RESEARCH ARTICLE

Transfusion-transmitted arboviruses: Update and systematic review

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Abstract

Background

The detection of the first cases of transfusion-transmitted West Nile virus in 2002 posed a new challenge for transfusion safety. Institutions like the World Health Organization have stated that blood transfusion centers need to know the epidemiology of the different emerging infectious agents and their impact on blood transfusion. The aim of the study is to review the published cases of arbovirus transmission through transfusion of blood or blood components and to analyze their main clinical and epidemiological characteristics.

Material and methods

Systematic literature searches were conducted in MEDLINE, Embase and Scopus. Pairs of review authors selected a variety of scientific publications reporting cases of transfusion-transmitted arboviruses. Main clinical and epidemiological characteristics were reviewed of the cases described. The study protocol was registered in PROSPERO CRD42021270355.

Results

A total of 74 cases of transfusion-transmitted infections were identified from 10 arboviruses: West Nile virus (n = 42), dengue virus (n = 18), Zika virus (n = 3), yellow fever vaccine virus (n = 3), tick-borne encephalitis virus (n = 2), Japanese encephalitis virus (n = 2), Powassan virus (n = 1), St. Louis encephalitis virus (n = 1), Ross River virus (n = 1) and Colorado tick fever virus (n = 1). The blood component most commonly involved was red blood cells (N = 35, 47.3%; 95% confidence interval [CI] 35.9% to 58.7%). In 54.1% (N = 40; 95% CI: 42.7%-65.47%) of the cases, the recipient was immunosuppressed. Transmission resulted in death in 18.9% (N = 14; 95% CI: 10.0%-27.8%) of the recipients. In addition, 18 additional arboviruses were identified with a potential threat to transfusion safety.



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Discussion

In the last 20 years, the number of published cases of transfusion-transmitted arboviruses increased notably, implicating new arboviruses. In addition, a significant number of arboviruses that may pose a threat to transfusion safety were detected. In the coming years, it is expected that transmission of arboviruses will continue to expand globally. It is therefore essential that all responsible agencies prepare for this potential threat to transfusion safety.

Author summary

The transfusion of blood and blood components entails some risks and potential complications, chief among them the transmission of infectious agents. Organizations like the American Association of Blood Banks have warned of the risks posed by emerging and reemerging viruses for transfusion safety and have classified transmission of several arboviruses as a high or very high risk to the transfusion of blood and blood components. Following recommendations by the World Health Organization (WHO), this study aims to enable safe blood transfusion services, by making available the latest updated information of transfusion-transmitted arboviruses and comprehensive knowledge of the current epidemiology of reported cases.

Research revealed case reports of transfusion transmission of 10 arboviruses, with West Nile virus and dengue virus as the most prevalent. Main clinical characteristics of reported cases were collected, including the type of blood component transfused and the state of immunosuppression of the recipient. Research also revealed 18 additional arboviruses with potential risk of transmission through other direct transmission routes. This systematic review provides an updated overview of the clinical characteristics of reported cases of transfusion-transmitted arboviruses. It is the most complete record published to date that assesses the risk posed by arboviruses to blood transfusion.

Background

Throughout history, transfusion safety has been threatened by the emergence of various infectious agents [1]. Currently, health service consumers expect a total absence of transfusionrelated transmission of infectious diseases. However, the 2002 notification of the first cases of West Nile virus transmission through transfusion of blood components, highlighted the threat posed to transfusion safety by emerging and re-emerging infectious agents [2,3].

Among the emerging viruses, arboviruses are especially relevant due to their known or theoretical potential for transmission through blood transfusion. These viruses are transmitted by arthropods, mainly mosquitoes (genera *Culex* spp. and *Aedes* spp.) and ticks (particularly the genus *Ixodes*) but also flies, dragonflies, and lice. In humans, they can cause clinical presentations ranging from asymptomatic infections or mild flu-like symptoms, to severe, potentially fatal hemorrhagic and neurological syndromes. Their epidemiology is highly variable and unpredictable and may manifest as isolated and localized cases; seasonal waves; or widespread, epidemic outbreaks. Originally from tropical regions, arboviruses have now spread widely, producing numerous epidemic outbreaks over the past 20 years and constituting a major public health problem all over the planet. Arboviruses are classified into different taxonomic families: *Flaviviridae*, *Bunyaviridae*, *Togaviridae*, *Rhabdoviridae*, *Reoviridae*, and *Asfarviridae*. The most important arboviruses that cause disease in humans belong to the families *Flaviviridae*, *Togaviridae* and *Bunyaviridae* [4].

Threats to transfusion safety arising from new infectious agents have traditionally been addressed through the development of methods to screen blood donors, and this also holds true for arboviruses. Screening methods used include serological and non-serological antigen rapid testing techniques as well as nucleic acid (PCR) tests. These tests may be used routinely or according to the epidemiological profile of the areas or countries in question. However, they may not be sufficient to fully guarantee transfusion safety in the case of donors with a very low viral load and/or who are in the window period after infection [5].

In recent years, various international organizations have created tools to assess the transfusion risk posed by these newly emerging and re-emerging infectious agents. In 2009, the AABB Transfusion-Transmitted Diseases Committee, in collaboration with members of the Food and Drug Administration (FDA), the Centers for Disease Control and Prevention (CDC), and other agencies, developed guidance to identify infectious agents that could pose a real or potential risk to transfusion safety. In this process, 68 infectious agents, including arboviruses, were classified according to their epidemiological profile and the public's awareness and perception of them. Of the arboviruses in this classification, dengue virus was assigned in the highest risk category, while chikungunya and St. Louis encephalitis viruses were included in the second-highest risk category [6]. In 2011, fact sheets were developed for miscellaneous arboviruses, including for yellow fever virus and yellow fever vaccine virus [7]. Various infectious agents were also classified in 2011 by the European Center for Disease Prevention and Control (ECDC) based on the epidemiological and environmental characteristics of these viruses in Europe. The agents given the highest priority include a number of arboviruses: West Nile virus, dengue virus, chikungunya virus and tick-borne encephalitis virus [8]. In Asia, the WHO also published the Asia Pacific Strategy for Emerging Diseases and Public Health Emergencies (APSED III) in 2017 with the aim of detecting and addressing the different emerging infectious diseases in the region [9].

With the development of these risk assessment tools, major global public health entities have expressed the need for comprehensive knowledge of the epidemiological characteristics of the various emerging transfusion-transmitted infections and to evaluate their potential impact on donor selection criteria and the supply of blood and blood products [10].

The main objective of this study is, therefore, to review the published cases of arbovirus transmission through the transfusion of blood or blood components throughout the history of transfusion and to study their main clinical and epidemiological characteristics. For the research question in PICOT format, please see (S1 Table). The secondary objective is to, additionally, identify arboviruses that can be a threat to transfusion safety through other non-vector, non blood transfusion related direct routes of transmission such as: organ and/or hematopoietic stem cell transplantation, mother-to-child transmission, direct blood contact or by their prevalence in blood donors.

Materials and methods

Design

This systematic literature review was designed and conducted in accordance with the *Cochrane Handbook of Systematic Reviews of Interventions* and reported in line with the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) Statement [11]. For the PRISMA checklist, please see the (<u>S1 PRISMA Checklist</u>). The protocol for this systematic review was published and registered in PROSPERO (www.crd.york.ac.uk/prospero, CRD42021270355).

First, a global descriptive analysis was completed including all cases of transfusion-transmitted arboviruses. Consequently, a brief study was conducted of the main clinical and epidemiological characteristics of reported cases of arboviral transfusion transmission. After a general analysis that included all cases, we specifically analyzed the arboviruses with the most reported cases of transmission through blood transfusion or blood components: West Nile virus and dengue virus. Finally, a descriptive analysis was performed of arboviruses that can be considered a potential threat to transfusion safety, based on published reports of any non-vector, non-blood related direct route of transmission or because of their prevalence in blood donors, regardless of reported evidence of transfusion transmission.

Source of data collection

Systematic literature searches were conducted in MEDLINE, Embase, Scopus using the server of University Miguel Hernández server.

Search strategy

The complete search strategy is described in more detail in the study protocol and (<u>S1 Text</u>). Initially, the authors have performed searches with free text terms in all three bibliographic databases using the term "transfusion" in combination with the name of the arboviruses being reviewed. In MEDLINE, Medical Subject Headings (MeSH) such as "Blood Transfusion"[-Mesh] OR "Transfusion Reaction"[Mesh] also served as search terms. In Embase the Entree thesaurus was used to search articles based on the following terms: "blood transfusion"/exp.

As a second search strategy, free text searches were performed in MEDLINE using the terms: "transplantation", "vertical transmission", "blood donors", while in Embase a free text search was conducted by entering the following terms in the Entree Thesaurus: "transplantation"/exp OR transplantation, "vertical transmission"/exp OR "vertical transmission" AND "blood donor"/exp OR "blood donor". In Scopus, we ran searches only using free text terms in the fields for title, abstract, and key words. Bibliographies of all included records to identify other potentially relevant publications were also hand-searched.

The authors included records published from the year of inception of each bibliographic database (start date—we did not apply any specific limits to the start date) to 10 November 2021 (end date). The records were entered into the Mendeley Desktop reference manager (Elsevier).

Study selection

In answer to the primary objective, selected publications were those in any language describing cases of transfusion-transmitted arboviruses transmitted through transfusion of blood or blood components. A wide variety of scientific studies (original articles, reviews, brief reports, case reports, letters to the editor, conference papers and others) were included reporting cases of arboviruses transmitted through transfusion of blood or blood components. Publications that collected cases of transmission in non-humans were excluded, as were publications that did not detail the specific type of arbovirus transmitted through the transfusion of blood or blo

Search records were independently evaluated by two review authors for inclusion in the publication overview. Any discrepancies were resolved by consensus. Inter-observer agreement was measured by determining the kappa index.

The bibliographic reference manager Mendeley Desktop was used to create folders containing records for each virus, eliminating duplicates. Titles and abstracts were screened to preselect records in any language that included general information on the possible transmission of arboviruses through the transfusion of blood or blood components. Consequently, full texts of all potentially relevant records were reviewed independently by two researchers. Records included, were those that reported cases of transfusion transmission of blood or blood components in humans over the entire period under study.

