Characterization of Phosphorus in Animal Manures Collected from Three (Dairy, Swine, and Broiler) Farms in China

Guohua Li^{1,2}, Haigang Li^{1*}, Peter A. Leffelaar², Jianbo Shen¹, Fusuo Zhang¹

1 Center for Resources, Environment and Food Security (CREFS), China Agricultural University, Beijing, China, 2 Plant Production Systems Group, Wageningen University, Wageningen, The Netherlands

Abstract

In order to identify the phosphorus species and concentration in animal manure, we comparatively characterized phosphorus in dairy manure, swine manure, and broiler litter, using a sequential procedure, a simplified two-step procedure (NaHCO₃/NaOH+EDTA), and a solution Phosphorus-31 Nuclear Magnetic Resonance (³¹P-NMR) spectroscopy procedure. In the sequential procedure, deionized water extracted 39, 22, and 32%; NaHCO₃ extracted 48, 26, and 37%; NaOH extracted 8, 9, and 13.8%; and HCI extracted 3, 42.8, and 17% of the total phosphorus in dairy manure, swine manure and broiler litter, respectively. Total phosphorus extracted by the NaHCO₃/NaOH+EDTA procedure was 7.5, 32.4, and 15.8 g P kg⁻¹ for dairy manure, swine manure, and broiler litter, respectively. The solution ³¹P-NMR procedure detected that 9, 34, and 29% of total phosphorus was phytic acid in dairy manure, swine manure, and broiler litter, respectively. These results show that phosphorus forms, availability, and quantities differ between animal manures, which provides valuable information for P characterization of animal manures in China.

Citation: Li G, Li H, Leffelaar PA, Shen J, Zhang F (2014) Characterization of Phosphorus in Animal Manures Collected from Three (Dairy, Swine, and Broiler) Farms in China. PLoS ONE 9(7): e102698. doi:10.1371/journal.pone.0102698

Editor: Dafeng Hui, Tennessee State University, United States of America

Received February 13, 2014; Accepted June 22, 2014; Published July 22, 2014

Copyright: © 2014 Li et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This research was supported by the National Basic Research Program of China (973-2013CB127402), the Innovative Group Grant of the National Natural Science Foundation of China (31121062), the National Natural Science Foundation of China (31201691, 30925024), and Chinese Universities Scientific Fund (2012YJ056). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* Email: haigangli@cau.edu.cn

Introduction

Due to the rapid economic development of China since 1980, the diet of residents has shifted from predominantly plant to more animal products [1]. Livestock and poultry breeding have increased considerably by the stimulation of consumer demand for animal products, and consequently a vast expansion in animal production has occurred. Modes of animal production have shifted gradually from conventional family-based animal rearing to intensive animal feeding operations, with more input of protein and energy into animal feeds. These differences may significantly impact forms, bioavailability, utilization, and cycling of manure nutrients and their potential impact on soil and water quality. For example, if diets of dairy cattle generally exceed animal phosphorus (P) requirements by 25% to 40%, a significant fraction (~80%) of the P consumed is excreted in the manure due to the low utilization efficiency of dietary P [2].

The excess of total P in animal feed is common throughout China, which has led to the production of high-concentration of animal manure P [3]. The capacity of soil to hold manure P mainly depends on the forms and bioavailability of P in manure [4]. If long-term manure application to the arable land is practiced without concerning the P forms and bioavailability, it may lead to soil P accumulation and an acceleration of eutrophication [5]. However, little detailed information about the relative quantities and forms of P in animal manure in China is available to farmers now. Thus, characterizing manure P becomes critical to better understand manure P dynamics and assist farmers in making optimal management of manure P for reducing water pollution.

Traditionally, the composition of animal manure P has been analyzed by three solution-based techniques, including sequential fractionation [6-10], enzymatic hydrolysis [11-15] and solution ³¹P-NMR spectroscopy [16-18]. The sequential procedure of Hedley et al. (1982) [19] originally developed for soil P analysis was adapted for characterizing P in three contrasting manures (broiler litter, dairy manure, swine manure) for the relative dissolution and fraction distribution with deionized water, 0.5 MNaHCO₃, 0.1 M NaOH, and 1.0 M HCl and concentrated sulfuric acid. Generally, H2O and NaHCO3 primarily extract labile P (i.e., plant-available) and NaOH primarily extracts organic P; HCl was assumed to extract inorganic forms of P, such as relatively stable Ca-bound phosphate. For example, Dou et al. (2000) [7] reported that deionized water, NaHCO₃, NaOH and HCl extracted 70, 14, 6, and 5% of the total P in the dairy manure, and 49, 19, 5, and 25% of the total P in the poultry manure sample, respectively. Ajiboye et al. (2004) [20] found the majority of P in fresh dairy, beef, and swine manure was in labile P pools, which can be extracted by NaHCO3 and NaOH. In order to further understand the structural information of manure P, solution ³¹ P-NMR was used to quantify P concentrations and compounds present in a simplified two-step fractionation procedure which involved extraction of readily soluble phosphorus in 0.5 M NaHCO₃ followed by extraction of stable phosphorus in a

solution containing 0.5 *M* NaOH and 50 m*M* EDTA [13,17,21,22]. The solution of NaOH-EDTA has a greater power to extract stable metal-P and organic-P species. Turner (2004) [17] used ³¹P-NMR to identify phytic acid in alkaline extracts of broiler litter and swine manure, while Leinweber et al. (1997a) [6] identified broad classes of P species, including orthophosphate monoesters and diesters, and pyrophosphates in swine and poultry manure. Information on the forms of phosphorus contained in animal manures is a pivotal factor determining its environmental fate [23].

