#### Check for updates

#### OPEN ACCESS

EDITED BY Behafarid Ghalandari, Shanghai Jiao Tong University, China

REVIEWED BY Arturo E. Aguilar-Rabiela, Friedrich-Alexander Universität, Germany Xiaojun Zhou, Donghua University, China

\*CORRESPONDENCE Dejian Li, lidejian880820@163.com Baoqing Yu, doctorybq@163.com

<sup>1</sup>These authors have contributed equally to this work and share second authorship

#### SPECIALTY SECTION

This article was submitted to Biomaterials, a section of the journal Frontiers in Bioengineering and Biotechnology

RECEIVED 28 July 2022 ACCEPTED 17 August 2022 PUBLISHED 12 September 2022

#### CITATION

Sarkari S, Khajehmohammadi M, Davari N, Li D and Yu B (2022), The effects of process parameters on polydopamine coatings employed in tissue engineering applications. *Front. Bioeng. Biotechnol.* 10:1005413. doi: 10.3389/fbioe.2022.1005413

#### COPYRIGHT

© 2022 Sarkari, Khajehmohammadi, Davari, Li and Yu. This is an open-access article distributed under the terms of the Creative Commons Attribution License

(CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# The effects of process parameters on polydopamine coatings employed in tissue engineering applications

# Soulmaz Sarkari<sup>1</sup>, Mehran Khajehmohammadi<sup>2,3†</sup>, Niyousha Davari<sup>4†</sup>, Dejian Li<sup>5</sup>\* and Baoqing Yu<sup>6</sup>\*

<sup>1</sup>Department of Biomedical Engineering, Science and Research Branch, Islamic Azad University, Tehran, Iran, <sup>2</sup>Department of Mechanical Engineering, Faculty of Engineering, Yazd University, Yazd, Iran, <sup>3</sup>Medical Nanotechnology and Tissue Engineering Research Center, Yazd Reproductive Sciences Institute, Shahid Sadoughi University of Medical Sciences, Yazd, Iran, <sup>4</sup>Department of Life Science Engineering, Faculty of New Sciences and Technologies, University of Tehran, Tehran, Iran, <sup>5</sup>Department of Orthopedics, Shanghai Pudong Hospital, Fudan University Pudong Medical Center, Shanghai, China, <sup>6</sup>Department of Orthopedics, Shanghai Pudong New Area People's Hospital, Shanghai, China

The biomaterials' success within the tissue engineering field is hinged on the capability to regulate tissue and cell responses, comprising cellular adhesion, as well as repair and immune processes' induction. In an attempt to enhance and fulfill these biomaterials' functions, scholars have been inspired by nature; in this regard, surface modification via coating the biomaterials with polydopamine is one of the most successful inspirations endowing the biomaterials with surface adhesive properties. By employing this approach, favorable results have been achieved in various tissue engineering-related experiments, a significant one of which is the more rapid cellular growth observed on the polydopamine-coated substrates compared to the untreated ones; nonetheless, some considerations regarding polydopamine-coated surfaces should be taken into account to control the ultimate outcomes. In this mini-review, the importance of coatings in the tissue engineering field, the different types of surfaces requiring coatings, the significance of polydopamine coatings, critical factors affecting the result of the coating procedure, and recent investigations concerning applications of polydopamine-coated biomaterials in tissue engineering are thoroughly discussed.

#### KEYWORDS

polydopamine, surface modification, process parameters, tissue engineering, biomaterial, coating

# 1 Introduction

Material selection and other critical qualities of scaffolds, including possessing an adhesive surface, biodegradability, biocompatibility, mechanical stability, etc., must be meticulously considered so that the various tissues can be successfully regenerated (Zhang et al., 2019; Ghorbani et al., 2020b). Concerning this subject, surface engineering of

biomaterials using various techniques is gaining increasing prominence in tissue engineering (TE) applications, and surface modification, as a popular surface engineering strategy, is utilized to alter the microenvironment of cells and transform the scaffolds' surface into an adhesive one (Arab-Ahmadi et al., 2021; Davari et al., 2022).

Compared to other coating substances, including proteins (Gennadios, 2002), particular peptide sequences (Pagel and Beck-Sickinger, 2017), ceramics (Montazerian et al., 2022), and metals (Yin et al., 2021), polydopamine (PDA), a melanin-like mussel-inspired coating polymer (Ghorbani et al., 2022), possesses a variety of desirable properties, such as outstanding adhesion features (Ghalandari et al., 2021; Feinberg and Hanks, 2022), extraordinary hydrophilicity (Li et al., 2018), biodegradability (Zhang and King, 2020), uniform shape (Saeed et al., 2021), biocompatibility (Zeng et al., 2020), and thermal stability (Yim et al., 2022). Furthermore, its biological properties, including enhanced cellular proliferation, improved bioactivity, free-radical scavenging activities, metal ion chelating capacity, and antibacterial capability, originate from its catecholamine and hydroxyl functional groups (Yu et al., 2017; Ghorbani et al., 2019b, 2019a; Deng et al., 2021). Notably, solvent selection is of paramount importance for solution-based chemical reactions associated with polymers like PDA. Water and ethanol have been broadly employed among various solvents (Jiang et al., 2014; Nieto et al., 2021). A self-assembled PDA layer can be formed on the surfaces of a variety of materials, like metals, ceramics, polymers, oxides, and semiconductors, when soaked in a PDA's weakly alkaline aqueous solution (10 mM Tris-HCl, pH~8.5) without the addition of sophisticated connecting agents (Kao et al., 2018; Yang et al., 2019; Lee et al., 2020; Li et al., 2021b; Park et al., 2021). Nonetheless, the volume ratio of water to alcohol strongly influences the PDA spheres' synthesis. It was illustrated that neither did PDA microspheres form in the pure ethanol nor had a perfect shape in the pure water; in essence, when ethanol and water were mixed with a ratio of 30:70, formation happened in the best way possible (Yan et al., 2013). Insufficient conductivity has ever been the major weakness of PDA (Mei et al., 2020). Since the pros of PDA outweigh its cons, various groups of scientists have modified the biomaterials' surfaces by employing PDA with the aim of augmenting their surface performance. Therefore, within the last couple of years, PDA has been in the spotlight, with diverse applications in the field of biomedical engineering (Lynge et al., 2011), including drug delivery (Huang et al., 2018), implants (Jia et al., 2019), surface engineering (Yang et al., 2015), cancer therapy (Abdollahi et al., 2022; Honmane et al., 2022), TE (bone (Huang et al., 2016; Kaushik et al., 2020), cartilage (Huang et al., 2021), muscle (Zhou et al., 2021), skin (Yazdi et al., 2022), tendon (Lin et al., 2019), and neuron (Qian et al., 2018; Yan et al., 2020)), and microfluidic systems (Niculescu et al., 2021).

The process parameters of PDA considerably impact the coating's quality and characteristics; thus, the experimental factors, such as coating time, temperature, pH, and DA's initial concentration, are properly adjusted so that a coating layer with outstanding features can be formed (Ghorbani et al., 2020c).

Within this paper, firstly, the coatings' significance in the TE field and various kinds of surfaces in need of coatings are comprehensively reviewed. Afterward, the PDA coatings' importance is exhaustively discussed. Ultimately, the parameters that are regulated to improve the properties of coatings and the employment of these interesting coatings in TE by introducing relevant case studies are explained.

