

Prostacyclin and thromboxane in benign and malignant breast tumours

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Summary 6-keto-PGF_{1α} and thromboxane B₂ were determined by radioimmunoassay in 37 extracts of breast carcinomata, 8 fibroadenomata, 12 sclerocystic-disease specimens and 51 normal breast tissues. More prostanoids were extracted from carcinomata than from normal specimens, fibroadenomata or sclerocystic-disease tissues ($P < 0.05$). The 6-keto-PGF_{1α}/TXB₂ ratio was higher in carcinomata than in normal tissues and fibroadenomata ($P < 0.05$) but was not significantly different from the ratio in sclerocystic disease. The prostaglandin levels and the 6-keto-PGF_{1α}/TXB₂ ratios from carcinomata did not correlate significantly with age, tumour size, differentiation, lymph node status, nuclear-cytoplasmic ratio, host cell reaction, mast cells, necrosis, elastosis, fibrosis or blood vessel density. Lower nuclear density was associated with lower 6-keto-PGF_{1α}/TXB₂ ratios ($P = 0.01$) whereas the latter value was higher when infiltration was lower ($P = 0.03$). There was a positive correlation between mitotic index and the 6-keto-PGF_{1α}/TXB₂ ratio ($P = 0.04$). Cumulation of variables revealed lower prostanoid ratios in tumours > 2 cm without lymph node metastasis than tumours < 2 cm with lymph node metastasis ($P = 0.05$). A first follow-up (14 months) showed a higher 6-keto-PGF_{1α}/TXB₂ ratio in patients who developed metastasis ($P = 0.04$). Our study does not confirm the hypothesis that high prostacyclin levels are a good prognostic index in breast cancer.

Honn *et al.* (1981) found that prostacyclin (PGI₂) had a beneficial influence against metastasis of B16 amelanotic melanoma tumours in mice, whereas inhibitors of PGI₂ synthesis enhanced the number of metastases. Other authors tested the antimetastatic potency of acetylsalicylic acid, indomethacin, dipyridamole, flurbiprofen, benorylate, heparin and warfarin (Gasic *et al.*, 1973; Elias *et al.*, 1973; Lione & Bosman, 1978; Bennett, 1982). Most of these studies are done on animals and showed no clearcut results.

Thromboxane (TX) A₂ is often a physiological antagonist of PGI₂ and an imbalance between them can disturb the (anti)haemostatic system. The anti-aggregating properties of nonsteroidal anti-inflammatory drugs (NSAID) can be explained by a stronger inhibition of the platelet cyclo-oxygenase in comparison with that of the vessel wall. As a result the release of TX will be lowered and aggregation will be blocked (Bunting *et al.*, 1983). Sloane *et al.* (1981) showed that some tumours are able to release cathepsin B. This enzyme stimulates the synthesis of TX and is produced in a variant of B16 melanoma which has high metastatic activity.

In order to study a possible prognostic value of the PGI₂/TX ratio in breast cancer, the stable

hydrolysis products of PGI₂ (6-keto-PGF_{1α}) and TXA₂ (TXB₂) were determined by RIA. 6-keto-PGF_{1α} and TXB₂ levels were examined in relation to the size of the tumour, axillary lymph node status, lymphatic vessel permeation, differentiation of the tumour, mitotic index density of nuclei of tumour cells, and age of the patient.

PG production can be influenced by inflammatory processes (Humes *et al.*, 1977; Brune *et al.*, 1978), and therefore the number of host-derived cells and the amount of necrosis was evaluated. Also the density of blood vessels was estimated as they could be a major source of 6-keto-PGF_{1α} (Moncada *et al.*, 1976), and platelets contribute considerably to the amounts of TXB₂ measured (Hamberg *et al.*, 1975).

