

Review Article

Potential Functional Food Products and Molecular Mechanisms of *Portulaca Oleracea* L. on Anticancer Activity: A Review

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Portulaca oleracea Linn. (*P. oleracea* L.) has recently gained attention as a functional food due to the chemical composition of this plant regarding bioactive compounds. The special attention to the use of *P. oleracea* as an ingredient in functional food products is also due to the promotion of sustainable food. It is an unconventional food plant, and its consumption may contribute to preserving biodiversity due to its cultivation in a polyculture system. Food sovereignty may be achieved, among other strategies, with the consumption of unconventional food plants that are more resistant in nature and easily cultivated in small places. *P. oleracea* grows spontaneously and may be found in streets and sidewalks, or it may be cultivated with seeds and cuttings propagation. The culinary versatility of *P. oleracea* opens up opportunities to explore the development of sustainable, functional food products. This mini-review shows that functional food products developed from *P. oleracea* are already available at the research level, but it is expected that more scientific literature focusing on the development of *P. oleracea* functional products with proven anticancer activities may be released in the near future. Polysaccharides, some phenolic compounds, alkaloids, and cerebrosides are associated with the inhibition and prevention of carcinogenesis through *in vitro* and *in vivo* investigations. The anticancer activities of *P. oleracea*, its bioactive compounds, and the involved molecular mechanisms have been reported in the literature. The importance of further elucidating the cancer inhibition mechanisms is in the interest of forthcoming applications in the development of food products with anticancer properties for implementation in the human diet.

1. Introduction

The common purslane (*P. oleracea* L.) is a herbaceous succulent annual plant from the Portulacaceae family, native to the Middle East and India [1, 2]. It may be found on roadsides, gardens, and cultivated areas in the tropical and subtropical regions [3, 4]. There are various cultivars of *P. oleracea* distributed worldwide, mainly with morphological

differences, with the common purslane having green-red stems, obovate leaves, yellow flowers, and single-layered petals, while the ornamental purslane produces flowers of different colors [1]. The stems and leaves have a slightly acid and salty taste and are usually consumed in salads, soups, and stews [5, 6]. It is an edible plant in regions of European, Mediterranean, African, and Asia countries and Australia [6]. In Brazil, *P. oleracea* is known as an “unconventional

food plant”, a term referring to plants that are not part of the usual consumption of most of the population in a particular region, country, or even the planet because basic food is very homogeneous, with the use of few food species [7].

P. oleracea has a high nutritional value and many antioxidant properties due to its phenolic compound and omega-3 fatty acid abundance, particularly α -linolenic acid. It is well-known in traditional Chinese medicine [2]. for its use in diuretic, febrifuge, antiseptic, antispasmodic, and vermifuge treatments [8]. Among its various pharmacological properties are its anti-inflammatory [9], antioxidative [10], renoprotective [11], neuroprotective [12], hepatoprotective [13], and muscle-relaxing effects [14].

Anticarcinogenic activities have been reported for *P. oleracea*. Investigations were carried out to screen the activities for antihepatocellular carcinoma [15, 16], colon cancer [17], glioblastoma multiforme [18], ovarian cancer [19] sarcoma [20], lung cancer [16], anti-cervical [21], gastric cancer [22], and pancreatic cancer [23]. *P. oleracea* contains bioactive compounds with antioxidant properties, act on metastasis and invasion, modulate the immune system, and inhibit tumor formation [19, 24, 25, 4].

Thus, this mini-review aimed to assemble the anticancer effects of bioactive compounds of *P. oleracea*, demonstrating the molecular mechanisms and the potential for the development of functional food products with anticancer properties.

2. The Nutritional Value and Bioactive Compounds of *P. Oleracea*

Proximate analyses of *P. oleracea* components including leaves, seeds, stems, buds, and flowers, have been performed. Ash, fiber, protein, and fat approximate contents of *P. oleracea* leaves as 20.56%, 36.27%, 12.82%, and 3.75%, respectively, are found on a dry matter basis [26]. *P. oleracea* also contains minerals in its leaves with concentration values approximate such as potassium (3710 mg/100 g of dry matter), calcium (2390 mg/100 g), nitrogen (2170 mg/100 g), magnesium (580 mg/100 g), phosphorus (350 mg/100 g), sulfur (200 mg/100 g), iron (32.4 mg/100 g), manganese (5.8 mg/100 g), boron (2.8 mg/100 g), zinc (2 mg/100 g), and copper (1.1 mg/100 g) [26]. This study showed higher levels of potassium, calcium, magnesium, phosphorus, and iron when compared to those of spinach (336 mg/100 g of dry matter, 98 mg/100 g, 82 mg/100 g, 25 mg/100 g, and 0.4 mg/100 g, respectively) [27].

P. oleracea contains high amounts of Omega-3 fatty acids, as discussed by Siriamornpun and Suttajit [28] that found higher levels of Omega-3 fatty acids in fresh leaves, with 523.146 ± 2.29 mg/100 g, while, for stems and flowers, the authors reported 148.87 ± 3.30 mg/100 g and 216.17 ± 1.16 mg/100 g, respectively. Other plants (analysis of leaves in dry matter) contain lower levels of Omega-3 fatty acids than *P. oleracea*, such as mint (194.9 mg/100 g), watercress (179.6 mg/100 g), spinach (129.2 mg/100 g), parsley (124.8 mg/100 g), and broccoli (110.3 mg/100 g) (analysis of leaves in dry matter) [29]. Omega-3 fatty acids may have pharmacological effects such as anti-hyperlipidemic, antimicrobial, anti-inflammatory, neuroprotective and nephropro-

TECTIVE activities [3, 30, 31, 32, 11]. *P. oleracea* also contains high levels of tocopherols, vitamin A, β -carotene and ascorbic acid [3, 32–34]. Antimicrobial and antioxidant activities were related to these compounds [3, 33].

