

Clinical Study

Preoperative Mean Platelet Volume and Platelet Distribution Width Predict Postoperative Sepsis in Patients with Colorectal Cancer

Xue-ting Li,¹ Zibo Yan,² Rui-tao Wang ^{3,4} and Kai-jiang Yu ^{4,5}

¹Department of Intensive Care Unit, Harbin Medical University Cancer Hospital, Harbin Medical University, Harbin, Heilongjiang 150081, China

²Department of Planned Immunization, Heilongjiang Provincial Center for Disease Control and Prevention, Harbin, Heilongjiang 150030, China

³Department of Internal Medicine, Harbin Medical University Cancer Hospital, Harbin Medical University, Harbin, Heilongjiang 150081, China

⁴Institute of Intensive Care Unit, Heilongjiang Academy of Medical Science, Harbin, Heilongjiang 150081, China

⁵Department of Intensive Care Unit, The First Affiliated Hospital of Harbin Medical University, Harbin Medical University, Harbin, Heilongjiang 150081, China

Correspondence should be addressed to Rui-tao Wang; ruitaowang@126.com and Kai-jiang Yu; kaijiangyu2019@gmail.com

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Purpose. Mean platelet volume (MPV) and platelet distribution width (PDW) have been used to reflect the platelet activity in clinics. We assessed initial serum MPV and PDW levels in colorectal cancer (CRC) patients, in predicting the development of sepsis in CRC patients postoperatively. **Patients and Methods.** This study included 220 patients diagnosed with CRC. 55 patients were stratified to one group that developed sepsis postoperatively, and 165 patients were stratified to the other group that did not develop sepsis postoperatively. Clinical and laboratory characteristics were collected 3 days before the operation. **Results.** MPV ($p < 0.001$) was significantly higher and PDW ($p < 0.001$) was significantly lower in the sepsis group than in the nonsepsis group. Either MPV or PDW is independently associated with ICU mortality in sepsis patients with CRC. MPV is independently associated with 14-day, 28-day, and 90-day mortality and PDW is independently associated with 90-day mortality in patients with CRC. The prevalence of sepsis increased as MPV tertiles increased ($p < 0.001$), and the prevalence of sepsis increased as PDW tertiles decreased ($p < 0.001$). **Conclusions.** Serum MPV and PDW levels between CRC patients with/without sepsis postoperatively are significantly different. The initial serum MPV or PDW levels can potentially serve as a predictor of sepsis in CRC patients postoperatively.

1. Introduction

Cancer is the leading cause of high mortality worldwide and causes heavy socioeconomic impact [1–3]. Among cancer patients, death due to sepsis-related multiorgan failure is more frequent than death due to cancer itself [4–7]. Colorectal cancer (CRC) is one of the commonest malignant diseases in China and is a frequent cause of cancer-related death [8].

Sepsis is identified as life-threatening organ dysfunction caused by a dysregulated host response to infection [9]. The

multiple organ failure caused by sepsis is the most lethal cause of surgery in ICU [10]. A study reported the mortality rate for cancer patients with septic shock admitted to the critical care unit (ICU) is nearly 54% [11].

It is well known that platelets are related to the hemostasis and coagulation function [12]. In recent years, more and more studies have confirmed that activated platelets are involved in the development and metastasis of tumors [13, 14]. Interaction between platelets and the tumor cells is not depending on the platelet quantity but on the volume and size because larger platelets have more granules and

receptors [15]. Platelet volume indices have been used to reflect the platelet activity in clinics, including mean platelet volume (MPV) and platelet distribution width (PDW) [16]. MPV is found as an indicator of activated platelets and associated with many cancers [17–19]. PDW, another platelet parameter, indicates variation in platelet size and differentially diagnoses thrombocytopenia [16].

Even though more and more precaution was paid to hospitalized patients, sepsis incidence in cancer patients after surgery still takes up a significant burden of illness [20]. Patients undergoing surgery for CRC are at particular risk of sepsis because of underlying malignancy, being immunocompromised associated with cancer management and the complexity of surgical procedures performed [21]. Therefore, it is urgent to find out biomarkers which can predict the occurrence of postoperative sepsis in CRC patients. This study aimed to address the role of MPV and PDW as biomarkers in predicting the development of postoperative sepsis in CRC patients.