The objective of this study was to identify the P species and concentration in dairy manure, swine manure, and broiler litter collected from three farms in China using three different methods, including sequential fractionation, NaHCO₃/NaOH+EDTA extraction procedure and solution ³¹P-NMR, respectively.

Materials and Methods

Sample collection

Three types of manure were collected from private livestock farms in China which did not involve endangered or protected species: swine manure (the swine was grain fed in housing system) from Beilangzhong farm, located at Zhaoquanying town, Shunyi district, Beijing (40°10' N, 116°35' E), Xuejun Liu was responsible to future permissions; dairy manure (the dairy was pasture fed in sheds) from an individual farm located at Shandong Agricultural University Experimental Station, Taishan district, Taian, Shandong Province (36°9' N, 117°9' E), Zhiguang Liu was responsible to future permissions; and broiler litter (the broiler chicken was grain fed in coop system) from an individual farm located at Xizhengqiao village, Jining, Shandong Province (35°20' N, 116°31' E), Dianting Li was responsible to future permissions. These three farms were all private farm. We had gotten the farmer's permission before collecting animal manure from their farms for P species analysis. The collected manure was stored immediately in a portable refrigerator, which could avoid the potential damage to public health. So there was no need to be approved by an Institutional Animal Care and Use Committee (IACUC) or equivalent animal ethics committee. For each type of manure, multiple samples were collected from animals at the same growth stage from different locations in the facility, then composited and subsampled. Animal feed samples were also collected 24 h before manure sampling to ensure that sampled feed represented the true source of manure P. Manure samples were stored below 4°C and analyzed within a week. Manure samples were dried at 65° C, ground to <2 mm, and stored in sealed containers at room temperature for the sequential extraction procedure. Manure pH was determined on a 10:1 water (mL) to dry matter ratio (g). Total elements in samples were determined by microwave digestion in concentrated HNO₃ and H_2O_2 (EPA Method 3052) with detection by inductively coupled plasma-optical-emission spectrometry (ICP-OES) (Table 1). The concentration of total P in feeds was analyzed after the samples were digested with concentrated H₂SO₄-HNO₃ for 75 min at 350°C [24], using the phosphomolybdate blue method [24].

Sequential Fractionation

Manure samples were sequentially extracted by a modified procedure developed by Dou et al. (2000) [7] (commonly applied to manure P), which was based on Hedley et al. (1982) [19] for fractionation of soil P. In the sequential fractionation, manure samples were extracted sequentially with deionized water, 0.5 M NaHCO₃, 0.1 M NaOH, 1.0 M HCl and concentrated H₂SO₄-HNO₃. Each extraction was performed in a 1:100 manure-to-

Manure	₽H†	AI‡	Ca	Fe	Mg	Мn	٩	U	z	C/N Ratio§
		g kg ⁻¹ dry weig	ht					mg kg ⁻¹		
Dairy manure	8.3	0.97	17.4	1.3	11	0.2	7	39	2	20
Swine manure	8.2	0.87	51.1	1.1	10	0.4	32	37	e	12
Broiler litter	6.4	0.59	24.2	0.7	7.6	0.2	14	36	5	7
Note: ¹ PH was determined on a 10:1 water (i ¹ ¹ btal elements in three animal manur ¹ fotal nitrogen and carbon in three an ⁴ fotal nitrogen and carbon 0102698.t001 doi:10.1371/journal.pone.0102698.t001	nL) to dry matter rati es determined by mid imal manures determ imal	io (g). rrowave digestion ined by Vario MAC	in concentrated F	4NO ₃ and H ₂	.O ₂ and detecti	on by ICP-OES				

study

in this s

manures used

1. Selected characteristics of the animal

Table

Table 2. The c	concentration of total P in anir	nal feed.		
ltem	Dairy feed	Swine feed	Broiler feed	
		g P kg ⁻¹		
Total P†	3.2	7.3	4.9	

Note:

I

[†]Total phosphorus was determined using H_2SO_4 - H_2O_2 digestion of original animal feed, and then tested using Murphy-Riley (1962). doi:10.1371/journal.pone.0102698.t002

solution ratio for 1 h [7]. For all procedures, extracts were made using 0.3 g dried, ground manure samples, shaken with 30 mL extractant at 20°C on a shaker at a speed of 150 excursions min⁻¹ for one hour, centrifuged at 10000×g for 10 min, and then suction-filtered through 0.45 μ m nitrocellulose membranes. The filtrate was analyzed for inorganic P (P_i) and total P (P_t). Organic P in the extracts was determined by difference of orthophosphate detected using the phosphomolybdate blue method of Murphy and Riley (1962) [25] before and after an aliquot filtrate was digested by sulfuric acid-persulfate [26,27]. At the end of the shaking/filtering cycle, residues on the filter membrane and the screw-cap centrifuge tube were transferred to a digestion tube using deionized water. Manure P in residues was determined by digestion by refluxing with concentrated H₂SO₄ and HNO₃ for 75 min at 350°C [24].

