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Mechanistic Insights into the FeCl₃-Catalyzed Oxidative Cross-Coupling of Phenols with 2-Aminonaphthalenes

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for preparing N,O-biaryl compounds that are widely used as ligands and catalysts. Based on a comprehensive kinetic investigation, a catalytic cycle involving a ternary complex that binds to both the coupling partners and the oxidant during the key oxidative coupling step is postulated. Furthermore, the studies showed that the reaction is regulated by off-cycle acid—base and ligand exchange processes.

■ INTRODUCTION

Iron-catalyzed oxidative phenol coupling reactions¹ bring together phenols with unfunctionalized C-H nucleophiles such as 1,3-dicarbonyl compounds,² conjugated alkenes,³ arenes, polyaromatic hydrocarbons (PAHs),^{2b,4} and a second phenolic coupling partner.⁵ This method is considered to be highly attractive in terms of step- and atom-economy for assembling new phenolic architectures.⁶ As part of our group research program, we aimed to extend this reaction for the coupling of anilines to afford N,O-biaryl compounds that are widely used in asymmetric transformations.7 Anilines and phenols share some of the same properties: they are both electron-rich cyclic π -systems that are prone to oxidation, generating a highly reactive electrophilic radical species.⁸ Therefore, the development of selective oxidative crosscoupling reactions between phenols and unprotected anilines is a challenging task that has rarely been achieved.⁹

In an early work, Kočovský studied the reaction between 2naphthol 1a and 2-aminonaphthalene 2a using a stoichiometric amount of a redox copper amine complex, ^{9k,1,n} affording (\pm)-2amino-2'-hydroxy-1,1'-binaphthyl 3 (NOBIN, Scheme 1A). Recently, the Shindo group has developed aerobic oxidative cross-coupling conditions based on a heterogeneous Rh/C catalyst for the reaction between tertiary *N*,*N*-dialkylamino-2naphthalenes and different nucleophiles, such as *N*,*N*dialkylanilines, arenes, and phenols (Scheme 1A).^{9f,10} Lately, our group has developed an M[TPP]Cl (M = Fe or Mn, TPP = 5,10,15,20-tetraphenyl-21H,23H-porphine)-catalyzed *para*selective oxidative amination of phenols by primary and secondary anilines (Scheme 1B).¹¹ We have demonstrated that, depending on the identity of the phenolic *para*-R group, the products of this coupling are either benzoquinone anils

(when R = H or OMe) or N,O-biaryl compounds (when R =alkyl). In a previous paper, we developed a two-step synthesis of optically pure NOBIN derivatives. The practical method is based on a stereoselective FeCl3-catalyzed oxidative crosscoupling between 2-naphthols (e.g., 1a, 1.5 equiv) and 2aminonaphthalenes with a labile chiral auxiliary group (such as 2c, 1 equiv, Scheme 1C),¹² affording a mixture of two separable NOBIN diastereoisomers [e.g., (R_a,S) -5 and (S_a,S) -5]. A simple hydrogenolysis of the auxiliary group $(H_2, Pd/C)$ offers a direct entry to the desirable (R_a) -3 and (S_a) -3 NOBINs in excellent chemical yields. Intrigued by the high degree of cross-coupling selectivity and the excellent yields imparted by the FeCl₃/TFA/t-BuOOt-Bu catalytic system, we were interested in probing the underlying mechanism and studying the generality of this method for the preparation of N,O-biaryl compounds (Scheme 1D).

tep E

TFA

(Fel=Fe(L),

Fel(2f) (30)

The general mechanistic line for the oxidative coupling of phenols by iron catalysts involves three key steps: (1) the formation of high-valent iron-phenolate complexes, (2) the generation of a ligated phenoxyl radical intermediate, and (3) coupling with a π -nucleophile or a radical species.^{1a} Recent mechanistic studies by our group for the FeCl₃-catalyzed oxidative homo- and cross-coupling reaction of phenols revealed a zero-order dependence on the [phenol].^{5b} Based on these results, it was suggested that a multicoordinated iron

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Scheme 1. (A) Oxidative Cross-Coupling between 2-Naphthols and 2-Aminonaphthalene Derivatives, (B) the M[TPP]Cl-Catalyzed Oxidative Amination of Phenols by Anilines, (C) the Stereoselective Synthesis of NOBINs, and (D) Mechanistic and Scope Studies of the FeCl₃-Catalyzed Oxidative Phenol-2-Aminonaphthalene Coupling (This Work)



catalyst mediates an inner-sphere oxidative radical—anion coupling between two neighboring ligands (Figure 1A).^{Sb} However, the partial order for the phenolic component is no longer zero when the catalyst has a limited number of vacant sites, as exemplified by Katsuki [(Fe[μ -OH][salen])₂ catalyst]¹³ and Pappo [Fe[phosphate]₃ catalyst] (Figure 1B).^{Sc}



Figure 1. Relationship between the coupling mechanisms and the catalyst structure.

Furthermore, when the reaction is mediated by Fe[TPP]Cl, which has only a single axial position available for binding, the coupling takes place between a ligated phenoxyl radical and a liberated phenoxyl^{5a} or an anilino radical¹¹ by an outer-sphere radical-radical coupling mechanism (Figure 1C). These studies show that the coupling mechanism changes as a function of the iron coordination sphere. Therefore, the selectivity and the efficiency of the oxidative coupling are expected to be affected by the relative binding strengths of the two coupling partners to the redox iron complex.

Herein, we report that $FeCl_3$ is an efficient catalyst for the oxidative coupling between readily oxidized phenols and primary, secondary, and tertiary 2-aminonaphthalene derivatives. The selective conditions were successfully applied for the synthesis of a long list of novel *N*,*O*-biaryl compounds that are needed as ligands in catalysis. Our comprehensive mechanistic studies support the existence of an inner-sphere coupling mechanism between a phenoxyl radical and a 2-aminonaphthalene ligand. Furthermore, initial rate kinetic experiments uncovered (a) the involvement of a ternary complex that binds to both the coupling partners and the oxidant during the key oxidative coupling step and (b) the existence of two off-cycle acid—base and ligand exchange processes that regulate the reaction rate.

RESULTS AND DISCUSSION

Method Development and Reaction Scope. Our research commenced by applying oxidative phenol-phenol coupling conditions [FeCl₃ (10 mol %), t-BuOOt-Bu (1.5 equiv), HFIP, and room temperature], which were developed by our group,^{1a,4,5b,14} for reacting 2-naphthol 1a (1.5 equiv) with 2-aminonaphthalene 2a (1 equiv). Fortunately, this reaction proceeded smoothly, affording (rac)-NOBIN 3 in 82% yield (Figure 2). However, when secondary and tertiary 2aminonaphthalene derivatives, N-butyl-2-aminonaphthalene 2d and piperidino-2-naphthalene 2e, respectively, were reacted with 2-naphthol 1a, poor conversions were observed. Our study revealed that the addition of TFA (1.25 equiv) to the reaction between 1a and the secondary 2-aminonaphthalene 2d significantly improved the reaction efficiency, affording NOBIN 6 in 79% yield (Figure 2).¹² However, with piperidino-2-naphthalene 2e, a higher concentration of TFA (3.75 equiv) was needed to ensure efficient and highly selective coupling, affording NOBIN 7 in 97% yield (Figure 2).

The scope of the reaction was further explored by reacting various 3- or 6-substituted-2-naphthols with 3-, 6-, and/or *N*-substituted-2-aminonaphthalene derivatives (Figure 2). Under the general conditions (*rac*)-NOBINS 3, 6–17 were prepared in moderate to excellent yields (60–97%). An oxidative coupling between substituted phenols and *N*-substituted-2-aminonaphthalene derivatives is also possible, affording *N*,*O*-biaryl compounds 18–30 with high chemoselectivity and with yields that varied between 38 and 98% (Figure 3). Importantly, oxidizable functional groups, such as *para*-methoxybenzyl (compounds 9, 10, 14, 23, and 25) and conjugate alkenes (13 and 28), survived the mild oxidation conditions. Finally, the scalability of the process was demonstrated by preparing compound 30 on a 2 mmol scale, and the structure of compound 20 was confirmed by X-ray diffraction analysis.

Mechanistic Studies. With the aim to elucidate a detailed catalytic cycle that will rationalize the observed reactivity and selectivity, a set of kinetic experiments were performed. The oxidative cross-coupling between 2,6-dimethylphenol (**1b**) and

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Figure 2. Scope of the oxidative coupling between 2-naphthols and 2aminonaphthalene derivatives. Reaction conditions: 2-naphthol (1.5 equiv), 2-aminonaphthalene (1 equiv), FeCl₃ (10 mol %), *t*-BuOOt-Bu (1.5 equiv), TFA (1.25 equiv), HFIP (0.5 M), room temperature, and 24 h. ^aThe reaction was performed without TFA. ^bThe reaction was performed with 3 equiv of 2-naphthol 1a, 4.5 equiv of *t*-BuOOt-Bu, and 3.75 equiv of TFA in total.

N-benzyl-2-aminonaphthalene (**2f**) was chosen since this transformation showed a high degree of cross-coupling selectivity.¹⁵ First, the dependence of [phenol **1b**], [*t*-BuOO*t*-Bu], and [FeCl₃] on the reaction rate was investigated. The results show (i) a saturation curve for phenol **1b**, first order at a low concentrations range (0.02–0.1 M; see Figure 4A and Figure S1 in the Supporting Information) and close to a zero-order dependency at a high concentrations range (0.1–0.3 M); (ii) a positive rate dependence for *t*-BuOO*t*-Bu is found when the experiment was performed at a high level of [**1b**] (0.25 M, Figure 5A); and (iii) first order in the catalyst was observed for FeCl₃ (Figure 6A).

