



Metabolic Profile, Bioactivities, and Variations in the Chemical Constituents of Essential Oils of the *Ferula* Genus (Apiaceae)

Priyankaraj Sonigra and Mukesh Meena*[†]

Laboratory of Phytopathology and Microbial Biotechnology, Department of Botany, Mohanlal Sukhadia University, Udaipur, India

The genus Ferula is the third largest and a well-known genus of the Apiaceae family. It is

OPEN ACCESS

Edited by:

Marcello Locatelli, University of Studies G d'Annunzio Chieti and Pescara, Italy

Reviewed by:

Majid Mohammadhosseini, Islamic Azad University, Shahrood, Iran Mohamad Reza Naghavi, University of Tehran, Iran

*Correspondence:

Mukesh Meena mukeshmeenamlsu@gmail.com drmukeshmeena321@mlsu.ac.in

> [†]ORCID: Mukesh Meena

orcid.org/0000-0002-6336-1140

Specialty section:

This article was submitted to Ethnopharmacology, a section of the journal Frontiers in Pharmacology

Received: 17 October 2020 Accepted: 11 December 2020 Published: 12 March 2021

Citation:

Sonigra P and Meena M (2021) Metabolic Profile, Bioactivities, and Variations in the Chemical Constituents of Essential Oils of the Ferula Genus (Apiaceae). Front. Pharmacol. 11:608649. doi: 10.3389/fphar.2020.608649 categorized in the Peucedaneae tribe and Ferulinae subtribe of the Apiaceae family. At present, about 180 Ferula species have been reported. The genus is mainly distributed throughout central and South-West Asia (especially Iran and Afghanistan), the far-East, North India, and the Mediterranean. The genus Ferula is characterized by the presence of oleo-gum-resins (asafoetida, sagapenum, galbanum, and ammoniacum) and their use in natural and conventional pharmaceuticals. The main phytochemicals present in the genus Ferula are as follows: coumarin, coumarin esters, sesquiterpenes, sesquiterpene lactones, monoterpene, monoterpene coumarins, prenylated coumarins, sulfur-containing compounds, phytoestrogen, flavonoids and carbohydrates. This genus is considered to be a valuable group of medicinal plants due to its many different biological and pharmacological uses as volatile oils (essential oils). Numerous biological activities are shown by the chemical components of the essential oils obtained from different Ferula species. Because this genus includes many bioactivities such as antimicrobial, insecticidal, antioxidant, cytotoxic, etc., researchers are now focusing on this genus. Several reviews are already available on this particular genus, including information about the importance and the uses of all the phytochemicals found in the species of Ferula. Despite this, no review that specifically provides information about the biological activities of Ferula-derived essential oils, has been published yet. Therefore, the present review has been conducted to provide important information about the chemical profile, factors affecting the chemical composition, and biological activities of essential oils of the Ferula species.

Keywords: antimicrobial, antioxidant, cytotoxic, essential oils, Ferula species, phytochemicals

INTRODUCTION

Medicinal plants are considered to be an invaluable and a constant source of biologically active phytochemicals. Amid all the other medicinal plants, spices are an important element of Ayurveda. Instead of being an irreplaceable part of food accessories, they have their history in the field of folk medicine since they possess different phytochemicals (Amalraj and Gopi, 2017). These phytochemicals have long been used to cure several ailments, and they can be a promising alternative to conventional medical therapies (Upadhyay, 2017). The Apiaceae family consists of about 455 genera and 3,600–3,751 plants species that belong to this family are often used in the form

1

of spices. *Ferula* is the 3rd largest genus of the Apiaceae family and is categorized in the Peucedaneae tribe and Ferulinae subtribe of the family. At present, about 180 *Ferula* species have been reported. *Ferula* is a Latin word meaning "vehicle" or "carrier" (Iranshahy and Iranshahi, 2011). The genus has a wide distribution all over central and South–West Asia (especially Iran and Afghanistan), the far-East, North India, and the Mediterranean (Hosseinzadeh et al., 2020a; Hosseinzadeh et al., 2020b), and some are distributed in desert areas. Most of the *Ferula* species grow in mountainous regions and arid climates (Yaqoob and Nawchoo, 2016). The *Ferula* species has been of great importance in folk and traditional medicine for more than a 1,000 years. The genus *Ferula* is characterized by the presence of oleo-gum-resins (asafoetida, sagapenum, galbanum, and ammoniacum) (Ahmadi et al., 2020).

The main chemical constituents present in the genus Ferula are as follows: coumarin (ferulenol, galbanic acid and umbelliprenin), coumarin esters (ferulone A, B), sesquiterpenes (germacranes, himachalanes, carotanes, humulanes, guaianes, daucane esters farnesiferol A and B, and sinkiangenorin C and E) (Zellagui et al., 2012), sesquiterpene lactones, monoterpene (α-pinene, β -pinene), monoterpene coumarins (auraptene), prenylated coumarins (ferprenin), sulfur-containing derivatives, phytoestrogen (ferutinin), flavonoids, carbohydrates (galactose, glucuronic acid, arabinose, rhamnose) (Iranshahi et al., 2018; Mohammadhosseini et al., 2019a; Salehi et al., 2019). Generally, aromatic acid lactones sesquiterpenes, coumarins, and sesquiterpene coumarins are present in the roots of Ferula species (Teng et al., 2013), whereas monoterpenes, oxygenated monoterpenoids sesquiterpenes and oxygenated sesquiterpenoid are the main chemical constituents of essential oil present in the aerial parts of Ferula (Mohammadhosseini et al., 2015). Due to the widespread therapeutic effects of this genus, it is being used in folk medicine to treat a variety of diseases and disorders, including skin infections, psychiatric disorders (especially seizure), hyperlipidemia, diabetes, arteries sclerosis, digestive disorders (dysentery), osteoporosis, arthritis, HIV, influenza type A, cancers (uterine cancer), muscle relaxant, rheumatism, headaches, hypertension, toothache and dizziness (Zhou et al., 2017; Meena et al., 2018; Arjmand and Dastan, 2020; Esmaeili et al., 2020).

In the last few decades, research on the phytochemicals of the Ferula species have gained momentum due to their natural origin, effectiveness and low to no unpleasant side effects. In recent years, several biological activities of phytochemicals, isolated from various Ferula species found in different geographical regions, have been reported, such as antimicrobial (Utegenova et al., 2018; Kahraman et al., 2019), insecticidal (Liu et al., 2020; Pavela et al., 2020), aphicidal (Stepanycheva et al., 2012), antihelmintic (Tavassoli et al., 2018), antiprotozoal activity (Bashir et al., 2014; Amin et al., 2016), antimycobacterial (Fallah et al., 2015), antiviral (Zhai et al., 2012; Ghannadi et al., 2014), antioxidant (Deveci et al., 2018; Rahali et al., 2019), anticancer (Asemani et al., 2018; Hosseinzadeh et al., 2020a; Hosseinzadeh et al., 2020b), antitumor (Alizadeh et al., 2018), cytotoxic (Iranshahy et al., 2019; Mahaki et al., 2019), antiproliferative (Verma et al., 2019), acetylcholinesterase inhibitory (Deveci et al., 2018; Karakaya et al., 2019a), antidepressant (Mohammadhosseini, 2016), antiulcer (Bagheri et al., 2018), muscarinic receptors inhibitory (Khazdair et al., 2015; Gholamnezhad et al., 2018), antihypertensive (Safaeian et al., 2015), anti-epileptic (Kiasalari et al., 2013), antispasmodic (Pavlović et al., 2012), antinociceptive (Bagheri et al., 2014a), phytotoxic (Dastan et al., 2014), hypnotic (Abbasnia and Aeinfar, 2016), antihemolytic and antioxidant (Nabavi et al., 2011), anticoagulant (Han et al., 2010), anticonvulsant (Bagheri at al., 2010; Bagheri et al., 2014a; Bagheri et al., 2014b; Bagheri et al., 2014c) relaxant (Bayrami et al., 2013; Bagheri et al., 2014a; Bagheri et al., 2014b; Bagheri et al., 2014c), memory enhancement (Upadhyay, 2017), increasing digestive enzyme activity (Safari et al., 2019), antigenotoxic (Ozkan et al., 2014; Rezaee et al., 2014), antihyperlipidemic (Yusufoglu et al., 2015; Latifi et al., 2019), antihyperglycemic (Iranshahi and Alizadeh, 2012; Yusufoglu et al., 2015), antidiabetic (Yarizade et al., 2017; Latifi et al., 2019), anxiolytics (Upadhyay et al., 2014; Batra et al., 2020) and antihepatotoxicity (Fatima et al., 2017; Deniz et al., 2019). In these activities, essential oils also make up a minimal but significant share.

Essential oils are defined as volatile aromatic compounds that provide a distinctive flavor, aroma, or scent to a plant (Raut and Karuppayil, 2014; Meena et al., 2017a; Chandran et al., 2020a, Chandran et al., 2020b). They are hydrophobic, lipid-soluble liquid comprised of volatile and non-volatile fractions (Hussain et al., 2008). The volatile fraction includes mono and sesquiterpene components, their oxygenated derivatives, alcohols, aliphatic aldehydes, and esters. On the other hand, non-volatile residues contain carotenoids, flavonoids, fatty acids, and waxes (Aziz et al., 2018). Essential oils are the byproducts of plants' secondary metabolic processes. Essential oils can be synthesized by any of the plant organs such as leaves, buds, flowers stems, fruits, seeds, wood or bark and roots, and are stored in epidermis cells, secretory cells, glandular hairs and plant-cell wall in the form of small droplets (Prakash et al., 2015; Barupal et al., 2019; Mohammadhosseini et al., 2019b; Chandran et al., 2020a). Generally, Ferula species found in warm to temperate climates such as tropical and Mediterranean countries possess essential oils. Ferula deputize an important part of the conventional pharmacopeia of those countries (Bakkali et al., 2008; Meena and Samal, 2019). In this review, we intend to cover different biological activities and clinical applications of essential oils of the Ferula genus described in recent years. Although, many reviews are available on the medicinal and bioactivity of the Ferula species, no review which specifically provides detailed and encompassing information about the biological activities and pharmaceutical applications of various essential oils has been published yet.

PHYTOCHEMISTRY OF ESSENTIAL OILS AND FACTORS AFFECTING THE CHEMICAL CONSTITUENTS OF THE ESSENTIAL OILS

Essential oils can be considered as liquids that are lighter than water and represent remarkable hydrophobic characteristics involving a

vast number of valuable natural compounds (Mohammadhosseini et al., 2019b). These secondary metabolites can be extracted from different parts of the plant materials using a variety of classical and advanced methods (Mohammadhosseini, 2017). Approximately 160 chemical compounds have been identified in the essential oils of the Ferula species; these chemical compounds are responsible for all biological activities represented by the essential oils and make them a better choice for industrial purposes (Figure 1). These chemical groups are monoterpene hydrocarbons: limonene, myrcene, γ -terpinene, p-cymene, δ -3-carene, α -pinene, and β -pinene (Znati et al., 2012; Ben et al., 2016; Khalifaev et al., 2018; Malekzadeh et al., 2018; Asilbekova et al., 2019; Karakava et al., 2019b; Meena and Swapnil, 2019; Tabari et al., 2019, Topdas et al., 2020); oxygenated monoterpenoids: α -terpinyl acetate, linalool, sabinene, α -terpineol, verbenone, neryl acetate and arcurcumene (Znati et al., 2012; Radulović et al., 2013; Essid et al., 2015; Khoury et al., 2018), sesquiterpene hydrocarbons: germacrene B and D, β -caryophyllene (E)-caryophyllene, bicyclogermacrene, α gurjunene, y-elemene, y-cadinene and δ -cadinene (Zellagui et al., 2012; Essid et al., 2015; Meena et al., 2017b; Topdas et al., 2020; Ahmadi et al., 2020); oxygenated sesquiterpenoids: α-cadinol, caryophyllene oxide, guaiol, α -eudesmol (Z)-ocimenone (E)nerolidol (E)-ocimenone, viridiflorol, carotol, epi-α-muurolol, hinesol, valerianol and spathulenol (Benchabane et al., 2012; Kasaian et al., 2016; Deveci et al., 2018; Akhzari and Saadatfar, 2019; Baccari et al., 2020; Meena et al., 2020a, Meena et al., 2020b) and sulfur-containing metabolites: dimethyl-trisulphide (E)-1propenylsec-butyl disulfide, sec-butyl-(Z)-propenyl-disulphide, disec-butyl-disulphide,phenol 2-methyl-5-(1-methylethyl), sec-butyl-(E)-propenyl-disulphide, 2,5-diethylthiophene, trimethylthiophene and 1-methylpropyl-(1E)-prop-1-en-1-yl-disulfide,bis-[(1methylthio) propyl]-disulfide and 1-methylpropyl-(Z)-prop-1-en-1-yl-disulfide (Kavoosi et al., 2013; Kasaian et al., 2016; Oüzek et al., 2017; Hassanabadi et al., 2019) (Table 1). The chemical profile of the essential oil (aerial parts) of Ferula orientalis L. was evaluated by Karakaya et al., (2019a). Pinene (α : 75.9% and β : 3.4%) was reported as an abundant constituent of the essential oil. Azarnivand et al., (2011) reported the variation in the chemical profile and yield of the essential oil extracted from the dry and fresh aerial parts of Ferula ovina (Boiss.) Boiss. In accordance with the research results, the amount of oil in the fresh aerial parts was higher than the dried aerial plant parts; the percentage of essential oil in the dry and fresh part of F. ovina was observed as 0.4% and 0.25%, respectively. The chemical profile of the essential oil in fresh (limonene, α -pinene, β -myrcene, *cis*- β -ocimene, isosylvestrene, β -pinene) and dried aerial parts spathulenol, germacrene D, β -caryophyllene, $(\alpha$ -pinene, α -terpineol and caryophyllene oxide) of *F. ovina* were also varied. In a report, it was observed that when F. ovina is consumed in its fresh form, it is poisonous while it is safe in dry form. The reason behind this was the presence of a high percentage of β -myrcene and limonene in new parts (Azarnivand et al., 2011). The environmental condition, soil texture, altitude, temperature, and precipitation rate also affect the oil content in Ferula species (Figure 2). Moghaddam and Farhadi (2015) revealed the correlation between mean annual temperatures and altitude; moreover, the effect of altitude on the yield and composition of essential oil was also studied. The maximum accumulations of essential oil and the

presence of a higher number of sulfur compounds in essential oil were found in *Ferula assa-foetida* L. The study of chemical profiles of nine samples revealed that (E)-propenyl sec-butyl disulfide (37–54%) was the abundant compound with (Z)-propenyl secbutyl disulfide (12–23%) in the essential oils (Hassanabadi et al., 2019). A study showed that decreasing temperature and increasing altitude had a negative correlation with the content of essential oils. In a cluster analysis, it was also concluded that genetic factors have had a greater effect on the chemical constituents of essential oil of *F. assa-foetida* L. compared to the environmental factor. The quality, quantity, and chemical profile of essential oils also differed in accordance with the methods of extraction (Mohammadhosseini and Nekoei, 2014; Kasaian et al., 2016).

The most commonly used essential oil extraction methods are steam distillation and hydro-distillation which are archaic methods that have been around for a long time (Mohammadhosseini, 2016). However, since the late twentieth century, microwave methods, such as solvent-free microwave extraction along with microwave-assisted hydrodistillation have been used for a faster and more effective extraction of plant essential oils (Mohammadhosseini and Nekoei, 2014; Meena et al., 2016; Meena et al., 2017c; Mohammadhosseini et al., 2017; Ragab et al., 2019). Akhzari and Saadatfar (2019) demonstrated the effect of stress conditions on the phytochemistry of essential oil of Ferula haussknechtii H. Wolff ex Rech. f. The result showed that the addition of lead nitrate (2 mM) decreased the myrcene, α -, and β -pinene concentration in the essential oil. The relation between the time of material collection (months) and the chemical composition of essential oil obtained from the oleo-gum-resin of F. assa-foetida, was evaluated by Kavoosi and Rowshan (2013). The oleo-gum-resins (OGRs) collected in three different collection times namely OGR1, OGR2, and OGR3 showed different essential oil chemical compositions (E)-1-propenyl sec-butyl disulfide (23.9%) and 10-epi-c-eudesmol (15.1%) were the major constituents in OGR1; whereas (Z)-1-propenyl sec-butyl disulfide and (E)-1-propenyl sec-butyl disulfide with a 27.7% and 20.3% quantity in OGR3; β-pinene (47.1%) and α -pinene (21.3%) were the major constituents in OGR2, respectively. Moreover, Karimian et al., (2020) observed a relationship between the collection time and the essential oil composition and quantity. The OGRs collected in July, August, September, and October showed 9.1%, 8.2%, 7.8%, and 7.4% oil content, respectively. The chemical compositions of the essential oil also varied in different age groups of plants. The essential oil of the young Ferula assa-foetida H. Karst. plant showed a higher amount of sulfur-containing and sesquiterpene compounds, whereas the essential oil of older plants displayed monoterpene (camphene) as the main chemical group of the essential oil profile (Mohammadi et al., 2019). Khalifaev et al., (2018) revealed the chemical profile of the Ferula kuhistanica Korovin essential oil using plants from two different locations of the central part of Tajikistan. The gas chromatography-flame ionization detector and gas chromatography-tandem mass spectrometry analysis of the hydrodistilled root oil, showed that monoterpene hydrocarbons (86.7%) had a greater proportion in oil; a-pinene (57.7-70.6%) was an abundant



monoterpene hydrocarbon, and others include β -pinene, β phellandrene and myrcene. Sagyndykova et al., (2019) identified 47 chemical components in the Ferula foetida (Bunge) Regel essential oil, using the GC-MS analysis. The samples were taken from two different populations (Tuyesu sands and Tynymbay Shoky hills) of the Mangyshlak peninsula which grew in two different types of soil (loamy and sandy). Both of the aforementioned populations were found to have respectable types of chemical components which were as follows; 2,5-dimethyl-2,5-dimethyl-thiophene, 3,4-dimethylthiophene, guaiol, myristicine, bulnesol, α -pinene, carvophyllene oxide, 2,5-dipropylthiophene, elemicine, 1heptatriacontanol, β -trans-caryophyllene, β -cis-caryophyllene, α -caryophyllene, β -pinene, dimethyl trisulfide, α -eudesmol, β -eudesmol, β -eudesmene and 2-ethylthieno [3.2-b]thiophene, 5,5-dimethyl-4-[(1E)-3-methyl-1,3-butadienyl]-1-oxaspiro [2.5] octane, S-9-thiabicyclo [3.3.1]non-6-en-2-yl. Asilbekova et al., (2019) assessed the chemical constituents of Ferula kuhistanica's essential oil with their enantiomeric excesses. The results of enantioselective and gas chromatography-tandem mass spectrometry analysis of the essential oil showed compounds with enantiomeric excesses as follows: $(+/-)-\alpha$ -pinene (61.9/ 38.1), (+/-)- β -pinene (28.6/71.4), (+/-)-sabinene (13.2/86.8), and (+/-)-limonene (82.7/17.3). The results of the study on the essential oils of Ferula aucheri Boiss. Piwczynski, Spalik, Panahi & Puchalka revealed the abundance of M. sesquiterpene hydrocarbons. The main components of flowering tops oil were germacrene B and β -caryophyllene

(14.96 and 12.87%, respectively) and neophytadiene (diterpene compound, 0.18%), whereas cis-dihydroagarofuran and δ -cadinene (9.02 and 8.28%, respectively); δ -cadinene and gurjunene were found in fruit and root oils (18.25 and 12.62%, respectively) (Ahmadi et al., 2020). In a study, Kasaian et al., (2016) reported an abundance of oxygenated sesquiterpenes (74.7%) and S-containing hydrocarbons (16.6%) in the hydrodistilled essential oil of Ferula alliacea Boiss. The major components were 10-epi-y-eudesmol, valerianol, hinesol, guaiol, and Z-propenyl-sec-butyl trisulphide with 22.3, 12.5, 8.3, 7.3, and 6.5% quantities, respectively. Malekzadeh et al., (2018) assessed the difference in yield and chemical components of the Ferula gummosa Boiss. Essential oils were growing in different types of bioclimates. The galbanum dry weight vs. essential oil amount was found to be the highest in the plants collected from Ebrahim Abad (16.9%) and the lowest in the plants collected from angouran (11%). F. gummosa oil contained a higher number of monoterpenes hydrocarbons such as 17–56.55% of α -pinene, 10.44–37.04% of β -pinene, 9.16–10.75% of δ -3-carene, and 0-13.23% of Limonene, moreover, a chemo-variation was observed in the oils. Kouvakhi et al., (2008) described that the plant samples (Ferula gummosa) collected from the tropical zone had a greater number and better type of flavoring combinations of essential oils as compared to the samples collected from the cold region. The samples collected from the same region, but at different altitudes could affect the chemical profile of a plant's essential oil and reflects the efficacy of environmental effects. Moreover, the samples collected from the different regions and at









Monoterpene hydrocarbons

·ΟΗ

 CH_2

`CH₃

 CH_3

H₃C Myrcene CH₃ H₃C CH₂

`CH₃

 CH_2

ĊH₂

H₃C p-Cymene ÇH₃ CH₃ ĊH₃

CH₃



Frontiers in Pharmacology | www.frontiersin.org

Ferula genus.