Materials and methods

We obtained 108 specimens from 67 patients who underwent surgery for a breast lump. Each specimen was divided into two representative parts and prepared as described earlier by Vergote *et al.* (1985). The tissues were immediately immersed either in acetone cooled by solid CO₂ (-70°C) for 6-keto-PGF_{1α} or TXB₂ analysis, or in Bouin's liquid for histopathological examination. The tissue samples for prostanoid investigation were stored at -30°C until radioimmunoassay was performed.

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Thirty-five tumours were diagnosed as primary breast cancer (patient age range: 36–82; mean 64). The patients were classified according to the pathological TNM system (UICC, Livre de Poche), pT1aN0 9; pT1aN1a 1; pT1aN1b 4; pT1aNx 2; pT2aN0 4; pT2aN1a 2; pT2aN1b 4; pT2aNx 2; pT3aN0 2; pT4bNx 1. No patient had overt metastases at the time of surgery and none was under current treatment with non-steroidal inflammatory drugs or corticosteroids. Five local relapses (ductal carcinomata) and one cellular intracanalicular fibroadenoma were included. Twenty tumours were benign: 8 fibroadenomata (age range: 17–47, mean: 34) and 12 sclerocystic-disease specimens (age range: 37–70, mean: 49). We investigated also histologically confirmed normal breast tissue of patients with malignant tumours and fibroadenomata. Furthermore, 12 specimens of patients who had neither benign nor malignant tumours were studied. The age of the patients and the tumour size at anatomopathological examination were recorded. The amounts of 6-keto-PGF_{1 α} and TXB₂ were expressed as ng mg⁻¹ protein and from these values the 6-keto-PGF_{1 α} /TXB₂ ratio was calculated.

Histopathology

The slides were independently reviewed by two of the authors and re-evaluated by a senior pathologist. In case of discordance, the results were not included in this study.

Tumours were classified according to the methodology used earlier (Vergote *et al.*, 1985). Subdivision of histopathological variables is shown in Table III.

Radioimmunoassay (RIA)

For the extraction of prostaglandins and the protein measurement the procedure of Vergote *et al.* (1985) was used. Acetone was evaporated under nitrogen and the weight of the tissue determined. Tris buffer (50 mM, pH=8.0 at 25°C) was added (3 ml g⁻¹ tissue) and sonicated for 90 min (Bransonic). Ice was regularly added to the bath fluid. The supernatant was separated from the tissue after centrifugation at 10,000 g (Eppendorf centrifuge). RIA was performed directly on the supernatant according to Granström and Kindahl (1978). The antisera were raised in rabbits. Cross reactivities on the 50% binding level of the curve were: for the 6-keto-PGF_{1 α} -antiserum: PGF_{1 α} , 1%; 15-HETE (hydroxyeicosatetraenoic acid), 0.01%; 15-HPETE (hydroperoxyeicosatetraenoic acid), 0.01%; PGE₂, 15-keto-PGE₂, TXB₂ and AA (arachidonic acid) <0.01%; for the TXB₂ antiserum: PGD₂, 8.9%; PGF_{2 α} , 1%; PGE₂, 0.9%; 6-keto-PGF_{1 α} , 0.1%; 15-

keto-13,14-dihydro-PGF_{2 α} , AA, 15-HETE and 15-HPETE, <0.01%.

The extraction recoveries for 6-keto-PGF_{1 α} were 112±10% (mean ±SE; n=3) and for TXB₂ 97±16 (mean ±SE; n=3). The intra-assay variation coefficient for RIA of 6-keto-PGF_{1 α} and TXB₂ were 15±1% (n=103) and 13±1% (n=111) respectively.

Reagents

TXB₂ and 6-keto-PGF_{1 α} (Upjohn), (³H)-radio-labelled 6-keto-PGF_{1 α} and (³H)-TXB₂ (NEN). Tris buffer was made with trizma base (Sigma) and HCl (Merk p.a.).