NaHCO₃/NaOH+EDTA procedure and solution ³¹P-NMR

The NaHCO₃/NaOH+EDTA procedure involved initial extraction of manure in 0.5 M NaHCO₃ for 4 h, followed by extraction overnight (16 h) in a solution containing 0.5 M NaOH

and 50 mM EDTA, which was first developed by Turner and Levtem (2004) [10]. Readily soluble P was extracted by NaHCO₃ followed by extraction of stable P in a NaOH+EDTA solution. In the procedure, the manure-to-solution ratio was 1:60 for the NaHCO₃ and 1:20 for the NaOH-EDTA extract [17]. Extractions were performed in the same way as the sequential fractionation. An aliquot (5 mL) of each extract was diluted and analyzed separately for P_i and P_t. The remainder of the solution of replicate extract was then frozen rapidly at -80° C, lyophilized, and ground to a fine powder. Before ICP-OES analysis and lyophilization, the solutions extracted by NaHCO3 were acidified with dilute HCl to approximately pH 3 to dissolve carbonates [10]. Approximately 100 mg of each lyophilized extract was subsequently dissolved in 0.9 mL of a solution containing 1 M NaOH and 0.1 M EDTA, and 0.1 mL deuterium oxide (D₂O) was added for signal lock before NMR spectroscopy. The inclusion of NaOH ensures consistent chemical shift at pH>13. Samples were then transferred to a 5-mm NMR tube and stored at 4°C before analysis within 12 h. Solution ³¹P-NMR spectra were obtained using a Bruker

Table 3. The concentration of phosphorus compounds in sequential extracts of animal manures from the modified Hedley fraction procedure determined by Murphy-Riley (1962).

P fraction†	Dairy manure	Swine manure	Broiler litter
		g P kg ⁻¹ of dry matter	
	Inorganic P		
H ₂ O-P _i	2.1 (31%)‡	5.0 (16%)	3.2 (29%)
NaHCO ₃ -P _i	2.4 (35%)	6.5 (21%)	1.3 (12%)
NaOH-P _i	0.07 (1%)	0.8 (3%)	0.3 (3%)
HCI-P _i	0.07 (1%)	5.7 (18%)	0.7 (6%)
$\sum P_i$ fractions	4.64 (68%)	18.0 (58%)	5.5 (50%)
	Organic P		
H ₂ O-P _o	0.5 (8%)	2.0 (6%)	0.3 (3%)
NaHCO ₃ -P _o	0.89 (13%)	1.6 (5%)	2.8 (25%)
NaOH-P _o	0.48 (7%)	1.9 (6%)	1.2 (10.8%)
HCI-P _o	0.15 (2%)	7.8 (24.8%)	1.3 (11%)
$\sum P_o$ fractions	2.0 (30%)	13.3 (41.8%)	5.6 (49.8%)
Residual P [¶]	0.15 (2%)	0.09 (0.2%)	0.02 (0.2%)
Total P	6.79	31.4	11.1

Note:

⁺H₂O-P_i and H₂O-P_o are water-extractable inorganic and organic phosphorus; NaHCO₃-P_i and NaHCO₃-P_o are NaHCO₃-extractable inorganic and organic phosphorus; NaOH-P_i and NaOH-P_o are NaOH-extractable inorganic and organic phosphorus; HCl-P_i and HCl-P_o are HCl-extractable inorganic and organic phosphorus. ⁺Inorganic phosphorus (P_i) in each extract was determined by Murphy-Riley (1962). Organic phosphorus (P_o) in each extract was calculated as the difference between P_t

and P_i. Values in parenthesis are the percentage of total manure phosphorus in each extract.

[¶]Residual P was determined using H₂SO₄-H₂O₂ digestion of the residual.

doi:10.1371/journal.pone.0102698.t003

Table 4. Concentration of phosphorus compounds in two steps extracts of animal manures determined by Murphy-Riley (1962).