2. Patients and Methods

2.1. Study Design. This was a prospective study. The study protocol was approved by the Institutional Review Board of the Harbin Medical University Cancer Hospital. All patients signed informed consent.

2.2. Patients and Data Collection. CRC patients were admitted to the Harbin Medical University Cancer Hospital for surgery between January 1, 2015, and December 30, 2017. All patients underwent complete surgical resection, and the pathologic diagnoses were histologically confirmed by two experienced pathologists. There were 55 sepsis cases after surgery (mean age 63.4 ± 9.7 years, range 57–86 years). Using incidence density sampling, we matched the 55 sepsis patients with 165 controls by age, sex, and BMI (mean age 63.4 ± 9.0 years, range 46–78 years). All sepsis patients fulfilled the criteria of sepsis (SEPSIS-3) [22]. Cases were included if they met the following criteria: (1) age > 18 years; (2) patients who underwent complete surgical resection and diagnosis of CRC cancers being confirmed by histology; (3) diagnosis of sepsis after surgery in the intensive care unit (ICU); and (4) patients who did not receive preoperative chemotherapy or radiation therapy. Exclusion criteria included HIV infection, neutropenia (<500 neutrophils/ mm^3), and the medical treatment with steroids. The Acute Physiology and Chronic Health Evaluation (APACHE) II scores and the Sequential Organ Failure Assessment (SOFA) scores were determined.

2.3. Clinical Examination and Biochemical Measurements. All subjects underwent physical examination. Body mass index (BMI) was calculated as the ratio of weight (kg) to height squared (m^2). Clinical data including medical history and medication use were recorded for each subject. Venous blood samples were collected 3 days before surgery. White blood cell (WBC), hemoglobin, and platelet indices were measured by an auto analyzer (Sysmex XE-2100, Kobe,

Japan). The normal ranges of MPV and PDW in our hospital are 7–11 fL and 11–17%, respectively. The inter- and intra-assay coefficients of variation (CVs) of all these assays were below 5%.

2.4. Outcomes. The primary outcome of this study was to assess the predicting role of MPV and PDW in CRC patients who will be subjected to sepsis. Secondary outcomes were to certify if MPV or PDW can be a biomarker to forecast the ICU mortality or 28-day and 90-day mortality in sepsis patients with CRC.

2.5. Statistical Analysis. All statistical analyses were performed using SPSS Statistics version 22.0 (SPSS Inc., Chicago, IL, USA). The descriptive statistics are presented as means \pm SD or medians (interquartile range) for continuous variables and percentages of the number for categorical variables. When baseline characteristics between two groups were compared, normally distributed continuous variables were compared with Student's *t*-test and skewed-distributed with the Mann–Whitney *U* test. The chi-square test was used for categorical variables. The odds ratios (ORs) and 95% confidence intervals (95% CIs) for sepsis were calculated using conditional logistic regression analysis. $p < 0.05$ was considered to indicate a statistically significant difference.

3. Results

The study cohort included a nonsepsis group ($n = 165$) and a sepsis group ($n = 55$). The nonsepsis group included 99 men and 66 women. The sepsis group included 33 men and 22 women. Their characteristics are shown in Table 1; the mean age in the sepsis group is 63.4 ± 9.7 years, compared with the mean age of 63.4 ± 9.0 years in the nonsepsis group. There was no significant difference between the sepsis group and the nonsepsis group with regard to age, gender, body mass index (BMI), smoking, drinking, WBC, platelet count, creatinine, ALP, fibrinogen, CEA, and comorbidities. It was observed that hemoglobin, albumin, and PDW were significantly decreased in the sepsis group than in the nonsepsis group. However, MPV, LDH, APACHE II score, SOFA score, length of ICU stay, and ICU mortality were significantly increased in the sepsis group than in the nonsepsis group.

