

Journal of Cerebral Blood Flow & Metabolism 2019, Vol. 39(5) 834–848 © Author(s) 2017



Article reuse guidelines: sagepub.com/journals-permissions DOI: 10.1177/0271678X17737909 journals.sagepub.com/home/jcbfm



Cerebral metabolism and vascular reactivity during breath-hold and hypoxic challenge in freedivers and healthy controls

Mark B Vestergaard and Henrik BW Larsson 1,2

Abstract

The goal of the present study was to examine the cerebral metabolism and vascular reactivity during extended breathholds (ranging from 2 min 32 s to 7 min 0 s) and during a hypoxic challenge in freedivers and non-diver controls. Magnetic resonance imaging was used to measure the global cerebral blood flow (CBF) and metabolic rate of oxygen (CMRO₂), and magnetic resonance spectroscopy was used to measure the cerebral lactate, glutamate+glutamine, N-acetylaspartate and phosphocreatine+creatine concentrations in the occipital lobe. Fifteen freedivers and seventeen non-diver controls participated. The freedivers showed remarkable increases in CBF (107%) during the breath-holds, compensating for arterial desaturation, and sustained cerebral oxygen delivery (CDO₂). CMRO₂ was unaffected throughout the breathholds. During the hypoxic challenge, the freedivers had larger increases in blood flow in the sagittal sinus than the non-divers, and could sustain normal CDO₂. No differences were found in lactate production, global CBF or CMRO₂. We conclude that the mechanism for sustaining brain function during breath-holding in freedivers involves an extraordinary increase in perfusion, and that freedivers present evidence for higher cerebrovascular reactivity, but not for higher lactate-producing glycolysis during a hypoxic challenge compared to controls.

Keywords

Cerebral autoregulation, cerebral blood flow, lactate, magnetic resonance imaging/spectroscopy, oxygen extraction fraction

Received 12 February 2017; Revised 18 September 2017; Accepted 25 September 2017

Introduction

Freedivers are capable of holding their breath for several minutes. At the 2016 AIDA (International Association for Development of Apnea, https://www.aidainternational.org) world championship, the longest breath-hold (BH) during static apnea (BH with one's face submerged in water without movement) was 9 min 34 s for men, and 7 min 59 s for women.

The brain requires a constant supply of oxygen to prevent neuronal insult. How freedivers maintain sufficient energy metabolism during extended BH and during a hypoxic challenge is investigated in this study. Humans have a number of coping mechanisms during diving that initiate during BH and constitute the so-called diver reflex. The diver reflex decreases the oxygen consumption in non-vital organs and muscles by means of peripheral vasoconstriction and the

redistribution of blood and resources to the vital organs, including the heart, lungs and brain.

The main coping mechanism against cerebral hypoxia or hypercapnia is increasing cerebral blood flow (CBF) from the cerebrovascular reserve capacity (CVR). A high CVR is correlated with optimal brain health and intact cognitive function in elderly healthy

Corresponding author:

Mark B Vestergaard, Department of Clinical Physiology, Nuclear Medicine and PET, Functional Imaging Unit, Rigshospitalet, Nordre Ringvej 57, Glostrup DK-2600, Denmark.

Email: mark.bitsch.vestergaard@regionh.dk

¹Department of Clinical Physiology, Nuclear Medicine and PET, Functional Imaging Unit, Copenhagen University Hospital, Rigshospitalet Glostrup, Glostrup, Denmark

²Institute of Clinical Medicine, The Faculty of Health and Medical Sciences, University of Copenhagen, Copenhagen, Denmark

adults¹⁻³; in contrast, impaired CVR is evident in a number of diseases correlated with stroke and cognitive decline. Patients with metabolic syndrome and related diseases, such as type 2 diabetes and obstructive sleep apnea, have reduced baseline CBF and CVR.4-9 Furthermore, CVR decreases with age, 10 and this decrease may play a role in the development of Alzheimer's disease. 11 These results suggest a correlation between high CVR and brain health. Previous studies in young, healthy subjects with normal CVR have found that a moderate-to-severe hypoxic challenge resulted in elevated CBF, which compensated, or almost compensated, for the arterial oxygen desaturation^{12–17} and sustained^{12–14}, or slightly increased, the cerebral metabolic rate of oxygen (CMRO₂). 15-17 Elevated CMRO2 is likely driven by hypocapnia from increased ventilation during hypoxia. 15,17

In addition to depending on oxygen, the brain relies on a continuous supply of glucose. In the adult brain, glucose is converted into energy mostly by oxidative phosphorylation; however, a small part of the glucose solely undergoes glycolysis where pyruvate is instead transformed to lactate as the end-product. Studies using magnetic resonance spectroscopy (MRS) have shown increased lactate concentrations in the resting brain during hypoxia^{17–19} and additionally, that a combination of visual stimulation and hypoxia results in an amplified lactate increase.¹⁷ The exact reason for the higher lactate concentration during hypoxia is not clear, but could be from increased astrocytic glycolysis. We hypothesize that increased glycolysis and lactate production constitute a rescue mechanism when hypoxia develops and threatens oxygen metabolism, and is a healthy sign of an adaptable metabolic system prepared to face a dangerous lack of oxygen. A similar shift towards anaerobic glycolysis in addition to high cerebral perfusion is observed in some diving marine mammals.20

During the prolonged BH performed by freedivers, the CVR is enhanced by both hypercapnia and hypoxia in the brain. Because freedivers can endure this critical environment in the brain, they are ideal candidates for the examination of the CVR in extreme situations. We hypothesize that the freediver will have a large increase in CBF to compensate for arterial desaturation. We conjecture further that if the increase in CBF is not sufficient to sustain cerebral oxygen supply, the freedivers will exhibit additional coping mechanisms, such as a shift towards more glycolysis, at least in astrocytes, which could manifest as a reduction in oxygen consumption.

The cerebrovascular reactivity and cerebral oxygen consumption in freedivers were examined by measuring the global CBF and global CMRO₂ at a high temporal resolution during extended BHs using magnetic

resonance imaging (MRI) techniques. To further investigate the effect of hypoxia on the cerebral metabolism, the participants underwent a 40 min hypoxic challenge by inhaling air with 10% fractional inspired oxygen (FiO₂). In addition to CBF and CMRO₂, cerebral lactate, glutamate+glutamine (Glu+Gln), N-acetylaspartate (NAA) and phosphocreatine+creatine (PCr+Cr) concentrations were measured by MRS before and during the hypoxic challenge. We hypothesize that a shift towards glycolysis is more pronounced in freedivers than in non-diver controls, and that during a hypoxic challenge, the freedivers will show a higher lactate concentration and thereby provide evidence of a cellular coping mechanism.

Methods

Fifteen freedivers (7 females, mean age=32.7 years, range=24.7–45.3 years) and 17 healthy controls (7 females, mean age=28.8 years, range=20.4–39.7 years) participated in the study. Only freedivers currently active in the sport were included in the study. The controls had no prior experience with freediving, diving in general or training BHs. The participants had no known heart, lung or neurological diseases.

The study was approved by the Danish National Committee on Health Research Ethics (H-15003589) and was conducted according to the Declaration of Helsinki. All subjects gave written informed consent prior to participation. All scans were performed on a Philips 3 Tesla Achieva dSTREAM MRI scanner (Philips Medical Systems, Best, The Netherlands) using a 32-channel phase array head coil.

The experiment consisted of one MRI session covering initial anatomical scans, followed by baseline normoxia functional scans, two to four scans during BHs, and lastly, functional scans during hypoxia (Figure 1). The functional scans consisted of acquisitions of global CBF, CMRO₂ and the concentrations of lactate, Glu+Gln, NAA and PCr+Cr. During BHs, global CBF or CMRO₂ was measured.