H₃Ç

H₃C

α-Pinene

Sabinene

H₃C

H₃C

Limonene



δ-3-Carene



Linalool

a-Terpineol

a-Terpinyl acetate

Verbenone (Continued on following page) Metabolic Profile and Bioactivities of Essential Oils of Ferula genus



QН

, _CH3

`CH₃

Metabolic Profile and Bioactivities of Essential Oils of Ferula genus

TABLE 1 (*Continued*) Chemical structures of some monoterpene hydrocarbons, oxygenated monoterpenoids, sesquiterpenes hydrocarbons, oxygenated sesquiterpenoids, sulfur containing compounds present in the essential of *Ferula* genus.



Neryl acetate

β-Caryophyllene

δ-Cadinene

Oxygenated monoterpenoids



CH₃ CH₃

Germacrene B



CH₃ H CH₃ H CH₃ CH₃ CH₃ CH₃ CH₃ CH₃

α-Gurjunene



Germacrene D

Bicyclogermacrene



H₂C H₃C CH₂ H₃C CH₃

γ-Cadinene

γ-Elemene (Continued on following page) TABLE 1 (*Continued*) Chemical structures of some monoterpene hydrocarbons, oxygenated monoterpenoids, sesquiterpenes hydrocarbons, oxygenated sesquiterpenoids, sulfur containing compounds present in the essential of *Ferula* genus.



(s)-Ar-curcumene

Sesquiterpene hydrocarbons







Caryophyllene oxide



a-Eudesmol





Guaiol





(Continued on following page)

(Z)-Ocimenone

(E)-Ocimenone





H₂C

H III

H₃C

H

Ч₃Н

H₃C −CH₃ OH H₃C H₃Ċ



Carotol

(E)-Nerolidol





Valerianol

Viridiflorol

Spathulenol

Epi-α-muurolol



. "″″И

CH₃

Hinesol

HO

March 2021 | Volume 11 | Article 608649

TABLE 1 (*Continued*) Chemical structures of some monoterpene hydrocarbons, oxygenated monoterpenoids, sesquiterpenes hydrocarbons, oxygenated sesquiterpenoids, sulfur containing compounds present in the essential of *Ferula* genus.

Oxygenated sesquiterpenoids



(E)-1-Propenyl-sec-butyl disulfide





2,5-Diethylthiophene

Trimethylthiophene
Sulfur-containing compounds





Dimethyl-trisulfide



(Z)-1-Propenyl sec-butyl disulfide CH₃ H₃C CH₃ CH₃ CH₃

Di-sec-butyl-disulfide

Bis-[(1-methylthio) propyl]-disulfide



the same altitude also showed variation in phytochemicals. Using an environmental metabolomics approach, the effect of edaphic factors (pH, texture, and iron, aluminum, and potassium content) and environmental factors (temperature latitude, altitude and longitude) were determined using the essential oils from the roots of 10 Iranian F. assa-foetida. Three different types of plant chemotypes were characterized with different major chemical compounds in their essential oils. A chemotype (I) possessed Z-1-propenyl sec-butyl disulfide and monoterpenes; α -agarofuran and eudesmane sesquiterpenoids by chemotype (II); and Z-1-propenyl sec-butyl and E-1propenyl sec-butyl by chemotype (III) Karimi et al., (2020). Fourteen Iranian and Afghan F. assa-foetida L. were investigated for their essential oil composition hydrodistilled from oleo gum-resin using a Clevenger type apparatus; a range of variations among 42 compounds was reported. The major constituent was (E)-1-propenyl sec-butyl disulfide (13.66–49.35%), β -pinene (Z)-1-propenyl sec-butyl disulfide, α -pinene, thiophene, and thiourea. The results of the analysis showed that the increase in altitude caused an increase in β -pinene and (Z)-1-propenyl sec-butyl disulfide content, but a reduction in the thiourea content in essential oils. This study showed that altitude was the environmental factor exerting the greatest effect and which caused a great number of variations in the essential oils' chemical components and yield (Hassanabadi et al., 2019). Ferula has many uses in ethnobotany due to its vast phytochemistry but the use of its essential oil has not been seen so much in folk and traditional medicine. Nevertheless, the volatile oils present in the gum of Ferula assa-foetida are released from the body through the lungs, thus, its essential oil is an excellent treatment for asthma. Apart from this, it is also very useful in discharge, breathing, flatus, and gastric erosions (Mahendra and Bisht, 2012).

BIOACTIVITIES OF THE ESSENTIAL OILS FROM THE FERULA SPECIES

The essential oil extracted from the *Ferula* species display different types of biological activities. These activities are related to the chemical group present in the essential oils. These bioactivities could be connected to a single compound or group of chemical compounds found in the essential oil. All-important biological activities and major chemical constituents of the essential oils from different species of the *Ferula* genus are shown in **Table 2**. These bioactivities of essential oils are discussed below.

INSECTICIDAL ACTIVITY

According to the obtained results, it was concluded that terpenes had active insecticidal properties, particularly as fumigants. Baccari et al., (2020) reported that Ferula tunetana Pomel ex Batt. Essential oils had insecticidal activity against Tribolium castaneum Herbst. The essential oils used as fumigants showed 93% repellency and toxicity against T. castaneum at 161.89 µL L/L LD₅₀ value. Bagheri and Rahimi (2014) observed that the Ferula assa-foetida H.Karst. essential oil had a valuable effect on the mortality rate of the black bean aphid at 1% probability level. The highest mortality rate (13.5) in aphids was reported at $300 \,\mu\text{L/ml}$ (leaf essential oil) and 500 µL/ml (seed essential oil) oil concentration, compared to the control (1.5). Furthermore, Goldansaz et al., (2012) evaluated the insecticidal activity of F. assa-foetida essential oil against carob moth (Ectomyelois ceratoniae) a polyphagous pest of pomegranate. All concentrations used in the experiment (oil: solvent, 1:1, 1:3, and 1:5), showed significant inhibition of insect with p < p

TABLE 2 | Tested living system, different bioactivities and major chemical components of the essential oil from different species of Ferula genus growing in different countries.

Plant name (province)	Used plant part (extraction method); essential oils major components	Activity	Tested living system	Result	References
<i>Ferula assa-foetida</i> L. (Iran)	Gum (HD)	Cytotoxic	Breast cancer 4T1 cells	Result showed that all constituents of essential oil could inhibit 4T1 cell proliferation in time and dose dependent manner	Bagheri et al., (2017a)
	Seeds (soxhlet)	Antinociceptive	Male albino mice	Exhibited valuable antinociceptive activity on acute and chronic pain in mice	Bagheri et al., (2017b)
		Relaxant	Male wistar rats ileum	Seed and assa-foetida essential oils (0.3% and 0.2%) could be significantly decrease ach (10^{-4} M) induced contractions by 35.6 (4.12) and 8 (2.4), <i>p</i> = 0.03; 43% and 12%, <i>p</i> = 0.02, respectively	Bagheri et al., (2014a)
	Gum (HD); 1-(2-methyl-1,3-oxathiolan-2-yl) ethanone, (thiophene, <i>trans</i> -dibenzylideneacetone and (Z)-pro- penyl sec butyl disulfide	Vasodilatory	Rats' thoracic aorta	AEO showed a significant vasodilatory effect which could be endothelium-independent or dependently	Esmaeili et al., (2017)
	Oleo-gum resin (HD)	Vasorelaxant	Rats' thoracic aorta	AEO displayed relaxant effect on the precontracted rings, dose-dependently at 23 μ L/L IC ₅₀ value	Esmaeili et al., (2020)
		Antiprotozoan	<i>Blastocystis</i> sps	The lowest and highest percentage inhibition value at which <i>Blastocystis</i> showed no growth and mul- tiplication were 16 and 40 mg/ml	El Deeb et al., (2012)
	Oleo-gum-resin (HD); OGR1: 10-Epi-c-eudesmol and (E)-1-propenyl sec-butyl disulfide; OGR2: (E)-1-propenyl sec-butyl disulfide (Z)-1-propenyl secbutyl disulfide; OGR3: α -pinene and, β -pinene	Antimicrobial	Gram-negative bacteria: Escherichia coli PTCC 1330 and Salmonella typhi PTCC 1609; foodborne gram-positive bacteria Bacillus subtilis PTCC 1023 and Staphylococcus aureus PTCC 1112; foodborne fungi: Candida albicans PTCC 5027 and Aspergillus niger PTCC 5010	MIC (minimal inhibitory concentration) for gram- negative and positive bacteria and fungi were 0.018–0.058, 0.027–0.107 and 0.028–0.111 mg/ml of essential oil, obtained from OGR3, OGR2 and OGR1, respectively	Kavoosi and Rowshan (2013), Kavoosi et al., (2015)
	Air-dried leaves latex (HD); E-1-propenyl-sec-butyl disulfide, β -pinene and β -ocimene	Scolicidal	Echinococcus granulosus	Results indicated that 10 min exposure of <i>F. assa- foetida</i> essential oil with 60 µg/ml or more concen- tration were killed all protoscolices	Kavoosi et al., (2013)
	Herbaceous plant (HD)	Insecticidal	Ectomyelois ceratoniae	All concentration showed complete inhibition of pest	Goldansaz et al., (2012)
	Dried aerial parts (HD)	Insecticidal	Callosobruchus maculatus	<i>F. assafoetida</i> L (150 uL) showed 100% mortality against adult <i>C. maculatus</i> , respectively	Estekhdami et al., (2020)
	Gum (HD); (E)-1-propenyl sec-butyl disulfide, β -pi- nene. (E)- β -ocimene, (Z)- β -ocimene and α -pinene	Insecticidal	Aphis gossypii	Exhibited appreciable insecticidal activity at 0.04μ L/L LC ₅₀ value and repellant activity at μ L/mL	Koorki et al., (2018)
	Gum (HD)	Insecticidal	Rhyzopertha dominica	Essential oil showed toxicity against <i>R. dominica</i> with 65.71 h persistence	Bahrami et al., (2016)
	Latex (HD)	Insecticidal	Agonoscena pistaciae	LC ₅₀ values of essential oil were obtained 5.62 mg/l	Izadi et al., (2012)
	Foliar parts (HD)	Anti-quorum sensing	Pseudomonas aeruginosa Chromobacterium violaceum	Essential oil caused reduction in pyocyanin, pyoyerdine, elastase, biofilm and homoserin lac- tones (HSL) production in <i>p. aeruginosa</i> and also inhibited violacein production in <i>C. violaceum</i>	Sepahi et al., (2015)
	Gum (HD); (E)-1-propenyl sec-butyl disulfide, (Z)-1- propenyl sec-butyl disulfide and 10-epi- γ -eudesmol	Antibiofilm	Candida albicans, C. dubliniensis, C. krusei	4 $\mu L/\text{ml}$ concentration of tested essential oil completely inhibited biofilm formation	Zomorodian et al., (2018)
	Seeds and oleo-gum-resin (HD); (E)-1-propenyl sec- butyl disulfide and (Z)-1-propenyl sec-butyl disulfide	Antimicrobial	Lactobacillus rhamnosus, Streptococcus sobrinus, S. mutans, S. salivarius and S. sanguis	The -resin oil had strongest antibacterial properties than the seed oil ($p < 0.001$)	Daneshkazemi et al., (2019)
	Resin (HD); guaiol (13.66%) and β -Pinene	Antibacterial	β-Lactamase producing Acinetobacter baumannii	Showed antibacterial effect with 18.75 mg/ml MIC value	Afshar et al., (2016)
	Dried plant materials (HD)	Antifungal	Penicillium digitatum and P. italicum	The essential oils showed growth inhibition along with conidia germination and germ tube elongation inhibition	Zahani and Khaledi (2018)

(Continued on following page)

Sonigra and Meena

Metabolic Profile and Bioactivities of Essential Oils of Ferula genus

TABLE 2 | (Continued) Tested living system, different bioactivities and major chemical components of the essential oil from different species of Ferula genus growing in different countries.

Plant name (province)	Used plant part (extraction method); essential oils major components	Activity	Tested living system	Result	References
<i>Ferula assa-foetida</i> L. (India)	Stem and root (HD); (E) and (Z) sec-butyl propenyl disulfide and methyl 1-(methylthio) propyl	Ovicidal and larvicidal	Larvae of Culex pipiens and Culex restuans	Result showed <i>Culex restuans</i> (LC ₅₀ : 10.1 mg/L) more sensitive to essential oil than <i>Cx. Pipiens</i> . The eggs exposed to essential oils were (55.8%) failed to hatch	Muturi et al., (2018)
	Latex (HD)	Anti-quorum sensing	Chromobacterium violaceum	Essential oil reduced the production of violacein and pyocyanin in <i>C. violaceum</i> and <i>P. aeruginosa</i> , respectively	Khambhala et al., (2016)
	Latex (HD); $\beta\text{-pinene, and 1,2-dithiolane and α-pinene}$	Antifungal and antibacterial	Salmonella typhi, E . coli, S. aureus, B . subtilis, A . niger, and C . albicans	MIC values 90 ± 11, 85 ± 5, 80 ± 12, 125 ± 17, >200, and >200, µg/mL were reported for S. typhi, E. coli, C. albicans, S. aureus, B. subtilis, A. niger, S. aureus, and B. subtilis, respectively.	Kavoosi et al., (2013)
	Gum (HD); pathani: 1-(1- propenylthio) propyl methyl disulfide and (E)-1- propenyl sec butyl disulphide; irani: (E)-1- propenyl sec-butyl disulfide and (Z)-1- propenyl sec-butyl disulfide	Antifungal and antibacterial	Aspergillus niger, A . flavus, A . ochraceus, Fusarium oxy- sporum, P. chrysogenum, S. aureus, Yersinia enteroco- litica, Salmonella paratyphi, S. typhi, Bacillus subtilis, B . cereus, Escherichia coli and Listeria monocytogenes	Essential oil from pathani and irani <i>F. assa-foetida</i> exhibited a good antibacterial (<i>Bacillus subtilis</i> and <i>Escherichia coli</i>) and antifungal (<i>Aspergillus ochra- ceus</i> and <i>Penicillium chrysogenum</i>) activity, respectively	Divya et al., (2014)
	Resin (soxhlet)	Antifungal	Alternaria alternata, A. solani, A. flavus, A. niger, A. wentii, Rhizoctonia spp. Drechslera tetramera, D. hawaiiensis, Fusarium semitectum, F. moniliforme and F. solani: Iso- lated from okra seeds	Essential oil exhibited significant inhibiton at 0.25% of essential oil concentration	Sitara et al., (2018)
Ferula aucheri boiss. Piwc- zynski, spalik, M. Panahi and puchalka (Iran)	Dried flowering tops, fruits, and roots (HD); β -caryophyllene and germacrene B (roots); δ -cadinene (flowering tops); gurjunene and δ -cadinene (fruit)	Antimicrobial	Pseudomonas aeruginosa, E. coli, Staphylococcus epi- dermidis, S. aureus, Bacillus subtilis, K. pneumonia, Shi- gella dysenteriae, Salmonella paratyphi, Proteus vulgaris, Candida albican, Aspergillus brasiliensis and A. niger	Exhibited lower MICs value for Klebsiella pneumo- nia, Bacillus subtilis, Salmonella paratyphi-A sero- type and Shigella dysenteriae compared to gentamicin, whereas fruit and root oils were most efficient against E. coli compared to gentamicin	Ahmadi et al., (2020)
Ferula assafoetida L. (kakan)	Leaves and seeds (HD)	Insecticidal	Black bean aphid	Highest death (13.5) rate in essential oil treated black bean aphid was reported than the con- trol (1.5)	Bagheri and Rahimi (2014
Ferula communis L. (Tunisia)	Aerial parts (HD); β-caryophyllene, β-myrcene, α-pi- nene, α-eudesmol, γ-curcumene, p-menthα-1,5-dien- 8-ol and γ-eudesmol	Antileishmanial	Leishmania infantum and Leishmania (L.) major promastigotes	Caryophyllene had high leishmaniacidal activity against <i>L. major</i> (1.33 \pm 0.52 µg/ml) and <i>L. infantum</i> (1.06 \pm 0.37 µg/ml)	Essid et al., (2015)
	Flowers, leaves, stems, and roots (HD); camphor, β -eudesmol, and α -pinene (flower); β -eudesmol, aeu- desmol and δ -eudesmol (Stems); dillapiole, guaiol and spathulenol (roots); α -eudesmol, deudesmol and β -eudesmol (leaf)	Antibacterial	Pseudomonas aeruginosa	The best results were showed by the essential oil of leaves against <i>P. aeruginosa</i> (MIC value 0.156 mg/ml)	Nguir et al., (2016)
Ferula communis L. (Turkey)	Aerial parts (HD)	Antibacterial	Chryseobacterium indologenes	Result showed the essential oil had a great poten- tial to control <i>C. indologenes</i>	Dadaşoğlu et al.,(2018)
<i>Ferula cupulari</i> s boiss. (Iran)	Flower leaves stem (HD); δ -2-carene (flower); β -ocimene and β -pinene (leaf); δ -3-carene and α -terpinyl isobutyrate (stem)	Antibacterial	Staphylococcus epidermidis, S. aureus, B . subtilis, E . coli, K . pneumonia	Showed 22.75, 5.69 and 2.85 mg/ml MIC value of flower, leaf and stem (except <i>P. aeruginosa</i>) es- sential oils against tested gram-positive bacteria	Alipour et al., (2015)
<i>Ferula elaeochytris</i> korovin (France)	Fruits (HD); $\alpha\text{-pinene},\ \beta\text{-phellandrene}$ and sabinene	Antibacterial and antifungal	S. aureus, E. coli, yeast, C. parapsilosis, C. albicans, Cryptococcus neoformans, T. violaceum, T. souda- nense, T. rubrum, T. mentagrophytes, T. tonsurans, A. spergillus and A. fumigatus	Presented the highest inhibition against Staphylo- coccus aureus Trichophyton species	Khoury et al., (2018)
Ferula ferulaeoides (steud.) korov. (China)	Root (HD)	Insecticidal	Plutella xylostella, Mythimna separate and Musca domestica	Guaiol showed great contact inhibition of <i>Plutella</i> <i>xylostella</i> and <i>Mythimna separate</i> at LD ₅₀ values: 8.9 and 0.07 mg/larva, respectively whereas, fu- migation activity against the <i>M. domestica</i> and <i>M.</i> <i>separata</i> , were observed at LC ₅₀ values of	Liu et al., (2013)
Ferula galbaniflua boiss. and buhse. (Brazil)	Leaves and stems (HD); methyl-8-pimaren-18-oate and ethyl phthalate	Antiprotozoan	Leishmania amazonensis	16.9 μ L/L and 3.5 μ L/L, respectively Essential oil had potent effect on against <i>Leish- mania amazonensis</i> and its IC ₅₀ /24 h value was 54.05–162.25 μ g/ml	Andrade et al., (2016)

Metabolic Profile and Bioactivities of Essential Oils of Ferula genus

Sonigra and Meena

(Continued on following page)

TABLE 2 | (Continued) Tested living system, different bioactivities and major chemical components of the essential oil from different species of Ferula genus growing in different countries.