## 2 Polydopamine-coated surfaces

Synthetic polymers, such as polylactic-co-glycolic acid (PLGA) (Ghorbani et al., 2020c), polyurethane (PU) (Merceron et al., 2015), polycaprolactone (PCL) (Perez-Puyana et al., 2021), polydimethylsiloxane (PDMS) (Cunniffe et al., 2019), poly-L-lactic acid (PLA) (Jenkins and Little, 2019), polyvinyl alcohol (PVA) (Rodríguez-Rodríguez et al., 2020), polyhydroxyalkanoates (PHAs) (Aguilar-Rabiela et al., 2021; Gregory et al., 2022), and polyamide (Razaviye et al., 2022) are frequently utilized in the fabrication of different scaffolds due to their superior characteristics, comprising customizable degradation rate, excellent processability, great mechanical properties, and wide in-vitro and in-vivo availability. Nevertheless, synthetic polymers typically have poor cell adhesion properties; consequently, combinations with natural polymers or various modifications are common approaches to tackle this obstacle (Terzopoulou et al., 2022). Besides, these modifications are also applied to natural polymers, such as chitosan (Bock et al., 2020), cellulose (Nishida and Koga, 2018), peptides (Ji and Parquette, 2020), etc., to enhance their features.

Boosting biomaterials' performance, modifying their physicochemical properties, and broadening the applications of them are possible by conducting the surface modification procedure (Salatin et al., 2015). Considerable effort has been dedicated to the substances' surface modification via employing various techniques, such as self-assembly monolayer (Kehr et al., 2015), plasma surface modification (Ghorbani et al., 2020d), and layer-by-layer assembly (Liu et al., 2016). Despite these approaches' multiple benefits, they suffer from some drawbacks: limited chemical characterization, restricted shape and dimension of the resultant substances, complicated equipment, and complex synthesis processes (Liu et al., 2016). As an alternative, a coating strategy based upon PDA, formed through covalent polymerization and non-covalent self-assembly (Hong et al., 2012), is widely employed.

The convoluted structure of PDA incorporates a mixture of diverse oligomers formed as a result of dopamine (DA) oxidation, including indole units with varying degrees of oxidation, carboxylic acid and amino groups, imine units, indolic/catecholic  $\pi$ -systems, quinone residues, phenol groups, and open-chain DA molecules (Liu et al., 2014; Ghorbani et al., 2019c). Of note, the functional groups in the PDA structure (catechol, imine, and amine) are employed to reinforce the interaction between the biomaterial and the tissue. What's more, within its irregular, cross-linked network with mixed bonding configurations, radicals are located at quinone residues (Du et al., 2017). Owing to the presence of a series of constant  $\pi$ -electron free radicals, PDA is paramagnetic and operates analogously to a radical trap (Kim et al., 2021b).

As a transitional surface, PDA enables the adhesion of other materials to scaffolds without demanding pre-functionalization. This phenomenon is mainly because of the quinone functional groups that are: 1) electron-deficient and 2) capable of participating in Schiff base reactions occurring on amines and stimulating the Michael addition reactions with nucleophilic groups, including amines and thiols (Rizzo and Kehr, 2021). Notably, the PDA surfaces' sensitivity towards the nucleophiles relies on the catechol/quinone equilibrium; therefore, the generated alkaline pH speeds up conjugation reactions (Batul et al., 2017). Other functional groups within the structure or generated by PDA serve pivotal roles; for instance, amine groups are involved in displacing hydrated minerals on the surface as well as bringing catechols closer to the surface to create strong molecular interactions (Lee et al., 2019). Another example is the hydroxyl and amine groups produced by PDA on the scaffolds that can promptly release signals to promote integrin binding, resulting in the increment of osteogenic activities (Ghorbani et al., 2020c). The more binding functional groups on the surface, the higher probability of the molecules and inorganic ions loading, leading to the improvement of cellular attachment (Richbourg et al., 2019).

# 3 Polydopamine coating's key process parameters in tissue engineering applications

Brimming with outstanding features, PDA coatings have been extensively employed in TE-related investigations. In this regard, several experiments have achieved favorable outcomes when utilizing this coating compared with the others (Sharma et al., 2019; Kopeć et al., 2022). Within a novel study, scaffolds were coated with PDA and gelatin to evaluate the anti-corrosion property. The PDA-coated and gelatin-coated samples were biocompatible and displayed corrosion current density (I<sub>corr</sub>) of  $2.95 \times 10^{-3}$  and  $3.72 \times 10^{-3} \frac{A}{cm^2}$ , respectively, indicating that the PDA coating layer could delay the corrosive process (Zhou et al., 2019).

For PDA polymerization on the surface of the biomaterials, multiple factors should be considered to achieve the desired outcomes in research cases. Among these parameters, coating time, temperature, pH of Tris-HCl, and initial DA concentration in the DA/Tris-HCl buffer solution are discussed in the following. Table 1 presents these determining factors and their impacts upon the formed PDA layer. Furthermore, Figure 1 illustrates the outline briefly describing the case studies in terms of the process parameters and their effects.

### 3.1 Coating time

The PDA coating process commonly occurs at room temperature in pH 8.5 for 24 h with the initial concentration of 2  $\frac{mg}{ml}$  in this regard, an investigation on the effects of PDA coating upon PVA/PU-polyaniline matrix is an interesting example in which the matrix was immersed into the DA solution under normal mentioned conditions. Scholars reported that the coating enhanced mechanical characteristics; more specifically, the resultant scaffold's tensile strength was  $34.06 \pm 1$  MPa compared to the  $29.51 \pm 1.63$  MPa strength shown without coating. Besides, the surface modification with PDA significantly improved the osseointegration and promoted the adhesion and differentiation of rat bone marrow mesenchymal stem cells (BMSCs), all of which were beneficial for bone TE (Ghorbani et al., 2020a). Nonetheless, the coating time is one of the main parameters influencing the produced PDA layer; in essence, this factor exerts a direct impact on the thickness of PDA. The more the coating time, the more PDA thickness (Li et al., 2009). As an instance, Ou and co-workers (Ou et al., 2009) coated modified silicon (Si) substrates with PDA and concluded that with the prolonging of deposition time, the coating got thicker, and the I<sub>corr</sub> decreased from 247.73  $\frac{A}{cm^2}$  in 2 h to 11.5.45  $\frac{A}{cm^2}$  in 10 h. The impact of coating time was analyzed in a research project to make phosphate glass fiber (PGF) suitable for the PCL/PGF composites formation. Firstly, PGF was dipped in the DA hydrochloride aqueous solution for varying periods (30 min-24 h), followed by the formation of PCL/PGF scaffolds utilizing the *in-situ* polymerization method. With increasing the coating time, the PDA quantity on the fiber surface enhanced; thus, when the time was set to 24 h, the PDA thickness augmented to 46 nm in comparison with 18 nm after 6 h. Furthermore, it was concluded that the combination of annealing and coating processes could yield better results for the bone TE applications compared to the sole coating procedure. Regarding this matter, flexural strength and modulus of annealed/coated, annealed/non-coated, and non-annealed fibers retained at nearly 70-75%, less than 20%, and less than 10% of the day one characteristics (strength: 120 MPa and

| PDA coa<br>paramete      | ating<br>ers | Thickness<br>increment | Thickness reduction | Normal<br>value        | Polymerization<br>rate<br>increase | Roughness<br>increment | Refs   |
|--------------------------|--------------|------------------------|---------------------|------------------------|------------------------------------|------------------------|--|
| Coating time             |              | V                      | -                   | 24 h                   | -                                  | v                      | Li et al. (2009); Ou et al. (2009); Dang et al.<br>(2015); Ghorbani et al. (2020a); Liu et al.<br>(2020); Kim et al. (2021a); Felfel et al.<br>(2021); Zia et al. (2021)   |
| Temperature              |              | 1                      | -                   | Ambient<br>temperature | V                                  | √                      | Ou et al. (2009); Jiang et al. (2011); Proks<br>et al. (2013); Zhou et al. (2014);<br>Schaubroeck et al. (2015); Zain et al.<br>(2015); Deng et al. (2018); Habibi Rad<br>et al. (2018); Malollari et al. (2019);<br>Oymaci et al. (2020); Davidsen et al.<br>(2021) |
| pH of<br>Tris-<br>HCl    | alkaline     | $\checkmark$           | -                   | 8.5                    | $\checkmark$                       | -                      | Yang et al. (2014); Hong et al. (2016);<br>Dong et al. (2018); Hong et al. (2018);<br>Salomäki et al. (2018); Mei et al. (2021);<br>Gan et al. (2022); Szewczyk et al. (2022)  |
|                          | acidic       | -                      | $\checkmark$        | -                      | -                                  | -                      |  |
| DA initial concentration |              | $\checkmark$           | -                   | 2 mg/ml                | 1                                  | $\checkmark$           | Ball et al. (2012); Ding et al. (2014); Hong<br>et al. (2016); Dong et al. (2018); Oymaci<br>et al. (2020); Wu et al. (2020); Li et al.<br>(2021a); Wang et al. (2022)   |

TABLE 1 PDA coating parameters and their effects on the formed PDA layer.

modulus: 14 GPa) after 56 days, respectively. The achieved features associated with annealed-coated fibers successfully met the target for bone repair for the upper extremities at least (Figure 2A) (Felfel et al., 2021).