The reaction does not take place in the absence of the redox catalyst, as confirmed by the zero intercept and our control experiments. The existence of free radical mechanisms was ruled out since the addition of butylated hydroxytoluene (BHT) to the reaction mixture had no effect on the coupling yield (see the Supporting Information). Furthermore, these kinetic results strengthen the premise that *t*-BuOO*t*-Bu binds to the iron catalyst prior to the slow oxidative coupling



Figure 3. Scope of the oxidative coupling between substituted phenols and 2-aminonaphthalene derivatives. Reaction conditions: phenol (1.5 equiv), 2-aminonaphthalene (1 equiv), FeCl₃ (10 mol %), *t*-BuOOt-Bu (1.5 equiv), TFA (1.25 equiv), HFIP (0.5 M), room temperature, and 24 h. ^{*a*}The reaction was performed with 3 equiv of *t*-BuOOt-Bu and 2.5 equiv of TFA in total. ^{*b*}The reaction was performed on a 2 mmol scale. ^{*c*}The conditions were similar, except for phenol (1 equiv).

step,^{2a,5c} ruling out its action as a terminal oxidant that regenerates Fe(III) from Fe(II) after the coupling step.

The initial rate kinetic study indicated that **1b**, *t*-BuOO*t*-Bu, and **2f**, which showed negative order dependency (vide infra), are bound to the iron prior to the irreversible oxidative coupling step. Therefore, a kinetic behavior that characterizes a ternary enzyme (E, Figure 7) was considered. The action of ternary enzymes has been comprehensively studied by Cleland¹⁵ and others.¹⁶ These studies indicate that the mechanistic scheme of Bi-substrate enzyme-catalyzed reactions is characterized by a reversible binding of two substrates (**A** and **B**) to the enzyme prior to the slow step (E·**A**·**B** \rightarrow products). The formation of E·**A**·**B** by the sequential binding of **A** and **B** can take place either via "random" or "ordered" sequential mechanisms. In a random mechanism, E·**A**·**B** is obtained from both E·**A** and E·**B**; i.e., the dissociation



Figure 4. Initial rate kinetic studies for phenol 1b. (A) Conditions: phenol 1b (0.02, 0.04, 0.08, 0.1, 0.15, 0.2, and 0.3 mmol), 2-aminonaphthalene 2f (0.1 mmol), TFA (0.125 mmol), FeCl₃ (0.01 mmol), *t*-BuOOt-Bu (0.15 mmol), and HFIP (1 mL). (B) Conditions: (i) phenol 1b (0.02, 0.04, 0.08, 0.2, and 0.3 mmol), 2-aminonaphthalene 2f (0.1 mmol), TFA (0.125 mmol), FeCl₃ (0.01 mmol), *t*-BuOOt-Bu (0.05 mmol), and HFIP (1 mL); (ii) phenol 1b (0.02, 0.04, 0.08, 0.2, and 0.3 mmol), 2-aminonaphthalene 2f (0.1 mmol), and HFIP (1 mL); (ii) phenol 1b (0.02, 0.04, 0.08, 0.2, and 0.3 mmol), 2-aminonaphthalene 2f (0.1 mmol), TFA (0.125 mmol), FeCl₃ (0.01 mmol), *t*-BuOOt-Bu (0.15 mmol), and HFIP (1 mL); (iii) phenol 1b (0.02, 0.04, 0.08, 0.2, and 0.3 mmol), 2-aminonaphthalene 2f (0.1 mmol), TFA (0.125 mmol), FeCl₃ (0.01 mmol), t-BuOOt-Bu (0.25 mmol), TFA (0.125 mmol),

constants of substrates to the free enzyme (K_{iA} for **A** and K_{iB} for **B**) and from E·**A**·**B** to enzymes E·**A** and E·**B** (K_A and K_B , respectively) are equal ($K_{iA} = K_A$ and $K_{iB} = K_B$).¹⁷ However, in an ordered sequential mechanism E·**A**·**B** is obtained solely from E·**A** if substrate **A** binds preferentially to the free enzyme E ($K_{iA} < K_A$ and $K_{iB} > K_B$) or solely from E·**B** if binding of substrate **A** to this complex occurs in higher affinity ($K_{iA} > K_A$ and $K_{iB} < K_B$).¹⁸

The order in which phenol **1b** and *t*-BuOO*t*-Bu bind to the iron catalyst (assuming that $E = [Fe](2f)_m$] was determined by performing a set of double-reciprocal analysis experiments.¹⁹ First, phenol **1b** (assigned as substrate **A**) was varied at fixed concentrations of *t*-BuOO*t*-Bu (0.05, 0.15, and 0.25 M, Figure 4B; see also Figure S1 in the Supporting Information) and then



Figure 5. Initial rate kinetic studies for t-BuOOt-Bu. (A) Conditions: t-BuOOt-Bu (0.05, 0.1, 0.15, 0.2, 0.25, and 0.3 mmol), phenol 1b (0.25 mmol), 2-aminonaphthalene 2f (0.1 mmol), TFA (0.125 mmol), FeCl₃ (0.01 mmol), and HFIP (1 mL). (B) Conditions: (i) t-BuOOt-Bu (0.05, 0.1, 0.15, 0.2, and 0.25 mmol), phenol 1b (0.15 mmol), 2-aminonaphthalene 2f (0.1 mmol), TFA (0.125 mmol), FeCl₃ (0.01 mmol), and HFIP (1 mL); (ii) t-BuOOt-Bu (0.05, 0.1, 0.15, 0.2, and 0.25 mmol), phenol 1b (0.25 mmol), 2-aminonaphthalene 2f (0.1 mmol), TFA (0.125 mmol), 2-aminonaphthalene 2f (0.1 mmol), TFA (0.125 mmol), FeCl₃ (0.01 mmol), and HFIP (1 mL); (iii) t-BuOOt-Bu (0.05, 0.1, 0.15, 0.2, and 0.25 mmol), phenol 1b (0.30 mmol), 2-aminonaphthalene 2f (0.1 mmol), TFA (0.125 mmol), FeCl₃ (0.01 mmol), and HFIP (1 mL). The rates of the formation of product 30 in the initial stage of the reaction were determined by HPLC, using mesitylene as the internal standard.

t-BuOO*t*-Bu (assigned as substrate **B**) was varied at fixed [phenol **1b**] values (0.15, 0.25, and 0.30 M, Figure 5B; see also Figure S2 in the Supporting Information). The Lineweaver–Burk plot for the phenol (Figure 4B) shows linear lines that intersect above the horizontal axis, whereas the position of the crossover point for the peroxide's linear lines (Figure 5B) is below the *x*-axis. According to Frieden analysis,¹⁹ these results indicate that $K_{iA} > K_A$ and $K_{iB} < K_B$ (Figure 7, Eq. 3),^{16,17,19,20} suggesting that [Fe]·(**2f**)_m·(**1b**)·(*t*-BuOO*t*-Bu) **III** is formed from [Fe]·(**2f**)_m **I** by a sequential binding of the peroxide (step A, Scheme 2) and the phenol (Step B).

The dependence of 2-aminonaphthalene **2f** on the reaction velocity was examined (Figure 6B). The negative relationship between the reaction rate and [**2f**] indicates the presence of a competitive off-cycle equilibrium.²¹ It is suggested that the association of the peroxide and the phenol to complex I (step A, Scheme 2) is suppressed by the competitive binding of **2f**, affording $[Fe] \cdot (\mathbf{2f})_{m+1}$ (**V**, off-cycle step, Scheme 2). Consequently, the rate of the coupling decelerates as [**2f**] increases. These results also support the assumption that 2-

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Figure 6. Initial rate kinetic studies for (A) conditions: FeCl₃ (0.001, 0.0025, 0.005, 0.01, and 0.015 mmol), phenol 1b (0.1 mmol), 2-aminonaphthalene 2f (0.1 mmol), TFA (0.125 mmol), t-BuOOt-Bu (0.15 mmol), and HFIP (1 mL). (B) Conditions: 2-aminonaphthalene 2f (0.05, 0.1, 0.15, 0.2, and 0.25 mmol), phenol 1b (0.1 mmol), TFA (1.25 equiv according to the concentration of 2f), FeCl₃ (0.01 mmol), t-BuOOt-Bu (0.15 mmol), and HFIP (1 mL). (C) Conditions: TFA (0.05, 0.1, 0.125, 0.15, 0.2, 0.25, and 0.3 mmol), phenol 1b (0.1 mmol), 2-aminonaphthalene 2f (0.1 mmol), FeCl₃ (0.01 mmol), t-BuOOt-Bu (0.15 mmol), and HFIP (1 mL). The rates of the formation of product 30 in the initial stage of the reaction were determined by HPLC, using mesitylene as the internal standard.



The general rate equation for a bi-substrate sequential mechanism:

Eq.1

$$V = \frac{V_{max}[A][B]}{K_{iA}K_B + K_A[B] + K_B[A] + [A][B]}$$

[4][D

The general rate equation in a double reciprocal form:

Eq.2

$$\frac{1}{v} = rate^{-1} = \left[\frac{K_A}{V_{max}} \left(1 + \frac{K_{iA}K_B}{K_A[B]}\right)\right] \frac{1}{[A]} + \frac{1}{V_{max}} \left(1 + \frac{K_B}{[B]}\right)$$

At the crossover point, the same $\frac{1}{n}$ value is obtained for different [**B**], therefore

Eq.3

$$\frac{1}{v} = \frac{1}{V_{max}} \left(1 - \frac{K_A}{K_{iA}} \right)$$

 $K_{A} = K_{IA}$ then the lines intersect on X-axis $K_{IA} > K_{A}$ then the lines intersect above X-axis $K_{A} < K_{A}$ then the lines intersect below X-axis

In a similar manner for different [A]:

$$\frac{1}{v} = \frac{1}{V_{max}} \left(1 - \frac{K_B}{K_{iB}} \right)$$

'Random' mechanism'Ordered' mechanism
$$K_{iA} = K_A$$
 and $K_{iB} = K_B$ Via E•A $K_{iA} < K_A$ and $K_{iB} > K_B$ Via E•B $K_{iA} > K_A$ and $K_{iB} < K_B$

Figure 7. Catalytic reaction involving a ternary complex.

aminonaphthalene 2f serves as a strong *N*-ligand that coordinates to the iron in preference to phenol 1b and *t*-BuOO*t*-Bu.

