


STANDARD ARTICLE

Strangles, convalescent *Streptococcus equi* subspecies *equi* M antibody titers, and presence of complications

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Background: *Streptococcus equi* subspecies *equi* infection elicits M protein antibody titers in equids. Interpretation of titers is not generally accepted.

Hypothesis: The magnitude of *S. equi* M protein (SeM) antibody titer after infection (titer $\geq 1:12\ 800$) will be useful to monitor for the presence of complications or the risk of development of complications.

Animals: Forty-eight horses on 1 farm involved in strangles outbreak.

Methods: Clinical and observational study. *S. equi* M protein antibody titers were measured on all horses 8 weeks after infection and select horses 12 and 28 weeks after infection. Horses were categorized: no disease, uncomplicated case, persistent guttural pouch (GP) infection, or complicated cases (metastatic abscesses, purpura hemorrhagica, secondary infections, and dysphagia). Category was compared to titer.

Results: Twenty-eight of 48 (58%) developed clinical signs of *S. equi* infection. Of those, 11 (39%) had uncomplicated strangles, 9 (21%) had persistent GP infection, 5 (18%) were complicated cases, and 3 (11%) had both persistent GP infection and complications. Thirty-three percent of horses (16 of 48) had SeM antibody titers $\geq 1:12\ 800$ eight weeks after infection. Of horses with titers $\geq 1:12\ 800$, 6 of 16 had evidence of complications. Of complicated cases, 6 of 8 had titers $\geq 1:12\ 800$. In this outbreak, the sensitivity (75%; 95% CI [confidence interval] 45-105) for a SeM antibody titer $\geq 1:12\ 800$ detecting complications was higher than the specificity (43%; 95% CI 23-64).

Conclusions and Clinical Importance: This outbreak demonstrates that SeM antibody titers can be increased after infection ($\geq 1:12\ 800$) in the absence of complications of strangles.

KEYWORDS

SeM antibody titer, strangles carrier, strangles complications, *Streptococcus equi* subspecies *equi*

Abbreviations: CI, confidence interval; *S. equi*, *Streptococcus equi* subspecies *equi*; SeM antibody titer, *Streptococcus equi* subspecies *equi* M protein antibody titer; GP, guttural pouch; PCR, polymerase chain reaction; QH, Quarter horse; TWH, Tennessee Walking horse.

1 | INTRODUCTION

Streptococcus equi subspecies *equi* is the causative agent of strangles, a highly contagious upper respiratory tract infection of horses. Typical clinical signs of disease include fever, inappetance, lethargy, submandibular or retropharyngeal lymphadenopathy or purulent drainage, or purulent nasal discharge. Complications of *S. equi* infection can occur

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and include airway obstruction from lymphadenopathy, disseminated abscesses from hematogenous spread, or purpura hemorrhagica and various diseases caused by immune-mediated processes.¹⁻⁵ *Streptococcus equi* M protein (SeM) antibody titers are typically measured to determine if a horse has developed a complication of strangles, such as purpura hemorrhagica or metastatic abscess formation, or to determine if a horse is at risk of purpura hemorrhagica if they were to be vaccinated. Both the 2005 and 2018 American College of Veterinary Internal Medicine consensus statements on strangles state that a very high titer ($\geq 1:12\ 800$) is associated with metastatic abscess formation or purpura hemorrhagica and that high titers (1:3200-1:6400) are detected 4-12 weeks after infection.^{1,2} Anecdotally, horses can have high titers ($\geq 1:12\ 800$) 4-8 weeks after infection and no signs of complications (authors' personal observations, KMD, LAB, ACT). The objective of this study was to measure SeM antibody titers on horses after outbreak to determine if titers detect the presence of complications.² An additional objective was to follow SeM antibody titers out to 7 months after infection to determine immunoglobulin decay and to monitor for development of additional complications. We hypothesized that the magnitude of SeM antibody titer after infection (SeM titer $\geq 1:12\ 800$) will be useful to monitor for the presence of complications or for the risk of development of complications.