P fraction†	Dairy manure	Swine manure	Broiler litter
		g P kg ⁻¹ of dry matter	
	Inorganic P		
NaHCO ₃ -P _i	3.6 (48%)	4.6 (14%)	3.5 (22%)
NaOH/EDTA-P _i	0.3 (4%)	2.3 (7%)	2.8 (18%)
$\sum P_i$ fractions	3.9 (52%)	6.9 (21%)	6.3 (40%)
	Organic P		
NaHCO ₃ -P _o	0.3 (4%)	0.3 (1%)	1.4 (9%)
NaOH/EDTA-P _o	2.2 (29%)	19.0 (59%)	5.7 (36%)
$\sum P_o$ fractions	2.5 (33%)	19.3 (60%)	7.1 (45%)
Residual P¶	1.1 (15%)	6.2 (19%)	2.4 (15%)
Total P	7.5	32.4	15.8

Note:

[†]NaHCO₃-P_i and NaHCO₃-P_o are NaHCO₃-extractable inorganic and organic phosphorus; NaOH/EDTA-P_i and NaOH/EDTA-P_o are NaOH/EDTA-extractable inorganic and organic phosphorus.

[‡]Inorganic phosphorus (P_i) in each extract was determined by Murphy-Riley (1962). Organic phosphorus (P_o) in each extract was calculated as the difference between P_t and P_i . Values in parenthesis are the percentage of total manure phosphorus in each extract.

[¶]Residual P was determined using H_2SO_4 - H_2O_2 digestion of the residual.

doi:10.1371/journal.pone.0102698.t004

Avance DRX 400 MHz spectrometer (Beijing, China) operating at 202.456 MHz for 31 P and 500.134 MHz for 1 H [10].

P compounds from the ³¹P-NMR spectra were identified by their chemical shifts of signals (ppm) relative to 85% H_3PO_4 [10,17,28,29]. Spectra were measured using Mestre-C software. The orthophosphate peak in each spectrum was factitiously standardized to 6 ppm to simplify comparisons among spectra. Chemical shift of signals was used to identify P compounds, including orthophosphate (6 ppm), orthophosphate monoesters between 5.8 and 3.94 ppm, pyrophosphate (-4.53 ppm) and orthophosphate diesters between -0.31 and 3.91 ppm [10,30,31,32]. The concentration of P compounds in extracts was calculated by multiplying the proportion of the spectral area assigned to the corresponding signal by the total phosphorus concentration (mg P kg⁻¹ dry manure) in the original extract.

Results and Discussion

Total P concentration of animal manures

Total P concentration of animal manures determined by ICP ranged from 32 g P kg⁻¹ in swine manure, 14 g P kg⁻¹ in broiler litter and 7 g P kg⁻¹ in dairy manure. Dou et al. (2003) [33] reported a total P concentration rang of 6 to 12 g P kg⁻¹ in dairy manures from 70 farms in USA. Turner and Leytem (2004) [10] reported a total P concentration of 15.9, 4.9, and 14.6 g P kg⁻¹ in broiler litter, cattle manure and swine manure in USA, respectively. Maguire et al. (2004) [34] reported a concentration range of 8.6 to 17.8 g $P kg^{-1}$ in broiler litter in USA. Pagliari and Laboski (2012) [35] reported that total P recovered in the sequential fractionation averaged 7.4, 6.5, 19.4, 21.2, and 33.9 g P kg⁻¹ in beef, dairy, turkey, chicken, and swine manure, respectively, collected from private farms and the University of Wisconsin Agricultural Research Station in Arlington, Wisconsin. Generally, the results in this study indicate a similar total P concentration for dairy manure and broiler litter, but a greater of total P concentration for swine manure in China compared with developed countries and much higher than those in China twenty years ago, which was only 9.0 g P kg⁻¹ [36]. The difference in

swine manure is likely to be caused by mineral P supplementation of the swines' diet [8]. The concentration of total P in swine feed is much higher than the feed standard which leads to the highconcentration of P in excreted manure (Table 2) [37]. In this study, the concentration of total P in swine manure was 4.4 times greater than that in dairy manure and 2.3 times that in broiler litter. So, more attention needs to be paid to the long-term swine manure application to the agricultural land which often leads to soil P accumulation above the amounts sufficient for optimal crop vields and an increased potential risk for P loss in runoff as well as in leachate [5]. Total Al, Fe, Mg, and Mn concentrations were greater in dairy manure and swine manure than in broiler litter, which may be the result of high pH in swine and dairy manure (Table 1). The concentration of Ca was significantly higher in swine manure than that in the other two types of manure due to the Ca-P additives in the diet. Organic C was similar among all three types of manure, whereas broiler litter contained the greatest amount of nitrogen (Table 1).

Sequential fractionation

In the sequential fractionation procedure, the percent of cumulative P in dairy manure was 39 (e.g., 39 is the sum of inorganic and organic P), 48, 8, 3 and 2% associated with extractants H₂O, NaHCO₃, NaOH, HCl and residues, respectively (Table 3). Corresponding P fractions for the swine manure and broiler litter, H_2O extracted 22 and 32% (as H_2O -P); NaHCO₃-P, 26 and 37%; NaOH-P, 9 and 13.8%; HCl-P, 42.8 and 17%; and residue-P, 0.2 and 0.2% of the total P, respectively (Table 3). In these three types of manure, more P was in the NaHCO3 and H2O extracted fractions followed by NaOH and HCl extracted fractions which suggested an increased risk of P loss through dissolving in rainwater or irrigation water from manure when surfaced applied. The majority of P components in dairy and swine manure were inorganic (68% and 58%), while the concentration of inorganic- and organic P was about equal in broiler litter. The concentration of P extracted by HCl from swine manure was higher than that from other manures. It is probable that Ca-P is the primary complex because Ca²⁺ is the dominating



