




ORIGINAL RESEARCH

The effect of water immersion and acute hypercapnia on ventilatory sensitivity and cerebrovascular reactivity

James R. Sackett , Zachary J. Schlader , Carol Cruz, David Hostler & Blair D. Johnson 

Center for Research and Education in Special Environments, Department of Exercise and Nutrition Sciences, University at Buffalo, Buffalo, New York

Keywords

Central chemoreceptors, elevated CO₂ pressure, head out water immersion, hemodynamics, ventilation.

Correspondence

Blair D. Johnson, Center for Research and Education in Special Environments, Department of Exercise and Nutrition Sciences, University at Buffalo, 208A Kimball Tower, Buffalo, NY 14214.
Tel: +1 (716) 829 6789
Fax: +1 (716) 829 2428
E-mail: blairjoh@buffalo.edu

Funding Information

This research was supported by the NASA Space Physiology Research Grant from the American College of Sports Medicine Foundation, the Mark Diamond Research Fund of the Graduate Student Association at the University at Buffalo, and the Office of Naval Research Award N00014-17-1-2665.

Received: 20 August 2018; Revised: 14 September 2018; Accepted: 19 September 2018

doi: 10.14814/phy2.13901

Physiol Rep, 6 (19), 2018, e13901,
<https://doi.org/10.14814/phy2.13901>

Abstract

The partial pressure of end tidal carbon dioxide (PETCO₂), ventilatory sensitivity to CO₂, and cerebral perfusion are augmented during thermoneutral head out water immersion (HOWI). We tested the hypotheses that HOWI and acute hypercapnia augments minute ventilation, ventilatory sensitivity to CO₂, cerebral perfusion, and cerebrovascular reactivity to CO₂. Twelve subjects (age: 24 ± 3 years, BMI: 25.3 ± 2.9 kg/m², 6 women) participated in two experimental visits: a HOWI visit (HOWI) and a matched hypercapnia visit (Dry + CO₂). A rebreathing test was conducted at baseline, 10, 30, 60 min, and post HOWI and Dry + CO₂. PETCO₂, minute ventilation, expired gases, blood pressure, heart rate, and middle cerebral artery blood velocity were recorded continuously. PETCO₂ increased throughout HOWI (baseline: 42 ± 2 mmHg; maximum at 10 min: 44 ± 2 mmHg, *P* ≤ 0.013) and Dry + CO₂ (baseline: 42 ± 2 mmHg; maximum at 10 min: 44 ± 2 mmHg, *P* ≤ 0.013) and was matched between conditions (condition main effect: *P* = 0.494). Minute ventilation was lower during HOWI versus Dry + CO₂ (maximum difference at 60 min: 13.2 ± 1.9 vs. 16.2 ± 2.7 L/min, *P* < 0.001). Ventilatory sensitivity to CO₂ and middle cerebral artery blood velocity were greater during HOWI versus Dry + CO₂ (maximum difference at 10 min: 2.60 ± 1.09 vs. 2.20 ± 1.05 L/min/mmHg, *P* < 0.001, and 63 ± 18 vs. 53 ± 14 cm/sec, *P* < 0.001 respectively). Cerebrovascular reactivity to CO₂ decreased throughout HOWI and Dry + CO₂ and was not different between conditions (condition main effect: *P* = 0.777). These data indicate that acute hypercapnia, matched to what occurs during HOWI, augments minute ventilation but not ventilatory sensitivity to CO₂ or middle cerebral artery blood velocity despite an attenuated cerebrovascular reactivity to CO₂.

Introduction

Thermoneutral (~35°C) head out water immersion (HOWI) induces central hypervolemia, which causes several hemodynamic alterations, such as immediate increases in stroke volume, cardiac output, pulmonary blood flow, and cerebral blood flow (Arborelius et al. 1972; Farhi et al. 1977; Carter et al. 2014), while heart

rate is slightly reduced (Farhi et al. 1977). Despite this, blood pressure does not change (Bonde-Petersen et al. 1992; Sramek et al. 2000; Watenpaugh et al. 2000) or is slightly attenuated (Craig and Dvorak 1966; Sackett et al. 2017) during HOWI due to a reduction in total peripheral resistance. The hydrostatic pressure of the water increases the work of breathing (i.e., static lung load) (Lambertsen 1977; Moon et al. 2009), although minute

ventilation does not change during HOWI compared to dry conditions (Sackett *et al.* 2017, 2018). Nevertheless, the partial pressure of end tidal carbon dioxide (PETCO₂) is increased during HOWI (Sackett *et al.* 2017, 2018), which indicates carbon dioxide (CO₂) retention (Lanphier and Bookspan 1999).