2 | MATERIALS AND METHODS

This was a clinical observational study of convalescent SeM antibody titers in a strangles outbreak with a high rate of complications. All samples were obtained with informed client consent. Approximately 8 weeks after initial diagnosis of infection with *S. equi*, serum was collected via jugular venipuncture on all 48 horses on the farm. *S. equi* M protein antibody titers were measured via ELISA for each horse at Equine Diagnostic Solutions LLC in Lexington, KY. At 12 weeks after initial diagnosis, serum was collected for repeat SeM antibody titers on select horses ($n = 18$). At 28 weeks after initial diagnosis, serum was once again collected for measurement of SeM antibody titers ($n = 36$). Physical examinations were performed at all time points to determine if horses were displaying any signs of disease.

Data on each horse on the property were collected from the initial diagnosis through follow-up to removal from quarantine. Data collected included signalment, clinical signs displayed (or absence of clinical signs), nasopharyngeal lavage or guttural pouch (GP) endoscopy and lavage results for *S. equi* culture and polymerase chain reaction (PCR), evidence of complications, vaccination status, and survival. Affected horses were categorized according to their clinical signs of disease into 4 categories: no disease, uncomplicated case, persistent GP infection, or complicated case. No disease was defined as no clinical evidence of *S. equi* infection. An uncomplicated case was defined as clinical signs of 1 or more of fever, inappetance, purulent nasal discharge, and submandibular or retropharyngeal lymphadenopathy or drainage. A persistent GP infection was defined as GP infection (positive nasopharyngeal or GP lavage *S. equi* culture or PCR) lasting >40 days.⁴ A complicated case of strangles was defined as any sequelae or atypical case including signs of immune-mediated purpura hemorrhagica, metastatic abscess formation (abscesses remote from

lymph nodes of the head), secondary infections, or dysphagia. Horses with evidence of persistent GP infection and complications were categorized dually, but no horse with uncomplicated strangles had persistent GP infection.

The frequency of titers $\geq 1:12\ 800$ in different disease categories was determined. Median SeM antibody titer for each category as well as for vaccination status was determined. A *t* test was used to analyze the difference in titer level for vaccination status. Correlations between disease category and SeM antibody titer level were analyzed by a Pearson's test. Sensitivities and specificities with 95% CI (confidence interval) for SeM antibody titers $\geq 1:12\ 800$ or $\geq 1:6400$ detecting complications or persistent GP infection were calculated. The mean reciprocal antibody titer for each time point was calculated, and regression was calculated to determine antibody decay over time. Values of $P < .05$ were considered statistically significant.

3 | RESULTS

3.1 | Clinical disease

The *S. equi* outbreak in a boarding facility began in late January 2017 and lasted approximately 1 month. Before *S. equi* was detected on the farm, 1 horse developed signs of colic, lethargy, and fever. This horse was isolated at a veterinary hospital. Fecal PCR testing was performed on this horse and was positive for equine coronavirus. After this horse was diagnosed, more horses were noted to have lethargy, fever, and occasional colic signs. Fifteen horses were presumed to be initially infected with coronavirus based on these clinical signs. Fecal PCR testing was performed on only 2 additional cases; however, they were negative for equine coronavirus. Approximately 2 weeks later, horses were noted to have submandibular lymphadenopathy and were confirmed to have *S. equi* infection. Quarantine of affected horses was attempted; however, during the course of the outbreak, horses in all housing areas had clinical signs. Movement on and off the property was stopped.

Forty-eight horses were on the property during the *S. equi* outbreak. Ages ranged from 6 to 36 years (mean 15.8 years). Breeds included Quarter horse (QH) ($n = 17$), Tennessee Walking horse (TWH) ($n = 4$), Arabian ($n = 4$), Paint ($n = 4$), Thoroughbred ($n = 2$), Warmblood ($n = 2$), Morgan ($n = 2$), and 12 other breeds ($n = 1$ each). Thirteen (27%) were females, and 35 (73%) were castrated males.

Twenty-eight (58%) horses developed clinical signs consistent with strangles. Of the affected horses, ages ranged from 6 to 36 years (mean 16.1 years). Breeds included QH ($n = 10$), Paint ($n = 4$), TWH ($n = 3$), Arabian ($n = 3$), Warmblood ($n = 2$), Morgan ($n = 2$), and 4 other breeds ($n = 1$ each). Seven (25%) were females, and 21 (75%) were castrated males. Signalment of affected horses was similar to the whole exposed population. Eight horses had a history of previous vaccination for *S. equi*. Five out of the 8 vaccinated horses (63%) did not develop clinical signs of disease, and 3 of 8 (38%) did develop clinical signs.