Figure 1. Solution ³¹P-nuclear magnetic resonance spectra of sequential extracts of dairy manure, swine manure, and broiler litter involving extraction in 0.5 *M* NaHCO₃ and a solution of 0.5 *M* NaOH and 50 m*M* EDTA. The main spectra were scaled to the full height of the phosphate signal at approximately 6.0 ppm. Spectra are expanded to show detailed peak assignments in the region from -6 to 9 ppm. doi:10.1371/journal.pone.0102698.g001

cation in swine manure [38,39,40,41]. He et al. (2006, 2008) [12,14] found some organic P presented in the HCl extracted fraction, which was also confirmed by Takahashi (2013) [42]. In this study, P_o -HCl was not determined that may cause underestimating of total organic P. The proportion of organic P to total P in dairy manure was 30%, which was lower than swine manure (41.8%) and broiler litter (49.8%) (Table 3). The reason may be the dairy manure was from ruminant animal, which could secrete phytases and other phosphatases from their gut to increase the organic P hydrolysis [34,43]. Another reason may be that there was less phytate in the diet for dairy, particularly when the dairy was grazed on grassland.

NaHCO₃/EDTA procedure

Total P extracted by the NaHCO₃/EDTA was 7.5, 32.4 and 15.8 g P kg⁻¹ in dairy manure, swine manure and broiler litter, respectively (Table 4). The recovery ratio of total P from manures was from 101% to 109% which confirmed that the procedure was an effective method to extract P from these manures (Table 4). The P extracted in this procedure was higher than that obtained with the sequential procedure. The probable reason was the P losses during the filtration through the 0.45 μ m nitrocellulose membranes in the sequential procedure. For the readily soluble P fraction, an initial NaHCO₃ extraction recovered less total P from three manures than the combined deionized water plus NaHCO₃ extraction in the sequential procedure (Table 4). This may be due to the presence of a high concentration of phosphate ions contained in the animal manure, while an initial NaHCO₃

res	
npa	
Ű,	
pr	
tior	
nat	
ctio	
frac	ses
ed	the
evis	ent
e Le	par
Ę	in.
⊿ i⊔	al P
1C	ota
1 El	of t
М	ns
50	forr
pu	Р
Чаї	e of
aO	age
Z Z	ent
5 2	erce
j 0.	; p(
jing	bm
tair	; pl
iuo	hift
с С	al s
ltio	nic
solt	her
g	e cl
vith	: th
2	ent
ctio	res
trac	rep
eX.	9
puq	to
ecc	0.31
аs)-) /
⊒.	(do
nds	osc
nou	ctre
dш	spe
8	Ge
rus	้าลท
ho	sor
osp	c re
hd	etic
of	agn
ses	шŝ
clas	ear
lal	ncli
tior	P-n
Incl	0
Ē.	by
ŝ	ted
ble	tec
Ta	dei

Manual Contractor	Inorganic P		a since									
	Orthophosphate	Pyrophosphate	Orthophosph	nate monoeste	ers						Orthophosph	ite diesters
	ø	-4.53	5.8	5.63-5.10	5.07	4.86	4.75	4.49	4.38	4.25-3.94	1.32	-0.31
						g P kg ⁻¹ (%	6 of P in extra	cts)				
Dairy manure	1.48 (20%)	ND	0.11 (1%)	DN	0.12 (2%)	0.23 (3%)	0.14 (2%)	0.21 (3%)	0.13 (2%)	DN	Ŋ	0.074 (1%)
Swine manure	8.84 (27%)	0.15 (0.5%)	2.09 (6%)	0.92 (3%)	QN	3.7 (11%)	DN	3.6 (11%)	1.9 (6%)	ND	ŊŊ	ND
Broiler litter	2.02 (13%)	0.085 (0.5%)	0.79 (5%)	0.82 (5%)	0.11 (1%)	1.42 (9%)	0.55 (3%)	1.62 (10%)	0.78 (5%)	0.28 (2%)	0.027 (0.2%)	DN
Note: ND: not detected. -0.31-6: the chemic doi:10.1371/journal	cal shift, ppm; percenta I.pone.0102698.t005	ige of P forms of tot:	al P in parenthe	ses.								

Phosphorus Characterization in Animal Manure

extraction could not supply sufficient carbonate ions to exchange the phosphate ions absorbed on the surface of the particles. Another reason may be that two extractions are more powerful than one extraction because of a dilution effect. We also noted that about 15–19% of total P was left in a residual P fraction, compared with that in the sequential procedure. It is probable that relatively stable Ca-bound phosphate, which was the major binding complex, could not be extracted completely by NaOH+EDTA in the procedure [39]. However, the alkaline solution improved organic P recovery from animal manure and facilitated the spectral resolution for subsequent ³¹P-NMR analysis [10,17,18].

