

Protective effects of alginate–chitosan microspheres loaded with alkaloids from *Coptis chinensis* Franch. and *Evodia rutaecarpa* (Juss.) Benth. (Zuojin Pill) against ethanol-induced acute gastric mucosal injury in rats

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Abstract: Zuojin Pill (ZJP), a traditional Chinese medicine formula, consists of *Coptis chinensis* Franch. and *Evodia rutaecarpa* (Juss.) Benth. in a ratio of 6:1 (w/w) and was first recorded in “Danxi’s experiential therapy” for treating gastrointestinal disorders in the 15th century. However, the poor solubility of alkaloids from ZJP restricted the protective effect in treating gastritis and gastric ulcer. The aim of the study was to investigate the protective mechanism of mucoadhesive microspheres loaded with alkaloids from *C. chinensis* Franch. and *E. rutaecarpa* (Juss.) Benth. on ethanol-induced acute gastric mucosal injury in rats. Surface morphology, particle size, drug loading, encapsulation efficiency, in vitro drug release, mucoadhesiveness, and fluorescent imaging of the microspheres in gastrointestinal tract were studied. The results showed that the mucoadhesive microspheres loaded with alkaloids could sustain the release of drugs beyond 12 hours and had gastric mucoadhesive property with 82.63% retention rate in vitro. The fluorescence tracer indicated high retention of mucoadhesive microspheres within 12 hours in vivo. The mucoadhesive microspheres loaded with alkaloids could reduce the gastric injury by decreasing the mucosal lesion index, increasing the percentage of inhibition and increasing the amount of mucus in the gastric mucosa in an ethanol-induced gastric mucosal injury rat model. Moreover, the mucoadhesive microspheres loaded with alkaloids reduce the inflammatory response by decreasing the levels of tumor necrosis factor- α (TNF- α), interleukin 1 β (IL-1 β), downregulating the mRNA expression of inducible nitric oxide synthase, TNF- α , and IL-1 β in gastric mucosa. All the results indicate that mucoadhesive microspheres loaded with alkaloids could not only increase the residence time of alkaloids in rat stomach, but also exert gastroprotective effects through reducing the inflammatory response on ethanol-induced gastric mucosal damage. Thus, these microspheres could be developed as a potential controlled release drug for treatment of gastric ulcer.

Keywords: alkaloids, mucoadhesive microspheres, alginate, chitosan, anti-inflammatory, gastric mucosal injury

Introduction

Peptic ulcer disease, a common disease among the gastrointestinal disorders, occurs in any part of the digestive tract, especially in the stomach and proximal duodenum. Gastric ulcer is one of the most common peptic ulcers and could develop with the disbalance of some endogenous aggressive factors (gastric acid, pepsin, leukotrienes) and gastroprotective factors, such as mucus, bicarbonate, prostaglandins, mucosal

blood flow, and some growth factors.¹ Gastric ulcer is a multifactorial disease, and various factors such as bacterial infection with *Helicobacter pylori* and overingestion of alcohol and nonsteroidal anti-inflammatory drugs may lead to the gastric injury.² Gastric ulcer could infringe on the submucosa, and therapeutic drugs are required to be able to penetrate the mucous layer and maintain high concentration in the stomach for better treatment. The conventional dosage forms, such as tablet, showed weak adhesion to the mucous surface because of its weight and rapid emptying from the stomach, thus restricting its use in the clinical treatment of gastric diseases. Mucoadhesive drug delivery systems, due to their ability to contact with the absorbing mucosa, could prolong residence time of drugs and improve the drug absorption and bioavailability in partial gastric mucosa, and microspheres which consist of one or more adhesive materials were chosen to treat gastric injury as they are lightweight and have smaller dose variation.³

Chitosan, a naturally derived polysaccharide, has been widely applied in biomedical fields as scaffolds for tissue engineering,⁴ wound dressing,⁵ and drug delivery systems.⁶ Alginate, another naturally occurring polysaccharide derived from the marine brown algae,⁷ shows a wide range of biomedical applications especially in drug delivery, cell immobilization, and tissue regeneration because of its outstanding properties such as biodegradability, biocompatibility, and chelating ability.⁸ Moreover, chitosan can form polyelectrolyte with alginate by electrostatic interactions, which could not only make up for the deficiency of alginate hydrogels but also improve the release performance and reinforce the bioadhesive property.⁹ Sodium alginate and chitosan are both marine-derived polysaccharides and offer many advantages over synthetic polymers since they interact under relatively mild temperature and pH. So, they have been chosen as the mucoadhesive materials as well as sustained delivery carriers.

Gastric ulcer is a complex pluricausal inflammatory disease and is known to develop due to the disruption of mucosal defense.¹⁰ In the inflammatory process, a number of inflammatory cytokines, chemokines, and other mediators, such as nitric oxide (NO), tumor necrosis factor- α (TNF- α), and interleukin 1 β (IL-1 β) are involved in the immune response.^{11,12} NO is a major product which is controlled by nitric oxide synthases (NOSs), such as inducible NOS (iNOS), endothelial NOS, and neuronal NOS.¹³ Most importantly, highly expressed iNOS could lead to organ destruction in some inflammatory and autoimmune diseases.¹⁴ NO could lead to oxidative burst,¹⁵ which would inflict

endothelial damage.¹⁶ Moreover, the enhanced generation of NO by iNOS may contribute to the pathogenesis of various gastroduodenal disorders including peptic ulcer.¹⁷ As many enzymes and free radicals damage the gastric mucosa, these are believed to contribute to ulcer formation. Prostaglandin E₂ (PGE₂) is also another important mediator which is produced from arachidonic acid metabolites catalyzed by cyclooxygenase (COX)-2 in inflammatory responses.¹⁸ Current evidence suggests that nonsteroidal anti-inflammatory drugs exert their impairing effects on ulcer healing through the inhibition of COX isoforms, such as COX-2-selective inhibitors,¹⁹ thus supporting a significant involvement of COX-2 in ulcer repair. Moreover, PGE₂ in ulcer tissue specimens has been reported to play a role in ulcer repair.²⁰

Zuojin Pill (ZJP), a typical traditional Chinese medicine formula, consists of Rhizoma Coptidis (*Coptis chinensis* Franch. was used in the study) and Fructus Evodiae (*Evodia rutaecarpa* (Juss.) Benth. was used in the study) in the ratio of 6:1 (w/w). ZJP was first recorded in "Danxi's Experiential Therapy" for treating gastrointestinal disorders in the 15th century. It is officially listed in the *Chinese Pharmacopoeia* as a prescription employed in patients suffering from gastric ulcer, gastroesophageal reflux disease, gastritis, and pyloric obstruction, among other disorders.²¹ Our previous study showed that alkaloids from *C. chinensis* Franch. and *E. rutaecarpa* (Juss.) Benth. could exert an anti-inflammatory effect by the inhibition of iNOS, COX-2, IL-6, IL-1 β , and TNF- α expression through preventing the nuclear translocation of the NF- κ B p50 and p65 subunits in RAW 264.7 cells,²² which provided evidence to understand the therapeutic effects of ZJP on gastritis, gastric ulcer, and other inflammatory diseases in clinic. Moreover, our results showed that alkaloids from *C. chinensis* Franch. and *E. rutaecarpa* (Juss.) Benth. produced an antidepressant-like effect via the central monoaminergic neurotransmitter system and 5-HT.²³ Although ZJP was used for treating gastrointestinal disorders in the 15th century, the poor solubility of the alkaloids from the two herbs in ZJP restricted the protective effect of ZJP in treating gastritis and gastric ulcer. Furthermore, the protective mechanism of the alkaloids from ZJP on gastritis and gastric ulcer has not been fully clarified in vivo.

In this study, total alkaloids extracted from *C. chinensis* Franch. and *E. rutaecarpa* (Juss.) Benth. were loaded into the mucoadhesive microspheres to increase the residence time of alkaloids in stomach. The protective mechanism of microspheres loaded with alkaloids on ethanol-induced acute gastric mucosal injury was investigated in rats.