Statistical analysis

Nonparametric statistical analysis was used to compare two (Wilcoxon test) or more groups (Kruskal & Wallis test; Sokal & Rohlf, 1981). Correlation coefficients were calculated by linear regression.

Results

PG levels in relation to the histopathological groups

6-keto-PGF_{1 α} levels were higher in carcinomata (CA) than in normal breast tissue (N), fibroadenomata (FA) and sclerocystic disease (SCD) ($P=0.0003$, Kruskal & Wallis). In FA, N and SCD the levels did not differ significantly ($P=0.17$, Kruskal & Wallis). CA-TXB₂ levels were significantly higher in comparison with the other groups ($P=0.05$, Kruskal & Wallis). The differences between N, FA and SCD were not significant ($P=0.83$, Kruskal & Wallis). The 6-keto-PGF_{1 α} /TXB₂ ratio in CA was higher than in N and FA ($P=0.002$, Kruskal & Wallis) which were similar ($P=0.39$). SCD and CA also gave similar ratios ($P=0.67$). These results are summarized in Table I and Figure 1. When local relapses (n=5) were calculated separately, 6-keto-PGF_{1 α} , TXB₂ and the 6-keto-PGF_{1 α} /TXB₂ ratio were respectively, median (limit values): 12.7 (0.6–15.0) ng mg⁻¹ protein, 2.5 (1.8–3.4) ng mg⁻¹ protein and 4.0 (0.3–7.3). They were similar to the ductal carcinomata ($P=0.95$; $P=0.40$ and $P=0.28$ respectively).

Histological type and differentiation

The infiltrating ductal carcinomata composed the substantial group. Statistical comparison between all the groups was difficult because some contained very few cases (Table II). We divided the tumours into two groups: undifferentiated and some degree of differentiation (small, moderate or high). Both

Table I PG-levels in relation to the pathology

Pathology	n	ng 6-keto-PGF _{1α} mg ⁻¹ protein median (semiquartiles)	ng TXB ₂ mg ⁻¹ protein median (semiquartiles)	6-keto-PGF _{1α} /TXB ₂ median (semiquartiles)
CA	37	4.4 (1.7–14.3)	0.6 (0.4–2.8)	4.6 (3.2–9.2)
N	51	1.2 (0.4–3.3)	0.4 (0.2–0.8)	2.6 (1.4–5.2)
FA	8	0.3 (0.2–1.8)	0.4 (0.2–0.5)	1.8 (0.7–4.9)
SCD	12	1.7 (0.9–3.5)	0.4 (0.2–1.0)	4.3 (2.7–8.7)

CA = carcinomata; N = normals; FA = fibroadenomata; SCD = sclerocystic disease.

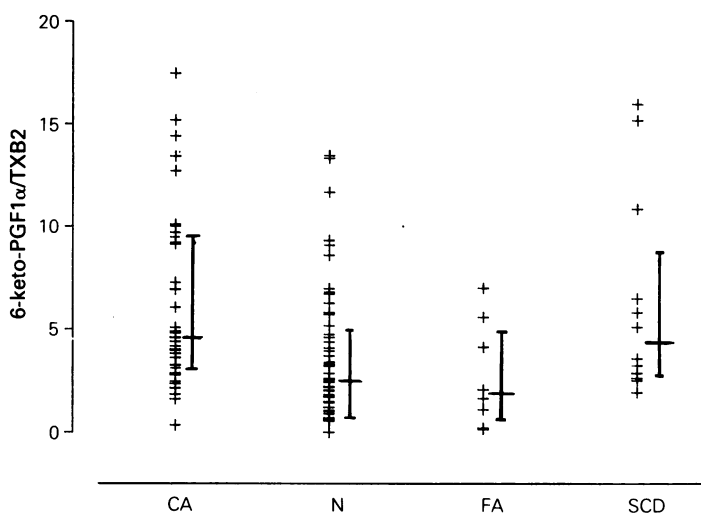


Figure 1 Individual 6-keto-PGF_{1α}/TXB₂ values in the pathological groups: CA (n=37; carcinomata), N (n=51; normal breast tissue), FA (n=8; fibroadenomata), SCD (n=12; sclerocystic disease). Medians and semiquartiles are indicated.