Plant name (province)	Used plant part (extraction method); essential oils major components	Activity	Tested living system	Result	References
Ferula gummosa boiss. (Iran)	Gum (HD); $\alpha\text{-pinene, carvacrol methyl ether, }\delta\text{-}3\text{-}$ carene and $\beta\text{-phellandrene}$	Cytotoxic	L929 mouse fibroblast cells	The average cell viability of L929 cells were ob- served about 88% at full concentration (50 μ L/m) of the essential oil	Abbaszadegan et al (2015)
	Gum (market product)	Hypoglycemic and hypolipidemic	Male wistar rats	Glucose, triglyceride, cholesterol LDL-C and HDL- C IC ₅₀ values for <i>F. gummosa</i> were recorded as 429.91 ± 46.14 , 105.18 \pm 12.13, 91.02 \pm 11.95, 29.59 \pm 3.76, 42.07 \pm 9.68 (mg/d) respectively	Karimlar et al., (2019)
	Market product	Insecticidal	Bemisia tabaci and Orius albidipennis	For Bernisia tabaci, 0.059 μ L ⁻¹ LD ₅₀ value was observed along with significant effect against <i>O</i> .	Zandi-Sohani et al., (201)
	Gum (HD)	Insecticidal	Callosobruchus maculatus	The essential oil showed 100% death rate at $500 \text{ µl}/l$ after 6 h exposure interval	Hosseinpour et al., (2011
	Root (HD)	Insecticidal	Ephestia kuehniella	Topical application presented drastic reduction in total hemocyte count	Ghasemi et al., (2014)
	Aerial part (HD)	Antibacterial	Enterococcus faecalis, Streptococcus sobrinus, S. sali- varius and S. mutans	The MIC value 1.0125 μ g/ml was reported for <i>E. faecalis</i> and other strains	Nazemisalman et a (2018)
	Resin (market product)	Antibacterial	Staphylococcus aureus	Showed the best antibacterial activity against methicillin resistant <i>S. aureus</i> (MRSA) and methi- cillin sensitive (MSSA) bacterial strains	Mahboubi et al., (2011)
	Resin (HD)	Antibacterial	Pseudomonas aeruginosa	Revealed the significant antibacterial activity of plant essential oils and extracts ($p < 0.001$)	Satarian et al., (2018)
	Resin (HD); $\alpha\text{-}$ pinene, $\beta\text{-pinene}$ and $\delta\text{-}3\text{-}$	Acaricidal	Tetranychus urticae	Essential oil displayed high toxicity on adults and eggs of <i>T. urticae</i> with LC_{50} value 6.52 and 6.98 ul /l , respectively	Fatemikia et al., (2017)
<i>Ferula haussknechtii</i> H. Wolff ex rech. f. (Iran)	Root and aerial parts (HD); camphene, isoverbanol and $\alpha\mbox{-pinene}$	Antibacterial	Bacillus cereus, B . subtilis, B . pumilus, E . coli, S. epi- dermidis, K . pneumonia, S. aureus, E . faecalis and p. aeruginosa	Essential oil exhibited efficient activity against Staphylococcus aureus, Staphylococcus epider- midis and Bacillus pumilus at 7.5 mg/ml concen- tration: 12–18 mm growth inhibition	Arjmand and Dastan (202
<i>Ferula hermonis</i> boiss. (France)	Root and rhizome (HD); $\alpha\text{-bisabolol, }\delta\text{-cadinene}$ and. $\beta\text{-farnesene}$	Antifungal	Aspergillus fumigatus, A. niger, Candida albicans, Micro- sporum gypseum, Penicillium purpurogenum, Saccha- romycescerevisiae andTrichophyton mentagrophytes	Essential oil exhibited the strongest antifungal activity against <i>T. mentagrophytes</i>	Al-Ja'fari et al., (2011)
Ferula heuffelii griseb. Ex heuff. (Serbia)	Underground part (HD); elemicin, phenyl propanoids, myristicin and $\alpha\text{-pinene}$	Antibacterial and anticandidal	Gram-positive bacteria: Staphylococcus aureus, S. epi- dermidis, Micrococcus luteus, Bacillus subtilus and B . cereus; gram-negative bacteria: Salmonella typhimurium, Pseudomonas aeruginosa and Escherichia coli; fungi:	Displayed antimicrobial potential against two strains of <i>Micrococcus luteus</i> , <i>Staphylococcus</i> <i>epidermidis</i> , <i>Micrococcus flavus</i> and <i>Candida albi-</i> <i>cans</i> ; MIC value 13.7, 13.7, 17.6, 21.1, 28.2 and 2.9, <i>cituare micrococcus</i>	Juglal et al., (2002 Pavlović et al., (2012)
<i>Ferula lycia</i> boiss. (Turkey)	$\alpha\text{-pinene, bornyl acetate, limonene and }\beta\text{-Pinene}$	Antibacterial	Candida gladrata, C albicans, C, tropicalis and C, sake Staphylococcus epidermidis, S. aureus, Enterococcus faecalis, E. cloacae, Klebsiella pneumonia, Escherichia coli, Serratia marcescens, Salmonella typhimurium, Pro- teus vulgaris, Haemophilus influenza and Pseudomonas aeruginosa	Haemophilus influenzae ATCC 49247 was reported as the most sensitive bacteria among all tested one with 14 mm inhibition zones	Kose et al., (2010)
<i>Ferula lutea</i> (poir.) maire. (Tunisia)	Roots (HD); delta-3-carene, $\alpha\text{-pinene},\beta\text{-myrcene}$ and $\alpha\text{-phellandrene}$	Cytotoxic	Human colon cells (HT-29 and HCT-116 cells)	The IC ₅₀ values were 26.39 \pm 3.98 $\mu g/ml$ and 81.00 \pm 12.81 $\mu g/ml$ for HT-29 and HCT-116, respectively	Ben et al., (2016)
<i>Ferula lutea</i> (poir.) maire. (France)	Roots (HD); delta-3-carene, alpha-phellandrene, myrcene and $\alpha\mbox{-}\mbox{pinene}$ -	Antibacterial	Staphylococcus epidermidis, S. aureus, Micrococcus luteus, Bacillus cereus, B . subtilus, E . coli, P. aeruginosa, Salmonella typhimurium, Candida albicans, C . glabrata C . tropicalis	The essential oil exhibited antibacterial and anti- candidal activity against <i>E. coli</i> , <i>Salmonella typhi-</i> <i>murium</i> , <i>Candida albicans</i> with MIC value; 39, 78 and 156 mg/ml, respectively	Ben et al., (2016)
	Flower (HD); delta-3-carene, 2, 3, 6-trimethylbenzal-dehyde and $\alpha\text{-pinene}$	Antibacterial and anticandidal	Gram-negative bacteria: Salmonella typhimurium Pseu- domonas aeruginosa and Escherichia coli; gram-positive bacteria: Staphylococcus aureus, S. epidermidis Micro- coccus luteus, Bacillus subtilis and B . cereus;Candida glabrata, C . albicans, C . tropicalis and C . sake	Exhibited antibacterial and anticandidal activity against S. epidermidis, S. aureus and E. coli (MIC = 39 µg/ml); and C. albicans (MIC = 156 µg/ml)	Znati et al., (2012)

TABLE 2 | (Continued) Tested living system, different bioactivities and major chemical components of the essential oil from different species of Ferula genus growing in different countries.

Plant name (province)	Used plant part (extraction method); essential oils major components	Activity		Tested living system	Result	References
Ferula orientalis L. (Turkey)	Stem (soxhlet); α-pinene, terpinolene, <i>o</i> -cymene, β-caryophyllene, limonene and bornyl acetate	Neuroprotective		Cortex neuron cells	Essential oil at 10 ⁻² concentration showed signif- icant neuroprotective activity	Topdas et al., (2020)
	Aerial parts (HD); α and $\beta\text{-Pinene}$	Antimicrobial		Staphylococcus aureus, Escherichia coli, Pseudomonas aeruginosa and Candida albicans	The oil of <i>F. orientalis</i> aerial parts was active against <i>Candida albicans</i> and <i>Staphylococcus aureus</i> strains	Karakaya et al., (2019a)
Ferula ovina (boiss.) boiss. (Iran)	Aerial parts (HD); $\alpha\text{-pinene},$ camphene, sabinene, $\beta\text{-pinene},$ sabinene, myrcene and dehydro-1, 8-cineole	Antinociceptive hyperalgesia	and	Mice and other animals	Essential oil constituents impart both antinocicep- tive and hyperalgesic potentially	Radulović et al., (2013)
	Dried root (HD)	Insecticidal		Sesamia cretica	Each concentration found to have significant effect on the immune ability of Sesamia cretica	Sadeghi et al., (2017)
Ferula persica willd. (Iran)	Gum (HD); alpha-pinene, (Z)-1-propenyl sec-butyl disulfide β-pinene, β-dihydroagarofuran, allo-aroma- dendrene, (Z)-β-ocimene, β-dihydrobenzofuran, α-caryophyllene and (E)-1-propenyl sec-butyl disulfide	Cytotoxic		Vero cell, murine colon carcinoma (CT26) cell lines	The viability was decreased in essential oil treated- Vero cell (0.125 \times 10 ⁻⁹ to 80 µL/ml) and CT26 cell line (0.125 \times 10 ⁻⁹ to 20 µL/ml), dose dependently	Hosseinzadeh et al. (2020a), Hosseinzadeh e al., (2020b)
Ferula tadshikorum pimenov. (Tajikistan)	Underground parts (HD); (Z)-sec-butyl propenyl disul- fide, (E)- sec-butyl propenyl disulfide and (E)-1-pro- penyl 1-(methylthio)propyl disulfide	Cytotoxic		CEM/ADR5000 and CCRF-CEM cell lines	Displayed 142.5 and 21.6 $\mu g/ml,$ IC $_{50}$ values for CEM/ADR5000 and CCRF-CEM cell lines, respectively	Sharopov et al., (2019)
	Underground (HD); (E)-1-propenyl sec-butyl disulfide, (Z)-1-propenyl sec-butyl disulfide, and (E)-1-propenyl 1-(methylthio)propyl disulfide	Antibacterial		Methicillin-resistant Staphylococcus aureus, Escherichia coli	Essential oil was not displayed any effective anti- bacterial activity up to 20 mg/ml concentration	Sharopov et al., (2019)
Ferula tingitana L. (Egypt)	Flowers and leaves (HD); $\alpha\text{-thujene, eudesmol, elemol}$ and cadinol	Antitumor		Hormone-responsive MCF7 breast cells, liver carcinoma (HePG2) cells, and cervical (HeLa) cells	IC_{50} % value for <i>in vitro</i> cytotoxicity of flower and leaves essential oil against liver (HEPG2) carcinoma cell, cervical (HELA) and breast (MCF7), lines were as follows; 4.4, 4.2; 8.6, 10.9 and 6.9, 4.8, µg/mL, respectively	Elghwaji et al., (2017)
	Flower and leaf (HD); terpinolene and α -thujene in leaf and flower derived oil, respectively	Antimicrobial		Bacteria: Staphylococcus aureus, Bacillus subtilis, Strep- tococcus faecalis, Escherichia coli, Pseudomonas aeru- ginosa and Neisseria gonorrhea; fungi: Candida albicans, Aspergillus flavus	Showed most efficient antibacterial activity against <i>Neisseria gonorrhoeae</i> and <i>Bacillus subtilis</i> with 41.9 and 48.3% potency compared to tetracycline	Elghwaji et al., (2017)
<i>Ferula tunetana</i> pomel ex batt. (Tunisia)	Seed oil (HD); α -pinene, myrcene, (Z)- β -ocimene, β -pinene, β -phellandrene and limonene	Antigerminative		Medicago sativa, Triticum aestivum and Lactuca sativa	Germination of seedlings were completely inhibited at 10 mg/5 ml and 20 mg/5 ml concentration	Znati et al., (2017)
	Seed (HD); a-pinene, (Z)- β -ocimene and β -pinene	Antimicrobial		Gram (+) bacteria: Staphyllococcus aureus ATCC 25923 and CIP106510, Bacillus subtilus ATCC 6633, B . cereus ATCC 14579, ATCC 11778 and <i>M</i> . luteus NCIMB 8166; gram (-) bacteria: Escherichia coli ATCC 25922, ATCC 35218, Salmonella typhimurium ATCC 13311, LT2DT104; <i>Pseudomonas aeruginosa</i> ATCC 27853, Candida albicans ATCC 90028 and C . glabrata ATCC 90030	Displayed valuable antimicrobial activity against S. typhimurium LT2 DT104 and B . cereus ATCC 14579 (inhibition zone 16.2–1.0 and 15.8–1.0 mm, respectively)	Znati et al., (2017)
	Flower (HD); $\alpha\text{-Pinene, epi-}\alpha\text{-muurolol, }\beta\text{-chenopodiol}$ and himachalol	Insecticidal		Tribolium castaneum	Essential oils fractions displayed 93% repellent activity	Baccari et al., (2020)
Ferula vesceritensis coss. and durieu ex trab. (Algeria)	Flowers and stems (HD); $\alpha\text{-pinene},\ \beta\text{-pinene},\ elixene$ $\alpha\text{-phellandrene},\ aristolene$ and fenchylacetate	Antibacterial		Pseudomonas aeruginosa, Enterobacter aerogenes, E. coli, S. aureus, Klebsiella pneumonia, Morganella morganii	Showed potent inhibition of tested bacterial strains	Labed-Zouad et al., (2015
Ferula vesceritensis coss. and durieu ex trab. (East of algiers)	Dried leaves (HD); shyobunol, (t-cadinol, $\delta\text{-}3\text{-cadinene},$ $\alpha\text{-cadinol}$ and aristolene	Insecticidal		Sitophilus oryzae	Contact and inhalation methods showed potent insecticidal activity with (LD ₅₀ = 16.4 μ L/ml, LD ₉₀ = 53.45 μ L/ml values	Benchabane (2014)
<i>Ferula vesceritensis</i> coss. and durieu ex trab. (Algeria)	Dried aerial parts (HD); germacrene D, 5, 9-tetrade-cadiyne, $\alpha\text{-bisabolene}$ and farnesene	Antibacterial		Escherichia coli, Staphylococcus aureus and Klebsiella pneumonia	The essential oil presented appreciable growth inhibition of listed bacteria	Zellagui et al., (2012)

0.001. In another study, Estekhdami et al., (2020) also evaluated the insecticidal activity of F. assa-foetida essential oil against Callosobruchus maculatus in an interval of 8, 24, and 48 h, respectively. F. assa-foetida essential oil was used with 0, 30, 60, 90, 120, and 150 µL concentration and a death rate reported at 8, 24, 48 h intervals. The mortality rate was observed to be 100% at a 30 µL concentration. Fumigant toxicity and persistence of the essential oil of F. assa-foetida with two others were investigated against adult insects (Rhyzopertha dominica F.). The result suggests that F. assa-foetida essential oil has more toxicity against R. dominica compared to other tested plants. The halflife of the essential oil of asafoetida was reported to be 65.71 h which is also greater than the others (Bahrami et al., 2016). Hosseinpour et al., (2011) investigated the fumigant insecticidal activity of two plants including galbanum (F. gummosa) essential oils extracted from the gum with a Clevenger apparatus. The fumigant insecticidal activity of essential oils was examined alongside adult Callosobruchus maculatus (1-7 days old) with different concentrations ranging from 7.1 to 57.1 µL/L air, at 27-28°C room temperature, 65-70% relative humidity and in dark conditions. Galbanum oil presented a 47.5%, 80%, and 100% mortality rate at 500 µL/L (air) after 2, 4, and 6 h exposure intervals, respectively.

Liu et al., (2013) evaluated an insecticidal sesquiterpene guaoil, found in the essential oils of many plants. To examine the insecticidal property of the compound, three insect larvae were used, namely Mythimna separata Walker, Plutella xylostella L., and Musca domestica L. An effective contact inhibition was observed in the 4th instar larvae of Mythimna separata and third instar larvae of P. xylostella at 0.07 and 8.9 mg/larva concentrations of guaoil while using the fumigation method, the growth inhibition was observed at 3.5 and 16.9 µL/L guaoil concentration, respectively. Koorki et al., (2018) tested F. assafoetida essential oil along with two other medicinally important plants to find their toxicity against Aphis gossypii Glover which has caused economic losses in tested medicinal plants. Lethal concentrations (LC₅₀) of the essential oil of F. assa-foetida were examined after 12 (9.04 µL/L air) and 24 (4.64 µL/L air) hours whereas at a 10 µL/ml concentration the essential oil showed repellent activity. The chemical composition of the essential oil was also evaluated by GC-MS and the major chemical components were (E)-sec-butyl propenyl disulfide (Z) and (E)- β -ocimene, β and α -pinene. The effect of essential oils from F. assa-foetida L. with some other plant species was studied on the growth and physical fitness of Trichogramma embryophagum (Hartig) and Trichogramma evanescens (West.)-parasitoids on the eggs of carob moths. Essential oils at 877 ppm (LC01) exhibited a significant reduction in longevity, wing normality, development, female's fecundity, the sex ratio of their progeny, and survivorship (Poorjavad et al., 2014).

ANTIQUORUM SENSING AND ANTIBIOFILM ACTIVITY

Sepahi et al., (2015) evaluated the potential of *F. assa-foetida* essential oil to inhibit the quorum sensing in *Pseudomonas*

aeruginosa. The essential oil (25 mg/ml) presented antiquorum activity and fully abolished pyocyanin, pyoyerdine, elastase, biofilm, and homoserine lactone (HSL) production. F. assa-foetida essential oil mediated las system inhibition was observed in Chromobacterium violaceum. Since the violacein production is related to the las system, violacein production inhibition was also observed in F. assa-foetida essential oil treated C. violaceum. The chemical profile of the essential oil extracted from the resin of F. assa-foetida, antibiofilm and antimicrobial activity were also investigated by Zomorodian et al., (2018). The main components of essential oil analyzed by the GC/MS method, contained 21.65% of (E)-sec-butyl propenyl disulfide and 19.5% of (10-epi-y-eudesmol and 10.20% of 2-[(Z)-prop-1-enyl] disulfanyl] butane. The essential oil exhibited partial inhibition (50%) of the biofilm formation in the standard strains of Candida krusei, C. tropicalis and C. albicans at concentrations of 0.06, 0.125, and 0.25 µL/ml, respectively. However, the essential oil showed 100% inhibition of biofilm formation at 4 µL/ml of concentration.

ANTIMICROBIAL ACTIVITY

It has been reported that the essential oils obtained from the root and rhizome of the Ferula hermonis Boiss. had a significant effect on many fungal strains such as dermatophytes namely Trichophyton mentagrophytes (Al-Ja'fari et al., 2011). The main chemical components of the essential oil analyzed by GC-MS and FID, and 13C NMR were α -bisabolol, β -farnesene, and δ -cadinene. The MIC and MFC values of essential oils for T. mentagrophytes were reported to be 8 µg/ml and 10.25 µg/ml for 3, 5-nonadiyne and JB73, respectively. In a study, the antibacterial activity of hydrodistilled essential oil from fresh stems (FS), fresh flowers (FF), and dry stems (DS), dry flowers (DF) of Ferula vesceritensis Coss. & Durieu ex Trab. were demonstrated. The essential oils possessed α and β -pinene, α -phellandrene as abundant chemical constituents along with a minute amount of caryophyllene oxide, aristolene, carotol, and elixene (Kim et al., 2004). Nine strains of foodborne and clinically isolated bacteria including Escherichia coli, Morganella morganii, Klebsiella pneumonia, Pseudomonas aeruginosa, Staphylococcus aureus, and Enterobacter aerogenes were used for the antibacterial activity analysis. The minimum inhibitory concentration (MIC) was reported at a concentration of 128 µg/ml against almost all foodborne pathogens and clinical isolates, whereas for the others the concentration was 16-80 µg/ml (Labed-Zouad et al., 2015). Satarian et al., (2018) also tested the extract and essential oils of Ferula gummosa for their antibacterial activity against clinically isolated from P. aeruginosa. In another study, bactericidal activity of essential oil from F. assa-foetida was reported by Kavoosi et al., (2013) together with some other activity including antioxidant, antiseptic, sedative. antispasmodic, analgesic, and carminative activity. The essential oils comprised pinene (B: 47.1%; a: 21.36%) and 1,2dithiolane (18.6%) as main compounds which showed a minimum inhibitory value at the concentration of 90 \pm 11, 85 ± 5 , 80 ± 12 , 125 ± 17 , >200 and >200 µg/ml against the

tested strains namely Candida albicans, Aspergillus niger, Bacillus subtilis, Staphylococcus aureus, Escherichia coli, and Salmonella *typhi*, respectively. The antifungal and antibacterial activity of the flower oil of Ferula elaeochytris Korovin along with some other plant species was demonstrated against some selected Grampositive, Gram-negative bacteria and fungal strains (Khoury et al., 2018). The essential oil was found to have the most antibacterial activity against the Staphylococcus aureus and dermatophytes (Trichophyton species) with a 8-64 µg/ml MIC value. The F. haussknechtii essential oil, which was extracted by the Clevenger apparatus contained camphene, α -pinene, and isoverbanol as abundant chemical constituents; these compounds exhibited antibacterial activity against nine bacterial strains. The result showed that Bacillus pumilus, Staphylococcus epidermidis and S. aureus were more sensitive to the essential oil among other tested bacterial strains (Arjmand and Dastan, 2020). Alipour et al., (2015) enumerated the bactericidal activity of the essential oil acquired from stem, leaf, and flower parts of the Ferula cupularis Boiss. against some Gram-positive bacteria. The main components that were analyzed by the GC-MS analysis displayed a great variation. The MIC value of flower, leaf, and stem (except P. aeruginosa) oils were 22.75 mg/ml, 5.69 mg/ml, and 2.85 mg/ml against tested Gram-positive bacteria. Ferula gummosa oil showed antibacterial potential against Enterococcus faecalis, Streptococcus sobrinus, S. salivarius, S. mutans with a 1.0125 mg/ml MIC value (Nazemisalman et al., 2018). Divya et al., (2014) investigated the antimicrobial activity of essential oils from two plant varieties (Pathani and Irani) of F. assa-foetida, against various food-borne bacterial and fungal species. Different types of plant varieties also possessed variations in their oil chemical profiles. In accordance with the result, a conclusion was made that the essential oil of the Pathani F. assa-foetida variety exhibited a good antibacterial activity (Escherichia coli and Bacillus subtilis); whereas, the essential oil of Irani the F. assa-foetida variety exhibited good fungicidal activity (Penicillium chrysogenum and Aspergillus ochraceus). Furthermore, a study revealed the antimicrobial properties of the essential oil of the resin and seed of F. assa-foetida against various oral bacteria such as Lactobacillus rhamnosus, Streptococcus salivarius, Streptococcus sanguis, Streptococcus sobrinus, and Streptococcus mutans (Daneshkazemi et al., 2019). The resulting analysis displayed that the oleogum-resin essential oil had significant and effective antibacterial activities as compared to the seed essential oil (p < 0.001). Jahani et al., (2015) reported the antibacterial activity of gelatin nano-capsules formulated from the essential oil of F. assa-foetida. Mallahi et al., (2018) studied the potential of Ferula essential oil to increase the shelf-life of Gerbera jamesonii (Gerbera Daisy) flower by inhibiting the pathogenic bacteria. Ahmadi et al., (2020) tested F. aucheri essential oil for its antimicrobial potential. All aerial parts of the plant used to extract oil had different major chemical components. All essential oils exhibited lower MIC for Salmonella paratyphi-A serotype, Shigella dysenteriae, Klebsiella pneumonia, and Bacillus subtilis but gentamicin exhibit greater MIC than essential oil which was used as a positive control. Additionally, fruit and root oils showed more efficacy against E. coli compared to gentamicin.