To answer to the secondary objective, publications in any language were first selected, describing cases stemming from direct non-vector or transfusion related routes of arbovirus transmission (e.g.: organ and/or hematopoietic stem cell transplantation, mother-to-child transmission, direct blood contact) or reports of prevalence of these arboviruses in blood donors. Consequently, a wide variety of records (original articles, reviews, brief reports, case reports, letters to the editor, conference papers and others) were included to accommodate this review. Lastly, for each encountered route of transmission and each arbovirus found prevalent in blood donors, one publication was selected for the produced publication overview, representing the routes of transmission or evidence of prevalence in screened blood donors. Representative publication, or publication in a journal with the highest impact factor. This evaluation was independently done by two review authors. Final selection for inclusion in the publication overview was determined by consensus. Inter-observer agreement was measured by determining the kappa index.

Data extraction and analysis

All data of cases of transfusion-transmitted arboviruses were extracted from the selected publications and directly entered into a Microsoft Excel database by a single review author. Data quality control was consequently done by a second review author. If a publication included more than one case of transfusion transmission, each case was recorded and entered separately in the database. Basic epidemiological and clinical data of the reported cases of transfusion transmission were then analyzed: type of blood component implicated, immunosuppression and clinical presentation in the recipient, fatality of the outcome following transmission as well as the plausibility of the respective transmission route. From the selected publications, study authors conclusions on transfusion transmission and mortality, attributable to the arboviral infection, were recorded. Determinants of potential alternative routes of transmission were additionally collected such as: exposure to the vector, prolonged hospital stay, additional patients infected by the same contaminated donated product or whether the authors themselves ruled out other potential explanations for the patient's infection.

Results were expressed as proportion with a 95% confidence interval (95% CI). The proportion was calculated as the number of the reported parameter under study divided by the total number of reported cases of transfusion transmission (Example: number of cases of transfusion-transmitted arboviruses related with the transfusion of red blood cells / total number of cases of transfusion-transmitted arboviruses x 100). The 95% CIs were for proportions $(p\pm Z_{\alpha/2}\sqrt{(p(1-p)/n)})$.

Quality assessment

Due to the mainly descriptive objective of this study and the type of publications included (case reports, letters to the editor), no standard quality control tool, such as ROBINS-I or STROBE checklist, was applied for assessing risk of bias in non-randomized studies.

Results

The search strategy yielded a total of 5158 records. After eliminating duplicates, the authors screened the title and abstract of 2918 records and retrieved the full text of 199. The authors

selected any records that suggest human transmission of arboviruses through blood transfusion or studies that display information that could facilitate the identification of other publications that report cases of transfusional transmission of arboviruses. Finally, 29 studies reporting cases of human transmission of arboviruses through transfusion of blood or blood components were included. Fig 1 presents the PRISMA flow chart, describing the selection process for including published studies of transfusion-transmitted arboviruses.

Of the 29 records included, 11 studies described transfusion-transmitted West Nile virus; 9, dengue virus; 2, Zika virus; 1 Japanese encephalitis virus; 1, St. Louis encephalitis virus; 1, tickborne encephalitis virus; 1, Powassan virus; 1, yellow fever virus; 1, Colorado tick fever virus; and 1, Ross River fever virus. Interobserver concordance was very high ($\kappa = 0.97$). For the PRISMA flow chart assessing each specific arbovirus, please see the (S1 Flow Chart).

For the PRISMA flow chart with the process of selecting records describing non-vector and non-transfusion related direct routes of transmission of arboviruses and publications reporting the prevalence of arboviruses in blood donors, please see (S2 Flow Chart). A total of 32 records were selected for the publication overview with an interobserver K value of 0.84.



Fig 1. PRISMA flow chart on selection of included studies covered by the literature review.

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Analysis of results

Included studies described 74 cases of suspected transfusion transmission of 10 arboviruses. Most cases involved transfusion transmission of West Nile virus (N = 42, 56.8%; 95% CI: 45.5–68.0%) and dengue virus (N = 18, 24.3%; 95% CI: 14.5–34.1%). Remaining cases described transfusion transmission of Zika virus, yellow Fever vaccine virus, tick-borne encephalitis virus, Japanese encephalitis virus, Powassan virus, St. Louis encephalitis virus, Ross River virus and Colorado tick fever virus (Fig 2).

The USA was the country with the most reported cases of transfusion transmission (N = 42, 62%.2%, 95% CI: 51.1–73,2%). The main type of blood component implicated was red blood cells (N = 35, 47.3%; 95% CI: 35.9–58.7%).

Over half of the reported transfusion-transmitted arbovirus cases, selected for the overview, were described in recipients classified as immunocompromised (N = 40, 54.1%; 95% CI: 42.7–65.5%). Arboviral morbidity and mortality was reported in 63.5% (N = 47; 95% CI: 52.5–74.5%) and 18.9% (N = 14; 95% CI: 10.0–27.8%) of cases described, respectively. Recipient mortality was mostly reported for the West Nile virus (N = 13; 92.9%; 95% CI: 79.4–100%) and 78.6% (N = 13; 95% CI: 57.1–100%). of fatal cases the recipient was classified as immuno-compromised. Plausibility of transfusion-transmission was confirmed by the authors of the selected records in 50.0% (N = 37; 95% CI: 38.6–61.4%) of cases. For the main clinical and epidemiological characteristics of selected cases, see Fig 3.

Regarding West Nile virus, 90.5% (N = 38; 95% CI: 81.6–99.4%) of reported cases were diagnosed in the USA, and the main type of blood component implicated was red blood cells 54.8% (N = 23; 95% CI: 39.7–69.8%). Symptoms as a result of transmission were reported in 73.8% (N = 31; 95% CI: 60.5–87.1%) of recipients and in 31.0% (N = 13; 95% CI: 17.0–44.9%) death was reported as the ultimate outcome of transmission. Fig 4 shows the main reported clinical and epidemiological characteristics of cases of West Nile virus transfusion transmission.

Of the reported transfusion transmitted cases related to dengue virus, 50% (N = 9; 95% CI: 26.9–73.1%) originated from Brazil. Transfusion of red blood cells was the main route of transmission described (N = 8, 44.4%; 95% CI: 21.5–67.4%). Symptoms were reported for 61.1% (N = 11; 95% CI: 38.6–83.6%) of recipients. None of the selected cases of transfusion



Fig 2. Reported transfusion-transmitted arbovirus cases.

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Country Australia (n=1) Brazil (n=12) China (n=3) Finland (n=2) Greece (n=4) Pakistan (n=2) Singapore (n=4) U.S.A. (n=42) Total (n=74)					Proportion [95% Cl] 0.01 [-0.01, 0.04] 0.16 [0.08, 0.24] 0.04 [0.00, 0.09] 0.03 [-0.01, 0.06] 0.05 [0.00, 0.11] 0.03 [-0.01, 0.06] 0.05 [0.00, 0.11] 0.05 [0.00, 0.11]
Blood product Red blood cells (n=35) Platelets (n=21) Fresh frozen plasma (r Granulocytes Not specified Total (n=74)	n=11) .∎ ∎		a		0.47 [0.36, 0.59] 0.28 [0.18, 0.39] 0.15 [0.07, 0.23] 0.01 [-0.01, 0.04] 0.08 [0.02, 0.14]
Status of immunosup Yes (n=40) No (n=24) Not specified (n=10) Total (n=74)	pression 				0.54 [0.43, 0.65] 0.32 [0.22, 0.43] 0.14 [0.06, 0.21]
Morbidity Yes (n=47) No (n=20) Not specified (n=7) Total (n=74)		.			0.64 [0.53, 0.75] 0.27 [0.17, 0.37] 0.09 [0.03, 0.16]
Mortality Yes (n=14) No (n=48) Not specified (n=12) Total (n=74)		••••••			0.19 [0.10, 0.28] 0.65 [0.54, 0.76] 0.16 [0.08, 0.25]
Plausibility of transfus Confirmed (n=37) Probable (n=21) Possible (n=5) Not specified (n=11) Total (n=74)	sion transn	nission 			0.50 [0.39, 0.61] 0.28 [0.18, 0.39] 0.07 [0.01, 0.13] 0.15 [0.07, 0.23]
	0,00	0,20	0,40	0,60	0,80

Fig 3. Main clinical and epidemiological characteristics of reported cases of transfusion-transmitted arboviruses.

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transmission of dengue virus were reported as fatal. In 38.9% (N = 7; 95% CI: 16.4–61.4%) of cases, the serotype involved was DENV-4. Fig 5 presents the main reported clinical and epidemiological characteristics of cases of transfusion transmission of dengue virus.

Tables 1, 2 and 3 provide an overview of the main the main characteristics of reported cases of transfusion-transmitted arboviruses apart from dengue and West Nile virus.

Arboviruses with a potential threat to transfusion safety

In addition to the 10 arboviruses involved in reported cases of transfusion-transmitted infections, 18 additional arboviruses were identified with a potential threat to transfusion safety by means of organ and/or hematopoietic stem cell transplantation, mother-to-child transmission, direct blood contact and/or prevalence in blood donors. The characteristics of published cases on these arboviruses are listed in <u>Table 4</u>. Reference studies on these other arboviruses describing the risk of direct transmission, are depicted in <u>Table 5</u>.

Discussion

Until the year 2000, transfusion transmission of arboviruses was almost anecdotal, with only one isolated case of transmission of Colorado tick fever virus and two cases of tick-borne



Fig 4. Main reported clinical and epidemiological characteristics of transfusion-transmitted West Nile virus.

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encephalitis virus. However, after the West Nile virus outbreak in the USA in 2002, the number of cases increased considerably [46]. Since then, more than 70 cases of arbovirus transmission have been published, including several cases of emerging arboviruses: Japanese encephalitis virus, Ross River virus, St. Louis encephalitis virus, Powassan virus, and Zika virus [13,15,17,19,26,27].

