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Integrative genomic analysis of N6-methyladenosine-single nucleotide polymorphisms (m⁶A-SNPs) associated with breast cancer

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ABSTRACT

Due to the important role of N6-methyladenosine (m⁶A) in breast cancer, single nucleotide polymorphisms (SNPs) in genes with m⁶A modification may also be involved in breast cancer pathogenesis. In this study, we used a public genome-wide association study dataset to identify m⁶A-SNPs associated with breast cancer and to further explore their potential functions. We found 113 m⁶A-SNPs associated with breast cancer that reached the genome-wide suggestive threshold (5.0E-05), and 86 m⁶A-SNPs had eQTL signals. Only six genes were differentially expressed between controls and breast cancer cases in GEO datasets (GSE15852, GSE115144, and GSE109169), and the SNPs rs4829 and rs9610915 were located next to the m⁶A modification sites in the 3 UTRs of *TOM1L1* and *MAFF*, respectively. In addition, we found that polyadenylate-binding protein cytoplasmic 1 might have a potential interaction with rs4829 (*TOM1L1*) and rs9610915 (*MAFF*). In summary, these findings indicated that the SNPs rs4829 and rs9610915 are potentially associated with breast cancer because they had eQTL signals, altered gene expression, and were located next to the m⁶A modification sites in the 3 UTRs of their coding genes. However, further studies are still needed to clarify how genetic variation affects the epigenetic modification, m⁶A, and its subsequent functions in the pathogenesis of breast cancer.

1 Introduction

Breast cancer is the most frequently diagnosed cancer and the leading cause of cancer-related death among women worldwide [1,2]. Numerous potential risk loci and susceptible genes have been identified by genome-wide association studies (GWASs) in breast cancer [3,4]. Presently, highpenetrance genes (BRCA1, BRCA2, CDH1, STK11, PTEN, and TP53) and moderate-penetrance genes (ATM, CHEK2, PALB2, and BRIP1) are known to genetically predispose individuals to the development of breast cancer [5]. Recently, a growing number of studies have suggested that genetic variations, especially single nucleotide polymorphisms (SNPs), contribute to breast cancer susceptibility [6,7] because SNPs located in protein-coding regions, as well as multiple SNPs located in noncoding regions, can alter gene expression and participate in epigenetic regulation.

most abundant internal modification of messenger RNA (mRNA) and non-coding RNA, such as microRNA, long non-coding RNA, and circular RNA in eukaryotes [8,9]. Emerging evidence has shown that m⁶A is involved in RNA metabolism and is dynamically regulated by methyltransdemethylases, and binding proteins ferases, [10, 11].Furthermore, it was confirmed that m⁶A plays an important role in breast cancer [12,13]. For example, the m⁶A demethylase FTO promoted breast cancer progression by inhibiting BNIP3¹³, and the HBXIP-elevated methyltransferase METTL3 promoted breast cancer progression by inhibiting the tumor suppressor let-7 g [12]. Recent studies have also suggested that SNPs may affect m⁶A by altering the RNA sequences of the target sites or crucial nucleotides [14] and also contribute to the progression of breast cancer.

N6-methyladenosine (m⁶A) modification is the

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m⁶A; single nucleotide polymorphisms (SNPs); breast cancer; GWAS; m⁶Avar Importantly, m⁶A-SNPs have the potential to regulate gene expression and mRNA stability, consequently leading to disease [15,16]. Yang et al. found that rs2416282 contributed to esophageal squamous cell carcinoma (ESCC) risk by regulating YTHDC2 expression [16]. Recently, another found that the SNP studv rs5746136 affected m⁶A modification and influenced the expression of SOD2 in bladder cancer [15], and some METTL14 SNPs were similarly related to neuroblastoma [17]. Zhuo also reported that WTAP SNPs may be genetic modifiers in hepatoblastoma [18]. Ren et al. reported that the rs8400 AA genotype was correlated with increased expression of ALKBH5 and may be a risk factor for hepatoblastoma in the clinical stage III + IV subgroup [19]. In glioma, studies found the m⁶A-SNPs, rs7766006 (WTAP), rs3738067 (YTHDF2), and rs9939609 (FTO), to increase cancer risk, whereas rs2293595 (YTHDC1), rs3813832 (YTHDC1), and rs8047395 (FTO) reduced the risk [20].