Zomorodian et al., (2018) uncovered the relationship between the antimicrobial activity and the presence of (E)-1-propenyl secbutyl disulfide and (Z)-1-propenyl sec-butyl disulfide as the major chemical compounds in the oil. In the same way, many studies have reported the link between antimicrobial activity of the essential oils with a high content of sulfur compounds (Iranshahi et al., 2008; Kavoosi and Rowshan, 2013; Divya et al., 2014). In this regard, Divya et al., (2014) demonstrated the morphological changes in fungal and bacterial cells tested with the F. assa-foetida essential oil using scanning electron microscopy. The morphological changes were like damaged bacterial membranes, irregular branching in fungal hyphae and sporulation inhibition and disruption of the cytoplasmic membrane, which result in leakage of ions and electrolytes. Additionally, a-pinene and 10-epi-y-eudesmol were also reported as active terpenoids against the broad range of microorganisms (Rivas da Silva et al., 2012). Terpenoids are highly lipophilic in nature, so they target cell membranes and cause toxicity through cell membrane integrity disruption (Kovač et al., 2015).

IMMUNOMODULATORY EFFECT

Oüzek et al., (2017) elucidated the chemistry and immunomodulatory activity of the essential oil procured from the dried plant material of the Ferula iliensis Krasn. ex Korovin material by the hydro-distillation method with Clevenger apparatus (Z) and (E)-propenyl sec-butyl disulfide 23.4-45.0% and 15.7-39.4%, respectively were the major chemical constituents of the extracted plant essential oil. On the basis of the results of the experiments, it was concluded that the essential oil had the potential to stimulate [Ca2+] ion mobilization and production of reactive oxygen species in the murine bone marrow phagocytes and human neutrophils. Furthermore, the effect of essential oil could be reversed by using capsazepine a TrpV1 channel antagonist, in a dose-dependent manner. This result indicated that TrpV1 channels were the target site for the essential oil component, which was likely to be (Z)-sec-butyl propenyl disulfide, aver by the molecular modeling method using a known TrpV1 agonist. Sadeghi et al., (2017) studies the effect of F. ovina essential oil on the immune system of Sesamia cretica Ledereer. The results proved that the essential oil had a visual effect on the S. cretica's immune system. Four main circulating hemocytes were identified in the fourth instar larvae which included oenocytoides, granulocytes (GRs), prohemocytes, and plasmatocytes. The 4th instar larvae were injected with 1 uL of each concentration of *F. ovina* oils (1,000, 2,500 and 7,000 ppm). The total number of hemocyte and GR count enhanced with 1,000 ppm concentration while decreased at 2,500 and 7,000 ppm concentration, dose-dependently. However, plasmatocyte numbers declined for all the treatment concentrations but more significantly with increased doses. However, the number of nodules and the phenol-oxidase activity was not affected by any tested essential oil concentration. Schepetkin et al., (2016) substantiated the effect of the Ferula akitschkensis B.Fedtsch. ex Koso-Pol. Essential oils on human neutrophils cells. Seed and

stem oil possessed 4(10)-thujene, α - and β -pinene; 4-terpineol, eremophilene, 2-himalachene-7 β -ol myristicin and (E)-6,10dimethylundeca-5,9-dien-2-one as their primary components. The report analysis showed that the major component of umbels seeds namely β -pinene, 4(10)-thujene, γ -terpinene, (E)-6,10-dimethylundeca-5,9-dien-2-one, pichtosin, and (E)non-2-enal stimulated calcium mobilization in neutrophils cells. Especially, geranylacetone and isobornyl acetate showed great potential with 7.6 \pm 1.9 and 6.4 \pm 1.7 μ M EC₅₀ value, respectively. Additionally, the treatment of neutrophil cells with the aforementioned components except (E)-2-nonenal, resulted in desensitization of the neutrophils due to fMLF (N-formyl-Met-Leu-Phe) and IL-8 (interleukin-8) induced calcium ions flux and inhibition of N-formyl-Met-Leu-Phe -induced chemotaxis in cells. The effect of these components on calcium ions flux in neutrophils could be inhibited by TRP channel blockers (transient receptor potential). A further study averred that geranylacetone was a TrpV1 agonist, and cause Ca²⁺ ions influx in TrpV1-transfected human embryonic kidney 293 cells (HEK293 cells); whereas myristicin was a TrpV1 antagonist, and inhibited fMLF and IL-8 mediated neutrophil $[Ca^{+2}]_i$ flux stimulation and abolished capsaicin (zostrix)induced Ca²⁺ions influx in TrpV1-transfected human embryonic kidney 293 cells.

ANTI-ACETYLCHOLINESTERASE, ANXIOLYTIC AND ANTISPASMODIC ACTIVITY

Ahmadi et al., (2020) unearthed the anti-acetylcholinesterase activity of the essential oils of flowering tops, fruits, and roots of the Ferula aucheri Boiss. Piwczynski, Spalik, M. Panahi & Puchalka with antimicrobial activity. The biochemistry of the oils was analyzed by GC-MS method; AChE inhibitory potential was assessed by Ellman's method with slight modification. The major portion of the chemical constituents of the essential oils belonged to the sesquiterpene hydrocarbon group (61.9%). The essential oil of roots and fruits showed weak acetylcholinesterase (AChE) inhibitory potential with 239.69 \pm 3.5 and 554.05 \pm 4.65 µg/ml half-maximal inhibitory concentration (IC₅₀) values, respectively whereas flowering tops essential oil presented moderate AChE inhibitory activity with 179.06 ± 4.3 µg/ml IC₅₀ values. Znati et al., (2012) also inquired about the anti-acetylcholinesterase and antimicrobial properties of flower oil from Ferula lutea (Poir.) Maire. The chemical compounds of the essential oils were analyzed by GC-MS. The major portion of essential oil chemical constituents was covered with monoterpene hydrocarbon and sulfur-containing compounds. The result showed that the flower oil exhibits significant AChE inhibitory activity with a IC₅₀ value of 70.25 \pm 5.41 µg/ml. Deveci et al., (2018) examined the anticholinesterase activity along with antioxidant and anti-tyrosinase activity of F. elaeochytris Korovin essential oils. The result of GC-MS and GC-FID showed the presence of 21.3% of β -cubebene, 17.5% of caryophyllene oxide, and 14.9% of β -caryophyllene. The antimeasured cholinesterase activity was by Ellman's

spectrophotometric method. The essential oil of F. elaeochytris exhibited the highest anticholinesterase and anti-tyrosinase activities. The anticholinesterase activity of the root's essential oil of Ferula lutea (Poir.) Maire. was also demonstrated by Ben et al., (2016). The constituents of hydrodistilled essential oils were analyzed by 13C-NMR, GC (FID) and GC-MS spectroscopy had a higher portion of monoterpene hydrocarbons viz. delta-car-3ene (72.6%), alpha-pinene, myrcene, and alpha-phellandrene. The results indicated that this essential oil exhibited efficient antiacetylcholinesterase bioactivity with a IC₅₀ value of 28.56 \pm 1.87 µg/ml. The effects of Ferula heuffelii Griseb. ex Heuff. essential oil on contractions (KCl and ACh induced or spontaneous) were examined by Pavlović et al., (2012). The essential oil has the potential to inhibit spontaneous contraction in rat ileum, dose-dependently. The essential oil exerted half of the atropine (positive control) effect at a ED_{50} value (median effective dose) of 86.64 µg/ml. The acetylcholine mediated induction of contractions in ileum was inhibited at 75.00 µg/ml of the essential oil concentration, whereas at 250.00 µg/ml of essential oil concentration, the spasmodic effect of KCl (80 mM) was almost abolished. Sadraei et al., (2001) examined the effect of F. gummosa essential oil with other extracts (methanolic extracts, petrolic hydro-alcoholic, and etheric). F. gummosa essential oil (FGEO) inhibited the response to KCl (80 mM) at 10-360 µg/ml concentration whereas at 180 µg/ml, concentration almost terminated the response to KCl. Two components of essential oil namely α and β -pinene (10 ng/mL-1.3 µg/ml and 2-138 ng/ml, serially) inhibited the tonic contraction induced by KCl in a dosedependent manner. These two components also inhibited the ACh (80 mM) induced contraction. α -Pinene (180 and 90 ng/ml), β -pinene (160, 80, 40, and 20 ng/ml) inhibited the acetylecholine induced contraction up to $45 \pm 9.7\%$, and $95 \pm 1.7\%$ to $79 \pm 7.7\%$; $0.8 \pm 0.8\%$, 11 ± 7.3 , 33 ± 7.3 and $95 \pm 2.3\%$ to $84 \pm 78.9\%$, sequentially (p < 0.05). The relaxant effect of oleo-gum-resin and seed oils of the F. assa-foetida on isolated rat's ileum was investigated by Bagheri et al., (2014a), Bagheri et al., (2014b), and Bagheri et al., (2014c). To reveal the relaxant effect of asafoetida resin and seed essential oil, the isolated ileum of rats treated with three doses, and isotonic contractions of the essential oil. The contractions of the specimen were induced by different doses (0.3, 0.2, and 0.1%) of asafoetida and essential oil. The amplitude of contraction was recorded before and after exposure to acetylcholine (ACh) cumulative concentration. Results showed that asafoetida (0.2% and 0.3% concentration) had an antispasmodic effect on acetylcholine-induced contraction. The essential oil also had effective antispasmodic acetylcholine-induced against contraction activity at concentrations 10^{-12} - 10^{-2} M of seed and asafoetida essential oils (0.2% and 0.3%). These oils could cause a significant reduction in acetylcholine (10⁻⁴ M) induced contractions at (4.12) and 8 (2.4) concentration, p = 0.03; up to 43% and 12%, p = 0.02, respectively. Batra et al., (2020) revealed the presence of triterpenoids with 32 other chemical components in the essential oil of Ferula sumbul Hook. roots. The essential oil (50 µL/kg) showed considerable anxiolytic activity in various tested models (light/dark, mirror chamber, elevated plus maze,

m-CPP-induced and open-field anxiety). The results showed that the anxiolytic effect of the essential oil was mediated primarily through the benzodiazepines site on GABA receptors and through 5-HT receptors.

GENOTOXIC AND ANTIGENOTOXIC PROPERTIES

Ozkan et al., (2014) planned to evaluate the genotoxic and antigenotoxic activities of essential oil extracted from the leaves and flowers of Ferula orientalis L. grown in Erzurum. The chemical constituents of essential oil were characterized by the GC-MS method. α -Cadinol (11.7%), γ -cadinene, germacrene D-4-ol, epi- γ -muurolol (α -pinene 9.3%, 11.9%, 6.1%) were recorded as the main chemical components in leaf (10.45, 8.1, 6.8, 5.9, and 5.7%, respectively) and flower (9.3, 11.9, 6.1, and 7.2%, respectively) essential oils. According to their results, the chemical constituents were responsible for biological activities. Bacterial strains such as Salmonella typhimurium TA1537, S. typhimurium TA1535, and E. coli WP2 uvrA were used to evaluate the mutagenic activity, using the bacterial reverse mutation assay method. The study showed that tested leaf and flower essential oil did not have any mutagenic activity on S. typhimurium and E. coli strains at any used concentration. Nevertheless, the essential oil showed antimutagenic property used mutagen, namely N-methyl-N'-nitro-Nagainst nitrosoguanidine (MNNG), 9-aminoacridine (9-AA), and sodium azide (NaN₃). Further investigations showed the potential of the essential oils to reduce the effect of mutagens on bacterial strains which were as follows: N-methyl-N'-nitro-Nnitrosoguanidine on E. coli WP2 uvrA (23-52%); 9aminoacridine on S. typhimurium TA1537 (40-68%), sodium azide on S. typhimurium TA1535 (29-36%).

SCOLICIDAL ACTIVITY

In a study, Tabari et al., (2019) revealed the effectiveness of F. gummosa's essential oil and its main components against Echinococcus granulosus protoscoleces. Results of GC/MS displayed β -pinene as an abundant chemical compound of the essential oil. Furthermore, the eosin staining method was used to measure mortality rate. The mean death rate of E. granulosus protoscoleces was recorded 100% at 50 µg/ml concentrations of the essential oil and 60 min exposure time. The essential oil of F. gummosa also showed a higher toxic effect on E. granulosus protoscoleces with 50% LC₅₀ values (lethal concentration) 17.18 µg/ml. Additionally, 10 µg/ml concentrations of only β -pinene resulted in the death of tested microorganisms with more than 80% mortality rate. The collective toxic effect of β -pinene was efficiently greater than the compressive effect of all chemical compounds presented in the essential oils of F. gummosa. On the basis of LC_{50} values (2.20 µg/ml) of β -pinene was considered as the most potent scolicidal agent in this study. Kavoosi et al., (2013) demonstrated the scolicidal effectiveness of essential oil from F. assa-foetida with a plant. A

sulfur-containing hydrocarbon (E)-sec-butyl propenyl disulfide, 62.7%) was found to be the main component in the essential oil analyzed by the gas chromatography method. The results proved that 10 min exposure of *F. assafoetida* essential oil with a concentration of $60 \,\mu$ g/ml or more could kill all *Echinococcus granulosus* protoscolices.

TOXICITY

Ye et al., (2011) studied the effect of chemical components separated from crude essential oil of F. sinkiangensis K.M.Shen, on acute toxicity in morphine-dependent animals. Two sulfur-containing compounds: 2-butyl cis-1-propenyl disulfide and 2-butyl trans-1-propenyl disulfide (SBD) were separated from the unrefined essential oil. To evaluate the effectiveness and naturally abstinent and naloxone-precipitated abstinent morphine-dependent models were applied and injected with 2-butyl trans-1-propenyl disulfide intraperitoneally. In addition, the antinociceptive effects, sedative effects, and acute toxicity of SBD were investigated by a writhing test, spontaneous activity test, and LD₅₀ values, respectively. The result showed that SBD could be inhibited by the abstinent syndromes, and the 2butyl trans-1-propenyl disulfide had great sedative and antinociceptive effects as well. Tempark et al., (2016) indicated that topical administration of F. assa-foetida oleo-gum-resin essential oil could cause contact dermatitis in infants. This high content of disulfide-containing hydrocarbons in asafoetida oil might cause skin-irritating effects. These types of pro-inflammatory side effects could be diminished by the elimination or reduction of disulfide compounds from the essential oil.

ANTIOXIDANT ACTIVITY

Kavoosi et al., (2012) demonstrated the radical scavenging properties of F. assa-foetida. The essential oil was prepared by hydrodistillation contained (E)-sec-butyl propenyl disulfide and β -ocimene and pinene. The essential oil was subjected to different radical scavenging activity assays to evaluate the antioxidant potential. The report displayed that F. assa-foetida essential oils did not have any significant antioxidant activity. Pavlović et al., (2012) effectuated antioxidant potential of essential oil obtained from Ferula heuffelii Griseb. ex Heuff. underground parts. The main compounds of the essential oil were elemicin and myristicin with 35.4 and 20.6% total concentration. L-Ascorbic acid was used as a reference substance. The resulting investigation delineated that essential oils showed antiradical activity, concentration-dependently. The SC50 value was obtained at 22.43 and 3.80 µg/ml for tested essential oil and reference substance, respectively. Ahmadvand et al., (2014) revealed the antioxidant potential and chemical profile of F. assa-foetida leaf essential oil. Analysis of hydro-distilled leaves' essential oil revealed major chemical compounds namely eremophilene (31.28%), δ -cadinene, longiborneol, dehydro aromadendrene, and isoledene. Antioxidant activity was examined by using a

0.01-1,000 µg/ml essential oil concentration. Antioxidant activity analysis showed that the IC₅₀ for DPPH (2,2-diphenyl-1picrylhydrazyl) free radicals was 2,375.66 ± 5.13 µg/ml. Benchabane et al., (2012) analyzed the antioxidant potential of Ferula vesceritensis Coss. & Durieu ex Trab. The chemical profile displayed viridiflorol (13.4%) as a major constituent of oil and delta-cadinene, trans-farnesol, alpha-fenchyl acetate, aristolene, cadinol, and fonenol were also found in the essential oil. Antioxidant activity was accomplished by using a 100-1,000 mg/L oil concentration. Butylated hydroxytoluene (BHT) was used as a positive control with 100-1,000 mg/L concentrations. F. vesceritensis essential oil exhibited a reduction in DPPH free radical concentration but with lower efficacy compared to BHT. Znati et al., (2017) assessed different bioactivities of seed oil from Ferula tunetana Pomel ex Batt. including antioxidant bioactivity. Pinene (α/β) and (Z)- β -ocimene were characterized as the main component of the essential oil by gas chromatography and carbon-13 nuclear magnetic resonance methods. The essential oil displayed antioxidant activity with moderate efficiency. The H₂O₂ assay exhibited the highest activity with 78.2 \pm 2.98 µg/ml IC₅₀ value, while ABTS, DPPH, and superoxide anion assay radical scavenging assays showed 234.2 \pm 12.9, 243.1 \pm 6.5 and 89.2 \pm 3.82, µg/mL IC₅₀ values, sequentially. Amiri (2014) elucidated the antioxidant activity of methanolic extracts and essential oil of Ferula microcolea (Boiss.). α -Pinene (27.3%), β -pinene, nonanaldehvde, β -caryophyllene, and 2-isopropyl-5methylphenol were recognized as primary chemical constituents. The IC₅₀ value for β -carotene-linoleic acid and DPPH were recorded at 55.2 \pm 0.4% and 253.1 \pm 2.2 µg/ml, for essential oil content, respectively. Kose et al.,(2010) accomplished the antioxidant potential of Ferula lycia Boiss. essential oil. The bleaching of the carotene-linoleate and DPPH was used to examine the antioxidant activity of the essential oil. The essential oil (0.4, 1.0 and 2.0 mg/ml) showed β -carotene linoleic acid abilities (5.69 ± 2.04, 16.16 ± 0.52, and 27.77 ± 2.37 mg/ml) and DPPH radical scavenging (11.05 ± 0.50 , 1.91 ± 0.43 and 2.81 ± 0.0 mg/ml), respectively. Dadaşoğlu et al., (2018) revealed the antibacterial and antioxidant activity of the essential oil of F. communis L. The essential oil showed antioxidant activity at 40.65% (0.1 ml) and 85.16% (0.2 ml) concentration for DPPH, ABTS assay, respectively. Jahani et al., (2015) synthesized gelatin nano-capsules using F. assafoetida essential oil (FAO) and tested their potential to exhibit antibacterial and antioxidant activity. Essential oils containing gelatin nano-capsules were synthesized with Ferula oil at 2, 4, 6, and 8% w/w concentrations; 25% w/w, glutaraldehyde (a crosslinker) and glycerol (plasticizer). Synthesized gelatin nanocapsules were evaluated by scanning electron microscopy. FAO incorporated gelatin nano-capsules exhibited excellent antioxidant and antibacterial at 8% of FAO concentration. Sharopov et al., (2015) examined the antioxidant activity, along with the anti-inflammatory activity of essential oils of some aromatic plants including Ferula clematidifolia Koso-Pol., Ferula foetida (Bunge) Regel, etc. The results showed that both species had moderate antioxidant activity. Kavoosi and Rowshan (2013) also investigated the chemical profile and antioxidant

potential of essential oil from F. assa-foetida oleo-gum-resin. F. assa-foetida resin (oleo-gum-resins, ORGs) was collected in three different times named as ORG1, ORG2, and OGR3 and subjected to hydro-distillation. The IC50 value for all listed scavenging methods were calculated as follows OGR1 (0.017 \pm 0.0019, 0.012 \pm 0.0020, 0.035 \pm 0.0027, and 0.022 \pm 0.0012 mg/ml; OGR2 (0.031 ± 0.0018 , 0.025 ± 0.0023 , $0.047 \pm$ 0.0028, and 0.033 ± 0.0043 mg/ml); OGR3 (0.047 ± 0.0028, 0.035 ± 0.0012 , 0.066 ± 0.0042 , and 0.055 ± 0.0038 mg/ml) for RNS, ROS, TBARS, and H₂O₂ scavenging assay, respectively. Antioxidant activity of OGR1 (18.16 \pm 1.2 mg), OGR2 (14.14 \pm 2.2 mg), and OGR3 (10.8 \pm 2.5 mg) were observed at mg ascorbic acid/g of essential oil, respectively. Topdas et al.,(2020) revealed the antioxidant activity of Ferula orientalis L. essential oil containing α -pinene, ortho-cymene, limonene, terpinolene, β -caryophyllene and isobornyl acetate as the major compounds. The antioxidant activity (in vitro) of the essential oil was analyzed. The report averred that the essential oil chemical group had significant antioxidant activity against the ABTS and DPPH free radicals. Sharopov et al.,(2019) revealed the antioxidant activities of hydrodistilled essential oil, extracted from the Ferula tadshikorum Pimenov. underground parts. Data analysis confirmed that the essential oil exerted lower antioxidant potential than the caffeic acid (positive control) with 17.8 and 8.2 mg/ml median inhibitory concentration (IC₅₀) for DPPH and ABTS, respectively.