At 4 weeks after initial detection of *S. equi*, no new horses developed clinical signs, and clinical signs of affected horses had resolved. Testing to detect persistent GP infection was performed at 8 weeks

after initial infection in conjunction with measurement of SeM antibody titer. Thirty-five horses were tested for persistent GP infection via either nasopharyngeal lavage ($n = 17$) or GP endoscopy and lavage ($n = 18$) *S. equi* PCR and culture. The decision to perform nasopharyngeal lavage or GP endoscopy was based on attending veterinarian recommendations and client preference. Thirteen horses were not tested for persistent GP infection status based on owners declining testing. Twelve of these horses were categorized as having no signs of disease, and 1 was categorized as an uncomplicated case of strangles.

Of the 28 horses affected, 11 (39%) had uncomplicated strangles, 9 (21%) had persistent GP infection, 5 (18%) had complicated cases, and 3 (11%) had both persistent GP infection and complications. Of the horses with persistent GP infection, 3 of 12 (25%) had endoscopically visible chondroids. Of the 8 complicated cases, 3 had purpura hemorrhagica, 3 had metastatic abscess formation, 1 had secondary pleuropneumonia, and 1 had dysphagia. The mean age was similar for each disease category. Of the 3 vaccinated horses that developed clinical signs, 1 had uncomplicated strangles, 1 had persistent GP infection, and 1 had both persistent GP infection and complications. Twenty-four of the 28 affected horses (86%) survived long term (>6 months after infection), whereas 4 of 28 (14%) were euthanized. The horses that were euthanized were all complicated cases, including metastatic abscess formation, infarctive purpura hemorrhagica, secondary pleuropneumonia, and dysphagia.

3.2 | *Streptococcus equi* subspecies *equi* M protein antibody titers

Streptococcus equi subspecies *equi* M antibody titers on all 48 horses at 8 weeks after infection are shown in Figure 1. One (2%) had a titer of 1:400, 7 (15%) had a titer of 1:800, 9 (19%) had a titer of 1:1600, 7 (15%) had a titer of 1:3200, 8 (17%) had a titer of 1:6400, 12 (25%) had a titer of 1:12 800, and 4 (8%) had a titer of $\geq 1:25 600$. The mean age was similar for each SeM antibody titer. Figure 1 shows the distribution of SeM antibody titers of horses with and without any clinical signs of *S. equi* infection. The median SeM antibody titers for horses with and without clinical signs were 1:12 800 and 1:1600, respectively. A correlation for horses with clinical signs to have SeM antibody titers $\geq 1:6400$ (Pearson's $R = 0.76$, $P < .05$) was noted. *S. equi* M

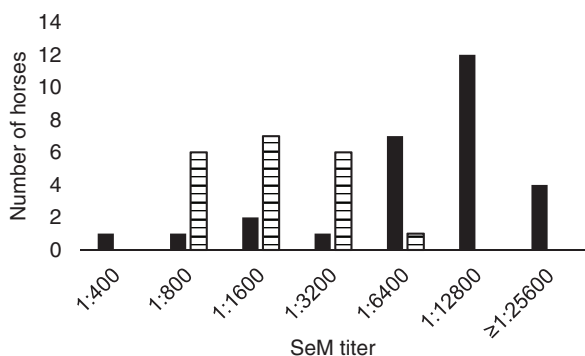


FIGURE 1 *Streptococcus equi* M protein (SeM) antibody titers of horses with any clinical signs (solid black, $n = 28$) versus horses without any clinical signs (horizontal lines, $n = 20$) 8 weeks after infection