High concentrations of oxalic acid have also been detected in *P. oleracea*. The intake of oxalic acid provided by the diet with *P. oleracea* may form complexes with minerals such as calcium and iron (insoluble salts) or sodium, magnesium, and potassium (soluble salts), reducing their bioavailability and possibly leading to the development of kidney stones through the formation of calcium oxalate crystals [35]. Thus, consumption of *P. oleracea* should be moderated by individuals with a propensity to develop kidney stones. Amounts of 23.45 ± 0.45 g, 5.58 ± 0.18 g, and 9.09 ± 0.12 g of total oxalates per kilogram of fresh weight oxalates were obtained in fresh leaves, stems, and buds, respectively, with 75.0% being soluble oxalates in the stems and buds, and only 27.5% in the leaves [36]. The authors reported a 66.7% reduction ($p < 0.001$) of soluble oxalates after cooking the leaves for a short time, discarding the water, and pickling them with white vinegar [36]. Some other bioactive compounds from secondary metabolism of *P. oleracea* such as flavonoids, alkaloids, terpenoids and their pharmacological activity can be seen in Table 1.

Flavonoids (a class of phenolic compounds) in *P. oleracea* were associated with anti-fertility, antimicrobial, antioxidant and antidiabetic effects [37–40]. Combined effects of polyunsaturated fatty acids, flavonoids and polysaccharides on hypoglycaemic, hypolipidaemic and insulin resistance reducer effects through ingestion of *P. oleracea* seeds in clinical test with humans were observed [40]. Other phenolic compounds (Polyphenols and phenolic acids) in *P. oleracea* have antioxidant and antimutagenic effects [41–43].

Other bioactive compounds with pharmacological importance in *P. oleracea* are alkaloids and terpenes. Anticancer, anti-inflammatory and antioxidant effects were described for alkaloids found in this plant while hepatoprotective, antibacterial, antifungal and anti-hypoxia effects were described for terpenes of *P. oleracea* [44–48].

3. Potential Antioxidant of the *P. Oleracea*

This plant is rich in antioxidants such as vitamin A, tocopherols, ascorbic acid, beta-carotene, and phenolic compounds [33, 49]. Beta-carotene was found in *P. oleracea* with content ranging from 21 μ g/g to 30 μ g/g of fresh mass in leaves and 3.6 μ g/g to 6.5 μ g/g of fresh mass in stems [50]. The antioxidant potential was measured at different growth stages (15, 30, 45, and 60 days) of aerial parts of *P. oleracea* [49]. The total phenolic content (TPC) for the young shoots at 15 days was significantly lower than at 30, 45, and 60 days, while the ascorbic acid content (AAC) did not show a significant decrease from the developing to the mature stage. According to the study, the IC₅₀ value of 1,1-diphenyl-2-picrylhydrazyl (DPPH) free radical scavenging activity ranged from 1.30 ± 0.04 mg/ml (60 days) to 1.71 ± 0.04 mg/ml (15 days), while the ascorbic acid equivalent antioxidant content (AEAC) values ranged from 229.5 ± 7.9 mg AA/100 g (15 days) to 319.3 ± 8.7 mg AA/100 g (60 days), the TPC varied

TABLE 1: Some classes of bioactive compounds from secondary metabolism of *P. oleracea* and their pharmacological activity.

Compounds	Plant structure	Form (fresh or dry)	Pharmacological activity	References
Flavonoids	Aerial part	Dry	Antifertility	[37]
	Aerial part	Dry	Antimicrobial	[38]
	Leaves	Fresh	Antioxidant	[39]
	Seeds	Dry	Antidiabetic	[40]
Polyphenols	Leaf, stem and flower	Dry	Antioxidant	[41]
	Whole plant	Fresh	Antimutagenic	[42]
Phenolic acids	Aerial parts	Dry	Antioxidant	[43]
	Aerial part	Dry	Anticancer	[44]
Alkaloids	Whole plant	Fresh	Anti-inflammatory	[45]
	Whole plant	Dry	Antioxidant	[46]
Terpenes	Whole plant	Dry	Hepatoprotective, antibacterial and antifungal	[47]
	Aerial part	Dry	Anti-hypoxia	[48]

from 174.5 ± 8.5 mg GAE/100 g (15 days) to 348.5 ± 7.9 mg GAE/100 g (60 days), the AAC varied from 60.5 ± 2.1 mg/100 g (60 days) to 86.5 ± 3.9 mg/100 g (15 days), and the ferric reducing antioxidant power (FRAP) ranged from 1.8 ± 0.1 mg GAE/g (15 days) to 4.3 ± 0.1 mg GAE/g (60 days). Thus, mature plants (60 days) of *P. oleracea* had higher TPC and antioxidant activities than immature plants.

The dry weights of the samples (leaves, flowers, and stems) from two different locations were investigated for potential antioxidant activity by Silva and Carvalho [41], who found that stems had a higher total phenolic content and total antioxidant activity than the flowers and leaves. The oil from seeds, leaves, and stems of *P. oleracea* were analyzed and found that the peroxide value was significantly higher for seed oil and the lowest for stem oil [51]. Furthermore, the highest ascorbic acid content was found for *P. oleracea* seed oil (41.67%), followed by leaf oil (32.29%), and the highest DPPH was obtained for leaf oil (12.55%), followed by seed oil (2.05%). Values for lettuce (IC₅₀ = 17.07 mg/ml), artichoke (IC₅₀ = 18.14 mg/ml), turmeric (IC₅₀ = 21.14 mg/ml), spinach (IC₅₀ = 22.87 mg/ml), and escarole (IC₅₀ = 32.2 mg/ml) were reported by Tiveron et al. [52], showing that *P. oleracea* presents the lowest IC₅₀ necessary to reduce 50% of DPPH free radicals.

4. Functional Food Products and *P. Oleracea*

The *P. oleracea* plant may be used as an ingredient in functional food products due to its nutritional value and bioactive compounds that will be incorporated into the formulations.

The use of the *P. oleracea* plant as food may not only enhance the nutrients and bioactive composition of functional products but also influence their sensory and technological characteristics. Although it is well-known that sensory acceptance by consumers is essential for a product's commercial success on the market, few studies in the literature have reported the application of *P. oleracea* in products and its performance or the sensory profile of such products.

