the possibility of bystander T cell activation in HIV infection was proposed following the discovery of activated CD8⁺ T cells specific to EBV, cytomegalovirus, and influenza virus during primary HIV Infection (56). In 2014, Bastidas *et al.* showed that this activation of CD8⁺ T cells is triggered mainly by IL-15 independent of antigen specificity (57). In addition to CD8⁺ T cells, cycling memory CD4⁺ T cells were proportionally more frequent in HIV infection and activated the expression of CD38 and PD-1 (29). During influenza A virus (IAV) infection causing acute severe pneumonia, IL-15 plays a critical role in the pathogenesis of acute lung injury implying bystander activation of CD8⁺ T cells in IAV infection (58). In addition, hepatitis A (HAV) infection causes acute hepatitis A (AHA) and induce IL-15 production, which can stimulate memory CD8⁺ T cells to acquire innate-like cytotoxicity via TCR-independent pathways (1). Recently, in the middle of the COVID-19 pandemic, Gregorova *et al.* reported increased frequencies of T-cells specific for unrelated antigens in a COVID-19 patient who developed recurring ventilator-associated pneumonia (VAP) caused by *Pseudomonas aeruginosa* strain, suggesting significant bystander activation (59).

Cancer

In 2006, Joncker *et al.* reported antigen-independent activation of T cells in human and murine tumors implying tumor infiltration of effector or memory T cells independent of tumor antigens (60). However, recent studies investigated the immunological role of tumor-antigen non-related T cells (3, 33, 61, 62). Simoni *et al.* reported that bystander CD8⁺ T cells, which recognize a wide range of epitopes unrelated to cancer, are abundant in human lung and colorectal cancer and can be distinguished from tumor antigen-specific CD8⁺ T cells, suggesting that bystander CD8⁺ T cells express low level of CD39 compared with tumor-related CD8⁺ T cells (3). Further, according to Ponzetta *et al.*, unconventional T cells can participate in anti-tumor response mediated by IL-12 generated from neutrophils

and macrophages and secreting IFN-γ (63). Interestingly, injecting viral peptides into tumors to reactivate antiviral memory CD8⁺ T cells can arrest the growth of checkpoint blockade-resistant and poorly immunogenic tumors in mice (64). Danahy *et al.* showed that compared with tumor-specific CD8⁺ T cells, pathogen-induced memory CD8⁺ T cells inside tumor increased the sensitivity to tumor-derived inflammation resulting in activation and better control of tumor growth in antitumor immunity in the absence of cognate antigen recognition (65). Interestingly, the tumor microenvironment can be reprogrammed by intratumoral treatment with seasonal influenza vaccine, which could convert “cold” tumors to “hot” types resulting in systemic CD8⁺ T cell-mediated antitumor immunity (66). Likewise, ongoing studies are investigating the role of bystander-activated T cells in tumors to repurpose as cancer immunotherapy.

CONCLUSION

Various types of T cell populations such as NKT cells, γδ T cells, MAIT cells, and conventional CD4⁺ and CD8⁺ T cells can be induced to exhibit innate-like effector function via bystander activation. Although antigen-specific T cell response is a hallmark of the adaptive immune system, antigen non-related T cells proliferate significantly and synthesize effector cytokines in inflammatory tissues. During pathogen infection, antigen-specific T cells are activated by cytokines to reject tumors or to ensure protective immunity. In addition, in the absence of cognate antigen recognition, bystander activation of various types of T cells to secrete inflammatory cytokines can also facilitate the clearance of pathogen or tumor cells. Otherwise, bystander T cell function is of limited interest in understanding the mechanism of autoimmune disease pathogenesis. Few studies in RA and EAE suggest infiltration of antigen-nonrelated T cells into inflammatory tissues and synergistically contribute to autoimmune diseases. The detailed mechanisms of inflammatory infiltration of antigen non-specific T cells and regulation of their effector functions have