Multiple studies have suggested the direct influence of coating time on the PDA layer roughness (Ou et al., 2009; Dang et al., 2015; Liu et al., 2020). Concerning this matter, Kim and co-workers engineered high robust PDA-coated polybenzimidazole membranes under different coating times (5, 10, and 30 min). The atomic force microscope image of the PDA layer illustrated that with increasing time, the roughness enhanced from nearly 1 nm in 5 min to 4 nm in 30 min (Kim et al., 2021a).

As an intermediate layer, PDA coating has been employed in various experiments. Recently, Zia et al. (Zia et al., 2021) used a PDA layer between chitosan and porous PLA via applying different polymerization times (1, 6, 12, 18, and 24 h). It was observed that the PDA layer thickness increased with time; to delineate, although some uncovered areas lacking the PDA layer were found after 1 h, a layer with an acceptable thickness appeared when polymerization continued for 6 h. Moreover, the scaffold's diameter and hydrophilicity were both enhanced by time; to elucidate on, the diameter increased from 1.2  $\mu$ m after 6 h to 1.8  $\mu$ m after 24 h, and the water contact angle decreased from 85° at 6 h to 70.6° at 24 h.

### 3.2 Temperature

The process temperature has been shown to improve thickness, roughness, as well as polymerization rate and shorten the polymerization time of the PDA layer. In an investigation conducted by a group of researchers (Zhou et al., 2014), it was illustrated that among various temperatures (25, 37, and 60°C), 60°C and 300  $\frac{r}{min}$  stirring could hasten the time of polymerization (30 min) without triggering chemical changes in the PDA layer compared to the 24-h normal approach. Regarding the comparison between dynamic and static conditions, PDA-coated surfaces prepared under the dynamic condition (stirring) in the range of 1–8 h displayed some superior features like higher thickness and lower contact angle compared to the ones fabricated under the static condition during 24 h. Furthermore, PDA coating endowed the scaffolds with favorable cytocompatibility since cultivated MG-63 osteoblast cells were highly viable.

Another effect of temperature is the increment of the PDA layer's thickness (Zain et al., 2015; Oymaci et al., 2020) and roughness (Schaubroeck et al., 2015; Oymaci et al., 2020), which was carefully assessed in a project. It was reported that the coating thickness on several materials, such as Si/polyvinylidene fluoride (PVDF), was augmented by raising the temperature; to be more precise, thickness increased from nearly 20 nm at 20°C to 65 nm at 45°C. It is also noteworthy to mention that the modified surfaces' water contact angle decreased with augmenting the reaction temperature (from  $63.9^{\circ}$  at  $20^{\circ}$ C to  $53.2^{\circ}$  at  $45^{\circ}$ C). Regarding the roughness, further tests indicated that the layer roughness increased from  $6.4 \pm 0.9$  nm ( $20^{\circ}$ C) to  $10.4 \pm 1.6$  nm ( $45^{\circ}$ C) (Jiang et al., 2011).

Concerning the influence of temperature on the polymerization rate, several studies have reported the rate enhancement with temperature raising (Deng et al., 2018; Habibi Rad et al., 2018). In a novel experiment in which PDA-coated Si surfaces were fabricated, the mentioned effect was investigated within a



#### FIGURE 1

The effects of PDA process parameters on the produced PDA layer, which are investigated in various experiments throughout the years (T: time, C: concentration, S: surface, Th: thickness, Temp: temperature, R: roughness, Au: aurum, Ti: titanium, Si: silicon, PU: polyurethane, PC: polycarbonate, CS: coating speed, PVDF: polyvinylidene fluoride, GR: coating layer growth rate, SS 316L: stainless steel 316L) (Li et al., 2009; Ou et al., 2009; Ball et al., 2012; Zain et al., 2015; Hong et al., 2016; Habibi Rad et al., 2018; Hong et al., 2018; Zhou et al., 2020; Szewczyk et al., 2022).

temperature range of 25–35°C. They found that higher temperature led to the accelerated rate; indeed, polymerization speed increased from 1.8  $\frac{nm}{hr}$  at 25°C to 2.2  $\frac{nm}{hr}$  at 35°C (Ou et al., 2009).

Improving the coating stability can be achieved by the coating's thermal treatment. In essence, the thermal treatment affects the number of quinone and amine groups at the PDA surface (Proks et al., 2013; Malollari et al., 2019). Within an intriguing study, titanium (Ti)/Si wafer samples were coated via PDA; specifically, some were thermally treated by storing at 121°C for 24 h, while others were post-treated by 2-week storing at room temperature. In both treatments, the numbers of the quinone and primary amine groups increased and decreased, respectively, improving layer stability and favoring cellular proliferation. The proliferation of fibroblast, human keratinocytes (HaCaT), and MC3T3-E1 cells on post-treated PDA-coated samples was more enhanced in comparison with the PDA-coated ones (Figure 2B) (Davidsen et al., 2021).

#### 3.3 pH of Tris-HCl

In the synthesis process of PDA coating, various experiments have reported the creation of a weak alkaline environment (pH > 8.5) with oxygen as the oxidant in the Tris-HCl buffer presence. When oxygen is constantly present as an oxidant in the reaction, an even coating of PDA is formed. As another synthesis method, creating an oxygen gradient can lead to the formation of a PDA gradient coating (Lee et al., 2019). In some synthesis approaches with other oxidants, the pH may vary between weakly acidic and neutral (Wei et al., 2010; Bernsmann et al., 2011; Ghorbani et al., 2021). Furthermore, PDA can be formed via self-polymerization within an acidic environment employing hydrothermal methods (160°C in 6 h) (Zheng et al., 2015).

The pH value directly affects the PDA layer thickness (Hong et al., 2018) since the initial DA oxidation depends on the amount of pH (Yang et al., 2014; Salomäki et al., 2018). Regarding this subject, the influence of pH on the PDA two-dimensional films' thickness was examined by choosing pH values of 7.5, 8.0, and 8.5. Interestingly, the lowest thickness (24 nm) was achieved at pH 7.5, whereas the highest one (35 nm) was obtained at pH 8.5. However, the most favorable result was found in pH 8.0 because of the fewer variations between the central and external regions of the film (Figure 2C) (Szewczyk et al., 2022).

Concerning the impact of pH on the polymerization rate, multiple investigations have revealed that the rate increases with the pH increment (Dong et al., 2018; Mei et al., 2021). Within an innovative project, sprayable PDA with ultra-fast polymerization was employed to coat solid surfaces. In this regard, the pH was selected in the range of 7–11, and when it went beyond 10.1, no PDA layer could be formed. Nevertheless, the polymerization speed enhanced from 1.48  $\frac{A^{\circ}}{\min}$  at pH 7.4 to 2.6  $\frac{A^{\circ}}{\min}$  at pH 9–10.1 (Hong et al., 2016).