During the scan, the subjects were fitted with a face-mask covering the mouth and nose. During normoxia measurements, the subjects inhaled atmospheric air from the surrounding environment. During the hypoxia measurements, the mask was connected by a tube and a one-way valve to an AltiTrainer system (SMTEC, Nyon, Switzerland) that provided hypoxic air. A one-way valve in the mask ensured that no rebreathing was possible. End-tidal CO₂ tension was measured from a tube connected to the mask by a Veris Monitor system (MEDRAD, Pittsburgh, Pennsylvania, USA). Heart rate and arterial oxygen saturation were measured continually throughout the scan by fingertip pulse oximetry, also using the Veris Monitor. Before the

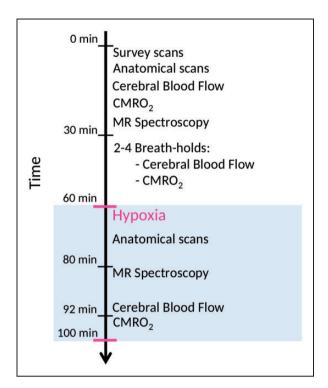


Figure 1. Timing of the measurement of parameters. Survey and anatomical scans, global CBF, combined oxygen saturation and blood flow in the sagittal sinus for calculation of CMRO₂ and MR spectroscopy were acquired during the initial 30 min of normoxia. Hereafter, the participants performed two to four breath-holds, with pauses between each breath-hold. During breath-holding, either global CBF or combined oxygen saturation and blood flow in the sagittal sinus for calculation of CMRO₂ was measured. After 60 min, the participants initiated the hypoxic challenge by inhaling 10% FiO₂ hypoxic air. After 20 min of hypoxic exposure, the MR spectroscopy measurements were repeated, and after 32 min, the acquisitions of CBF and CMRO₂ were repeated.

scan, a venous blood sample was drawn and analysed immediately for haemoglobin and haematocrit using a Radiometer ABL800 Flex (Radiometer, Copenhagen, Denmark).

Breath-hold paradigm

Acquisition of CBF or CMRO₂ was initiated approximately 2–3 min prior to breath-holding, and was thereafter continuously measured until cessation of the BH period. After approximately 2 min of resting for baseline measurements, the subjects were asked to prepare for breath-holding and subsequently start the breath-holding. The preparation consisted of one to three deep inhalations depending on the divers' personal preference. The participants were encouraged not to hyperventilate or to perform lungpacking. The breath-holding was initiated after a full inhalation. The contribution

of breath holding to CBF or CMRO₂ measurements was inferred by comparing data acquired during the 2-min baseline period with the subsequent breath-holding phase.

Magnetic resonance imaging methods

Anatomical scan. Anatomical scans for estimating brain weight were obtained with a 3D T1-weighted turbo field echo sequence (150 slices, field of view (FOV)= $241 \times 180 \times 16 \,\mathrm{mm}^3$ voxel size= $1.09 \times 0.81 \times 1.10 \,\mathrm{mm}^3$, echo time (TE)= $2.78 \,\mathrm{ms}$, repetition time (TR)= $6.9 \,\mathrm{ms}$, flip angle= 9°). Brain volume was estimated from anatomical images using FSL BET and FAST software (FMRIB Software Library, Oxford University, Oxford, UK), 21 and brain weight was calculated by assuming a brain density of $1.05 \,\mathrm{g/ml}$. 22

Magnetic resonance spectroscopy. The lactate, Glu+Gln, NAA and PCr+Cr concentrations were measured in the occipital lobe using MRS. It is not possible to differentiate between the concentrations of glutamate and glutamine or between phosphocreatine and creatine when using 3 Tesla MRS in vivo in humans. Therefore, only the combined concentrations of glutamate+glutamine and phosphocreatine+creatine are reported. A water-suppressed point-resolved spectroscopy (PRESS) pulse sequence was used (TR = $3000 \, \text{ms}$, TE = $36.5 \, \text{ms}$, voxel size = $30 \times 35 \times 30 \, \text{mm}^3$, $160 \, \text{acquisitions}$, total scan time = $8 \, \text{min} \, 54 \, \text{s}$). Post-processing and quantification of the spectra were performed using LCModel (LCModel (Version 6.3-1F), Toronto, Canada).

Cerebral blood flow. The mean global CBF was calculated by measuring the mean velocity and the cross-sectional area of the carotid arteries and basilar artery by phase contrast mapping. Blood velocity maps were acquired with a turbo field echo sequence (FOV= $240 \times 240 \text{ mm}^2$, voxel size= $0.75 \times 0.75 \times 8 \text{ mm}^3$, 1 slice, TE=7.33 ms, TR=27.63 ms, flip angle= 10° , velocity encoding=100 cm/s).

For acquiring global CBF before and during the hypoxic challenge, two measurements were performed. The first measurement was performed with an imaging slice placed orthogonal to the carotid arteries and was followed by a second measurement with an imaging slice placed orthogonal to the basilar artery. Each measurement was repeated five times and averaged, resulting in total scan-time of 3 min 19 s. To measure global CBF during BH, only one imaging slice was used that acquired flow in the carotid and vertebral arteries simultaneously, thus enabling a high temporal resolution of 10.9 s but at the cost of slightly decreasing the accuracy of the quantification. ²³ Examples of

velocity maps acquired during a BH are shown in the supplementary material (Figure S1). Regions of interest (ROIs) were drawn corresponding to each cerebral artery, and the mean velocities and cross-sectional areas were calculated from the ROIs. The ROIs were initially drawn on the magnitude image and thereafter copied to each velocity map and corrected for misalignment due to possible motion, if necessary. Flow was calculated by multiplying the mean velocity and cross-sectional areas for all arteries. Quantitative global CBF was calculated by normalizing the total flow to the whole brain tissue weight.

The cerebral delivery of oxygen (CDO₂) was calculated by multiplying CBF by the arterial oxygen concentration (CaO₂). CaO₂ was calculated as the arterial saturation multiplied by the oxygen carrying haemoglobin concentration.

Post-processing was performed using Matlab (Mathworks, Natick, MA, USA) scripts developed inhouse.

Cerebral metabolic rate of oxygen. CMRO₂ was calculated using Fick's Principle (equation (1))

$$CMRO_2 = [Hgb] \cdot BF_{SS} \cdot (SaO_2 - SvO_2)$$
 (1)

where [Hgb] is the oxygen carrying haemoglobin concentration (mmol/l), BF_{ss} is the blood flow in the sagittal sinus in ml/min, SaO₂ is the arterial oxygen saturation, and SvO2 is the oxygen saturation of the venous blood in the sagittal sinus. SaO2 was measured by pulse oximetry. SvO₂ and BF_{ss} were measured by a sequence combining susceptibility-based MRI for oxygen saturation and phase contrast for flow.²⁴ By simultaneously measuring venous saturation and blood flow in the sagittal sinus, we ensured that flow and saturation corresponded in time and originated from the same brain tissue. The calculated CMRO2 will derive from approximately one half of the brain drained by the sagittal sinus, which we assume to be representative of the entire brain. During steady-state baseline, the CMRO₂ measurement was normalized to individual brain volumes to compare values with prior published results by scaling the CMRO2 calculated from equation (1) with the ratio between global CBF and BFss measured at resting baseline in each individual subject and normalised to brain weight.9 During physiological challenges, the ratio between CBF and BF_{ss} may vary and scaling of CMRO₂ was therefore omitted when comparing baseline CMRO2 with values during breath-holding and the hypoxic challenge.