Solution ³¹P-NMR spectra

Solution ³¹P-NMR spectra of two extracts from three types of animal manure in the NaHCO₃/EDTA procedure were shown in Figure 1. An orthophosphate peak was identified in spectra from all extracts. However, peaks assigned to orthophosphate monoesters and diesters were much less and weaker in NaHCO3 extract spectra than the spectrum from NaOH-EDTA, which indicated the relatively lower concentration compared with those phosphates in the NaHCO₃ extracts. The results were similar to those of Turner and Leytem (2004) [10] and He et al. (2009) [15]. In NaHCO₃ extract spectra, P species, except orthophosphate, were almost undetectable at low resolution. However, P species identified in the NaOH-EDTA extract spectrum of broiler litter and swine manure were abundant (Figure. 1). All peaks presented in the spectrum from NaOH-EDTA were subsequently grouped into P species or P classes if specific identification could not be made, according to the literature (Table 5). Inorganic P species identified from the spectrum included orthophosphate at 6 ppm and pyrophosphate at -4.53 ppm. Detected organic P species included orthophosphate monoesters between 5.8 and 3.94 ppm, while orthophosphate diesters were between -0.31 and 1.32 ppm [29,30,32,44]. The dominant P species in the NaOH-EDTA extracts was orthophosphate at 6 ppm which accounted for 13-27% of total P extracted from three different manures. The following most abundant of P species were phytate (mainly myoinositol hexakisphosphate and α -glycerophosphate) at 5.8, 4.86, 4.49, and 4.38 ppm in the orthophosphate monoesters region (4.38 to 5.8 ppm) in all manures (Table 5). The total concentration of phytate (P species at 5.8, 4.86, 4.49, and 4.38 ppm) was 0.68, 11.29, and 4.61 g kg⁻¹ accounting for 9, 34, and 29% of total P in dairy manure, swine manure and broiler litter, respectively. A similar result was also reported in Toor et al. (2005b) [45]. The possible explanation for this difference was that the ruminant animal of dairy cattle secretes phytase to digest phytate contained in feed which resulted in a reduced quantity of phytate in manure [34,45,46]. Some organic P species such as phytate were more strongly sorbed to soil particles while many others (e.g., orthophosphate diesters) were poorly absorbed to soil, which could lead to localized areas of soil P saturation and P loss [47]. To overcome the low digestibility of phytate P in nonruminant diets and decrease the total P content and organic P species, feed additives such as phytase and other enzymes are necessary, and thus could greatly reduce surpluses of manure P in areas of intensive animal production. For example, Maguire et al. (2004) [34] reported that phytase added to feed could decrease phytate P in broiler litter by 25 to 38%. Pyrophosphate was identified in swine manure and broiler litter, accounting for 0.5% of total P. The concentration of orthophosphate diesters detected in dairy manure and broiler litter has seldom been compared with monoesters which would include intact phospholipids.

In general, The NaHCO₃/NaOH-EDTA procedure was suitable to be used as a simple and efficient test method to

estimate the magnitude of vulnerable P with environmental relevance in manures which could help farmers identify optimal management techniques and treatments to reduce potential P runoff on farms. Based on the accurate analysis of organic P in NaOH-EDTA extracts by ³¹P-NMR, orthophosphate monoesters, especially phytate, were identified as the dominant P species in swine manure and broiler litter. Thus, feeding management including the use of feed additives such as phytase and reducing over-feeding to minimize excessive dietary P could be a necessary and critical practice to reduce phytate and total P excretion, respectively, in manures to reduce the potential risk of P loss to the environment associated with surface application of manures. This would particularly be useful for areas where limited arable land was combined with intensive animal farming. In the future, we must not only promote the development of analytical methods to measure manure P to better understand the manure P dynamics and enhance our capability in management, but also we should be concerned about the changes of P forms, bioavailability, and potential for transport in runoff and animal manure interaction with soil.

Conclusion

In this study, the average total P determined by two different methods was 31.9 g P kg^{-1} in swine manure, 13.5 g P kg^{-1} in