This study confirms the possibility that arboviruses can be transmitted through transfusion of blood and blood components. A total of 74 reported cases of suspected transfusion-transmitted infections from 10 arboviruses were identified. Infections were reported to be symptomatic in 63.5% and fatal in 18.9% of the reported cases. This transmission route therefore poses a considerable threat to transfusion safety, reportedly causing significant morbidity and mortality and an increased health spending. As more than 50% of affected patients were classified as immunocompromised, transmission of arboviruses through blood or blood components can additionally be considered a threat to a particularly vulnerable section of the population. Analysis of reported cases revealed that transfusion transmission occurred through various blood products. It is observed that transmission through platelet transfusion was found to be higher in cases of dengue virus transmission, compared to other arboviral transmission cases, including the West Nile virus cases. This finding could be related to signs of tropism of the dengue virus against platelets or at least some of their components, such as glycoprotein Ib (CD42b) [82].

In addition to the published cases of transfusion transmission of arboviruses, our review identified 18 additional arboviruses with potential threat to transfusion safety through solid

Table 1. T	ransfusion-	transmitted	arboviri	Ises cas	es.		-	-									
Virus	Publication	Type of	Country	Year	Blood	Recipient	C	Morbidity	Mortality	Diagnostic		Alternative e:	vplanation				Other non-
		publication			product					method	Exposure to Vector	Prolonged hospital stay	Other infected recipients	Non- vector vias rejected	Plausibility	Additional data	vector, non- blood transfusion related transmission
Colorado tick fever virus	Randall 1975 [12]	Case report	U.S.A.	1975	Not specified (WB?)	Colon carcinoma	Yes	Yes	No	Clinical Not specified	No	I	I	I	I	I	Vertical transmission[13]
Japanese encephalitis virus	Cheng 2018 [14]	Original article	China	2017	RBC	Double lung transplantation	Yes	Yes	Yes	RT-PCR Sanger sequencing Phylogenetic analysis	N.P.	Yes	Yes	Yes	Confirmed	Same donor No data available about PL recipient	Screening in blood donors [15] Vertical transmission[12]
					Platelets	Acute myeloid leukemia	Yes	No	No	Serology IgM	I	I	Yes	I			
Ross River virus	Hoad 2015 [16]	Case reports	Australia	2014	RBC	Myelodysplastic syndrome	Yes	°N	No	Serology IgM RT-PCR	N.P.	1	No	1	1	I	Screening in blood donors [17] Vertical transmission[12]
St. Louis encephalitis virus	Venkat 2017 [18]	Original article	U.S.A.	2015	RBC	Kidney transplant	Yes	Yes	I	Serology IgM+ PRNT	N.P.	Yes	No	Yes	Possible	No data available about PL recipient.	Screening in blood donors [19]
Powassan virus	Taylor 2020 [20]	Original article	U.S.A.	2018	RBC	Kidney transplant	Yes	Yes	No	Serology IgM+ PRNT RT-PCR	No	Yes	No *	Yes	Probable	1	1
Tick-borne encephalitis	Wahlberg 1989 [21]	Original article	Finland	1959- 1987	Not specified	Not specified	1	Yes	No	Not specified	N.P.	I	I	I	I	I	Blood contact [22]
virus					Not specified	Not specified	1	Yes	No	Not specified	N.P.	1	1	1	1		S.O.T. transmission[23] Screening in blood donors [24]
Yellow fever vaccine virus	Lederman 2010 [25]	Original article	U.S.A.	2009	Platelets (irradiated)	Wilm's tumor (relapsed), QMT	Yes	No	No	Serology IgM+ PRNT	1	I	I	I	I	2 exposed recipients	Screening in blood donors
					Platelets	Kidney/liver transplantation, DM	Yes	No	No		1	1	1	1	I	without evidence of transmission	[26] Vertical transmission[12]
					FFP (2 units)	CKF, TBC, psoriasis (infliximab)	Yes	No	No		1	I	I	1	1		
Zika virus	Barjas- Castro 2016 [27]	Case report	Brazil	2015	Platelets (Pool)	Liver transplantation	Yes	No	No	Virus isolation RT-PCR Phylogenetic analysis	No	No	No *	I	Probable		Blood contact [29] Screening in blood donors
	Motta 2016 [28]	Letter to the editor	Brazil	2016	Platelets (apheresis)	Primary myelofibrosis	Yes	No	No	Serology IgG/ IgM+	I	I	Yes	I	Confirmed	Same donor	[30] Vertical
					Platelets (apheresis)	Acute myeloid leukemia—BMT	Yes	Ŷ	No	PRNT RT-PCR Molecular sequencing Phylogenetic analysis	No	Yes	Yes	1			TCHORSTINGING
BMT: bone neutralizati virus. RNA	marrow tra on test, TBC Ribonuclei	insplantation 2 Tuberculos 6 Acid	n, CKD: (sis, RBC 1	chronic red blo	kidney dis [,] od cells, IC:	ease, CTX: cherr Immunocompi	other omise	apy, DM: ed, RT-PC	diabetes R Revers	mellitus, FFP ie transcriptioi	Fresh froza n polymera	en plasma, ase chain r	NP: not pi eaction, SC	obable, I)T: Solid	PL: plasma, organ tran	PRNT Plaque splantation, Zl	reduction .KV: Zika

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* PL not transfused was positive

– Data not available

	Other non-	vector, non- blood transfusion related transmission		Blood contact (needle-stick	related transmissión)	[41]HSCTtransmission[42]Screening in	blood donors [43] S.O.T. transmission [44]	Vertical transmission [45]											(Constand)
	Additional	data		Same donor					Different	donors						Same donor RBC	recipient was not tested for dengue virus	Different donors	
	Plausibility		Confirmed	I	I	1	Confirmed	Confirmed	Probable	Probable	Probable	Probable	Probable	Possible	Confirmed	Possible	Possible	Probable	Possible
		Non- vector vias rejected	I	I	I	1	1	1	I	I	I	I	I	I	1	Yes	Yes	I	1
	planation	Other infected recipients	1	Yes	Yes	Yes	No *	No	I	I	I	I	I	I	1	Yes	Yes	I	I
	Alternative ex	Prolonged hospital stay	1	I	I	Yes	Yes	Yes	I	1	1	1	1	I	Yes	Yes	No	1	1
	4	Exposure to Vector	1	I	1	N.P.	°Z	N.P.	I	1	1	1	1	I	N.P.	N.P.	N.P.	I	1
	Serotype		-	2			2	-	4						1	I		4	1
	Diagnostic	method	Serology IgM + HI RT-PCR	Serology IgG/IgM	PCR Sequencing		Serology IgG/IgM RT-PCR Sanger sequencing	Serology IgG/IgM RT-PCR	Serology	IgG/IgM	NAT				Serology IgM RT-PCR Sequencing NS1 Phylogenetic analysis	Serology IgM RT-PCR		Serology IgM/IgG RT-PCR	Serology IgM
	Mortality		No	No	No	No	No	No	I	I	1	I	1	I	1	I	1	Ŋ	No
	Morbidity		Yes	Yes	Ŷ	Yes	Yes	Yes	I	1		I	1	I	Yes	Yes	Yes	Yes	Yes
	IC. Wo		No	Yes	Yes	Yes	No	Yes	I	1	1	Т	Т	I	Yes	No	°Z	Yes	Yes
	Recipient		Bronchiectasis, vitamin B12 deficiency anaemia	DM, ischemic heart disease	Hepatocellular carcinoma	DM, hypertension, ischemic heart disease, CRI.	Gastrointestinal bleeding.	Aplastic anemia and neutropenic colitis.	Not specified	Not specified	Not specified	Not specified	Not specified	Not specified	Lymphoblastic lymphoma— autologus H.S.C. T.	Coronary artery bypass grafting	Coronary artery bypass surgery	Lung cancer, coronary heart disease	Pancreatic cancer, DM, dementia.
VII US CASES	Blood	Product RBC RBC Platelets FFP RBC RBC RBC RBC RBC RBC RBC RBC RBC RBC		Platelets (apheresis)	Platelets	FFP	RBC	RBC	Platelets	FFP (2 units)	Platelets	Platelets	FFP	RBC	RBC				
anguan n	Year		2002	2007		-	Not specified, 2014?	2014	2014 1 ((2012			Not specified, 2016? 2016?	2015	·	2010	2011	
ין מוואוווונונ	Country		China	Singapore			Singapore	Brazil	Brazil						Brazil	Pakistan		U.S.A.	
-IIOISUUSIUS	Type of	publication	Original article	Letter to the editor			Case report	Original article	Original	article					Conference abstract	Original article		Original article	
1 aute 2. 11	Publication		Chuang 2008 [32]	Tambyah 2008 [33]			Boon Oh 2015 [34]	Levi 2015 [35]	Sabino	2015 [36]					Alves 2016 [37]	Karim 2016 [38]		Matos 2016 [39]	

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Other non-	vector, non- blood transfusion related transmission					
Additional	data					
Plausibility		Confirmed				
	Non- vector vias rejected	I				
xplanation	Other infected recipients	I				
Alternative e	Prolonged hospital stay	I				
	Exposure to Vector	I				
Serotype		2				
Diagnostic	method	RT-PCR	In house	sequencing	Phylogenetic	analysis
Mortality		No				
Morbidity		Yes				
IC.		Yes				
Recipient		Sickle cell	disease on	chronic	transfusion	therapy
Blood	product	RBC				
Year		2019				
Country		Brazil				
Type of	publication	Case report				
Publication		Santos 2020	[40]			

CKD: chronic kidney disease, DM: diabetes mellitus, FFP: fresh frozen plasma, HI: hemagglutination inhibition assay, HSCT: hematopoietic stem cell transplantation, IC: immunocompromised, PL: plasma, NAT: nucleic acid testing, NP: not probable, RBC: red blood cells, RT-PCR: reverse transcription polymerase chain reaction, SOT: solid organ transplantation.