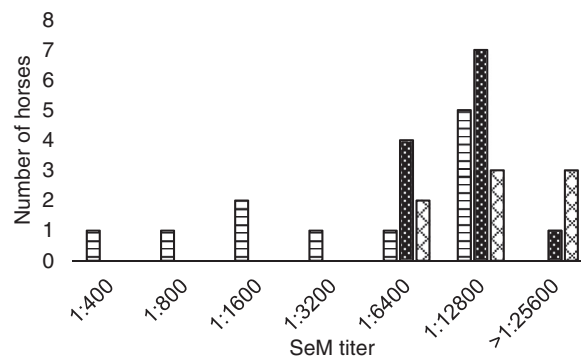


FIGURE 2 *Streptococcus equi* M protein (SeM) antibody titers of affected horses according to clinical syndromes 8 weeks after infection. Horizontal lines, uncomplicated cases ($n = 11$); black with white dots, persistent guttural pouch (GP) infection ($n = 12$); diamonds, complicated cases ($n = 8$)

protein antibody titers of affected horses were compared to their specific clinical syndromes (Figure 2). The median SeM antibody titer was 1:6400 for uncomplicated cases and was 1:12 800 for both persistent GP infection and complicated cases. A correlation for horses with persistent GP infection or complicated cases to have SeM antibody titers $\geq 1:6400$ (Pearson $R = 0.58$, $P < .05$; Pearson $R = 0.45$, $P < .05$, respectively) was also found. Six out of 16 (38%) horses with very high SeM antibody titers $\geq 1:12 800$ had evidence of complicated cases (Figure 2). *S. equi* M protein antibody titers of nonsurvivors ($n = 4$) were all $\geq 1:6400$. For this outbreak, sensitivity and specificity for a SeM antibody titer $\geq 1:12 800$ detecting complications were 75% (95% CI 45-105) and 43% (95% CI 23-64), respectively. Using a cutoff of $\geq 1:6400$ instead, the sensitivity and specificity for detecting complications were 100% (95% CI 100-100) and 22% (95% CI 5-39), respectively. All horses with persistent GP infection had SeM antibody titers $\geq 1:6400$, and 8 of 12 had a SeM antibody titer $\geq 1:12 800$. Sensitivity and specificity for SeM antibody titer $\geq 1:12 800$ detecting persistent GP infection were 67% (95% CI 40-93) and 42% (95% CI 20-64), respectively. Using a cutoff of $\geq 1:6400$ instead, the sensitivity and specificity for detecting persistent GP infection were 100% (95% CI 100-100) and 26% (95% CI 7-46), respectively.

Of the 8 vaccinated horses, 2 had a titer of 1:800, 1 had a titer of 1:1600, 2 had a titer of 1:3200, 1 had a titer of 1:6400, and 2 had a titer of 1:12 800. The median titer of vaccinated horses (1:3200) compared to unvaccinated horses (1:6400) was not significantly different ($P = .19$).

At 12 weeks after infection, SeM antibody titers were measured on 18 horses. Previous titers on these horses were 1:800 ($n = 1$), 1:3200 ($n = 1$), 1:6400 ($n = 3$), 1:12 800 ($n = 11$), and $> 1:25 600$ ($n = 1$). At the 12 week time point, 1 had a titer of 1:400, 5 had a titer of 1:3200, 9 had a titer of 1:6400, and 3 had a titer of 1:12 800 (Figure 3). One horse with a titer of 1:12 800 was being treated for metastatic abscess formation, the second horse with a titer of 1:12 800 had a persistent GP infection with no evidence of other complications, and the third horse with a titer of 1:12 800 had no clinical evidence of disease or complications. Two other horses at 12 weeks after infection had a persistent GP infection and had a SeM antibody titers $\leq 1:6400$. Fifteen out of 18 horses at this time point had a decrease in SeM antibody titer. At 28 weeks after infection, SeM antibody titers were measured on

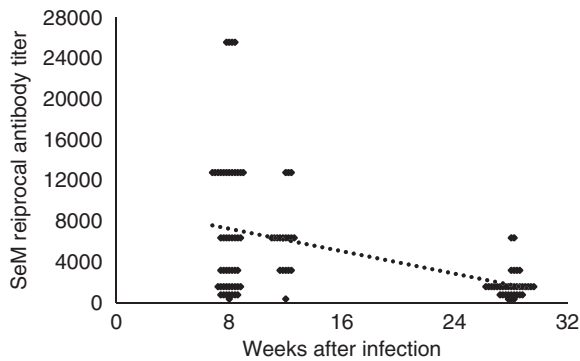


FIGURE 3 Scatterplot of *Streptococcus equi* M protein (SeM) reciprocal antibody titers at 8 weeks after infection ($n = 48$), 12 weeks after infection ($n = 12$), and 28 weeks after infection ($n = 36$). Regression line illustrates immunologic decay of mean SeM reciprocal antibody titers at each time point ($r = -0.44$, $P = .001$)