Given that m⁶A-SNPs play a pivotal role in cancer [18,21-23] and the effect of m⁶A -SNPs in breast cancer is unclear, identifying m⁶A -SNPs associated with breast cancer is necessary and could provide a new annotation for the pathogenic mechanism of breast cancer risk loci identified by GWAS. Therefore, this study aimed to identify m⁶A-SNPs associated with breast cancer using a public GWAS database and the m⁶AVar database, and to demonstrate their potential functions.

2 Materials and methods

2.1 Screening for m⁶A -SNPs associated with breast cancer

The m⁶AVar database provides information for the annotation, visualization, and exploration of variants associated with m⁶A [24,25]. In the m⁶AVar database, variants were defined at three confidence levels, namely high confidence level when RNA modification sites were directly obtained from the m⁶A-Label-seq, DART-seq, m-⁶A-REF-seq, or miCLIP experiment; medium confidence level when RNA modification sites were obtained from the MeRIP-Seq or m⁶A-Seal-seq experiment; and low confidence level when RNA modification sites were obtained by prediction of the whole transcriptome.

The intersection of SNPs, which reached the genome-wide suggestive threshold (5.0E-05), between the breast cancer GWAS and m⁶Avar databases was used to explore m⁶A-SNPs associated with breast cancer. In this study, breast cancer GWAS data were downloaded from the website (ftp://ftp.ebi.ac.uk/pub/databases/gwas/summary_statistics/MichailidouK_29059683_GCST004988/), and the m⁶A-SNP list was obtained from the m⁶Avar database (http://rmvar.renlab.org/download.html).

2.2 Expression quantitative trait loci (eQTL) analysis of m⁶A-SNPs

eQTL analysis can be used to evaluate the potential function of m⁶A-SNPs with eQTL signals in transcription regulation, such as altering protein binding, changing motifs, and affecting deoxyribonuclease. Hence, eQTL analysis was performed to investigate the breast cancer-associated m⁶A-SNPs that could affect RNA modification using the HaploReg browser (https://www.encode project.org/software/haploreg/) [26,27].

2.3 Gene Ontology (GO) enrichment analysis

GO enrichment analysis was performed using the Metascape tools (https://metascape.org/gp/index. html#/main/step1). GO biological process, GO molecular function, and GO cellular component were used to classify these genes according their functional annotation [28].

2.4 Differential gene expression analysis

The GSE15852, GSE115144, and GSE109169 datasets were downloaded from NCBI GEO (https:// www.ncbi.nlm.nih.gov/geo/) [29,30]. GEO2R (https://www.ncbi.nlm.nih.gov/geo/geo2r/) was used to identify the differentially expressed genes corresponding to m⁶A-SNPs with eQTL between controls and breast cancer cases. A significance level of 0.05 and $|LogFC| \ge 2$ were used for differential expression analysis.

2.5 Prediction of m⁶A-SNPs affecting RNA modification

To determine whether the m^6A -SNPs affected RNA modification and predicted changes in m^6A modification, we used the sequencebased RNA adenosine methylation site predictor (SRAMP), which is an online m^6A modification prediction tool [31]. Further, the USCS browser (GRCh37/hg19; https://genome.ucsc.edu/) includes abundant assemblies and annotations of the genomes of vertebrates and model organisms. Therefore, we used it to analyze the potential functions of m^6A -SNPs [32,33].

3 Results

3.1 Identification of m⁶A-SNPs associated with breast cancer

To identify m^6A -SNPs associated with breast cancer, the intersection of the GWAS and m^6AVar databases was analyzed. Of the 11,637,797 SNPs and 1,401,814 m^6A -SNPs in the breast cancer GWAS and m^6A Var databases (Figures 1 and 2), we identified 113 m^6A -SNPs, which reached the genome-wide suggestive threshold (5.0E-05), to be associated with breast cancer (Table S1).