Materials and methods

Materials and animals

Sodium alginate, chitosan (with 90% deacetylation degree), and fluorescein isothiocyanate (FITC) were obtained from Sangon Biotech, Co., Ltd. (Shanghai, People's Republic of China). Liquid paraffin was supplied by Tianjin Beifang Tianyi Chemical Reagent Company (Tianjin, People's Republic of China). Span-80 and Tween-80 were purchased from Sigma-Aldrich (St Louis, MO, USA). Micronized CaCO_3 (40 nm) was provided by Beijing Wangyong Technology Co., Ltd. (Beijing, People's Republic of China). The rhizomes of *C. chinensis* Franch. and the dry ripe fruit of *E. rutaecarpa* (Juss.) Benth. were obtained from Baokang Hospital of Tianjin University of Traditional Chinese Medicine (Tianjin, People's Republic of China) and authenticated by Prof Yuan-Lu Cui at Tianjin University of Traditional Chinese Medicine according to the *Chinese Pharmacopoeia* (2010 edition). The standards of berberine chloride, evodiamine, and rutaecarpine with purity of over 98% were all purchased from the National Institute for Control of Pharmaceutical and Biological Products (Beijing, People's Republic of China). Epiberberine (98%) and coptisine (98%) were supplied by Chengdu Must-Technology Co., Ltd. (Chengdu, People's Republic of China). Jatrorrhizine (98%) and palmatine (98%) were provided by Phytomarker Company (Tianjin, People's Republic of China). Omeprazole capsules were obtained from Ruikang pharmaceutical Co., Ltd. (Beijing, People's Republic of China). Methanol, ethanol, and acetonitrile of HPLC (high-performance liquid chromatography) grade were supplied by Tianjin Concord Technology Co., Ltd. (Tianjin, People's Republic of China). Phosphoric acid and triethylamine of analytical grade were obtained from Tianjin Chemical Reagent Company (Tianjin, People's Republic of China). TNF- α and IL-1 β rat ELISA Kit were obtained from eBioscience (San Diego, CA, USA). Prostaglandin E_2 Parameter Assay Kit was obtained from R&D Systems (Minneapolis, MN, USA). BCA protein assay kit was obtained from Thermo (San Jose, CA, USA). Mammalian cell lysis kit and UNIQ-10 column Trizol total RNA extraction kit were bought from Sangon Biotech. Improm-II Reverse Transcription System was purchased from Promega Corporation (Madison, WI, USA). FastStart Universal SYBR Green Master (ROX) kit was purchased from Roche (Mannheim, Germany).

Male Sprague-Dawley (SD) rats (weighing 200–220 g) were obtained from Beijing Vital River Laboratories Animal Technology Co., Ltd. (Beijing, People's Republic of China).

Male ICR mice (weighing 18–22 g) were purchased from Beijing HuaFuKang Bio-Technology Co., Ltd. (Beijing, People's Republic of China). The animals were kept under controlled light (12 hours light/12 hours dark, lights on at 7 am). Ambient temperature and relative humidity were maintained at $24^\circ\text{C}\pm 1^\circ\text{C}$ and $55\%\pm 5\%$, respectively. Animal experiments were performed according to the National Institutes of Health Guide for Care and Use of Laboratory Animals, and the protocol was approved by the Animal Ethics Committee of Tianjin University of Traditional Chinese Medicine (TCM-2009-037-E15).

Total alkaloids extracted from *C. chinensis* and *E. rutaecarpa* and quantitative analysis of seven alkaloids

Preparation and quantitative analysis of seven alkaloids from *C. chinensis* and *E. rutaecarpa* have been published in our previous paper.²²

Preparation of mucoadhesive microspheres loaded with alkaloids

Alginate–chitosan microspheres loaded with alkaloids were prepared by internal gelation of alginate with ion (Ca^{2+}) and coating chitosan outside. Sodium alginate solution (1.5%, w/v) was dissolved in deionized water with magnetic stirring for 6 hours at 45°C . Nano- CaCO_3 (1%, w/v) and alkaloids (0.15 g) were added into 20 mL sodium alginate solution in turn by a homogenizer, the mixture was dropped into 100 mL paraffin oil containing 1% of Span-80 to form uniform-sized droplets. After 15 minutes emulsification, 0.5 mL glacial acetic (dispersed in 5 mL paraffin oil) was added, which led to dissolution of insoluble CaCO_3 to release free Ca^{2+} , and the gelation process was continued for 15 minutes with stirring at 300 rpm. The calcium alginate gel microspheres were collected and washed three times with 1% (volume fraction) Tween 80 solution and two times with deionized water by centrifugation (1,000 rpm, 3 minutes, 25°C), followed by a membrane forming step where the particles were dispersed in a 1.0% (w/v) chitosan solution (pH 5.5) and allowed to react for 10 minutes under magnetic stirring. Finally, the alginate–chitosan microspheres loaded with alkaloids were washed three times with deionized water and lyophilized for 24 hours.

Characterization of the microspheres

Morphological examination

The surface morphology of alginate–chitosan microspheres loaded with alkaloids was observed using an optical

microscope (Nikon Ti-U inverted biological microscope, Tokyo, Japan). Environmental scanning electron microscopy (ZEISS EVOLS-15, Jena, Germany) was used to compare the surface characteristics of dry microspheres.

Particle size measurement

The average size of microspheres and size distribution curves were determined by using a particle-size analyzer. Dry microspheres (4 mg) were suspended in distilled water and ultrasonicated for 10 seconds. A drop of suspension was placed on a clean glass slide, and the microspheres were counted under an optical microscope. A minimum of 200 microspheres was counted per batch.

Differential scanning calorimetry analysis

Differential scanning calorimetry (DSC) curves were recorded using a DSC7 differential scanning calorimeter (Perkin-Elmer, Norwalk, CT, USA). The samples of sodium alginate, chitosan, chitosan and alginate physical mixture, and chitosan-coated alginate microspheres were freeze-dried. Lyophilized samples (5–8 mg) were heated from 30°C to 350°C at a heating rate of 10°C/minute under constant purging of nitrogen at 20 mL/minute. An empty cup was used as reference and thermograms were then obtained.

Fourier transform infrared spectroscopy

The Fourier transform infrared spectroscopy (FTIR) transmission spectra were recorded using a FTS 3000 spectrophotometer (Bio-Rad, Richmond, CA, USA) in the wavenumber range of 4,000–400 cm^{-1} using KBr pellets. The samples were freeze-dried, and a total of 2% (w/w) of samples were mixed with potassium bromide. The mixtures were ground into powders, and disks were compressed for scanning. Each sample was assessed in triplicate at a minimum.

Drug release study of mucoadhesive microspheres loaded with alkaloids in vitro

The release behaviors of seven alkaloids loaded in microspheres were investigated in the simulated gastric fluid (hydrochloric acid–sodium chloride solution containing pepsin at pH 1.2±0.1) as described by Anal and Stevens.⁹ The detailed experiment was carried out according to *Chinese Pharmacopoeia* (2010). Alginate–chitosan microspheres loaded with alkaloids (0.7 g) were placed in 200 mL of dissolution medium with magnetic stirring at 50 rpm and 37°C±0.5°C. At scheduled time intervals, 1 mL of sample solution was withdrawn and filtered through a 0.22 μm pore size Millipore filter, and then an equal volume of fresh dissolution medium was added.

The samples were diluted suitably and analyzed by HPLC to calculate the accumulated release percentage. Determination of alkaloid content in microspheres was performed by HPLC method as mentioned in the previous study.²²

Evaluating the mucoadhesive properties of microspheres loaded with alkaloids in vitro and in vivo

The mucoadhesive properties of the microspheres in vitro were assessed according to the method designed by Akiyama et al²⁴ with slight modifications. Briefly, the mucoadhesive properties were analyzed by determination of the amount of microspheres sticking to mucosa after being rinsed using the rat's stomach in vitro. Three SD rats were fasted overnight with free access to water and dissected after sacrifice. The stomachs were removed and opened along the greater curvature, then rinsed thoroughly with normal saline and fixed on the microscopic slide. A certain amount of microspheres (10 mg) were scattered uniformly on the surface of stomach mucosa. Then, the stomach mucosa with microspheres was put in an airtight container and maintained at a certain relative humidity and room temperature. The mucosa was taken out and fixed on a plastic foam support at an angle of 45° after 20 minutes. The stomach tissue was rinsed with pH 1.2 simulated gastric fluids for 5 minutes at a rate of 6 mL/minute. The microspheres adhering on the gastric mucosa were weighed and the percentage of remaining microspheres calculated.

FITC-labeled chitosan-coated microspheres were prepared to evaluate the mucoadhesive properties of microspheres loaded with alkaloids in vivo. Fifteen ICR mice were divided into five groups randomly and fasted overnight with free access to water. Mice were orally administrated microspheres coated with FITC-labeled chitosan (6 mg/20 g) and observed at 0, 2, 4, 8, and 12 hours. Mice were anesthetized by intraperitoneal injection of chloral hydrate before the test and sacrificed at scheduled time points, then the stomach, duodenum, and jejunum were observed with a Kodak in vivo Imaging System.