Scheme 2. Postulated Mechanism for the Oxidative Cross-Coupling between 2,6-Dimethylphenol 1b and N-Benzyl-2aminonaphthalene 2f

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Product inhibition experiments offer useful inputs when deciding the kinetic mechanism of a ternary complex ($E \cdot \mathbf{Q} \rightleftharpoons E + \mathbf{Q}$, Figure 7). The experiments were performed by monitoring the formation rate of product **30** in the presence of increasing concentrations of NOBIN **6** or *t*-BuOH. NOBIN **6** was chosen for practical reasons associated with the fact that NOBINs **30** and **6** have different retention times in the HPLC. Figure 8 shows that although *t*-BuOH acts as a weak inhibitor, at saturating values of NOBIN **6**, the catalyst's activity approaches zero. Unsurprisingly, these results indicate that the coupling product acts as a competitive ligand. It is expected that the concentration of complex IV ($[Fe] \cdot (2f)_{m-1}(30)$, Scheme 2) builds up as the reaction proceeds; consequently, the velocity of the coupling decreases.

Based on these kinetic results, a detailed mechanistic scheme was postulated and is presented in Scheme 2. The catalytic cycle begins with the reversible binding of the peroxide and the phenol to $[Fe] \cdot (2f)_m$ (I), affording complex III ($[Fe] \cdot (2f)_m \cdot$ (1b) · (*t*-BuOO*t*-Bu)), steps A and B). The homolytic cleavage of the peroxide bond by the iron, followed by an inner-sphere coupling between a phenoxyl radical and a neighboring 2aminonaphthalene ligand, will afford complex IV and two molecules of *t*-BuOH (step C).^{2d,e,5b,14} The catalytic cycle is terminated by a reversible ligand exchange process that involves the liberation of *N*,*O*-biaryl product **30**, along with the binding of 2-aminonaphthalene **2f** (step D).



Figure 8. Product inhibition experiments. The effect of [NOBIN 6] (green diamonds) and [*t*-BuOH] (purple triangles) on the reaction rate. Conditions: phenol 1b (0.1 mmol), 2-aminonaphthalene 2f (0.1 mmol), TFA (0.125 mmol), FeCl₃ (0.01 mmol), NOBIN 6, or *t*-BuOH (0.02, 0.03, 0.05, 0.1, and 0.3 mmol), *t*-BuOOt-Bu (0.15 mmol), and HFIP (1 mL). The rates of the formation of product 30 in the initial stage of the reaction were determined by HPLC, using mesitylene as the internal standard.

Our study implies that the reaction kinetics is strongly influenced by the relative binding strength of the substrates (2aminonaphthalene, phenol, and peroxide) and the coupling product to the iron. As mentioned previously, the addition of TFA is mandatory when secondary and tertiary 2-aminopubs.acs.org/joc

naphthalenes are being reacted. It is expected that TFA, which forms an acid-base adduct with the latter coupling partners, interferes in the net of ligand exchange processes. To clarify the role of the acid, we performed additional sets of kinetic experiments.

The dependence of the initial rate on [TFA] (Figure 6C) revealed that, although no reaction occurs in the absence of the acid, the maximum reactivity is achieved when [2f] and [TFA] are equalized (ca. a 1:1 ratio). However, as the acid concentration increases, the reaction velocity diminishes. These results can be rationalized by the existence of ligandto-metal exchange and acid-base net reactions (Scheme 2). It is suggested that the entire catalytic process is regulated by TFA, which forms an acid-base adduct with 2f. Accordingly, as the acid concentration increases, the concentration of free 2aminonaphthalene 2f drops (step E). Consequently, the offcycle equilibrium inclines toward complex I and the rate accelerates (0.05 M < [TFA] < 0.12 M). On the other hand, at high concentrations of TFA ([TFA] > 0.12 M), the concentration of 2f diminishes. Consequently, the catalytic cycle termination step (IV \rightarrow I, step D), which includes the reversible ligand exchange of the N,O-biaryl product 30 with 2f, is discouraged, and the reaction rate declines.

The strength of the TFA-based adduct depends on the basicity of the 2-aminonapthalene molecule. Therefore, different amounts of acid should be used to regulate the coupling of primary, secondary, or tertiary 2-aminonaphthalenes. To support this claim, a set of competitive experiments that studied the coupling of 2-aminonaphthalenes 2a, 2d, or 2e (1 equiv) and 2-naphthol (1a, 1.5 equiv) either with or without 2 equiv of TFA were performed (Figure 9). The results show that the addition of TFA to the reaction of 2a, which is a weaker base in comparison with 2d and 2e, negatively affects the reaction rate (Figure 9A). On the other



Figure 9. Reaction progress of the oxidative coupling of 2-naphthol 1a with N-substituted-2-aminonaphthalenes (A) 2a, (B) 2d, and (C) 2e with and without TFA (2 equiv). Conditions: 2-naphthol 1a (0.375 mmol), 2-aminonaphthalene 2a, 2d, or 2e (0.25 mmol), FeCl₃ (10 mol %), TFA (0 or 0.5 mmol), *t*-BuOOt-Bu (0.375 mmol), and HFIP (0.5 mL) at rt. The formation of products 3, 6, or 7 was determined by HPLC, using mesitylene as the internal standard.

hand, the reaction of 2d in the presence of TFA resulted in a significant improvement in the reactivity (Figure 9B) and cross-coupling selectivity (see Figure S3 in the Supporting Information). Finally, tertiary 2-aminonaphthalene 2e exhibited only a mild improvement in the rate upon the addition of TFA (Figure 9C). This is probably because 2 equiv of TFA are insufficient to regulate the inhibiting off-cycle process. Indeed, almost twice the amount of TFA (3.75 equiv) is needed to ensure efficient cross-coupling, affording NOBIN 7 in 97% yield (Figure 2). Ultimately, the coupling of primary 2-aminonaphthalene takes place at a high efficiency without TFA (see the inserted table, Figure 9), whereas the successful coupling of secondary and tertiary 2-aminonaphthalenes relies on the addition of TFA (1.25 equiv and 3.75 equiv, respectively).

The changes in the catalytic activity at high concentrations of TFA may also be attributed to the generation of iron trifluoroacetate complexes $[Fe(CF_3CO_2)_n(Cl)_m]$. To examine this hypothesis, the $Fe(CF_3CO_2)_3$ complex²² was prepared and used as a catalyst (10 mol %) in the coupling between **1b** and **2f** (Figure 10). Fe(CF_3CO_2)_3 exhibited almost no catalytic



Figure 10. Effect of chloride anions [TBAC] on the catalytic activity of the Fe(TFA)₃ catalyst. Conditions: phenol **1b** (0.1 mmol), 2-aminonaphthalene **2f** (0.1 mmol), TFA (0.125 mmol), FeCl₃ (0.01 mmol) or Fe(CF₃CO₂)₃ and Bu₄N⁺Cl⁻ (0, 10, 20, and 30 mol %), *t*-BuOO*t*-Bu (0.15 mmol), and HFIP (1 mL). The rates of the formation of product **30** in the initial stage of the reaction were determined by HPLC, using mesitylene as the internal standard.

activity. However, the reactivity was enhanced with the addition of 10 mol % of tetrabutylammonium chloride (TBAC). Interestingly, almost a complete recovery of the catalytic activity (in comparison to $FeCl_3$) was achieved with 1:2 and 1:3 iron to chloride ratios. These results suggest that the chloride anions play a key role during the reaction.

CONCLUSIONS

In conclusion, the FeCl₃-catalyzed oxidative phenol coupling reaction was applied to combine readily oxidized phenols with primary, secondary, and tertiary 2-aminonaphthalenes. This sustainable and practical method enables a highly selective and efficient synthesis of N,O-biaryl compounds that are not readily available by other means.

Our mechanistic data, which include control experiments and comprehensive kinetic studies, revealed the existence of a catalytic cycle that involves the formation of a ternary iron complex $[Fe] \cdot (2f) \cdot (1b) \cdot (t-BuOOt-Bu)$ (III) from $[Fe] \cdot (2f)_m$ (I) by the sequential binding of peroxide and phenol. The irreversible rate-determining oxidative coupling step comprises the conversion of complex III to $IV\;([\text{Fe}]{\cdot}(2f)_{\text{m-1}}(30))$ and the liberation of two molecules of t-BuOH. In this transformation, a reaction between an iron-bound phenoxyl radical and a neighboring 2-aminonaphthalene ligand takes place. The velocity of the reaction is regulated by a net of acid-base and ligand exchange processes. The reaction rate is highly sensitive to changes in the concentrations of the substrates (2aminonaphthalene, phenol, and peroxide), the acid (TFA), and the N,O-biaryl product. Furthermore, the chloride anions have a strong effect on the reaction efficiency. Finally, this study is a part of our laboratory ongoing research that aims to develop selective oxidative cross-coupling reactions for the coupling of anilines by first-row metal catalysts.

EXPERIMENTAL SECTION

General Methods. All reagents were of reagent-grade quality, purchased commercially from Sigma-Aldrich, Alfa-Aesar, or Fluka, and used without further purification. FeCl₃ (anhydrous 98%) was purchased from Strem Chemicals. Purification by column chromatography was performed on Merck chromatographic silica gel (40-63 μ m). Thin-layer chromatography (TLC) analyses were performed using Merck silica gel glass plates 60 F254. NMR spectra were recorded on Bruker DPX400 or DMX500 instruments; chemical shifts are relative to Me₄Si as the internal standard or to the residual solvent peak. High-resolution mass spectrometry (HRMS) data were obtained using an LTQ Orbitrap XL ETD (Thermo Fisher Scientific, Germany and USA) high-resolution mass spectrometer. The reactions in the microwave were performed using a CEM Discover SP microwave synthesizer. IR spectra were recorded on a JASCO FT/IR-460 Plus FT-IR instrument. HPLC analysis was carried out on an Agilent 1260 instrument equipped with a G4212-60008 photodiode array detector and an Agilent reverse phase ZORBAX Eclipse plus C18 3.5 μ m column (4.6 × 100 mm).

General Procedures for the Synthesis of *N*-Alkyl-2-aminonaphthalenes. *Method A*. A mixture of 2-naphthol derivative (1 equiv) and alkyl/arylamine (5 equiv) was irradiated in a microwave for 20 h (sealed reaction vessel, temperature of 275 °C was monitored by using an external surface sensor and a power of 200 W). The volatiles were removed under reduced pressure, and the crude residue was further purified by silica-gel column chromatography (silica gel 40–63 μ m). This method was used for the preparation of 2aminonaphthalene derivatives 2d, 2e, 2i, 2j, 2n, and 2o.