The sequence uses a dual gradient-echo sequence to generate susceptibility-weighted phase-difference maps by subtracting phase maps from the two echoes $(FOV = 220 \times 189 \text{ mm}^2, 1 \text{ slice, voxel size} = 0.7 \times 0.7 \times 8 \text{ mm}^3, TR = 23 \text{ ms, TE1} = 8.03 \text{ ms, TE2} = 17.73 \text{ ms, flip angle} = 30^{\circ}, \text{ velocity encoding} = 100 \text{ cm/s, SENSE-factor} = 2). The differences in phase-difference values in venous blood and brain tissue can be related to saturation using an exact formulation (equation <math>(2))^{24,25}$

$$SvO_{2} = \left(1 - \frac{2|\phi_{ss} - \phi_{tissue}|}{\gamma \Delta \chi_{do} B_{0} \Delta TE(\cos^{2}\theta - \frac{1}{3})Hct} + \frac{\Delta \chi_{oxy}}{\Delta \chi_{do}}\right)$$

$$\cdot 100\%$$
(2)

where ϕ_{ss} is the phase-difference value in the venous blood in the sagittal sinus, ϕ_{tissue} is the phase-difference value in the surrounding brain tissue, θ is the angle between the sagittal sinus and the B₀ field acquired from MRI angiography, Hct is the haematocrit, $\Delta \chi_{do} = 4\pi \cdot 0.27$ ppm is the susceptibility difference between fully deoxygenated blood and fully oxygenated blood, and $\Delta \chi_{\text{oxy}} = 4\pi (-0.008)$ ppm is the susceptibility difference between fully oxygenated blood and water.²⁶ The phase-difference values in venous blood and surrounding brain tissue were determined by drawing ROIs on the phase-difference maps. For each echo, the measurement was repeated with velocity encoding for the simultaneous measurement of blood velocity in the sagittal sinus. Examples of phase-difference maps and velocity maps perpendicular to the sagittal sinus acquired during a BH are shown in the supplementary material (Figure S1). The blood flow in the sagittal sinus was determined by drawing a ROI in the sagittal sinus and multiplying the mean velocity and cross-sectional area from the ROI, similar to the post-processing used to calculate CBF described earlier. To determine the baseline normoxia CMRO₂ and hypoxia CMRO₂, each measurement was repeated 10 times and averaged. To determine the CMRO₂ during BH, the measurements were repeated throughout the BH with a time resolution of 6.9 s. ROIs were corrected for misalignment due to possible motion for each repeated measurement. If the motion caused severe artefacts and the sagittal sinus could not be delineated, the measurements were discarded. In some subjects, we observed artefacts of high velocity at the edge of the vein, probably related to motion and non-linear flow; these voxels were removed from the ROIs.

SaO₂ was continuously sampled by pulse oximetry. Prior to the study, the pulse oximeter was calibrated with simultaneously drawn arterial blood samples from 23 subjects during normoxia and hypoxia from a previous study with a similar set-up and equipment.¹⁷

Arteriovenous oxygen saturation difference (A-V O_2) was calculated by SaO_2 - SvO_2 for the measurements from normoxia, the hypoxic challenge, and the time series from the BHs. The time series of A-V O_2

saturation and BF_{ss} acquired during BHs were low-pass filtered with a five-sample mean filter to reduce noise in the time series before the CMRO₂ calculations.

Post-processing was performed using Matlab scripts developed in-house.

Statistics

The time series acquired during BH were aligned to the beginning of the BHs (example in Figures 2, 3 and 5). To test for significant changes during BH, the time series were stretched or narrowed by linear interpolation to the mean BH duration (example in Figure 2(c) and (d)). Each interpolated time point (Y_t) was tested against baseline values before the BHs by a mixed model linear regression (equation (3)) with a categorical variable indicating BH or baseline $(C_{\text{Baseline}|\text{BH}})$, subject grouping (u, matrix indicating)

subject as most subjects contributed with multiple BHs) as a random variable to account for between-subject variability and ε as noise (equation (3)).

$$Y_t = \beta_0 + \beta_1 \cdot C_{\text{Baseline}|BH} + u + \varepsilon \tag{3}$$

Significance of β_1 will indicate difference between baseline values and values during BHs. Two tests were performed for the measurement of CMRO₂: one for all BHs longer than 2 min 30 s (example on Figure 5(a)) and a separate test only for BHs longer than 4 min (example on Figure 5(b)). The *p*-values from all time points were corrected for multiple comparisons using Holm–Bonferroni corrections.²⁷ The effects of breathholding are summarized in the supplementary material (Table S1).

The average rates at which CBF increased during the BHs of the non-diver controls were compared with the

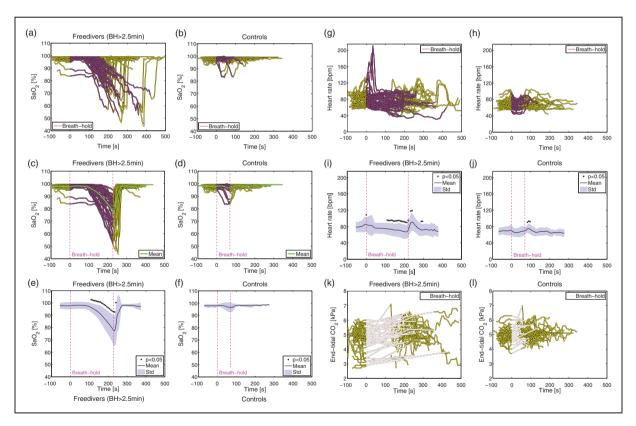


Figure 2. Time series of blood gasses and heart rate during breath-holds. Time series (yellow and purple lines) of arterial saturation during breath-holds (BH) in freedivers (n = 38) (a) and non-diver controls (n = 30) (b). Breath-holds, indicated by the purple line, were aligned to start at time equal zero. Individual time series of the arterial saturation were stretched or narrowed by linear interpolation to the average breath-hold duration (3 min 50 s) with mean saturation (green line) for freedivers (c) and for non-divers (d). Mean (black line) and standard deviation (blue shade) from the interpolated time series and indication of significant differences (black star) from baseline values from before breath-hold for freedivers (e) and non-diver controls (f). Time series of heart rate in freedivers (g) and non-diver controls (h). Mean heart rate (black line) and standard deviation (blue shade) and indication of significant differences (black star) for freedivers (i) and non-diver controls (j). Time series of end-tidal CO_2 from expired air from freedivers (k) and non-diver controls (l). End-tidal CO_2 could not be measured during the breath-hold because of no expired air (indicated by the gray lines).

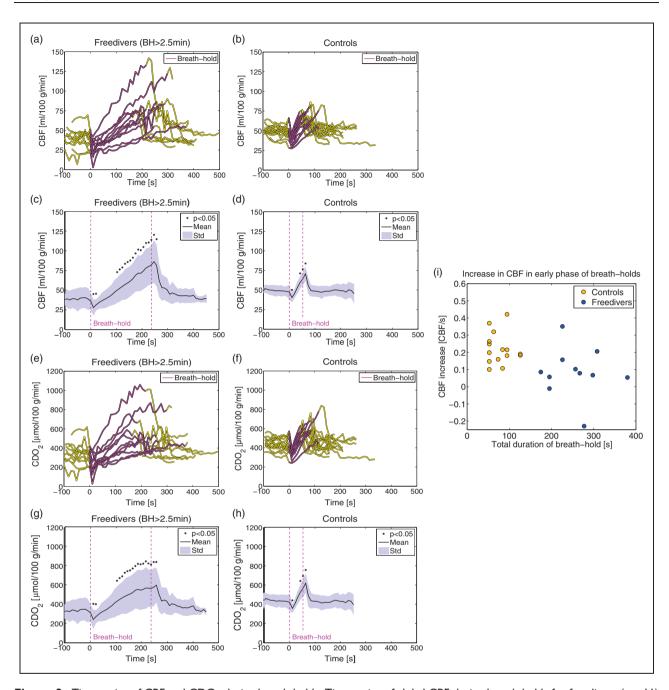


Figure 3. Time series of CBF and CDO₂ during breath-holds. Time series of global CBF during breath-holds for freedivers (n = 11) (a) and non-diver controls (n = 15) (b) and mean CBF (black line) and standard deviation (blue shade) for interpolated time series with indications of significant differences (black star) for freedivers (c) and non-divers controls (d). Global CDO₂ during breath-hold for freedivers (e) and non-diver controls (f) and mean CDO₂ and standard deviation for interpolated time-series with indication of significant differences (black star) for freedivers (g) and non-diver controls (h). (i) Rate of increase in CBF from the breath-holds performed by the non-diver controls (yellow) and from the early phase of the breath-holds from freedivers (blue) against total duration of breath-hold. Student's t-test between the rates from freedivers and controls yielded a significant difference (p = 0.0058).

rates observed in freedivers in the early phase of their BH, which corresponded to the mean duration of the controls' BHs. The rate of CBF increase is shown against the total BH duration for the two groups in Figure 3(i). The rates of CBF increase between the

two groups were compared using Student's *t*-test. Post-processing of measurements from BHs was blinded regarding subject.