Gastroprotective study in rats Ethanol-induced gastric mucosal injury and administration in rats

The SD rats were deprived of food, but had free access to water for 24 hours before ulcer induction. Gastric mucosal lesions were induced with absolute ethanol at a dose of 5 mL/kg by oral administration. Rats were divided into six groups randomly: normal control (normal saline), model group (ethanol), omeprazole (20 mg/kg) + ethanol, microspheres (0.02 mg/kg) + ethanol, microspheres (0.05 mg/kg) + ethanol,

and microspheres (0.18 mg/kg) + ethanol. Microspheres were suspended in normal saline. All animals were pretreated with omeprazole or microspheres orally once a day for 3 days and were fasted overnight with free access to water until experiment. Two hours after the last administration, rats were gavaged with ethanol. The rats were sacrificed and their stomachs were immediately removed after 4 hours.

Determination of mucosal lesion index and percentage inhibition

The rats' stomachs were opened along the greater curvature and rinsed thoroughly with normal saline and then fixed to determine the mucosal lesion index (MLI) and percentage inhibition. An observer who was unfamiliar with the treatment regimen performed this operation. MLI and percentage inhibition were calculated according to Liu et al's²⁵ study and scored as follows: if hemorrhagic erosion <1 mm, three small dot hemorrhages (score 1 each); calculated by multiplying the length (mm) and width (mm) when hemorrhagic erosion >1 mm (score 1 every mm²), the sum of the MLI for each animal was calculated as mean mucosal lesions score. The percentage of inhibition (%) was calculated according to the following formula:²⁶

$$\text{Inhibition \%} = \frac{(\text{MLI}_{\text{model}} - \text{MLI}_{\text{treated}})}{\text{MLI}_{\text{model}}} \times 100\%$$

Measurement of mucus production

The gastric mucosa of each rat was gently scraped using a glass slide and any extra moisture was removed using a filter paper. The collected mucus was weighed by precision electronic balance.²⁷

Histopathology

For pathological examinations, all gastric tissues were fixed in 10% buffered formalin solution followed by dehydration in a graded series of alcohols and xylene, and then immersed in paraffin. The pathological sections were sliced along the longitudinal axis, and 5 μm thick sections were prepared and stained with hematoxylin–eosin by standard procedure for histological examination. The stained sections of each test sample were examined using light microscopy (OLYMPUS, Tokyo, Japan).

Effects of mucoadhesive microspheres loaded with alkaloids on inflammatory response in ethanol-induced acute gastric mucosal injury

Following the macroscopic analysis, production of PGE₂, TNF-α, and IL-1β as well as the mRNA expression of

iNOS, TNF-α, and IL-1β were determined in rat stomach tissues.

Production of PGE₂, TNF-α, and IL-1β in gastric mucosa

Production of PGE₂, TNF-α, and IL-1β in gastric mucosa were detected by enzyme-linked immunosorbent assay. Gastric mucosal homogenates were prepared by treating mixed mucosal tissues with lysis buffer at a ratio of 1:10 (mg/μL), and these were then homogenized on ice using a homogenizer and incubated for 30 minutes on ice. Then, the samples were centrifuged at 12,000 rpm at 4°C for 20 minutes and the supernatants were collected for biochemical analysis. The protein concentrations of the homogenized mucosa solutions were determined using a BCA protein assay kit according to the manufacturer's instruction. Samples were analyzed and the concentrations of PGE₂, TNF-α, and IL-1β were corrected as picograms per milligram protein.

Quantitative real-time RT-PCR for the mRNA expression of iNOS, TNF-α, and IL-1β

Total RNA was isolated using a Sangon UNIQ-10 column Trizol total RNA extraction kit according to the instructions of the manufacturer. RNA was reversely transcribed using an ImProm-II Reverse Transcription System cDNA synthesis kit (Promega Corporation, Madison, WI, USA). The real-time reverse transcription polymerase chain reaction (RT-PCR) oligonucleotide primers used for rat iNOS, TNF-α, and IL-1β are shown in Table 1. The reactions were set up in duplicates in 25 μL total volumes with 1 μL of each primer (0.3 μM final concentrations), 12.5 μL of FastStart Universal SYBR Green Master (ROX) (Roche), and 1 μL of template. The PCR cycle was as follows: 95°C for 10 minutes, 40 cycles of 95°C for 15 seconds, 60°C for 1 minute, and a melt curve analysis was performed at the end of each experiment to verify that a single product per primer pair was amplified. The amplification and analysis were performed using an ABI Prism 7,500 real-time PCR system. Samples were analyzed using the relative C_T method. The fold increase or decrease was determined relative to a blank control after normalizing to a housekeeping gene using 2^{-ΔΔC_T}.^{28,29}

Statistical analysis

All values are represented as mean ± standard deviation excluding results given in gastroprotective study in rats that were expressed with mean ± standard error. Statistically significant differences between groups were determined by one-way analysis of variance followed by Scheffe's multiple

Table 1 Real-time RT-PCR oligonucleotide primers

Gene	Primer	Sequence (5'-3')	PCR product (bp)
GAPDH (NM_017008.4)	Forward	AGACAGCCGCATCTTCTTGT	142
	Reverse	TGATGGCAACAATGTCCACT	
iNOS (NM_012611.3)	Forward	CACCTTGGAGTTCACCCAGT	170
	Reverse	ACCACTCGTACTTGGGATGC	
TNF- α (NM_012675.3)	Forward	TGACCCCCATTACTCTGACC	142
	Reverse	CGTGTGTTTCTGAGCATCGT	
IL-1 β (NM_031512.2)	Forward	AAAAATGCCTCGTGCTGTCT	127
	Reverse	GGGATTTTGTGCTTGCTTGT	

Abbreviations: TNF- α , tumor necrosis factor- α ; IL-1 β , interleukin 1 β ; iNOS, inducible nitric oxide synthase; RT-PCR, reverse transcription polymerase chain reaction.

range test. The criterion for statistical significance was $P < 0.01$ or $P < 0.05$.

Results

Characterization of alginate–chitosan microspheres loaded with alkaloids

As shown in Figure 1, the scanning electron microphotographs showed that the surface of microsphere after freeze-drying became rough and had many wrinkles. The particle size of the microspheres was approximately 114

μm , indicating that microspheres showed a narrow size distribution (Figure 1C), which would improve the adhesion and mobility in vivo.

The DSC characteristics of (a) alginate, (b) chitosan, (c) physical mixture of chitosan and sodium alginate, and (d) chitosan-coated alginate microspheres were recorded and are shown in Figure 2. The DSC curve of alginate showed a sharp exothermic peak at 246.4°C which was attributed to rupture of polymer framework, and stood for the alginate degradation temperature (Figure 2a). The thermogram of chitosan exhibited an