ANTIPROTOZOAL ACTIVITY

Essid et al., (2015) investigated the antileishmanial activity of essential oil from medicinal plants. This study included Ferula communis L. with 11 medicinal plants to identify the antileishmanial potential against Leishmania infantum and L. major promastigotes. The major components of F. communis were: β -caryophyllene (15.22%), β -myrcene alpha-eudesmol, alpha-pinene, para-mentha-1, 5-dien-8-ol and y-curcumene. Amphotericin B was applied as a form of positive control in the experimental setup. Data collected from the experiment setup showed that the essential oils had potent antileishmanial activity against L. infantum and L. major promastigotes at IC₅₀ value <1 µg/ml. After result analysis, it was concluded that L. infantum promastigotes (IC₅₀ value $0.80 \pm 0.18 \,\mu\text{g/ml}$) were more sensitive to the essential oil and their constituents as compared to L. major (IC₅₀ value 0.22 \pm 0.09 µg/ml). According to Essid et al., caryophyllene had high leishmanicidal activity against L. infantum and L. major (1.06 \pm 0.37 and 1.33 \pm 0.52 µg/ml, respectively). Andrade et al.,(2016) effectuated the in vitro anti-leishmanial activity of different plants' essential oils including Ferula galbaniflua Boiss. & Buhse. GC-MS analysis of the essential oil showed methyl pimar-8-en-18-oate (41.82%) and diethyl phthalate (13.09%) as main components. In vitro leishmaniacidal activity of essential oil was examined on Leishmania amazonensis promastigotes forms at 30-500 µg/ml oil concentrations. F. galbaniflua essential oil was more potent against L. amazonensisat at 95.70 \pm 1.82 µg/ml (IC₅₀/24 h). El-Deeb et al., (2012) substantiated in vitro inhibitory effects of F.

assa-foetida essential oil on *Blastocystis* species. The volatile oil of powdered assa-foetida was extracted by hydrodistillation and tested against the *Blastocystis* sp. subtype. Various concentrations such as 5, 10, 25, 40, and 50 mg/ml were used for 24, 72, and 144 h. Metronidazole was used as the reference antiprotozoan drug including a 10, 100, and 500 μ g/ml concentration. The results confirmed by microscopy described that extracted oil decreased the viability and counts of all the *Blastocystis* sp. The lowest and highest percentage inhibition values at which blastocysts showed no growth and multiplication were 16 and 40 mg/ml, respectively. At the aforementioned concentration, the mean count was the same for the oil extract and the reference drug. Furthermore, recultivation of *Blastocystis* in oil-free medium did not display any growth even after 48, 72, and 144 h of cultivation.

NEUROPROTECTIVE

Topdas et al.,(2020) elucidated the neuroprotective potential of various types of extracts and essential oils of *F. orientalis* L. Neuroprotective potential of the essential oil was investigated in cortex neuron cells by 2-(3,5-diphenyltetrazol-2-ium-2-yl)-4,5-dimethyl-1,3-thiazole; bromide (MTT) assay. The essential oil concentrations ranging from 10^{-1} to 10^{-8} were used for the experiments. The cell groups treated with essential oil clearly exhibited the highest cell viability rates. The viability rates were 92.57 ± 4.23 , 91.29 ± 4.12 , and $83.60 \pm 3.98\%$ at 10^{-2} , 10^{-3} , and 10^{-4} of the oil concentration (p > 0.05), respectively. Moreover, the result clearly showed that at 10^{-2} concentration, the cell viability was at its peak, after this mentioned concentration of essential oil, the viability rates started to fall down slightly.

ANTIGERMINATIVE ACTIVITY

The *in vitro* antigerminative property of *Ferula tunetana* Pomel ex Batt. seed oil was demonstrated by Znati et al.,(2017). Four doses (1.25, 5, 10, and 20 mg) of the oil were prepared by diluting it in the emulsion in 5 ml deionized H₂O. The essential oil exerted significant toxicity against *Medicago sativa* L., *Triticum aestivum* L. and *Lactuca sativa* Linn. Seven days of exposure to essential oil showed the maximum toxic effect with 0% germination. Furthermore, the germination of *M. sativa*, *T. aestivum* and *L. sativa*, seedlings were efficiently inhibited at a 20 and 10 (mg/ 5 ml) concentration. The authors suggested that α -pinene (39.8%) was responsible for the toxic effect.

VASODILATORY OR VASORELAXANT ACTIVITY

Esmaeili et al.,(2020) studied the role and significance of the K^+ channels in vasorelaxant and the effect of essential oil from asafoetida (AEO). The AEO obtained from *F. assa-foetida* oleoresin was subject to the vasodilation effect examination. This effect had two types (endothelium-independent and dependent). This

research was designed to demonstrate whether intracellular Ca²⁺ release and K⁺ channels had a contribution in the essential oilmediated vasodilation or not. For the experiments, isolated rats' thoracic aortas were denuded, and the concentration-response curve was plotted after induction of contraction by KCl (60 mM) and addition of 0.625-80 µL/L of AEO in the medium. The vasodilatory effect of the essential oil was studied by the addition of essential oil in the medium before and after the addition of phenylephrine and potassium channel blocking chemicals viz. barium chloride (BaCl₂), glibenclamide (GL) and 4-aminopyridine (4A). AEO displayed a relaxant effect on the precontracted rings, concentration-dependently at a 23 µL/L IC₅₀ value. Furthermore, it was observed that K⁺ channel blockers significantly abolished the AEO mediated vasodilatory effect, if it was added before the addition of KCl to the ring medium. In contrast to potassium channel blockers, the tension was significantly decreased with the addition of the AEO before or after phenylephrine addition. The result interpreted that the inhibition of Ca⁺² channels and the activation of smooth muscle membrane K⁺ channels were responsible for the vasodilatory effect of the essential oil on the denude y-endothelium aortic ring. Esmaeili et al.,(2017) investigated the vasodilatory activity of AEO using rat aorta ring as a living system. The contribution of Ca⁺² channel, prostacyclin and NO (nitric oxide) in the vasodilation process were additional. In an experiment, thoracic aorta rings were stretched in an organ bath apparatus, after that the rings precontracted using 80 mM of KCl with or without the AEO portion. To reveal the role of nitric oxide and prostacyclin in the AEO vasodilatory effect, indomethacin (blocker of cyclooxygenase) and L-NAME (NO synthase blocker) were used. The AEO effect on the influx of Ca^{+2} ions were also evaluated. Data showed that the essential oil exerted significant effects on the aorta rings' vasodilation; the IC50 values for denuded and intact endothelium cells were 19.2 and 1.6 µL/L, sequentially. The AEO mediated vasodilatory could reduce by L-NAME or indomethacin but could not be abolished. On the basis of the result, it was concluded that the AEO had an effective vasodilation activity, which could be endothelium-dependent or independent. AEO also decreased the influx of Ca⁺² ions from the calcium channels of the plasma membrane into the cell.

ANTIEPILEPTIC OR ANTICONVULSANT AND HYPOTENSIVE EFFECT

Sayah et al.,(2001) used the fruit essential oil of the *F. gummosa* to evaluate antiepileptic activity. The result showed that the essential oil had no measurable effect on maximal electroshock-induced seizures, but it can reduce the effect of pentylenetetrazole-induced tonic seizures in mice. GC analyses of the essential oil presented pinene (β : 50.1%; α : 18.3%), delta-3-carene (6.7%), origanene (3.3%), and 4(10)-thujene (3.1%) as the main components. It was also suggested that toxic and anticonvulsant effects of the essential oil might be related to the compounds α -thujene and β -pinene. Ghanbari et al.,(2012) aimed to reveal the chronic and acute effects of *Ferula persica* Willd. on hypertensive rats' blood pressure. Their study presented that the hypotensive effect might

be due to the presence of safranal, a component of the *F. persica* EO. Furthermore, it was found that *F. persica*'s essential oil might exert a hypotensive effect by the induction of nitrous oxide release and muscarinic receptors targeting in rats.

HYPOGLYCEMIC AND HYPOLIPIDEMIC EFFECT

Heydari-Majd et al., (2019) performed the synthesis of F. gummosa essential oil or barije essential oil (BEO) incorporated zein nanofibre, subjected to α -amylase and α -glucosidase inhibitory action. GC/MS analysis of BEO revealed the presence of alphapinene, guai-1 (10)-en-11-ol, champacol and β -myrcene as the major components of the oil. Morphological analysis of prepared zein nanofibers done by SEM and FT-IR showed that the essential oil components were successfully entangled in the ribbon structured zein fibers with ~95% encapsulation efficiency. BEOloaded (1–4% w/w) zein nano-fibers exhibited α -amylase (IC₅₀: 1.09 \pm 0.02 to 1.64 \pm 0.01 mg/ml) and α -glucosidase (IC_{50}:0.78 \pm 0.01 to 1.25 ± 0.03 mg/ml) inhibition activity. The model-fitting results showed that BEO-loaded zein nano-fibers could be a delivery vehicle for diabetes control. Karimlar et al.,(2019) exposed the hypoglycemic and hypolipidemic effects of three medicinal plants' essential oils including F. gummosa on streptozotocin-induced diabetic rats. For the experimental setup the streptozotocin-induced (45 mg/kg doses, intraperitoneally) male wistar rats were used and treated with the essential oil (200 mg/kg/day). After 30 days, rats' lipid profiles and serum glucose were assessed. Data were examined by the Tukey test and one-way ANOVA test. The value of HDL-C, LDL-C, cholesterol, triglyceride, and glucose in F. gummosa essential oil treated rats' blood were recorded as 42.07 ± 9.68, 29.59 ± 3.76, 91.02 ± 11.95, 105.18 ± 12.13, and 429.91 ± 46.14 µM, respectively. The F. gummosa essential oil significantly reduced low-density lipoprotein cholesterol and triglycerides in diabetic rats; even though the essential oil tested group did not display any significant difference in glucose level from the diabetic group. Yarizade et al., (2017) studied the antidiabetic activity of F. assafoetida. In this study, it was observed that F. assa-fetida showed its antidiabetic activity by inhibiting a-glucosidase and DPP-IV (Dipeptidyl peptidase- IV).

ANTINOCICEPTIVE AND HYPERALGESIA EFFECT

In a study, Radulović et al.,(2013) reported hyperalgesia induction in mice with a chemical compound isolated from *F. ovina* (Boiss.) Boiss. The chemical profile of aerial parts of the essential oil of the *F. ovina* revealed the presence of a rare aromatic ester of monoterpenic alcohol named bornyl 4-methoxybenzoate, and its structure was evaluated by X-ray crystallographic analysis. The analgesic effect (the hot plate and tail immersion tests) and antinociceptive activity (abdominal writhings test) of the new compound with other oil constituents were elucidated. To determine the effect of essential oil and bornyl 4methoxybenzoate, an experiment was performed using male BALB/c mice as laboratory models. The results showed that bornyl 4-methoxybenzoate induced hyperalgesia in mice which is revealed by a hot plate test. The transient receptor channels (TrpV3) could have a target for tested substances and is a possible reason for hyperalgesia. The oil was found to have exerted a modest central and significant peripheral analgesic effect. The oil rendered a significant antinociceptive activity, dose-dependently, and abolished acetic acid-induced abdominal writhings. The number of writhings was reduced by up to 92% at 200 mg/kg essential oil concentration and up to 83% at 200 mg/kg bornyl 4methoxybenzoate concentration in treated mice was observed. Some of the major chemical constituents such as myrcene, limonene, and α -pinene were ascribed to possess certain analgesic potential. The result showed that essential oil and bornyl 4-methoxybenzoate could have caused inhibition of prostaglandin synthesis. Bagheri et al.,(2016) studied the antinociceptive potential of F. assa-foetida seed essential oil in mice. To evaluate the antinociceptive effect of the essential oil (2.5, 5 and 10 mg/kg), acetic acid-induced writhing and a hot plate test were used and for the control group morphine sulfate (8 mg/kg) or sodium diclofenac (30 mg/kg) were applied. Hot plate testing results showed that the percentage of the MPE (maximum possible effect) was higher for all used concentrations of the essential oil than morphine sulfate and sodium diclofenac. The writhes numbers were significantly less in the essential oil treated mice as compared to the control group. This research findings indicated that the essential oil reduces acetic acid-induced writhes numbers dose-dependently and presented a potent antinociceptive effect on acute/chronic pain in mice. The analgesic effect of the essential oil is thought to be either due to its action on acetic acidsensitive visceral receptors or inhibition of synthesis and action of prostaglandins and also cyclooxygenase and/or lipoxygenase in the arachidonic acid cascade at the peripheral route.

ANTICANCER AND ANTITUMOR ACTIVITY

Hosseinzadeh et al.,(2020a), Hosseinzadeh et al.,(2020b) performed the synthesis of gold nanoparticles using essential oil obtained from the gum of F. persica Willd. and evaluated their in vitro anticancer effects. Phytochemistry profiles were effectuated by the GC- MS method and displayed 27 constituents such as α -pinene (27.1%), (Z)-sec-butyl propenyl disulfide (20.2%) and β -pinene (10.6%) as the major component. The gold nanoparticles (Au NPs), which were characterized by ultraviolet-visible spectroscopy showed absorption at 530 nm. The shape (spherical) and size (37.05-78.6 nm) of Au NPs were confirmed by TEM image. The presence of reducing and capping essential oil compounds on the gold ions and metal crystal structure was revealed by the FTIR spectrum and XRD pattern, respectively. The apoptosis and cytotoxicity assessment were performed by MTT [3-(4,5-dimethylthiazol-2-yl)-2,5diphenyl tetrazolium bromide] assay and AO/EB (acridine orange/ethidium bromide) staining using non-cancerous (Vero cells) cells and cancerous (Murine colon carcinoma CT26) cells. The result showed that the cytotoxicity effect of Au NPs was dosedependent and exhibited cytotoxicity against Vero cell and murine colon carcinoma CT26 lines with IC50 values 0.0024 and 0.0307 mg/ml, respectively. Further, AuNPs inhibited colony formation in the aforementioned cells and induced apoptosis. The effect of AuNPs was reported more intensively against CT26 cells. The result clearly indicated the cytotoxic, apoptotic, and antiproliferative potential of the synthesized Au NPs. Elghwaji et al.,(2017) aimed to study the antitumor potentiality of the Ferula tingitana L. of essential oil. The essential oils were obtained from the hydrodistillation of flower and leaves possessing sesquiterpenes and oxygenated sesquiterpenes as their major chemical components, respectively. Cytotoxicity against human tumor cells viz. hormone-responsive breast (MCF7) cells, cervical (HeLa) cells and liver carcinoma (HePG2) cells were performed by sulforhodamine B (SRB) method using different doses of the essential oil (0.0-50.0 µg/ml). The IC₅₀% value for in vitro cytotoxicity of flower and leaf derived essential oils against liver carcinoma (HEPG2), cervical (HELA), and breast (MCF7) cell lines (two different) were 4.4, 4.2; 8.6, 10.9 and 6.9, 4.8 µg/ml, respectively. Dithiolane found in high concentrations in F. assa-foetida essential oil; this compound exhibited antiproliferative activity in two human liver carcinoma cell lines (SK-Hep1and HepG2), dose-dependently. Two signaling molecules: NF-kB and TGF-ß altered after the use of bioactive compounds of F. assa-foetida; moreover, an increase in caspase-3 and TNF-a expression was observed and caused induction of apoptosis (Verma et al., 2019).

CYTOTOXIC ACTIVITY

The chemical profile and bioactivity of essential oil from underground parts of the Ferula tadshikorum Pimenov. were investigated (Sharopov et al., 2019). The chemical profile analysis revealed sulfur-containing hydrocarbon as a major component. The assessment of the cytotoxic effect of the essential oil was done on CEM/ADR5000 and CCRF-CEM cancer cell lines by using an MTT assay. Data displayed IC₅₀ values were 142.5 µg/ml and 21.6 µg/ml for CEM/ADR5000 and CCRF-CEM cell lines, respectively. The essential oil shows a reduced cytotoxicity effect on CEM/ADR5000, due to the presence of substrates P-glycoprotein (p-gp) and overexpression of ATP-binding cassette transporter p-gp, and it rapidly pumped all the active molecules of essential oil out of the cells. Ben et al.,(2016) elucidated the cytotoxic effect of the root oil obtained from the Ferula lutea (Poir.) Maire. The chemical profile of the oil was investigated by GC-MS/FID and 13C-NMR spectroscopy revealed the major component viz. delta-3-carene (~73%). The cytotoxic effect of the essential oil was carried out using the MTT method on human colon cell lines (HCT-116 and HT-29 cells). For the testing, Paclitaxel was added as a positive control. The result of MTT assay delineated that the hydrodistilled F. lutea roots' essential oil has a moderate cytotoxic effect on HT-29 and HCT-116 cells (Khajeh et al., 2005). The IC₅₀ values were 26.39 \pm 3.98 µg/ml and 81.00 ± 12.81 µg/ml for HT-29 and HCT-116, respectively. The result showed that HCT-116 cells were extra sensitive to the isolated essential oil compared to HT-29 cells. The cytotoxicity of gum essential oil from F. persica Willd. on Vero cell lines and murine colon carcinoma (CT26) was demonstrated by MTT assay (Hosseinzadeh et al., 2020a; Hosseinzadeh et al., 2020b). The primary component of the essential oil was alpha-pinene (27%), (Z)-sec-butyl propenyl disulfide (20%), β -pinene (11%), transdihydroagarofuran (6%), allo-aromadendrene (5%), (Z)- β -ocimene (4.5%) and α -caryophyllene (3%). The viability was decreased in the Vero cell $(0.125 \times 10^{-9}$ to $80 \,\mu\text{L/ml})$ and CT26 cell line (0.125 \times 10⁻⁹ to 20 μ L/ml) significantly with all different concentrations of the essential oil treatment, dose-dependently. The IC_{50} value for the CT26 cell line (0.3247 μ L/ml) was relatively greater than Vero cells (0.0010 μ L/ml). The essential oil treatment exhibited valuable inhibition of colony formation in Vero cells and CT26 compared to the control. Moreover, some measurable changes such as morphological changes, the monolayer of the cells with areas devoid of cells, nucleus condensation, blebbing of the cell membrane, and apoptotic body formation were observed in fluorescence microscope by AO/EB staining. Bagheri et al.,(2017a) studied the cytotoxic effect of ferulic acid and essential oil obtained from oleogum resin of F. assa-foetida on 4T1 breast cancer cells. Data analysis enlightened the fact that the incubation of breast cancer 4T1 cells with the essential oil at the concentration ranging from 1 to $1,000 \mu g/$ ml for 24 h did not show any significant cytotoxicity. Additionally, the viability of cancer cells started to gradually decrease after 48 and 72 h of the incubation period. Nevertheless, a 10% cell viability rate was remarked even after the incubation with the highest concentration (1,000 µg/ml) of the essential oil for 72 h. On the basis of the result, it could be concluded that the cytotoxic effect of the essential oil was time and concentration-dependent. Nguir et al., (2016) used the Hela cervix cell and A549 human lung epithelial carcinoma cell lines to assess the cytotoxicity of F. communis L. essential oil (flowers, roots, leaves, and stems). The MTT test method was used with slight modifications.