Typically, increases in arterial CO₂ pressure stimulate the peripheral (Prabhakar and Peng 2004; Kumar and Prabhakar 2012) and central chemoreceptors (Nattie and Li 2012; MacKay *et al.* 2016) to increase ventilation in an attempt to lower arterial CO₂ pressure. The absent increase in ventilation during HOWI, in the presence of elevated PETCO₂, suggests that a small increase in PETCO₂ is not sufficient to stimulate the chemoreceptors and cause a rise in ventilation. We have previously demonstrated that peripheral chemosensitivity to hypercapnia is not altered during thermoneutral HOWI (Sackett *et al.* 2017), whereas central chemosensitivity (*i.e.*, ventilatory sensitivity to CO₂) is augmented during thermoneutral HOWI (Sackett *et al.* 2018). Yet, it is not currently known if the increase in ventilatory sensitivity to CO₂ during HOWI is due to the background of elevated PETCO₂ alone or the combined effects of HOWI (*i.e.*, elevated PETCO₂, central hypervolemia, increased cerebral perfusion, increased work of breathing, *etc.*).

Increases in arterial CO₂ pressure augment cerebral blood flow to enhance hydrogen “washout” at the site of the central chemoreceptors (Ainslie and Duffin 2009). The concomitant increase in PETCO₂ and cerebral perfusion suggests that a link between the two could exist during HOWI (Carter *et al.* 2014). However, it is not clear if the rise in cerebral perfusion during HOWI is due to acute hypercapnia or the combined effects of HOWI, including elevated PETCO₂. Furthermore, alterations in cerebral blood flow appear to modulate changes in cerebrovascular reactivity to CO₂, such that increases in cerebral blood flow enhance cerebrovascular reactivity to CO₂, while decreases in cerebral blood flow blunt cerebrovascular reactivity to CO₂. Moreover, an attenuated cerebrovascular reactivity to CO₂ enhances ventilatory sensitivity to CO₂ (Fan *et al.* 1985; Xie *et al.* 2005; Ainslie *et al.* 2007), which indicates an important link between cerebrovascular reactivity to CO₂ and ventilatory sensitivity to CO₂. However, the effect of an augmented cerebrovascular reactivity to CO₂ on ventilatory sensitivity to CO₂ remains equivocal (Chapman *et al.* 1979). It is possible that an augmented cerebral blood flow, which occurs in the presence of elevated PETCO₂ during HOWI, may enhance cerebral CO₂ delivery and contribute to subsequent alterations in cerebrovascular reactivity to CO₂ and ventilatory sensitivity to CO₂. Yet, the independent effects of acute hypercapnia and HOWI on cerebrovascular reactivity to CO₂ are not currently known.

The purpose of our study was to determine the effect of water immersion and acute hypercapnia on ventilatory sensitivity to CO₂ and cerebrovascular reactivity to CO₂. To test our hypotheses, we compared a HOWI visit (HOWI) to a dry condition where we matched the hypercapnia that occurred during HOWI (Dry + CO₂). We hypothesized that: (1) minute ventilation and ventilatory sensitivity to CO₂ are augmented during both HOWI and Dry + CO₂ and (2) cerebral perfusion and cerebrovascular reactivity to CO₂ are increased during both HOWI and Dry + CO₂.