Conditional logistic regression analysis showed that albumin (HR, 0.883; CI, 0.829–0.940; $p < 0.001$), MPV (HR, 2.125; CI, 1.677–2.694; $p < 0.001$), and PDW (HR, 0.756; CI, 0.645–0.886; $p < 0.001$) were independently associated with postoperative sepsis in CRC patients, as shown in Table 2. In addition, hemoglobin (HR, 1.029; CI, 1.002–1.057; $p = 0.038$), albumin (HR, 0.884; CI, 0.805–0.970; $p = 0.01$), MPV (HR, 1.841; CI, 1.369–2.476; $p < 0.001$), and PDW (HR, 0.669; CI, 0.537–0.833; $p < 0.001$) were independently associated with ICU mortality in sepsis patients with CRC, as shown in Table 3. Moreover, it is shown in Table 4 that albumin (HR, 0.842; CI, 0.772–0.917; $p < 0.001$) and MPV (HR, 0.372; CI, 1.042–1.807; $p = 0.024$) were independently associated with 28-day mortality. Albumin (HR, 0.867; CI,

TABLE 1: Baseline characteristics of CRC patients stratified by development of sepsis status after operation.

| Variables | Sepsis, <i>n</i> = 55 | Nonsepsis, <i>n</i> = 165 | <i>p</i> value |
|--------------------------------------|-----------------------|---------------------------|----------------|
| Age (years) | 63.4 ± 9.7 | 63.4 ± 9.0 | 0.993 |
| Gender (male, %) | 33 (60.0) | 99 (60.0) | 1.000 |
| BMI (kg/m ²) | 23.5 ± 3.1 | 23.5 ± 3.3 | 0.927 |
| Smoker (%) | 9 (16.4) | 33 (20.0) | 0.552 |
| Drinking (%) | 8 (14.5) | 35 (21.2) | 0.280 |
| WBC (×10 ⁹ /L) | 8.58 ± 6.08 | 7.15 ± 2.63 | 0.093 |
| Hemoglobin (g/L) | 113.4 ± 26.7 | 130.6 ± 24.6 | <0.001 |
| Platelet count (×10 ⁹ /L) | 272.5 ± 131.8 | 272.5 ± 84.4 | 0.999 |
| MPV (fL) | 9.6 ± 1.3 | 8.4 ± 1.7 | <0.001 |
| PDW (%) | 14.6 ± 2.8 | 17.2 ± 1.1 | <0.001 |
| Creatinine (umol/L) | 84.1 ± 37.4 | 88.0 ± 20.9 | 0.460 |
| Albumin (g/L) | 35.0 ± 6.9 | 44.7 ± 6.8 | <0.001 |
| LDH (U/L) | 166 (146–193) | 145 (125.5–179) | 0.003 |
| ALP (U/L) | 83.0 (68.0–108.0) | 87.0 (75.0–109.0) | 0.196 |
| Fibrinogen (g/L) | 3.44 ± 1.02 | 3.69 ± 1.03 | 0.126 |
| CEA (ng/ml) | 5.16 (2.19–11.64) | 4.72 (2.26–15.78) | 0.779 |
| Comorbidities, <i>n</i> (%) | | | |
| Coronary artery disease | 10 (18.2) | 33 (20.0) | 0.768 |
| COPD | 0 | 0 | |
| Hypertension | 12 (21.8) | 27 (16.4) | 0.359 |
| Diabetes mellitus | 10 (18.2) | 31 (18.8) | 0.920 |
| APACHE II score | 17 (12–22) | 13 (11–14) | <0.001 |
| SOFA score | 9 (6–10) | 6 (6–8) | 0.005 |
| Length of ICU stay, median (IQR) | 4 (1–7) | 1 (1–1) | <0.001 |
| ICU mortality (%) | 15 (27.3) | 0 (0) | <0.001 |
| Tumor location | | | 0.392 |
| Colon | 31 (56.4) | 82 (49.7) | |
| Rectum | 24 (43.6) | 83 (50.3) | |
| Tumor size (cm) | | | 0.576 |
| <5.0 | 32 (58.2) | 103 (62.4) | |
| ≥5.0 | 23 (41.8) | 62 (37.6) | |
| Histology differentiation | | | 0.322 |
| Well/moderately | 47 (85.5) | 131 (79.4) | |
| Poorly | 8 (14.5) | 34 (20.6) | |

Data are expressed as mean (SD) or percentage. SD, standard deviation; BMI, body mass index; WBC, white blood cell; BUN, blood urea nitrogen; CEA, carcinoembryonic antigen; FPG, fasting plasma glucose; LDH, lactate dehydrogenase; ALP, alkaline phosphatase; ICU, intensive care unit; SOFA, Sequential Organ Failure Assessment; APACHE, Acute Physiology and Chronic Health Evaluation; IQR, 25% and 75% interquartile range; COPD, chronic obstructive pulmonary disease.