Method B.²³ A mixture of 2-aminonaphthalene (1 equiv) and benzaldehyde (1.1 equiv) was stirred in methanol (0.17 M) for 1 h, and then NaBH₄ (1.5 equiv) was added. The reaction was stirred for 20 min, and the volatiles were removed under reduced pressure. NaOH (1 M, 30 mL) was added and extracted with diethyl ether (3 × 20 mL). The combined organic phase was dried over MgSO₄ and evaporated under reduced pressure. The crude residue was further purified by silica-gel column chromatography (silica gel 40–63 μ m). This method was used for the preparation of 2-aminonaphthalene derivatives 2f, 2g, 2h, 2k, 2l, 2 m, and 2p.

N-Heptyl-3-methoxy-2-aminonaphthalene (2i). This compound was prepared from 3-methoxy-2-naphthol (1 g, 5.74 mmol) and 1-heptylamine (4.24 mL, 28.7 mmol) according to method A. The crude residue was purified by silica-gel column chromatography (hexane/ethyl acetate 99:1) to afford compound **2i** (1.49 g, 96% yield) as a dark red oil. ¹H NMR (CDCl₃, 400 MHz): δ 0.93 (t, 3H, J = 6.9 Hz), 1.25–1.56 (m, 8H), 1.75 (quin, 2H, J = 7.2 Hz), 3.24 (t, 2H, J = 7.2 Hz), 3.98 (s, 3H), 6.79 (s, 1H), 7.02 (s, 1H), 7.21 (ddt, 1H, J = 8.0, 7.0, 1.2 Hz), 7.29 (ddt, 1H, J = 8.0, 6.9, 1.2 Hz), 7.62 (d,

1H, J = 4.1 Hz), 7.64 (d, 1H, J = 4.0 Hz). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 14.2, 22.8, 27.4, 29.3, 29.4, 32.0, 43.8, 55.6, 103.4, 104.5, 122.1, 124.1, 125.4, 126.4, 127.2, 130.6, 138.8, 148.2. HRMS (ESI) m/z: [M + H]⁺ calcd for C₁₈H₂₆NO 272.2009, found 272.2014.

N-*Heptyl-6-methoxy-2-aminonaphthalene* (**2***j*). This compound was prepared from 6-methoxy-2-naphthol (697 mg, 4 mmol) and 1-heptylamine (2.84 mL, 20 mmol) according to method A. The crude residue was purified by silica-gel column chromatography (hexane/ethyl acetate 98:2) to afford compound **2***j* (787 mg, 72% yield) as a dark gray solid. ¹H NMR (CDCl₃, 400 MHz): δ 0.90 (t, 3H, *J* = 6.9 Hz), 1.21–1.53 (m, 8H), 1.68 (quin, 2H, *J* = 8.0 Hz), 3.19 (t, 2H, *J* = 8.0 Hz), 3.88 (s, 3H), 6.83 (d, 1H, *J* = 2.3 Hz), 6.90 (dd, 1H, *J* = 8.7, 2.4 Hz), 7.03 (d, 1H, *J* = 2.5 Hz), 7.07 (dd, 1H, *J* = 8.9, 2.6 Hz), 7.53 (s, 1H), 7.55 (s, 1H). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 14.3, 22.8, 27.3, 29.3, 29.5, 32.0, 44.7, 55.4, 105.6, 106.3, 118.6, 118.8, 127.6, 127.8, 128.3, 130.6, 144.4, 155.2. HRMS (ESI) *m*/*z*: [M + H]⁺ calcd for C₁₈H₂₆NO 272.2009, found 272.2003.

N-(*4*-*t*-*Butylbenzyl*)-2-*aminonaphthalene* (**2m**). This compound was prepared from 2-naphthylamine (716 mg, 5 mmol) and 4-*t*-butylbenzaldehyde (0.92 mL, 5.5 mmol) according to method B. The crude residue was purified by silica-gel column chromatography (hexane/ethyl acetate 98:2) to afford compound **2m** (928 mg, 64% yield) as a white solid. ¹H NMR (CDCl₃, 400 MHz): δ 1.34 (s, 9H), 4.41 (s, 2H), 6.87 (d, 1H, *J* = 2.3 Hz), 6.92 (dd, 1H, *J* = 8.7, 2.4 Hz), 7.20 (ddd, 1H, *J* = 8.1, 6.8, 1.2 Hz), 7.33–7.42 (m, 5H), 7.61 (dd, 1H, *J* = 8.3, 0.7 Hz), 7.64 (d, 1H, *J* = 9.0 Hz), 7.68 (d, 1H, *J* = 8.2 Hz). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 31.5, 34.7, 48.2, 104.7, 118.0, 122.1, 125.8, 126.1, 126.4, 127.6, 127.7, 127.8, 129.1, 135.4, 136.2, 146.0, 150.5. HRMS (ESI) *m*/*z*: [M + H]⁺ calcd for C₂₁H₂₄N 290.1903, found 290.1899.

N-Benzyl-3-methoxy-2-aminonaphthalene (**2n**). This compound was prepared from 3-methoxy-2-naphthol (1 g, 5.74 mmol) and benzylamine (3.14 mL, 28.7 mmol) according to method A. The crude residue was purified by silica-gel column chromatography (hexane/ethyl acetate 99:1) to afford compound **2n** (470 mg, 31% yield) as a white solid. ¹H NMR (CDCl₃, 400 MHz): δ 3.99 (s, 3H), 4.48 (s, 2H), 4.94 (bs, 1H), 6.82 (s, 1H), 7.06 (s, 1H), 7.20–7.35 (m, 3H), 7.39 (t, 2H, *J* = 7.3 Hz), 7.46 (d, 2H, *J* = 7.5 Hz), 7.59 (d, 1H, *J* = 8.0 Hz), 7.65 (d, 1H, *J* = 7.8 Hz). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 48.1, 55.6, 103.9, 104.7, 122.4, 124.1, 125.6, 126.4, 127.4 (× 2C), 127.8, 128.8, 130.4, 138.5, 139.2, 148.2. HRMS (ESI) *m/z*: [M + H]⁺ calcd for C₁₈H₁₈NO 264.1383, found 264.1382.

General Procedure for the Oxidative Cross-Coupling of 2-Naphthols/Phenols with 2-Aminonaphthalenes. To a stirred solution of 2-naphthol or phenol derivative (1.5 equiv), 2-aminonaphthalene derivative (1 equiv), TFA (1.25 equiv), and FeCl₃ (10 mol %) in HFIP (0.5 M), *t*-BuOO*t*-Bu (1.5 equiv) was added dropwise at room temperature. The reaction was stirred for 24 h until full consumption of 2-aminonaphthalene. The volatiles were removed under reduced pressure, and the crude was further purified by silicagel column chromatography (silica gel 40–63 μ m), affording pure NOBIN products.

2-Amino-2'-hydroxy-1,1'-binaphthyl (3). 2-Naphthol 1a (54.1 mg, 0.375 mmol) and 2-aminonaphthalene 2a (35.8 mg, 0.25 mmol) were reacted according to the general procedure. In this reaction, TFA was not added. The crude residue was purified by column chromatography (hexane/ethyl acetate 90:10) to afford compound 3 (58.5 mg, 82% yield) as a pale brown solid. ¹H NMR (CDCl₃, 400 MHz): δ 7.03–7.08 (m, 1H), 7.13 (d, 1H, *J* = 8.8 Hz), 7.16–7.20 (m, 1H), 7.20–7.30 (m, 3H), 7.33–7.38 (m, 1H), 7.39 (d, 1H, *J* = 8.9 Hz), 7.81 (dd, 1H, *J* = 7.4, 1.9 Hz), 7.85 (d, 1H, *J* = 8.6 Hz), 7.89 (d, 1H, *J* = 8.0 Hz), 7.93 (d, 1H, *J* = 8.8 Hz). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 108.7, 114.4, 117.8, 118.3, 122.9, 123.8, 123.9, 124.7, 127.1, 127.4, 128.4 (x 3C), 129.6, 130.5, 130.7, 133.3, 134.2, 143.8, 151.9. HRMS (ESI) *m/z*: [M-H]⁺ calcd for C₂₀H₁₄NO 284.1070, found 284.1058.

2'-(Butylamino)-[1,1'-binaphthalen]-2-ol (6). 2-Naphthol 1a (54.1 mg, 0.375 mmol) and N-butyl-2-aminonaphthalene 2d (49.8 mg, 0.25 mmol) were reacted according to the general procedure. The crude residue was purified by column chromatography (hexane/ethyl

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acetate 95:5) to afford compound **6** (67.4 mg, 79% yield) as a yellow syrup. ¹H NMR (CDCl₃, 400 MHz): δ 0.80 (t, 3H, *J* = 7.3 Hz), 1.11–1.23 (m, 2H), 1.33–1.44 (m, 2H), 3.16 (td, 2H, *J* = 7.1, 1.9 Hz), 6.92–6.98 (m, 1H), 7.14 (d, 1H, *J* = 8.6 Hz), 7.16–7.22 (m, 2H), 7.24 (ddd, 2H, *J* = 8.0, 4.7, 1.7 Hz), 7.30–7.35 (m, 1H), 7.37 (d, 1H, *J* = 8.9 Hz), 7.75–7.79 (m, 1H), 7.86 (d, 1H, *J* = 8.0 Hz), 7.90 (d, 1H, *J* = 5.5 Hz), 7.92 (d, 1H, *J* = 5.4 Hz). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 13.9, 20.0, 31.9, 43.7, 107.9, 114.1, 114.3, 117.8, 122.2, 123.6, 123.8, 124.7, 127.0, 127.3, 127.5, 128.3, 128.4, 129.7, 130.5, 130.8, 133.6, 134.3, 145.6, 152.1. HRMS (ESI) *m*/*z*: [M + H]⁺ calcd for C₇₄H₂₄NO 342.1852, found 342.1845.