All post-processing of data from the hypoxic challenge was blinded regarding gas challenge, subject and

group. The results from the hypoxic challenge are presented as the mean±standard deviation (Table 1). If the subject had pronounced hyperventilation during the hypoxic challenge and the arterial saturation did not decrease to less than 85%, the subject was excluded from further analysis with regard to the hypoxic challenge. The effects on the measured parameters were tested with a paired Student's *t*-test (Table 1). Subject response to the hypoxic challenge may vary in terms of the degree of hypoxia, as measured by CaO₂; therefore, a linear regression model was utilized to test for a

correlation between CaO_2 , and the measured parameter of interest. A mixed model linear regression was used with the measured parameter modelled as the response variable (Y), CaO_2 as the fixed variable, subject grouping (u) as the random variable to account for between-subject variability, and ε as random noise (equation (4)).

$$Y = \beta_0 + \beta_1 \cdot \text{CaO}_2 + u + \varepsilon \tag{4}$$

To test for differences in responses to the hypoxic challenge between freedivers and non-diver controls, the

Table 1. Summary of the results and statistics from the hypoxic challenge.

	Normoxia	Нурохіа	Absolute change	Relative change (%)	t-test p-value	Mixed model regression correlations	
						$CaO_2^a (\beta_1)$ p-value	CaO_2^b · group (β_3) <i>p</i> -value
Freedivers							
Blood gases							
SaO ₂ (%)	$97.4\pm~2.0$	69.7 ± 9.0	-27.7	-28.4	< 10 ⁻⁶ *		
SvO ₂ (%)	69.7 ± 5.0	54.4 ± 7.1	-15.3	-21.9	< 10 ⁻⁶ *		
PaCO ₂ (kPa)	5.1 ± 0.4	$4.2\pm~0.5$	-0.9	−17.1	$< 10^{-5}*$		
Cerebral metabolism							
Heart rate (bpm)	68.0 ± 12.7	81.7 ± 16.7	13.7	20.2	0.003*	0.003*	
CBF (ml/100 g/min)	$49.3 \pm \hspace{1.5mm} 8.0$	67.I ± 13.2	17.8	36.1	$< 10^{-3}*$	< 10 ⁻⁶ *	0.20
CDO ₂ (µmol/100 g/min)	415.9 ± 57.3	389.4 ± 51.8	-26.5	-6.4	0.11	0.094	0.24
BF _{ss} (ml/min)	310.0 ± 54.6	527.8 ± 154.6	217.8	70.3	$< 10^{-4}*$	< 10 ⁻⁶ *	0.003*
A-V O ₂ saturation (%)	$31.0\pm~5.4$	19.2 ± 4.9	-11.8	−38.I	$< 10^{-5}*$	< 10 ⁻⁶ *	0.11
CMRO ₂ (µmol/min)	$829.8 \pm\ 203.4$	$853.7 \pm\ 288.6$	23.9	2.8	0.66	0.58	0.53
Cerebral lactate (mmol/l)	$\textbf{0.44} \pm \ \textbf{0.18}$	$0.67 \pm\ 0.22$	0.23	52.0	0.010*	$< 10^{-3}*$	0.21
Glu+Gln (mmol/l)	$7.5\pm~1.0$	7.3 ± 1.4	-0.2	-3.0	0.41	0.44	0.74
PCr+Cr (mmol/l)	5.7 ± 0.3	$5.4\pm~0.3$	-0.3	-4.5	0.007*	$< 10^{-3}*$	0.44
NAA (mmol/l)	8.01 ± 0.4	$7.7 \pm~0.6$	-0.3	-4.2	0.031*	0.014*	0.61
Controls							
Blood gases							
SaO ₂ (%)	97.5 \pm 1.1	69.6 ± 6.5	-27.9	-28.6	< 10 ⁻⁶ *		
SvO ₂ (%)	69.5 ± 5.0	$53.6 \pm \ 6.5$	-16.0	-23.0	< I0 ⁻⁶ *		
PaCO ₂ (kPa)	$5.0\pm~0.5$	$3.5\pm~0.9$	-1.5	-29.7	$< 10^{-5}*$		
Cerebral metabolism							
Heart rate (bpm)	62.2 ±7.0	79.1 ± 10.7	16.9	27.1	< 10 ⁻⁶ *	< 10 ⁻⁶ *	
CBF (ml/100 g/min)	$48.5\pm~4.5$	62.7 ± 11.1	14.4	29.2	< I0 ⁻⁴ *	< I0 ⁻⁶ *	
CDO ₂ (µmol/100 g/min)	434.4 ± 58.8	393.0 ± 59.0	-41.4	-9.5	0.015*	0.003*	
BF _{ss} (ml/min)	$\textbf{304.4} \pm \textbf{54.6}$	453.0 ± 109.1	148.5	48.8	$< 10^{-5}*$	< 10 ⁻⁶ *	
A-V O ₂ saturation (%)	30.9 ± 5.0	21.6 ± 5.4	-9.3	−30.I	< I0 ⁻⁴ *	$< 10^{-6}*$	
CMRO ₂ (µmol/min)	$859.8 \pm\ 217.3$	873.0 ± 244.0	13.1	1.5	0.79	0.87	
Cerebral lactate (mmol/l)	$0.44 \pm\ 0.23$	$0.80 \pm\ 0.36$	0.36	81.4	$< 10^{-5}*$	$< 10^{-6}*$	
Glu+Gln (mmol/l)	$7.1\pm~1.2$	$7.1\pm~1.5$	0.1	0.9	0.65	0.81	
PCr+Cr (mmol/l)	$5.4 \pm\ 0.4$	$5.3 \pm\ 0.6$	-0.9	-2.6	0.017*	0.005*	
NAA (mmol/l)	$8.1 \pm~0.4$	$7.8\pm~0.7$	-0.4	-4.4	0.08	0.29	

Note: Results are reported as mean \pm SD.

^{*}A statistically significant change (p < 0.05).

^ap-value for correlation with CaO₂ from mixed model linear regression (equation (4)).

^bp-value for correlation with the interaction term from mixed model (equation (5)) with CaO₂, subject grouping and their interaction (β_3) as fixed parameters.

grouping and interaction between grouping and CaO₂ were added to the model (equation (5)).

$$Y = \beta_0 + \beta_1 \cdot \text{CaO}_2 + \beta_2 \cdot \text{group} + \beta_3 \cdot \text{group} \cdot \text{CaO}_2 + u + \varepsilon$$
 (5)

The effect of the interaction parameter (β_3) will describe differences in responses to the hypoxic challenge between the freedivers and non-diver controls.²⁸

Results

Data collection

In total, 47 BHs were performed by 15 freedivers. Eight BHs were shorter than 2 min 30 s and were excluded from further analysis. This exclusion resulted in 11 BHs during measurement of global CBF and 28 during measurements of combined flow and oxygen saturation in the sagittal sinus for the calculation of CMRO₂.

Seventeen non-diver controls performed a total of 30 BHs: 15 corresponding to global CBF measurements, and the remaining 15 corresponding to the combined flow and oxygen saturation CMRO₂ calculation.

Fourteen divers and all controls completed the hypoxic challenge. One freediver had pronounced hyperventilation with arterial saturation over 85% during the hypoxic challenge, likely due to discomfort from ventilating through the mask and valves in the oxygen delivery system, and was excluded from further analysis. The same subject also demonstrated pronounced hyperventilation before breath-holding. One control had a bifurcation of the sagittal sinus making it unfeasible to measure SvO₂. Measurements of BF_{ss}, A-V O₂ saturation and CMRO₂ were excluded from this subject.

Blood gases

The mean duration of all the BHs performed by the freedivers was $3 \min 50 \text{ s} \pm 1 \min 5 \text{ s}$ (range= $2 \min 32 \text{ s}$ to $7 \min 0 s$) and $1 \min 9 s \pm 26 s$ (range=42 s to $2 \min 9 s \pm 26 s$) 19 s) for the non-diver controls. On average, the arterial oxygen saturation decreased significantly $75.3\% \pm 13.2\%$ (range=43.6-94.7%) in the BHs from the freedivers (Figure 2(a) and (e)). The arterial oxygen saturation at the end (or a few seconds after the end) of the BHs decreased to less than 60% for seven divers and decreased to less than 50% for two divers. In the non-divers, the arterial saturation decreased slightly for two subjects and was unchanged for the remaining subjects (Figure 2(b) and (f)).