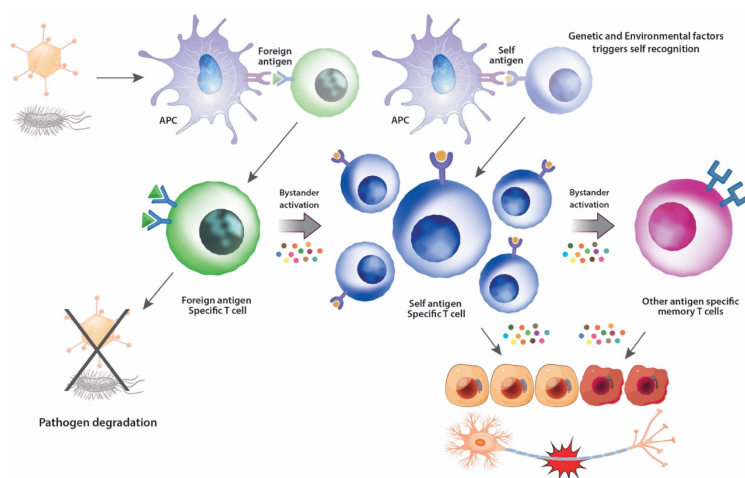


Fig. 2. Hypothesis of bystander T cell functions in autoimmune diseases.

yet to be elucidated. Based on the summary of previous studies, we propose the mechanism of autoimmune disease pathogenesis via bystander activation of either self-antigen-specific T cells or non-related T cells (Fig. 2). Further studies are needed to distinguish antigen-specific T cells from bystander-activated T cells to better understand disease pathogenesis. Collectively, the elucidation of bystander-activated T cell function along with antigen-stimulated T cells is required to understand the pathologic mechanism of autoimmune disease and to develop novel drug candidates targeting autoimmune disease.

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CONFLICTS OF INTEREST

The authors have no conflicting interests.