Regarding the technological aspect, the incorporation of the durum wheat flour with 5% of *P. oleracea* to bread resulted in the improvement of the rheological characteristics, an increase in antioxidant properties, and a decrease in the Omega-6-to-Omega-3 ratio, which is beneficial for human health, in addition to improving the sensorial quality [53].

The durum wheat spaghetti fortified with 10% of *P. oleracea*, a potential functional food, was appreciated by consumers. It showed a high concentration of α -linolenic acids (Omega-3), total phenolic compounds, and antioxidant properties, so that, considering 100 g of pasta per day, it is possible to obtain 75 mg of essential linoleic acid and 9 mg of linolenic acid, along with a four-fold increase in total phenolic compounds [54]. The Omega-3 fatty acids can also inhibit carcinogenesis and slow tumor growth, as demonstrated by *in vitro*, *in vivo*, and clinical investigations [55].

The analysis of bread incorporated with four different concentrations of *P. oleracea* powder (0%, 5%, 10%, and 15%) showed increasing water absorption capacity, stability under the mixer, and softening levels as the *P. oleracea* powder concentration in the samples increased. The protein, fat, total ash, moisture, and fiber contents also increased along with the *P. oleracea* concentrations [56]. However, the bread with 15% of *P. oleracea* powder showed a decreased farinograph quality number and presented the lowest scores for sensory properties and color, taste, texture, and overall liking. The optimized formulation containing 10% of *P. oleracea* powder had the highest acceptance.

P. oleracea has also been used to produce powder mixtures with two other plant species, *Amaranthus hybridus* L. and *Chenopodium berlandieri* L. The powder mixtures containing *P. oleracea* showed more significant contents of phenolic compounds, with an increase in the antioxidant activity [57].

Another innovative functional product assessed was a fermented *P. oleracea* juice added with a selected lactic acid bacteria. Results demonstrated an increase in total antioxidants, preserved vitamin C, A, and E levels, and increased contents of vitamin B2 and phenolic compounds. In addition, decreased levels of pro-inflammatory mediators and

TABLE 2: Bioactive compounds of *P. oleracea*, types of extracts, and molecular mechanisms for cancer inhibition.

<i>In vitro</i>	Experimental model	<i>In vivo</i>	Compounds	Types of extract	Types of cancer inhibited	Mechanisms and results	References
		Rats	Polysaccharides	Aqueous extract	Ovarian	Scavenge superoxide anion, (DPPH-), nitric oxide, and hydroxyl radicals Inhibit RBC hemolysis	[19]
	Human cancer cell lines SF-268, NCI-H460, K-562, SGC-7901, and SMMC-7721		Homoisoflavonoids	Hydroalcoholic extract		Spleen, thymocyte, T and B lymphocyte proliferation	[61]
		Mice	Polysaccharides	Aqueous extract	Cervical	Sub-G1 phase cell cycle arrest, triggering DNA damage Inducing apoptosis	[21]
	Treatment of HeLa cell	Mice	Polysaccharides	Aqueous extract		Inhibit the growth of transplantable sarcoma 180 Increase in the number of white blood cells (WBC) and CD4+ T-lymphocytes	[20]
		Rats	Polysaccharides	Aqueous extract	Gastric	Increase in the CD4 ⁺ /CD8 ⁺ ratio Interleukin-2 (IL-2), interleukin-4 (IL-4), and tumor necrosis factor-alpha (TNF- α) was enhanced Provide dose-dependent protection against MNGG-induced oxidative injury by enhancing SOD, CAT, GSH-Px	[22]
	Human lung (K562 and A549) and breast (MCF-7 and MDA-MB-435) cancer cell lines		Alkaloids	Hydroalcoholic extract	Lung Breast	Moderate cytotoxic activities against A549 and weak cytotoxic activities against K562. The compounds showed low cytotoxic activity against MCF-7 and MDA-MB-435 cells.	[54]
	Human hepatocellular carcinoma cells			Seed alcoholic extract	Hepatocellular	Significantly reduced the cell viability of HepG2.	[15]
	The uterine cervical carcinoma (U14) cell line		Polysaccharides	Aqueous extract	Cervical	Upregulated the expression of CD80, CD86, CD83 Increase in IL-12, TLR-4, Decrease in IL-10	[29]
	Human HL60 cell line		Portulacerebroside A	Aqueous extract	Leukemia	Mitochondrial membrane potential ROS accumulated Increase in RNA expressions and protein levels of Bax/Bcl-2, caspase-3, and caspase-9	[49]
	HepG2 and A-549 cell lines			Seed oil	Liver Lung	ERK1/2, JNK1/2 and p38 MAPK pathway were blocked Significant cytotoxicity and inhibition of growth of the liver cancer (HepG2) and lung cancer (A-549) cell lines	[20]
	Human liver cancer HCCLM3 cells		Portulacerebroside A	Aqueous extract	Liver	Increase in RNA and protein expression levels of TIMP-2 and nm23-H1 Inhibition of the mRNA expression of MTA1, MMP-2, and MMP-9	[23]

TABLE 2: Continued.