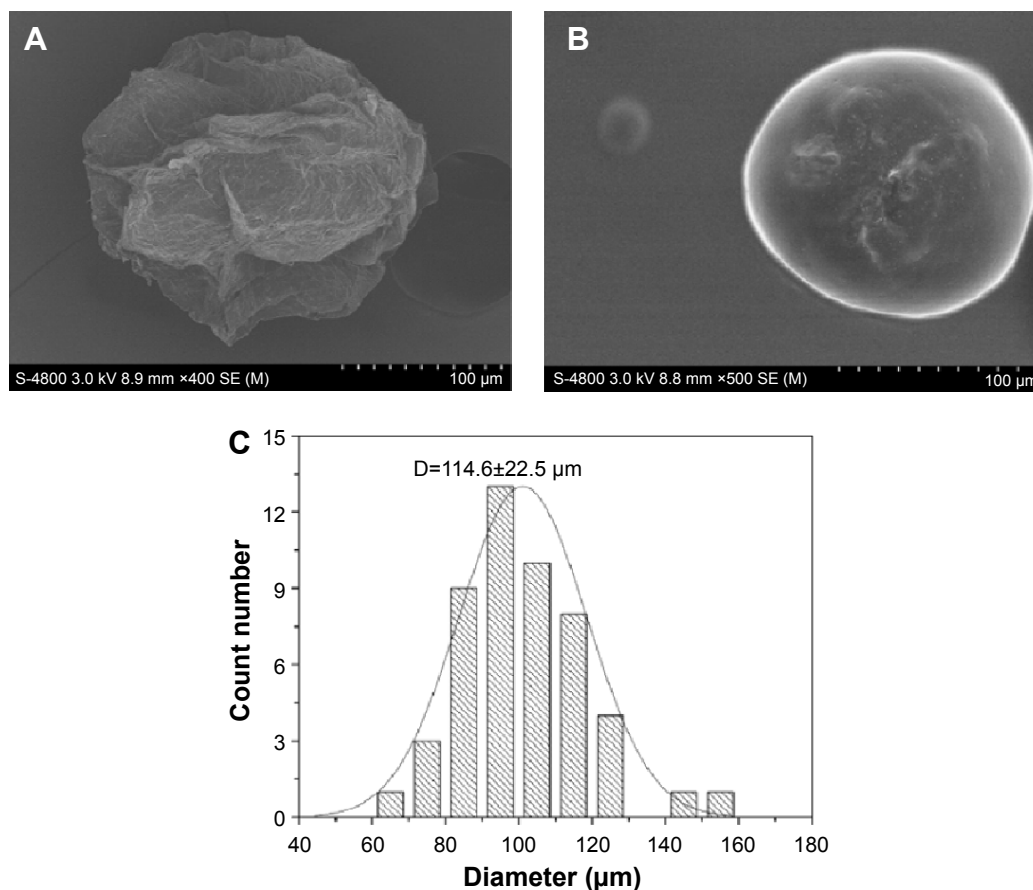


Figure 1 Optical micrographs and SEM micrographs of alginate–chitosan microspheres: (A) freeze-dried microsphere; (B) microsphere loaded with alkaloids; (C) particle size distribution of alkaloid-loaded microspheres.

Abbreviation: SEM, scanning electron microscope.

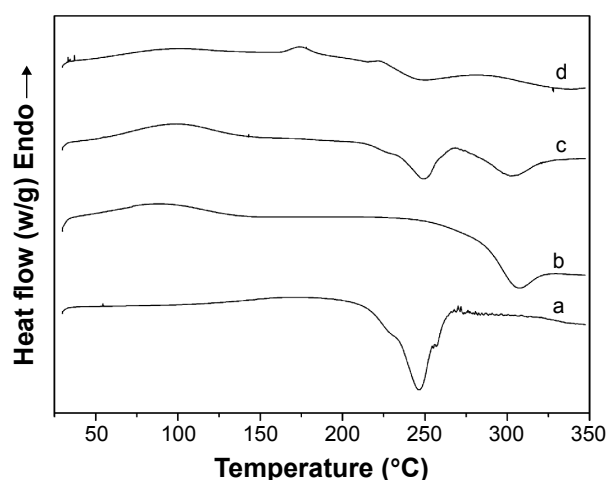


Figure 2 DSC thermograms of (a) sodium alginate, (b) chitosan, (c) physical mixture of chitosan and sodium alginate, and (d) alkaloid-loaded microspheres.

Abbreviation: DSC, differential scanning calorimetry.

exothermic peak at approximately 306.12°C, which indicated that the chitosan was degraded at this temperature (Figure 2b). From the DSC scanning curve of the physical mixture of chitosan and sodium alginate, we found that it was the simple sum of the two polymers' absorption peaks (Figure 2c). However, the characteristic peaks of the two polymers disappeared and new peaks emerged in the thermogram of chitosan-coated alginate microspheres, which indicated the interaction between chitosan and alginate (Figure 2d).

FTIR analysis allows for the observation of vibration modes for the specific groups of compounds analyzed. The spectra of (a) alginate, (b) chitosan, (c) physical mixture of chitosan and sodium alginate, and (d) chitosan-coated alginate microsphere are shown in Figure 3. The IR spectrum of alginate showed a wide band at 3,430 cm^{-1} , which related to the characteristic absorption of hydroxyl groups; the peaks near 1,615 and 1,418 cm^{-1} were attributed to asymmetric and symmetric stretching vibrations of carboxyl groups (COO^-), respectively. A band around 1,029 cm^{-1} (C–O–C stretching) was observed corresponding to its saccharide structure. The peaks at 1,615 cm^{-1} and 1,418 cm^{-1} shifted a little similarly when alginate interacted with chitosan. In the spectrum of chitosan, a broad band at 3,411 cm^{-1} as well as peaks at 1,651, 1,593, and 1,419 cm^{-1} belonged to the characteristic absorption of amino groups ($-\text{NH}_3^+$); the wide absorption peak observed at 1,086 cm^{-1} was the characteristic peak of $-\text{CH}-\text{OH}$. When chitosan interacted with alginate, there was a new absorption peak at 1,609 cm^{-1} . The peaks at 1,593 cm^{-1} and 1,086 cm^{-1} disappeared, suggesting that the amine groups and carboxyl groups reacted, which would prove that the polyelectrolyte complex was formed between alginate and chitosan.

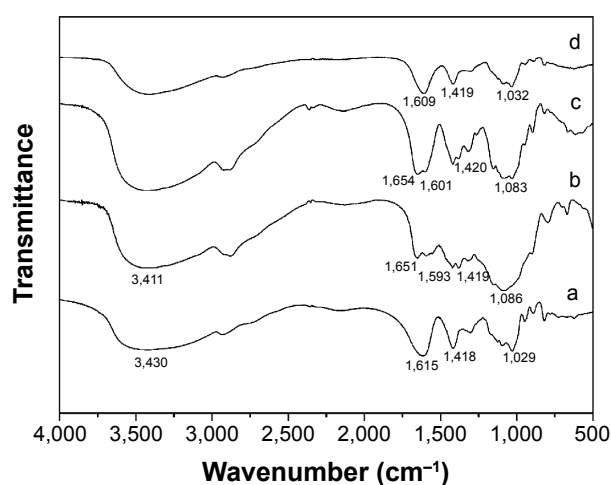


Figure 3 FTIR spectra of (a) sodium alginate, (b) chitosan, (c) physical mixture of chitosan and sodium alginate, and (d) alkaloid-loaded microspheres.

Abbreviation: FTIR, fourier transform infrared spectroscopy.

There are mainly seven alkaloids in *C. chinensis* and *E. rutaecarpa*. To characterize the drug-loading properties of microspheres, the contents of seven alkaloids in microspheres were detected by HPLC. The results are shown in Table 2, and the content of total alkaloids loaded in microspheres was 1.21 mg/g.

Drug release studies in vitro

As shown in Figure 4, the release behavior of seven alkaloids from mucoadhesive microspheres was analyzed by HPLC. Seven alkaloids, except evodiamine, had similar release behavior and maintained sustained release at 12 hours in the simulated gastric fluid (pH 1.2±0.1). The delivery of drugs from the microspheres tended to stabilize after 8 hours and there was a small increase after 12 hours. Approximately 83.55% of rutaecarpine, 77.48% of epiberberine, 70.83% of palmatine, 60.97% of jatrorrhizine, 57.16% of coptisine, 53.13% of berberine, and 16.08% of evodiamine in the microspheres were released respectively after 12 hours.

Table 2 The contents of seven alkaloids in microspheres (n=3)

Number	Seven alkaloids	Content, mean ± SD (mg/kg)	Total alkaloid (mg/kg)
1	Epiberberine	64.7±5.3	1,210.0
2	Jatrorrhizine	37.6±5.1	
3	Coptisine	67.2±9.1	
4	Palmatine	45.9±5.8	
5	Berberine	256.1±31.8	
6	Evodiamine	33.3±10.4	
7	Rutaecarpine	705.3±156.4	

Abbreviation: SD, standard deviation.