Methods

Subjects

Twelve subjects (age: 24 ± 3 years, BMI: 25.3 ± 2.9 kg/m², 6 women) completed three visits: a screening visit and two experimental visits. Subjects reported to be recreationally active, nonsmokers, not taking medications, and free from any known cardiovascular, metabolic, neurological, or psychological disease. Women were not pregnant (confirmed via a urine pregnancy test). To control for menstrual cycle hormones, women were tested during the first 10 days following self-identified menstruation (Minson *et al.* 2000). During the screening visit, subjects gave written consent following a comprehensive explanation of the experimental procedures and possible risks. The study was approved by the Institutional Review Board at the University at Buffalo and the study was performed in accordance with the standards set forth by the latest version of the Declaration of Helsinki.

Instrumentation and measurements

Height and weight were measured with a stadiometer and scale (Sartorius Corp., Bohemia, NY, USA) and urine specific gravity was measured using a refractometer (Atago USA, Inc., Bellevue, WA, USA) prior to each experimental visit. Previous reports indicate that urine specific gravity is a valid indicator of hydration status (Armstrong *et al.* 1998). The CO₂ pressure waveform was measured via a capnograph (Nonin Medical, Inc., Plymouth, MN, USA), of which the minimum value represented the inspired pressure of CO₂ and the maximum value (*i.e.*, end tidal) represented the expired pressure of CO₂ (*i.e.*, PETCO₂). The sample rate of the capnograph was 4 Hz. PETCO₂ was used as a marker of arterial CO₂ pressure since it reflects arterial CO₂ pressure during water immersion (Dunworth *et al.* 2017) and throughout a wide range of physiological dead space (McSwain *et al.* 2010), which may be increased during water immersion (Cherry *et al.* 2009). Inspired and expired ventilation was

measured using a heated pneumotachometer (Hans Rudolph, Inc., Shawnee, KS, USA) that was attached to a silicone mouthpiece (Hans Rudolph, Inc., Shawnee, KS, USA). The fraction of expired CO₂ was measured via a CO₂ analyzer (Vacu Med, Venture, CA, USA), which was sampled at a 3 L mixing chamber. Beat to beat blood pressure was measured via photoplethysmography (ccNexfin Bmeye NA, St. Louis, MO, USA) on the left hand, which was suspended slightly above the water during HOWI. Blood pressure was corrected to the heart level using a height correction sensor. Heart rate was measured from a three-lead electrocardiograph (DA100C, Biopac Systems, Inc., Goleta, CA, USA). Stroke volume was determined via the arterial pressure waveform using Modelflow (ccNexfin Bmeye NA, St. Louis, MO, USA) (Wesseling *et al.* 1993). Cardiac output was calculated as the product of heart rate and stroke volume. Total peripheral resistance was calculated as mean arterial pressure divided by cardiac output. Right middle cerebral artery blood velocity was measured via transcranial Doppler sonography using a 2 MHz probe (DWL USA, Inc., Germany, Europe) by the same research technician throughout all experimental visits. After a quality Doppler signal of middle cerebral artery blood velocity was obtained, the depth, gain, and location were recorded to be used for the second study visit. Middle cerebral artery conductance was calculated as middle cerebral artery blood velocity divided by mean arterial pressure.

Minute ventilation, tidal volume, and respiratory rate were determined using the breath by breath respiratory analysis feature of the data analysis software (AcqKnowledge 4.2, Goleta, CA, USA). Abhorrent breaths (e.g., sigh, breath hold, cough, etc.) were manually excluded and ventilation data are presented in body temperature and pressure, saturated (BTPS). The fraction of inspired CO₂ was calculated as the inspired pressure of CO₂ multiplied by the barometric pressure. The rate of CO₂ production ($\dot{V}CO_2$) was calculated as minute ventilation multiplied by the fraction of expired CO₂ minus the fraction of inspired CO₂. Alveolar ventilation was calculated as the product of $\dot{V}CO_2$ and 863 divided by PETCO₂ (West 2012), and dead space ventilation was calculated as minute ventilation minus alveolar ventilation. The alveolar ventilation to cardiac output ratio was calculated as an index of the alveolar ventilation to pulmonary perfusion ratio (Derion *et al.* 1992; Levitzky 2013).

Rebreathing test

A rebreathing test was used to assess central chemoreceptor function (Read 1967; Rebuck 1976; MacKay *et al.* 2016). Subjects rebreathed 7% CO₂ and 93% O₂ for 3.5 min from a custom made 10 L anesthesia bag (Read

1967). The volume of the gas in the bag was equal to predicted vital capacity (Casio, 2018) plus 1 L. Data from the first 30 sec of the test were not used during analysis as this is when the CO₂ pressure waveform is entering equilibrium.