2'-(Piperidin-1-yl)-[1,1'-binaphthalen]-2-ol (7). 2-Naphthol 1a (54.1 mg, 0.375 mmol) and piperidino-2-naphthalene 2e (52.8 mg, 0.25 mmol) were reacted according to the general procedure. In this reaction, addition of 2-naphthol (1.5 equiv), TFA (2×1.25 equiv), and t-BuOOt-Bu $(2 \times 1.5 \text{ equiv})$ was needed to ensure full consumption of 2-aminonaphthalene. Then, the reaction was quenched by the addition of CH₂Cl₂ and sat. NaHCO₃. The organic phase was separated, dried over MgSO4, and evaporated under reduced pressure. The crude residue was purified by column chromatography (hexane/ethyl acetate 98:2) to afford compound 7 (85.8 mg, 97% yield) as a brown solid. ¹H NMR (CDCl₃, 400 MHz): δ 1.46 (s, 6H), 2.88–3.06 (m, 4H), 7.09 (dd, 2H, J = 8.0, 0.5 Hz), 7.13–7.23 (m, 2H), 7.34 (dtd, 2H, J = 8.0, 6.6, 1.3 Hz), 7.46 (d, 1H, J = 8.8 Hz), 7.51 (d, 1H, J = 8.9 Hz), 7.86 (dd, 1H, J = 8.2, 0.5 Hz), 7.88-7.91 (m, 1H), 7.91 (d, 1H, J = 8.9 Hz), 7.98 (d, 1H, J = 8.8 Hz), 8.26 (bs, 1H). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 24.1, 26.4, 53.5, 118.7, 118.8, 120.5, 123.2, 124.6, 125.3, 126.1, 126.2, 126.3, 126.6, 128.0, 128.3, 129.4, 129.7, 129.9, 130.6, 133.8, 134.3, 149.1, 151.8. IR (KBr pellet): 3431 (s, broad), 2934 (s), 1224 (s), 749 (s) cm⁻¹. HRMS (ESI) m/z: $[M + H]^+$ calcd for C₂₅H₂₄NO 354.1852, found 354.1847.

2'-((4-Chlorobenzyl)amino)-[1,1'-binaphthalen]-2-ol (8). 2-Naphthol 1a (54.1 mg, 0.375 mmol) and N-(4-chlorobenzyl)-2aminonaphthalene 2 g (66.9 mg, 0.25 mmol) were reacted according to the general procedure. The crude residue was purified by column chromatography (hexane/ethyl acetate 85:15) to afford compound 8 (94.7 mg, 92% yield) as a brown solid. ¹H NMR (CDCl₃, 400 MHz): δ 4.36 (s, 2H), 5.16 (bs, 1H), 6.97–7.03 (m, 1H), 7.09 (d, 1H, J = 9.0 Hz), 7.12 (d, 2H, J = 8.4 Hz), 7.17 (dd, 1H, J = 8.3, 0.4 Hz), 7.18-7.25 (m, 4H), 7.30 (ddd, 1H, J = 8.2, 6.8, 1.4 Hz), 7.37 (ddd, 1H, J = 8.1, 6.8, 1.3 Hz), 7.41 (d, 1H, J = 8.9 Hz), 7.75-7.80 (m, 1H), 7.84 (d, 1H, J = 8.8 Hz), 7.90 (d, 1H, J = 7.9 Hz), 7.95 (d, 1H, J = 8.9 Hz). $^{13}C{^{1}H}$ NMR (CDCl₃, 100 MHz): δ 47.0, 108.5, 114.0, 117.8, 122.6, 123.6, 123.9, 124.7, 127.2, 127.5, 127.8, 128.2, 128.4, 128.5, 128.9 (x 2C), 129.7, 130.7, 130.9, 132.9, 133.5, 134.2, 138.1, 144.7, 152.2. HRMS (ESI) m/z: $[M + H]^+$ calcd for C₂₇H₂₁ClNO 410.1306, found 410.1299.

6-Bromo-2'-((3-hydroxy-4-methoxybenzyl)amino)-[1,1'-binaphthalen]-2-ol (9). 6-Bromo-2-naphthol (83.6 mg, 0.375 mmol) and N-(3-hydroxy-4-methoxybenzyl)-2-aminonaphthalene 2h (69.8 mg, 0.25 mmol) were reacted according to the general procedure. The crude residue was purified by column chromatography (hexane/ethyl acetate 95:5) to afford compound 9 (100.4 mg, 80% yield) as a brown solid. ¹H NMR (CDCl₃, 400 MHz): δ 3.82 (s, 3H), 4.14 (bs, 1H), 4.29 (s, 2H), 5.28 (bs, 1H), 5.58 (s, 1H), 6.64 (dd, 1H, J = 8.2, 2.0 Hz), 6.71 (d, 1H, J = 8.2 Hz), 6.76 (d, 1H, J = 2.0 Hz), 6.90–6.96 (m, 1H), 7.05 (d, 1H, J = 9.0 Hz), 7.16 (d, 1H, J = 9.0 Hz), 7.18– 7.25 (m, 2H), 7.34 (dd, 1H, J = 9.0, 2.0 Hz), 7.41 (d, 1H, J = 8.9 Hz), 7.74-7.79 (m, 1H), 7.84 (d, 2H, J = 8.8 Hz), 8.03 (d, 1H, J = 2.0Hz). $^{13}\mathrm{C}\{^{1}\mathrm{H}\}$ NMR (CDCl₃, 100 MHz): δ 47.2, 56.1, 107.6, 110.8, 113.2, 114.4, 114.6, 117.6, 118.2, 119.0, 122.5, 123.3, 126.7, 127.5, 127.7, 128.4, 129.6, 130.3, 130.4, 130.8, 131.0, 132.1, 132.6, 134.1, 145.1, 145.8 (x 2C), 152.5. HRMS (ESI) m/z: $[M + H]^+$ calcd for C28H23BrNO3 500.0856, found 500.0856.

3-Bromo-2'-((3-hydroxy-4-methoxybenzyl)amino)-[1,1'-binaphthalen]-2-ol (10). 3-Bromo-2-naphthol (83.6 mg, 0.375 mmol) and N-(3-hydroxy-4-methoxybenzyl)-2-aminonaphthalene 2 h (69.8 mg, 0.25 mmol) were reacted according to the general procedure. The crude residue was purified by column chromatography (hexane/ethyl acetate 95:5) to afford compound **10** (101.4 mg, 81% yield) as a brown solid. ¹H NMR (CDCl₃, 400 MHz): δ 3.81 (s, 3H), 3.95 (s, 1H), 4.30 (s, 2H), 5.58 (bs, 1H), 6.66 (dd, 1H, *J* = 8.3, 1.6 Hz), 6.71 (d, 1H, *J* = 8.2 Hz), 6.76 (d, 1H, *J* = 1.7 Hz), 6.95 (dd, 1H, *J* = 8.7, 4.5 Hz), 7.13–7.24 (m, 4H), 7.28–7.36 (m, 1H), 7.36–7.45 (m, 1H), 7.75–7.79 (m, 1H), 7.81 (d, 1H, *J* = 8.1 Hz), 7.85 (d, 1H, *J* = 9.0 Hz), 8.25 (s, 1H). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 47.3, 56.0, 108.5, 110.3, 110.8, 111.9, 113.2, 114.4, 116.2, 118.2, 122.4, 123.3, 124.8, 125.0, 127.4, 127.5, 127.6, 128.4, 130.2, 130.9, 132.6, 132.8 (x 2C), 133.8, 144.8, 145.7, 145.8, 148.7. HRMS (ESI) *m*/*z*: [M + H]⁺ calcd for C₁₈H₂BrNO₃ 500.0856, found 500.0880.

Methyl 2'-(Heptylamino)-2-hydroxy-3'-methoxy-[1,1'-binaphthalene]-3-carboxylate (11). Methyl 3-hydroxy-2-naphthoate (75.8 mg, 0.375 mmol) and N-heptyl-3-methoxy-2-aminonaphthalene 2i (67.8 mg, 0.25 mmol) were reacted according to the general procedure. The crude residue was purified by column chromatography (hexane/ethyl acetate 85:15) to afford compound 11 (81.9 mg, 70% yield) as a yellow solid. ¹H NMR (CDCl₃, 400 MHz): δ 0.84 (t, 3H, J = 7.2 Hz), 0.88-1.12 (m, 6H), 1.22 (ddd, 4H, J = 21.9, 16.5, 9.1 Hz), 2.52-2.71 (m, 1H), 2.71-2.86 (m, 1H), 4.06 (s, 3H), 4.07 (s, 3H), 6.98 (d, 1H, J = 8.4 Hz), 7.08 (t, 1H, J = 7.6 Hz), 7.23 (d, 2H, J = 4.7 Hz), 7.28 (d, 1H, J = 4.3 Hz), 7.35 (dd, 2H, J = 6.1, 2.4 Hz), 7.74 (d, 1H, J = 8.0 Hz), 7.86-7.99 (m, 1H), 8.70 (s, 1H). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 14.1, 22.6, 26.7, 28.9, 30.7, 31.8, 46.6, 52.8, 55.6, 105.8, 114.0, 115.7, 119.8, 123.2, 124.0, 124.2 (x 2C), 125.5, 126.8, 127.1, 129.0, 129.4, 129.7 (x 2C), 132.7, 137.7, 138.2, 150.3, 154.4, 170.6. HRMS (ESI) m/z: [M + H]⁺ calcd for C₃₀H₃₄NO₄ 472.2482, found 472.2477.