To check whether these m⁶A-SNPs have eQTL effects, the HaploReg browser was used, finding



Figure 1. Flow chart of study design and analysis.



Figure 2. Manhattan plot of identified m⁶A-SNPs associated with breast cancer. The Manhattan plot showed – log10(p.value) for each of 17,599 m⁶A-SNPs associated with breast cancer. There were 285 m⁶A-SNPs associated with breast cancer (p < 0.001), and 113 m⁶A-SNPs associated with breast cancer (p < 5.0E-5).

that 86 of these breast cancer-associated m- 6 A-SNPs had eQTL signals. Subsequently, we found that of the 86 m⁶A-SNPs with eQTL signals, 31 belonged to the high confidence category, 20 to the medium confidence category, and 35 to the low confidence category. Additionally, among these 86 m⁶A-SNPs, 72 were loss-of-function SNPs and 14 were gain-of-function SNPs (Table S1).

According to GO enrichment analysis, we that molecular found the functions of these m⁶A-SNPs were associated with rRNA binding, calcium-dependent protein binding, DNA-binding transcription activator activity, and RNA polymerase II-specific binding. In addithe functions of the genes tion, with these m⁶A-SNPs were enriched in various biological processes, such as the activation of protein kinase A activity, mammary gland duct morphogenesis, and autophagosome maturation (Figure 3).

3.2 Six corresponding genes of eight m⁶A-SNPs were differentially expressed in breast cancer

To study whether these m⁶A-SNPs may affect the expression level of their corresponding genes, differential gene expression analysis using GEO2R was performed, finding that only six genes, namely TOM1L1 (rs1802212; rs4829), ESR1 (rs2747655), MAFF (rs2267372; rs9610915), TNS1 (rs1424916), EFEMP2 (rs2234458), and TRIM31 (rs2023472), were differentially expressed in the GSE15852 dataset (p < 0.05) (Table 1). Specifically, TOM1L1 and ESR1 were highly expressed, but MAFF, TNS1, EFEMP2, and TRIM31 were significantly downregulated in breast cancer (Figure 4). Moreover, we found that only TNS1 was differentially expressed in the GSE115144 and GSE109169 datasets (Figure 5). Taken together, these results suggest that eight m⁶A-SNPs (rs1802212, rs4829, rs2747655, rs2267372, rs9610915, rs1424916, rs2234458, and rs2023472) may affect the expression level of their corresponding genes.



Figure 3. GO enrichment analysis was performed to annotate the potential function of genes corresponding to identified m⁶A-SNPs.

Table 1. Six	genes	of eight	m ⁶ A-SNPs	associated	with	breast	cancer	were	differentially	expressed	between	controls	and	breast
cancer in the	e GSE15	852.												

				Gene_				Modification_			
SNP ID	Chr	Position	m6A_ID	Gene	region	eQTL Hit	Confidence_level	function	Pvalue		
rs1802212	17	54961293	RMVar_ID_782932	TOM1L1	3′UTR	32	m6A-Seal-seq:(Medium)	Functional Loss	3.20E-19		
rs2747655	6	152126186	RMVar_ID_1369593	ESR1	3'UTR	3	Prediction:(Low)	Functional Loss	8.62E-14		
rs2267372	22	38202227	RMVar_ID_520365	MAFF	exon	35	MeRIP-seq:(Medium)	Functional Loss	2.49E-10		
rs9610915	22	38215073	RMVar_ID_778207	MAFF	3'UTR	9	MeRIP-seq:(Medium)	Functional Loss	1.05E-08		
rs1424916	2	217874577	RMVar_ID_814993	TNS1	intron	4	m6A-Label-seq:(High)	Functional Loss	3.36E-08		
rs4829	17	54961954	RMVar_ID_781699	TOM1L1	3′UTR	14	miCLIP:(High)	Functional Loss	1.00E-06		
rs2234458	11	65871903	RMVar_ID_919384	EFEMP2	intron	5	Prediction:(Low)	Functional Loss	4.68E-06		
rs2023472	6	30108087	RMVar_ID_1375857	TRIM31	CDS	1	Prediction:(Low)	Functional Gain	1.83E-05		

Abbreviations: m⁶A, N6-methyladenosine; UTR, untranslated region; Chr, chromosome; eQTL, expression quantitative trait loci.