TABLE 2: Conditional logistic regression analysis of variables independently associated with postoperative sepsis in CRC patients.

| | β | HR (95% CI) | <i>p</i> value |
|---------------------------|---------|---------------------|----------------|
| WBC (×10 ⁹ /L) | 0.102 | 1.107 (0.979–1.252) | 0.104 |
| Hemoglobin (g/L) | 0.006 | 1.006 (0.991–1.022) | 0.420 |
| LDH (U/L) | 0.000 | 1.000 (0.999–1.002) | 0.573 |
| Albumin (g/L) | -0.125 | 0.883 (0.829–0.940) | <0.001 |
| MPV (fL) | 0.754 | 2.125 (1.677–2.694) | <0.001 |
| PDW (%) | -0.280 | 0.756 (0.645–0.886) | 0.001 |

HR, hazard ratio; CI, confidence interval; WBC, white blood cell; LDH, lactate dehydrogenase.

0.809–0.929; *p* < 0.001), MPV (HR, 1.626; CI, 1.309–2.019; *p* < 0.001), and PDW (HR, 0.830; CI, 0.710–0.970; *p* = 0.019) were independently associated with 90-day mortality in patients with CRC.

The association between MPV levels and the prevalence rate of sepsis (%) is shown in Figure 1. All the 220 participants (nonsepsis group + sepsis group) were stratified into tertiles according to their MPV levels. Tertile 1 (T1) was

TABLE 3: Conditional logistic regression analysis of variables independently associated with ICU mortality in patients with colorectal cancer.

| | β | HR (95% CI) | <i>p</i> value |
|---------------------------|---------|---------------------|----------------|
| WBC (×10 ⁹ /L) | -0.165 | 0.848 (0.646–1.113) | 0.234 |
| Hemoglobin (g/L) | 0.028 | 1.029 (1.002–1.057) | 0.038 |
| LDH (U/L) | 0.001 | 1.001 (0.999–1.002) | 0.409 |
| Albumin (g/L) | -0.124 | 0.884 (0.805–0.970) | 0.010 |
| MPV (fL) | 0.610 | 1.841 (1.369–2.476) | <0.001 |
| PDW (%) | -0.402 | 0.669 (0.537–0.833) | <0.001 |

HR, hazard ratio; CI, confidence interval; WBC, white blood cell; LDH, lactate dehydrogenase.

MPV ≤ 8.0 fL, tertile 2 (T2) was MPV 8.1–9.0 fL, and tertile 3 (T3) was MPV ≥ 9.1 fL. The prevalence rate of sepsis in T1, T2, and T3 was 8.75%, 18.31%, and 50.72%, respectively.

The association between PDW levels and the prevalence rate of sepsis (%) is shown in Figure 2. Participants were stratified into tertiles according to their PDW levels. Tertile 1 (T1) was PDW ≤ 16.5%, tertile 2 (T2) was PDW 16.6–17.4%,

TABLE 4: Conditional logistic regression analysis of variables independently associated with 14-day mortality, 28-day mortality, and 90-day mortality in patients with colorectal cancer.