Given that better outcomes concerning the PDA polymerization have been produced in the alkaline environment, most studies have concentrated on this pH range. For example, as a modification factor, PDA was grafted onto the hyaluronic acid (HA) chains under a pH value of 8.0, followed by the production of hydrogel scaffolds through incorporating PDA/HA complex in a dual cross-linked collagen type I (Col I) matrix for use in cartilage regeneration. The final results showed that the employed PDA coating enhanced BMSCs' proliferation, improved cellular affinity, and induced chondrogenic differentiation of cells compared to those of the cell-seeded scaffolds without the PDA layer. Additionally, it augmented anti-inflammation capacity, immune modulation ability, and *in-vivo* cartilage repair in the rabbit model (Gan et al., 2022).

#### 3.4 Concentration

Like other discussed parameters, concentration is another critical factor impacting the final PDA layer. Of note, the concentration of the DA monomer must be at least 2  $\frac{mg}{ml}$  to achieve a layer of PDA. In a novel project, magnesium discs were coated with a calcium-deficient HA (CDHA) layer using hydrothermal treatment. Afterward, vascular endothelial growth factor was bound on the CDHA surface through PDA's covalent bonds (2  $\frac{mg}{ml}$  and pH 8.5). It was concluded that the surface morphology altered when coating with the PDA layer was smoother, and the coating decreased the rate of magnesium degradation; therefore, a desirable environment was created for the growth factor, indicating the good biocompatibility of this scaffold for prolonged usage (Li et al., 2021a).

For investigating the effect of concentration on the PDA layer, the porous Col I-HA scaffold was modified via immersing into various DA concentrations (0.5, 1.0, and  $2.0 \frac{mg}{ml}$ ) prepared in 10 mM Tris-HCl buffer solution at ambient temperature. All coated sponges maintained their porous structure, and the scaffold with 2.0  $\frac{mg}{ml}$  DA concentration had the highest swelling rate, largest reactive oxygen species scavenging capacity, excellent antioxidant property resulting from the PDA's catechol groups, good procoagulant effect, and higher promotion of diabetic wound healing in the rat model compared to other groups (Wang et al., 2022).

With respect to the effect of concentration on the polymerization rate, the rate boosts with the concentration increment (Hong et al., 2016). For instance, in the DA concentration range of  $0.1-5 \frac{g}{L}$  the highest polymerization speed,  $4 \frac{nm}{hr}$ , was obtained in the concentration of  $5 \frac{g}{s}$  (Ball et al., 2012).

The thickness and roughness of the PDA layer can be controlled by tuning this factor; regarding this matter, several studies have stated that lower concentrations result in lower PDA's thickness (Dong et al., 2018; Wu et al., 2020) and roughness (Ball et al., 2012; Ding et al., 2014). To evaluate these phenomena, a team of scientists (Oymaci et al., 2020) coated the polyethersulfone membranes with PDA possessing



#### FIGURE 2

(A) The influence of PDA coating time upon roughness and thickness of the PDA layer on the PGF (reproduced content is open access) (Felfel et al., 2021). (B) Post-treatment of the PDA layer present on the PDA-coated Ti samples and the quantification of fibroblast, HaCaT, and MC3T3-E1 cells on the surfaces (reproduced content is open access) (Davidsen et al., 2021). (C) The different pH values of Tris buffer and their effects on the film thickness illustrated in the comparative chart (I), mapping chart (II), and line chart (III) (reproduced content is open access) (Szewczyk et al., 2022). (D) The DA concentration impact on the morphology of coating displayed by field emission scanning electron microscopy images. Surface and cross-section images of (I,V) non-coated polyethersulfone membranes as well as membranes coated with PDA possessing DA concentrations of (II,VI) 0.5  $\frac{9}{L}$ , (III,VII) 2.0  $\frac{9}{L}$  and (IV,VIII) 3.0  $\frac{9}{L}$  at 10<sup>5</sup> X and 25 × 10<sup>4</sup> X magnifications, respectively (reproduced content is open access) (Oymaci et al., 2020).

diverse concentrations  $(0.5-3.0 \frac{g}{L})$ , and it was demonstrated that the coating thickness enhanced with the DA concentration increment. Specifically, using the 0.5  $\frac{g}{L}$  DA solution resulted in a thin coating layer formation which was not readily visible, while a PDA layer with higher thickness was produced employing DA solution with the concentration of 3.0  $\frac{g}{L}$ . Furthermore, the layer roughness increased when the concentration of DA was augmented, as observed in the cross-section images in Figure 2D.

## 4 Conclusion

According to the ability of PDA as a surface modifier, this nature-inspired material has been widely employed in TE applications. Notably, certain conditions and parameters should be considered for the PDA polymerization process, including coating time, temperature, pH of Tris-HCl, and initial DA concentration, to acquire beneficial results. Even though the mentioned parameters have been given fixed values (24 h, room temperature, pH 8.5, and 2  $\frac{mg}{ml}$ ) in most investigations, some researchers think outside the box and change the factors to regulate the coating layer's features.

There is still a long way to go regarding translating these innovative coated biomaterials into the clinic; consequently, successfully managing parameters together for furnishing the modified scaffolds with the desired properties has become a prominent goal in the preclinical experiments.

# Author contributions

Conceptualization, SS; Methodology, SS and MKH; Validation, SS, MKH, DL, and ND; Investigation, SS, MKH,

#### References

Abdollahi, J., Davari, N., Panahi, Y., and Gardaneh, M. (2022). Detection of metastatic breast cancer from whole-slide pathology images using an ensemble deep-learning method. *Archives Breast Cancer* 9 (3), 364–376. doi:10.32768/abc.202293364-376

Aguilar-Rabiela, A. E., Leal-Egaña, A., Nawaz, Q., and Boccaccini, A. R. (2021). Integration of mesoporous bioactive glass nanoparticles and curcumin into PHBV microspheres as biocompatible composite for drug delivery applications. *Molecules* 26 (11), 3177. doi:10.3390/molecules26113177

Arab-Ahmadi, S., Irani, S., Bakhshi, H., Atyabi, F., and Ghalandari, B. (2021). Immobilization of carboxymethyl chitosan/laponite on polycaprolactone nanofibers as osteoinductive bone scaffolds. *Polym. Adv. Technol.* 32 (2), 755–765. doi:10.1002/ pat.5128

Ball, V., Frari, D., Toniazzo, V., and Ruch, D. (2012). Kinetics of polydopamine film deposition as a function of pH and dopamine concentration: Insights in the polydopamine deposition mechanism. *J. Colloid Interface Sci.* 386 (1), 366–372. doi:10.1016/j.jcis.2012.07.030

Batul, R., Tamanna, T., Khaliq, A., and Yu, A. (2017). Recent progress in the biomedical applications of polydopamine nanostructures. *Biomater. Sci.* 5 (7), 1204–1229. doi:10.1039/C7BM00187H

Bernsmann, F., Ball, V., Addiego, F., Ponche, A., Michel, M., Gracio, J. J. d. A., et al. (2011). Dopamine–Melanin film deposition depends on the used oxidant and buffer solution. *Langmuir* 27 (6), 2819–2825. doi:10.1021/la104981s

DL, and ND; Resources, SS and MKH; Data Curation, SS, MKH, DL, and ND; Writing "original draft preparation, SS, MKH, DL, and ND; Writing " review and editing, BY; Visualization, MKH; Supervision, BY; Project Administration, BY; Funding Acquisition, BY. All authors have read and agreed to the published version of the manuscript.

# Funding

This work was supported by the Outstanding Clinical Discipline Project of Shanghai Pudong (Grant No. PWYgy 2021-08), the Joint project of Shanghai Pudong New Area Health Commission (PW2021D-12), and the National Natural Science Foundation of China (Grant No. 81971753, 82170897).

# Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

# Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Bock, N., Pham, T. L.-B., Nguyen, T. B., Tran, H. A., and Tran, P. A. (2020). Polydopamine coating of uncrosslinked chitosan as an acellular scaffold for full thickness skin grafts. *Carbohydr. Polym.* 245, 116524. doi:10.1016/j.carbpol.2020.116524

Cunniffe, G. M., Díaz-Payno, P. J., Sheehy, E. J., Critchley, S. E., Almeida, H. V., Pitacco, P., et al. (2019). Tissue-specific extracellular matrix scaffolds for the regeneration of spatially complex musculoskeletal tissues. *Biomaterials* 188, 63–73. doi:10.1016/j.biomaterials.2018.09.044

Dang, Y., Xing, C.-M., Quan, M., Wang, Y. B., Zhang, S. P., Shi, S. Q., et al. (2015). Substrate independent coating formation and anti-biofouling performance improvement of mussel inspired polydopamine. *J. Mat. Chem. B* 3 (20), 4181–4190. doi:10.1039/C5TB00341E

Davari, N., Bakhtiary, N., Khajehmohammadi, M., Sarkari, S., Tolabi, H., Ghorbani, F., et al. (2022). Protein-based hydrogels: Promising materials for tissue engineering. *Polymers* 14 (5), 986. doi:10.3390/polym14050986

Davidsen, M. B., Teixeira, J. F. L., Dehli, J., Karlsson, C., Kraft, D., Souza, P. P., et al. (2021). Post-treatments of polydopamine coatings influence cellular response. *Colloids Surfaces B Biointerfaces* 207, 111972. doi:10.1016/j.colsurfb. 2021.111972

Deng, Z., Shang, B., and Peng, B. (2018). Polydopamine based colloidal materials: Synthesis and applications. *Chem. Rec.* 18 (4), 410–432. doi:10.1002/tcr.201700051

Deng, Z., Wang, W., Xu, X., Nie, Y., Liu, Y., Gould, O. E. C., et al. (2021). Biofunction of polydopamine coating in stem cell culture. *ACS Appl. Mat. Interfaces* 13 (9), 10748–10759. doi:10.1021/acsami.0c22565

Ding, Y., Weng, L.-T., Yang, M., Yang, Z., Lu, X., Huang, N., et al. (2014). Insights into the aggregation/deposition and structure of a polydopamine film. *Langmuir* 30 (41), 12258–12269. doi:10.1021/la5026608

Dong, L., Liu, X., Xiong, Z., Sheng, D., Lin, C., Zhou, Y., et al. (2018). Fabrication of highly efficient ultraviolet absorbing PVDF membranes via surface polydopamine deposition. *J. Appl. Polym. Sci.* 135 (4), 45746. doi:10.1002/app. 45746

Du, Q., An, J., Li, J., Zhou, L., Li, N., and Wang, X. (2017). Polydopamine as a new modification material to accelerate startup and promote anode performance in microbial fuel cells. *J. Power Sources* 343, 477–482. doi:10.1016/j.jpowsour.2017. 01.093

Feinberg, H., and Hanks, T. W. (2022). Polydopamine: A bioinspired adhesive and surface modification platform. *Polym. Int.* 71 (5), 578-582. doi:10.1002/pi.6358

Felfel, R. M., Parsons, A. J., Chen, M., Stuart, B. W., Wadge, M. D., and Grant, D. M. (2021). Water resistant fibre/matrix interface in a degradable composite: Synergistic effects of heat treatment and polydopamine coating. *Compos. Part A Appl. Sci. Manuf.* 146, 106415. doi:10.1016/j.compositesa.2021.106415

Gan, D., Jiang, Y., Hu, Y., Wang, X., Wang, Q., Wang, K., et al. (2022). Musselinspired extracellular matrix-mimicking hydrogel scaffold with high cell affinity and immunomodulation ability for growth factor-free cartilage regeneration. *J. Orthop. Transl.* 33, 120–131. doi:10.1016/j.jot.2022.02.006

Gennadios, A. (2002). Protein-based films and coatings. 1st Ed. Boston: CRC Press. doi:10.1201/9781420031980

Ghalandari, B., Yu, Y., Ghorbani, F., Warden, A. R., Ahmad, K. Z., and Sang, X. (2021). Polydopamine nanospheres coated with bovine serum albumin permit enhanced cell differentiation: Fundamental mechanism and practical application for protein coating formation. *Nanoscale* 47. doi:10.1039/D1NR07469E

Ghorbani, F., Ghalandari, B., Khan, A. L., Li, D., Zamanian, A., and Yu, B. (2020a). Decoration of electrical conductive polyurethane-polyaniline/polyvinyl alcohol matrixes with mussel-inspired polydopamine for bone tissue engineering. *Biotechnol. Prog.* 36 (6), e3043. doi:10.1002/btpr.3043

Ghorbani, F., Ghalandari, B., and Liu, C. (2021). A facile method to synthesize 3D pomegranate-like polydopamine microspheres. *Front. Bioeng. Biotechnol.* 9, 737074. doi:10.3389/fbioe.2021.737074

Ghorbani, F., Kim, M., Monavari, M., Ghalandari, B., and Boccaccini, A. R. (2022). Mussel-inspired polydopamine decorated alginate dialdehyde-gelatin 3D printed scaffolds for bone tissue engineering application. *Front. Bioeng. Biotechnol.* 10, 940070. doi:10.3389/fbioe.2022.940070

Ghorbani, F., Zamanian, A., and Aidun, A. (2019a). Bioinspired polydopamine coating-assisted electrospun polyurethane-graphene oxide nanofibers for bone tissue engineering application. *J. Appl. Polym. Sci.* 136 (24), 47656. doi:10.1002/app.47656

Ghorbani, F., Zamanian, A., Behnamghader, A., and Daliri-Joupari, M. (2019c). Bone-like hydroxyapatite mineralization on the bio-inspired PDA nanoparticles using microwave irradiation. *Surfaces Interfaces* 15, 38–42. doi:10.1016/j.surfin. 2019.01.007

Ghorbani, F., Zamanian, A., Behnamghader, A., and Joupari, M. D. (2019b). A facile method to synthesize mussel-inspired polydopamine nanospheres as an active template for *in situ* formation of biomimetic hydroxyapatite. *Mater. Sci. Eng. C* 94, 729–739. doi:10.1016/j.msec.2018.10.010

Ghorbani, F., Zamanian, A., Kermanian, F., and Shamoosi, A. (2020b). A bioinspired 3D shape olibanum-collagen-gelatin scaffolds with tunable porous microstructure for efficient neural tissue regeneration. *Biotechnol. Prog.* 36 (1), e2918–12. doi:10.1002/btpr.2918

Ghorbani, F., Zamanian, A., and Sahranavard, M. (2020c). Mussel-inspired polydopamine-mediated surface modification of freeze-cast poly (ε-caprolactone) scaffolds for bone tissue engineering applications. *Biomed. Eng./ Biomed. Tech.* 65 (3), 273–287. doi:10.1515/bmt-2019-0061

Ghorbani, F., Zamanian, A., and Torabinejad, B. (2020d). The effect of oxygen plasma pretreatment on the properties of mussel-inspired polydopamine-decorated polyurethane nanofibers. *J. Polym. Eng.* 40 (2), 109–119. doi:10.1515/polyeng-2019-0219

Gregory, D. A., Taylor, C. S., Fricker, A. T. R., Asare, E., Tetali, S. S., Haycock, J. W., et al. (2022). Polyhydroxyalkanoates and their advances for biomedical applications. *Trends Mol. Med.* 28 (4), 331–342. doi:10.1016/j.molmed.2022. 01.007

Habibi Rad, M., Zamanian, A., Hadavi, S. M. M., and Khanlarkhani, A. (2018). A two-stage kinetics model for polydopamine layer growth. *Macromol. Chem. Phys.* 219 (9), 1700505. doi:10.1002/macp.201700505