Figure 4. Expression levels of selected genes in GSE15852 dataset. TOM1L1 and ESR1 were highly expressed, but MAFF, TNS1, EFEMP2, and TRIM31 were significantly downregulated in breast cancer.



Figure 5. Expression levels of selected genes in GSE115144 and GSE109169 dataset, and we found only *TNS1* was differentially expressed in GSE115144 (a) and GSE109169 (b) datasets.

3.3 TOM1L1 (rs4829) and *MAFF* (rs9610915) may affect m⁶A modification in breast cancer

To investigate the possible m⁶A methylation sites of *TOM1L1* (rs1802212; rs4829), *MAFF* (rs2267372; rs9610915), and *TNS1* (rs1424916), their transcript sequences were predicted at the SRAMP website.

The results showed that five m^6A -SNPs belonged to the high or medium confidence category, including rs1802212 (*TOM1L1*), rs4829 (*TOM1L1*), rs2267372 (*MAFF*), rs9610915 (*MAFF*), and rs1424916 (*TNS1*) (Table 1). Based on the SRAMP website, we found moderately-highly convincible m^6A -modified predicted peaks near TOM1L1 (rs4829) and a moderately convincible m⁶A-modified predicted peak near MAFF (rs9610915) (Figure 6).

We next used the USCS browser to analyze the potential functions of the m⁶A-SNPs. An integrative analysis of the potential functions of rs4829 and rs9610915 is shown in Figure 7. The SNP rs4829 was located in the 3'UTR of *TOM1L1* on chromosome 17 and rs9610915 was located in the 3'UTR of *MAFF* on chromosome 22 (Figure 7). This finding suggested that these two SNPs may affect the m6A modification site and regulate gene

expression to participate in breast cancer pathogenesis.

4 Discussion

 m^6A plays an important role in breast cancer, but the effect of m^6A -SNPs in breast cancer is unclear. Identifying m^6A -SNPs associated with breast cancer will provide a new annotation for the pathogenic mechanism of breast cancer risk loci identified by GWAS; therefore, we identified m^6A -SNPs associated with breast



#	Position	Sequence context	Score(binary)	Score(knn)	Score(spectrum)	Score(combined)	Decision
1	60	GTGCC AACTC TCTAA AACGT AGACT CTGTG CAGCT TTGAA GCCTG	0.644	0.592	0.652	0.645	m ⁶ A site (High confidence)
2	87	GTGCA GCTTT GAAGC CTGGA AGACA ATACC TACCA ACATG TCAAA	0.470	0.494	0.717	0.570	m ⁶ A site (Moderate confidence)
3	205	AACAG CGTGA GATTT CAACA GAACT TGTTT GGAAC AAATA CTCAC	0.61	0.778	0.586	0.609	m ⁶ A site (High confidence)



#	Position	Sequence context	Score(binary)	Score(knn)	Score(spectrum)	Score(combined)	Decision
1	171	TCTTG CAGCC CCCCA AACTG GGACC GAATG ACCCT GGGAA GGGGA	0.531	0.54	0.554	0.540	m ⁶ A site (Low confidence)
2	194	CCGAA TGACC CTGGG AAGGG GAACT TGGGT AGGTT GGGGA TGGGG	0.621	0.624	0.522	0.581	m ⁶ A site (Moderate confidence)

Figure 6. The genomic sequence of TOM1L1 and MAFF transcripts were used to predict the m⁶A modification on website (http://www.cuilab.cn/sramp). There were moderately-highly convincible m⁶A-modified predicted peaks near *TOM1L1* (rs4829) and a moderately convincible m⁶A-modified predicted peak near *MAFF* (rs9610915).