References

- Wang F, Dou Z, Ma L, Ma W, Sims JT, et al. (2010) Nitrogen Mass Flow in China's Animal Production System and Environmental Implications. J Environ Qual 39: 1537.
- Knowlton KF, Kohn R (1999) Feeding management to reduce phosphorus losses from dairy farms. Proceedings of the Mid Atlantic Dairy Management Conference 24–25.
- Wang F, Sims JT, Ma L, Ma W, Dou Z, et al. (2011) The Phosphorus Footprint of China's Food Chain: Implications for Food Security, Natural Resource Management, and Environmental Quality. J Environ Qual 40: 1081.
- Williams PH, Haynes RJ (1995) Effect of sheep, deer and cattle dung on herbage production and soil nutrient content. Grass Forage Sci 50: 263–271.
- Sims JT, Edwards AC, Schoumans OF, Simard RR (2000) Integrating soil phosphorus testing into environmentally based agricultural management practices. J Environ Qual 29: 60–71.
- Leinweber P, Haumaier L, Zech W (1997) Sequential extractions and ³¹P-NMR spectroscopy of phosphorus forms in animal manures, whole soils and particlesize separates from a densely populated livestock area in northwest Germany. Biol Fertil Soils 25: 89–94.
- Dou Z, Toth JD, Galligan DT, Ramberg CF, Ferguson JD (2000) Laboratory procedures for characterizing manure phosphorus. J Environ Qual 29: 508–514.
- Sharpley A, Moyer B (2000) Phosphorus forms in manure and compost and their release during simulated rainfall. J Environ Qual 29: 1462–1469.
- Sharpley A, Tunney H (2000) Phosphorus research strategies to meet agricultural and environmental challenges of the 21st century. J Environ Qual 29: 176–181.
- Turner BL, Leytem AB (2004) Phosphorus compounds in sequential extracts of animal manures: Chemical speciation and a novel fractionation procedure. Environ Sci Technol 38; 6101–6108.
- He Z, Griffin TS, Honeycutt CW (2004) Enzymatic hydrolysis of organic phosphorus in swine manure and soil. J Environ Qual 33: 367–372.
- He Z, Toor GS, Honeycutt CW, Sims JT (2006) An enzymatic hydrolysis approach for characterizing labile phosphorus forms in dairy manure under mild assay conditions. Bioresource Technol 97: 1660–1668.
- He Z, Cade-Menun BJ, Toor GS, Fortuna AM, Honeycutt CW, et al. (2007) Comparison of Phosphorus Forms in Wet and Dried Animal Manures by Solution Phosphorus-31 Nuclear Magnetic Resonance Spectroscopy and Enzymatic Hydrolysis. J Environ Qual 36: 1086–1095.
- He Z, Honeycutt CW, Cade-Menun BJ, Senwo ZN, Tazisong IA (2008) Phosphorus in Poultry Litter and Soil: Enzymatic and Nuclear Magnetic Resonance Characterization. Soil Sci Soc Am J 72:1425–1433.
- He Z, Honeycutt CW, Griffin TS, Cade-Menun BJ, Pellechia PJ, et al. (2009) Phosphorus forms in conventional and organic dairy manure identified by solution and solid state P-31 NMR spectroscopy. J Environ Qual 38: 1909–1918.
- Hansen JC, Cade-Menun BJ, Strawn DG (2004) Phosphorus speciation in manure-amended alkaline soils. J Environ Qual 33: 1521–1527.
- Turner BL (2004) Optimizing phosphorus characterization in animal manures by solution phosphorus-31 nuclear magnetic resonance spectroscopy. J Environ Qual 33: 757–766.

broiler litter and 7.1 g P kg⁻¹ in dairy manure collected from three farms, respectively, which was similar with developed countries, and much higher than concentrations reported in China in the 1990s (8.5–9.5 g P kg⁻¹ in swine manure, 8.7–9.9 g P kg⁻¹ in broiler litter and 4.1–4.5 g P kg⁻¹ in dairy manure in 1990s) [36]. Our results show that more than 48% of total P in swine manure, 87% of total P in dairy manure and 69% of total P in broiler litter was water-soluble P (P_t extracted by Water and NaHCO₃), which is easily lost into the environment. It is reported that 1582 Gg of animal P was presumably lost to the environment in 2006 in China, which is a big threat to water quality [3]. Therefore, an integrated management practice is necessary to reduce manure P losses at animal farm scale, including animal diet modification, livestock manure collection, storage and field application.

Author Contributions

Conceived and designed the experiments: HL JS FZ. Performed the experiments: GL. Analyzed the data: GL. Contributed reagents/materials/ analysis tools: HL FZ. Wrote the paper: GL HL. Extensive review and editing of manuscript: PAL JS.