Methyl 2'-(Heptylamino)-2-hydroxy-6'-methoxy-[1,1'-binaphthalene]-6-carboxylate (12). Methyl 6-hydroxy-2-naphthoate (75.8 mg, 0.375 mmol) and N-heptyl-6-methoxy-2-aminonaphthalene 2j (67.8 mg, 0.25 mmol) were reacted according to the general procedure. The crude residue was purified by column chromatography (hexane/ethyl acetate 90:10) to afford compound 12 (111.1 mg, 94% yield) as a brown solid. ¹H NMR (CDCl₃, 400 MHz): δ 0.83 (t, 3H, J = 7.0 Hz), 1.03–1.24 (m, 8H), 1.33–1.46 (m, 2H), 3.14 (t, 2H, J = 7.1 Hz), 3.88 (s, 3H), 3.94 (s, 3H), 6.84 (d, 1H, J = 9.2 Hz), 6.90 (dd, 1H, J = 9.2, 2.6 Hz), 7.15 (d, 1H, J = 2.6 Hz), 7.16 (d, 1H, J = 8.9 Hz), 7.24 (d, 1H, J = 9.0 Hz), 7.43 (d, 1H, J = 8.9 Hz), 7.79-7.88 (m, 2H), 8.03 (d, 1H, J = 8.9 Hz), 8.63 (d, 1H, J = 1.6 Hz). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 14.2, 22.6, 26.8, 29.0, 29.7, 31.8, 44.2, 52.2, 55.5, 106.9, 108.0, 114.8, 114.9, 118.7, 119.9, 124.9, 125.0, 125.4, 126.5, 128.4, 128.6, 129.5, 129.8, 131.6, 132.0, 136.1, 144.0, 154.3, 155.4, 167.4. HRMS (ESI) m/z: [M + H]⁺ calcd for C₃₀H₃₄NO₄ 472.2482, found 472.2459.

2'-(Cinnamylamino)-6-isopropyl-[1,1'-binaphthalen]-2-ol (13). 6-Isopropyl-2-naphthol^{5c} (69.8 mg, 0.375 mmol) and N-cinnamyl-2aminonaphthalene 2k (64.8 mg, 0.25 mmol) were reacted according to the general procedure. The crude residue was purified by column chromatography (hexane/ethyl acetate 95:5) to afford compound 13 (66.3 mg, 60% yield) as a yellow solid. ¹H NMR (CDCl₃, 400 MHz): δ 1.32 (dd, 6H, J = 6.9, 1.9 Hz), 3.03 (hept, 1H, J = 6.9 Hz), 4.00 (d, 2H, J = 4.5 Hz), 5.10 (bs, 1H), 6.15 (dt, 1H, J = 15.9, 4.8 Hz), 6.44 (d, 1H, J = 15.9 Hz), 7.00-7.03 (m, 1H), 7.13 (d, 1H, J = 8.7 Hz),7.17-7.24 (m, 4H), 7.26 (d, 3H, J = 2.3 Hz), 7.28 (d, 2H, J = 9.1 Hz), 7.37 (d, 1H, J = 8.9 Hz), 7.69 (d, 1H, J = 1.4 Hz), 7.78-7.81 (m, 1H), 7.87–7.94 (m, 2H). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 24.0, 24.1, 34.0, 45.7, 108.7, 114.1, 114.3, 117.7, 122.4, 123.7, 124.7 (x 2C), 126.4, 127.0, 127.2, 127.3, 127.6, 127.8, 128.3, 128.6, 129.9, 130.2, 130.8, 131.1, 132.0, 134.3, 136.8, 144.2, 145.1, 151.6. HRMS (ESI) m/z: $[M + H]^+$ calcd for $C_{32}H_{30}NO$ 444.2322, found 444.2319.

2'-((4-Methoxybenzyl)amino)-5,6,7,8-tetrahydro-[1,1'-binaphthalen]-2-ol (14). 5,6,7,8-Tetrahydro-2-naphthol (55.6 mg, 0.375 mmol) and N-(4-methoxybenzyl)-2-aminonaphthalene 2l (65.8 mg, 0.25 mmol) were reacted according to the general procedure. The crude residue was purified by column chromatography (hexane/ethyl acetate 95:5) to afford compound 14 (88.1 mg, 86% yield) as a brown solid. ¹H NMR (CDCl₃, 500 MHz): δ 1.80–1.89 (m, 4H), 2.68–2.90 (m, 4H), 3.78 (s, 3H), 4.40 (s, 2H), 6.85 (ddd, 3H, J = 6.8, 5.3, 2.8 Hz), 6.91 (s, 1H), 7.09–7.15 (m, 1H), 7.18–7.25 (m, 3H), 7.29– 7.33 (m, 2H), 7.72 (d, 1H, J = 8.1 Hz), 7.75 (d, 1H, J = 9.0 Hz). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 23.3, 23.6, 28.7, 29.6, 47.6, 55.4, 114.2, 114.4, 116.0, 119.3, 122.4, 123.1, 123.9, 127.0, 127.7, 128.2, 128.3, 130.1, 130.2, 131.4, 132.3, 134.0, 139.2, 144.0, 151.7, 158.9. HRMS (ESI) m/z: $[M + H]^+$ calcd for C₂₈H₂₈NO₂ 410.2115, found 410.2104.

2'-((4-(t-Butyl)benzyl)amino)-6-(4-(t-butyl)phenyl)-[1,1'-binaphthalen]-2-ol (15). 6-(4-t-Butylphenyl)-2-naphthol^{5c} (103.6 mg, 0.375 mmol) and N-(4-t-butylbenzyl)-2-aminonaphthalene 2m (72.4 mg, 0.25 mmol) were reacted according to the general procedure. The crude residue was purified by column chromatography (hexane/ethyl acetate 98:2) to afford compound 15 (99.9 mg, 71% yield) as a brown solid. ¹H NMR (CDCl₃, 400 MHz): δ 1.27 (s, 9H), 1.38 (s, 9H), 4.33-4.46 (m, 2H), 7.05 (dd, 1H, J = 6.0, 3.3 Hz), 7.13 (d, 2H, J = 8.1 Hz), 7.18–7.24 (m, 3H), 7.27 (d, 3H, J = 8.1 Hz), 7.43 (d, 1H, J = 8.9 Hz, 7.50 (d, 2H, J = 8.1 Hz), 7.57 (d, 1H, J = 8.7 Hz), 7.63 (d, 2H, J = 8.2 Hz), 7.78 (dd, 1H, J = 5.9, 3.4 Hz), 7.87 (d, 1H, J = 9.1 Hz), 7.99 (d, 1H, J = 9.0 Hz), 8.08 (s, 1H). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 31.5 (x 2C), 34.6, 34.7, 47.3, 108.2, 114.2, 114.4, 118.2, 122.4, 123.6, 125.3, 125.6, 125.9, 126.2, 126.6, 126.8, 127.0, 127.4, 127.7, 128.4, 130.0, 130.9 (x 2C), 132.7, 134.2, 136.4, 136.5, 138.3, 145.2, 150.1, 150.3, 152.2. HRMS (ESI) m/z: $[M + H]^+$ calcd for C41H42NO 564.3261, found 564.3267.

2'-(Benzylamino)-3-(4-(t-butyl)phenyl)-[1,1'-binaphthalen]-2-ol (16). 3-(4-t-Butylphenyl)-2-naphthol¹² (103.6 mg, 0.375 mmol) and N-benzyl-2-aminonaphthalene 2f (58.3 mg, 0.25 mmol) were reacted according to the general procedure. The crude residue was purified by column chromatography (hexane/ethyl acetate 98:2) to afford compound 16 (80.1 mg, 63% yield) as a brown solid. ¹H NMR (CDCl₃, 400 MHz): δ 1.39 (s, 9H), 4.43 (s, 2H), 7.05–7.11 (m, 1H), 7.14–7.27 (m, 9H), 7.27–7.32 (m, 1H), 7.38 (ddd, 1H, *J* = 8.1, 6.8, 1.3 Hz), 7.49–7.55 (m, 2H), 7.69–7.74 (m, 2H), 7.75–7.81 (m, 1H), 7.86 (d, 1H, *J* = 8.6 Hz), 7.91 (dd, 1H, *J* = 8.0, 0.5 Hz), 8.00 (s, 1H). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 31.5, 34.8, 47.7, 108.6, 114.3, 114.9, 122.4, 123.7, 124.1, 124.7, 125.4, 126.9 (x 2C), 127.2, 127.4, 127.7, 128.3, 128.5, 128.7, 129.4, 129.8, 130.6, 130.7, 130.9, 132.9, 134.2, 135.1, 139.6, 145.1, 149.9, 150.6. HRMS (ESI) *m*/*z*: [M + H]⁺ calcd for C₃₇H₃₄NO 508.2635, found 508.2628.

2'-(Benzylamino)-3-(4-(t-butyl)phenyl)-3'-methoxy-[1,1'-binaphthalen]-2-ol (17). 3-(4-t-Butylphenyl)-2-naphthol¹² (103.6 mg, 0.375 mmol) and N-benzyl-3-methoxy-2-aminonaphthalene **2n** (65.8 mg, 0.25 mmol) were reacted according to the general procedure. The crude residue was purified by column chromatography (hexane/ethyl acetate 85:15) to afford compound **1**7 (122.7 mg, 91% yield) as a brown solid. ¹H NMR (CDCl₃, 400 MHz): δ 1.46 (s, 9H), 3.95 (q, 2H, *J* = 13.6 Hz), 4.08 (s, 3H), 6.95–7.07 (m, 2H), 7.19 (dt, 4H, *J* = 16.2, 7.3 Hz), 7.23–7.45 (m, 6H), 7.53–7.64 (m, 2H), 7.71–7.85 (m, 3H), 7.88–8.12 (m, 2H). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 31.5, 34.7, 50.8, 55.9, 106.6, 114.4, 117.3, 123.9, 124.0, 124.5, 124.8, 125.3, 125.4, 126.7, 126.8, 127.1, 127.8, 128.3, 128.4, 129.3, 129.4 (x 2C), 129.5, 130.4, 130.8, 133.5, 135.3, 138.5, 139.9, 149.6, 150.0, 150.4. HRMS (ESI) *m/z*: [M + H]⁺ calcd for C₃₈H₃₆NO₂ 538.2741, found 538.2748.