Ventilatory sensitivity to CO₂ was determined by plotting mean minute ventilation versus mean PETCO₂ every 30 sec throughout the rebreathing test. Ventilatory sensitivity to CO₂ data are reported as the slope of the linear regression line of minute ventilation versus PETCO₂. The ventilatory threshold to CO₂ is represented by the *x*-intercept of the linear regression line of mean minute ventilation versus mean PETCO₂. This value represents the theoretical PETCO₂ value at which minute ventilation is 0 L/min (Read 1967). We also calculated the rate at which minute ventilation and PETCO₂ increased over time throughout each rebreathing test. These data provide insight as to which variable (i.e., minute ventilation or PETCO₂) is contributing to changes in ventilatory sensitivity to CO₂. The increase in minute ventilation and PETCO₂ over time were calculated as the change in minute ventilation and PETCO₂, respectively, from the beginning to the end of the test, divided by the change in time (i.e., 3.5 min) (Fowle and Campbell 1964).

We calculated cerebrovascular reactivity to CO₂ during the rebreathing test as an indicator of the sensitivity of middle cerebral artery blood velocity to changes in PETCO₂ (MacKay *et al.* 2016). This was determined by plotting mean middle cerebral artery blood velocity versus mean PETCO₂ every 30 sec throughout the rebreathing test. Cerebrovascular reactivity to CO₂ data are reported as the slope of the linear regression line of middle cerebral artery blood velocity versus PETCO₂ (MacKay *et al.* 2016).

Experimental approach

The two experimental visits, in order, included: (1) a HOWI visit (HOWI) and (2) a matched hypercapnia visit performed in dry conditions (Dry + CO₂). The HOWI visit always occurred first in order to establish the level of hypercapnia that was matched during Dry + CO₂. The experimental visits did not occur on consecutive days and took place within a maximum of 6 days for all subjects. Subjects arrived at the laboratory having refrained from exercise, alcohol, and caffeine for 12 h, and food for 2 h for both experimental visits. Subjects also arrived to the laboratory euhydrated for both HOWI (urine specific gravity: 1.009 ± 0.005) and Dry + CO₂ (urine specific gravity: 1.009 ± 0.007) visits. All data collection took place in a temperature controlled laboratory (24 ± 2°C, 41 ± 14% relative humidity) while subjects viewed a

nonstimulating documentary and were encouraged to breathe normally. Following at least 10 min of seated rest, a baseline rebreathing test commenced. Upon completion of the baseline measurements, the subjects emptied their bladder and either entered the pool (HOWI) or continued seated rest (Dry + CO₂) for 1 h. Over the next hour, a rebreathing test was performed at 10, 30, and 60 min. During HOWI, subjects were seated in thermoneutral water (35.0 ± 0.1°C) up to the neck. During Dry + CO₂, small amounts of CO₂ were added to the inspirate from a pre-mixed gas tank (i.e., 13% CO₂, 21% O₂, and 66% N₂) to match PETCO₂ values that were obtained during HOWI. CO₂ was added at the inspired side of a two-way nonrebreathing valve (Hans Rudolph, Inc., Shawnee, KS, USA), such that the fraction of inspired CO₂ induced hypercapnia. The flow of the added CO₂ gas mixture was titrated to match the PETCO₂ that occurred during the HOWI visit. After one hour of HOWI or Dry + CO₂, subjects exited the pool or the addition of CO₂ to the inspirate ceased. At this point, the subjects emptied their bladder and a rebreathing test commenced after 10 min of seated rest (i.e., post).