| | β | HR (95% CI) | <i>p</i> value |
|-------------------------|---------|---------------------|----------------|
| 14-day mortality | | | |
| WBC ($\times 10^9/L$) | 0.064 | 1.066 (0.958–1.186) | 0.241 |
| Hemoglobin (g/L) | -0.031 | 0.969 (0.942–0.997) | 0.032 |
| LDH (U/L) | 0.002 | 1.002 (1.000–1.004) | 0.045 |
| Albumin (g/L) | -0.185 | 0.831 (0.750–0.922) | <0.001 |
| MPV (fL) | 0.467 | 1.595 (1.151–2.211) | 0.005 |
| PDW (%) | 0.109 | 1.115 (0.890–1.397) | 0.344 |
| 28-day mortality | | | |
| WBC ($\times 10^9/L$) | 0.052 | 1.053 (0.954–1.163) | 0.307 |
| Hemoglobin (g/L) | -0.018 | 0.982 (0.960–1.005) | 0.127 |
| LDH (U/L) | 0.001 | 1.001 (1.000–1.003) | 0.059 |
| Albumin (g/L) | -0.172 | 0.842 (0.772–0.917) | <0.001 |
| MPV (fL) | 0.316 | 1.372 (1.042–1.807) | 0.024 |
| PDW (%) | 0.128 | 1.136 (0.923–1.399) | 0.228 |
| 90-day mortality | | | |
| WBC ($\times 10^9/L$) | 0.103 | 1.109 (0.985–1.249) | 0.088 |
| Hemoglobin (g/L) | 0.011 | 1.011 (0.993–1.029) | 0.226 |
| LDH (U/L) | 0.001 | 1.001 (0.999–1.002) | 0.307 |
| Albumin (g/L) | -0.143 | 0.867 (0.809–0.929) | <0.001 |
| MPV (fL) | 0.486 | 1.626 (1.309–2.019) | <0.001 |
| PDW (%) | -0.187 | 0.830 (0.710–0.970) | 0.019 |

HR, hazard ratio; CI, confidence interval; WBC, white blood cell; LDH, lactate dehydrogenase.

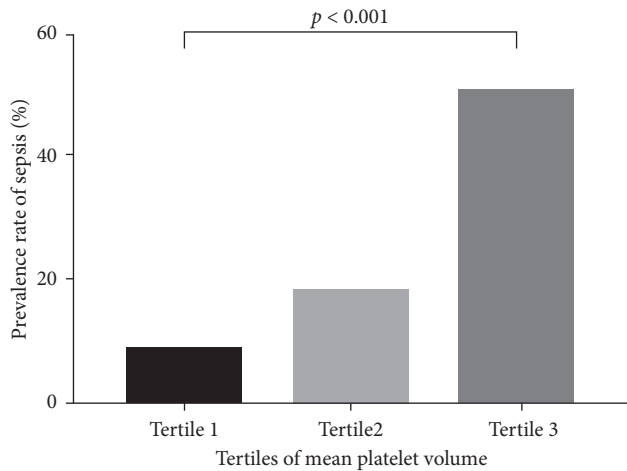


FIGURE 1: Association between MPV levels and the prevalence rate of sepsis (%). Participants were stratified into tertiles according to their MPV levels. Tertile 1 (T1) was $MPV \leq 8.0$ fL, tertile 2 (T2) was $MPV 8.1-9.0$ fL, and tertile 3 (T3) was $MPV \geq 9.1$ fL. The prevalence rate of sepsis in T1, T2, and T3 was 8.75%, 18.31%, and 50.72%, respectively.

and tertile 3 (T3) was $PDW \geq 17.5\%$. The prevalence rate of sepsis in T1, T2, and T3 was 50.00%, 15.79%, and 8.57%, respectively.

4. Discussion

In this study, we found that elevated MPV levels or reduced PDW levels before surgery are associated with postoperative sepsis events. In addition, MPV or PDW was independently

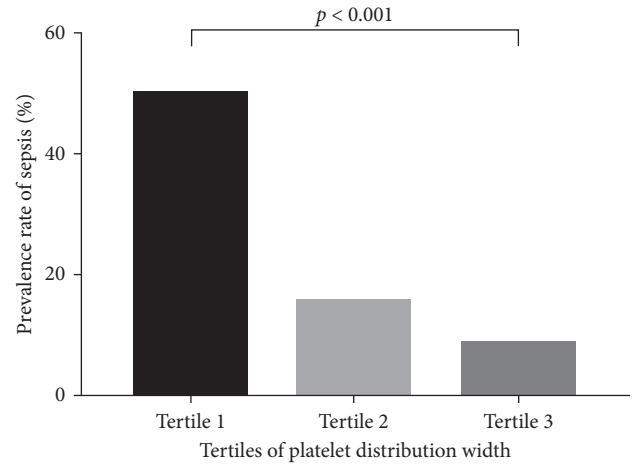


FIGURE 2: Association between PDW levels and the prevalence rate of sepsis (%). Participants were stratified into tertiles according to their PDW levels. Tertile 1 (T1) was $PDW \leq 16.5\%$, tertile 2 (T2) was $PDW 16.6-17.4\%$, and tertile 3 (T3) was $PDW \geq 17.5\%$. The prevalence rate of sepsis in T1, T2, and T3 was 50.00%, 15.79%, and 8.57%, respectively.