36 horses. This included all horses that were still on the property or in the area. Eight horses were lost to follow-up and 4 had been euthanized secondary to *S. equi* infection. At the 28 week time point, 3 of 36 (8%) had a titer of 1:400, 9 (25%) had a titer of 1:800, 18 (50%) had a titer of 1:1600, 4 (11%) had a titer of 1:3200, 2 (6%) had a titer of 1:6400, and 0 (0%) had titers $\geq 1:12\ 800$ (Figure 3). No horse at this time point had evidence of clinical disease. Thirty out of 36 horses demonstrated a decline in SeM antibody titer. There was a decline ($r = -0.44$, $P = .001$) in mean SeM reciprocal antibody titer at each time point: 7292 (range 400–25 600) at 8 weeks after infection, 6244 (range 400–12 800) at 12 weeks after infection, and 1744 (range 400–6400) at 28 weeks after infection (Figure 3).

4 | CONCLUSIONS AND CLINICAL IMPLICATIONS

Thirty-three percent of horses in this outbreak had SeM antibody titers $\geq 1:12\ 800$ eight weeks after infection. This is in contrast to typical outbreaks and previous SeM antibody titer interpretation.^{1,2} A recent study measured SeM antibody titers 1.5 to 27.5 months after natural infection.⁶ At 1.5 months after infection, none of the 45 horses tested had a SeM antibody titer $\geq 1:12\ 800$.⁶ SeM antibody titers $\geq 1:12\ 800$ occur after experimental commingling infections.⁷ SeM antibody titers peak at 5 weeks after experimental infection, in which SeM-specific IgGa titers rise to $1:38\ 400 \pm 14\ 021$ and SeM-specific IgGb titers rise to $1:102\ 400 \pm 56\ 089$.⁷ That experimental study did not describe if horses developed any complications after outbreak.⁷ In our outbreak, 6 out of 8 horses with complications had SeM antibody titers $\geq 1:12\ 800$, and 2 of 8 complicated cases had SeM antibody titers of 1:6400. One of these cases with dysphagia caused by severe GP disease was later euthanized. The other case had mild signs of purpura hemorrhagica which resolved without the use of immunosuppressive medications. These cases illustrate that SeM antibody titers after infection might not follow the typical interpretations of a titer $\geq 1:12\ 800$ supporting a diagnosis of metastatic abscess formation or purpura hemorrhagica^{1,2}; however, an increased titer would warrant further diagnostics to rule out *S. equi* complications.

Of the horses in this outbreak with SeM antibody titers $\geq 1:12\ 800$ at 8 weeks after infection, 5 of 16 (31%) had uncomplicated cases, 5 of 16 (31%) had persistent GP infection, 3 of 16 (19%) had complicated cases, and 3 of 16 (19%) had both persistent GP infection and complicated cases (Figure 2). Based on these data and the calculated sensitivity and specificity, SeM antibody titers might be useful for monitoring for complications and risk of complications after outbreak because of high sensitivity but do not necessarily confirm complications are present because of low specificity. *S. equi* M protein antibody titers do not correlate with a persistent GP infection.^{1,8,9} In our outbreak, 12 horses had persistent GP infection. At 8 weeks after infection, all of these horses (100%) had SeM antibody titers $\geq 1:6400$ and 8 of 12 (67%) had SeM antibody titers $\geq 1:12\ 800$. The high SeM antibody titers in these horses likely relate to continual exposure to the organism eliciting an immune response. At a SeM antibody titer $\geq 1:12\ 800$, the sensitivity and specificity of detecting persistent GP infection were low (67% and 42%, respectively); however, when using the cutoff of $\geq 1:6400$, the sensitivity was 100%, but the specificity was unacceptable (26%). A correlation applicable to other populations or outbreaks cannot be determined based on this outbreak, but an increased SeM antibody titer after outbreak could warrant additional diagnostics including testing for carrier status via nasopharyngeal or GP lavage.