REFERENCES

1. Kim J, Chang DY, Lee HW et al (2018) Innate-like cytotoxic function of bystander-activated CD8(+) T cells is associated with liver injury in acute hepatitis A. *Immunity* 48, 161-173.e165
2. Lee HG, Lee JU, Kim DH, Lim S, Kang I and Choi JM (2019) Pathogenic function of bystander-activated memory-like CD4(+) T cells in autoimmune encephalomyelitis. *Nat Commun* 10, 709
3. Simoni Y, Becht E, Fehlings M et al (2018) Bystander CD8(+) T cells are abundant and phenotypically distinct in human tumour infiltrates. *Nature* 557, 575-579
4. Tough DF, Borrow P and Sprent J (1996) Induction of bystander T cell proliferation by viruses and type I interferon in vivo. *Science* 272, 1947-1950
5. Tough DF, Sun S and Sprent J (1997) T cell stimulation in vivo by lipopolysaccharide (LPS). *J Exp Med* 185, 2089-2094
6. Gangappa S, Deshpande SP and Rouse BT (1999) Bystander activation of CD4(+) T cells can represent an exclusive means of immunopathology in a virus infection. *Eur J Immunol* 29, 3674-3682
7. Yang HY, Dundon PL, Nahill SR and Welsh RM (1989) Virus-induced polyclonal cytotoxic T lymphocyte stimulation. *J Immunol* 142, 1710-1718
8. Unutmaz D, Pileri P and Abrignani S (1994) Antigen-independent activation of naive and memory resting T cells by a cytokine combination. *J Exp Med* 180, 1159-1164
9. Zarozinski CC and Welsh RM (1997) Minimal bystander activation of CD8 T cells during the virus-induced polyclonal T cell response. *J Exp Med* 185, 1629-1639
10. Murali-Krishna K, Altman JD, Suresh M et al (1998) Counting antigen-specific CD8 T cells: a reevaluation of bystander activation during viral infection. *Immunity* 8, 177-187
11. Polley R, Zubairi S and Kaye PM (2005) The fate of heterologous CD4+ T cells during *Leishmania donovani* infection. *Eur J Immunol* 35, 498-504
12. Guo L, Wei G, Zhu J et al (2009) IL-1 family members and STAT activators induce cytokine production by Th2, Th17, and Th1 cells. *Proc Natl Acad Sci U S A* 106, 13463-13468
13. Guo L, Huang Y, Chen X, Hu-Li J, Urban JF Jr and Paul WE (2015) Innate immunological function of TH2 cells in vivo. *Nat Immunol* 16, 1051-1059
14. Pellicci DG, Koay HF and Berzins SP (2020) Thymic development of unconventional T cells: how NKT cells, MAIT cells and $\gamma\delta$ T cells emerge. *Nat Rev Immunol* 20, 756-770
15. Leite-De-Moraes MC, Hameg A, Arnould A et al (1999) A distinct IL-18-induced pathway to fully activate NK T lymphocytes independently from TCR engagement. *J Immunol* 163, 5871-5876
16. Terashima A, Watarai H, Inoue S et al (2008) A novel subset of mouse NKT cells bearing the IL-17 receptor B responds to IL-25 and contributes to airway hyperreactivity. *J Exp Med* 205, 2727-2733
17. Doisne JM, Soulard V, Bécourt C et al (2011) Cutting edge: crucial role of IL-1 and IL-23 in the innate IL-17 response of peripheral lymph node NK1.1-invariant NKT cells to bacteria. *J Immunol* 186, 662-666
18. Ma R, Yuan D, Guo Y, Yan R and Li K (2020) Immune effects of $\gamma\delta$ T cells in colorectal cancer: a review. *Front Immunol* 11, 1600
19. Haas JD, González FH, Schmitz S et al (2009) CCR6 and NK1.1 distinguish between IL-17A and IFN- γ -producing gammadelta effector T cells. *Eur J Immunol* 39, 3488-3497
20. Kjer-Nielsen L, Patel O, Corbett AJ et al (2012) MR1 presents microbial vitamin B metabolites to MAIT cells. *Nature* 491, 717-723
21. Ussher JE, Bilton M, Attwod E et al (2014) CD161+ CD8+ T cells, including the MAIT cell subset, are specifically activated by IL-12+IL-18 in a TCR-independent manner. *Eur J Immunol* 44, 195-203
22. van Wilgenburg B, Schervitzl I, Hutchinson EC et al (2016) MAIT cells are activated during human viral infections. *Nat Commun* 7, 11653
23. Swain SL, Hu H and Huston G (1999) Class II-independent generation of CD4 memory T cells from effectors. *Science* 286, 1381-1383
24. Eberl G, Brawand P and MacDonald HR (2000) Selective bystander proliferation of memory CD4+ and CD8+ T cells upon NK T or T cell activation. *J Immunol* 165, 4305-4311
25. Chakir H, Lam DK, Lemay AM and Webb JR (2003) "Bystander polarization" of CD4+ T cells: activation with high-dose IL-2 renders naive T cells responsive to IL-12 and/or IL-18 in the absence of TCR ligation. *Eur J Immunol* 33, 1788-1798
26. Chung Y, Chang SH, Martinez GJ et al (2009) Critical regulation of early Th17 cell differentiation by interleukin-1 signaling. *Immunity* 30, 576-587
27. Ben-Sasson SZ, Caucheteux S, Crank M, Hu-Li J and Paul WE (2011) IL-1 acts on T cells to enhance the magnitude of in vivo immune responses. *Cytokine* 56, 122-125
28. Lee YK, Landuyt AE, Lobionda S, Sittipo P, Zhao Q and Maynard CL (2017) TCR-independent functions of Th17 cells mediated by the synergistic actions of cytokines of the IL-12 and IL-1 families. *PLoS One* 12, e0186351