associated with ICU mortality in CRC patients. Moreover, MPV was independently associated with 14-day mortality, 28-day mortality, and 90-day mortality in CRC patients; meanwhile, PDW was independently associated with 90-day mortality in CRC patients.

The mechanisms to explain the association between preoperative MPV or PDW and postoperative sepsis events remain unclear. The MPV is on behalf of an average size of platelets. Increased MPV levels are involved in the mechanism of platelet activation [23]. PDW reflected the heterogeneity in platelet size. A high value of PDW suggests a large range of platelet size due to swelling, destruction, and immaturity [24]. PDW and MPV are common indices which are associated with platelet activation and function [25]. In our study, we observed the patients who developed postoperative sepsis have higher preoperative MPV levels but lower PDW levels. This is maybe due to the cancer-associated inflammation. Studies have shown that the original chronic inflammatory disease directly causes at least 20% of cancers, especially in colorectal cancer [26, 27]. More cytokines are generated during the inflammation course in cancer, such as IL-6 and IL-11, and highly upregulated in many cancers [28]. In addition, previous research confirmed that tumors could promote platelet production and activation through IL-6 signaling [29]. TLR2 is a toll-like receptor protein which is expressed on megakaryocytes and platelets and is involved in innate immune responses. Interactively, inflammation mediated platelet activation through cytokines [30].

Consistent with Li et al.'s study, elevated MPV levels in CRC patients predicted the worse outcomes [31]. One clinical study uncovered a phenomenon where there were increased MPV levels and the thrombocyte consumption in infections [32]. Cekmez et al. showed that the MPV levels have positive relation with the incidence of early sepsis [33]. A study of neonatal sepsis confirmed that patients with neonatal sepsis

had high levels of MPV [34]. Orak et al. found high MPV levels were significant in terms of prognosis and mortality in sepsis patients [32]. In our study, the elevated MPV levels were associated with ICU mortality in CRC patients similar to those reported in other studies. Furthermore, we had firstly confirmed the predictive effect of preoperative MPV levels on postoperative patients with sepsis. However, Zhu et al. reported the change in PDW in colorectal cancer patients was significantly higher versus that in both colorectal adenoma patients and healthy participants [35]. Our conflicting data are maybe due to sepsis, small sample sizes, failure to exclude confounding factors, different tumor types, and selected populations. Therefore, further research is needed for PDW.

Our study has a few limitations: Firstly, our study is single-centered, and there were only 220 patients included. In addition, this study lacks mechanistic data to explain the role of MPV and PDW in the development of sepsis. Moreover, we only assessed the initial MPV and PDW levels, and we had no serial data. Time-dependent changes in MPV and PDW levels would be more valuable.

5. Conclusions

In conclusion, either increased MPV or decreased PDW is an independent predictor of postoperative sepsis after CRC surgery. Our data shed light on the value of MPV and PDW in sepsis prevention.

Data Availability

The clinical data of patients used to support the findings of this study are available from the corresponding author upon request.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