Data not available
* PL not transfused was positive

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Publication	Type of	Country	Year	Blood product	ases. Recipient	IC.	Morbility	Mortality	Diagnostic		Alternative e	xplanation		Plausibility	Addtitional	Other non-vector,
	publication								method	Exposure to Vector	vProlonged hospital stay	Other infected recipients	Non- vector vias rejected		data	non-blood transfusion related transmission
Pealer 2003	Original	U.S.A.	2002	RBC	Post partum	No	Yes	No	Serology	Possible	I	I	I	Confirmed	16 implicated	S.O.T.
[46]	article			Platelets	Liver transplantation	Yes	Yes	No	PT_DCP	I	I	I	I	Confirmed	blood donors	transmission [57] Screening in
				RBC	Post partum	No	Yes	No	VOJ-IV	I	I	I	I	Confirmed		blood donors [58]
				Platelets (2 units)	Acute myeloid leukemya	Yes	Yes	No		I	1	1	1	Confirmed		Vertical transmission [12]
				RBC	Lung cancer	Yes	Yes	Yes		1	1	1	1	Confirmed		
				FFP	Breast cancer	Yes	Yes	No		1	1	Yes	1	Confirmed		
				Platelets	Acute myeloid leukemya—BMT	Yes	Yes	Yes		I	1	1	1	Confirmed		
				FFP	Aneurysm repair	No	No	No			ı	Yes	1	Confirmed		
				RBC	Acute myeloid leukemya—HSCT	Yes	No	No		I	1	Yes	1	Confirmed		
				Platelets	Rhabdomyosarcoma	Yes	Yes	No		I	I	I	I	Confirmed		
				RBC	Pneumonia, septic shock	No	No	No		I	I	Yes	I	Confirmed		
				RBC	Coronary-artery bypass grafting	ů	Yes	Yes		N.P.	Yes	I	I	Confirmed		
				Platelets	Aortic-valve replacement	No	No	No		I	1	Yes	1	Confirmed		
				Platelets	Coronary-artery bypass grafting	No	Yes	No		I	1	1	1	Confirmed		
				RBC	Acute myloid leukemya—BMT	Yes	No	Yes		I	1	1	I	Confirmed		
				Platelets	Congenital bone marrow failure— HSCT	Yes	Yes	No		1	1	1	1	Confirmed		
				FFP	Motor vehicle accident	ů	No	No		I	1	1	I	Confirmed		
				RBC	Lumbar laminectomy	No	Yes	No		I	I	I	I	Confirmed		
				RBC	Femur-fracture repair	No	No	No		I	I	Yes	I	Confirmed		
				RBC	Motor vehicle accident	No	No	Yes		I	I	I	I	Confirmed		
				RBC	Kidney transplantation	Yes	Yes	Yes		I	I	I	I	Confirmed		
				RBC	Gastrointestinal bleeding	No	Yes	No		1	1	1	1	Confirmed		
				RBC	Aplastic anemia	Yes	Yes	Yes		1	ı	1	ı	Confirmed		
Caglioti 2004 [47]	Case report	U.S.A.	2004	RBC	Above-knee amputation—DM	Yes	Yes	Yes	Serology IgM RT-PCR	Possible	No	No *	Yes	Probable		
Kightlinger 2006 [48]	Case report	U.S.A.	2006	RBC	Kidney transplantation	Yes	Yes	No	Serology IgM	N.P.	Yes	Yes	Yes	Probable	Traceback case 1 RBC	
				FHP	Kidney transplantation, DM, spinal fracture	Yes	Yes	No		1	I	Yes	I	Probable		
																(Continued)

			;	· •					;					1 111		
Fublication	1 ype or publication	Country	I car	biood product	kecipient	<u></u>	Morbuilty	Mortauty	Diagnostic method	Exposure to Vector	Allemanye e: vProlonged hospital stay	xplanation Other infected recipients	Non- vector vias rejected	Flausioury	data	Other non-vector, non-blood transfusion related transmission
Montgomery 2006 [49]	Original article	U.S.A.	2003	RBC	Coronary artery bypass graft surgery	Ŋ	Yes	No	Serology IgM+	N.P.	Yes	No	Yes	Confirmed	Different donors	
				RBC	Myelodysplastic syndrome	Yes	Yes	Yes	PRNT RT-PCR	I	I	No *	Yes	Probable	MP-NAT nonreactive.	
				RBC	Motor vehicle accident	No	Yes	No		I	I	No *	Yes	Probable		
				RBC	Coronary bypass, aortic valve replacement	No	Yes	No		I	No	No *	Yes	Probable		
				RBC	Coronary artery bypass graft surgery	No	No	No		I	I	No *	Yes	Probable	1	
				Not specified	Coronary artery bypass graft surgery	No	Yes	No		I	I	I	Yes	Probable		
Stanley 2009	Case report	U.S.A.	2008	FFP	Heart transplant	Yes	Yes	No	Serology	N.P.	I	Yes	Yes	Probable	Traceback	
[50]				RBC	Atrial fibrillation, anemia	No	No	No	lgG+ PRNT	I	I	Yes	I	Probable	case 1 FFP	
Meny 2011 [51]	Letter to the editor	U.S.A.	2010	Granulocytes (apheresis)	B-lymphoblatic leukemia	Yes	Yes	Yes	Serology IgG/IgM NAT	I	Yes	No	I	I	NAT results after transfusion	
Politis 2011 [52]	Conference abstract	Greece	2010	Not specified (RBC?)	Thalassaemia	No	Yes	I	Serology IgG/IgM	I	I	I	I	Confirmed	Different donors.	
				Not specified (RBC?)	Thalassaemia	No	Yes	I	PCR	1	I	I	I	Confirmed	One aditional inconclusive case	
Kelly 2013 [53]	Case report	U.S.A.	2012	Platelets (apheresis)	Non-Hodgkin's lymphoma—HSCT	Yes	Yes	Yes	Serology IgM PRNT RT-PCR	N.P.	Yes	No *	Yes	Probable	MP-NAT nonreactive.	
Pervanidou 2014 [54]	Original article	Greece	2012	Not specified	Not specified	1	Yes	No	Not specified	N.P.	I	Yes	I	I	Same donor	
				Not specified	Not specified	1	No	No	Not specified	I	I	Yes	I			
Groves 2017 [55]	Original article	U.S.A.	2016	FFP	Aortic valve replacement and coronary bypass surgery	Ň	Yes	Yes	Serology IgG/IgM	N.P.	No	I	Yes	Probable	MP-NAT nonreactive No data available about RBC recipient.	
Hayes 2018 [56]	Conference abstract	U.S.A.	2017	Platelets (apheresis)	Heart transplant	Yes	Yes	Yes	Serology IgG/IgM PCR	I	No	No	Yes	Probable	ID-NAT negative	
BMT: bone 1	narrow tran	splantatio	on, CF	XD: chronic ki	lney disease, DM: d	iabete	s mellitus	, FFP: fre	sh frozen pla	ısma, HSC'	T: hematopo	ietic stem c	ell transp	lantation, M	4P: minipool,]	NAT: nucleic

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* PL not transfused was positive

transplantation – Data not available

Source of threat to transfusion safety	Arboviruses
Transmission through organ—and/or hematopoietic stem cell transplantation	Dengue virus, eastern equine encephalitis virus, heartland virus, Jamestown Canyon virus, tick-borne encephalitis virus, and West Nile virus
Mother-to-child transmission	Colorado tick fever virus, Japanese encephalitis virus, Ross River virus, yellow fever virus, Zika virus, dengue virus, West Nile virus, chikungunya virus, Crimean-Congo hemorrhagic virus, La Crosse virus, O'nyong'nyong virus, Rift Valley fever virus, Sindbis virus, Venezuelan equine encephalitis virus and Western equine encephalitis virus.
Prevalence in blood donors	Japanese encephalitis virus, Ross River virus, St. Louis encephalitis virus, tick-borne encephalitis virus, yellow fever virus, Zika virus, dengue virus, West Nile virus, Barmah Forest Virus, chikungunya virus, Crimean-Congo hemorrhagic virus, eastern equine encephalitis virus, Heartland virus, Jamestown Canyon virus, Mayaro virus, Murray Valley encephalitis virus, O'nyong'nyong virus, Rift Valley fever virus, Huaiyangshan banyangvirus (Severe fever with thrombocytopenia syndrome Virus), Sindbis virus, Tahyna virus, Toscana virus, and Piry vesiculovirus.
Other direct routes of transmission	Blood contact: tick-borne encephalitis virus, dengue virus, chikungunya virus, Crimean-Congo haemorrhagic virus, La Crosse virus, Zika virus

Table 4. Arboviruses that could pose a threat to transfusion safety due to direct non-vector related transmission mechanisms.

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organ transplantation, mother-to-child transmission, blood contact or prevalence in blood donors. The results of this review further confirm identification of arboviruses with a high-risk of future transfusion transmission, as they were reported to be involved in direct non-vector related transmission and prevalent among blood donors. These are: Chikungunya virus, Crimean-Congo haemorrhagic virus, Eastern equine encephalitis virus, Heartland virus and Jamestown Canyong virus.

Based on the results of this review, the reported rapid growth of arboviral cases in recent years can also be confirmed and seems to correspond to the expanding endemicity of arboviruses globally. According to literature, the increased frequency of outbreaks can be explained by multiple environmental and anthropogenic factors, including increased travel and trade, population growth, changing land use patterns, and the urbanization of rural areas [83].

These facts come on top of a growing list of emerging infectious diseases. It is estimated that from 1940 to 2004, 5.3 new viruses were discovered each year, of which at least two-thirds were zoonoses [84]. In consonance with their emerging nature, there is very little knowledge on some of these new arboviruses, such as the SFTS virus, which has already caused multiple deaths in South Korea [85].