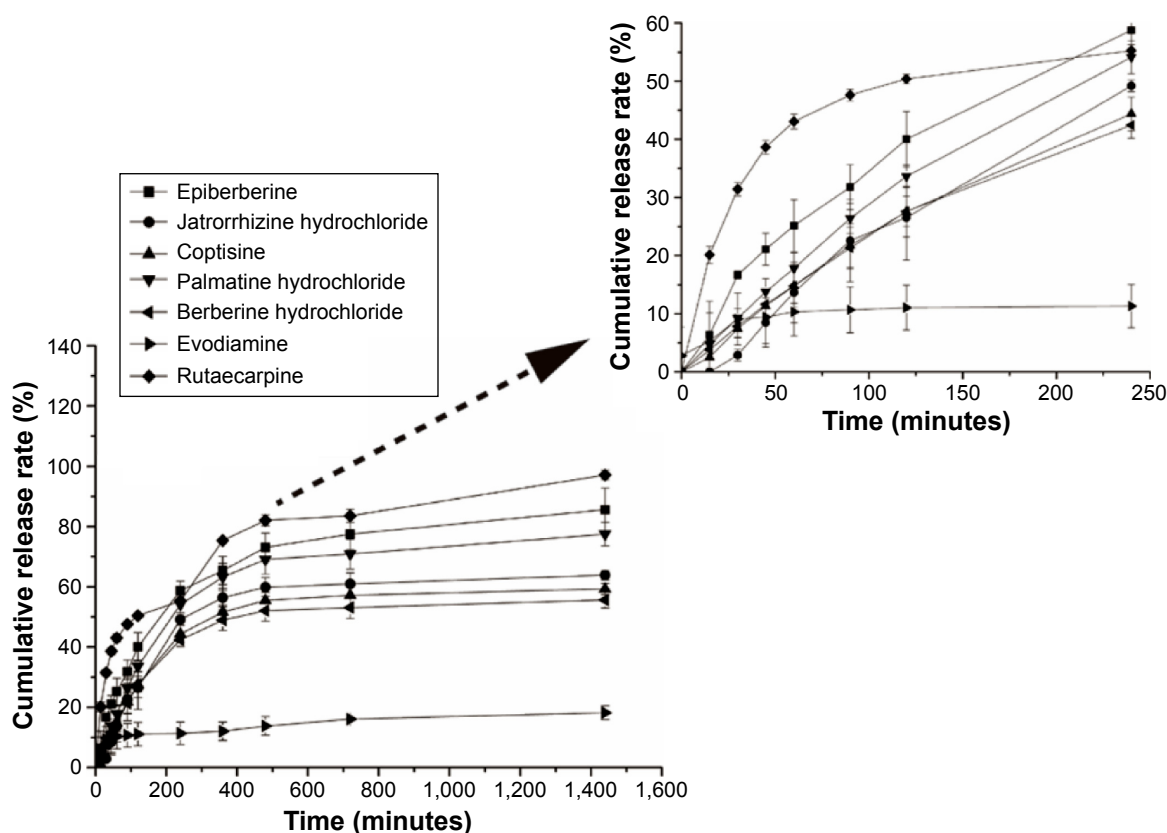


Figure 4 Release curves of seven alkaloids from alginate–chitosan microspheres loaded with total alkaloids extracted from *Coptis chinensis* and *Evodia rutaecarpa*.

Evaluation of mucoadhesive properties in vitro

Mucoadhesive properties of microspheres were evaluated by wash-off test in vitro. As shown in Figure 5A, there was still a mass of microspheres sticking to gastric mucosa after being rinsed with simulated gastric fluid. The percentage of microspheres remaining on the gastric mucosa was $82.63\% \pm 1.42\%$, as determined by the adhesion test in vitro.

Evaluation of mucoadhesive properties in vivo

The FITC-labeled chitosan was used to study the mucoadhesive properties of microspheres in vivo. The labeling efficiency of FITC/chitosan was $38.5 \mu\text{g (FITC)}/\text{mg (chitosan)}$. As shown in Figure 5B, the microspheres could stick on the gastrointestinal tract when mice were orally administrated microspheres coated with FITC-labeled chitosan. At 8 hours, the fluorescence intensities of FITC-labeled microspheres on the surface of gastric mucosa were still relatively high, which indicated that the microspheres possess enhanced mucoadhesive properties. A small amount of microspheres could still be found in the stomach even after 12 hours, which indicated that the microspheres had good adhesive properties to stay on the gastric mucosal surface to protect it from damage.

Gastroprotective studies

MI, percentage inhibition, and mucus production

Since ethanol could easily penetrate into the gastric mucosa,^{30,31} and then influence blood circulation within the mucosa by altering protective factors,^{32,33} ethanol is regarded as the greatest contributor to gastric ulcer formation. The severity of gastric mucosal injury induced by ethanol is shown in Figure 6A. The model group showed macroscopic morphological changes, such as glandular area hyperemia, mucosal edema accompanied by dot and linear hemorrhage necrosis. A pathology score was used to evaluate whether pretreatment with microspheres loaded with alkaloids could protect against ethanol-induced gastric mucosal injury. As shown in Table 3, the model group displayed a significant increase of MI compared to control group ($P < 0.05$); however, the group pretreated orally with omeprazole or mucoadhesive microspheres loaded with alkaloids could decrease the MI ($P < 0.05$) and increase the percentage of inhibition significantly. As shown in Table 3, oral administration of ethanol could decrease the mucus production significantly compared to the normal group ($P < 0.05$), but pretreatment with omeprazole or microspheres loaded with alkaloids could increase the amount of mucus in the gastric mucosa compared to model group.

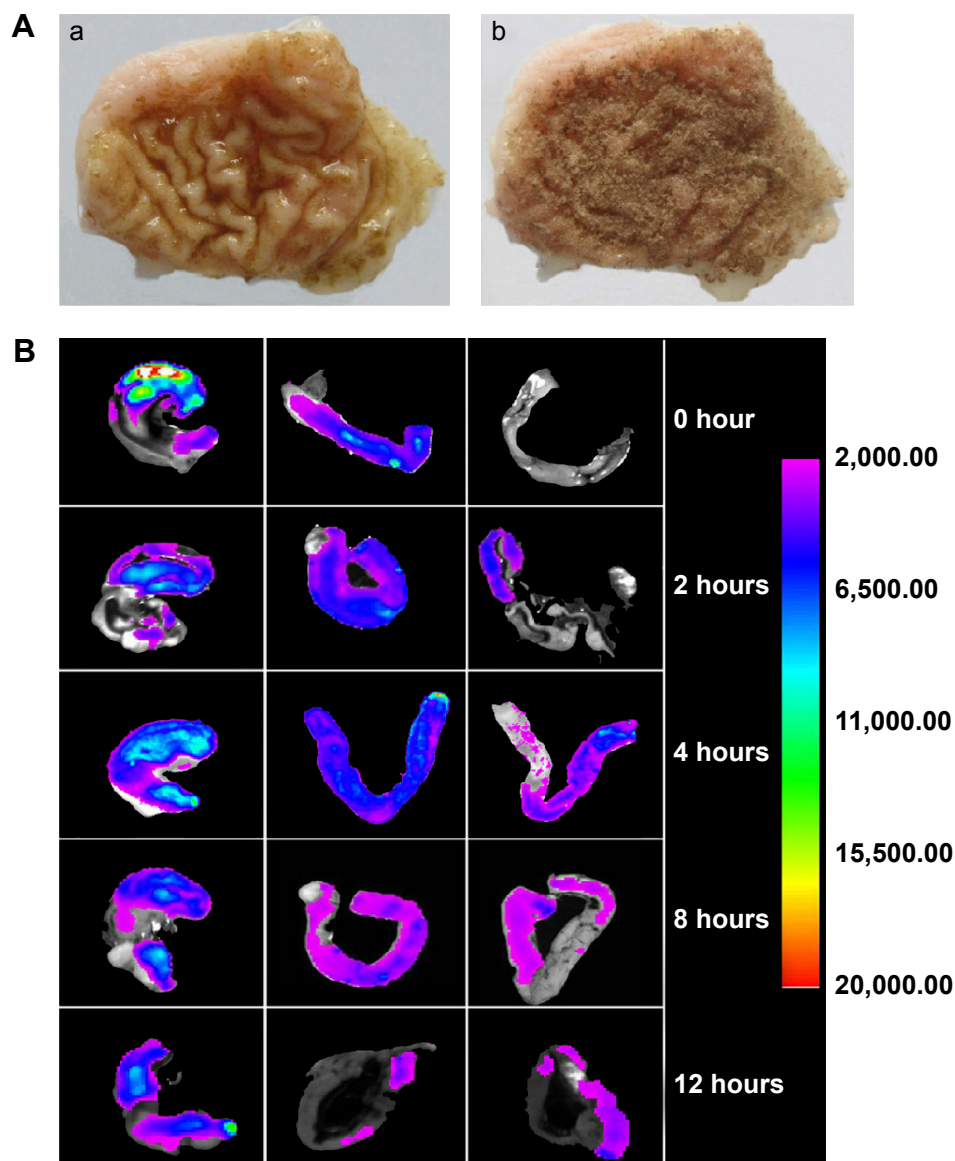


Figure 5 Mucoadhesive properties (A) in vitro and (B) in vivo.

Notes: (A) Microspheres sticking to stomach mucosa (a) after rinsing and (b) before rinsing. (B) Characterizations of mucoadhesive properties of FITC-labeled chitosan-coated alkaloid-loaded microspheres in vivo (Left→Right: stomach, duodenum, and jejunum) varied from 0 to 12 hours.