This reported outbreak is novel because there was a high proportion of horses with SeM antibody titers $\geq 1:12\ 800$ (33%). There was also a high proportion of horses with complicated disease (29%) compared to reported complication rates (2%–20%).^{1,2,4} In this outbreak, case fatality in complicated cases (50%) was high compared to reported case fatality in complicated cases (up to 40%).² Lastly, there was a high proportion of horses developing persistent GP infection (43%); this is higher than previous reports (up to 10%)² but consistent with a more recent report (up to 40%).^{1,4} Possible explanations for the higher complication rates and higher convalescent SeM antibody titers include a higher dose of exposure, a more virulent *S. equi* strain,^{10,11} or a more naïve population. The causative *S. equi* organism was not sequenced for the M protein gene in this outbreak. The role of possible concurrent equine enteric coronavirus infections contributing to high complication rates in this outbreak is unknown. It could have resulted in delayed detection and quarantine specific for *S. equi* transmission, or it could have resulted in immunosuppression allowing more sequelae.

Repeat SeM antibody titers revealed immunologic decay in most cases at each time point consistent with resolution of disease, discontinued exposure to the organism, and are consistent with previous reports of experimental infection.⁷ As late as 12 weeks after infection, 3 horses had SeM antibody titers of 1:12 800. One horse was being treated for metastatic abscess formation, and the titer was declining consistent with response to treatment. At 28 weeks after infection, all horses had SeM antibody titers $< 1:12\ 800$. This is similar to a recent outbreak that reported 4.2% of horses having SeM antibody titers $\geq 1:12\ 800$ after outbreak in which serologic testing was performed at 8 months after infection.¹² High titers (1:3200 or 1:6400) persisted in 6 horses at 28 weeks after infection which would have been previously interpreted as consistent with 4–12 weeks after infection.² Declining but persistently increased serum SeM-specific IgGb titer levels were previously reported at 28 weeks after experimental infection.⁷

Limitations of interpreting data from this outbreak are that the exact dates of infection and times of resolution of clinical signs in each horse were unknown and SeM antibody titers were unknown before the outbreak. Peak SeM antibody titers could not have been detected as SeM antibody titers peak at 5 weeks after experimental infection,⁷ and serologic testing and testing for carrier status was performed 8 weeks after initial infection in this report. Additionally a selection bias likely existed at the 12 week after infection time point at which the majority of horses rechecked had a previous titer of $\geq 1:6400$. Lastly, 13 horses were not tested for carrier status and were presumed not to have persistent GP infection; however, this could have led to misclassification. At initial testing for carrier status, some horses were tested only once with nasopharyngeal lavage largely based on economics; however, the current recommendations for detection of carrier status are endoscopy and GP lavage.^{1,4} This limitation could have led to underestimating the number of horses with persistent GP infection in this outbreak. It could have also led to overestimating the sensitivity and specificity of SeM antibody titers for detecting persistent GP infection as all of these horses had a SeM antibody titer $\leq 1:3200$; however, this outbreak and other studies have indicated that SeM titers are not useful in this manner.^{1,8,9}

This outbreak illustrates the utility of SeM antibody titers after infection with *S. equi*. This study demonstrates that a horse may have complications of strangles without a SeM antibody titer $\geq 1:12\ 800$ and that a horse may have a SeM antibody titer $\geq 1:12\ 800$ without complications. A convalescent SeM antibody titer $\geq 1:12\ 800$ warrants additional investigation for complications or persistent GP infection but does not necessarily confirm a horse has complicated disease.

ACKNOWLEDGMENTS

Results of this study were presented as an abstract at the 2018 American College of Veterinary Internal Medicine Forum, Seattle, WA.

OFF-LABEL ANTIMICROBIAL DECLARATION

There was not off-label antimicrobial use.

CONFLICT OF INTEREST DECLARATION

Dr John Timoney shares a patent on the SeM sequence (US Patent # 6458358 October 1, 2002). Dr Jennifer Morrow is a co-owner of Equine Diagnostic Solutions LLC, Lexington, KY where sample analysis (SeM antibody titer ELISA) was performed.

INSTITUTIONAL ANIMAL CARE AND USE COMMITTEE (IACUC) OR OTHER APPROVAL DECLARATION

Authors declare no IACUC or other approval was needed. Informed client consent was obtained.

HUMAN ETHICS APPROVAL DECLARATION

Authors declare human ethics approval was not needed for this study.

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