- McDowell RW, Dou Z, Toth JD, Cade-Menun BJ, Kleinman PJA, et al. (2008) A Comparison of Phosphorus Speciation and Potential Bioavailability in Feed and Feces of Different Dairy Herds Using ³¹P Nuclear Magnetic Resonance Spectroscopy. J Environ Qual 37: 741–752.
- Hedley MJ, Stewart JWB, Chauhan BS (1982) Changes in Inorganic and Organic Soil Phosphorus Fractions Induced by Cultivation Practices and by Laboratory Incubations. Soil Sci Soc Am J 46: 970.
- Ajiboye B, Akinremi OO, Racz GJ (2004) Laboratory characterization of phosphorus in fresh and oven-dried organic amendments. J Environ Qual 33: 1062–1069.
- Cade-Menun BJ, Liu CW, Nunlist R, McColl JG (2002) Soil and litter phosphorus-31 nuclear magnetic resonance spectroscopy: Extractants, metals, and phosphorus relaxation times. J Environ Qual 31: 457–465.
- Cade-Menun BJ, Navaratnam JA, Walbridge MR (2006) Characterizing dissolved and particulate phosphorus in water with ³¹P nuclear magnetic resonance spectroscopy. Environ Sci Technol 40: 7874–7880.
- Leinweber P, Lunsmann F, Eckhardt KU (1997b) Phosphorus sorption capacities and saturation of soils in two regions with different livestock densities in northwest Germany. Soil Use and Manage 13: 82–89.
- American Public Health Association (1995) Standard Methods for the Examination of Water and Wastewater. Washington, DC: American Public Health Association.
- Murphy J, Riley JP (1962) A modified single solution method for the determination of phosphate in natural waters. Anal Chim Acta 27: 31–36.
- Watanabe FS, Olsen SR (1965) Test of an ascorbic acid method for determining phosphorus in water and NaHCO₃ extracts from soil. Soil Sci Soc Am J 29: 677–678.
- McDowell RW, Stewart I (2005) Phosphorus in fresh and dry dung of grazing dairy cattle, deer, and sheep: Sequential fraction and phosphorus-31 nuclear magnetic resonance analyses. J Environ Qual 34: 598–607.
- Hill JE, Cade-Menun BJ (2009) Phosphorus-31 nuclear magnetic resonance spectroscopy transect study of poultry operations on the Delmarva Peninsula. J Environ Qual 38: 130.
- Turner B L, Mahieu N, Condron LM (2003) Phosphorus-31 nuclear magnetic resonance spectral assignments of phosphorus compounds in soil NaOH-EDTA extracts. Soil Sci Soc Am J 67: 497–510.
- Cade-Menun BJ, Benitez-Nelson CR, Pellechia P, Paytan A (2005) Refining ³¹P nuclear magnetic resonance spectroscopy for marine particulate samples: Storage conditions and extraction recovery. Mar Chem 97: 293–306.
- Toor GS, Cade-Menun BJ, Sims JT (2005a) Establishing a linkage between phosphorus forms in dairy diets, feces, and manures. J Environ Qual 34: 1380– 1391.
- Cade-Menun BJ, Paytan A (2010) Nutrient temperature and light stress alter phosphorus and carbon forms in culture-grown algae. Mar Chem 121: 27–36.
- Dou Z, Ferguson JD, Fiorini J, Toth JD (2003) Phosphorus feeding levels and critical control points on dairy farms. J Dairy Sci 86: 3787–3795.

- Maguire RO, Sims JT, Saylor WW, Turner BL, Angel R, et al. (2004) Influence of phytase addition to poultry diets on phosphorus forms and solubility in litters and amended soils. J Environ Qual 33: 2306–2316.
- Pagliari PH, Laboski CA (2012) Investigation of the Inorganic and Organic Phosphorus Forms in Animal Manure. J Environ Qual 41: 901–910.
- NATEas Center (1999) China organic fertilizer. Beijing, China. p. 6. [In Chinese].
- Huaitalla RM, Gallmann E, Liu X, Hartung E (2011) Nutrients and Trace Elements in a Pig Farm in Beijing: Chinese and German Recommendations. J Agr Sci Tech 2: 33–50.
- Safley LM, Westerman PW, Barker JC (1986) Fresh dairy manure characteristics and barnlot nutrient losses. Agr Waste 17: 203–215.
- Kuo S, Hummel RL, Jellum EJ, Winters D (1999) Solubility and leachability of fishwaste compost phosphorus in soilless growing media. J Environ Qual 28: 164–169.
- He Z, Zhang H, Toor GS, Dou Z, Honeycutt CW, et al. (2010) Phosphorus Distribution in Sequantial Extracted Fractions of Biosolids, Poultry Litter, and Granulated Products. Soil Sci 175: 154–161.

- He Z, Honeycutt CW, Griffin TS (2003) Comparative Investigation of Sequentially Extracted Phosphorus Fractions in a Sandy Loam Soil and a Swine Manure. Comm Soil Sci Plant Anal 34: 1729–1742.
- Takahashi S (2013) Phosphorus characterization of manure composts and combined organic fertilizers by a sequential-fractionation method. J. Plant Nutr Soil Sci 176: 494–496.
- Maguire RO, Sims JT, Applegate TJ (2005) Phytase supplementation and reduced-phosphorus turkey diets reduce phosphorus loss in runoff following litter application. J Environ Qual 34: 359–369.
- Cade-Menun BJ (2005) Characterizing phosphorus in environmental and agricultural samples by ³¹P nuclear magnetic resonance spectroscopy. Talanta 66: 359–371.
- 45. Toor GS, Sims JT, Dou Z (2005b) Reducing phosphorus in dairy diets improves farm nutrient balances and decreases the risk of nonpoint pollution of surface and ground waters. Agr Ecosystem Environ 105: 401–411.
- Bieleski RL (1973) Phosphate pools, phosphate transport, and phosphate availability. Ann Rev Plant Physiol 24: 225–252.
- Leytem A, Mikkelsen RL, Gilliam JW (2002) Sorption of organic phosphorus compounds in Atlantic coastal plain soils. Soil Sci 167: 652–658.