29. Jiang W, Younes SA, Funderburg NT et al (2014) Cycling memory CD4+ T cells in HIV disease have a diverse T cell receptor repertoire and a phenotype consistent with bystander activation. *J Virol* 88, 5369-5380
30. van Aalst S, Ludwig IS, van der Zee R, van Eden W and Broere F (2017) Bystander activation of irrelevant CD4+ T cells following antigen-specific vaccination occurs in the presence and absence of adjuvant. *PLoS One* 12, e0177365
31. Zhang X, Sun S, Hwang I, Tough DF and Sprent J (1998) Potent and selective stimulation of memory-phenotype CD8+ T cells in vivo by IL-15. *Immunity* 8, 591-599
32. Lu J, Giuntoli RL 2nd, Omiya R, Kobayashi H, Kennedy R and Celis E (2002) Interleukin 15 promotes antigen-independent in vitro expansion and long-term survival of anti-tumor cytotoxic T lymphocytes. *Clin Cancer Res* 8, 3877-3884
33. Tietze JK, Wilkins DE, Sckisel GD et al (2012) Delineation of antigen-specific and antigen-nonspecific CD8(+) memory T-cell responses after cytokine-based cancer immunotherapy. *Blood* 119, 3073-3083
34. Chu T, Tyznik AJ, Roepke S et al (2013) Bystander-activated memory CD8 T cells control early pathogen load in an innate-like, NKG2D-dependent manner. *Cell Rep* 3, 701-708
35. Crosby EJ, Goldschmidt MH, Wherry EJ and Scott P (2014) Engagement of NKG2D on bystander memory CD8 T cells promotes increased immunopathology following *Leishmania* major infection. *PLoS Pathog* 10, e1003970
36. Younes SA, Freeman ML, Mudd JC et al (2016) IL-15 promotes activation and expansion of CD8+ T cells in HIV-1 infection. *J Clin Invest* 126, 2745-2756
37. Seo IH, Eun HS, Kim JK et al (2021) IL-15 enhances CCR5-mediated migration of memory CD8(+) T cells by upregulating CCR5 expression in the absence of TCR stimulation. *Cell Rep* 36, 109438
38. Lees JR, Sim J and Russell JH (2010) Encephalitogenic T-cells increase numbers of CNS T-cells regardless of antigen specificity by both increasing T-cell entry and preventing egress. *J Neuroimmunol* 220, 10-16
39. Walsh JT, Hendrix S, Boato F et al (2015) MHCII-independent CD4+ T cells protect injured CNS neurons via IL-4. *J Clin Invest* 125, 699-714
40. Van Kaer L, Postoak JL, Wang C, Yang G and Wu L (2019) Innate, innate-like and adaptive lymphocytes in the pathogenesis of MS and EAE. *Cell Mol Immunol* 16, 531-539
41. Jones RE, Kay T, Keller T and Bourdette D (2003) Nonmyelin-specific T cells accelerate development of central nervous system APC and increase susceptibility to experimental autoimmune encephalomyelitis. *J Immunol* 170, 831-837
42. Reynolds JM, Pappu BP, Peng J et al (2010) Toll-like receptor 2 signaling in CD4(+) T lymphocytes promotes T helper 17 responses and regulates the pathogenesis of autoimmune disease. *Immunity* 32, 692-702
43. Reynolds JM, Martinez GJ, Chung Y and Dong C (2012) Toll-like receptor 4 signaling in T cells promotes autoimmune inflammation. *Proc Natl Acad Sci U S A* 109, 13064-13069
44. Ferreira TB, Hygino J, Wing AC et al (2018) Different interleukin-17-secreting Toll-like receptor(+) T-cell subsets are associated with disease activity in multiple sclerosis. *Immunology* 154, 239-252
45. Nogai A, Siffrin V, Bonhagen K et al (2005) Lipopolysaccharide injection induces relapses of experimental autoimmune encephalomyelitis in nontransgenic mice via bystander activation of autoreactive CD4+ cells. *J Immunol* 175, 959-966
46. Tan LC, Mowat AG, Fazou C et al (2000) Specificity of T cells in synovial fluid: high frequencies of CD8(+) T cells that are specific for certain viral epitopes. *Arthritis Res* 2, 154-164
47. Kobayashi M, Yasui N, Ishimaru N, Arakaki R and Hayashi Y (2004) Development of autoimmune arthritis with aging via bystander T cell activation in the mouse model of Sjögren's syndrome. *Arthritis Rheum* 50, 3974-3984
48. Sobek V, Birkner N, Falk I et al (2004) Direct Toll-like receptor 2 mediated co-stimulation of T cells in the mouse system as a basis for chronic inflammatory joint disease. *Arthritis Res Ther* 6, R433-446
49. Brennan FM, Smith NM, Owen S et al (2008) Resting CD4+ effector memory T cells are precursors of bystander-activated effectors: a surrogate model of rheumatoid arthritis synovial T-cell function. *Arthritis Res Ther* 10, R36
50. Horwitz MS, Bradley LM, Harbertson J, Krahl T, Lee J and Sarvetnick N (1998) Diabetes induced by Coxsackie virus: initiation by bystander damage and not molecular mimicry. *Nat Med* 4, 781-785
51. Pane JA and Coulson BS (2015) Lessons from the mouse: potential contribution of bystander lymphocyte activation by viruses to human type 1 diabetes. *Diabetologia* 58, 1149-1159
52. Simoni Y, Gautron AS, Beaudoin L et al (2011) NOD mice contain an elevated frequency of iNKT17 cells that exacerbate diabetes. *Eur J Immunol* 41, 3574-3585
53. Markle JG, Mortin-Toth S, Wong AS, Geng L, Hayday A and Danska JS (2013) $\gamma\delta$ T cells are essential effectors of type 1 diabetes in the nonobese diabetic mouse model. *J Immunol* 190, 5392-5401
54. Rouxel O, Da Silva J, Beaudoin L et al (2017) Cytotoxic and regulatory roles of mucosal-associated invariant T cells in type 1 diabetes. *Nat Immunol* 18, 1321-1331
55. Lertmemongkolchai G, Cai G, Hunter CA and Bancroft GJ (2001) Bystander activation of CD8+ T cells contributes to the rapid production of IFN-gamma in response to bacterial pathogens. *J Immunol* 166, 1097-1105
56. Doisne JM, Urrutia A, Lacabaratz-Porret C et al (2004) CD8+ T cells specific for EBV, cytomegalovirus, and influenza virus are activated during primary HIV infection. *J Immunol* 173, 2410-2418
57. Bastidas S, Graw F, Smith MZ, Kuster H, Günthard HF and Oxenius A (2014) CD8+ T cells are activated in an antigen-independent manner in HIV-infected individuals. *J Immunol* 192, 1732-1744
58. Nakamura R, Maeda N, Shibata K, Yamada H, Kase T and Yoshikai Y (2010) Interleukin-15 is critical in the pathogenesis of influenza a virus-induced acute lung injury. *J Virol* 84, 5574-5582
59. Gregorova M, Morse D, Brignoli T et al (2020) Post-acute COVID-19 associated with evidence of bystander T-cell activation and a recurring antibiotic-resistant bacterial pneumonia. *Elife* 9, e63430
60. Joncker NT, Marloie MA, Chernysheva A et al (2006) Antigen-independent accumulation of activated effector/memory T lymphocytes into human and murine tumors. *Int J Cancer*