<i>In vitro</i>	Experimental model	<i>In vivo</i>	Compounds	Types of extract	Types of cancer inhibited	Mechanisms and results	References
	Cervical cancer HeLa cells, esophageal cancer Eca-109 cells and breast cancer MCF-7 cells			Seed oil	Cervical Esophageal Breast	Suppression of the protein expression of MTA1, RhoA, Rac1/Cdc42, MMP-2, but not RhoC and MMP-9 Stronger inhibitory effect on the proliferation of MCF-7 cells and significantly inhibited the proliferation of HeLa cells and Eca-109 cells	[51]
	PANC-1 cancer cell line			Aqueous extract	Pancreatic	Significant effect on apoptosis in pancreatic cell line and high expression of P53 and reduction of CDK gene expression	[23]
	Human colon adenocarcinoma (HCT-15) and normal (Vero) cell line			Chloroform extract	Colon adenocarcinoma	Chloroform extract does not have cytotoxic activity and was not safe to normal Vero cell line.	[67]
	Colon cancer cells (HT-29) and HT-29 cancer stem cells			Ethyl alcohol extract	Colon Stem cells	Inhibited the proliferation of both HT-29 cancer cells and HT-29 cancer stem cells Significantly decreased the expression of the Notch1 and β -catenin genes in both cell types Decrease HeLa cell proliferation	[52]
	The human cervical cancer HeLa cells.			Aqueous extract	Cervical	Upregulate Bax level and downregulate Bcl-2 level in a concentration-dependent manner Inhibit the protein expression levels of TLR4, MyD88, TRAF6, AP-1 and NF- κ B subunit P65 Reduce the production of cytokine/chemokine	[50]
	The mouse cervical carcinoma U14 cells			Aqueous extract	Intestinal	Dendritic cell (DC) apoptosis in U14-bearing mice Increase intestinal DC survival Stimulate the TLR4-PI3K/AKT-NF- κ B signaling pathway	[48]
	Human glioblastoma cancer cell line (U-87)			Hydroethanolic extract		Cytotoxicity and apoptogenic effects Anti-NF- κ B activity along with two upstream ROS and NO mechanisms	[17]

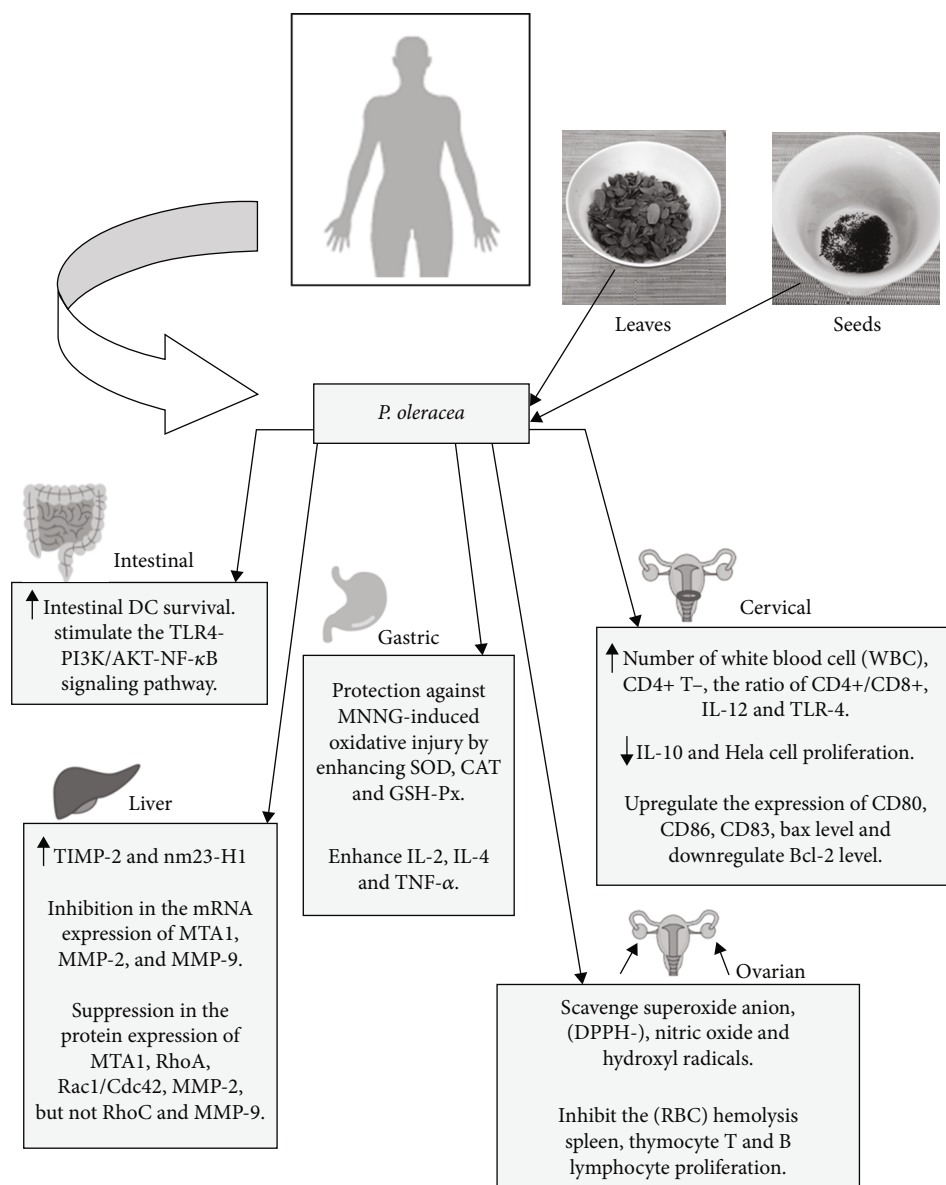


FIGURE 1: Some possible mechanisms of *P. oleracea* for anticancer activity.

reactive oxygen species were observed, with a consequent increase in the restorative characteristics of the use of *P. oleracea* juice for intestinal inflammation and epithelial injury [58].

The combination of yogurt or coconut plant extract or coconut cream with fresh leaves of *P. oleracea* reduced the overall oxalate content by simple dilution. The soluble oxalate content decreased from 53.0% to 10.7% when *P. oleracea* leaves were added to yogurt. However, the coconut plant extract and coconut cream had no effect on the percentage of soluble oxalate content but provided the mixture with an acceptable flavor [59].

The addition of fresh purslane leaves (ranging from 1% to 10%, w/w) to tomato sauces resulted in a decrease of total soluble solids from 9.57°Bx to 9.20°Bx, beneficially impacting sugar reduction. On the other hand, the amount of protein significantly increased from 0.12% to 1.83% from the lowest to the highest concentrations, respectively [60].

5. Bioactive Compounds of *P. Oleracea* on Anticancer Activity

P. oleracea presents phytochemicals and nutrients associated with anticarcinogenic properties. The 12% reduction in the activity of the mutagenic nitrosation mixture may be attributed to the ascorbic acid (vitamin C), α and β -carotene, chlorophyll, and polyphenols of the *P. oleracea* extract obtained through a standard juice extractor [42].