To improve transfusion safety, blood banks have taken a number of measures. One common measure is the exclusion of donors presenting symptoms. This is a measure with limited effectiveness given the high rate of asymptomatic infections. Another measure taken is the cancellation of blood donations in affected regions, which is at times difficult to apply, if the epidemic outbreak is extensive and no contingency plans are in place to guarantee the supply of blood and blood components from other geographical regions. So far, the most useful measure is the application of nucleic acid testing (NAT) in donor screening. One example illustrating this success comes from the significant reduction in transmission of West Nile virus in the USA, after the implementation of NAT screening in 2003. However, NAT tests are not infallible: for instance, one case of West Nile virus transmission was reported in a donor with very low viremia who had undergone NAT screening, using a simple sample [56]. Another

Virus	Publication	Type of publication	Threat to transfusion safety-Direct transmission (non-vector related transmission)
Barmah Forest Virus	Faddy 2014 [59]	Short communication	Screening in blood donors: 5.791 samples screened in 2011 by ELISA IgM: 68 positives (1,17%).
Chikungunya virus	Parola 2006 [60]	Original article	Blood contact: A 60-year-old female nurse developed symptom 3 days after contact with a patient with dengue. Interview and examination did not show recent travel abroad, mosquito bite, accidental skin puncture during blood sampling, skin lesion, or eczema. She noted direct contact with patient blood during hemostasis.
	Simmons 2016 [61]	Original article	Screening in blood donors: 3.007 samples screened by NAT: 56 (1.86%) were confirmed positive.
	Villamil-Gómez 2015 [<u>62</u>]	Original article	Vertical transmission: From September 2014 to February 2015, 7 pregnant women with serological and reverse transcription-polymerase chain reaction-positive test for CHIK delivered 8 infants with CHIK.
	Couderc 2012 [63]	Original article	Other: 4 infected corneas from apparently uninfected donors.
Crimean-Congo haemorrhagic virus	Tsergouli 2019[64]	Original article	Blood contact: Between 1953–2016: 158 cases of nosocomial transmission (2 needle-stick injury or abrasion).
	Charlier 2017 [12]	Original article	Vertical transmission: Transmission documented: miscarriages, stillbirths with maternal death and 1 case of documented fatal neonatal infection.
	Barabás 2018 [<u>65</u>]	Conference abstract	Screening in blood donors: Screening of 1885 blood donors with an IFA in-house method, 10 were positive (0.53%).
Eastern equine encephalitis virus	Leiby 2014 [66]	Conference abstract	Screening in blood donors: 1.314 samples have been screened by PRNT: 2 (0.15%) were confirmed positive.
	Pouch 2019 [<u>67</u>]	Original article	Solid organ transplantation: In 2017, 3 solid organ (heart, liver and lung) transplant recipients from a common donor developed encephalitis by Eastern equine encephalitis virus (EEEV).
Heartland virus	Hevey 2019 [<u>68</u>]	Case report	Solid organ transplantation: Heartland virus infection in a heart transplant recipient from the Heartland (U.S.A.).
	Lindsey 2019 [69]	Orginal article	Screening in blood donors: Of the 487 blood donors tested, 12 were positives, and 7 of those were confirmed for Heartland virus neutralizing antibodies by PRNT.
Jamestown Canyong virus	Askar 2019 [70]	Case report	Solid organ transplantation: Jamestown Canyon virus encephalitis in a heart transplant patient.
	Mayo 2001 [71]	Letter to the Editor	Screening in blood donors: Of the 1,086 sera collected from blood donors in 1990, 164 (15%) were positive by IFA.
La Crosse virus	R. Bowen 2013 [72]	Conference abstract	Blood contact: During the 2011 outbreak in China it was discovered that the virus was able to be transmitted person to person through blood contact, indicating the possibility this virus could be transmitted through a tainted blood product.
	Charlier 2017 [12]	Original article	Vertical transmission: Transmission documented: 1 asymptomatic mother-to-child transmission documented serologically in cord serum, after a maternal infection.
Mayaro virus	Romeiro 2020 [73]	Original article	Screening in blood donors: 5608 blood donor samples were tested. IgM and IgG antibodies to MAYV were detected in 36 and 11 samples, respectively.
Murray valley encephalitis virus	Williams 2013 [74]	Original article	Screening in blood donors: 592 samples from blood donors from the Murray Valley region, 6 positives for IgG antibodies (1%; 95% CI: 0.2–1.8%)
O'nyong'nyong virus	Clements 2019 [75]	Original article	Screening in blood donors: 24 of 552 CHIKV IgG-positive samples (4.4%) were tested for their ability to neutralize CHIKV and other related alphaviruses. 23 of the samples showed higher titers against ONNV than CHIKV.
	Charlier 2017 [12]	Original article	Vertical transmission: Transmission uncertain; 2 miscarriages reported.
Rift Valley fever virus	Azhar 2010 [76]	Original article	Screening in blood donors: 2 of 1260 (0.16%) blood donors were RVFV IgG positive.
	Charlier 2017 [12]	Original article	Vertical transmission: Transmission documented; increased risk of miscarriage. Two symptomatic cases of mother-to-child transmission.
SFTS Virus	Zeng 2015 [77]	Original article	Screening in blood donors: 14,764 donor samples were tested for anti-SFTSV IgG antibodies, 86 of which were repeat-reactive and defined as seropositive.
Sindbis virus	Jöst 2011 [<u>78</u>]	Letter to the Editor	Screening in blood donors: 389 investigated blood donor samples were tested for SINV-specific-IgG antibodies, 4 were positive.
	Charlier 2017 [12]	Original article	Vertical transmission: Transmission uncertain; two stillbirths reported.

Table 5. Reference studies on arboviruses that could be a threat to transfusion safety.

(Continued)

Virus	Publication	Type of publication	Threat to transfusion safety–Direct transmission (non-vector related transmission)
Tahyna virus	Sonnleitner 2014 [79]	Original article	Screening in blood donors: IgG antibodies were determined in sera of 1630 blood donors. 10 (0.6%) reacted positive by TAHV IFA, 5 of which (0.3%) were confirmed by neutralization test.
Toscana virus	Brisbarre 2011 [80]	Letter to the Editor	Screening in blood donors: 84 (11.5%) of 729 plasma samples were positive for IgG. 24 (3.3%) were positive for IgM, and 5 (0.7%) were positive for IgG and IgM.
Venezuelan equine encephalitis virus	Charlier 2017 [12]	Original article	Vertical transmission: virus documented in the brains of ten aborted fetuses; developmental brain lesions in infants borne by infected mothers.
Vesiculovirus Piry	Tavares-Neto 1990 [81]	Original article	Screening in blood donors: frequency of neutralizing antibodies 8% (13/162).
Western equine encephalitis virus	Charlier 2017 [12]	Original article	Vertical transmission: three cases with severe encephalitis.

Table 5. (Continued)

IFA: indirect fluorescent antibody, NAT: nucleic acid testing, PRNT: plaque reduction neutralization test, SFTS Virus: Severe fever with thrombocytopenia syndrome virus (Huaiyangshan banyangvirus).

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compromising aspect is the expense: the implementation of NAT testing for West Nile virus in the USA, had an estimated cost of USD 483,000–1,300,000 per quality-adjusted life year (QALY), clearly exceeding the widely accepted threshold of USD 50,000–100,000 per QALY [3,86]. Furthermore, implementing NAT tests requires human and logistical resources that are not available in all settings. As an alternative, techniques for the inactivation of pathogens such as methylene blue and visible light, could also be applied where available, as these methods were proven effective against several arboviruses [87,88]. In general, and despite limitations, the best probable strategy is probably to screen using NAT techniques and apply pathogen inactivation techniques to as many blood components as resources allow.

In addition, blood transfusion centers should develop robust hemovigilance systems with the ability to trace blood products from infected donors, so that these can be destroyed prior to use or, in case these have already been transfused, the physician responsible for clinical management of the recipient, can be informed in a timely fashion.

The risk of transfusion-transmitted arbovirus infections is of particular concern in lowincome countries, usually in endemic areas or areas where extensive epidemic outbreaks occur. Particularly since blood bank systems are usually not based on voluntary donations, but rather on replacement donations. In addition, testing for infectious agents is not optimal, due to technical and resource constraints or a lack of qualified personnel, often relying on rapid diagnostic tests, known to have a poor sensitivity [89,90].

One limitation of this systematic review is the inherent difficulty of confirming the transmission of the arbovirus through the transfusion of blood components. Unequivocally concluding that an infection was transmitted through transfusion would require demonstrating that the infectious agent was found in samples from the donor or in residual blood components from the donation, that the recipient was not infected before receiving the transfusion, and that the virus was not transmitted through the bite of a vector. This is especially complicated in endemic areas, where the population is exposed to vector bites (mainly mosquitoes), with the possible exception of patients admitted to specific units such as: units specialized in transplantation and intensive care units. However, even in these "mosquito free" units, patients could have been infected before hospital admission. Phylogenetic analysis may be relevant to exclude the transfusion hypothesis, but if the donor and recipient reside in the same geographic area, the same genetic sequence may not be enough to confirm this exclusion [91]. Therefore, definitively classifying a case as transfusion transmission is often unfeasible.

Country Australia (n=1) Brazil (n=12) China (n=3) Finland (n=2) Greece (n=4)	· · · · · · · · · · · · · · · · · · ·				Proportion [95% Cl] 0.01 [-0.01, 0.04] 0.16 [0.08, 0.24] 0.04 [0.00, 0.09] 0.03 [-0.01, 0.06] 0.05 [0.00, 0.11]
Pakistan (n=2)					0.03 [-0.01, 0.06]
Singapore (n=4)	·····	•		_	0.05 [0.00, 0.11]
U.S.A. (n=42) Total (n=74)					
Blood product					
Red blood cells (n=35)					0.47 [0.36, 0.59]
Platelets (n=21)					0.28 [0.18, 0.39]
Fresh frozen plasma (n	=11)	•••••			0.15 [0.07, 0.23]
Granulocytes Not specified					
Total (n=74)					0.08 [0.02, 0.14]
Status of immunosupp	ression				
Yes (n=40)					0.54 [0.43, 0.65]
No (n=24)		_	••••		0.32 [0.22, 0.43]
Not specified (n=10) Total (n=74)		•••••			0.14 [0.06, 0.21]
Morbidity					
Yes (n=47)					0.64 [0.53, 0.75]
No (n=20)		······			0.27 [0.17, 0.37]
Not specified (n=7) Total (n=74)					0.09 [0.03, 0.16]
Mortality					
Yes (n=14)					0.19 [0.10, 0.28]
No (n=48)				••••••	0.65 [0.54, 0.76]
Not specified (n=12) Total (n=74)					0.16 [0.08, 0.25]
Plausibility of transfus	ion transr	mission			
Confirmed (n=37)					0.50 [0.39, 0.61]
Probable (n=21)			·····		0.28 [0.18, 0.39]
Possible (n=5)					0.07 [0.01, 0.13]
Not specified (n=11) Total (n=74)					0.15 [0.07, 0.23]
	0.00		1	1	



https://doi.org/10.1371/journal.pntd.0010843.g005

Similarly, a significant number of transfusion-transmitted arbovirus infections are likely to go undetected. If, despite the transmission, the recipient does not present symptoms, or if the transmission occurs in a geographical region during an epidemic outbreak, where transmission through the vector cannot be ruled out, transfusion transmission may go unnoticed.