Abbreviation: FITC, fluorescein isothiocyanate.

Histological results

Histological observations results are shown in Figure 6B. There was no damage on the stomach of normal control group, but the model group treated with absolute ethanol showed serious injury of gastric mucosa, and the superficial gastric epithelium was disrupted and exfoliated. Moreover, extensive hemorrhage, edema, necrosis, and leukocyte infiltration were also observed. In some areas, damage cells could be observed in the submucosa. The gastric mucosa damages were markedly attenuated with only slight injury observed in superficial gastric epithelium on treatment with omeprazole or microspheres loaded with alkaloids. There was little damage in gastric mucosa of animals pretreated

with omeprazole (20 mg/kg) or microspheres loaded with alkaloids (0.05, 0.18 mg/kg).

Effect of mucoadhesive microspheres loaded with alkaloids on production of PGE₂, TNF- α , and IL-1 β in ethanol-induced acute gastric mucosal injury

PGE₂ was an effective vasodilator, which could not only inhibit the gastric acid secretion, but also stimulate mucus and bicarbonate secretion in the stomach to enhance the mucosal defense. As shown in Figure 7, the production of PGE₂ in the gastric mucosa treated with ethanol was downregulated significantly ($P < 0.05$) compared to normal control. Pretreatment with omeprazole or microspheres loaded with alkaloids

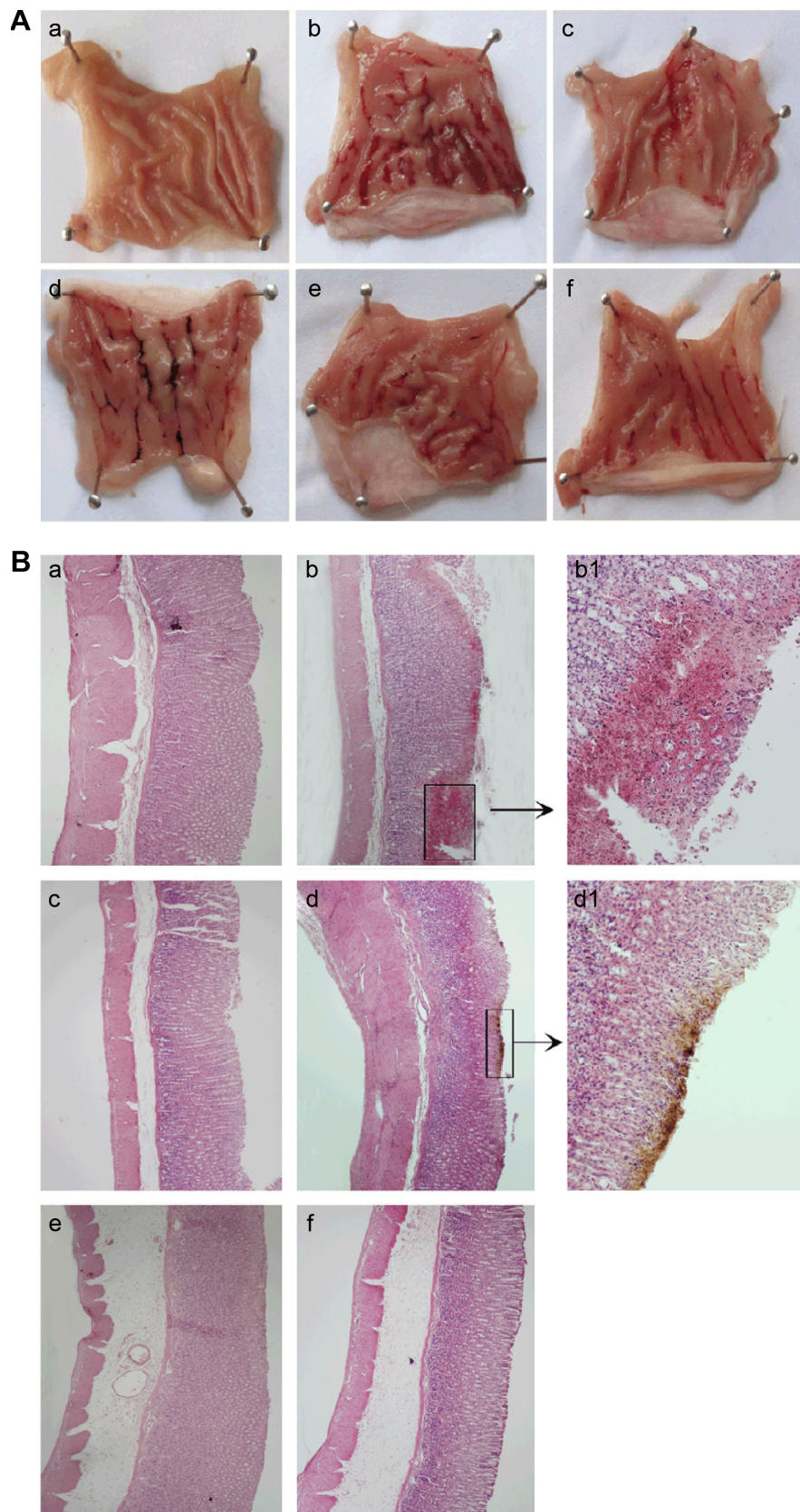


Figure 6 Effects of alkaloid-loaded microspheres on **(A)** gross appearance and **(B)** H&E-stained micrographs of stomach.

Notes: **(A)** (a) Normal control group, (b) model group, (c) omeprazole group (20 mg/kg), microsphere groups (d) at 0.02 mg/kg, (e) at 0.05 mg/kg, and (f) at 0.18 mg/kg. **(B)** H&E-stained micrographs of stomach from (a) normal control (40 \times), (b) model group (b-40 \times , b1-100 \times), (c) omeprazole at 20 mg/kg (40 \times), microsphere groups (d) at 0.02 mg/kg (d-40 \times , d1-100 \times), (e) at 0.05 mg/kg (40 \times), (f) at 0.18 mg/kg (40 \times).

Abbreviation: H&E, hematoxylin and eosin.

Table 3 Macroscopic analysis results of mucosa lesions induced by ethanol

Groups	Pretreatment	Mucosal lesion index (score)	Percentage of inhibition (%)	Mean mucus weight (mg)
1	Normal control	0	–	186.8±1.62
2	Model group	115.99±8.74*	–	140.1±4.11*
3	Omeprazole (20 mg/kg)	24.52±1.90**	78.9	147.7±10.0
4	Microspheres (0.02 mg/kg)	46.47±11.14**	59.9	135.3±8.87
5	Microspheres (0.05 mg/kg)	30.71±2.35**	73.5	152.0±2.77
6	Microspheres (0.18 mg/kg)	38.73±6.07**	66.6	171.6±6.10**

Notes: Data are expressed as mean ± standard error mean. * $P<0.05$ compared to the control group; ** $P<0.05$ compared to the model group.

could increase the PGE_2 levels significantly ($P<0.05$) in the gastric mucosa compared to model group. The administration of ethanol produced plentiful gastric mucosal injuries, especially an increase in inflammatory mediators. In the process of acute-phase inflammation, a variety of mediators such as $\text{TNF-}\alpha$ and $\text{IL-1}\beta$ were involved.^{34–36} As shown in Figure 7, the production of $\text{TNF-}\alpha$ and $\text{IL-1}\beta$ in the model group was increased significantly ($P<0.05$) compared with normal group. In the groups pretreated with omeprazole (20 mg/kg) and microspheres loaded with alkaloids (0.05, 0.18 mg/kg), the production of $\text{TNF-}\alpha$ and $\text{IL-1}\beta$ decreased

significantly ($P<0.05$). The anti-inflammatory property on gastric ulcer was in accordance with the previous study of the ethanol extract from alkaloids on inhibiting the production of inflammatory factors.²²

Effect of mucoadhesive microspheres loaded with alkaloids on mRNA expression of iNOS, $\text{TNF-}\alpha$, and $\text{IL-1}\beta$ in ethanol-induced acute gastric mucosal injury
In the digestive systems, NO generated from iNOS is cytotoxic,³⁷ and the effects of microspheres on mRNA expression of iNOS were investigated in this study. As shown