Phenolic compounds such as kaempferol and apigenin from a hydroethanolic extract of *P. oleracea* have effects *in vitro* against human glioma cells, and homoisoflavonoids showed *in vitro* selective cytotoxic activity for SF-268, NCI-H460, and SGC-7901 cell lines, as shown Table 2 [18, 61].

Polysaccharides from *P. oleracea* act on free radicals through the antioxidant mechanism, modulating the immune system, which may be preventive and therapeutic

in rat ovarian and gastric cancer and mouse cervical cancer and sarcomas, as shown Table 2 [19–22, 62].

Another bioactivity from *P. oleracea* is portulacacerebroside A, a cerebroside compound that suppresses the invasion and metastasis of liver cancer HCCLM3 cells and acts in leucocythemia treatment are shown in Table 2 [24, 63].

Polysaccharides showed activity against ovarian cancer by inhibiting the red blood cell (RBC) hemolysis in the spleen, thymocyte, and T and B lymphocyte proliferation [19]. These compounds also act against cervical cancer through Sub-G1 phase cell cycle arrest triggering DNA damage, inhibit the growth of transplantable sarcoma 180, increase the number of white blood cells (WBC), CD4+ T-, the CD4+/CD8+ ratio, IL-12, and TLR-4, decrease IL-10 and HeLa cell proliferation, reduce the production of cytokine/chemokine and the expression levels of CD80, CD86, CD83, Bax, and downregulate the Bcl-2 level in a concentration-dependent manner. In addition, polysaccharides inhibit the protein expression levels of TLR4, myeloid differentiation primary response 88 (MyD88), TNF receptor associated Factor 6 (TRAF6), activator protein-1 (AP-1), and factor nuclear kappa B (NF- κ B) subunit P65 [20, 21, 63, 64].

In gastric cancer, interleukins (IL-2 and IL-4) and TNF- α were enhanced by polysaccharides that also provide dose-dependent protection against N-methyl-N'-nitro-N-nitrosoguanidine (MNNG) induced oxidative injury by enhancing Superoxide dismutase (SOD), catalase (CAT), and glutathione (GSH-Px) [22]. In addition to acting against ovarian, gastric, and cervical cancer, polysaccharides also work against intestinal cancer by stimulating the TLR4-PI3K/AKT-NF- κ B signaling pathway and Anti-NF- κ B activity along with two upstream ROS and NO mechanisms [18, 62], showing the importance of studying these molecules in *P. oleracea* matrices.

The cerebroside compound, Portulacacerebroside A, affects leukemia and cervical, liver, esophageal, breast, and colon cancer and cancer stem cells [16, 24, 63, 65, 66]. Some mechanisms involved with Portulacacerebroside A have increased RNA expressions and protein levels of Bax/Bcl-2, caspase-3, and caspase-9, protein expression levels of TIMP-2 and nm23-H1, inhibition of the mRNA expression of MTA1, MMP-2, and MMP-9, RhoA, Rac1/Cdc42, MMP-2, and downregulation of the expression of the Notch1 and β -catenin genes.

Alkaloids inhibited lung and breast cancer through moderate cytotoxic activities against A549, weak cytotoxic activities against K562, and low cytotoxic activity against MCF-7 and MDA-MB-435 cells.

Some possible mechanisms of *P. oleracea* for anticancer activity are represented in Figure 1. The bioactivity of *P. oleracea* and the potential to develop new products from this underused plant in some regions deserve attention regarding its valorization as a functional food and its pharmacological properties. Different anticancer mechanisms of *P. oleracea* were explored and reported in this review. Aqueous extracts, seed oil, and hydroethanolic extracts present cytotoxicity to cancer cell lines while chloroform extract does not have cytotoxic activity [67]. Further studies will be needed to determine anticancer activity in particular food matrices and beverages.

6. Conclusion

The *P. oleracea* plant may be promising for developing and innovating potential functional food products. The high levels of antioxidants such as phenolic compounds, carotenoids, and other nutrients such as minerals and Omega-3 fatty acids are supported by functional food studies. Research has indicated the anticancer activity of *P. oleracea* extracts. Polysaccharides, some phenolic compounds, alkaloids, and cerebroside detected in *P. oleracea* and contained in aqueous extracts, seed oil, and hydroethanolic extracts are associated with inhibition and prevention of carcinogenesis. However, more studies are needed to prove the anticancer activity of food products containing *P. oleracea* as an ingredient to promote health benefits to the consumers.

Data Availability

The data used to support the findings of this study are included within the article.

Conflicts of Interest

The authors declare that there is no conflict of interest.

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References

- [1] M. Alam, A. S. Juraimi, M. Y. Rafii et al., "Genetic improvement of purslane (*Portulaca oleracea* L.) and its future prospects," *Molecular Biology Reports*, vol. 41, no. 11, pp. 7395–7411, 2014.
- [2] M. Iranshahy, M. Javadi, M. Iranshahi et al., "A review of traditional uses, phytochemistry and pharmacology of *Portulaca oleracea* L.," *Journal of Ethnopharmacology*, vol. 205, pp. 158–172, 2017.
- [3] M. K. Uddin, A. S. Juraimi, M. S. Hossain, M. A. U. Nahar, M. E. Ali, and M. M. Rahman, "Purslane weed (*Portulaca oleracea*): A prospective plant source of nutrition, omega-3 fatty acid, and antioxidant attributes," *The Scientific World Journal*, vol. 2014, Article ID 951019, 6 pages, 2014.
- [4] V. B. Rahimi, F. Ajam, H. Rakhshandeh, and V. R. Askari, "A pharmacological review on *Portulaca oleracea* L.: focusing on anti-inflammatory, anti-oxidant, immuno-modulatory and antitumor activities," *Korean Pharmacopuncture Institute*, vol. 22, no. 1, pp. 7–15, 2019.
- [5] Y. Y. Lim and E. P. L. Quah, "Antioxidant properties of different cultivars of *Portulaca oleracea*," *Food Chemistry*, vol. 103, no. 3, pp. 734–740, 2007.
- [6] B. Nemzer, F. Al-TaHER, and N. Abshiru, "Phytochemical composition and nutritional value of different plant parts in two cultivated and wild purslane (*Portulaca oleracea* L.) genotypes," *Food Chemistry*, vol. 320, article 126621, 2020.

