

# Genome Sequence of *Bacillus thuringiensis* subsp. *kurstaki* Strain HD-1

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**We report here the complete genome sequence of *Bacillus thuringiensis* subsp. *kurstaki* strain HD-1, which serves as the primary U.S. reference standard for all commercial insecticidal formulations of *B. thuringiensis* manufactured around the world.**

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*Bacillus thuringiensis* is a ubiquitous Gram-positive bacterium that has been isolated from a variety of ecological niches. The most distinctive property of *B. thuringiensis* is its production of parasporal insecticidal proteins (Cry toxins) whose entomopathogenicity spans a number of insect species among the orders *Lepidoptera* (moths and butterflies), *Diptera* (mosquitoes and blackflies), *Coleoptera* (beetles), and *Hymenoptera* (wasps, ants, and others) (1). These features of *B. thuringiensis* have been exploited for more than 50 years to control a number of insects, including agriculturally and medically important pest and disease vector insects. Indeed, *B. thuringiensis* exhibits highly specific and selective entomopathogenicity and is safe for humans (2–5). Among all strains of *B. thuringiensis*, the HD-1 strain (serotypes 3a and 3b) has been the most intensely used environmentally compatible biopesticide worldwide, and it has been designated the primary U.S. reference standard for the standardization of all commercial formulations of *B. thuringiensis* produced globally (6).

Total genomic DNA was extracted using the high-salt SDS method (7), followed by purification using the phenol-chloroform-isoamyl alcohol protocol (24:24:1 [vol/vol/vol]), as described previously (8). Sequencing was performed on an Illumina MiSeq, using a 300-bp paired-end library, by the Oklahoma Medical Research Foundation Core Facility (<http://omrf.org/research-faculty/core-facilities/>), generating 39,682,366 reads. Read quality was assessed with FastQC (<http://www.bioinformatics.babraham.ac.uk/projects/fastqc/>). The MiSeq reads were *de novo* assembled using the A5 assembly pipeline (9), yielding 194 contigs, for a total contig length of 6,391,935 bp, with 34.75% G+C content and an  $N_{50}$  of 96,479 bp. Automated annotation was performed using the RAST server (10). The resulting contig database contains 11 plasmids, including the previously identified pBMB2062 (11), six plasmids highly similar to those found in *B. thuringiensis* subsp. *kurstaki* HD73 (12), two plasmids similar to those found in *B. thuringiensis* subsp. *thuringiensis* 5056 (13), and one plasmid similar to pBMB9741, found in *B. thuringiensis* subsp. *kurstaki* YBT-1520 (14). We also identified a linear bacteriophage very similar to

GIL16c (15). Using the *glpF*, *gmk*, *ilvD*, *pta*, *pur*, *pycA*, and *tpi* genes, we confirmed that strain HD1 is in sequence type 8 (ST8) of the *Bacillus cereus* multilocus sequence type database (16). A comparison of strain HD1 with *B. thuringiensis* HD73 indicated that HD1 lacks approximately 500 kb of DNA found in HD73 while containing approximately 450 kb of DNA not found in strain HD73. The remainder of strain HD1 is highly collinear with HD73. In HD-1, there are 25 rRNA genes, 12 of which code for the 16S ribosomal subunit, 4 for the 23S ribosomal subunit, and 9 for the 5S rRNA subunit. The genome has 84 tRNAs, including five pseudo-tRNAs, which may be tRNA remnants that do not function in translation but may be involved in the maintenance of other cellular functions, such as cell wall biosynthesis and antibiotic resistance, among others (17). These data will aid in-depth explorations of the unique phenotypic properties of this important biocontrol agent.

**Nucleotide sequence accession numbers.** This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession no. [JMHW00000000](https://www.ncbi.nlm.nih.gov/nuclink/JMHW00000000). The version described in this paper is version [JMHW01000000](https://www.ncbi.nlm.nih.gov/nuclink/JMHW01000000).

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## REFERENCES

1. Ibrahim MA, Griko N, Junker M, Bulla LA. 2010. *Bacillus thuringiensis*: a genomics and proteomics perspective. *Bioeng. Bugs* 1:31–50. [http://dx.doi.org/10.4161/bbug.1.1.10519](https://doi.org/10.4161/bbug.1.1.10519).
2. Ellis RT, Stockhoff BA, Stamp L, Schnepf HE, Schwab GE, Knuth M, Russell J, Cardineau GA, Narva KE. 2002. Novel *Bacillus thuringiensis* binary insecticidal crystal proteins active on western corn rootworm, *Diabrotica virgifera virgifera* LeConte. *Appl. Environ. Microbiol.* 68:1137–1145. [http://dx.doi.org/10.1128/AEM.68.3.1137-1145.2002](https://doi.org/10.1128/AEM.68.3.1137-1145.2002).
3. Pearce M, Habbick B, Williams J, Eastman M, Newman M. 2002. The effects of aerial spraying with *Bacillus thuringiensis* *kurstaki* on children with asthma. *Can. J. Public Health* 93:21–25.
4. Valadares De Amorim G, Whittome B, Shore B, Levin DB. 2001. Identification of *Bacillus thuringiensis* subsp. *kurstaki* strain HD1-like bacteria from environmental and human samples after aerial spraying of Vic-