For the activity assessment, the cell lines were treated with the essential oil at different concentrations. It was observed that activity was increased in both cell lines with a higher essential oil concentration, but at 500 µg/ml concentration significant activities were observed in both cell lines. Moreover, the Hela cells were reported to be more sensitive than A549 cells and displayed 79.05% and 77.52% inhibition at 500 and 250 µg/ml concentration of stem essential oil, respectively; flower essential oil showed 74.89% inhibition. Abbaszadegan et al.,(2015) demonstrated the cytotoxicity of Ferula gummosa essential oil against L929 mouse fibroblasts using a colorimetric, MTT assay, and the Sigma-Aldrich method. Chlorhexidine (CHX: 0.2%) and sodium hypochlorite (NaOCl: 5%) solutions were used as a control. The cytocompatibility of the essential oil was estimated on L929 fibroblast cells in comparison to the control. The chemical profile analysis of the essential oil displayed the presence of 27 chemical constituents. β -Pinene (51.83%) was the main component in the essential oil. In the experiment, culture medium and H₂O₂ (35%) were used as the negative and positive controls, respectively. The cytotoxicity assessment showed that the full concentration (50 μ L/ ml) of the essential oil had the ability to keep the mean cell viability of L929 mouse fibroblast cells at about 88%. There was no measurable difference in the mean viability, of the negative control group (H₂O₂) and in NaOCl (5%) and CHX (0.2%) treated cells. Additionally, no significant variation between the

cytotoxic effect of the essential oil and CHX or NaOCl was observed. Kavoosi and Purfard (2013) considered phenolic monoterpenes as a target site for essential oils to show their cytotoxic effect which was the cytoplasmic and mitochondrial membrane. The oil molecules pass through the cytoplasmic membrane and increase permeabilization, and ions leakage (especially, potassium and calcium) from membranes reduces membrane electric potential, ATP, amino acids, proteins synthesis, and cell death.

MISCELLANEOUS ACTIVITY

Rashidi et al.,(2014) demonstrated the galbanum prophylactic effect on caffeine teratogenic effects. For the experiment, four groups of pregnant rats were selected: one group (control) was injected with saline, two groups with galbanum (200 mg/kg), caffeine (80 mg/kg), and the remaining group with both compounds (galbanum + caffeine), intraperitoneally for 9-11 days of gestation. To obtain the data, fetuses (20th day of gestation) were collected and stained using Alizarin red-Alcian blue method. The report established that galbanum decreased caffeine-induced cleft palate incidence by 8.3% in the galbanum group as compared to the caffeine group (33.3%). Kavoosi et al.,(2013) tried to develop F. assa-foetida essential oil incorporated wound dressing film with potent antibacterial and antioxidant properties. The film was prepared with 10% w/v gelatin solutions containing different concentrations of the essential oil. The result analysis displayed that the entraption of essential oil into gelatin films showed a valuable reduction in tensile strength swelling, elastic modulus and vapor penetrability; enhancement in solubility and resistance. However, essential oil incorporated gelatin film displayed appreciable antioxidant and antimicrobial activities compared to gelatin film without essential oil. In a study, Esmaeili et al., (2018) investigated the effect of the essential oil on a myocardial ischemic-reperfusion injury obtained from F. assafoetida (AEO). Three concentrations of AEO (0.50, 0.25 and 0.125 µL/g heart) were used. The results of the analysis showed that the AEO treated group exhibited severe myocardial dysfunction with a significant increase in left ventricular enddiastolic pressure and a reduction in left ventricular developed pressure as compared to the control group. The markers of myocardial injury (lactate dehydrogenase and creatine kinase) were also significantly active in the treated group compared to the control. Moreover, the essential oil exerted an effect on perfusion in isolated rat hearts at 0.5 µL/g heart concentration, but not below these concentrations.

CONCLUSION AND FUTURE PERSPECTIVES

The trend of natural products in the medical and other fields is growing and replacing the use of chemically synthesized drugs. Therefore, numerous scientific investigations have been conducted over a few decades. On the basis of the literature, the genus Ferula is a well-known genus of the Apiaceae family. It is widely used as an aroma spice in different foods all over the world. To date, a large number of biologically active components have been separated from the essential oils of Ferula which have shown the many different activities such as antimicrobial, insecticidal, antioxidant, antigerminative, cytotoxic, antitumor, antidiabetic activities, etc. All these activities have been discussed in this review. A large number of reports have been conducted on the variability in the chemical composition (quality and quantity) of essential oil extracted from the members of the genus Ferula. In these reports, it has clearly been stated that environmental factors, genetic factors, geographical area, and collection time have a great effect on essential oil composition. Ferula essential oil showed great anti-cancerous activity against various tested cancer cell lines. After a literature survey on the antimicrobial and insecticidal potential of this genus, it can be concluded that the essential oils are effective drug candidates for developing natural or semi-synthetic derivatives against drug-resistant microbes and insects and protozoan (Leishmania). Along with all the positive points, there are some negative points associated with essential oils derived from this genus which should be considered. Some species of Ferula show toxicity in humans and animals. Further, research is therefore needed in order to develop a befitting standard of the safe use of essential oils. The pharmaceutical potential of the essential oil of the Ferula species is still not completely understood. The revealed bioactivities are reported *in vitro*, therefore their practical application should be focused on and detailed studies are also needed to find new chemical constituents in the genus.

AUTHOR CONTRIBUTIONS

MM and PS conceived the idea of the review, provided the general concept and inputs for each specific section, and drafted part of the manuscript. PS and MM wrote the review after collecting literature. MM edited, compiled, and finalized the draft. Finally, both the authors read and approved it for publication.

FUNDING

This study was supported by the Startup Research Grant (UGC Faculty Research Promotion Scheme; FRPS) and sustained by Mohanlal Sukhadia University, Udaipur, Rajasthan, India.

ACKNOWLEDGMENTS

The author, MM is thankful to Mohanlal Sukhadia University, Udaipur for providing the necessary facilities during the course of the study.

REFERENCES

- Abbasnia, V. S., and Aeinfar, H. (2016). Anxiolytic and hypnotic effect of Ferula assafoetida aqueous extract in mice. Int. J. Pharm. Technol. 8, 15974–15979.
- Abbaszadegan, A., Gholami, A., Mirhadi, H., Saliminasab, M., Kazemi, A., and Moein, M. R. (2015). Antimicrobial and cytotoxic activity of *Ferula gummosa* plant essential oil compared to NaOCl and CHX: a preliminary *in vitro* study. *Restor. Dent. Endod.* 40 (1), 50–57. doi:10.5395/rde.2015.40.1.50
- Afshar, F. F., Saffarian, P., Hosseini, H. M., Sattarian, F., Amin, M., and Fooladi, A. A. I. (2016). Antimicrobial effects of *Ferula gummosa* Boiss gum against extended-spectrum β-lactamase producing *Acinetobacter* clinical isolates. Iran. J. Microbiol. 8 (4), 263. doi:10.2174/1574891X13666180426163427
- Ahmadi, K. S., Hadjiakhoondi, A., Delnavazi, M. R., Tofighi, Z., Ajani, Y., and Kiashi, F. (2020). Chemical composition and biological activity of *Ferula* aucheri essential oil. Res. J. Pharmacogn. 7 (2), 21–31. doi:10.22127/RJP. 2020.210354.1537
- Ahmadvand, H., Amiri, H., Dehghani Elmi, Z., and Bagheri, S. (2014). Chemical composition and antioxidant properties of *Ferula-assa-foetida* leaves essential oil. Iran. J. Pharm. Therapeut. 12 (2), 52–57. http://ijpt.iums.ac.ir/article-1-262-en.html.
- Akhzari, D., and Saadatfar, A. (2019). Growth, phytoremediation potency and essential oil component of *Ferula haussknechtii* H. Wolff ex Rech. f. grown under lead stress conditions. *J. Rangel. Sci.* 9 (4), 392–401. doi:10.3390/ biom10040592
- Al-Ja'fari, A. H., Vila, R., Freixa, B., Tomi, F., Casanova, J., Costa, J., et al. (2011). Composition and antifungal activity of the essential oil from the rhizome and roots of *Ferula hermonis*. *Phytochemistry*. 72 (11–12), 1406–1413. doi:10.1016/j. phytochem.2011.04.013
- Alipour, Z., Taheri, P., and Samadi, N. (2015). Chemical composition and antibacterial activity of the essential oils from flower, leaf and stem of *Ferula cupularis* growing wild in Iran. *Pharm. Biol.* 53 (4), 483–487. doi:10. 3109/13880209.2014.924149
- Alizadeh, M. N., Rashidi, M., Muhammadnejad, A., Zanjani, T. M., and Ziai, S. A. (2018). Antitumor effects of umbelliprenin in a mouse model of colorectal cancer. *Iran J. Pharm. Res. IJPR.* 17 (3), 976–985. doi:10.3390/molecules24234278
- Amalraj, A., and Gopi, S. (2017). Biological activities and medicinal properties of Asafoetida: a review. J. Tradit. Complement. Med. 7 (3), 347–359. doi:10.1016/j. jtcme.2016.11.004
- Amin, A., Tuenter, E., Cos, P., Maes, L., Exarchou, V., Apers, S., et al. (2016). Antiprotozoal and antiglycation activities of sesquiterpene coumarins from *Ferula narthex* exudate. *Molecules*. 21 (10), 1287. doi:10.3390/molecules21101287
- Amiri, H. (2014). Chemical composition and antioxidant activity of essential oil and methanolic extracts of *Ferula microcolea* Boiss (Apiaceae). *Int. J. Food Prop.* 17 (4), 722–730. doi:10.1080/10942912.2012.665403
- Andrade, M. A., Azevedo, C., Motta, F. N., Santos, M. L., Silva, C. L., Santana, J. M., et al. (2016). Essential oils: *In vitro* activity against *Leishmania amazonensis*, cytotoxicity and chemical composition. *BMC Compl. Alternative Med.* 16 (1), 444. doi:10.1186/s12906-016-1401-9
- Arjmand, Z., and Dastan, D. (2020). Chemical characterization and biological activity of essential oils from the aerial part and root of *Ferula haussknechtii*. *Flavour Fragr. J.* 35 (1), 114–123. doi:10.1002/ffj.3544
- Asemani, Y., Azadmehr, A., Hajiaghaee, R., and Amirghofran, Z. (2018). Anticancer potential of *Ferula hezarlalehzarica* Y. Ajani fraction in Raji lymphoma cell line: induction of apoptosis, cell cycle arrest, and changes in mitochondrial membrane potential. *Daru.* 26 (2), 143–154. doi:10.1007/ s40199-018-0219-z
- Asilbekova, D. T., Ozek, G., Ozek, T., Bobakulov, K. M., Baser, K. H. C., and Sagdullaev, S. S. (2019). Essential oil and lipids from leaves of *Ferula* kuhistanica. Chem. Nat. Compd. 55 (6), 993–998. doi:10.1007/s10600-019-02877-3
- Azarnivand, H., Alikhah-Asl, M., Jafari, M., Arzani, H., Amin, G., and Mousavi, S. S. (2011). Comparison of essential oils from *Ferula ovina* (Boiss.) aerial parts in fresh and dry stages. *J. Essent. Oil-Bear. Plants.* 14 (2), 250–254. doi:10.1080/ 0972060X.2011.10643929
- Aziz, Z. A., Ahmad, A., Setapar, S. H. M., Karakucuk, A., Azim, M. M., Lokhat, D., et al. (2018). Essential oils: extraction techniques, pharmaceutical and therapeutic potential—a review. *Curr. Drug Metabol.* 19 (13), 1100–1110. doi:10.2174/1389200219666180723144850

- Baccari, W., Znati, M., Zardi-Bergaoui, A., Chaieb, I., Flamini, G., Ascrizzi, R., et al. (2020). Composition and insecticide potential against *Tribolium castaneum* of the fractionated essential oil from the flowers of the Tunisian endemic plant *Ferula tunetana* Pomel ex Batt. *Ind. Crop. Prod.* 143, 111888. doi:10.1016/j. indcrop.2019.111888
- Bagheri, M., and Rahimi, M. (2014). Effect of *Ferula assafoetida* essential oil in controlling the black bean aphid. *Int. J. Biosci.* 5 (12), 350–356. doi:10.12692/ijb/ 5.12.350-356
- Bagheri, S. M., Asl, A. A., Shams, A., Mirghanizadeh-Bafghi, S. A., and Hafizibarjin, Z. (2017a). Evaluation of cytotoxicity effects of oleo-gum-resin and its essential oil of *Ferula assa-foetida* and ferulic acid on 4T1 breast cancer cells. *Indian J. Med. Paediatr. Oncol.* 38 (2), 116. doi:10.4103/ijmpo.ijmpo_60_16
- Bagheri, S. M., Dashti-R, M. H., and Morshedi, A. (2014a). Antinociceptive effect of Ferula assa-foetida oleo-gum-resin in mice. Res. Pharm. Sci. 9 (3), 207.
- Bagheri, S. M., Rezvani, M. E., Vahidi, A. R., and Esmaili, M. (2014b). Anticonvulsant effect of *Ferula assa-foetida* oleo gum resin on chemical and amygdala-kindled rats. *N. Am. J. Med. Sci.* 6 (8), 408–412. doi:10.4103/1947-2714.139296
- Bagheri, S. M., Hejazian, S. H., and Dashti-R, M. H. (2014c). The relaxant effect of seed fs essential oil and oleo-gum-resin of *Ferula assa-foetida* on isolated rat's ileum. *Ann. Med. Health Sci. Res.* 4 (2), 238–241. doi:10.4103/2141-9248. 129050
- Bagheri, S. M., Hedesh, S. T., Mirjalili, A., and Dashti-R, M. H. (2016). Evaluation of anti-inflammatory and some possible mechanisms of antinociceptive effect of *Ferula assa foetida* oleo gum resin. J. Evid. Based Complem. Altern. Med. 21 (4), 271–276. doi:10.1177/2156587215605903
- Bagheri, S. M., Sahebkar, A., Gohari, A. R., Saeidnia, S., Malmir, M., and Iranshahi, M. (2010). Evaluation of cytotoxicity and anticonvulsant activity of some Iranian medicinal *Ferula* species. *Pharm. Biol.* 48 (3), 242–246. doi:10.3109/ 13880200903081796
- Bagheri, S. M., Mohamadsadeghi, H., and Hejazian, E. S. (2017b). Antinociceptive effect of seed's essential oil of *Ferula assa-foetida* in mice. *Int. J. Clin. Exp. Pathol.* 4 (1), 34–37. doi:10.4103/2141-9248.129050
- Bagheri, S. M., Yadegari, M., Zare-Mohazabiye, F., Momeni-Asl, H., Mirjalili, A., Anvari, M., et al. (2018). Effect of *Ferula assa foetida* oleo-gum-resin on gastric ulcer in indomethacin-ulcerated rats. *Int. J. Curr. Res. Sci. Med.* 4 (1), 42–46. doi:10.4103/jcrsm.jcrsm_48_17
- Bahrami, R., Kocheili, F., and Ziaee, M. (2016). Fumigant toxicity and persistence of essential oils from asafetida, geranium, and walnut on adults of *Rhyzopertha dominica* (Col.: bostrichidae). *Toxin Rev.* 35 (3–4), 63–68. doi:10.1080/ 15569543.2016.1217542
- Bakkali, F., Averbeck, S., Averbeck, D., and Idaomar, M. (2008). Biological effects of essential oils--a review. *Food Chem. Toxicol.* 46 (2), 446–475. doi:10.1016/j. fct.2007.09.106
- Barupal, T., Meena, M., and Sharma, K. (2019). Inhibitory effects of leaf extract of Lawsonia inermis on Curvularia lunata and characterization of novel inhibitory compounds by GC–MS analysis. Biotechnol. Rep. 23, e00335. doi:10.1016/j.btre. 2019.e00335
- Bashir, S., Alam, M., Adhikari, A., Shrestha, R. L. S., Yousuf, S., Ahmad, B., et al. (2014). New antileishmanial sesquiterpene coumarins from *Ferula narthex* Boiss. *Phytochem. Lett.* 9, 46–50. doi:10.1016/j.phytol.2014.04.009
- Batra, S., Kumar, A., and Sharma, A. (2020). GABA and 5-HT receptor mediated anxiolytic effect of essential oil of *Ferula sumbul* Hook. roots. *Nat. Prod. J.* 10 (3), 262–271. doi:10.2174/2210315509666190211123646
- Bayrami, G., Boskabady, M. H., Iranshahi, M., and Gholamnezhad, Z. (2013). Relaxant effects of asafoetida extract and its constituent umbelliprenin on Guinea-pig tracheal smooth muscle. *Chin. J. Integr. Med.* 1–6, 28 doi:10.1007/ s11655-013-1550-3
- Ben Salem, S., Znati, M., Jabrane, A., Casanova, J., and Ben Jannet, H. (2016). Chemical composition, antimicrobial, anti-acetylcholinesterase and cytotoxic activities of the root essential oil from the Tunisian *Ferula lutea* (Poir.). *Maire* (*Apiaceae*). 19 (4), 897–906. doi:10.1080/0972060X.2015.1137238
- Benchabane, O., Hazzit, M., Baaliouamer, A., and Mouhouche, F. (2012). Analysis and antioxidant activity of the essential oils of *Ferula vesceritensis* Coss.et Dur. and *Thymus munbyanus* Desf. J. Essent. Oil-Bear. Plants. 15 (5), 774–781. doi:10.1080/0972060X.2012.10644119
- Benchabane, O. T. (2014). Chemical composition and insecticidal activities of essential oils of two Algerian endemic plants: *Ferula vesceritensis* Coss. et Dur.

and Thymus pallescens de Noe. Int. J. Agric. Sci. Res. 4 (6), 185–191. doi:10.1080/10942912.2017.1381112

- Chandran, H., Meena, M., and Sharma, K. (2020a). Microbial biodiversity and bioremediation assessment through omics approaches. *Front. Environ. Chem.* 1, 570326. doi:10.3389/fenvc.2020.570326
- Chandran, H., Meena, M., Barupal, T., and Sharma, K. (2020b). Plant tissue culture as a perpetual source for production of industrially important bioactive compounds. *Biotechnol. Rep. (Amst).* 26, e00450. doi:10.1016/j. btre.2020.e00450
- Dadaşoğlu, E., Öztekin, A., and Dadaşoğlu, F. (2018). Antibacterial and antioxidant activity of essential oil and extracts of *Ferula communis* and determination of chemical composition of its essential oil. *Fresenius Environ. Bull.* 27 (6), 4186–4191. doi:10.1080/14786419.2015.1071365
- Daneshkazemi, A., Zandi, H., Davari, A., Vakili, M., Emtiazi, M., Lotfi, R., et al. (2019). Antimicrobial activity of the essential oil obtained from the seed and oleo-gum-resin of *Ferula assa-foetida* against oral pathogens. *Front. Dent.* 16 (2), 113. doi:10.18502/fid.v16i2.1362
- Dastan, D., Salehi, P., Ghanati, F., Gohari, A. R., Maroofi, H., and Alnajar, N. (2014). Phytotoxicity and cytotoxicity of disesquiterpene and sesquiterpene coumarins from *Ferula pseudalliacea*. *Ind. Crop. Prod.* 55, 43–48. doi:10.1016/j. indcrop.2014.01.051
- Deniz, G. Y., Laloglu, E., Koc, K., and Geyikoglu, F. (2019). Hepatoprotective potential of *Ferula communis* extract for carbon tetrachloride induced hepatotoxicity and oxidative damage in rats. *Biotech. Histochem.* 94 (5), 334–340. doi:10.1080/10520295.2019.1566831
- Deveci, E., Tel-Çayan, G., and Duru, M. E. (2018). Phenolic profile, antioxidant, anticholinesterase, and anti-tyrosinase activities of the various extracts of *Ferula elaeochytris* and *Sideritis stricta*. *Int. J. Food Prop.* 21 (1), 771–783. doi:10.1080/ 10942912.2018.1431660
- Divya, K., Ramalakshmi, K., Murthy, P. S., and Rao, L. J. M. (2014). Volatile oils from *Ferula asafoetida* varieties and their antimicrobial activity. *LWT Food Sci. Technol. (Lebensmittel-Wissenschaft -Technol.).* 59 (2), 774–779. doi:10.1016/j. lwt.2014.07.013
- El-Deeb, H. K., Al Khadrawy, F. M., and Abd El-Hameid, A. K. (2012). Inhibitory effect of *Ferula asafoetida* L. (Umbelliferae) on *Blastocystis* sp. subtype 3 growth *in vitro*. *Parasitol. Res.* 111 (3), 1213–1221. doi:10.1007/s00436-012-2955-1
- Elghwaji, W., El-Sayed, A. M., El-Deeb, K. S., and ElSayed, A. M. (2017). Chemical composition, antimicrobial and antitumor potentiality of essential oil of *Ferula tingitana* L. Apiaceae grow in Libya. *Phcog. Mag.* 13 (3), 446–451. doi:10.4103/ pm.pm_323_15
- Esmaeili, H., Esmailidehaj, M., Entezari Zarch, S., and Azizian, H. (2020). Role of the potassium channels in vasorelaxant effect of asafoetida essential oil. *Avicenna J. Phytomed.* 10 (4), 407–416. doi:10.1016/j.jep.2004.01.002
- Esmaeili, H., Hafezimoghadam, Z., Esmailidehaj, M., Rezvani, M. E., and Hafizibarjin, Z. (2018). The effect of asafoetida essential oil on myocardial ischemic-reperfusion injury in isolated rat hearts. *Avicenna J. Phytomed.* 8 (4), 338–349. doi:10.22038/AJP.2018.10315
- Esmaeili, H., Sharifi, M., Esmailidehaj, M., Rezvani, M. E., and Hafizibarjin, Z. (2017). Vasodilatory effect of Asafoetida essential oil on rat aorta rings: the role of nitric oxide, prostacyclin, and calcium channels. *Phytomedicine*. 36, 88–94. doi:10.1016/j.phymed.2017.10.002
- Essid, R., Rahali, F. Z., Msaada, K., Sghair, I., Hammani, M., Bouratbine, A., et al. (2015). Antileishmanial and cytotoxic potential of essential oils from medicinal plants in Northern Tunisia. *Ind. Crop. Prod.* 77, 795–802. doi:10.1016/j.indcrop. 2015.09.049
- Estekhdami, P., Dehsorkhi, A. N., and Kalvandi, R. (2020). Insecticidal efficacy of essential oils from Cinnamomum zeylanicum, Thymus vulgaris, Ferula assafoetida L on Callosobruchus maculatus F. Asian J. Agric. Res. 13 (2), 52–62. doi:10.9734/ajaar/2020/v13i230103
- Fallah, F., Emadi, F., Ayatollahi, A., Taheri, S., Yazdi, M. K., and Rad, P. K. (2015). The anti-mycobacterial activity of the extract of *Ferula gummosa*. *Int. J. Mycobacteriol.* 4 (1), 166. doi:10.1016/j.ijmyco.2014.11.054
- Fatemikia, S., Abbasipour, H., and Saeedizadeh, A. (2017). Phytochemical and acaricidal study of the galbanum, *Ferula gummosa* boiss. (Apiaceae) essential oil against *Tetranychus urticae* koch (tetranychidae). *J. Essent. Oil-Bear. Plants.* 20 (1), 185–195. doi:10.1080/0972060X.2016.1257957
- Fatima, N., Fatmi, N., Shahzada, M., Sharma, S., Kumar, R., and Ali, M. (2017). Hepatoprotective effect of *Ferula assafoetida* against arsenic induced toxicity in