2,6-Dimethyl-4-(2-(piperidin-1-yl)naphthalen-1-yl)phenol (18). 2,6-Dimethylphenol 1b (45.8 mg, 0.375 mmol) and piperidino-2naphthalene 2e (52.8 mg, 0.25 mmol) were reacted according to the general procedure. In this reaction, addition of TFA (1.25 equiv) and t-BuOOt-Bu (1.5 equiv) was needed to ensure full consumption of 2aminonaphthalene. Then, the reaction was quenched by the addition of CH₂Cl₂ and sat. NaHCO₃. The organic phase was separated, dried over MgSO4, and evaporated under reduced pressure. The crude residue was purified by column chromatography (hexane/ethyl acetate 95:5) to afford compound 18 (78.6 mg, 95% yield) as a pale brown solid. ¹H NMR (CDCl₃, 400 MHz): δ 1.35-1.50 (m, 6H), 2.35 (s, 6H), 2.83–2.92 (m, 4H), 4.71 (bs, 1H), 7.08 (s, 2H), 7.29-7.37 (m, 2H), 7.39 (d, 1H, J = 8.8 Hz), 7.70-7.76 (m, 1H), 7.77–7.84 (m, 2H). $^{13}C{^{1}H}$ NMR (CDCl₃, 100 MHz): δ 16.1, 24.5, 26.5, 53.0, 120.4, 122.4, 123.7, 125.7, 125.8, 127.8, 128.1, 130.2, 130.3, 131.4, 131.6, 133.7, 149.3, 150.8. HRMS (ESI) *m/z*: [M + H]⁺

calcd for $C_{23}H_{26}NO$ 332.2009, found 332.2009. IR (KBr pellet): 3566 (s), 2932 (s), 1231 (s), 749 (s) cm⁻¹.

2-(2-(Benzylamino)-3-methoxynaphthalen-1-yl)-3,5,6-trimethylphenol (**19**). 2,3,5-Trimethylphenol (51.1 mg, 0.375 mmol) and Nbenzyl-3-methoxy-2-aminonaphthalene **2n** (65.8 mg, 0.25 mmol) were reacted according to the general procedure. The crude residue was purified by column chromatography (hexane/ethyl acetate 95:5) to afford compound **19** (60.6 mg, 61% yield) as a pale brown solid. ¹H NMR (CDCl₃, 400 MHz): δ 1.89 (s, 3H), 2.23 (s, 3H), 2.36 (s, 3H), 4.04 (s, 3H), 4.05 (d, 2H, *J* = 2.1 Hz), 6.77 (s, 1H), 7.13–7.34 (m, 9H), 7.72 (dd, 1H, *J* = 7.8, 0.6 Hz). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 12.0, 19.7, 20.2, 50.5, 55.7, 106.2, 115.9, 120.4 (x 2C), 123.5, 123.8, 123.9, 124.8, 126.7, 127.1, 128.0, 128.5, 129.1, 129.3, 135.6, 137.6, 137.7, 140.3, 150.0, 151.8. HRMS (ESI) *m*/*z*: [M + H]⁺ calcd for C₂₇H₂₈NO₂ 398.2115, found 398.2109.

2-(2-(Dimethylamino)naphthalen-1-yl)-6-methoxy-4-methylphenol (20). 2-Methoxy-4-methylphenol (48 µL, 0.375 mmol) and N,N-dimethyl-2-aminonaphthalene^{9f} 2b (42.8 mg, 0.25 mmol) were reacted according to the general procedure. In this reaction, addition of TFA (1.25 equiv) and t-BuOOt-Bu (1.5 equiv) was needed to ensure full consumption of the 2-aminonaphthalene. Then, the reaction was quenched by the addition of CH₂Cl₂ and sat. NaHCO₃. The organic phase was separated, dried over MgSO₄, and evaporated under reduced pressure. The crude residue was purified by column chromatography (hexane/ethyl acetate 90:10) to afford compound 20 (75.1 mg, 98% yield) as a pale brown solid. ¹H NMR (CDCl₃, 400 MHz): δ 2.37 (s, 3H), 2.72 (s, 6H), 3.96 (s, 3H), 6.73–6.83 (m, 2H), 7.32-7.40 (m, 2H), 7.44 (d, 1H, J = 8.9 Hz), 7.80 (dt, 2H, J = 5.6, 2.5 Hz), 7.86 (d, 1H, J = 8.8 Hz). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 21.4, 43.9, 56.1, 111.8, 117.8, 124.5, 125.4, 125.7, 126.2, 126.4, 127.9, 128.3, 128.9, 129.1, 131.0, 133.7, 142.1, 146.9, 149.8. HRMS (ESI) m/z: $[M + H]^+$ calcd for $C_{20}H_{22}NO_2$ 308.1645, found 308.1640.

2-(2-(Benzylamino)naphthalen-1-yl)-6-methoxy-4-methylphenol (21). 2-Methoxy-4-methylphenol (48 μL, 0.375 mmol) and Nbenzyl-2-aminonaphthalene 2f (58.3 mg, 0.25 mmol) were reacted according to the general procedure. The crude residue was purified by column chromatography (hexane/ethyl acetate 85:15) to afford compound 21 (50 mg, 54% yield) as a brown solid. ¹H NMR (CDCl₃, 400 MHz): δ 2.38 (s, 3H), 3.97 (s, 3H), 4.51 (s, 2H), 6.69 (d, 1H, *J* = 0.9 Hz), 6.83 (d, 1H, *J* = 1.8 Hz), 7.11 (d, 1H, *J* = 9.0 Hz), 7.19–7.28 (m, 2H), 7.29–7.36 (m, 6H), 7.73 (d, 1H, *J* = 8.0 Hz), 7.75 (d, 1H, *J* = 8.9 Hz). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 21.3, 48.0, 56.1, 111.9, 114.2, 122.0, 122.1, 124.0, 124.4, 126.5, 127.0 (x 3C), 127.5, 128.1, 128.7, 129.5, 130.3, 133.6, 139.9, 141.7, 143.4, 147.4. HRMS (ESI) *m/z*: [M + H]⁺ calcd for C₂₅H₂₄NO₂ 370.1802, found 370.1803.

2-(2-((4-Chlorobenzyl)amino)naphthalen-1-yl)-6-methoxy-4*methylphenol* (**22**). 2-Methoxy-4-methylphenol (48 µL, 0.375 mmol) and N-(4-chlorobenzyl)-2-aminonaphthalene 2 g (66.9 mg, 0.25 mmol) were reacted according to the general procedure. The crude residue was purified by column chromatography (hexane/ethyl acetate 85:15) to afford compound 22 (52.6 mg, 52% yield) as a pale brown solid. ¹H NMR (CDCl₃, 400 MHz): δ 2.36 (s, 3H), 3.97 (s, 3H), 4.45 (s, 2H), 6.64 (dd, 1H, J = 1.9, 0.8 Hz), 6.82 (d, 1H, J = 1.7 Hz), 7.01 (d, 1H, J = 9.0 Hz), 7.17-7.23 (m, 1H), 7.25 (s, 4H), 7.27-7.29 (m, 2H), 7.71 (d, 1H, J = 8.0 Hz), 7.72 (d, 1H, J = 9.0 Hz). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 21.3, 47.4, 56.1, 111.8, 114.0, 114.5, 121.9, 122.1, 124.1, 124.3, 126.6, 127.5, 128.2, 128.3, 128.8, 129.6, 130.4, 132.7, 133.6, 138.5, 141.6, 143.0, 147.4. HRMS (ESI) m/z: $[M + H]^+$ calcd for $C_{25}H_{23}ClNO_2$ 404.1412, found 404.1411. IR (KBr pellet): 3419 (s, broad), 2921 (s), 1506 (s), 1244 (m), 810 (s) cm^{-1}

2-Methoxy-6-(2-((4-methoxybenzyl)amino)naphthalen-1-yl)-4methylphenol (23). 2-Methoxy-4-methylphenol (48 μ L, 0.375 mmol) and N-(4-methoxybenzyl)-2-aminonaphthalene 21 (65.8 mg, 0.25 mmol) were reacted according to the general procedure. The crude residue was purified by column chromatography (hexane/ethyl acetate 85:15) to afford compound 23 (54.9 mg, 55% yield) as a brown solid. ¹H NMR (CDCl₃, 400 MHz): δ 2.38 (s, 3H), 3.80 (s, 3H), 3.99 (s, 3H), 4.45 (s, 2H), 6.67 (dd, 1H, J = 1.9, 0.8 Hz), 6.84 (d, 1H, *J* = 1.8 Hz), 6.85 (d, 1H, *J* = 2.1 Hz), 6.87 (d, 1H, *J* = 2.1 Hz), 7.13 (d, 1H, *J* = 9.0 Hz), 7.19–7.27 (m, 3H), 7.28–7.32 (m, 2H), 7.75 (t, 2H, *J* = 8.7 Hz). $^{13}C{}^{1}H$ NMR (CDCl₃, 100 MHz): δ 21.3, 47.5, 55.4, 56.1, 111.9, 114.0, 114.1, 114.3, 121.9, 122.1, 124.0, 124.4, 126.5, 127.4, 128.1, 128.2, 129.5, 130.3, 131.8, 133.6, 141.6, 143.5, 147.4, 158.7. HRMS (ESI) *m*/*z*: [M + H] + calcd for C₂₆H₂₆NO₃ 400.1907, found 400.1900.

2-(2-(Heptylamino)naphthalen-1-yl)-6-methoxy-4-methylphenol (24). 2-Methoxy-4-methylphenol (48 μL, 0.375 mmol) and Nheptyl-2-aminonaphthalene 2o (60.3 mg, 0.25 mmol) were reacted according to the general procedure. The crude residue was purified by column chromatography (hexane/ethyl acetate 85:15) to afford compound 24 (66 mg, 70% yield) as a brown syrup. ¹H NMR (CDCl₃, 400 MHz): δ 0.91 (t, 3H, J = 6.9 Hz), 1.25–1.35 (m, 8H), 1.52–1.60 (m, 2H), 2.38 (s, 3H), 3.25 (t, 2H, J = 7.1 Hz), 3.99 (s, 3H), 6.66 (dd, 1H, J = 1.9, 0.8 Hz), 6.84 (d, 1H, J = 1.8 Hz), 7.18– 7.24 (m, 2H), 7.27–7.33 (m, 2H), 7.76 (d, 1H, J = 8.1 Hz), 7.83 (d, 1H, J = 8.9 Hz). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 14.2, 21.3, 22.7, 27.0, 29.1, 29.8, 31.9, 44.4, 56.1, 111.8, 113.9, 114.2, 121.8, 122.2, 124.0, 124.4, 126.5, 127.3, 128.1, 129.5, 130.2, 133.6, 141.6, 143.9, 147.4. HRMS (ESI) m/z: [M + H]⁺ calcd for C₂₅H₃₂NO₂ 378.2428, found 378.2421.