Hong, S. H., Hong, S., Ryou, M.-H., Choi, J. W., Kang, S. M., and Lee, H. (2016). Sprayable ultrafast polydopamine surface modifications. *Adv. Mat. Interfaces* 3 (11), 1500857. doi:10.1002/admi.201500857

Hong, S., Na, Y. S., Choi, S., Song, I. T., Kim, W. Y., and Lee, H. (2012). Noncovalent self-assembly and covalent polymerization Co-contribute to polydopamine formation. *Adv. Funct. Mat.* 22 (22), 4711–4717. doi:10.1002/ adfm.201201156

Hong, S., Wang, Y., Park, S. Y., and Lee, H. (2018). Progressive fuzzy cation- $\pi$  assembly of biological catecholamines. *Sci. Adv.* 4 (9), eaat7457. doi:10.1126/sciadv. aat7457

Honmane, S. M., Charde, M. S., Salunkhe, S. S., Choudhari, P. B., and Nangare, S. N. (2022). Polydopamine surface-modified nanocarriers for improved anticancer activity: Current progress and future prospects. *OpenNano* 7, 100059. doi:10.1016/j. onano.2022.100059

Huang, J., Xiong, J., Wang, D., Zhang, J., Yang, L., Sun, S., et al. (2021). 3D bioprinting of hydrogels for cartilage tissue engineering. *Gels* 7 (3), 144. doi:10. 3390/gels7030144

Huang, L., Liu, M., Huang, H., Wen, Y., Zhang, X., and Wei, Y. (2018). Recent advances and progress on melanin-like materials and their biomedical applications. *Biomacromolecules* 19 (6), 1858–1868. doi:10.1021/acs.biomac.8b00437

Huang, S., Liang, N., Hu, Y., Zhou, X., and Abidi, N. (2016). Polydopamine-Assisted surface modification for bone biosubstitutes. *Biomed. Res. Int.* 2016, 2389895. doi:10.1155/2016/2389895

Jenkins, T. L., and Little, D. (2019). Synthetic scaffolds for musculoskeletal tissue engineering: Cellular responses to fiber parameters. *npj Regen. Med.* 4 (1), 15. doi:10.1038/s41536-019-0076-5

Ji, M., and Parquette, J. R. (2020). Enhanced stability of peptide nanofibers coated with a conformal layer of polydopamine. *Chem. Eur. J.* 26 (39), 8572–8578. doi:10. 1002/chem.202000403

Jia, L., Han, F., Wang, H., Zhu, C., Guo, Q., Li, J., et al. (2019). Polydopamineassisted surface modification for orthopaedic implants. *J. Orthop. Transl.* 17, 82–95. doi:10.1016/j.jot.2019.04.001

Jiang, J., Zhu, L., Zhu, L., Zhu, B., and Xu, Y. (2011). Surface characteristics of a self-polymerized dopamine coating deposited on hydrophobic polymer films. *Langmuir* 27 (23), 14180–14187. doi:10.1021/la202877k

Jiang, X., Wang, Y., and Li, M. (2014). Selecting water-alcohol mixed solvent for synthesis of polydopamine nano-spheres using solubility parameter. *Sci. Rep.* 4 (1), 6070. doi:10.1038/srep06070

Kao, C.-T., Chen, Y.-J., Ng, H.-Y., Lee, A., Huang, T. H., Lin, T. F., et al. (2018). Surface modification of calcium silicate via mussel-inspired polydopamine and effective adsorption of extracellular matrix to promote osteogenesis differentiation for bone tissue engineering. *Materials* 11 (9), 1664. doi:10.3390/ma11091664

Kaushik, N., Nhat Nguyen, L., Kim, J. H., Choi, E. H., and Kumar Kaushik, N. (2020). Strategies for using polydopamine to induce biomineralization of hydroxyapatite on implant materials for bone tissue engineering. *Int. J. Mol. Sci.* 21 (18), 6544. doi:10.3390/ijms21186544

Kehr, N. S., Atay, S., and Ergün, B. (2015). Self-assembled monolayers and nanocomposite hydrogels of functional nanomaterials for tissue engineering applications. *Macromol. Biosci.* 15 (4), 445–463. doi:10.1002/mabi.201400363

Kim, S. D., Won, G. Y., Shah, A. A., Park, A., Park, Y. I., Nam, S. E., et al. (2021a). Reinforcing the polybenzimidazole membrane surface by an ultrathin cocrosslinked polydopamine layer for organic solvent nanofiltration applications. *J. Membr. Sci.* 636, 119587. doi:10.1016/j.memsci.2021.119587

Kim, S. Y., Park, C. S., Hosseini, S., Lampert, J., Kim, Y. J., and Nazar, L. F. (2021b). Inhibiting oxygen release from Li-rich, Mn-rich layered oxides at the surface with a solution processable oxygen scavenger polymer. *Adv. Energy Mat.* 11 (30), 2100552. doi:10.1002/aenm.202100552

Kopeć, K., Wojasiński, M., Eichler, M., Genc, H., Friedrich, R. P., Stein, R., et al. (2022). Polydopamine and gelatin coating for rapid endothelialization of vascular scaffolds. *Biomater. Adv.* 134, 112544. doi:10.1016/j.msec.2021.112544

Lee, H. A., Ma, Y., Zhou, F., and Hong, S. (2019). Material-Independent surface chemistry beyond polydopamine coating. *Acc. Chem. Res.* 52 (3), 704–713. doi:10. 1021/acs.accounts.8b00583

Lee, H. A., Park, E., and Lee, H. (2020). Polydopamine and its derivative surface chemistry in material science: A focused review for studies at kaist. *Adv. Mat.* 32 (35), 1907505. doi:10.1002/adma.201907505

Li, B., Liu, W., Jiang, Z., Dong, X., Wang, B., and Zhong, Y. (2009). Ultrathin and stable Active layer of dense composite membrane enabled by poly(dopamine). *Langmuir* 25 (13), 7368–7374. doi:10.1021/la900262p

Li, J.-H., Ni, X.-X., Zhang, D.-B., Zheng, H., Wang, J. B., and Zhang, Q. Q. (2018). Engineering a self-driven PVDF/PDA hybrid membranes based on membrane micro-reactor effect to achieve super-hydrophilicity, excellent antifouling properties and hemocompatibility. *Appl. Surf. Sci.* 444, 672–690. doi:10.1016/j. apsusc.2018.03.034

Li, J., Cao, F., Wu, B., Yang, J., Xu, W., Wang, W., et al. (2021a). Immobilization of bioactive vascular endothelial growth factor onto Ca-de fi cient hydroxyapatite-coated Mg by covalent bonding using polydopamine. *J. Orthop. Transl.* 30, 82–92. doi:10.1016/j.jot.2021.06.002

Li, S., Hou, X., Lu, S., Xu, W., Tao, J., Zhao, Z., et al. (2021b). Fabrication and simulation of a layered ultrahigh thermal conductive material made of self-assembled graphene and polydopamine on a copper substrate. *RSC Adv.* 11 (55), 34676–34687. doi:10.1039/D1RA05252G

Lin, Y., Zhang, L., Liu, N. Q., Yao, Q., Van Handel, B., Xu, Y., et al. (2019). *In vitro* behavior of tendon stem/progenitor cells on bioactive electrospun nanofiber membranes for tendon-bone tissue engineering applications. *Int. J. Nanomedicine* 14, 5831–5848. doi:10.2147/IJN.S210509

Liu, M., Zeng, G., Wang, K., Wan, Q., Tao, L., Zhang, X., et al. (2016). Recent developments in polydopamine: An emerging soft matter for surface modification and biomedical applications. *Nanoscale* 8 (38), 16819–16840. doi:10.1039/C5NR09078D