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References

- [1] J. D. Albano, E. Ward, A. Jemal et al., "Cancer mortality in the United States by education level and race," *JNCI: Journal of the National Cancer Institute*, vol. 99, no. 18, pp. 1384–1394, 2007.
- [2] J. Ferlay, D. M. Parkin, and E. Steliarova-Foucher, "Estimates of cancer incidence and mortality in Europe in 2008," *European Journal of Cancer*, vol. 46, no. 4, pp. 765–781, 2010.
- [3] G. H. Lim, K. Y. Chow, and H. P. Lee, "Singapore cancer trends in the last decade," *Singapore Medical Journal*, vol. 53, no. 1, pp. 3–10, 2012.
- [4] G. T. Bird, P. Farquhar-Smith, T. Wigmore, M. Potter, and P. C. Gruber, "Outcomes and prognostic factors in patients with haematological malignancy admitted to a specialist cancer intensive care unit: a 5 yr study," *British Journal of Anaesthesia*, vol. 108, no. 3, pp. 452–459, 2012.
- [5] F. I. Hawari, L. H. Nazer, A. Addassi, D. Rimawi, and K. Jamal, "Predictors of ICU admission in patients with cancer and the related characteristics and outcomes," *Critical Care Medicine*, vol. 44, no. 3, pp. 548–553, 2016.
- [6] V. Allareddy, S. Prakasam, V. Allareddy et al., "Poor oral health linked with increased risk of infectious complications in adults with leukemia," *Journal of the Massachusetts Dental Society*, vol. 64, no. 3, pp. 38–42, 2015.
- [7] N. Obeng-Nkrumah, A.-K. Labi, M. E. Acquah, and E. S. Donkor, "Bloodstream infections in patients with malignancies: implications for antibiotic treatment in a Ghanaian tertiary setting," *BMC Research Notes*, vol. 8, no. 1, p. 742, 2015.
- [8] J. Zhu, Z. Tan, K. Hollis-Hansen, Y. Zhang, C. Yu, and Y. Li, "Epidemiological trends in colorectal cancer in China: an ecological study," *Digestive Diseases and Sciences*, vol. 62, no. 1, pp. 235–243, 2017.
- [9] A. Rhodes, L. E. Evans, W. Alhazzani et al., "Surviving sepsis campaign: international guidelines for management of sepsis and septic shock: 2016," *Critical Care Medicine*, vol. 45, no. 3, pp. 486–552, 2017.
- [10] C. W. Seymour, F. Gesten, H. C. Prescott et al., "Time to treatment and mortality during mandated emergency care for sepsis," *New England Journal of Medicine*, vol. 376, no. 23, pp. 2235–2244, 2017.
- [11] C. J. Regazzoni, C. I. Irrazabal, C. M. Luna, and J. J. Poderoso, "Cancer patients with septic shock: mortality predictors and neutropenia," *Supportive Care in Cancer*, vol. 12, no. 12, pp. 833–839, 2004.
- [12] A. Osman, W. E. Hitzler, and P. Provost, "The platelets' perspective to pathogen reduction technologies," *Platelets*, vol. 29, no. 2, pp. 140–147, 2018.
- [13] N. M. Bambace and C. E. Holmes, "The platelet contribution to cancer progression," *Journal of Thrombosis and Haemostasis*, vol. 9, no. 2, pp. 237–249, 2011.
- [14] H. A. Goubran, J. Stakiw, M. Radosevic, and T. Burnouf, "Platelet-cancer interactions," *Seminars in Thrombosis and Hemostasis*, vol. 40, no. 3, pp. 296–305, 2014.
- [15] J. F. Martin, P. M. W. Bath, and M. L. Burr, "Influence of platelet size on outcome after myocardial infarction," *The Lancet*, vol. 338, no. 8780, pp. 1409–1411, 1991.
- [16] K. Kaito, H. Otsubo, N. Usui et al., "Platelet size deviation width, platelet large cell ratio, and mean platelet volume have sufficient sensitivity and specificity in the diagnosis of immune thrombocytopenia," *British Journal of Haematology*, vol. 128, no. 5, pp. 698–702, 2005.
- [17] Y. Kemal, G. Demirağ, K. Ekiz, and İ. Yücel, "Mean platelet volume could be a useful biomarker for monitoring epithelial ovarian cancer," *Journal of Obstetrics and Gynaecology*, vol. 34, no. 6, pp. 515–518, 2014.
- [18] S. Kilincalp, F. Ekiz, O. Başar et al., "Mean platelet volume could be possible biomarker in early diagnosis and monitoring of gastric cancer," *Platelets*, vol. 25, no. 8, pp. 592–594, 2014.
- [19] Y. Kumagai, S. Gilmour, E. Ota et al., "Estimating the burden of foodborne diseases in Japan," *Bulletin of the World Health Organization*, vol. 93, no. 8, pp. 540–549C, 2015.
- [20] A. Hiong, K. A. Thursky, B. W. Teh, G. M. Haeusler, M. A. Slavin, and L. J. Worth, "Sepsis following cancer surgery: the need for early recognition and standardised clinical care," *Expert Review of Anti-infective Therapy*, vol. 14, no. 4, pp. 425–433, 2016.