Figure 7. Integrative analysis of the potential function of rs4829 and rs9610915 SNPs by querying USCS. The SNP rs4829 was located on the 3'UTR of the *TOM1L1* gene on chromosome 17 and rs9610915 was located on the 3'UTR of the *MAFF* gene on chromosome 22. Further, RIP-chip GeneST from ENCODE/SUNY Albany data suggested that polyadenylate-binding protein cytoplasmic 1 (*PABPC1*) might have a potential interaction with rs4829 (*TOM1L1*) and rs9610915 (MAFF).

cancer and explored their potential functions. We found 113 m⁶A-SNPs that were associated with breast cancer, and 86 m⁶A-SNPs that had eQTL signals.

Additionally, there were 31 m⁶A-SNPs belonging to the high confidence category; further, 75% of the m⁶A-SNPs were distributed in intron regions, 20% were distributed in the 3'UTR region, and 5% were distributed in the 5'UTR region. However, there were no SNPs of m⁶A regulators (*METTL3*, *METTL14*, *METTL16*, *WTAP*, *VIRMA*, *RBM15*, *FTO*, *ALKBH5*, *YTHDC1*, *YTHDC2*, *YTHDF1*, *YTHDF2*, *YTHDF3*, *IGF2BP1*, *IGF2BP2*, *IGF2BP3*, *HNRNPA2B1*, and *EIF3A*) in the m⁶A-SNPs associated with breast cancer (Table S1). This finding indicated that SNPs of these 18 m⁶A regulators may not affect breast cancer.

We also found that six corresponding genes of eight m⁶A-SNPs were differentially expressed in breast cancer, and the SNPs rs4829 and rs9610915 located were next to the m⁶A modification site in the 3'UTR of TOM1L1 and MAFF, respectively, suggesting that these two SNPs may affect the m⁶A modification site and regulate gene expression to play a role in breast cancer. Studies have shown that TOM1L1 is associated with breast cancer. For example, Chevalier et al. found that TOM1L1 amplification may enhance the metastatic progression of ERBB2-positive breast cancers by regulating the ERBB2-driven proteolytic invasion [34,35]. There is also evidence that MAFF regulates cancer pathogenesis. For instance, Martínez-Hernández et al. found 17 variants of MAFF in chronic myeloid leukemia (CML), and the rs9610915 SNP of MAFF was significantly associated with CML Further, RIP-chip GeneST from the [36]. ENCODE/SUNY Albany data suggested that polyadenylate-binding protein cytoplasmic 1 (*PABPC1*) might have a potential interaction with 2396 👄 Z. XUAN ET AL.

rs4829 (*TOM1L1*) and rs9610915 (*MAFF*) (Figure 5). It is known that PABP1 participates in the initiation of translation and stabilization of mRNA and that *PABPC1* can encode PABP1 protein and promote tumor progression of gliomas and hepatocellular carcinomas [37,38]. In breast cancer, studies have indicated that *PABPC1* mediates *SNHG14*-induced oncogenic effects [39].

5 Conclusion

In this study, we used a comprehensive analysis of GWAS raw data and summary statistics combined with eQTL and differential gene expression analyses to identify breast cancer-associated m-⁶A-SNPs. We found that two SNPs (rs4829 and rs9610915) had eQTL signals, altered gene located expression, and were next to the m⁶A modification sites in the 3'UTRs of their coding genes and that PABPC1 might have a potential interaction with these two SNPs. These findings indicated that the SNPs rs4829 and rs9610915 may be potentially associated with breast cancer. However, the relationship between these m⁶A-SNPs and breast cancer risk and their potential roles in the pathogenesis of breast cancer still require clarification.

Highlights

- 113 m⁶A-SNPs associated with breast cancer (5.0E-05), and 86 m⁶A-SNPs had eQTL signals.
- (2) SNPs rs4829 and rs9610915 had eQTL signals, altered the expression, located next to the m6A 3'UTR modification site of their coding genes.
- (3) SNPs rs4829 and rs9610915 may be potentially associated with breast cancer.
- (4) *PABPC1* might have a potential interaction with SNPs rs4829 and rs9610915.

Disclosure statement

No potential conflict of interest was reported by the author(s).

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Author contributions

GB.Z and P.H designed the research. ZX.X, YW.Z, JY.J, and XW.Z performed the experiments. XP.H, XL.Y and YF.S analysed the data. All authors drafted and reviewed the manuscript.

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