- [7] V. F. Kinnup and H. Lorenzi, *Plantas Alimentícias Não-Convencionais (PANC) no Brasil*, Instituto Plantarum de Estudos da Flora, São Paulo, 2014.
- [8] L. Xiang, X. Dongming, W. Wang, R. Wang, Y. Ding, and L. Du, "Alkaloids from *Portulaca oleracea* L.," *Phytochemistry*, vol. 66, no. 21, pp. 2595–2601, 2005.
- [9] K. Chan, M. W. Islam, M. Kamil et al., "The analgesic and anti-inflammatory effects of *Portulaca oleracea* L. subsp. *sativa* (Haw.) Celak," *Journal of Ethnopharmacology*, vol. 73, no. 3, pp. 445–451, 2000.
- [10] M. A. Dkhil, A. E. A. Moniem, S. Al-Quraishy, and R. A. Saleh, "Antioxidant effect of purslane (*Portulaca oleracea*) and its mechanism of action," *Journal of Medicinal Plants Research*, vol. 5, no. 9, pp. 1589–1593, 2011.
- [11] W. Hozayen, M. Bastawy, and H. Elshafeey, "Effects of aqueous purslane (*Portulaca Oleracea*) extract and fish oil on gentamicin nephrotoxicity in albino rats," *Nature and Science*, vol. 9, no. 2, pp. 47–62, 2011.
- [12] C. Q. Wang and G. Q. Yang, "Betacyanins from *Portulaca oleracea* L. ameliorate cognition deficits and attenuate oxidative damage induced by D-galactose in the brains of senescent mice," *Phytomedicine*, vol. 17, no. 7, pp. 527–532, 2010.
- [13] A. Eidi, P. Mortazavi, J. Z. Moghadam, and M. Mardani, "Hepatoprotective effects of *Portulaca oleracea* extract against CCl₄-induced damage in rats," *Pharmaceutical Biology*, vol. 53, no. 7, pp. 1042–1051, 2015.
- [14] M. Gonnella, M. Charfeddine, G. Conversa, and P. Santamaria, "Purslane: a review of its potential for health and agricultural aspects," *European Journal of Plant Science and Biotechnology*, vol. 4, no. 1, pp. 131–136, 2010.
- [15] N. N. Farshori, E. S. S. Al-Sheddi, M. M. Al-Oqail, J. Musarrat, A. A. Al-Khedhairi, and M. A. Siddiqui, "Cytotoxicity assessments of *Portulaca oleracea* and *Petroselinum sativum* seed extracts on human hepatocellular carcinoma cells (HepG2)," *Asian Pacific Journal of Cancer Prevention*, vol. 15, no. 16, pp. 6633–6638, 2014.
- [16] E. S. Al-Sheddi, N. N. Farshori, M. M. Al-Oqail, J. Musarrat, A. A. Al-Khedhairi, and M. A. Siddiqui, "*Portulaca oleracea* seed oil exerts cytotoxic effects on human liver cancer (HepG2) and human lung cancer (A-549) cell lines," *Asian Pacific Journal of Cancer Prevention*, vol. 16, no. 8, pp. 3383–3387, 2015.
- [17] G. P. Asnani and C. R. Kokare, "In vitro and in vivo evaluation of colon cancer targeted epichlorohydrin crosslinked *Portulaca*-alginate beads," *Biomolecular Concepts*, vol. 9, no. 1, pp. 190–199, 2018.
- [18] V. B. Rahimi, S. H. Mousavi, S. Haghghi, S. Soheili-Far, and V. R. Askari, "Cytotoxicity and apoptogenic properties of the standardized extract of *Portulaca oleracea* on glioblastoma multiforme cancer cell line (U-87): a mechanistic study," *EXCLI Journal*, vol. 18, pp. 165–186, 2019.
- [19] C. Youguo, S. Zongji, and C. XiaoPing, "Evaluation of free radicals scavenging and immunity-modulatory activities of Purslane polysaccharides," *International Journal of Biological Macromolecules*, vol. 45, no. 5, pp. 448–452, 2009.
- [20] H. Shen, G. Tangg, G. Zeng et al., "Purification and characterization of an antitumor polysaccharide from *Portulaca oleracea* L.," *Carbohydrate Polymers*, vol. 93, no. 2, pp. 395–400, 2013.
- [21] R. Zhao, X. Gao, Y. Cai et al., "Antitumor activity of *Portulaca oleracea* L. polysaccharides against cervical carcinoma in vitro and in vivo," *Carbohydrate Polymers*, vol. 96, no. 2, pp. 376–383, 2013.