Table II PG-levels in relation to the histological type

Type	n	ng 6-keto-PGF _{1α} mg ⁻¹ protein	ng TXB ₂ mg ⁻¹ protein	6-keto-PGF _{1α} /TXB ₂
Infiltrating ductal carcinoma	23	4.4 (2.3–16.6) ^a	1.0 (0.3–3.4) ^a	4.6 (3.3–9.6) ^a
Lobular	2	5.1–8.1	0.4–0.5	12.8–15.2
Comedo	4	0.4–2.2 ^b	0.1–0.7 ^b	0.6–8.33 ^b
Medullary	1	1.4	0.4	4.0
Mucoid	1	7.7	3.6	2.1

^aMedian (semiquartiles); ^bLimit values

groups showed similar 6-keto-PGF_{1 α} , TXB₂ and 6-keto-PGF_{1 α} /TXB₂ levels (Table III).

Lymph node metastasis and lymphatic vessel permeation

The median amount of 6-keto-PGF_{1 α} was slightly higher in tumour extracts from patients without lymph node metastasis ($P=0.23$). Median TXB₂ amounts and the 6-keto-PGF_{1 α} /TXB₂ ratio were comparable.

The groups without lymphatic vessel permeation had more 6-keto-PGF_{1 α} ($P=0.04$), but the TXB₂ values and the 6-keto-PGF_{1 α} /TXB₂ ratio were similar (Table III).

Size and density of nuclei of carcinoma cells

Tumours with moderate density had significantly less 6-keto-PGF_{1 α} ($P=0.04$) and TXB₂ ($P=0.01$) than those with low or high density. Higher 6-keto-PGF_{1 α} /TXB₂ ratios were found in moderate and high density groups ($P=0.01$; Table III).

Carcinomata with large nuclei tended to yield less TXB₂ than those having moderate size nuclei ($P=0.18$; Table III).

Mitotic index

There was at most a weak correlation between the number of mitoses per high power field (HPF) and the 6-keto-PGF_{1 α} or TXB₂ levels ($r=-0.167$, $P=0.18$ and -0.212 , $P=0.13$ respectively), but a positive correlation occurred with the 6-keto-PGF_{1 α} /TXB₂ ratio ($r=0.33$, $P=0.04$).

Nuclear and cellular polymorphism and the nuclear cytoplasmic ratio

Tumours with a low nuclear and cellular polymorphism showed at most a weak tendency to higher 6-keto-PGF_{1 α} levels ($P=0.19$) no other relationships were seen.

Host cell reaction, necrosis and mast cells

Host cell reaction, necrosis and presence of mast cells did not correlate with the amounts of extracted prostanoids.

Elastosis, fibrosis and infiltration

No significant tendencies were observed between PG-levels and elastosis or fibrosis. Infiltration was inversely related to the 6-keto-PGF_{1 α} /TXB₂ ratio ($P=0.03$), but no significant differences were seen for the 6-keto-PGF_{1 α} or TXB₂ (Table III).

Blood vessel density

Presence of blood vessels did not correlate with prostanoid yields in the tumour biopsies. the 6-keto-PGF_{1 α} /TXB₂ ratio even tended to be lower when more blood vessels were present ($P=0.18$; Table III).

Age

No correlations between prostanoids and age were seen: 6-keto-PGF_{1 α} , $r=-0.023$ ($P=0.45$), TXB₂, $r=0.157$ ($P=0.20$), 6-keto-PGF_{1 α} /TXB₂, $r=0.179$ ($P=0.20$).