Swiss albino mice. J. Drug Discov. Dev. Deliv. 4 (1), 1030. doi:10.7324/JAPS. 2019.91107

- Ghanbari, M., Zahedi Khorasani, M., and Vakili, A. (2012). Acute and chronic effects of *Ferula persica* on blood pressure of hypertensive rats and its possible mechanism of action. J. Med. Plants. 3 (43), 62–68. doi:10.31925/farmacia.2020.3.9
- Ghannadi, A., Fattahian, K., Shokoohinia, Y., Behbahani, M., and Shahnoush, A. (2014). Anti-viral evaluation of sesquiterpene coumarins from *Ferula assa-foetida* against HSV-1. *Iran. J. Pharm. Res. (IJPR)*. 13 (2), 523. doi:10.22037/ijpr. 2014.1497
- Ghasemi, V., Yazdi, A. K., Tavallaie, F. Z., and Sendi, J. J. (2014). Effect of essential oils from *Callistemon viminalis* and *Ferula gummosa* on toxicity and on the hemocyte profile of *Ephestia kuehniella* (Lep.: Pyralidae). Arch. Phytopathol. Pflanzenschutz. 47 (3), 268–278. doi:10.1080/03235408.2013.808856
- Gholamnezhad, Z., Ghorani, V., Saadat, S., Shakeri, F., and Boskabady, M. H. (2018). The effects of medicinal plants on muscarinic receptors in various types of smooth muscle. *Phytother Res.* 32 (12), 2340–2363. doi:10.1002/ptr.6179
- Goldansaz, S. H., Talaei, L., Poorjavad, N., and Dehghani, Y. H. (2012). "Inhibition of carob moth damage using *Ferula assafoetida* essential oil in pomegranate orchards of Iran", in 2nd International symposium on the pomegranate, vol. 103, 129–131.
- Han, H. Y., Li, G. Y., Wang, H. Y., Ma, Q. D., Gao, J. B., and Wang, J. H. (2010). Anticoagulant activity of radiatinol and scopoletin from *Ferula dissecta* (Ledeb.) Ledeb. *in vitro*. J. Nongken Med. 2, 5. doi:10.11889/j.1000-3436.2016.rrj.34.020203
- Hassanabadi, M., Ebrahimi, M., Farajpour, M., and Dejahang, A. (2019). Variation in essential oil components among Iranian *Ferula assa-foetida* L. accessions. *Ind. Crop. Prod.* 140, 111598. doi:10.1016/j.indcrop.2019.111598
- Heydari-Majd, M., Rezaeinia, H., Shadan, M. R., Ghorani, B., and Tucker, N. (2019). Enrichment of zein nanofibre assemblies for therapeutic delivery of Barije (*Ferula gummosa* Boiss) essential oil. J. Drug Deliv. Sci. Technol. 54, 101290. doi:10.1016/j.jddst.2019.101290
- Hosseinpour, M. H., Askarianzadeh, A., Moharramipour, S., and Jalali Sendi, S. (2011). Insecticidal activity of essential oils isolated from Rue (*Ruta graveolens* L.) and Galbanum (*Ferula gummosa* Bioss.) on *Callosobruchus maculatus* (F). *Integr. Prot. Stored Prod.* 69, 271–275. doi:10.3390/molecules24224047
- Hosseinzadeh, N., Shomali, T., Hosseinzadeh, S., Raouf Fard, F., Pourmontaseri, M., and Fazeli, M. (2020a). Green synthesis of gold nanoparticles by using *Ferula persica* Willd. gum essential oil: production, characterization and *in vitro* anti-cancer effects. *J. Pharm. Pharmacol.* 72 (8), 1013–1025. doi:10.1111/jphp. 13274
- Hosseinzadeh, N., Shomali, T., Hosseinzadeh, S., Raouf Fard, F., Jalaei, J., and Fazeli, M. (2020b). Cytotoxic activity of *Ferula persica* gum essential oil on murine colon carcinoma (CT26) and Vero cell lines. *J. Essent. Oil Res.* 32 (2), 169–177. doi:10.1080/10412905.2020.1729880
- Hussain, A. I., Anwar, F., Hussain Sherazi, S. T., and Przybylski, R. (2008). Chemical composition, antioxidant and antimicrobial activities of basil (*Ocimum basilicum*) essential oils depends on seasonal variations. *Food Chem.* 108 (3), 986–995. doi:10.1016/j.foodchem.2007.12.010
- Iranshahi, M., Rezaee, R., Najaf Najafi, M., Haghbin, A., and Kasaian, J. (2018). Cytotoxic activity of the genus *Ferula* (Apiaceae) and its bioactive constituents. *Avicenna J. Phytomed.* 8 (4), 296–312. doi:10.22038/AJP.2018.26953.1963
- Iranshahi, M., and Alizadeh, M. (2012). Antihyperglycemic effect of asafoetida (*Ferula assafoetida* oleo-gum-resin) in streptozotocin-induced diabetic rats. World Appl. Sci. J. 17 (2), 157–162. doi:10.1016/j.jtcme.2016.11.004
- Iranshahi, M., Fata, A., Emami, B., Shahr, B. M. J., and FazlyBazzaz, B. S. (2008). In vitro antifungal activity of polysulfides-rich essential oil of Ferula latisecta fruits against human pathogenic dermatophytes. Nat. Prod. Commun. 3, 1543–1546. doi:10.1177/2F1934578X0800300929
- Iranshahy, M., Farhadi, F., Paknejad, B., Zareian, P., Iranshahi, M., Karami, M., et al. (2019). Gummosin, a sesquiterpene coumarin from *Ferula assa-foetida* is preferentially cytotoxic to human breast and prostate cancer cell lines. *Avicenna J. Phytomed.* 9 (5), 446–453. doi:10.22038/AJP.2019.12598
- Iranshahy, M., and Iranshahi, M. (2011). Traditional uses, phytochemistry and pharmacology of asafoetida (*Ferula assa-foetida* oleo-gum-resin)—a review. J. Ethnopharmacol. 134 (1), 1–10. doi:10.1016/j.jep.2010.11.067
- Izadi, H., Sarnevesht, M., Sadeghi, R., Mahdian, K., and Jalai, M. A. (2012). Toxic effects of pyriproxyfen, neemarin, acetamiprid and *Ferula assafoetida* essential oil on the common *pistachio psylla*, *Agonoscena pistaciae*. Arch. Phytopathol. Pflanzenschutz. 45 (18), 2236–2242. doi:10.1080/03235408.2012.724973

- Jahani, S., Salehi, M., Shakiba, A., Moradipour, A., and Forouzandeh, F. (2015). Production and study of antioxidant and antibacterial activities of gelatin nanocapsules containing *Ferula assa-foetida* essential oil. *J. Arak Uni. Med. Sci.* 18 (5), 33–48. doi:10.18502/fid.v16i2.1362
- Juglal, S., Govinden, R., and Odhav, B. (2002). Spice oils for the control of cooccurring mycotoxin-producing fungi. J. Food Protect. 65, 683–687. doi:10. 4315/0362-028x-65.4.683
- Kahraman, C., Topcu, G., Bedir, E., Tatli, I. I., Ekizoglu, M., and Akdemir, Z. S. (2019). Phytochemical screening and evaluation of the antimicrobial and antioxidant activities of *Ferula caspica*. M. Bieb. extracts. Saudi. Pharm. J. 27 (4), 525–531. doi:10.1016/j.jsps.2019.01.016
- Karakaya, S., Göger, G., Bostanlik, F. D., Demirci, B., Duman, H., and Kilic, C. S. (2019a). Comparison of the essential oils of *Ferula orientalis L., Ferula gosandrasica* Peşmen and Quézel, and *Hippomarathrum microcarpum* Petrov and their antimicrobial activity. *Turk. J. Pharm. Sci.* 16 (1), 69–75. doi:10.4274/tjps.77200
- Karakaya, S., Koca, M., Sytar, O., and Duman, H. (2019b). Determination of natural phenolic compounds of *Ferula longipedunculata* Peşmen and assessment their antioxidant and anticholinesterase potentials. *Nat. Prod. Res.* 14 (5), 1–3. doi:10.1080/14786419.2019.1619728
- Karimi, A., Krähmer, A., Herwig, N., Hadian, J., Schulz, H., and Meiners, T. (2020). Metabolomics approaches for analysing effects of geographic and environmental factors on the variation of root essential oils of *Ferula assafoetida* L. J. Agric. Food Chem. 37, 1–31. doi:10.1021/acs.jafc.0c03681
- Karimian, V., Sepehry, A., Barani, H., Ebrahimi, S. N., and Mirjalili, M. H. (2020). Productivity, essential oil variability and antioxidant activity of *Ferula assa-foetida* L. oleo-gum-resin during the plant exploitation period. *J. Essent. Oil Res.* 1–11, 28. doi:10.1080/10412905.2020.1794988
- Karimlar, S., Naderi, A., Mohammadi, F., Moslehishad, M., Delrish, E., Aghajanpour, L., et al. (2019). Hypoglycemic and hypolipidemic effects of *Myrtus communis, Trachyspermum copticum* and *Ferula gummosa* essential oils on streptozotocin induced diabetic rats. *Int. J. Food Sci. Nutr.* 6 (1), 1–8. doi:10. 29252/nfsr.6.1.1
- Kasaian, J., Asili, J., and Iranshahi, M. (2016). Sulphur-containing compounds in the essential oil of *Ferula alliacea* roots and their mass spectral fragmentation patterns. *Pharm. Biol.* 54 (10), 2264–2268. doi:10.3109/13880209.2016.1152279
- Kavoosi, G., Purfard, A. M., and Aram, F. (2012). Radical scavenging properties of essential oils from *Zataria multiflora* and *Ferula assafoetida*. Asian Pac. J. Trop. Biomed. 2 (3), 28. doi:10.1016/S2221-1691(12)60415-8
- Kavoosi, G., and Purfard, A. M. (2013). Scolicidal effectiveness of essential oil from Zataria multiflora and Ferula assafoetida: disparity between phenolic monoterpenes and disulphide compounds. Comp. Clin. Pathol. 22 (5), 999–1005. doi:10.1007/s00580-012-1518-2
- Kavoosi, G., and Rowshan, V. (2013). Chemical composition, antioxidant and antimicrobial activities of essential oil obtained from *Ferula assa-foetida* oleogum-resin: effect of collection time. *Food Chem.* 138 (4), 2180–2187. doi:10. 1016/j.foodchem.2012.11.131
- Kavoosi, G., Shakiba, A., Ghorbani, M., Dadfar, S. M. M., and Purfard, A. M. (2015). Antioxidant, antibacterial, water binding capacity and mechanical behavior of gelatin-ferula oil film as a wound dressing material. *Galen Medical J.* 4 (2), 103–114.
- Kavoosi, G., Tafsiry, A., Ebdam, A. A., and Rowshan, V. (2013). Evaluation of antioxidant and antimicrobial activities of essential oils from *Carum copticum* seed and *Ferula assafoetida* latex. J. Food Sci. 78 (2), 46. doi:10.1111/1750-3841.12020
- Khajeh, M., Yamini, Y., Bahramifar, N., Sefidkon, F., and Pirmoradei, M. R. (2005). Comparison of essential oils compositions of *Ferula assa-foetida* obtained by supercritical carbon dioxide extraction and hydrodistillation methods. *Food Chem.* 91 (4), 639–644. doi:10.1016/j.foodchem.2004.06.033
- Khalifaev, P. D., Sharopov, F. S., Safomuddin, A., Numonov, S., Bakri, M., Habasi, M., et al. (2018). Chemical composition of the essential oil from the roots of *Ferula kuhistanica* growing wild in Tajikistan. *Nat. Prod. Commun.* 13 (2), 220–222. doi:10.1177/1934578X1801300226
- Khambhala, P., Verma, S., Joshi, S., Seshadri, S., and Kothari, V. (2016). Inhibition of bacterial quorum-sensing by *Ferula asafoetida* essential oil. *Adv. Genet. Eng.* 5 (2), 2169–0111. doi:10.4172/2169-0111.1000i105
- Khazdair, M. R., Boskabady, M. H., Kiyanmehr, M., Hashemzehi, M., and Iranshahi, M. (2015). The inhibitory effects of *Ferula assa-foetida* on muscarinic receptors of Guinea-Pig tracheal smooth muscle. *Jundishapur J. Nat. Pharm. Prod.* 10 (3), e20008 doi:10.17795/jjnpp-20008

- Khoury, M., El Beyrouthy, M., Eparvier, V., Ouaini, N., and Stien, D. (2018). Chemical diversity and antimicrobial activity of the essential oils of four Apiaceae species growing wild in Lebanon. J. Essent. Oil Res. 25–31, 55. doi:10.1080/10412905.2017.1372314
- Kiasalari, Z., Khalili, M., Roghani, M., Heidari, H., and Azizi, Y. (2013). Antiepileptic and antioxidant effect of hydroalcoholic extract of *Ferula assa foetida* gum on pentylentetrazole-induced kindling in male mice. *Basic Clin. Neurosci.* 4 (4), 299–306.
- Kim, J. W., Huh, J. E., Kyung, S. H., and Kyung, K. H. (2004). Antimicrobial activity of alk(en)yl sulfides found in essential oils of garlic and onion. *Food Sci. Biotechnol.* 13 (2), 235–239.
- Koorki, Z., Shahidi-Noghabi, S., Mahdian, K., and Pirmaoradi, M. (2018). Chemical composition and insecticidal properties of several plant essential oils on the melon aphid, *Aphis gossypii* Glover (Hemiptera: aphididae). *J. Essent. Oil-Bear. Plants.* 21 (2), 420–429. doi:10.1080/0972060X.2018.1435308
- Kose, E. O., Akta, Ö., Deniz, I. G., and Sarikürkçü, C. (2010). Chemical composition antimicrobial and antioxidant activity of essential oil of endemic *Ferula lycia* Boiss. J. Med. Plants Res. 4 (17), 1698–1703. doi:10. 5897/JMPR09.439
- Kouyakhi, E. T., Naghavi, M. R., and Alayhs, M. (2008). Study of the essential oil variation of *Ferula gummosa* samples from Iran. *Chem. Nat. Compd.* 44 (1), 124–126. doi:10.1007/s10600-008-0038-4
- Kovač, J., Šimunović, K., Wu, Z., Klančnik, A., Bucar, F., Zhang, Q., et al. (2015). Antibiotic resistance modulation and modes of action of (-)-α-pinene in *Campylobacter jejuni. PloS One.* 10, e0122871. doi:10.1371/journal.pone.0122871
- Labed-Zouad, I., Labed, A., Laggoune, S., Zahia, S., Kabouche, A., and Kabouche, Z. (2015). Chemical compositions and antibacterial activity of four essential oils from *Ferula vesceritensis* Coss. and Dur. against clinical isolated and food-borne pathogens. *Record Nat. Prod.* 9 (4), 518–525. doi:10.1177/1934578X1801300226
- Latifi, E., Mohammadpour, A. A., Fathi, B., and Nourani, H. (2019). Antidiabetic and antihyperlipidemic effects of ethanolic *Ferula assa-foetida* oleo-gum-resin extract in streptozotocin-induced diabetic wistar rats. *Biomed. Pharmacother*. 110, 197–202. doi:10.1016/j.biopha.2018.10.152
- Liu, T., Wang, C. J., Xie, H. Q., and Mu, Q. (2013). Guaiol-a naturally occurring insecticidal sesquiterpene. *Nat. Prod. Commun.* 8 (10), 1353–1354. doi:10.1177/ 1934578X1300801001
- Liu, T., Wang, L., Zhang, L., Jiang, H., Zhang, Y., and Mao, L. (2020). Insecticidal, cytotoxic and anti-phytopathogenic fungal activities of chemical constituents from the aerial parts of *Ferula sinkiangensis*. *Nat. Prod. Res.* 34 (10), 1430–1436. doi:10.1080/14786419.2018.1509328
- Mahaki, H., Tanzadehpanah, H., Abou-Zied, O. K., Moghadam, N. H., Bahmani, A., Salehzadeh, S., et al. (2019). Cytotoxicity and antioxidant activity of Kamolonol acetate from *Ferula pseudalliacea*, and studying its interactions with calf thymus DNA (ct-DNA) and human serum albumin (HSA) by spectroscopic and molecular docking techniques. *Process Biochem*. 79, 203–213. doi:10.1016/j.procbio.2018.12.004
- Mahboubi, M., Kazempour, N., and Mahboubi, M. (2011). Antimicrobial activity of Rosemary, Fennel and Galbanum essential oils against clinical isolates of *Staphylococcus aureus. Biharean Biol.* 5 (1), 4–7.
- Mahendra, P., and Bisht, S. (2012). Ferula asafoetida: traditional uses and pharmacological activity. Pharmacogn. Rev. 6 (12), 141. doi:10.4103/0973-7847.99948
- Malekzadeh, M., Angourani, H. R., Yazdinezhad, A., Hassan, M., Abiodun, F., and Hazrati, S. (2018). Evaluation of volatile oil in indigenous populations of *Ferula* gummosa boiss. J. Essent. Oil-Bear. Plants. 21 (1), 206–213. doi:10.1080/ 0972060X.2016.1244495
- Mallahi, T., Ramezanian, A., Saharkhiz, M. J., Javanmardi, J., and Iraji, A. (2018). Antimicrobial activities of asafoetida and shirazi thyme essential oils improve the vase life of gerbera cut flowers. *Acta Ecol. Sin.* 38 (3), 228–233. doi:10.1016/j. chnaes.2017.08.009
- Meena, M., Aamir, M., Vikas, K., Swapnil, P., and Upadhyay, R. S. (2018). Evaluation of morpho-physiological growth parameters of tomato in response to Cd induced toxicity and characterization of metal sensitive NRAMP3 transporter protein. *Environ. Exp. Bot.* 148, 144–167. doi:10.1016/ j.envexpbot.2018.01.007
- Meena, M., and Samal, S. (2019). Alternaria host-specific (HSTs) toxins: an overview of chemical characterization, target sites, regulation and their toxic effects. Toxicol. Rep. 6, 745–758. doi:10.1016/j.toxrep.2019.06.021