- 118, 1205-1214
61. Duhén T, Duhén R, Montler R et al (2018) Co-expression of CD39 and CD103 identifies tumor-reactive CD8 T cells in human solid tumors. *Nat Commun* 9, 2724
 62. Scheper W, Kelderman S, Fanchi LF et al (2019) Low and variable tumor reactivity of the intratumoral TCR repertoire in human cancers. *Nat Med* 25, 89-94
 63. Ponzetta A, Carriero R, Carnevale S et al (2019) Neutrophils driving unconventional T cells mediate resistance against murine sarcomas and selected human tumors. *Cell* 178, 346-360.e324
 64. Rosato PC, Wijeyesinghe S, Stolley JM et al (2019) Virus-specific memory T cells populate tumors and can be repurposed for tumor immunotherapy. *Nat Commun* 10, 567
 65. Danahy DB, Berton RR and Badovinac VP (2020) Cutting edge: antitumor immunity by pathogen-specific CD8 T cells in the absence of cognate antigen recognition. *J Immunol* 204, 1431-1435
 66. Newman JH, Chesson CB, Herzog NL et al (2020) Intratumoral injection of the seasonal flu shot converts immunologically cold tumors to hot and serves as an immunotherapy for cancer. *Proc Natl Acad Sci U S A* 117, 1119-1128