The average end-tidal CO_2 was 4.8 ± 0.8 kPa (range=2.7-6.0 kPa) before the BH and 5.1 ± 1.0 kPa

(range=3.2–7.0 kPa) immediately after the BH in the freedivers, and these values in the non-divers were 5.0±0.5 kPa (range=4.2–6.2 kPa) and 5.2±0.6 kPa (range=4.0–7.0 kPa), respectively (Figure 2(k) and (l)).

The heart rate decreased gradually throughout the BH in the freedivers but increased immediately before and after the BHs (Figure 2(g) to (j)).

Cerebral blood flow

The mean duration of the freedivers' BHs during the measurement of the global CBF was $4 \min 2 \text{ s} \pm 1 \min 0 \text{ s}$ (range= $2 \min 43 \text{ s}$ to $6 \min 4 \text{ s}$) and the arterial saturation decreased on average to $78.3\% \pm 9.6\%$ (range=56.8-93.0%). The global CBF increased significantly by 107.8% from $40.2 \pm 12.5 \text{ ml}/100 \text{ g/min}$ before the BH to $81.1 \pm 24.1 \text{ ml}/100 \text{ g/min}$ at the very end of the BH (Figure 3(a) and (c)). During the initial phase of the BHs, CBF decreased significantly by 30.6%. The three longest BHs lasted $4 \min 47 \text{ s}$, $4 \min 59 \text{ s}$, and $6 \min 4 \text{ s}$, and revealed increases in CBF of 137%, 165% and 132%, respectively.

The mean duration of the BHs of the non-diver controls was 1 min $7 \, \text{s} \pm 26 \, \text{s}$ (range= $42 \, \text{s}$ to 1 min $56 \, \text{s}$). The global CBF increased significantly by 25.6% from $49.7 \pm 6.2 \, \text{ml}/100 \, \text{g/min}$ before the BH to $62.0 \pm 8.1 \, \text{ml}/100 \, \text{g/min}$ at the end of the BH (Figure 3(b) and (d)). During the initial phase of the BHs, CBF also decreased significantly for the controls by 19.3%. The two longest BHs for the controls were both 1 min $56 \, \text{s}$ and CBF increased by 47.4% and 47.2%. The increased CBF compensated for desaturation, and caused increased CDO₂ throughout the BHs for both freedivers and controls (Figure 3(e) to (h)).

For both the freedivers and non-diver controls, CBF decreased immediately after the end of BHs and normalized in less than 1 min (Figure 3(c) and (d)).

The rates at which CBF increased during the early phase of the BHs were significantly lower in the free-divers than the non-divers (p=0.0058) (Figure 3(i)).

Cerebral metabolic rate of oxygen

Examples of time series of BF_{ss}, A-V O₂ saturation and calculated CMRO₂ for a BH from a freediver with an average duration (4 min 8 s), and for the longest BH (duration=7 min 0 s) are shown in Figure 4. The mean duration of the BHs when measuring CMRO₂ was 3 min 44 s±1 min 5 s (range=2 min 32 s to 7 min 0 s), and the average lowest arterial saturation was 74.1±14.4% (range=43.6–94.7%). BF_{ss} increased significantly by 115.0% from 250.6±65.3 ml/min to 525.5±120.1 ml/min (Figure 5(a) and (d)). The A-V O₂ saturation decreased significantly by 54.4% from 34.2±11.3% before the BH to 15.5±6.8% at the very

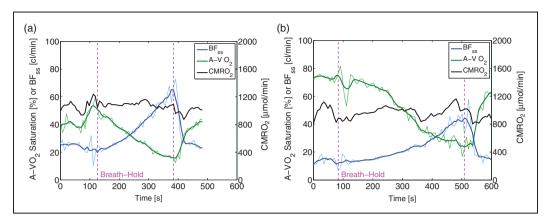


Figure 4. Example of concurrent measurement of blood flow in the sagittal sinus (blue line) and A-V O_2 saturation (green) from the sagittal sinus and CMR O_2 (black) calculated from Fick's principle for an average freediver breath-hold (duration=4 min 8 s) (a) and for the longest freediver breath-hold (duration=7 min 0 s) (b). The raw time series (thin line) and the low-pass mean filtered time series (thick line) are shown.

end of the BHs (Figure 5(g) and (j)). CMRO₂ was sustained throughout the BHs (Figure 5(m) and (p)).

When only BHs over 4 min were included in the analysis, nine BHs remained, with an average duration of 4 min $56 \, \text{s} \pm 1 \, \text{min} \, 6 \, \text{s}$ (range=4 min $8 \, \text{s}$ to 7 min $0 \, \text{s}$), and the average lowest arterial saturation was $59.3 \pm 11.3\%$ (range=43.6–80.0%). For these BHs, BF_{ss} increased by 144% from 226.0 $\pm 49.3 \, \text{ml/min}$ to $542.1 \pm 116.6 \, \text{ml/min}$, and A-V O₂ saturation decreased by 60.8% from $37.8 \pm 16.5\%$ to $14.7 \pm 7.9\%$ (Figure 5(b), (e), (h) and (k)). CMRO₂ remained unaffected (Figure 5(n) and (q)).

In non-divers, the mean BH duration was 1 min $12 \text{ s}\pm27\text{s}$ (range=48 s to 2 min 19 s), and BF_{ss} increased significantly by 35.6% (Figure 5(c) and (f)), the A-V O₂ saturation decreased significantly by 18.8% (Figure 5(i) and (l)), at the end of the BHs and CMRO₂ was unchanged (Figure 5(o) and (r)).

The results of breath-holding are summarized in in the supplementary material (Table S1).

Hypoxic challenge

The results of the hypoxic challenge are shown in Table 1. Baseline normoxia $CMRO_2$ scaled to global CBF and normalised to individual brain volumes yielded on average $129.8\pm18.6\,\mu\text{mol}/100\,\text{g/min}$ for the freediver group and $136.9\pm27.0\,\mu\text{mol}/100\,\text{g/min}$ for the non-diver group, which compare favourably with previous published values. 13,17,24

We observed increased global CBF, BF_{ss} and lactate concentration, decreased PCr+Cr concentration, and unaffected CMRO₂ and Glu+Gln concentration (Figure 6). The mixed model regression showed significant negative correlations between global CBF, BF_{ss},

lactate concentration, and CaO₂ and a significant positive correlation between PCr+Cr and CaO₂ for both freedivers and controls.

NAA concentration was significantly positively correlated with CaO_2 (p=0.014) in freedivers but not in controls. CDO_2 was significantly positively correlated with CaO_2 in the non-divers (p=0.003) but was unaffected in the freedivers (p=0.094). Comparisons of the correlation with CaO_2 (the interaction parameter β_3 (equation 5)) between the two groups yielded a significantly (p=0.003) higher increase in BF_{ss} in freedivers than in non-diver controls, but no significant differences were evident in the remaining parameters (Table 1).

Discussion

Despite being in an unfamiliar and dry environment, the freedivers were still able to demonstrate extraordinary long BHs. During most BHs, the effect of the diver reflex was apparent as a slowing of the heart rate. The arterial oxygen saturation was highly dependent on the duration of the BH, and decreased to remarkably low values for long BHs.