Liu, W., Sun, J., Sun, Y., Xiang, Y., Yan, Y., Han, Z., et al. (2020). Multifunctional injectable protein-based hydrogel for bone regeneration. *Chem. Eng. J.* 394, 124875. doi:10.1016/j.cej.2020.124875

Liu, Y., Ai, K., and Lu, L. (2014). Polydopamine and its derivative materials: Synthesis and promising applications in energy, environmental, and biomedical fields. *Chem. Rev.* 114 (9), 5057–5115. doi:10.1021/cr400407a

Lynge, M. E., van der Westen, R., Postma, A., and Stadler, B. (2011). Polydopamine—A nature-inspired polymer coating for biomedical science. *Nanoscale* 3 (12), 4916. doi:10.1039/c1nr10969c

Malollari, K. G., Delparastan, P., Sobek, C., Vachhani, S. J., Fink, T. D., Zha, R. H., et al. (2019). Mechanical enhancement of bioinspired polydopamine nanocoatings. *ACS Appl. Mat. Interfaces* 11 (46), 43599–43607. doi:10.1021/acsami.9b15740

Mei, H., Gao, Z., Wang, Q., Sun, H., Zhao, K., Zhang, P., et al. (2021). Ultrasound expands the versatility of polydopamine coatings. *Ultrason. Sonochemistry* 74, 105571. doi:10.1016/j.ultsonch.2021.105571

Mei, S., Xu, X., Priestley, R. D., and Lu, Y. (2020). Polydopamine-based nanoreactors: Synthesis and applications in bioscience and energy materials. *Chem. Sci.* 11 (45), 12269–12281. doi:10.1039/D0SC04486E

Merceron, T. K., Burt, M., Seol, Y.-J., Kang, H. W., Lee, S. J., Yoo, J. J., et al. (2015). A 3D bioprinted complex structure for engineering the muscle-tendon unit. *Biofabrication* 7 (3), 035003. doi:10.1088/1758-5090/7/3/035003

Montazerian, M., Hosseinzadeh, F., Migneco, C., Fook, M. V., and Baino, F. (2022). Bioceramic coatings on metallic implants: An overview. *Ceram. Int.* 48 (7), 8987–9005. doi:10.1016/j.ceramint.2022.02.055

Niculescu, A.-G., Chircov, C., Bîrcă, A. C., and Grumezescu, A. M. (2021). Fabrication and applications of microfluidic devices: A review. *Int. J. Mol. Sci.* 22 (4), 2011. doi:10.3390/ijms22042011

Nieto, C., Marcelo, G., Vega, M., and Martin del Valle, E. M. (2021). Antineoplastic behavior of polydopamine nanoparticles prepared in different water/alcohol media. *Colloids Surfaces B Biointerfaces* 199, 111506. doi:10.1016/j. colsurfb.2020.111506

Nishida, H., and Koga, F. (2018). Potential of oxidative polymerization coating of cellulose nano-fiber by dopamine. *IOP Conf. Ser. Mat. Sci. Eng.* 368, 012043. doi:10. 1088/1757-899X/368/1/012043

Ou, J., Wang, J., Liu, S., Zhou, J., Ren, S., and Yang, S. (2009). Microtribological and electrochemical corrosion behaviors of polydopamine coating on APTS-SAM modified Si substrate. *Appl. Surf. Sci.* 256 (3), 894–899. doi:10.1016/j.apsusc.2009. 08.081

Oymaci, P., Nijmeijer, K., and Borneman, Z. (2020). Development of polydopamine forward osmosis membranes with low reverse salt flux. *Membranes* 10 (5), 94. doi:10.3390/membranes10050094

Pagel, M., and Beck-Sickinger, A. G. (2017). Multifunctional biomaterial coatings: Synthetic challenges and biological activity. *Biol. Chem.* 398 (1), 3–22. doi:10.1515/ hsz-2016-0204

Park, J., Lee, S. J., Jung, T. G., Lee, J. H., Kim, W. D., Lee, J. Y., et al. (2021). Surface modification of a three-dimensional polycaprolactone scaffold by polydopamine, biomineralization, and BMP-2 immobilization for potential bone tissue applications. *Colloids Surfaces B Biointerfaces* 199, 111528. doi:10.1016/j. colsurfb.2020.111528

Perez-Puyana, V., Wieringa, P., Yuste, Y., Portilla, F., Guererro, A., Romero, A., et al. (2021). Fabrication of hybrid scaffolds obtained from combinations of PCL with gelatin or collagen via electrospinning for skeletal muscle tissue engineering. *J. Biomed. Mat. Res. A* 109 (9), 1600–1612. doi:10.1002/jbm.a.37156 Proks, V., Brus, J., Pop-Georgievski, O., Vecernikova, E., Wisniewski, W., Kotek, J., et al. (2013). Thermal-induced transformation of polydopamine structures: An efficient route for the stabilization of the polydopamine surfaces. *Macromol. Chem. Phys.* 214 (4), 499–507. doi:10.1002/macp.201200505

Qian, Y., Zhao, X., Han, Q., Chen, W., Li, H., and Yuan, W. (2018). An integrated multi-layer 3D-fabrication of PDA/RGD coated graphene loaded PCL nanoscaffold for peripheral nerve restoration. *Nat. Commun.* 9 (1), 323. doi:10.1038/s41467-017-02598-7

Razaviye, M. K., Tafti, R. A., and Khajehmohammadi, M. (2022). An investigation on mechanical properties of PA12 parts produced by a SLS 3D printer: An experimental approach. *CIRP J. Manuf. Sci. Technol.* 38, 760–768. doi:10.1016/j. cirpj.2022.06.016

Richbourg, N. R., Peppas, N. A., and Sikavitsas, V. I. (2019). Tuning the biomimetic behavior of scaffolds for regenerative medicine through surface modifications. J. Tissue Eng. Regen. Med. 13 (8), 1275–1293. doi:10.1002/term.2859

Rizzo, F., and Kehr, N. S. (2021). Recent advances in injectable hydrogels for controlled and local drug delivery. *Adv. Healthc. Mat.* 10 (1), 2001341. doi:10.1002/adhm.202001341

Rodríguez-Rodríguez, R., Espinosa-Andrews, H., Velasquillo-Martínez, C., and Garcia-Carvajal, Z. Y. (2020). Composite hydrogels based on gelatin, chitosan and polyvinyl alcohol to biomedical applications: A review. *Int. J. Polym. Mater. Polym. Biomaterials* 69 (1), 1–20. doi:10.1080/00914037.2019.1581780

Saeed, M., Beigi-Boroujeni, S., Rajabi, S., Rafati Ashteiani, G., Dolatfarahi, M., and Ozcan, M. (2021). A simple, green chemistry technology for fabrication of tissueengineered scaffolds based on mussel-inspired 3D centrifugal spun. *Mater. Sci. Eng.* C 121, 111849. doi:10.1016/j.msec.2020.111849

Salatin, S., Maleki Dizaj, S., and Yari Khosroushahi, A. (2015). Effect of the surface modification, size, and shape on cellular uptake of nanoparticles. *Cell Biol. Int.* 39 (8), 881–890. doi:10.1002/cbin.10459

Salomäki, M., Marttila, L., Kivelä, H., Ouvinen, T., and Lukkari, J. (2018). Effects of pH and oxidants on the first steps of polydopamine formation: A thermodynamic approach. J. Phys. Chem. B 122 (24), 6314–6327. doi:10.1021/acs.jpcb.8b02304

Schaubroeck, D., Mader, L., Dubruel, P., and Vanfleteren, J. (2015). Surface modification of an epoxy resin with polyamines and polydopamine: Adhesion toward electroless deposited copper. *Appl. Surf. Sci.* 353, 238–244. doi:10.1016/j. apsusc.2015.06.114