- toria, British Columbia, Canada, with Foray 48B. *Appl. Environ. Microbiol.* 67:1035–1043. <http://dx.doi.org/10.1128/AEM.67.3.1035-1043.2001>.
5. van Netten C, Teschke K, Leung V, Chow Y, Bartlett K. 2000. The measurement of volatile constituents in Foray 48B, an insecticide prepared from *Bacillus thuringiensis* var. *kurstaki*. *Sci. Total Environ.* 263: 155–160. [http://dx.doi.org/10.1016/S0048-9697\(00\)00696-3](http://dx.doi.org/10.1016/S0048-9697(00)00696-3).
  6. Dulmage HT. 1973. *B. thuringiensis* U.S. assay standard: report on the adoption of a primary U.S. reference standard for assay of formulations containing the  $\delta$ -endotoxin of *Bacillus thuringiensis*. *Bull. Entomol. Soc. Am.* 19:200–202.
  7. Zhou J, Bruns MA, Tiedje JM. 1996. DNA recovery from soils of diverse composition. *Appl. Environ. Microbiol.* 62:316–322.
  8. Green M, Sambrook J. 2012. Preparation of plasmid DNA by alkaline lysis with SDS: minipreps, p 11. *Molecular cloning: a laboratory manual*. Cold Spring Harbor Press, Cold Spring Harbor, NY.
  9. Tritt A, Eisen JA, Facciotti MT, Darling AE. 2012. An integrated pipeline for *de novo* assembly of microbial genomes. *PLoS One* 7:e42304. <http://dx.doi.org/10.1371/journal.pone.0042304>.
  10. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:75. <http://dx.doi.org/10.1186/1471-2164-9-75>.
  11. Zhong C, Peng D, Ye W, Chai L, Qi J, Yu Z, Ruan L, Sun M. 2011. Determination of plasmid copy number reveals the total plasmid DNA amount is greater than the chromosomal DNA amount in *Bacillus thuringiensis* YBT-1520. *PLoS One* 6:e16025. <http://dx.doi.org/10.1371/journal.pone.0016025>.
  12. Liu G, Song L, Shu C, Wang P, Deng C, Lereclus D, Wang X, Huang D, Zhang J. 2013. Complete genome sequence of *Bacillus thuringiensis* subsp. *kurstaki* strain HD73. *Genome Announc.* 1(2):e00080-13. <http://dx.doi.org/10.1128/genomeA.00080-13>.
  13. Murawska E, Fiedoruk K, Bideshi DK, Swiecicka I. 2013. Complete genome sequence of *Bacillus thuringiensis* subsp. *thuringiensis* strain IS5056, an isolate highly toxic to *Trichoplusia ni*. *Genome Announc.* 1(2): e00108-13. <http://dx.doi.org/10.1128/genomeA.00108-13>.
  14. Zhang Q, Sun M, Xu Z, Yu Z. 2007. Cloning and characterization of pBMB9741, a native plasmid of *Bacillus thuringiensis* subsp. *kurstaki* strain YBT-1520. *Curr. Microbiol.* 55:302–307. <http://dx.doi.org/10.1007/s00284-006-0623-3>.
  15. Verheust C, Fornelos N, Mahillon J. 2005. GIL16, a new Gram-positive tectiviral phage related to the *Bacillus thuringiensis* GIL01 and the *Bacillus cereus* pBClin15 elements. *J. Bacteriol.* 187:1966–1973. <http://dx.doi.org/10.1128/JB.187.6.1966-1973.2005>.
  16. Jolley KA, Maiden MC. 2010. BIGSdb: scalable analysis of bacterial genome variation at the population level. *BMC Bioinformatics* 11:595. <http://dx.doi.org/10.1186/1471-2105-11-595>.
  17. Rogers TE, Ataide SF, Dare K, Katz A, Seveau S, Roy H, Ibba M. 2012. A pseudo-tRNA modulates antibiotic resistance in *Bacillus cereus*. *PLoS One* 7:e41248. <http://dx.doi.org/10.1371/journal.pone.0041248>.