2-(t-Butyl)-6-(2-((3-hydroxy-4-methoxybenzyl)amino)naphthalen-1-yl)-4-methoxyphenol (25). 2-t-Butyl-4-methoxyphenol (67.6 mg, 0.375 mmol) and N-(3-hydroxy-4-methoxybenzyl)-2aminonaphthalene 2h (69.8 mg, 0.25 mmol) were reacted according to the general procedure. The crude residue was purified by column chromatography (hexane/ethyl acetate 85:15) to afford compound 25 (105.2 mg, 92% yield) as a pale brown solid. ¹H NMR (CDCl₃, 400 MHz): δ 1.47 (s, 9H), 3.77 (s, 3H), 3.84 (s, 3H), 4.34 (d, 1H, J = 15.4 Hz), 4.39 (d, 1H, J = 15.5 Hz), 6.63 (d, 1H, J = 3.1 Hz), 6.73-6.80 (m, 2H), 6.87 (d, 1H, J = 1.6 Hz), 7.02 (d, 1H, J = 3.1 Hz), 7.14 (d, 1H, I = 9.0 Hz), 7.23 (ddd, 1H, I = 8.0, 6.4, 1.6 Hz), 7.26-7.35(m, 2H), 7.74 (d, 1H, J = 8.0 Hz), 7.78 (d, 1H, J = 8.9 Hz). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 29.6, 35.3, 47.6, 55.8, 56.1, 110.8, 112.3 (x 2C), 113.5, 114.3, 114.9, 118.5, 122.3, 122.4, 123.7, 127.1, 127.6, 128.2, 130.4, 132.7, 133.9, 138.5, 144.2, 145.8, 145.9, 146.7, 153.4. HRMS (ESI) m/z: $[M + H]^+$ calcd for C₂₉H₃₂NO₄ 458.2326, found 458.2311.

2-(2-(Butylamino)naphthalen-1-yl)-4,5-dimethoxyphenol (26). 3,4-Dimethoxyphenol (57.8 mg, 0.375 mmol) and N-butyl-2-aminonaphthalene 2d (49.8 mg, 0.25 mmol) were reacted according to the general procedure. The crude residue was purified by column chromatography (hexane/ethyl acetate 80:20) to afford compound 26 (60 mg, 68% yield) as a brown syrup. ¹H NMR (CDCl₃, 400 MHz): δ 0.91 (t, 3H, *J* = 7.3 Hz), 1.34 (sex, 2H, *J* = 7.5 Hz), 1.48–1.58 (m, 2H), 3.22 (t, 2H, *J* = 7.1 Hz), 3.81 (s, 3H), 3.96 (s, 3H), 6.70 (s, 1H), 6.75 (s, 1H), 7.17 (d, 1H, *J* = 9.0 Hz), 7.22 (ddd, 1H, *J* = 8.0, 6.0, 2.0 Hz), 7.27–7.35 (m, 2H), 7.74 (d, 1H, *J* = 8.1 Hz), 7.82 (d, 1H, *J* = 8.9 Hz). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 14.0, 20.2, 31.9, 43.9, 56.1, 56.5, 100.7, 111.6, 112.0, 114.0, 114.2, 122.2, 123.6, 127.1, 127.4, 128.3, 130.3, 134.1, 143.7, 144.7, 148.3, 150.2. HRMS (ESI) *m/z*: [M + H]⁺ calcd for C₂₂H₂₆NO₃ 352.1907, found 352.1909.

4-(2-((4-(t-Butyl)benzyl)amino)naphthalen-1-yl)-2-methoxy-6methylphenol (27). 2-Methoxy-6-methylphenol (51.8 mg, 0.375 mmol) and N-(4-t-butylbenzyl)-2-aminonaphthalene **2m** (72.4 mg, 0.25 mmol) were reacted according to the general procedure. The crude residue was purified by column chromatography (hexane/ethyl acetate 92:8) to afford compound **27** (75.5 mg, 71% yield) as a pale brown solid. ¹H NMR (CDCl₃, 400 MHz): δ 1.34 (s, 9H), 2.36 (s, 3H), 3.86 (s, 3H), 4.45 (s, 2H), 5.84 (bs, 1H), 6.72 (d, 1H, *J* = 1.3 Hz), 6.76 (d, 1H, *J* = 0.5 Hz), 7.18 (d, 1H, *J* = 9.0 Hz), 7.20–7.25 (m, 3H), 7.26–7.32 (m, 1H), 7.35 (d, 3H, *J* = 8.4 Hz), 7.74 (d, 1H, *J* = 8.0 Hz), 7.75 (d, 1H, *J* = 8.9 Hz). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 15.7, 31.5, 34.6, 48.1, 56.2, 111.0, 114.1, 120.4, 121.8, 124.4, 124.8, 125.6, 125.8, 126.2, 126.9, 127.3, 127.7, 128.0, 128.7, 134.0, 136.9, 143.1, 143.2, 147.0, 150.0. HRMS (ESI) *m/z*: [M + H]⁺ calcd for C₂₉H₃₂NO₂ 426.2428, found 426.2420.

(E)-2-(2-(Cinnamylamino)naphthalen-1-yl)-4,6-dimethylphenol (28). 2,4-Dimethylphenol (45 μL, 0.375 mmol) and N-cinnamyl-2-

aminonaphthalene **2k** (64.8 mg, 0.25 mmol) were reacted according to the general procedure. The crude residue was purified by column chromatography (hexane/ethyl acetate 97:3) to afford compound **28** (53.9 mg, 57% yield) as a yellow solid. ¹H NMR (CDCl₃, 400 MHz): δ 2.31 (s, 3H), 2.33 (s, 3H), 4.05 (dd, 2H, *J* = 5.4, 1.6 Hz), 6.25 (dt, 1H, *J* = 15.9, 5.4 Hz), 6.54 (d, 1H, *J* = 15.9 Hz), 6.85–6.89 (m, 1H), 7.07 (dd, 1H, *J* = 1.4, 0.6 Hz), 7.17–7.36 (m, 9H), 7.74 (d, 1H, *J* = 8.4 Hz), 7.82 (d, 1H, *J* = 8.9 Hz). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 16.4, 20.7, 46.0, 112.5, 114.2, 121.1, 122.3, 123.8, 125.0, 126.4, 127.1, 127.3, 127.6 (x 2C), 128.2, 128.7, 129.7, 130.1, 130.2, 131.2, 132.1, 133.9, 136.8, 144.2, 150.0. HRMS (ESI) *m*/*z*: [M + H]⁺ calcd for C₂₇H₂₆NO 380.2009, found 380.2001.

4-(2-((Furan-2-ylmethyl)amino)naphthalen-1-yl)-2,5-dimethylphenol (**29**). 2,5-Dimethylphenol (45.8 mg, 0.375 mmol) and N-(2-furylmethyl)-2-aminonaphthalene **2p** (55.8 mg, 0.25 mmol) were reacted according to the general procedure. The crude residue was purified by column chromatography (hexane/ethyl acetate 90:10) to afford compound **29** (33 mg, 38% yield) as a brown solid. ¹H NMR (CDCl₃, 400 MHz): δ 1.85 (s, 3H), 2.25 (s, 3H), 4.41 (s, 2H), 6.11 (dd, 1H, *J* = 3.2, 0.8 Hz), 6.26 (dd, 1H, *J* = 3.2, 1.9 Hz), 6.81 (s, 1H), 6.88 (s, 1H), 7.08–7.14 (m, 1H), 7.18–7.32 (m, 4H), 7.72–7.75 (m, 1H), 7.77 (d, 1H, *J* = 8.8 Hz). ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 15.5, 19.1, 41.9, 106.8, 110.4, 114.5, 117.1, 120.2, 122.1 (x 2C), 124.3, 126.3, 127.7, 127.8, 128.0, 128.5, 133.8 (x 2C), 137.4, 141.8, 142.5, 153.4, 153.6. HRMS (ESI) *m/z*: [M + H]⁺ calcd for C₂₃H₂₂NO₂ 344.1645, found 344.1646.

4-(2-(Benzylamino)naphthalen-1-yl)-2,6-dimethylphenol (**30**). 2,6-Dimethylphenol **1b** (45.8 mg, 0.375 mmol) and N-benzyl-2aminonaphthalene **2f** (58.3 mg, 0.25 mmol) were reacted according to the general procedure. The crude residue was purified by column chromatography (hexane/ethyl acetate 90:10) to afford compound **30** (66.9 mg, 76% yield) as an orange syrup. The reaction was also performed on a 2 mmol scale, affording compound **30** (523 mg, 74% yield). ¹H NMR (CDCl₃, 400 MHz): δ 2.29 (s, 6H), 4.43 (s, 2H), 6.92–6.94 (m, 2H), 7.08 (d, 1H, *J* = 8.9 Hz), 7.16 (ddd, 1H, *J* = 8.0, 6.5, 1.5 Hz), 7.20–7.32 (m, 7H), 7.67 (dd, 1H, *J* = 1.9, 1.1 Hz), 7.69 (s, 1H). ¹³C{¹H}NMR (CDCl₃, 100 MHz): δ 16.1, 48.5, 114.4, 120.4, 121.8, 124.1, 124.5, 126.1, 127.1 (x 2C), 127.4, 127.9, 128.5, 128.6, 128.7, 131.3, 134.1, 140.0, 143.0, 151.8. HRMS (ESI) *m/z*: [M + H]⁺ calcd for C₂₅H₂₄NO 354.1852, found 354.1848. IR (KBr pellet): 3419 (s, broad), 2924 (m), 1200 (s), 746 (m) cm⁻¹.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acs.joc.0c00874.

Kinetic data and mechanistic studies as well as full spectroscopic data for all compounds (PDF)

X-ray diffraction analysis results of compound 20 (CIF)

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Notes

The authors declare no competing financial interest.

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