Cumulation of variables and follow up

Tumour size showed little or no relationship to tissue prostanoids: 6-keto-PGF_{1 α} , $r=-0.182$ ($P=0.16$), TXB₂, $r=-0.126$ ($P=0.41$), 6-keto-PGF_{1 α} /TXB₂, $r=-0.038$ ($P=0.42$). Tumours with strong metastatic potential (<2 cm and lymph node metastasis, $n=5$) had a median 6-keto-PGF_{1 α} /TXB₂ ratio of 3.8 (2.0–9.4), which was higher ($P<0.05$) than with tumours having a relatively weak metastatic potential (>2 cm and no lymph node metastasis, $n=7$; 2.8 (0.5–4.6)).

A preliminary analysis of 19 patients with a follow-up of 14 months revealed metastases in 5 patients. These 5 patients had a median 6-keto-PGF_{1 α} /TXB₂ ratio of 9.2 (4.0–15.3) which was higher than the ratio of nonmetastatic patients 4.8 (1.6–13.5) ($P=0.04$).

Discussion

Little is known about the role of PGI₂ and TX in human malignant tumours, and only 2 studies on extracted breast tissues have been published.

Karmali *et al.* (1983) measured 6-keto-PGF_{1 α} and TXB₂ in 24 breast tumours and expressed their results as log ng g⁻¹ wet weight. They obtained more TXB₂ from large tumours and those with lymph node metastasis. These results were interpreted as supporting the findings of Honn (1981) that a higher TXA₂/PGI₂ ratio has a worse prognosis in terms of metastasis.

However, Karmali *et al.* (1983) did not give any data about the 6-keto-PGF_{1 α} /TXB₂ ratio which is important in the regulation of blood platelet aggregation. Furthermore, the size of the tumour is not necessarily an indication of the metastatic potential. In addition, Karmali *et al.* (1983) observed no correlation between TXB₂ levels and metastasis. Aitokallio-Tallberg *et al.* (1985) studied the *in vitro* production of 6-keto-PGF_{1 α} and TXB₂ by 23 breast tumour tissues, but as their investigations were directed towards steroid receptor status

