

WILEY

Biopsy-based single-cell transcriptomics reveals MAIT cells as potential targets for controlling fibrosis-related liver inflammation due to chronic hepatitis-B infection

Dear Editor,

Chronic hepatitis B virus (HBV) infection affects about 844 million people worldwide, with two million annual deaths and a rising incidence. Persistent HBV infection may lead to liver inflammation and fibrosis, a leading risk for liver transplantation and hepatocellular carcinoma.¹ However, few treatment options for controlling HBVrelated liver inflammation are currently available. The limited understanding on corresponding mechanisms impedes the discovery of rational therapeutics targeting on liver inflammation and fibrosis. Thus, we conducted a single-cell transcriptomic study² to better understand the mechanisms underlying chronic HBV-related liver inflammation and to enable the discovery of therapeutic targets.

Liver biopsies from six chronic HBV patients with different grades of liver inflammation were included in the present study (Figure S1; Table S1). Single-cell RNA sequencing of biopsied liver tissues was performed. Functions and cross-talks of single cells were also investigated (provided in Supplementary Materials).

Based on around 24,000 single cells, we obtained 12 cell clusters. Among them, T cell population was the most abundant (Figure 1; Figure S2). By assigning the samples to different grades of inflammation (G), no apparent perturbation was observed in respect to the fractions of cells between grades G1 and G2 (Table S2). We further classified the 16, 386 T cells into nine cell subclusters, six of which varied significantly as liver inflammation progresses (Figure 2A,B). We then assigned them to known lineages based on marker genes (Figure 2C,D; Table S3). Concisely, T7 and T6 were annotated as CD8⁺ mucosal-associated invariant T (MAIT) cells. T2 and T0 were identified as activated and exhausted CD8⁺ T cells, respectively. Most interestingly, MAIT cells, especially T7, decreased, while the ratio of T6 versus T7 increased in grade G2. Moreover, T7 dominated in grade G1 (95.20%) (Figure 2E and

Figure S3A,B). The above findings were confirmed by additional single-cell RNA sequencing data from liver samples of HBV-infected patients with grades G1 (n = 4) and G2 (n = 4) (Figure S3C,D).² T6 was assumed to be developed from T7 (Figure 2F), with genes such as *CCL3L1*, *PTMS*, *KLRG1* and *GADD45B* responsible for the transition (Figure S3E). Additionally, macrophage subcluster Ma3, inclined to have M2 signature, increased in grade G2 (Figure S4). Although no apparent perturbations were observed for the fractions of different cell types, T cell subpopulations were remodeled during the progression from grades G1 to G2.

We further confirmed the presence of two MAIT cell populations by immunofluorescence, and annotated them as CD3+SLC4A10+TNFAIP3+ (T7) and CD3⁺SLC4A10⁺TNFAIP3⁻ (T6) MAIT cells (Figure 3). T7 may have stronger direct anti-viral functions. Firstly, intrahepatic MAIT cells have been reported to be the most important innate effector cells secreting anti-viral cytokine IFN- γ ,³ and T7 demonstrated up-regulated pathways 'interferon gamma secretion' and 'regulation of response to interferon gamma' (Figure 4A). Secondly, T7 may have stronger type-17 function of MAIT cell populations that might have a certain antiviral role,⁴ evidenced by the up-regulation of pathways such as 'T helper 17 type immune response'. Moreover, T7 may be stronger mediators of immune responses. On one hand, T7 may increase the anti-viral capability of hepatocytes by ligand-receptor pairs such as 'FcRn complex_ALB' (Figure 4B), where FcRn was reported to increase the sensitivity of hepatocyte to toxicity.⁵ Pathways such as 'Toll-like receptor 4 signaling pathways' that can reduce HBV replication in an IFN-independent manner in hepatocytes⁶ was also up-regulated in T7. On the other hand, T7 may be more active in cytokine production and recruiting activated CD4⁺ and CD8⁺ T cells,⁷ evidenced by up-regulated pathways such as 'positive regulation of CD4

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

^{© 2022} The Authors. Clinical and Translational Medicine published by John Wiley & Sons Australia, Ltd on behalf of Shanghai Institute of Clinical Bioinformatics.