End-tidal CO₂ was only marginally increased after the BH. This result is probably due to the measurement setup, which was not ideal for measuring CO₂ from a single expiration because the expired air was not collected in a bag but instead analysed continuously. A more elaborate setup with arterial blood sampling or collection of expired air was not possible in the MRI-scanner environment. Previous studies on arterial blood gases in freedivers have shown an average increase in arterial CO₂ tension of approximately

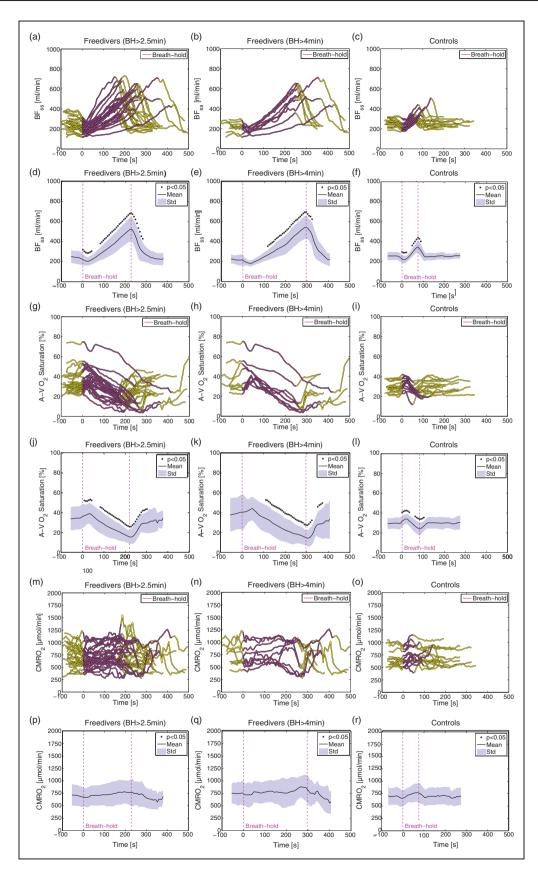


Figure 5. Time series of BF_{ss} , A-V O_2 saturation and $CMRO_2$ during breath-holds. Time series of blood flow in the sagittal sinus (a), arteriovenous oxygen saturation difference (g) and $CMRO_2$ for breath-holds longer than 2 min 30 s from freedivers (n = 28) (m), for breath-holds longer than 4 min from freedivers (n = 9) (b,h,n) and for non-diver controls (n = 15) (c, i, o). Mean (black) and standard deviation (blue shade) of interpolated time-series with indication of significant differences (black star) for blood flow in the sagittal sinus (d), A-V O_2 saturation(j) and $CMRO_2$ (p) for all breath-holds from freedivers, for breath-holds over 4 min from freedivers (e,k,q) and from non-diver controls (f, I, r).

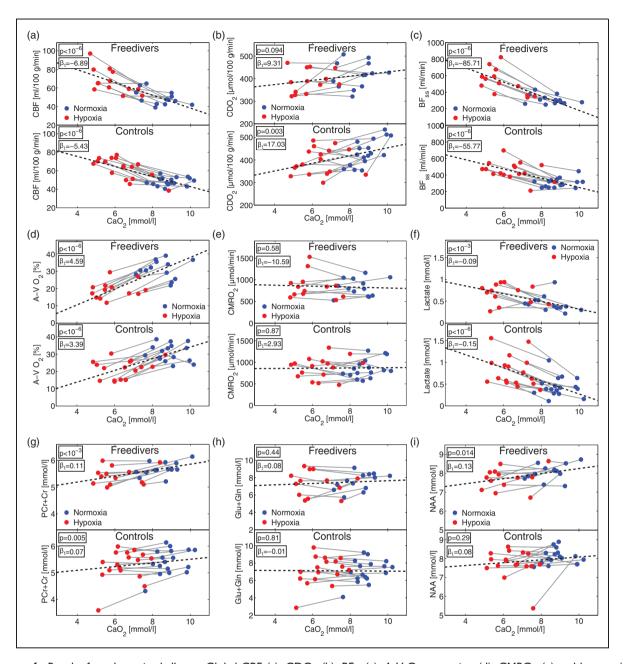


Figure 6. Results from hypoxic challenge. Global CBF (a), CDO₂ (b), BF_{ss} (c), A-V O₂ saturation (d), CMRO₂ (e) and lactate, (f) PCr+Cr, (g), Glu+Gln (h) and NAA (i) concentrations in the occipital lobe during normoxia (blue dots) and during the hypoxic challenge (red dots). Correlations between the parameters and CaO₂ from the mixed model linear regression are shown as black dotted lines. *P*-values and regression-coefficients (β_1 in equation (4)) from each regression are noted.

50% by blood sampling at the end of extended BHs.²⁹ This is probably an accurate depiction of the level of hypercapnia during the extended BHs.

Cerebrovascular reactivity

Freedivers presented remarkable increases in CBF throughout the entire BH, resulting in on average a more than two-fold increase of CBF. The elevated

CBF compensated for the decreased oxygen content in the blood, maintaining the oxygen delivery to the brain. Additionally, CMRO₂ was not impaired, corresponding to the absence of oxygen deprivation in the brain throughout the BHs. At the beginning of a BH, the arterial oxygen saturation was not decreased. Thus, increase in CBF must have been due to the increasing CO₂ tension causing vasodilation. During a BH, when arterial blood began to desaturate, the CVR was

further stimulated by hypoxia. In the non-divers, we observed little or no desaturation, and the increased BF must have been due to hypercapnia.

One may speculate that the non-diver controls could be capable of an equal elevation in CBF, as was seen in the freedivers, and that the main difference with freedivers is due to their ability to suppress the CO₂ stimulus to breathe. This in turn enables them to prolong the BH into more severe state of hypercapnia and hypoxia. However, we did find a correlation between the rate at which the CBF increases and the duration of the BH. For long BHs, the rate at which CBF increased was smaller, especially at the beginning of the BH (Figure 3(i)). This may explain the differences in regulation of CBF between the freedivers and controls. If the non-divers were able to prolong their BHs, the CBF would increase at a higher rate and reach maximal CBF sooner than in the freedivers. It is not clear why the freedivers had a slower rate of increase in CBF. The freedivers could have a less sensitive vasodilating response to hypercapnia, which traditionally means a reduced reactivity. On the contrary, during the hypoxic challenge, the freedivers had the same or slightly higher increases in CBF as the controls with normal reactivity. There could be a decline in the selective sensitivity to hypercapnia, but not to hypoxia. The difference in the rate of CBF increase could also be due to difference in end-tidal CO₂ in the freedivers (4.6 kPa) before the BH compared to that of the controls (5.1 kPa). The CO₂ build-up starts at a lower CO₂ tension at the beginning of the BH, the stimulation from pronounced hypercapnia would manifest later during the BH, and the sensitivity to CO₂ would not be reduced. We expect that the rate of CO₂ build-up was similar between groups as CMRO₂ was equal and sustained in both groups. High CVR is positively correlated with brain health and cognitive function, and negatively correlated with a number of diseases, such as small and large vessel brain diseases, including stroke and cognitive decline.^{2,5-7} It is unclear whether the overall ability of cerebral vasodilation or the selective sensitivity to CO₂ or hypoxia is the most important factor for brain health, but in patients with type 2 diabetes, the decrease in CVR is present regardless of the stimulation.^{6,7} We therefore conjecture that the capacity for vasodilation is an important component for the beneficial effects in the prevention of cerebral decline, and that strengthening the CVR with BHs has potential as a beneficial preventative activity.

Previous studies have quantified CBF and oxygen consumption during BHs by freedivers. ^{29,30} Two studies found that CBF compensated completely for decreased oxygen in the blood, and that the delivery of oxygen to the brain was not impaired. Results are similar to our findings. One study found significantly

decreased CMRO₂ at the very end of the BH but not during the BH.³⁰ The authors suggested cause for this decrease is hypercapnia.^{30,31} In contrast, we did not find decreased CMRO₂ in our study. Differences in experimental setup could account for some of the discrepancy, i.e. study by Bain et al.³⁰ allowed lung packing and had a longer BH preparation phase. Also, study by Bain et al.³⁰ had on average 1 min longer BHs when comparing with our group of BHs longer than 4 min, which could cause difference in blood gasses at end of breath-holding; however, the average desaturations were similar.

A further reason could be that we found a larger increase in CBF of 107.0% compared to an 80% increase, which will manifest itself in the CMRO₂ calculation using Fick's principle. Methodological differences could also account for the discrepancy observed as we used MRI techniques rather than Doppler ultrasound. Doppler ultrasound is not ideal for measuring quantitative flow, and it presents errors mainly related to user-dependent biases and difficulty measuring the blood velocity and cross-sectional area in all of the feeding arteries simultaneously. On contrary, phase contrast mapping can potentially overestimate the flow quantification due to partial volume errors in small vessels. ^{23,43,44} All of which could account for the inconsistency between the studies. ^{32,33}

Breath-hold breakpoint

During freediver competitions, loss of consciousness or motor control can occur.³⁴ In these situations, there must be insufficient cerebral oxygen delivery and therefore decreased CMRO₂. Saturations less than 40–50% will often result in loss of consciousness and neuronal function. In this study, the arterial saturation decreased on average to 59.7% in BHs over 4 min and to less than 50% for two BHs, which we must expect is close to a hypoxic loss of neuronal function, yet in these BHs, the CMRO₂ was not decreased. Our data therefore suggest that CMRO₂ is sustained until, or very close to, the potential loss of consciousness or neuronal function.