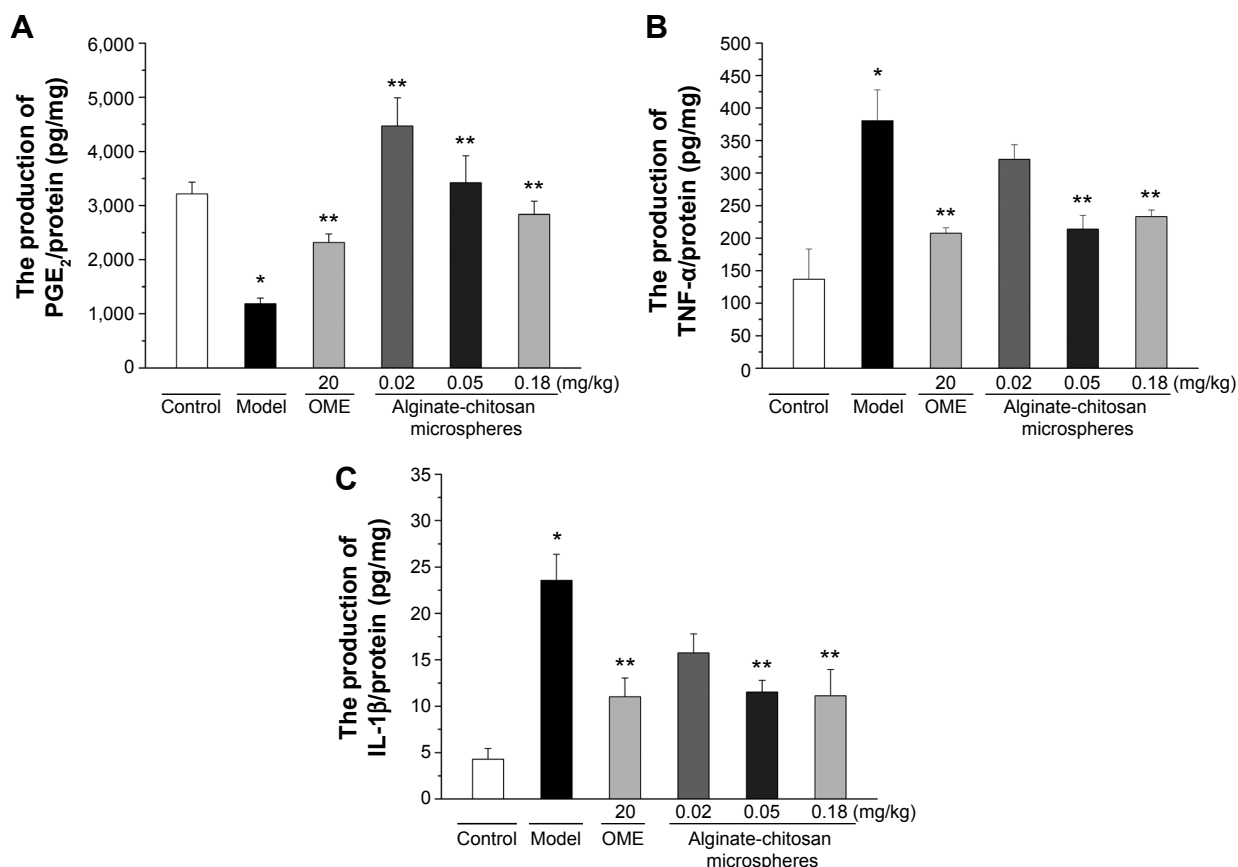


Figure 7 Effects of alkaloid-loaded microspheres on production of PGE_2 , $\text{TNF-}\alpha$, and $\text{IL-1}\beta$ in ethanol-induced gastric mucosa lesions.

Notes: (A) PGE_2 , (B) $\text{TNF-}\alpha$, and (C) $\text{IL-1}\beta$ in gastric mucosal homogenates were measured by ELISA. Data were expressed with mean ± SE from three independent experiments. * $P<0.05$ ($n=4$) compared to the normal control group, ** $P<0.05$ ($n=4$) compared to the model group.

Abbreviations: PGE_2 , Prostaglandin E₂; OME, omeprazole; $\text{TNF-}\alpha$, tumor necrosis factor- α ; $\text{IL-1}\beta$, interleukin 1 β ; ELISA, enzyme-linked immunosorbent assay; SE, standard error.

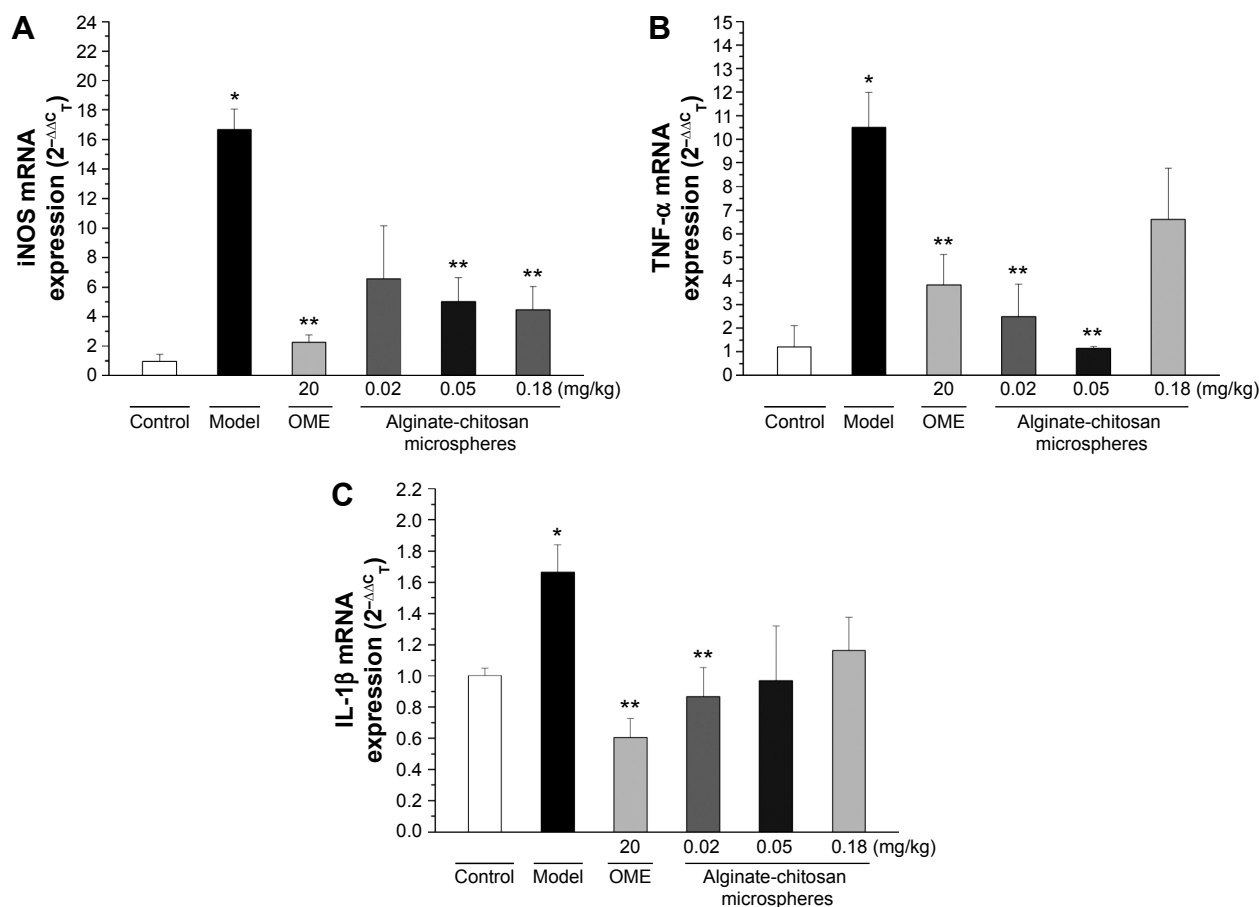


Figure 8 Effects of alkaloid-loaded microspheres on mRNA expression of iNOS, TNF- α , and IL-1 β in ethanol-induced gastric mucosa lesions.

Notes: The mRNA expressions of (A) iNOS, (B) TNF- α , and (C) IL-1 β were measured by real-time RT-PCR. Data were expressed with mean \pm SE from three independent experiments. * $P < 0.05$ ($n = 4$) compared to the normal control group; ** $P < 0.05$ ($n = 4$) compared to the model group.