Another limitation of the study is that conclusions of authors of the reviewed records on the plausibility of transfusion transmission and mortality as a consequence of the arboviral infection, were copied with limited information available to confirm these independently. In addition, each author may have applied different criteria in establishing the plausibility of arboviral transmission through the transfusion of blood or blood components and mortality as a possible outcome.

In conclusion, this review updated, characterized and classified cases of arbovirus transmission through transfusion. In addition, a significant number of arboviruses were identified with a potential threat to future transfusion safety. In recent years a global increase was seen in the number of epidemic outbreaks of various arboviruses. In the coming years, it is expected that this number will continue to increase. For this reason, it is important that each system or organization responsible for transfusion safety has adequate surveillance systems, contingency plans, and protocols in place to guarantee transfusion safety. The different regulatory agencies, public health bodies, medical communities, and the medical industry should collaborate closely and coordinate efforts to provide a rapid and effective response to any threat to transfusion safety.

Supporting information

S1 Table. PICOT research question. (DOCX)

S1 PRISMA Checklist. (DOCX)

S1 Text. Search strategy. (DOCX)

S1 Flow Chart. Flow chart on selection of case studies of transfusion-transmitted arboviruses.

(DOCX)

S2 Flow Chart. Flow chart for the process of selecting records describing other mechanisms of direct transmission. (DOCX)

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References

- Perkins HA, Busch MP. Transfusion-associated infections: 50 years of relentless challenges and remarkable progress. Transfusion. 2010; 50(10):2080–2099. <u>https://doi.org/10.1111/j.1537-2995.2010</u>. 02851.x PMID: 20738828
- Stramer SL, Dodd RY. AABB Transfusion-Transmitted Diseases Emerging Infectious Diseases Subgroup. Transfusion-transmitted emerging infectious diseases: 30 years of challenges and progress. Transfusion. 2013; 53(10 Pt 2):2375–2383. https://doi.org/10.1111/trf.12371 PMID: 23926897
- Petersen LR, Busch MP. Transfusion-transmitted arboviruses. Vox Sang. 2010; 98(4):495–503. https://doi.org/10.1111/j.1423-0410.2009.01286.x PMID: 19951309
- Go YY, Balasuriya UB, Lee CK. Zoonotic encephalitides caused by arboviruses: transmission and epidemiology of alphaviruses and flaviviruses. Clin Exp Vaccine Res. 2014; 3(1):58–77. <u>https://doi.org/10.</u> 7774/cevr.2014.3.1.58 PMID: 24427764
- Giménez-Richarte Á, de Salazar MO, Arbona C, Giménez-Richarte MP, Collado M, Fernández PL, et al. Prevalence of Chikungunya, Dengue and Zika viruses in blood donors: a systematic literature review and meta-analysis Blood Transfus. 2022; 20(4):267–280. <u>https://doi.org/10.2450/2021.0106-21</u> PMID: 34694219
- Stramer SL, Hollinger FB, Katz LM, Kleinman S, Metzel PS, Gregory KR, et al. Emerging infectious disease agents and their potential threat to transfusion safety. Transfusion. 2009; 49(Suppl 2):1S–29S. https://doi.org/10.1111/j.1537-2995.2009.02279.x PMID: 19686562
- American Association of Blood Banks. Fact Sheets Created or Updated Post Publication Of The TRANSFUSION August 2009 Supplement. 2020 Available from http://www.aabb.org/tm/eid/Pages/eidpostpub.aspx.
- European Centro for Disease Prevention and Control. Assessing the risk of communicable diseases transmissible through substances of human origin. 2011. Available from https://www.ecdc.europa.eu/ sites/default/files/media/en/publications/Publications/diseases-communicable-by-substances-ofhuman-origin-SoHo.pdf.
- World Health Organization. Asia Pacific Strategy for Emerging Diseases and Public Health Emergencies (APSED III). 2017. Available from https://iris.wpro.who.int/bitstream/handle/10665.1/13654/ 9789290618171-eng.pdf.
- 10. World Health Organization. Blood donor selection: guidelines on assessing donor suitability for blood donation. 2012. Available from https://www.who.int/bloodsafety/publications/guide_selection_assessing_suitability.pdf.
- 11. Higgins JPT, Green S. Cochrane Handbook for Systematic Reviews of Interventions Version 5.1.0 [updated March 2011]. The Cochrane Collaboration, 2011. Available from www.cochrane-handbook. org.
- Centers for Disease Control and Prevention (CDC). Transmission of Colorado tick fever virus by blood transfusion–Montana. MMWR Morb Mortal Wkly Rep. 1975; 24(50):422–427.
- Charlier C, Beaudoin MC, Couderc T, Lortholary O, Lecuit M. Arboviruses and pregnancy: maternal, fetal, and neonatal effects. Lancet Child Adolesc Health. 2017; 1(2):134–146. https://doi.org/10.1016/ S2352-4642(17)30021-4 PMID: 30169203
- Cheng VCC, Sridhar S, Wong SC, Wong SCY, Chan JFW, Yip CCY et al. Japanese Encephalitis Virus Transmitted Via Blood Transfusion, Hong Kong, China. Emerg Infect Dis. 2018; 24(1):49–57. https:// doi.org/10.3201/eid2401.171297 PMID: 29043965
- Sayama Y, Suksakhone C, Keokhamphoui C, Kongphaly D, Mazda T, Hoshi Y, et al. Prevalence of mosquito-borne viral infection (dengue fever, Japanese encephalitis, and Chikungunya fever) among blood donors in Lao PDR. Vox Sang. 2013; 105(SUPPL.2):98–99
- Hoad VC, Speers DJ, Keller AJ, Dowse GK, Seed CR, Lindsay MD, et al. First reported case of transfusion-transmitted Ross River virus infection. Med J Aust. 2015; 202(5):267–270. <u>https://doi.org/10.5694/</u> mja14.01522 PMID: 25758699

- Faddy H, Dunford M, Seed C, Olds A, Harley D, Dean M, et al. An evidence-based evaluation of the risk posed by ross river virus to the safety of the australian blood supply. Vox Sang. 2012; 103(SUPPL. 1):184–185.
- Venkat H, Adams L, Sunenshine R, Krow-Lucal E, Levy C, Kafenbaum T, et al. St. Louis encephalitis virus possibly transmitted through blood transfusion-Arizona, 2015. Transfusion. 2017; 57(12):2987– 2994. https://doi.org/10.1111/trf.14314 PMID: 28905395
- Spinsanti L, Re V, Aguilar J, Contigiani M. An indirect immunofluorescence assay to detect antibodies against St. Louis Encephalitis virus. Rev Inst Med Trop Sao Paulo. 2001; 43(6):339–340. https://doi. org/10.1590/s0036-46652001000600008 PMID: 11781605
- Taylor L, Stevens T, Destrampe EM, Brown JA, McGavic J, Gould CV, et al. Powassan virus infection likely acquired through blood transfusion presenting as encephalitis in a kidney transplant recipient [published online ahead of print, 2020 Jun 15]. Clin Infect Dis. 2020;ciaa738. https://doi.org/10.1093/ cid/ciaa738 PMID: 32539111
- Wahlberg P, Saikku P, Brummer-Korvenkontio M. Tick-borne viral encephalitis in Finland. The clinical features of Kumlinge disease during 1959–1987. J Intern Med. 1989; 225(3):173–177. <u>https://doi.org/ 10.1111/j.1365-2796.1989.tb00059.x PMID: 2703799</u>
- Laboratory safety for arboviruses and certain other viruses of vertebrates. The Subcommittee on Arbovirus Laboratory Safety of the American Committee on Arthropod-Borne Viruses. Am J Trop Med Hyg. 1980; 29(6):1359–1381. https://doi.org/10.4269/ajtmh.1980.29.1359
- Lipowski D, Popiel M, Perlejewski K, Nakamura S, Bukowska-Osko I, Rzadkiewicz E, et al. A Cluster of Fatal Tick-borne Encephalitis Virus Infection in Organ Transplant Setting. J Infect Dis. 2017; 215 (6):896–901. https://doi.org/10.1093/infdis/jix040 PMID: 28453842
- Marvik Å, Tveten Y, Pedersen AB, Stiasny K, Andreassen ÅK, Grude N. Low prevalence of tick-borne encephalitis virus antibodies in Norwegian blood donors [published online ahead of print, 2020 Sep 14]. Infect Dis (Lond). 2020;1–8. https://doi.org/10.1080/23744235.2020.1819561 PMID: 32924695
- Centers for Disease Control and Prevention (CDC). Transfusion-Related Transmission of Yellow Fever Vaccine Virus—California, 2009. MMWR Morb Mortal Wkly Rep. 2010; 59(2):34–37. PMID: 20094025
- 26. Ergünay K, Saygan MB, Aydoğan S, Litzba N, Niedrig M, Pınar A, et al. Orta/Kuzey Anadolu Bölgesi kan donörlerinde Dengue virusu ve sarı humma virusu seropozitifliğinin araştırılması [Investigation of dengue virus and yellow fever virus seropositivities in blood donors from Central/Northern Anatolia, Turkey]. Mikrobiyol Bul. 2010; 44(3):415–424.
- Barjas-Castro ML, Angerami RN, Cunha MS, Suzuki A, Nogueira JS, Rocco IM, et al. Probable transfusion-transmitted Zika virus in Brazil. Transfusion. 2016; 56(7):1684–1688. <u>https://doi.org/10.1111/trf.</u> 13681 PMID: 27329551
- Motta IJ, Spencer BR, Cordeiro da Silva SG, Arruda MB, Dobbin JA, Gonzaga YB, et al. Evidence for Transmission of Zika Virus by Platelet Transfusion. N Engl J Med. 2016; 375(11):1101–1103. <u>https:// doi.org/10.1056/NEJMc1607262</u> PMID: 27532622
- 29. Hills SL, Morrison A, Stuck S, Sandhu K, Mason KL, Stanek D, et al. Case Series of Laboratory-Associated Zika Virus Disease, United States, 2016–2019. Emerg Infect Dis. 2021; 27(5):1296–1300. https://doi.org/10.3201/eid2705.203602 PMID: 33900178
- Beau F, Lastère S, Mallet HP, Mauguin S, Broult J, Laperche S. Impact on blood safety of the last arboviruses outbreaks in French Polynesia (2012–18). Transfus Clin Biol 2020; 27:4–9. <u>https://doi.org/10.</u> 1016/j.tracli.2019.12.001 PMID: 31889619
- Marbán-Castro E, Goncé A, Fumadó V, Romero-Acevedo L, Bardají A. Zika virus infection in pregnant women and their children: A review. Eur J Obstet Gynecol Reprod Biol. 2021; 265:162–168. https://doi. org/10.1016/j.ejogrb.2021.07.012 PMID: 34508989
- Chuang V, Wong TY, Leung YH, Ma E, Law YL, Tsang O, et al. Review of dengue fever cases in Hong Kong during 1998 to 2005. Hong Kong Med J. 2008; 14(3):170–177. PMID: 18525084
- Tambyah PA, Koay ES, Poon ML, Lin RV, Ong BK. Transfusion-Transmitted Dengue Infection Study Group. Dengue hemorrhagic fever transmitted by blood transfusion. N Engl J Med. 2008; 359 (14):1526–1527. https://doi.org/10.1056/NEJMc0708673 PMID: 18832256
- Oh HB, Muthu V, Daruwalla ZJ, Lee SY, Koay ES, Tambyah PA. Bitten by a bug or a bag? Transfusiontransmitted dengue: a rare complication in the bleeding surgical patient. Transfusion. 2015; 55 (7):1655–1661. https://doi.org/10.1111/trf.13054 PMID: 25728040
- Levi JE, Nishiya A, Félix AC, Salles NA, Sampaio LR, Hangai F, et al. Real-time symptomatic case of transfusion-transmitted dengue. Transfusion. 2015; 55(5):961–964. <u>https://doi.org/10.1111/trf.12944</u> PMID: 25605570