The breaking point of the BH is determined by numerous factors, including lung stretch, volitional cognitive factors, and chemoreceptor stimulation by oxygen and CO₂. To the non-diver controls, increased stimulation from CO₂ tension caused a breaking point before any hypoxic stress in the brain. The freedivers, in contrast, seemed to be capable of suppressing the CO₂ stimulation to stop the BH and could even continue during hypoxia. For the majority of BHs observed in freedivers, we saw a breaking point before severe hypoxia in the brain, but for the very long BHs, over 4 min, the arterial blood desaturated on average to less than 60%. The ability to reach severe

hypoxia during a BH is probably determined by volitional cognitive factors.

By hyperventilating before the BH, freedivers can decrease the CO₂ tension in the blood and delay the CO₂ drive to end the BH. This capability is often the reason for hypoxic loss of consciousness or motor function because the BH can be extended into dangerous hypoxic levels.³⁴ In the longest BH in this study lasting 7 min 0 s, the freediver had pronounced hyperventilation before the BH, a feature not observed with the other subjects within the study. This effect was accompanied by low end-tidal CO₂, large A-V O₂ saturation and low CBF before the BH (Figure 4(b)). The hypocapnic environment from hyperventilation could cause the slow rate of increase in CBF; however, a large A-V O₂ saturation was required to account for the low CBF and continued throughout the BH. This BH showed that through hyperventilation, the point of maximal CBF can be substantially delayed to prolong the BH. However, the extraction of oxygen will be increased and make the brain more sensitive to desaturation, thus shortening the BH. There seems to be an advantageous effect from hyperventilation through lowering of the CO₂ tension and CBF but an opposite effect from higher oxygen extraction; however, the total effect seems to result in a prolonged BH.

Hypoxic challenge

During the hypoxic challenge, both groups presented increased CBF and BFss. We observed significantly larger BF_{ss} in the freedivers when compared to the non-divers, which suggests increased hypoxic reactivity in the freedivers. The freedivers also had a larger increase in global CBF compared to non-divers, but this was not significantly different. The reason we observed significant differences in BFss and not global CBF could be due to relatively greater increases in perfusion in the areas of the brain drained by the sagittal sinus during the hypoxic challenge. The freedivers sustained normal CDO₂ during the hypoxic challenge (Table 1, Figure 6(b)), which also suggests increased cerebrovascular reactivity, at least to hypoxic stimulation; however, the mixed model regression showed no significant difference between the freedivers and controls. Both groups showed unchanged CMRO2. Similar studies have previously shown no effect 12,13 or small increases in CMRO2 that might be related to different levels of hypocapnia. 15-17 The discrepancy with Vestergaard et al. 17 is probably due to the more homogenous ventilatory responses in this study, where the subjects on average demonstrated greater desaturation. The discrepancy with the increased CMRO₂ observed by Smith et al. 15 and Xu et al. 16 could also be due to the greater desaturation found in this study.

For both groups, we observed increased lactate concentrations, similar to previously published results. 19 The exact reason for the increased lactate is not obvious as CMRO₂ is maintained. It could be hypothesized that lactate is produced by increased glycolysis in astrocytes, a defence mechanism that potentially conserves oxygen for the more oxygen-dependent neurons. We would expect that increased astrocytic glycolysis would cause a decrease in CMRO2, which we did not observe. However, this finding could be because the global CMRO₂ measurement is not sensitive enough, and variation could not be Additionally, astrocytes only account for 5-15% of the global oxygen consumption, ^{39,40} and a small shift towards glycolysis in astrocytes could be smaller than our detection limit. We did not observe differences in lactate concentrations between the two groups, and therefore found no evidence for a larger shift towards glycolysis as a coping mechanism in freedivers compared to controls.

The NAA concentration was slightly decreased in the freedivers during the hypoxic challenge. NAA is related to neuronal density; thus, we would not expect an effect from this short hypoxic challenge, and we believe the decrease is more likely a result of a blood oxygenation level dependent (BOLD) effect on the measured spectrum from the increased flow than a physiological change during the hypoxic challenge. A possible BOLD effect will most likely be present in the strongest singlet in the spectrum, e.g. the NAA peak, and negligible in the weaker metabolite peaks.

The Glu+Gln concentrations were unchanged for both groups, and the PCr+Cr concentrations decreased for both groups. Hypoxic challenges have previously been shown to increase glutamate concentration and decrease PCr+Cr, but during additional neuronal simulation, making comparisons with this study difficult. It is hypothesized that the decrease is due to a shift towards creatine kinase-mediated ATP catalysis as a potential coping mechanism during hypoxia.

We found no differences between the freedivers and controls in CMRO₂ or lactate, Glu+Gln or PCr+Cr concentrations during the hypoxic challenge and no effect on CMRO₂ during the BH. Therefore, the only coping mechanism we found evidence for in freedivers was a remarkable increase in CBF.

Strengths and limitations

The main strength of this study was our ability to measure quantitative blood flow and CMRO₂ throughout entire BHs with a high temporal resolution, and our ability to measure quantitative CBF and CMRO2 along with lactate and glutamate concentration during a hypoxic challenge. Furthermore, we had a relatively

large group of freedivers and a non-diver control group, who together provided a large number of BHs.

A limitation of this study is the lack of arterial CO₂ tension measurements during BHs. Such knowledge on CO₂ tension would have provided further insight into the individual characteristics of BHs.

Measurement of flow by phase mapping has potential errors regarding artery geometry in-plane resolution and cardiac pulsation; however, these errors have been shown to be minor with the present setup. ^{23,43,44}

A further limitation is that we only measured the global values of CBF and CMRO₂, and the values of MRS parameters in the occipital lobe. The combination of position-specific lactate and CMRO₂ measurements will be especially advantageous to further examine possible anaerobic glycolysis at a regional level.

Conclusion

The freedivers presented a remarkable increase in blood flow during BHs, while simultaneously maintaining oxygen consumption throughout. However, in the early phase of the BHs, the freedivers demonstrate a slower increase in CBF compared to controls. This difference could be due to hyperventilation before breathholding or decreased sensitivity to hypercapnic vasodilation. During the hypoxic challenge, the freedivers, unlike the controls, sustained CDO₂. Additionally, we observed greater increases in blood flow in the sagittal sinus but found no evidence for differences in the lactate production, CMRO₂, Glu+Gln or PCr+Cr concentrations when comparing freedivers to controls. We conclude that the main coping mechanism against hypoxia in freedivers is a remarkable aptitude for increasing CBF with no evidence for increased lactate-producing glycolysis compared to controls.

Funding

The author(s) received no financial support for the research, authorship, and/or publication of this article.

Acknowledgements

We express our gratitude to the freedivers who participated in the study and to Københavns Fridykkerklub. We also thank Helle Juhl Simonsen for assistance with blood sampling and analysis.

Declaration of conflicting interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Authors' contributions

MBV and HBWL designed the experiment, conducted the data processing and analysis, and wrote the manuscript.

MBV collected the data. Both authors approved the final version of the manuscript.

Supplementary material

Supplementary material for this paper can be found at the journal website: http://journals.sagepub.com/home/jcb

Data availability

The data derived from the MRI-images and all other data supporting the findings of this study are available from the corresponding author upon request.