Table III Prostanoid levels and anatomopathological variables

Variable (n)	ng 6-keto-PGF _{1α} mg ⁻¹ prot. median (limit values)		ng TXB ₂ mg ⁻¹ prot. median (limit values)		6-keto-PGF _{1α} TXB ₂ median (limit values)		P
		P		P			
Differentiation							
no diff. (15)	5.0 (0.6–27.4)	0.44	0.5 (0.1–17.0)	0.71	4.3 (0.3–17.5)	0.68	
diff. (16)	4.4 (0.9–62.9)		0.8 (0.1–12.8)		5.0 (2.4–14.4)		
Lymph node metastasis							
positive (11)	3.9 (0.7–45.3)	0.23	0.5 (0.3–6.5)	0.70	4.8 (2.8–10.0)	0.89	
negative (15)	5.4 (0.5–62.9)		0.7 (0.1–12.8)		4.3 (1.8–9.2)		
Lymphatic vessel permeation							
positive (23)	4.0 (0.6–62.9)	0.04 ^a	0.5 (0.1–17.0)	0.17	4.7 (0.3–17.5)	0.53	
negative (8)	9.3 (4.3–29.9)		2.1 (0.3–9.6)		5.2 (3.1–15.2)		
Density of nuclei of carcinoma cells							
low (8)	12.7 (1.4–62.9)	0.04 ^a	3.5 (0.4–12.8)	0.01 ^a	2.9 (2.1–4.6)	0.01 ^a	
moderate (16)	4.0 (0.6–16.7)		0.4 (0.1–2.7)		6.1 (0.3–17.5)		
high (7)	7.7 (2.2–27.4)		0.8 (0.3–17.0)		7.0 (1.6–14.4)		
Size of nuclei of carcinoma cells							
small (3)	8.1 (0.6–14.3)	0.44	0.5 (0.2–1.8)	0.18	7.7 (0.3–15.2)	0.34	
moderate (23)	5.0 (0.9–27.4)		0.8 (0.1–17.0)		4.4 (1.6–17.5)		
large (5)	4.0 (1.2–8.3)		0.4 (0.3–0.9)		9.7 (4.6–13.4)		
Nuclear and cellular polymorphism							
low (8)	11.2 (0.6–62.9)	0.19	1.2 (0.2–17.0)	0.44	5.1 (0.3–15.2)	0.98	
moderate (15)	4.4 (0.9–29.9)		0.8 (0.1–11.2)		4.4 (2.4–17.5)		
high (8)	3.1 (1.2–8.3)		0.4 (0.3–3.6)		4.9 (2.1–13.4)		
Nuclear cytoplasmic ratio							
low (4)	5.0 (1.2–16.6)	0.93	1.6 (0.3–3.6)	0.74	4.9 (2.1–6.1)	0.77	
moderate (20)	4.3 (0.9–62.9)		0.7 (0.1–17.0)		4.2 (1.6–17.5)		
high (6)	6.6 (0.6–24.6)		0.4 (0.2–5.3)		5.1 (0.3–15.2)		
Host cell reaction							
low (19)	5.0 (0.6–62.9)	0.59	0.8 (0.1–12.8)	0.39	4.0 (0.3–17.5)	0.67	
moderate (11)	4.2 (1.0–27.4)		0.5 (0.2–17.0)		5.1 (1.6–13.4)		
Mast cells							
positive (11)	4.1 (1.2–62.9)	0.56	0.7 (0.3–12.8)	0.77	4.6 (2.1–13.4)	0.34	
negative (20)	5.4 (0.6–29.9)		0.6 (0.1–17.0)		5.1 (0.3–17.5)		
Necrosis							
positive (15)	4.0 (1.2–26.5)	0.21	0.5 (0.1–11.2)	0.59	4.8 (2.1–17.5)	0.97	
negative (16)	7.9 (0.6–62.9)		0.8 (0.1–17.0)		4.4 (0.3–15.2)		
Elastosis							
positive (21)	5.4 (1.4–62.9)	0.32	0.9 (0.2–12.8)	0.57	5.1 (0.3–17.5)	0.13	
negative (10)	4.3 (0.6–27.4)		0.6 (0.1–17.0)		3.2 (2.4–13.4)		
Fibrosis							
negative (2)	10.4–15.0	0.52	2.9–3.4	0.40	3.6–4.4	0.55	
low (6)	5.9 (0.6–8.3)		0.7 (0.3–1.8)		9.4 (0.3–15.2)		
moderate (16)	4.4 (1.2–62.9)		0.7 (0.1–17.0)		4.6 (1.6–17.5)		
high (7)	4.2 (0.9–29.9)		0.5 (0.1–9.6)		8.1 (2.1–10.1)		
Infiltration							
low (3)	10.4 (1.7–15.0)	0.92	2.9 (0.4–3.4)	0.49	4.4 (3.6–4.6)	0.03 [*]	
moderate (14)	4.4 (0.9–26.5)		0.5 (0.1–11.2)		8.0 (2.4–17.5)		
high (14)	5.0 (0.6–62.9)		1.0 (0.2–17.0)		3.2 (0.3–10.0)		
Blood vessel density							
low (14)	5.3 (0.9–29.9)	0.49	0.8 (0.1–9.6)	0.38	8.0 (2.1–17.5)	0.18	
moderate (8)	6.0 (1.2–27.4)		0.9 (0.3–17.0)		3.7 (1.6–15.2)		
high (7)	2.2 (0.6–62.9)		0.4 (0.2–12.8)		4.8 (0.3–13.4)		

^aSignificantly different

they did not include control tissue superfusion in their protocol. The prostanoid yields were similar from metastasized and non-metastasized cancers (follow-up of at least 3 years).