FIGURE 1 scRNA-seq reveals cell types in liver biopsies of chronic hepatitis B virus (HBV) patients. (A) A schematic diagram of single-cell sequencing for liver biopsies. (B) uniform manifold approximation and projection (UMAP) plots of single-cell clusters coloured by samples. (C) UMAP plots of single-cell clusters coloured by cell types. (D) Fraction of cells for all cell types in each sample. (E) Marker genes for T cells



FIGURE 2 Cell populations fluctuate as inflammation progresses. (A) UMAP plots of T cell subclusters. (B) Volcano plot illustrating fold change and *p* values for T cell subclusters, where circles in grey mean insignificant. (C) Heatmap showing marker genes of T cell clusters. (D) Dotplot showing the expression level of *CD4*, *CD8A* and *CD8B* in different T cell subsets. (E) Fraction of cells for the six differential T cell subclusters coloured by grade. (F) Developmental trajectory for differential T cell subclusters



FIGURE 3 Characteristics of two mucosal-associated invariant T (MAIT) cell populations. (A) Immunofluorescence of genes *CD3* (red), *SLC4A10* (rose red), *DAPI* (blue) and *TNFAIP3* (green) for liver biopsies obtained from samples S1 (grade G1), S2 (grade G2) and S5 (grade G2). (B) Heatmap of differential genes between two MAIT cell populations. (C) gene set enrichment analysis (GSEA) enriched terms for the genes differentially expressed between two MAIT cell populations



FIGURE 4 Gene set variation analysis (GSVA) and cross-talk among T cell subclusters. (A) Variation of pathways for two mucosal-associated invariant T (MAIT) cell populations using GSVA. (B) The number of interactions and ligand-receptor pairs between MAIT cell subclusters (T7, T6) and others

positive alpha beta T cell activation'. Then, T7 may assist virus infection control via activating NK cell and CD8⁺ T cell mediated immune responses through cross-talks for T7-NK cells and T7-T2, evidenced by ligand-receptor pairs such as 'HLA-C_KIR2DL3' and 'HLA-A_KIR3DL1', which were reported to function as helper masters to antiviral T cell responses.⁸ Such results indicated that MAIT cell population T7 may have active antiviral functions for reducing HBV replication in grade G1. Immunofluorescence results for additional two biopsies obtained from two G1 patients (Figure S5) further validated the presence of T7 in grade G1.

We speculated impaired antiviral roles of T6 in grade G2 from the down-regulation of corresponding pathways mentioned above, and the decrease in pathways 'regulation of defense response to virus' and 'innate immune response' (Figure 4A). Such results implied the existence of MAIT cell dysfunction in grade G2. We also observed a sharp decrease of activated CD8⁺ T cells, and a sharp increase of CD8⁺ exhausted T cells in grade G2. T cell exhaustion, in together with immune dysfunction of T6, may contribute to the inadequate HBV-specific immune responses and inadequate infection control in grade G2. The increased M2-type macrophage activation (Ma3) also implied the impaired immune response⁹ in grade G2. Bacterial superantigens resulting from the leaky gut of chronic HBV patients can make MAIT cells hyperactive and exhausted, which will lead to immunosuppression and increases the risk of secondary opportunistic infections.¹⁰ Thus, the dysfunction of MAIT cells T6 and exhaustion of CD8⁺ T cells may also provide an explanation for the progression of HBV-related inflammation in grade G2. Moreover, MAIT cells demonstrated extensive interactions with other immune cells (provided in Supplementary Materials).

In conclusion, our study reported the remodelling of T cell subpopulations and the presence of two MAIT cell populations that may contribute to HBV-related liver inflammation for the first time. MAIT cell population T7 may have active roles in assisting virus clearance via direct anti-viral functions and mediating immune responses, which was impaired in T6, the proportion of which in CD8⁺ MAIT cells increased in grade G2. The sharp decrease of active CD8⁺ T cells, MAIT cell dysfunction in together may contribute to the inadequate HBV-specific immune responses in grade G2. Inspired from the decrease of MAIT cells and increased ratio of T6 versus T7 during liver inflammation progression, MAIT cells may act as a potential target for controlling HBV-related liver inflammation in future studies. However, studies using more samples are required to confirm the findings between G1 and G2 reported in this study.