Sharma, D., Jia, W., Long, F., Pati, S., Chen, Q., Qyang, Y., et al. (2019). Polydopamine and collagen coated micro-grated polydimethylsiloxane for human mesenchymal stem cell culture. *Bioact. Mater.* 4, 142–150. doi:10.1016/j. bioactmat.2019.02.002

Szewczyk, J., Pochylski, M., Szutkowski, K., Kempinski, M., Mrowczynski, R., Iatsunskyi, I., et al. (2022). *In-situ* thickness control of centimetre-scale 2D-Like polydopamine films with large scalability. *Mater. Today Chem.* 24, 100935. doi:10. 1016/j.mtchem.2022.100935

Terzopoulou, Z., Zamboulis, A., Koumentakou, I., Michailidou, G., Noordam, M. J., and Bikiaris, D. N. (2022). Biocompatible synthetic polymers for tissue engineering purposes. *Biomacromolecules* 23 (5), 1841–1863. doi:10.1021/acs. biomac.2c00047

Wang, Y., Chen, L., Ren, D.-Y., Feng, Z. X., Zhang, L. Y., Zhong, Y. F., et al. (2022). Mussel-inspired collagen-hyaluronic acid composite scaffold with excellent antioxidant properties and sustained release of a growth factor for enhancing diabetic wound healing. *Mater. Today Bio* 15, 100320. doi:10.1016/j.mtbio.2022. 100320

Wei, Q., Zhang, F. L., Li, J., Li, B., and Zhao, C. (2010). Oxidant-induced dopamine polymerization for multifunctional coatings. *Polym. Chem.* 1 (9), 1430–1433. doi:10.1039/c0py00215a

Wu, M., Wang, T., Müller, L., and Muller, F. A. (2020). Adjustable synthesis of polydopamine nanospheres and their nucleation and growth. *Colloids Surfaces A Physicochem. Eng. Aspects* 603, 125196. doi:10.1016/j.colsurfa. 2020.125196

Yan, J., Wu, R., Liao, S., Jiang, M., and Qian, Y. (2020). Applications of polydopamine-modified scaffolds in the peripheral nerve tissue engineering. *Front. Bioeng. Biotechnol.* 8, 590998–591007. doi:10.3389/fbioe.2020.590998

Yan, J., Yang, L., Lin, M., Ma, J., Lu, X., and Lee, P. S. (2013). Polydopamine spheres as active templates for convenient synthesis of various nanostructures. *Small* 9 (4), 596–603. doi:10.1002/smll.201201064

Yang, C., Huang, X., Huang, Y., Chen, Y., Wang, L., Zheng, X., et al. (2019). Characterization and *in vitro* experiments of composite membrane materials that polydopamine-loaded on the surface of collagen modified by a novel nanomaterial graphene oxide. *Int. J. Polym. Analysis Charact.* 24 (8), 741–751. doi:10.1080/ 1023666X.2019.1670396 Yang, H.-C., Luo, J., Lv, Y., Shen, P., and Xu, Z. K. (2015). Surface engineering of polymer membranes via mussel-inspired chemistry. *J. Membr. Sci.* 483, 42–59. doi:10.1016/j.memsci.2015.02.027

Yang, J., Cohen Stuart, M. A., and Kamperman, M. (2014). Jack of all trades: Versatile catechol crosslinking mechanisms. *Chem. Soc. Rev.* 43 (24), 8271–8298. doi:10.1039/C4CS00185K

Yazdi, M. K., Zare, M., Khodadadi, A., Seidi, F., Sajadi, S. M., Zarrintaj, P., et al. (2022). Polydopamine biomaterials for skin regeneration. *ACS Biomater. Sci. Eng.* 8 (6), 2196–2219. doi:10.1021/acsbiomaterials.1c01436

Yim, W., Borum, R. M., Zhou, J., Mantri, Y., Wu, Z., Zhou, J. V. et al. (2022). Ultrasmall gold nanorod-polydopamine hybrids for enhanced photoacoustic imaging and photothermal therapy in second near-infrared window. *Nanotheranostics* 6 (1), 79–90. doi:10.7150/ntno.63634

Yin, P., Liu, Y., Xiao, L., and Zhang, C. (2021). Advanced metallic and polymeric coatings for neural interfacing: Structures, properties and tissue responses. *Polymers* 13 (16), 2834. doi:10.3390/polym13162834

Yu, B., Pei, P., Yu, B., Li, D., Zhang, X., Huang, J., et al. (2017). Enhance the bioactivity and osseointegration of the polyethylene-terephthalate-based artificial ligament via poly(dopamine) coating with mesoporous bioactive glass. *Adv. Eng. Mat.* 19 (5), 1600708. doi:10.1002/adem.201600708

Zain, N. M., Hussain, R., and Abdul Kadir, M. R. (2015). Quinone-rich polydopamine functionalization of yttria stabilized zirconia for apatite biomineralization: The effects of coating temperature. *Appl. Surf. Sci.* 346, 317–328. doi:10.1016/j.apsusc.2015.04.007

Zeng, J., Wang, Y., Sun, Z., Chang, H., Cao, M., Zhao, J., et al. (2020). A novel biocompatible PDA/IR820/DAP coating for antibiotic/photodynamic/ photothermal triple therapy to inhibit and eliminate Staphylococcus aureus biofilm. *Chem. Eng. J.* 394, 125017. doi:10.1016/j.cej.2020.125017

Zhang, F., and King, M. W. (2020). Biodegradable polymers as the pivotal player in the design of tissue engineering scaffolds. *Adv. Healthc. Mat.* 9 (13), 1901358. doi:10.1002/adhm.201901358

Zhang, L., Yang, G., Johnson, B. N., and Jia, X. (2019). Three-dimensional (3D) printed scaffold and material selection for bone repair. *Acta Biomater.* 84, 16–33. doi:10.1016/j.actbio.2018.11.039

Zheng, W., Fan, H., Wang, L., and Jin, Z. (2015). Oxidative self-polymerization of dopamine in an acidic environment. *Langmuir* 31 (42), 11671–11677. doi:10.1021/acs.langmuir.5b02757

Zhou, L., Ge, J., Wang, M., Chen, M., Cheng, W., Ji, W., et al. (2021). Injectable muscle-adhesive antioxidant conductive photothermal bioactive nanomatrix for efficiently promoting full-thickness skeletal muscle regeneration. *Bioact. Mater.* 6 (6), 1605–1617. doi:10.1016/j.bioactmat.2020.11.005

Zhou, L., Li, X., Wang, K., Shen, F., Zhang, L., Li, P., et al. (2020). Cu<sup>I</sup>-loaded polydopamine coatings with *in situ* nitric oxide generation function for improved hemocompatibility. *Regen. Biomater.* 7 (2), 153–160. doi:10.1093/rb/rbz043

Zhou, P., Deng, Y., Lyu, B., Zhang, R., Zhang, H., Ma, H., et al. (2014). Rapidly-Deposited polydopamine coating via high temperature and vigorous stirring: Formation, characterization and biofunctional evaluation. *PLoS ONE* 9 (11), e113087. doi:10.1371/ journal.pone.0113087

Zhou, X., OuYang, J., Li, L., Liu, Q., Liu, C., Tang, M., et al. (2019). In vitro and in vivo anti-corrosion properties and bio-compatibility of  $5\beta$ -TCP/Mg-3Zn scaffold coated with dopamine-gelatin composite. *Surf. Coatings Technol.* 374, 152–163. doi:10.1016/j.surfcoat.2019.05.046

Zia, Q., Tabassum, M., Meng, J., Xin, Z., Gong, H., and Li, J. (2021). Polydopamine-assisted grafting of chitosan on porous poly (L-lactic acid) electrospun membranes for adsorption of heavy metal ions. *Int. J. Biol. Macromol.* 167, 1479–1490. doi:10.1016/j.ijbiomac.2020.11.101