References

- Brown AD, McMorris CA, Longman RS, et al. Effects of cardiorespiratory fitness and cerebral blood flow on cognitive outcomes in older women. *Neurobiol Aging* 2010; 31: 2047–2057.
- 2. Eskes GA, Longman S, Brown AD, et al. Contribution of physical fitness, cerebrovascular reserve and cognitive stimulation to cognitive function in post-menopausal women. *Front Aging Neurosci* 2010; 2: 137.
- Hajjar I, Marmarelis V, Shin DC, et al. Assessment of cerebrovascular reactivity during resting state breathing and its correlation with cognitive function in hypertension. *Cerebrovasc Dis* 2015; 38: 10–16.
- Tyndall A V, Davenport MH, Wilson BJ, et al. The brain-in-motion study: effect of a 6-month aerobic exercise intervention on cerebrovascular regulation and cognitive function in older adults. BMC Geriatr 2013; 13: 21.
- 5. Tyndall AV, Argourd L, Sajobi TT, et al. Cardiometabolic risk factors predict cerebrovascular health in older adults: results from the Brain in Motion study. *Physiol Rep* 2016; 4: e12733.
- Dandona P, James IM, Newbury PA, et al. Cerebral blood flow in diabetes mellitus: evidence of abnormal cerebrovascular reactivity. Br Med J 1978; 2: 325–326.
- 7. Fülesdi B, Limburg M, Bereczki D, et al. Cerebrovascular reactivity and reserve capacity in type II diabetes mellitus. *J Diab Complicat* 1999; 13: 191–199.
- 8. Placidi F, Diomedi M, Cupini LM, et al. Impairment of daytime cerebrovascular reactivity in patients with obstructive sleep apnoea syndrome. *J Sleep Res* 1998; 7: 288–292.
- Rodgers ZB, Leinwand SE, Keenan BT, et al. Cerebral metabolic rate of oxygen in obstructive sleep apnea at rest and in response to breath-hold challenge. J Cereb Blood Flow Metab 2016; 36: 755–767.
- Galvin SD, Celi LA, Thomas KN, et al. Effects of age and coronary artery disease on cerebrovascular reactivity to carbon dioxide in humans. *Anaesth Intens Care* 2010; 38: 710–717.
- 11. Girouard H and Iadecola C. Neurovascular coupling in the normal brain and in hypertension, stroke, and Alzheimer disease. *J Appl Physiol* 2006; 100: 328–335.
- Kety SS and Schmidt CF. The effects of altered arterial tensions of carbon dioxide and oxygen on cerebral blood flow and cerebral oxygen consumption of normal young men. J Clin Invest 1948; 27: 484–492.

- Cohen PJ, Alexander SC, Smith TC, et al. Effects of hypoxia and normocarbia on cerebral blood flow and metabolism in conscious man. *J Appl Physiol* 1967; 23: 183–9.
- Ainslie PN, Shaw AD, Smith KJ, et al. Stability of cerebral metabolism and substrate availability in humans during hypoxia and hyperoxia. *Clin Sci* 2014; 126: 661–70.
- Smith ZM, Krizay E, Guo J, et al. Sustained high-altitude hypoxia increases cerebral oxygen metabolism. J Appl Physiol 2013; 114: 11–8.
- Xu F, Liu P, Pascual JM, et al. Effect of hypoxia and hyperoxia on cerebral blood flow, blood oxygenation, and oxidative metabolism. *J Cereb blood flow Metab* 2012; 32: 1909–18.
- Vestergaard MB, Lindberg U, Aachmann-Andersen NJ, et al. Acute hypoxia increases the cerebral metabolic rate

 a magnetic resonance imaging study. J Cereb Blood Flow Metab 2016; 36: 1046–1058.
- Harris AD, Roberton VH, Huckle DL, et al. Temporal dynamics of lactate concentration in the human brain during acute inspiratory hypoxia. *J Magn Reson Imaging* 2013; 37: 739–45.
- 19. Arngrim N, Schytz HW, Britze J, et al. Migraine induced by hypoxia: an MRI spectroscopy and angiography study. *Brain* 2016; 139: 723–737.
- Ramirez J-M, Folkow LP and Blix AS. Hypoxia tolerance in mammals and birds: from the wilderness to the clinic. *Annu Rev Physiol* 2007; 69: 113–43.
- 21. Jenkinson M, Beckmann CF, Behrens TEJ, et al. Fsl. *Neuroimage* 2012; 62: 782–790.
- Torack RM, Alcala H, Gado M, et al. Correlative assay of computerized cranial tomography (CCT), water content and specific gravity in normal and pathological postmortem brain. *J Neuropathol Exp Neurol* 1976; 35: 385–392.
- Peng S-L, Su P, Wang F-N, et al. Optimization of phasecontrast MRI for the quantification of whole-brain cerebral blood flow. *J Magn Reson Imaging* 2015; 42: 1126–1133.
- Rodgers ZB, Jain V, Englund EK, et al. High temporal resolution MRI quantification of global cerebral metabolic rate of oxygen consumption in response to apneic challenge. J Cereb Blood Flow Metab 2013; 33: 1514–22.
- Fernández-Seara MA, Techawiboonwong A, Detre JA, et al. MR susceptometry for measuring global brain oxygen extraction. Magn Reson Med 2006; 55: 967–73.
- Jain V, Abdulmalik O, Propert KJ, et al. Investigating the magnetic susceptibility properties of fresh human blood for noninvasive oxygen saturation quantification. *Magn Reson Med* 2012; 68: 863–867.
- 27. Holm S. A Simple sequentially rejective multiple test procedure. *Scand J Stat* 1979; 6: 65–70.
- 28. Gujarati D. Use of dummy variables in testing for equality between sets of coefficients in linear regressions: a generalization author. *Am Stat* 1970; 24: 18–22.
- Willie CK, Ainslie PN, Drvis I, et al. Regulation of brain blood flow and oxygen delivery in elite breath-hold divers. *J Cereb blood flow Metab* 2015; 35: 66–73.

- 30. Bain AR, Ainslie PN, Hoiland RL, et al. Cerebral oxidative metabolism is decreased with extreme apnoea in humans; impact of hypercapnia. *J Physiol* 2016; 594: 5317–5328.
- 31. Bain AR, Ainslie PN, Barak OF, et al. Hypercapnia is essential to reduce the cerebral oxidative metabolism during extreme apnea in humans. *J Cereb Blood Flow Metab* 2017; 37: 3231–3242.
- 32. Schöning M, Walter J and Scheel P. Estimation of cerebral blood flow through color duplex sonography of the carotid and vertebral arteries in healthy adults. *Stroke* 1994; 25: 17–22.
- 33. Thomas KN, Lewis NCS, Hill BG, et al. Technical recommendations for the use of carotid duplex ultrasound for the assessment of extracranial blood flow. *Am J Physiol Regul Integr Comp Physiol* 2015; 309: R707–R720.
- Lindholm P. Loss of motor control and/or loss of consciousness during breath-hold competitions. *Int J Sports Med* 2007; 28: 295–299.
- 35. Parkes MJ. Breath-holding and its breakpoint. *Exp Physiol* 2006; 91: 1–15.
- 36. Godfrey S and Campbell EJM. Mechanical and chemical control of breath holding. *Q J Exp Physiol Cogn Med Sci* 1969; 54: 117–128.
- 37. Skow RJ, Day TA, Fuller JE, et al. The ins and outs of breath holding: simple demonstrations of complex respiratory physiology. *Adv Physiol Educ* 2015; 39: 223–231.
- 38. Overgaard K, Friis S, Pedersen RB, et al. Influence of lung volume, glossopharyngeal inhalation and PET O2 and PET CO2 on apnea performance in trained breathhold divers. *Eur J Appl Physiol* 2006; 97: 158–164.
- Bélanger M, Allaman I and Magistretti PJ. Brain energy metabolism: Focus on Astrocyte-neuron metabolic cooperation. *Cell Metab* 2011; 14: 724–738.
- Attwell D and Laughlin SB. An energy budget for signaling in the grey matter of the brain. J Cereb Blood Flow Metab 2001; 21: 1133–1145.
- 41. Zhu XH and Chen WD. Observed BOLD effects on cerebral metabolite resonances in human visual cortex during visual stimulation: A functional H-1 MRS study at 4 T. *Magn Reson Med* 2001; 46: 841–847.
- Mangia S, Tkác I, Gruetter R, et al. Sensitivity of single-voxel 1H-MRS in investigating the metabolism of the activated human visual cortex at 7 T. Magn Reson Imaging 2006; 24: 343–8.
- 43. Vestergaard MB, Lindberg U, Aachmann-Andersen NJ, et al. Comparison of global cerebral blood flow measured by phase-contrast mapping MRI with 15O-H2O positron emission tomography. *J Magn Reson Imaging* 2017; 45: 692–699.
- 44. Enzmann DR, Marks MP and Pelc NJ. Comparison of cerebral artery blood flow measurements with gated cine and ungated phase-contrast techniques. *J Magn Reson Imaging* 1993; 3: 705–712.