- [22] Y. Li, Y. Hu, S. Shi, and L. Jiang, "Evaluation of antioxidant and immuno-enhancing activities of Purslane polysaccharides in gastric cancer rats," *International Journal of Biological Macromolecules*, vol. 68, pp. 113–116, 2014.
- [23] S. Alipour, L. Pishkar, and V. Chaleshi, "Cytotoxic effect of *Portulaca Oleracea* extract on the regulation of CDK1 and P53 gene expression in pancreatic cancer cell line," *Nutrition and Cancer*, vol. 74, no. 5, pp. 1792–1801, 2022.
- [24] Q. Ji, G. Y. Zheng, W. Xia et al., "Inhibition of invasion and metastasis of human liver cancer HCCLM3 cells by portulacerebroside a," *Pharmaceutical Biology*, vol. 53, no. 5, pp. 773–780, 2015.
- [25] V. R. Askari, S. A. R. Rezaee, K. Abnous, M. Iranshahi, and M. H. Boskabady, "The influence of hydro-ethanolic extract of *Portulaca oleracea* L. on Th₁/Th₂ balance in isolated human lymphocytes," *Journal of Ethnopharmacology*, vol. 194, pp. 1112–1121, 2016.
- [26] D. C. S. Oliveira, C. Wobeto, M. R. Zanuzo, and C. Severgnini, "Composição mineral e teor de ácido ascórbico nas folhas de quatro espécies olerícolas não-convencionais," *Horticultura Brasileira*, vol. 31, no. 3, pp. 472–475, 2013.
- [27] NEPA/UNICAMP, *Tabela Brasileira de Composição de Alimentos*, NEPA/UNICAMP, Campinas, 2011.
- [28] S. Siriamornpun and M. Suttajit, "Microchemical components and antioxidant activity of different morphological parts of Thai wild purslane (*Portulaca oleracea*)," *Weed Science*, vol. 58, no. 3, pp. 182–188, 2010.
- [29] C. Pereira, D. Li, and A. J. Sinclair, "The alpha-linolenic acid content of green vegetables commonly available in Australia," *International Journal for Vitamin and Nutrition Research*, vol. 71, no. 4, pp. 223–228, 2001.
- [30] A. P. Simpopoulos, "Omega-3 fatty acids and antioxidants in edible wild plants," *Biological Research*, vol. 37, no. 2, pp. 263–277, 2004.
- [31] F. Naem and S. H. Khan, "Purslane (*Portulaca oleracea* L.) as phytochemical substance—a review," *Journal of Herbs, Spices and Medicinal Plants*, vol. 19, no. 3, pp. 216–232, 2013.
- [32] R. Zhao, T. Zhang, H. Zhao, and Y. Cai, "Effects of *Portulaca oleracea* L. Polysaccharides on phenotypic and functional maturation of murine bone marrow derived dendritic cells," *Nutrition and Cancer*, vol. 67, no. 6, pp. 987–993, 2015.
- [33] S. Petropoulos, A. Karkanis, N. Martins, and I. C. F. R. Ferreira, "Phytochemical composition and bioactive compounds of common purslane (*Portulaca oleracea* L.) as affected by crop management practices," *Trends in Food Science & Technology*, vol. 55, pp. 1–10, 2016.
- [34] M. M. S. Viana, L. A. Carlos, E. C. Silva, S. M. F. Pereira, D. B. Oliveira, and M. L. V. Assis, "Composição fitoquímica e potencial antioxidante de hortaliças não convencionais," *Horticultura Brasileira*, vol. 33, no. 4, pp. 504–509, 2015.
- [35] U. R. Palaniswamy, B. Bible, and R. J. McAvoy, "OXALIC acid concentrations in purslane (*Portulaca oleracea* L.) is altered by the stage of harvest and the nitrate to ammonium ratios in hydroponics," *Acta Horticulturae*, vol. 629, no. 629, pp. 299–305, 2004.
- [36] G. Y. Poeydomenge and G. P. Savage, "Oxalate content of raw and cooked purslane," *Journal of Food, Agriculture and Environment*, vol. 5, no. 1, pp. 124–128, 2007.
- [37] H. B. Nayaka, R. L. Londonkar, M. K. Umesh, and A. Tukappa, "Antibacterial attributes of apigenin, isolated from *Portulaca oleracea* L.," *International Journal of Bacteriology*, vol. 2014, Article ID 175851, 8 pages, 2014.