ACKNOWLEDGEMENTS

The present study was funded by grants from the National Natural Science Foundation of China (grant numbers: 31870839 and 81870426), the Natural Science Foundation of Zhejiang Province (grant numbers: LZ20H290002 and LY22C010001), the National Youth Top-notch Talent Support Program (grant number: W02070098), the HangZhou Medical and Health Technology Project (grant number: Z20200052), and the Innovation Team and Talents Cultivation Program of National Administration of Traditional Chinese Medicine (grant number: ZYYCXTD-D-202002).

CONFLICT OF INTEREST

The authors declare that they have no conflict of interest.

ACKNOWLEDGEMENTS

This work was supported by the key medical disciplines of Hangzhou and the Westlake Laboratory (Westlake Laboratory of Life Sciences and Biomedicine). The authors thank all the participants who helped collecting liver biopsies.

> Li Shao¹ Hong Zhao³ Rongfang Guo² Jinlin Cheng³ Xiaoyan Lu² Xiaohui Fan^{2,4,5} (**b**)

 ¹School of Clinical Medicine, Hangzhou Normal University, The Affiliated Hospital of Hangzhou Normal University, Hangzhou, China
²Pharmaceutical Informatics Institute, College of Pharmaceutical Sciences, Zhejiang University, Hangzhou, China
³The First Affiliated Hospital, School of Medicine, Zhejiang University, Hangzhou, China
⁴Innovation Center in Zhejiang University, State Key Laboratory of Component-Based Chinese Medicine, Hangzhou, China
⁵Westlake Laboratory of Life Sciences and Biomedicine, Hangzhou, China

Correspondence

Xiaohui Fan, Pharmaceutical Informatics Institute, College of Pharmaceutical Sciences, Zhejiang University, Hangzhou, 310058 Zhejiang, China. Email: fanxh@zju.edu.cn

Li Shao, Hong Zhao and Rongfang Guo contributed equally to this work.

Xiaoyan Lu and Xiaohui Fan are the lead contact.

Xiaohui Fan b https://orcid.org/0000-0002-6336-3007

REFERENCES

- 1. Seeger C, Mason WS. Molecular biology of hepatitis B virus infection. Virology. 2015;479-480:672-686.
- 2. Zhang C, Li J, Cheng Y, et al. Gut. 2022;1-15. https://doi.org/10. 1136/gutjnl-2021-325915
- 3. Jo J, Tan AT, Ussher JE, et al. Toll-like receptor 8 agonist and bacteria trigger potent activation of innate immune cells in human liver. PLoS Pathog. 2014;10(6):e1004210.
- 4. Wu W, Li J, Chen F, et al. Circulating Th17 cells frequency is associated with the disease progression in HBV infected patients. J Gastroenterol Hepatol. 2010;25(4):750-757.
- 5. Pyzik M, Rath T, Kuo TT, et al. Hepatic FcRn regulates albumin homeostasis and susceptibility to liver injury. Proc Natl Acad Sci USA. 2017;114(14):E2862-E2871.
- 6. Zhang E, Lu M. Toll-like receptor (TLR)-mediated innate immune responses in the control of hepatitis B virus (HBV) infection. Med Microbiol Immunol. 2015;204(1):11-20.

CLINICAL AND TRANSLATIONAL MEDICINE

- 7. Meierovics A, Yankelevich WJ, Cowley SC. MAIT cells are critical for optimal mucosal immune responses during in vivo pulmonary bacterial infection. Proc Natl Acad Sci U S A. 2013;110(33):E3119-E3128.
- Zheng M, Sun R, Wei H, et al. NK cells help induce anti-8. hepatitis B virus CD8+ T cell immunity in mice. J Immunol. 2016;196(10):4122-4131.
- 9. Li Y, Li S, Duan X, et al. Macrophage phenotypes and hepatitis B virus infection. J Clin Transl Hepatol. 2020;8(4):424-431.
- 10. Shaler CR, Choi J, Rudak PT, et al. MAIT cells launch a rapid, robust and distinct hyperinflammatory response to bacterial superantigens and quickly acquire an anergic phenotype that impedes their cognate antimicrobial function: defining a novel mechanism of superantigen-induced immunopathology and immunosuppression. PLoS Biol. 2017;15(6):e2001930.

SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.