- Sabino EC, Loureiro P, Lopes ME, Capuani L, McClure C, Chowdhury D, et al. Transfusion-Transmitted Dengue and Associated Clinical Symptoms During the 2012 Epidemic in Brazil. J Infect Dis. 2016; 213 (5):694–702. https://doi.org/10.1093/infdis/jiv326 PMID: 26908780
- 37. Alves M, Infante V, Zenatti C, Vieira G, Silva I, Kayano S, et al. Blood-borne dengue virus: A case report of severe dengue in a hematopoietic stem cell recipient. Open Forum Infect. Dis. 2016; 3(Supp1).
- Karim F, Nasir N, Moiz B. Transfusion transmitted dengue: One donor infects two patients. Transfus Apher Sci. 2017; 56(2):151–153. https://doi.org/10.1016/j.transci.2016.10.013 PMID: 27825640
- 39. Matos D, Tomashek KM, Perez-Padilla J, Muñoz-Jordán J, Hunsperger E, Horiuchi K, et al. Probable and possible transfusion-transmitted dengue associated with NS1 antigen-negative but RNA confirmed-positive red blood cells. Transfusion. 2016; 56(1):215–222. <u>https://doi.org/10.1111/trf.13288</u> PMID: 26469514
- Santos FLS, Slavov SN, Bezerra RS, Santos EV, Silva-Pinto AC, Morais ALL, et al. Vaso-occlusive crisis in a sickle cell patient after transfusion-transmitted dengue infection. Transfusion. 2020; 60 (9):2139–2143. https://doi.org/10.1111/trf.15968 PMID: 32735379
- De Wazières B, Gil H, Vuitton DA, Dupond JL. Nosocomial transmission of dengue from a needlestick injury. Lancet. 1998; 351(9101):498. https://doi.org/10.1016/S0140-6736(05)78686-4 PMID: 9482448
- Yadav A, Rastogi N, Upasana K, Arora S, Thakkar D, Yadav SP. Dengue virus transmission from donor to recipient during haploidentical stem cell transplantation. IDCases. 2021; 25:e01220. <u>https://doi.org/ 10.1016/j.idcr.2021.e01220</u> PMID: 34295644
- **43.** Ashshi AM. The prevalence of dengue virus serotypes in asymptomatic blood donors reveals the emergence of serotype 4 in Saudi Arabia. Virol J 2017; 14:107. <u>https://doi.org/10.1186/s12985-017-0768-7</u> PMID: 28599678
- Shaji Mathew J, Menon VP, Menon VP, Mallick S, Sivasankara Pillai Thankamony Amma B, et al. Dengue virus transmission from live donor liver graft. Am J Transplant. 2019; 19(6):1838–1846. <u>https://doi.org/10.1111/ajt.15270 PMID: 30672135</u>
- 45. Evans-Gilbert T. Vertically transmitted Chikungunya, Zika and dengue virus infections: The pathogenesis from mother to fetus and the implications of co-infections and vaccine development. Int J Pediatr Adolesc Med. 2020; 7(3):107–111. https://doi.org/10.1016/j.ijpam.2019.05.004 PMID: 33094137
- 46. Pealer LN, Marfin AA, Petersen LR, Lanciotti RS, Page PL, Stramer SL, et al. Transmission of West Nile virus through blood transfusion in the United States in 2002. N Engl J Med. 2003; 349(13):1236– 1245. https://doi.org/10.1056/NEJMoa030969 PMID: 14500806
- Centers for Disease Control and Prevention (CDC). Transfusion-associated transmission of West Nile virus—Arizona, 2004. MMWR Morb Mortal Wkly Rep. 2004; 53(36):842–844. PMID: 15371966
- Centers for Disease Control and Prevention (CDC). West Nile virus transmission through blood transfusion—South Dakota, 2006. MMWR Morb Mortal Wkly Rep. 2007; 56(4):76–79. PMID: 17268405
- Montgomery SP, Brown JA, Kuehnert M, Smith TL, Crall N, Lanciotti RS, et al. Transfusion-associated transmission of West Nile virus, United States 2003 through 2005. Transfusion. 2006; 46(12):2038– 2046. https://doi.org/10.1111/j.1537-2995.2006.01030.x PMID: 17176314
- Centers for Disease Control and Prevention (CDC). West Nile virus transmission via Organ Transplantation and Blood Transfusion—Louisiana, 2008. MMWR Morb Mortal Wkly Rep. 2009; 58(45):1263– 1267. PMID: 19940831
- Meny GM, Santos-Zabala L, Szallasi A, Stramer SL. West Nile virus infection transmitted by granulocyte transfusion. Blood. 2011; 117(21):5778–5779. https://doi.org/10.1182/blood-2011-02-335901 PMID: 21617013
- Politis C, Pappa A, Hassapopoulou H, Pantelidou D, Perifanis V, Teli A, et al. Transfusion transmitted west Nile virus (WNV) infection in thalassaemic patients in Greece. Vox Sanguinis. 2011; 101 (Supp.1):236.
- Centers for Disease Control and Prevention (CDC). Fatal West Nile Virus Infection After Probable Transfusion-Associated Transmission—Colorado, 2012. MMWR Morb Mortal Wkly Rep. 2013; 62 (31):622–624. PMID: 23925171
- Pervanidou D, Detsis M, Danis K, Mellou K, Papanikolaou E, Terzaki I, et al. West Nile virus outbreak in humans, Greece, 2012: third consecutive year of local transmission [published correction appears in Euro Surveill. 2014;19(14):pii/20766]. Euro Surveill. 2014; 19(13):20758. <u>https://doi.org/10.2807/1560-7917.es2014.19.13.20758</u> PMID: 24721540
- 55. Groves JA, Shafi H, Nomura JH, Herron RM, Baez D, Dodd RY, et al. A probable case of West Nile virus transfusion transmission. Transfusion. 2017; 57(3pt2):850–856. <u>https://doi.org/10.1111/trf.14018</u> PMID: 28164314