- [38] Y.-K. Du, L. Jing, X.-M. Li et al., "Flavonoids extract from *Portulaca oleracea* L. induce Staphylococcus aureus death by apoptosislike pathway," *International Journal of Food Properties*, vol. 20, no. 1, pp. 534–542, 2017.
- [39] V. Scari, M. R. Loizzo, R. Tundis, A. Mincione, and T. M. Pellicano, "*Portulaca oleracea* L. (purslane) extracts display antioxidant and hypoglycaemic effects," *Journal of Applied Botany and Food Quality*, vol. 91, pp. 39–46, 2018.
- [40] M.-I. K. El-Sayed, "Effects of *Portulaca oleracea* L. seeds in treatment of type-2 diabetes mellitus patients as adjunctive and alternative therapy," *Journal of Ethnopharmacology*, vol. 137, no. 1, pp. 643–651, 2011.
- [41] R. Silva and I. S. Carvalho, "In vitro antioxidant activity, phenolic compounds and protective effect against DNA damage provided by leaves, stems and flowers of *Portulaca oleracea* (purslane)," *Natural Product Communications*, vol. 9, no. 1, pp. 45–50, 2014.
- [42] S. Caballero-Salazar, R. Riverón-Negrete, M. G. Ordáz-Tellez, F. Abdullaev, and J. J. Espinosa-Aguirre, "Evaluation of the antimutagenic activity of different vegetable extracts using an *In Vitro* screening test," *Proceedings of the Western Pharmacology Society*, vol. 45, pp. 101–103, 2022.
- [43] N. Erkan, "Antioxidant activity and phenolic compounds of fractions from *Portulaca oleracea* L.," *Food Chemistry*, vol. 133, no. 3, pp. 775–781, 2012.
- [44] J. L. Tian, X. Liang, P. Y. Gao et al., "Two new alkaloids from *Portulaca oleracea* and their cytotoxic activities," *Journal of Asian Natural Products Research*, vol. 16, no. 3, pp. 259–264, 2014.
- [45] Y. Meng, Z. Ying, Z. Xiang et al., "The anti-inflammation and pharmacokinetics of a novel alkaloid from *Portulaca oleracea* L.," *Journal of Pharmacy and Pharmacology*, vol. 68, no. 3, pp. 397–405, 2016.
- [46] Z. Yang, C. Liu, L. Xiang, and Y. Zheng, "*Phenolic alkaloids as a new class of antioxidants in Portulaca oleracea*," *Phytotherapy Research*, vol. 23, no. 7, pp. 1032–1035, 2009.
- [47] E. S. Elkhayat, S. R. M. Ibrahim, and M. A. Aziz, "Portulene, a new diterpene from *Portulaca oleracea* L.," *Journal of Asian Natural Products Research*, vol. 10, no. 11, pp. 1039–1043, 2008.
- [48] C.-J. Chen, W.-Y. Wang, X.-L. Wang et al., "Anti-hypoxic activity of the ethanol extract from *Portulaca oleracea* in mice," *Journal of Ethnopharmacology*, vol. 124, pp. 246–250, 2009.
- [49] M. K. Uddin, A. S. Juraimi, M. E. Ali, and M. R. Ismail, "Evaluation of antioxidant properties and mineral composition of purslane (*Portulaca oleracea* L.) at different growth stages," *International Journal of Molecular Sciences*, vol. 13, no. 8, pp. 10257–10267, 2012.
- [50] L. Liu, P. Howe, Y. F. Zhou, Z. Q. Xu, C. Hocart, and R. Zhang, "Fatty acids and β -carotene in Australian purslane (*Portulaca oleracea*) varieties," *Journal of Chromatography A*, vol. 893, no. 1, pp. 207–213, 2000.
- [51] M. Desta, A. Molla, and Z. Yusuf, "Characterization of physico-chemical properties and antioxidant activity of oil from seed, leaf and stem of purslane (*Portulaca oleracea* L.)," *Biotechnology Reports*, vol. 27, pp. 1–5, 2020.
- [52] A. P. Tiveron, P. S. Melo, K. B. Bergamaschi, T. M. F. S. Vieira, M. A. B. Regitano-d'Arce, and S. M. Alencar, "Antioxidant activity of Brazilian vegetables and its relation with phenolic composition," *International Journal of Molecular Sciences*, vol. 13, no. 7, pp. 8943–8957, 2012.
- [53] M. G. Melilli, V. D. Stefano, F. Sciacca et al., "Improvement of fatty acid profile in durum wheat breads supplemented with *Portulaca oleracea* L. quality traits of purslane-fortified bread," *Food*, vol. 9, no. 6, 2020.
- [54] M. G. Melilli, A. Pagliaro, S. Scandurra, C. Gentile, and V. di Stefano, "Omega-3 rich foods: durum wheat spaghetti fortified with *Portulaca oleracea*," *Food Bioscience*, vol. 37, article 100730, 2020.
- [55] J. Y. Lee, T. B. Sim, J. E. Lee, and H. K. Na, "Chemopreventive and chemotherapeutic effects of fish oil derived omega-3 polyunsaturated fatty acids on colon carcinogenesis," *Clinical Nutrition Research*, vol. 6, no. 3, pp. 147–160, 2017.
- [56] L. N. Delvarianzadeh, M. Nafchi, and H. Ebrahimi, "Physicochemical, rheological, and sensory evaluation of voluminous breads enriched by purslane (*Portulaca oleracea* L.)," *Italian Journal of Food Science*, vol. 32, pp. 815–830, 2020.
- [57] Y. O. Santiago-Saenz, C. U. López-Palestina, J. Gutiérrez-Tlahque, R. Monroy-Torres, J. M. Pinedo-Espinoza, and A. D. Hernández-Fuentes, "Nutritional and functional evaluation of three powder mixtures based on mexican quelites: alternative ingredients to formulate food supplements," *Food Science and Technology*, vol. 40, no. 4, pp. 1029–1037, 2020.
- [58] R. Di Cagno, P. Filannino, O. Vincentini, V. Cantatore, I. Cavoski, and M. Gobbetti, "Fermented *portulaca oleracea* L. juice: a novel functional beverage with potential ameliorating effects on the intestinal inflammation and epithelial injury," *Nutrients*, vol. 11, no. 2, p. 248, 2019.
- [59] A. G. Moreau and G. P. Savage, "Oxalate content of purslane leaves and the effect of combining them with yoghurt or coconut products," *Journal of Food Composition and Analysis*, vol. 22, no. 4, pp. 303–306, 2009.
- [60] L. C. Apostol, S. Ropciuc, A. E. Prisacaru, and E. Albu, "Characterization of tomato sauce enriched with purslane (*Portulaca oleracea*) leaves," *Journal of Hygienic Engineering and Design*, vol. 31, pp. 127–132, 2020.
- [61] J. Yan, L. R. Sun, Z. Y. Zhou et al., "Homoisoflavonoids from the medicinal plant *Portulaca oleracea*," *Phytochemistry*, vol. 80, pp. 37–41, 2012.
- [62] R. Zhao, X. Shao, G. Jia et al., "Anti-cervical carcinoma effect of *Portulaca oleracea* L. polysaccharides by oral administration on intestinal dendritic cells," *BMC Complementary and Alternative Medicine*, vol. 19, no. 1, pp. 1–10, 2019.
- [63] Q. Ye, N. Zhang, K. Chen, J. Zhu, and H. Jiang, "Effects of portulacerebroside a on apoptosis of human leukemia HL60 cells and p38/JNK signaling pathway," *International Journal of Clinical and Experimental Pathology*, vol. 8, no. 11, pp. 13968–13977, 2015.
- [64] R. Zhao, T. Zhang, B. Ma, and X. Li, "Antitumor activity of *Portulaca Oleracea* L. polysaccharide on HeLa cells through inducing TLR4/NF- κ B signaling," *Nutrition and Cancer*, vol. 69, no. 1, pp. 131–139, 2017.
- [65] G. Guo, L. Yue, S. Fan, S. Jing, and L. H. Yan, "Antioxidant and antiproliferative activities of purslane seed oil," *Journal of Hypertension*, vol. 5, no. 2, pp. 01–19, 2016.
- [66] H. Jin, L. Chen, S. Wang, and D. Chao, "*Portulaca oleracea* extract can inhibit nodule formation of colon cancer stem cells by regulating gene expression of the notch signal transduction pathway," *Tumor Biology*, vol. 39, no. 7, pp. 101042831770869–101042831770869, 2017.
- [67] P. Y. Mali, "Assessment of cytotoxicity of *Portulaca oleracea* Linn. Against human colon adenocarcinoma and vero cell line," *Ayu*, vol. 36, no. 4, pp. 432–436, 2015.