Abbreviations: OME, omeprazole; TNF- α , tumor necrosis factor- α ; IL-1 β , interleukin 1 β ; ELISA, enzyme-linked immunosorbent assay; SE, standard error; iNOS, inducible nitric oxide synthase; RT-PCR, reverse transcription polymerase chain reaction.

in Figure 8, ethanol ingestion could induce severe gastric mucosal damage compared with the normal group by increasing the mRNA expression of iNOS. The mRNA expression of iNOS was significantly ($P < 0.05$) downregulated by pre-treating with omeprazole (20 mg/kg) or microspheres loaded with alkaloids (0.05, 0.18 mg/kg) compared to model group. There was no significance on concentration of 0.02 mg/kg compared to model group.

Since microspheres loaded with alkaloids were found to inhibit the production of TNF- α and IL-1 β , the effects on mRNA expression of TNF- α and IL-1 β were evaluated by real-time RT-PCR. As shown in Figure 8, the mRNA expression of TNF- α could be inhibited significantly ($P < 0.05$) by pretreating with alkaloid-loaded microspheres (0.02, 0.05 mg/kg) or omeprazole (20 mg/kg) compared to model group. The mRNA expression of IL-1 β was downregulated by alkaloid-loaded microspheres significantly ($P < 0.05$) only at the concentration of 0.02 mg/kg.

Discussion

Various formulations for oral drug delivery in the treatment of gastric ulcers, such as tablets and capsules, have been challenged because of their incomplete eradication of *H. pylori*. One reason for the incomplete eradication of *H. pylori* may be due to the short residence time of drugs in the stomach and because effective treatment concentrations cannot be achieved in the gastric mucous layer or epithelial cell surfaces where *H. pylori* exists.³⁸ Another reason might be the poor stability of drugs in gastric acid or poor permeability of the drugs across the mucus layer.³⁹ In order to increase the residence time and adhesion of alkaloids from *C. chinensis* and *E. rutaecarpa* in stomach, alkaloids were loading into the alginate-chitosan microspheres, and the protective mechanism of microspheres on ethanol-induced acute gastric mucosal injury in rats was investigated.

The surface of microspheres after freeze-drying became rough and had many wrinkles, which would not only improve

the adhesion and mobility *in vivo*, but also accelerate the release of drugs in targeted areas, such as gastric mucosa, which has many wrinkled areas. The DSC characteristics of microspheres showed that the characteristic peaks of the two polymers disappeared and new peaks emerged in the thermogram of chitosan-coated alginate microspheres, which indicated that the interaction between chitosan and alginate. FTIR analysis suggested that the amine groups and carboxyl groups reacted, which would prove that the polyelectrolyte complex was formed between alginate and chitosan. The release study showed that alginate–chitosan microspheres loaded with alkaloids had good controlled-release properties and could be an ideal candidate for drug delivery carrier.

Mucoadhesive properties of microspheres played an important role in the sustained release of drug. Evaluation of mucoadhesive properties showed that there was still a mass of microspheres sticking to stomach mucosa after rinsing with simulated gastric fluid *in vitro*. The results indicated that microspheres had strong adhesion performance as most microspheres still remained on the gastric mucosa. Microspheres could stick to the stomach easily because of the natural biological folds of stomach and the adhesion properties of gastric mucosa. The FITC-labeled chitosan was used to investigate the mucoadhesive properties of microspheres *in vivo*. The results showed that a small amount of microspheres still could be found in the digestive tract even at 12 hours. Studies showed that chitosan could cross-link with mucin and the strong electrostatic interaction between them contributed to the good mucoadhesive property of chitosan-coated alginate microspheres. Hydrophilic functional groups such as hydroxyl groups in chitosan may form hydrogen bonds with sialic acid in mucin molecules. This interaction may be responsible for enhancing permeability of locally delivery drugs and facilitating more effective absorption.^{40,41}

Ethanol could also produce acute hemorrhagic gastric erosion and lead to mucosal edema, cellular exfoliation, and inflammatory cell infiltration, which was the symbol of acute inflammatory reaction.⁴² The group pretreated with omeprazole or alginate–chitosan microspheres loaded with alkaloids could decrease the MI and increase the percentage of inhibition significantly. It was reported that stimulating the mucus production was also one of the mucosal defense mechanisms for protecting the surface epithelial cells and the mucosa against damage.⁴³ In order to further investigate the possible mechanism of the protective effect in gastric mucosa, the effects of microspheres loaded with alkaloids on mucus production were investigated. Oral administration of ethanol

could decrease the mucus production significantly compared to the normal group, but pretreatment with omeprazole or microspheres loaded with alkaloids could increase the amount of mucus in the gastric mucosa compared to the model group. The increase of gastric mucus may be one of the protective mechanisms involved with the antiulcer effects.

Neutrophils inflicted endothelial damage by generating various free radicals including NO that influenced neutrophil oxidative burst.⁴⁴ The enhanced generation of NO by the iNOS might contribute to the pathogenesis of various gastroduodenal disorders including peptic ulcer.¹⁷ An increase in iNOS activity in the gastric mucosa was closely related to the development of gastric mucosal lesions. Ethanol ingestion could induce severe gastric mucosal damage by increasing the expression of the proinflammatory mediators such as iNOS mRNA. The mRNA expression of iNOS was downregulated by microspheres loaded with alkaloids compared to model group. The gastric mucosa has specific defense mechanisms for holding its structural integrity, and its epithelial surface can secrete a barrier consisting of mucin bicarbonate and prostaglandins.⁴⁵ PGE₂ was considered as a key defense factor in the gastrointestinal mucosa for gastric ulcer healing.⁴⁶ Pretreatment with microspheres could increase the PGE₂ level significantly in the gastric mucosa compared to model group. Moreover, PGE₂ also could promote gastric ulcer healing by influencing other inflammatory mediators such as TNF- α , histamine, and IL-1 β in macrophages and platelet-activating factor in mast cells.⁴⁷ The administration of ethanol produced plentiful gastric mucosal injuries, especially an increase in inflammatory mediators. In the process of acute phase inflammation, a variety of mediators such as TNF- α and IL-1 β were involved.^{36,48} The production of TNF- α and IL-1 β were significantly decreased by pretreating with microspheres. The anti-inflammatory property on gastric ulcer was in accordance with the previous study of alkaloids on inhibiting the production of inflammatory factors. The mRNA expression of TNF- α and IL-1 β was inhibited significantly when pretreated with microspheres. Alkaloids could decrease the production of TNF- α and IL-1 β by inhibiting the mRNA expression of TNF- α and IL-1 β .

It is noteworthy that the effects of microspheres loaded with alkaloids on the inflammatory response induced by ethanol were not dose-dependent. One reason may be attributed to the multiple pharmacological effects based on the interactions of its complex active constituents in alkaloids, as we can find that there are seven main alkaloids by HPLC method. Another reason was that the intestinal absorption mechanisms of alkaloids were differently related with

P-glycoprotein, and their intestinal absorption was limited as there was involvement of P-glycoprotein *in vivo*. For example, the study of berberine indicated that it possessed a biphasic effect on regulation of P-glycoprotein ATPase activity (stimulation at low concentration while inhibition at high concentration).⁴⁹ Furthermore, coptisine, jateorrhizine, palmatine, and berberine are all P-glycoprotein substrates, and no obvious inhibitory effect was found on the activity of P-glycoprotein at concentrations of 1–100 μM ,⁵⁰ which may be one of the reasons for their low bioavailability. Finally, the pharmacokinetics of alkaloids could be another influencing factor. Studies have shown that after intravenous administration, plasma concentrations of jatrorrhizine showed a biphasic decline, dose-independent clearance and half-life of terminal elimination phase reduction, and a relatively large distribution volume. The metabolic property of jatrorrhizine was suggested to multiple metabolic pathways.⁵¹ Further research on metabolic processes of alkaloids will be developed *in vivo* in the future.

In conclusion, the mucoadhesive microspheres loaded with alkaloids were prepared with chitosan and alginate and possessed good release performances and gastric mucosa adhesion. The mucoadhesive alkaloid-loaded microspheres could decrease the production of TNF- α and IL-1 β , downregulate the mRNA expression of iNOS, TNF- α , IL-1 β and increase the production of gastroprotective factor PGE₂ in gastric mucosa in an ethanol-induced gastric mucosal injury model in rats. Moreover, the mucoadhesive alkaloid-loaded microspheres could maintain sustained release at 12 hours and play an important role in reducing the gastric injury by decreasing the MI, increasing the percentage of inhibition, and increasing the amount of mucus in the gastric mucosa. Therefore, the results of our studies will provide strong scientific evidence for the mucoadhesive microspheres loaded with total alkaloids of *C. chinensis* and *E. rutaecarpa* to be developed as a new method for the prevention and treatment of gastric ulcer.

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Disclosure

The authors report no conflicts of interest in this work.

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