We studied the PG-levels in the tumours at the time of resection. Treatment with acetone at -70°C inactivated the tumour enzymes and stopped the conversion of arachidonic acid to prostaglandins (Vergote *et al.*, 1985). Prostanoids were measured in the tumour extracts, but the recoveries were about 100%, we can consider the RIA-results as reflecting the prostanoid production at the time of resection. Since the surgical manipulation might induce PGI_2 and TXA_2 production, we also analyzed normal breast tissue taken from the neighbourhood of the cancer. The tumour specimen was always taken first, so that artefactual prostanoid might be higher in the normal tissue because of the longer trauma or lower because of prostanoid metabolism. For example 6,15-diketo 13,14-dihydro $\text{PGF}_{1\alpha}$ is formed from PGI_2 in plasma and vascular tissue (Peskar *et al.*, 1980).

However, continued enzymatic conversion would be blocked by the acetone treatment.

Non-malignant sources of the tumour prostanoids are the host cells, but no correlation was found between the amounts of the prostanoids and the host cell reaction or the presence of mast cells. Furthermore, the 6-keto- $\text{PGF}_{1\alpha}$ and TXB_2 tissue levels did not correlate with the density of blood vessels.

We preferred to express the prostanoid yields in ng mg^{-1} protein, rather than to use ng g^{-1} wet or dry weight, in an attempt to reduce the variation of the results. One of the main conclusions from our study is that carcinomata had higher median 6-keto- $\text{PGF}_{1\alpha}$ and TXB_2 levels than N, FA or SCD, but even so, the ranges overlapped considerably.

As can be seen from Table III, most of the differences were not significant, but a high metastatic potential and mitotic index correlated with higher 6-keto- $\text{PGF}_{1\alpha}/\text{TXB}_2$ ratios. Furthermore, with 42 comparisons, 2 would be expected by chance to show P values <0.05 when no difference really exists.

No firm conclusions can yet be made, although follow-up over 14 months revealed a higher 6-keto- $\text{PGF}_{1\alpha}/\text{TXB}_2$ ratio in patients with metastasis.

These data should be interpreted very cautiously, as the number of patients evaluated is low and the follow-up period is limited. Nevertheless, the histopathology results do not support the suggestion of Karmali *et al.* (1983), for a protective action of prostacyclin against metastasis in human breast cancer, or the hypothesis of Honn (1981) that TXA_2 promotes metastasis.

A common observation by several authors is the greater prostaglandin production by malignant breast tumours compared with normal breast tissues. This conclusion is independent of the prostaglandin measured or the ways of performing the incubations, extractions or measurements (Bennett *et al.*, 1975; Kibbey *et al.*, 1979; Greaves *et al.*, 1980; Rolland *et al.*, 1980; Malachi *et al.*, 1981; Campbell *et al.*, 1983; Karmali *et al.*, 1983). The relationship between high prostaglandin levels and malignancy of breast tumours could lead to the systematic incorporation of cyclo-oxygenase inhibitors (NSAID) in cancer therapy. However, when histopathological prognostic variables of malignant tissues are examined, the prognosis seems to be better when the $\text{PGF}_{2\alpha}$ levels were higher (Vergote *et al.*, 1985). Although this was not found for 6-keto- $\text{PGF}_{1\alpha}$ and TXB_2 in our present study, the mitotic index, prognosis and the first follow-up results indicate that tumour 6-keto- $\text{PGF}_{1\alpha}$ at the time of surgery was often higher in patients with bad prognosis or metastasis. Using NSAID could theoretically worsen the prognosis, by depressing the 6-keto- $\text{PGF}_{1\alpha}$ levels, but we do not know what effect this would have on the 6-keto- $\text{PGF}_{1\alpha}/\text{TXB}_2$ ratio. Therefore we can neither support nor recommend the use of NSAID in the treatment of breast cancer, particularly since other actions of prostaglandins, e.g. in immunological functions, may also be affected.

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