- Meena, M., Sonigra, P., and Yadav, G. (2020a). Biological-based methods for the removal of volatile organic compounds (VOCs) and heavy metals. *Environ. Sci. Pollut. Res.* 11, 124. doi:10.1007/s11356-020-11112-4
- Meena, M., Swapnil, P., Divyanshu, K., Kumar, S., Harish, Y. N., et al. (2020b). PGPR-mediated induction of systemic resistance and physiochemical alterations in plants against the pathogens: current perspectives. J. Basic Microbiol. 60 (8), 1–34. doi:10.1002/jobm.202000370
- Meena, M., and Swapnil, P. (2019). Regulation of WRKY genes in plant defense with beneficial fungus *Trichoderma*: current perspectives and future prospects. *Arch. Phytopathol. Plant Protect.* 52 (1-2), 1–17. doi:10.1080/03235408.2019.1606490
- Meena, M., Swapnil, P., Zehra, A., Dubey, M. K., and Upadhyay, R. S. (2017a). Antagonistic assessment of *Trichoderma* spp. by producing volatile and nonvolatile compounds against different fungal pathogens. Arch. Phytopathol. Plant Protect. 50 (13–14), 629–648. doi:10.1080/03235408.2017.1357360
- Meena, M., Gupta, S. K., Swapnil, P., Zehra, A., Dubey, M. K., and Upadhyay, R. S. (2017b). Alternaria toxins: potential virulence factors and genes related to pathogenesis. Front. Microbiol. 8, 1451. doi:10.3389/fmicb.2017.01451
- Meena, M., Swapnil, P., and Upadhyay, R. S. (2017c). Isolation, characterization and toxicological potential of tenuazonic acid, alternariol and alternariol monomethyl ether produced by *Alternaria* species phytopathogenic on plants. *Sci. Rep.* 7, 8777. doi:10.1038/s41598-017-09138-9
- Meena, M., Zehra, A., Dubey, M. K., Aamir, M., Gupta, V. K., and Upadhyay, R. S. (2016). Comparative evaluation of biochemical changes in tomato (*Lycopersicon esculentum* Mill.) infected by *Alternaria alternata* and its toxic metabolites (TeA, AOH, and AME). *Front. Plant Sci.* 7, 1408. doi:10.3389/fpls.2016.01408
- Moghaddam, M., and Farhadi, N. (2015). Influence of environmental and genetic factors on resin yield, essential oil content and chemical composition of *Ferula* assa-foetida L. populations. J. Appl. Res. Med. Aromat. Plants. 2 (3), 69–76. doi:10.1016/j.jarmap.2015.04.001
- Mohammadhosseini, M. (2016). A Comprehensive Review on new methods for processing, separation and identification of the essential oils. Shahrood, Iran: Islamic Azad University of Shahrood Press, 61–73. doi:10.1016/j.indcrop.2017. 05.009
- Mohammadhosseini, M., Akbarzadeh, A., and Flamini, G. (2017). Profiling of compositions of essential oils and volatiles of *Salvia limbata* using traditional and advanced techniques and evaluation for biological activities of their extracts. *Chem. Biodivers.* 14 (5), e1600361. doi:10.1002/cbdv.201600361
- Mohammadhosseini, M. (2017). Essential oils extracted using microwave-assisted hydrodistillation from aerial parts of eleven Artemisia species: chemical compositions and diversities in different geographical regions of Iran. *Record Nat. Prod.* 11 (2), 114–129. https://www.acgpubs.org/doc/ 2018080513061416-RNP-EO_1602-010.pdf.
- Mohammadhosseini, M., Mahdavi, B., and Shahnama, M. (2015). Chemical composition of essential oils from aerial parts of *Ferula gummosa* (Apiaceae) in Jajarm Region, Iran using traditional hydrodistillation and solvent-free microwave extraction methods: a comparative approach. J. Essent. Oil. Bear. Plants. 18 (6), 1321–1328. doi:10.1080/0972060X.2015.1024445
- Mohammadhosseini, M., and Nekoei, M. (2014). Chemical compositions of the essential oils and volatile compounds from the aerial parts of *Ferula ovina* using hydrodistillation, MAHD, SFME and HS-SPME methods. *J. Essent. Oil Bear. Plants.* 17 (5), 747–757. doi:10.1080/0972060X.2014.884951
- Mohammadhosseini, M., Venditti, A., and Akbarzadeh, A. (2019a). The genus Perovskia Kar.: ethnobotany, chemotaxonomy and phytochemistry: a review. Toxin Rev. 28, 1–22. doi:10.1080/15569543.2019.1691013
- Mohammadhosseini, M., Venditti, A., Sarker, S. D., Nahar, L., and Akbarzadeh, A. (2019b). The genus *Ferula*: ethnobotany, phytochemistry and bioactivities–A review. *Ind. Crop. Prod.* 129, 350–394. doi:10.1016/j.indcrop.2018.12.012
- Mohammadi, S., Ebrahimzadeh, H., Niknam, V., and Zahed, Z. (2019). Agedependent responses in cellular mechanisms and essential oil production in sweet *Ferula assafoetida* under prolonged drought stress. *J. Plant Interact.* 14 (1), 324–333. doi:10.1080/17429145.2019.1632946
- Muturi, E. J., Ramirez, J. L., Zilkowski, B., Flor-Weiler, L. B., and Rooney, A. P. (2018). Ovicidal and larvicidal effects of garlic and asafoetida essential oils against West Nile virus vectors. J. Insect Sci. 18 (2), 1–6. doi:10.1093/jisesa/iey036
- Nabavi, S. M., Ebrahimzadeh, M. A., Nabavi, S. F., Eslami, B., and Dehpour, A. A. (2011). Antioxidant and antihaemolytic activities of *Ferula foetida* regel (Umbelliferae). *Eur. Rev. Med. Pharmacol. Sci.* 15 (2), 157–164. https:// www.europeanreview.org/article/887.

- Nazemisalman, B., Vahabi, S., Yazdinejad, A., Haghghi, F., Jam, M. S., and Heydari, F. (2018). Comparison of antimicrobial effect of *Ziziphora tenuior*, *Dracocephalum moldavica*, *Ferula gummosa*, and *Prangos ferulacea* essential oil with chlorhexidine on *Enterococcus faecalis*: an *in vitro* study. J. Dent. Res. 15 (2), 111–116. doi:10.4103/1735-3327.226525
- Nguir, A., Mabrouk, H., Douki, W., Ismail, M. B., Jannet, H. B., and Flamini, G. (2016). Chemical composition and bioactivities of the essential oil from different organs of *Ferula communis* L. growing in Tunisia. *Med. Chem. Res.* 25 (3), 515–525. doi:10.1007/s00044-016-1506-1
- Oüzek, G., Schepetkin, I. A., Utegenova, G. A., Kirpotina, L. N., andrei, S. R., Oüzek, T., et al. (2017). Chemical composition and phagocyte immunomodulatory activity of *Ferula iliensis* essential oils. *J. Leukoc. Biol.* 101 (6), 1361–1371. doi:10.1189/jlb.3A1216-518RR
- Ozkan, H., Yanmis, D., Karadayi, M., Bal, T., Baris, O., and Gulluce, M. (2014).
 Determination of genotoxic and antigenotoxic properties of essential oil from *Ferula orientalis* L. using Ames/Salmonella and E. coli WP2 bacterial test systems. *Toxicol. Ind. Health.* 30 (8), 714–723. doi:10.1177/0748233712462479
- Pavela, R., Morshedloo, M. R., Lupidi, G., Carolla, G., Barboni, L., Quassinti, L., et al. (2020). The volatile oils from the oleo-gum-resins of *Ferula assa-foetida* and *Ferula gummosa*: a comprehensive investigation of their insecticidal activity and eco-toxicological effects. *Food Chem. Toxicol.* 140, 111312. doi:10.1016/j.fct.2020.111312
- Pavlović, I., Petrović, S., Radenković, M., Milenković, M., Couladis, M., Branković, S., et al. (2012). Composition, antimicrobial, antiradical and spasmolytic activity of *Ferula heuffelii* Griseb. ex Heuffel (Apiaceae) essential oil. *Food Chem.* 130 (2), 310–315. doi:10.1016/j.foodchem.2011.07.043
- Poorjavad, N., Goldansaz, S. H., and Dadpour, H. (2014). Effect of Ferula assafoetida essential oil on some biological and behavioral traits of Trichogramma embryophagum and T. evanescens. Biocontrol. 59 (4), 403–413. doi:10.1007/s10526-014-9583-x
- Prakash, B., Kedia, A., Mishra, P. K., and Dubey, N. K. (2015). Plant essential oils as food preservatives to control moulds, mycotoxin contamination and oxidative deterioration of agri-food commodities–potentials and challenges. *Food Contr.* 47, 381–391. doi:10.1016/j.foodcont.2014.07.023
- Radulović, N. S., Zlatković, D. B., Randjelović, P. J., Stojanović, N. M., Novaković, S. B., and Akhlaghi, H. (2013). Chemistry of spices: bornyl 4-methoxybenzoate from *Ferula ovina* (Boiss.) Boiss.(Apiaceae) induces hyperalgesia in mice. *Food Funct.* 4 (12), 1751–1758. doi:10.1039/C3FO60319A
- Ragab, T. I., El Gendy, A. N. G., Saleh, I. A., and Esawy, M. A. (2019). Chemical composition and evaluation of antimicrobial activity of the Origanum majorana essential oil extracted by microwave-assisted extraction, conventional hydro-distillation and steam distillation. J. Essent. Oil-Bear. Plants. 22 (2), 563–573. doi:10.1080/0972060X.2019.1611486
- Rahali, F. Z., Kefi, S., Bettaieb Rebey, I., Hamdaoui, G., Tabart, J., Kevers, C., et al. (2019). Phytochemical composition and antioxidant activities of different aerial parts extracts of *Ferula communis* L. *Plant Biosyst.* 153 (2), 213–221. doi:10. 1080/11263504.2018.1461696
- Rashidi, F., Khaksary, M. M., Ranjbar, R., and Najafzadeh, V. H. (2014). The Effects of essential oil of galbanum on caffeine induced-cleft palate in rat embryos. *Zahedan. J. Res. Med. Sci.* 16 (2), 37–41. doi:10.3329/bjp.v11i4.26915
- Raut, J. S., and Karuppayil, S. M. (2014). A status review on the medicinal properties of essential oils. *Ind. Crop. Prod.* 62, 250–264. doi:10.1016/j. indcrop.2014.05.055
- Rezaee, R., Behravan, E., Behravan, J., Soltani, F., Naderi, Y., Emami, B., et al. (2014). Antigenotoxic activities of the natural dietary coumarins umbelliferone, herniarin and 7-isopentenyloxy coumarin on human lymphocytes exposed to oxidative stress. *Drug Chem. Toxicol.* 37 (2), 144–148. doi:10.3109/01480545. 2013.834352
- Rivas da Silva, A. C., Lopes, P. M., Barros de Azevedo, M. M., Costa, D. C., Alviano, C. S., and Alviano, D. S. (2012). Biological activities of alpha-pinene and betapinene enantiomers. *Molecules*. 17, 6305–6316.
- Sadeghi, R., Hadizadeh Raeisi, N., and Jamshidnia, A. (2017). Immunological responses of Sesamia cretica to Ferula ovina essential oil. J. Insect Sci. 17 (1), 1–5. doi:10.1093/jisesa/iew124
- Sadraei, H., Asghari, G. R., Hajhashemi, V., Kolagar, A., and Ebrahimi, M. (2001). Spasmolytic activity of essential oil and various extracts of *Ferula gummosa* Boiss. on ileum contractions. *Phytomedicine*. 8 (5), 370–376. doi:10.1078/0944-7113-00052

- Safaeian, L., Ghannadi, A., Javanmard, S. H., and Vahidian, M. H. (2015). The effect of hydroalcoholic extract of *Ferula foetida* stems on blood pressure and oxidative stress in dexamethasone-induced hypertensive rats. *Res. Pharm. Sci.* 10 (4), 326. doi:10.4103/1735-5362.220964
- Sagyndykova, M. S., Imanbayeva, A. A., Suleimen, Y. M., and Ishmuratova, M. Y. (2019). Chemical composition and properties of essential oil of *Ferula foetida* (Bunge) Regel growing on Mangyshlak peninsula. 496, 27–34. doi:10.31489/ 2019Ch4/25-34
- Salehi, M., Naghavi, M. R., and Bahmankar, M. (2019). A review of *Ferula* species: biochemical characteristics, pharmaceutical and industrial applications, and suggestions for biotechnologists. *Ind. Crop. Prod.* 139, 111511. doi:10.1016/j. indcrop.2019.111511
- Satarian, F., Hosseini, H. M., Ghadaksaz, A., Amin, M., and Fooladi, A. A. (2018). Multi-drug resistant clinical *Pseudomonas aeruginosas* inhibited by *Ferula gummosa* Boiss. *Recent Pat. Anti-Infect. Drug Discov.* 13 (1), 89–99. doi:10. 2174/1574891X13666180426163427
- Sayah, M., Kamalinezhad, M., Roustaeian, A., and Bahrami, H. R. (2001). Antiepileptic potential and composition of the fruit essential oil of *Ferula gummosa* boiss. *Iran. Biomed. J.* 5 (2–3), 69–72.
- Schepetkin, I. A., Kushnarenko, S. V., Özek, G., Kirpotina, L. N., Sinharoy, P., Utegenova, G. A., et al. (2016). Modulation of human neutrophil responses by the essential oils from *Ferula akitschkensis* and their constituents. *J. Agric. Food Chem.* 64 (38), 7156–7170. doi:10.1021/acs.jafc.6b03205
- Sepahi, E., Tarighi, S., Ahmadi, F. S., and Bagheri, A. (2015). Inhibition of quorum sensing in *Pseudomonas aeruginosa* by two herbal essential oils from Apiaceae family. J. Microbiol. 53 (2), 176–180. doi:10.1007/s12275-015-4203-8
- Sharopov, F., Braun, M. S., Gulmurodov, I., Khalifaev, D., Isupov, S., and Wink, M. (2015). Antimicrobial, antioxidant, and anti-inflammatory activities of essential oils of selected aromatic plants from Tajikistan. *Foods.* 4 (4), 645–653. doi:10. 3390/foods4040645
- Sharopov, F. S., Khalifaev, P. D., Satyal, P., Sun, Y., Safomuddin, A., Musozoda, S., et al. (2019). The chemical composition and biological activity of the essential oil from the underground parts of *Ferula tadshikorum* (Apiaceae). *Record Nat. Prod.* 13 (1), 18–23. doi:10.25135/rnp.65.18.02.089
- Sitara, U., Akbar, A., Abid, M., and Ahmad, A. (2018). Essential oils show antifungal activity against seed-borne mycoflora associated with okra seeds. *Int. J. Biotechnol. Mol. Biol. Res.* 15, 855–863. doi:10.3923/ajppaj.2012.66.74
- Stepanycheva, E. A., Chakaeva, A. S., Savelieva, E. I., and Chermenskaya, T. D. (2012). Aphicidal activity of substances from roots of *Ferula foetida* (Bunge), Regel. against grain aphid, *Schizaphis graminum* (Rondani). *Biopestic. Int.* 8, 18–25.
- Tabari, M. A., Youssefi, M. R., Nasiri, M., Hamidi, M., Kiani, K., Samakkhah, S. A., et al. (2019). Towards green drugs against cestodes: effectiveness of *Pelargonium roseum* and *Ferula gummosa* essential oils and their main component on *Echinococcus granulosus* protoscoleces. *Vet. Parasitol.* 266, 84–87. doi:10.1016/j.vetpar.2018.12.019
- Tavassoli, M., Jalilzadeh-Amin, G., Fard, V. R. B., and Esfandiarpour, R. (2018). The *in vitro* effect of *Ferula asafoetida* and *Allium sativum* extracts on *Strongylus* spp. Ann. Parasitol. 64 (1), 59–63. doi:10.17420/ap6401.133
- Tempark, T., Chatproedprai, S., and Wananukul, S. (2016). Localized contact dermatitis from *Ferula assa-foetida* oleo-gum-resin essential oil, a traditional topical preparation for stomach ache and flatulence. *Indian J. Dermatol. Venereol. Leprol.* 82 (4), 467. doi:10.4103/0378-6323.182969
- Teng, L., Ma, G. Z., Li, L., Ma, L. Y., and Xu, X. Q. (2013). Karatavicinol a, a new anti-ulcer sesquiterpene coumarin from *Ferula sinkiangensis. Chem. Nat. Compd.* 49 (4), 606–609. doi:10.1007/s10600-013-0690-1
- Topdas, E. F., Sengul, M., Taghizadehghalehjoughi, A., and Hacimuftuoglu, A. (2020). Neuroprotective potential and antioxidant activity of various solvent extracts and essential oil of *Ferula orientalis* L. J. Essent. Oil-Bear. Plants. 23 (1), 121–138. doi:10.1080/0972060X.2020.1729247
- Upadhyay, P. K. (2017). Pharmacological activities and therapeutic uses of resins obtained from *Ferula asafoetida* Linn.: a review. *Int. J. Green Pharm.* 11, 2. doi:10.22377/ijgp.v11i02.1033
- Upadhyay, P. K., Singh, O., Yadav, A., and Sharma, R. (2014). A review on anxiolytic and antiepileptic effects of oleo–gum-resin of *Ferula asafoetida*. Am. J. Pharm. Heath Res. 2, 1–11. doi:10.4103/1947-2714.139296

- Utegenova, G. A., Pallister, K. B., Kushnarenko, S. V., Özek, G., Özek, T., Abidkulova, K. T., et al. (2018). Chemical composition and antibacterial activity of essential oils from *Ferula L.* species against methicillin-resistant *Staphylococcus aureus. Molecules*. 23 (7), 1679. doi:10.3390/molecules23071679
- Verma, S., Khambhala, P., Joshi, S., Kothari, V., Patel, T., and Seshadri, S. (2019). Evaluating the role of dithiolane rich fraction of *Ferula asafoetida* (apiaceae) for its antiproliferative and apoptotic properties: *in vitro* studies. *Exp. Oncol.* 4141 (22), 90–94. doi:10.32471/exp-oncology.2312-8852
- Yaqoob, U., and Nawchoo, I. A. (2016). Distribution and taxonomy of Ferula L.: a review. Res. Rev. J. Biotechnol. 5 (3), 15–23. doi:10.3390/plants9060740
- Yarizade, A., Kumleh, H. H., and Niazi, A. L. I. (2017). *In vitro* antidiabetic effects of *Ferula assa-foetida* extracts through dipeptidyl peptidase IV and α-glucosidase inhibitory activity. *In Vitro*. 10 (5), 357–360. doi:10.22159/ ajpcr.2017.v10i5.16648
- Ye, B., Wang, S., and Zhang, L. (2011). Studies on the detoxification effects and acute toxicity of a mixture of cis-sec-butyl-1-propoenyl disulphide and transsec-butyl-1-propoenyl disulphide isolated from crude essential oil of *Ferula sinkiangensis* KM Shen, a Chinese traditional herbal medicine. Nat. Prod. Res. 25 (12), 1161–1170. doi:10.1080/14786419.2010.550027
- Yusufoglu, H., Soliman, G., Abdel-Rahman, R., Abdel-Kader, M., Ganaie, M., Bedir, E., et al. (2015). Antihyperglycemic and antihyperlipidemic effects of *Ferula duranii* in experimental type 2 diabetic rats. *Int. J. Pharmacol.* 11 (6), 532–541. doi:10.3923/ijp.2015.532.541
- Zahani, F. H., and Khaledi, N. (2018). Biological effects of various essential oils on citrus decay pathogens. Int. J. New Technol. Res. 4 (4), 129–139. doi:10.1155/ 2017/9268468
- Zandi-Sohani, N., Rajabpour, A., Yarahmadi, F., and Ramezani, L. (2018). Sensitivity of *Bemisia tabaci* (Hemiptera: aleyrodidae) and the generalist predator *Orius albidipennis* (Hemiptera: anthocoridae) to vapors of essential oils. *J. Entomol. Sci.* 53 (4), 493–502. doi:10.18474/JES17-113.1
- Zellagui, A., Gherraf, N., and Rhouati, S. (2012). Chemical composition and antibacterial activity of the essential oils of *Ferula vesceritensis* Coss et Dur. leaves, endemic in Algeria. *Bioorg. Med. Chem. Lett.* 2 (1), 2191–2858. doi:10. 1186/2191-2858-2-31
- Zhai, L. L., Liu, T., Xie, H. Q., Xie, Y. H., and Mu, Q. (2012). Inhibition effects on Hepatitis B virus replication by hydrophobic extracts from *Ferula ferulaeoides* (Steud.) Korov. *J. Med. Plants Res.* 6 (8), 1486–1488. doi:10. 5897/JMPR11.553
- Zhou, Y., Xin, F., Zhang, G., Qu, H., Yang, D., and Han, X. (2017). Recent advances on bioactive constituents in *Ferula*. Drug Dev. Res. 78 (7), 321–331. doi:10.1002/ ddr.21402
- Znati, M., Filali, I., Jabrane, A., Casanova, J., Bouajila, J., and Ben Jannet, H. (2017). Chemical composition and *in vitro* evaluation of antimicrobial, antioxidant and antigerminative properties of the seed oil from the Tunisian endemic *Ferula tunetana* Pomel ex Batt. *Chem. Biodivers.* 14 (1), e1600116. doi:10.1002/cbdv. 201600116
- Znati, M., Jabrane, A., Hajlaoui, H., Harzallah-Skhiri, F., Bouajila, J., Casanova, J., et al. (2012). Chemical composition and *in vitro* evaluation of antimicrobial and anti-acetylcholinesterase properties of the flower oil of *Ferula lutea*. *Nat. Prod. Commun.* 7 (7), 947–950. doi:10.1177/1934578X1200700738
- Zomorodian, K., Saharkhiz, J., Pakshir, K., Immeripour, Z., and Sadatsharifi, A. (2018). The composition, antibiofilm and antimicrobial activities of essential oil of *Ferula assa-foetida* oleo-gum-resin. *Biocatal. Agric. Biotechnol.* 14, 300–304. doi:10.1016/j.bcab.2018.03.014

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2021 Sonigra and Meena. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.