- [21] H. Brody, "Colorectal cancer," *Nature*, vol. 521, no. 7551, p. S1, 2015.
- [22] J. A. Russell, T. Lee, J. Singer, J. H. Boyd, and K. R. Walley, "The septic shock 3.0 definition and trials," *Critical Care Medicine*, vol. 45, no. 6, pp. 940–948, 2017.
- [23] J. Yang, X. Zhou, X. Fan et al., "mTORC1 promotes aging-related venous thrombosis in mice via elevation of platelet volume and activation," *Blood*, vol. 128, no. 5, pp. 615–624, 2016.
- [24] Z. J. Ren, P.-W. Ren, B. Yang et al., "Mean platelet volume, platelet distribution width and platelet count in erectile dysfunction: a systematic review and meta-analysis," *Andrologia*, vol. 49, no. 10, Article ID e12777, 2017.
- [25] J. D. Eicher, G. Lettre, and A. D. Johnson, "The genetics of platelet count and volume in humans," *Platelets*, vol. 29, no. 2, pp. 125–130, 2018.
- [26] S. I. Grivennikov, K. Wang, D. Mucida et al., "Adenoma-linked barrier defects and microbial products drive IL-23/IL-17-mediated tumour growth," *Nature*, vol. 491, no. 7423, pp. 254–258, 2012.
- [27] K. Wang, M. K. Kim, G. Di Caro et al., "Interleukin-17 receptor signaling in transformed enterocytes promotes early colorectal tumorigenesis," *Immunity*, vol. 41, no. 6, pp. 1052–1063, 2014.
- [28] K. Taniguchi and M. Karin, "IL-6 and related cytokines as the critical lynchpins between inflammation and cancer," *Seminars in Immunology*, vol. 26, no. 1, pp. 54–74, 2014.
- [29] R. J. Lin, V. Afshar-Kharghan, and A. I. Schafer, "Paraneoplastic thrombocytosis: the secrets of tumor self-promotion," *Blood*, vol. 124, no. 2, pp. 184–187, 2014.
- [30] R. B. Undi, S. Sarvothaman, K. Narasaiah, U. Gutti, and R. K. Gutti, "Toll-like receptor 2 signalling: significance in megakaryocyte development through wnt signalling cross-talk and cytokine induction," *Cytokine*, vol. 83, pp. 245–249, 2016.
- [31] N. Li, Z. Yu, X. Zhang et al., "Elevated mean platelet volume predicts poor prognosis in colorectal cancer," *Scientific Reports*, vol. 7, no. 1, p. 10261, 2017.
- [32] M. Orak, Y. Karakoç, M. Ustundag, Y. Yildirim, M. K. Celen, and C. Güloğlu, "An investigation of the effects of the mean platelet volume, platelet distribution width, platelet/lymphocyte ratio, and platelet counts on mortality in patients with sepsis who applied to the emergency department," *Nigerian Journal of Clinical Practice*, vol. 21, no. 5, pp. 667–671, 2018.
- [33] Y. Cekmez, M. Dizdar Güleçoğlu, C. Özcan, L. Karadeniz, and G. Kiran, "The utility of maternal mean platelet volume levels for early onset neonatal sepsis prediction of term infants," *Ginekologia Polska*, vol. 88, no. 6, pp. 312–314, 2017.
- [34] C. H. Patrick and J. Lazarchick, "The effect of bacteremia on automated platelet measurements in neonates," *American Journal of Clinical Pathology*, vol. 93, no. 3, pp. 391–394, 1990.
- [35] X. Zhu, Y. Cao, P. Lu et al., "Evaluation of platelet indices as diagnostic biomarkers for colorectal cancer," *Scientific Reports*, vol. 8, no. 1, p. 11814, 2018.