- 56. Hayes C, Stephens L, Fridey JL, Snyder RE, Groves JA, Stramer SL, et al. Probable transfusion transmission of West Nile virus from an apheresis platelet that screened non-reactive by individual donor-nucleic acid testing. Transfusion. 2020; 60(2):424–429. https://doi.org/10.1111/trf.15568 PMID: 31633814
- 57. Winston DJ, Vikram HR, Rabe IB, Dhillon G, Mulligan D, Hong JC, et al. Donor-derived West Nile virus infection in solid organ transplant recipients: report of four additional cases and review of clinical, diagnostic, and therapeutic features. Transplantation. 2014; 97(9):881–889. <u>https://doi.org/10.1097/TP.00000000000024</u> PMID: 24827763
- Busch MP, Caglioti S, Robertson EF, McAuley JD, Tobler LH, Kamel H, et al. Screening the blood supply for West Nile virus RNA by nucleic acid amplification testing. N Engl J Med. 2005; 353(5):460–467. https://doi.org/10.1056/NEJMoa044029 PMID: 16079369
- 59. Faddy H, Dunford M, Seed C, Olds A, Harley D, Dean M, et al. Seroprevalence of Antibodies to Ross River and Barmah Forest Viruses: Possible Implications for Blood Transfusion Safety After Extreme Weather Events. Ecohealth. 2015; 12(2):347–353. https://doi.org/10.1007/s10393-014-1005-0 PMID: 25537629
- 60. Parola P, de Lamballerie X, Jourdan J, Rovery C, Vaillant V, Minodier P, et al. Novel Chikungunya virus variant in travelers returning from Indian Ocean islands. Emerg Infect Dis. 2006; 12(10):1493–1499. https://doi.org/10.3201/eid1210.060610 PMID: 17176562
- Simmons G, Brès V, Lu K, Liss NM, Brambilla DJ, Ryff KR, et al. High Incidence of Chikungunya Virus and Frequency of Viremic Blood Donations during Epidemic, Puerto Rico, USA, 2014. Emerging Infectious Diseases. 2016; 22(7):1221–1228. https://doi.org/10.3201/eid2207.160116 PMID: 27070192
- Villamil-Gómez W, Alba-Silvera L, Menco-Ramos A, Gonzalez-Vergara A, Molinares-Palacios T, Barrios-Corrales M, et al. Congenital Chikungunya Virus Infection in Sincelejo, Colombia: A Case Series. J Trop Pediatr. 2015; 61(5):386–392. https://doi.org/10.1093/tropej/fmv051 PMID: 26246086
- Couderc T, Gangneux N, Chrétien F, Caro V, Le Luong T, Ducloux B, et al. Chikungunya virus infection of corneal grafts. J Infect Dis. 2012; 206(6):851–859. <u>https://doi.org/10.1093/infdis/jis296</u> PMID: 22706183
- Tsergouli K, Karampatakis T, Haidich AB, Metallidis S, Papa A. Nosocomial infections caused by Crimean-Congo haemorrhagic fever virus. J Hosp Infect. 2020; 105(1):43–52. <u>https://doi.org/10.1016/j.jhin.2019.12.001</u> PMID: 31821852
- 65. Barabás E, Magyar N, Palyi B, Nagy A, Henczko J, Takacs M, et al. An emerging virus in a new geographic area–serological evidence of the Crimean-Congo haemorrhagic fever virus among hungarian blood donors. Vox Sanguinis. 2018; 113(Suppl. 1):217.
- Leiby DA, Hapip CA, Kenney JL, Mutebi J, Ahmed A, Stramer SL. Preliminary investigations of donor exposure to eastern equine encephalitis virus and implications for risk. Transfusion. 2014; 54(Suppl. 2):214A.
- Pouch SM, Katugaha SB, Shieh WJ, Annambhotla P, Walker WL, Basavaraju SV, et al. Transmission of Eastern Equine Encephalitis Virus From an Organ Donor to 3 Transplant Recipients. Clin Infect Dis. 2019; 69(3):450–458. https://doi.org/10.1093/cid/ciy923 PMID: 30371754
- Hevey MA, O"Halloran JA, Jagger BW, Staples JE, Lambert AJ, Panella AJ, et al. Heartland virus infection in a heart transplant recipient from the Heartland. Transpl Infect Dis. 2019; 21(4):e13098. https://doi.org/10.1111/tid.13098 PMID: 31009160
- Lindsey NP, Menitove JE, Biggerstaff BJ, Turabelidze G, Parton P, Peck K, et al. Seroprevalence of Heartland Virus Antibodies in Blood Donors, Northwestern Missouri, USA. Emerg Infect Dis. 2019; 25 (2):358–360. https://doi.org/10.3201/eid2502.181288 PMID: 30511916
- Askar W, Menaria P, Thohan V, Brummitt CF. Jamestown Canyon virus encephalitis in a heart transplant patient. Transpl Infect Dis. 2020; 22(1):e13210. https://doi.org/10.1111/tid.13210 PMID: 31713971
- Mayo D, Karabatsos N, Scarano FJ, Brennan T, Buck D, Fiorentino T, et al. Jamestown Canyon virus: seroprevalence in Connecticut. Emerg Infect Dis. 2001; 7(5):911–912. https://doi.org/10.3201/eid0705. 017529 PMID: 11747714
- 72. Keil D, Marschner S, Gosney EJ, Bowen R. Photochemical inactivation of la crosse virus (LCV) using the mirasol PRT system. Transfusion. 2013; 53(Suppl. 2):217A.
- 73. Romeiro MF, Fumagalli MJ, Dos Anjos AB, Figueiredo LTM. Serological evidence of Mayaro virus infection in blood donors from São Carlos, São Paulo, Brazil. Trans R Soc Trop Med Hyg. 2020; 114(9):686–689. https://doi.org/10.1093/trstmh/traa016 PMID: 32333001
- 74. Williams SA, Richards JS, Faddy HM, Leydon J, Moran R, Nicholson S, et al. Low seroprevalence of Murray Valley encephalitis and Kunjin viruses in an opportunistic serosurvey, Victoria 2011. Aust N Z J Public Health. 2013; 37(5):427–433. https://doi.org/10.1111/1753-6405.12113 PMID: 24090325

- 75. Clements TL, Rossi CA, Irish AK, Kibuuka H, Eller LA, Robb ML, et al. Chikungunya and O'nyongnyong Viruses in Uganda: Implications for Diagnostics. Open Forum Infect Dis. 2019; 6(3):ofz001. https://doi.org/10.1093/ofid/ofz001 PMID: 31660384
- Azhar EI, Madani TA, Kao M, Jamjoom G. A seroepidemiological study of rift valley fever virus among slaughter houses' workers in Saudi Arabia. Int. J. Med. Med. Sci. 2010; 2(4):111–115.
- 77. Zeng P, Ma L, Gao Z, Wang J, Liu J, Huang X, et al. A study of seroprevalence and rates of asymptomatic viremia of severe fever with thrombocytopenia syndrome virus among Chinese blood donors. Transfusion. 2015; 55(5):965–971. https://doi.org/10.1111/trf.12953 PMID: 25496479
- Jöst H, Bürck-Kammerer S, Hütter G, Lattwein E, Lederer S, Litzba N, et al. Medical importance of Sindbis virus in south-west Germany. J Clin Virol. 2011; 52(3):278–279. https://doi.org/10.1016/j.jcv.2011. 08.002 PMID: 21893428
- 79. Sonnleitner ST, Lundström J, Baumgartner R, Simeoni J, Schennach H, Zelger R, et al. Investigations on California serogroup orthobunyaviruses in the Tyrols: first description of Tahyna virus in the Alps. Vector Borne Zoonotic Dis. 2014; 14(4):272–277. <u>https://doi.org/10.1089/vbz.2013.1360</u> PMID: 24689784
- Brisbarre N, Attoui H, Gallian P, Di Bonito P, Giorgi C, Cantaloube JF, et al. Seroprevalence of Toscana virus in blood donors, France, 2007. Emerg Infect Dis. 2011; 17(5):941–943. https://doi.org/10.3201/ eid1705.101052 PMID: 21529423
- 81. Tavares-Neto J, Travassos DR, Amélia PA, Morais-Souza H, Vasconcelos P, Travassos da Rosa. Freqüência de anticorpos neutralizantes contra o vesiculovirus Piry, em doadores de sangue de Uberaba, Minas Gerais, Brasil. [Frequency of neutralizing antibodies to the vesiculovirus Piry, in blood donors of Uberaba, Minas Gerais, Brazil]. Rev Inst Med Trop Sao Paulo. 1990; 32(3):211–214.
- Attatippaholkun N, Kosaisawe N, U-Pratya Y, Supraditaporn P, Lorthongpanich C, Pattanapanyasat K, et al. Selective Tropism of Dengue Virus for Human Glycoprotein Ib [published correction appears in Sci Rep. 2018 Apr 12;8(1):6000]. Sci Rep. 2018; 8(1):2688. https://doi.org/10.1038/s41598-018-20914-z PMID: 29426910
- Petersen LR, Marfin AA. Shifting epidemiology of Flaviviridae. J Travel Med 2005; 12(Suppl. 1):S3– S11. https://doi.org/10.2310/7060.2005.12052 PMID: 16225801
- Jones KE, Patel NG, Levy MA, Storeygard A, Balk D, Gittleman JL, et al. Global trends in emerging infectious diseases. Nature. 2008; 451(7181):990–993. https://doi.org/10.1038/nature06536 PMID: 18288193
- Kim KH, Yi J, Kim G, Choi SJ, Jun KI, Kim NH, et al. Severe fever with thrombocytopenia syndrome, South Korea, 2012. Emerg Infect Dis 2013; 19:1892–4. https://doi.org/10.3201/eid1911.130792 PMID: 24206586
- Busch MP, Bloch EM, Kleinman S. Prevention of transfusion-transmitted infections. Blood. 2019; 133 (17):1854–1864. https://doi.org/10.1182/blood-2018-11-833996 PMID: 30808637
- Fryk JJ, Marks DC, Hobson-Peters J, Prow NA, Watterson D, Hall RA, et al. Dengue and Chikungunya viruses in plasma are effectively inactivated after treatment with methylene blue and visible light. Transfusion. 2016; 56:2278–2285. https://doi.org/10.1111/trf.13729 PMID: 27456861
- Papin JF, Floyd RA, Dittmer DP. Methylene blue photoinactivation abolishes West Nile virus infectivity in vivo. Antiviral Res. 2005; 68:84–87. https://doi.org/10.1016/j.antiviral.2005.07.001 PMID: 16118025
- Laperche S. Francophone African Group for Research in Blood Transfusion. Multinational assessment of blood-borne virus testing and transfusion safety on the African continent. Transfusion. 2013; 53 (4):816–826. https://doi.org/10.1111/j.1537-2995.2012.03797.x PMID: 22804482
- **90.** Prugger C, Laperche S, Murphy EL, Bloch EM, Kaidarova Z, Tafflet M, et al. Screening for transfusion transmissible infections using rapid diagnostic tests in Africa: a potential hazard to blood safety? Vox Sang. 2016; 110(2):196–198. https://doi.org/10.1111/vox.12327 PMID: 26646317
- Musso D, Gould E, Lanteri MC. Documentation of transfusion-transmitted arbovirus infections in endemic areas. Transfusion. 2016; 56(12):3143–3144. https://doi.org/10.1111/trf.13861 PMID: 27933623