Data and statistical analyses

Ventilatory data were captured at 62 Hz and hemodynamic data were obtained at 1 kHz by a data acquisition system (Biopac MP 150, Goleta, CA, USA). Data were stored on a personal computer for offline analyses. Data were assessed for approximation to a normal distribution, and an outlier analysis was performed. Outliers were identified and removed using the ROUT method (Motulsky and Brown 2006), in which the Q value, or the false discovery rate, was set conservatively (i.e., 0.1%) so that only definitive outliers were removed. Outliers were only removed from statistical analyses for ventilatory threshold to CO₂ ($n = 2$). Following the outlier analysis, data were analyzed using a two-way repeated measures ANOVA. If a significant interaction or main effect was found, the Holm-Sidak multiple comparisons post hoc test was used to determine where differences existed. All data were analyzed using Prism software (Version 6, GraphPad Software Inc., La Jolla, CA, USA) and data are reported as mean ± SD. Significance was set a priori to $P < 0.05$ and exact P -values are reported where possible.

Results

Body weight and urine loss

Reductions in body weight were not different following HOWI versus Dry + CO₂ (0.68 ± 0.37 vs. 0.58 ± 0.35

kg, $P = 0.125$). However, urine loss was greater during HOWI versus Dry + CO₂ (0.57 ± 0.38 vs. 0.44 ± 0.37 L, $P = 0.046$).

Ventilation

PETCO₂ (Fig. 1A) was greater than baseline during HOWI and Dry + CO₂ at 10 min ($P < 0.001$), 30 min ($P < 0.001$), 60 min ($P < 0.001$), and post ($P = 0.013$) and was successfully matched between conditions so that there was not a condition effect (condition main effect: $P = 0.494$) or an interaction effect (interaction main effect: $P = 0.674$). Minute ventilation (Fig. 1B) and alveolar ventilation (Fig. 1C) were lower during HOWI versus Dry + CO₂ at 10 min ($P \leq 0.001$), 30 min ($P \leq 0.003$), and 60 min ($P \leq 0.001$). Dead space ventilation (Fig. 1D) was greater than baseline during Dry + CO₂ at 10 min ($P = 0.011$) and 30 min ($P = 0.009$) but was not different during HOWI versus Dry + CO₂ at any time point (condition main effect: $P = 0.193$). Tidal volume (Fig. 1E) was greater during HOWI versus Dry + CO₂ at baseline ($P = 0.030$) and lower during HOWI versus Dry + CO₂ at 10 min ($P < 0.001$), 30 min ($P < 0.001$), and 60 min ($P < 0.001$). Respiratory rate (Fig. 1F) was greater than baseline throughout HOWI ($P \leq 0.003$) and Dry + CO₂ ($P \leq 0.026$) and was not different between HOWI and Dry + CO₂ (condition main effect: $P = 0.831$).

Hemodynamics

Mean arterial pressure (Fig. 2A) was lower during HOWI versus Dry + CO₂ at 10 min ($P < 0.001$), 30 min ($P < 0.001$), and 60 min ($P < 0.001$). Meanwhile, cardiac output (Fig. 2B) was greater than baseline during HOWI at 10 min ($P < 0.001$), 30 min ($P < 0.001$), and 60 min ($P < 0.001$), while total peripheral resistance (Fig. 2C) was lower than baseline during HOWI at 10 min ($P < 0.001$), 30 min ($P < 0.001$), and 60 min ($P < 0.001$). Cardiac output was greater during HOWI versus Dry + CO₂ at 10 min ($P < 0.001$), 30 min ($P < 0.001$), and 60 min ($P < 0.001$), while total peripheral resistance was lower during HOWI versus Dry + CO₂ at 10 min ($P < 0.001$), 30 min ($P < 0.001$), and 60 min ($P < 0.001$). Heart rate (Fig. 2D) was not different across time during HOWI ($P \geq 0.105$) or Dry + CO₂ ($P \geq 0.553$) or between conditions (condition main effect: $P = 0.419$). Stroke volume was greater during HOWI versus Dry + CO₂ at 10 min ($P < 0.001$), 30 min ($P < 0.001$), and 60 min ($P < 0.001$). The alveolar ventilation to cardiac output ratio (Fig. 2F) was less than baseline during HOWI at 10 min ($P < 0.001$), 30 min ($P < 0.001$), and 60 min ($P < 0.001$) and was lower during HOWI versus Dry + CO₂ at 10 min ($P < 0.001$), 30 min ($P < 0.001$), and 60 min ($P < 0.001$).

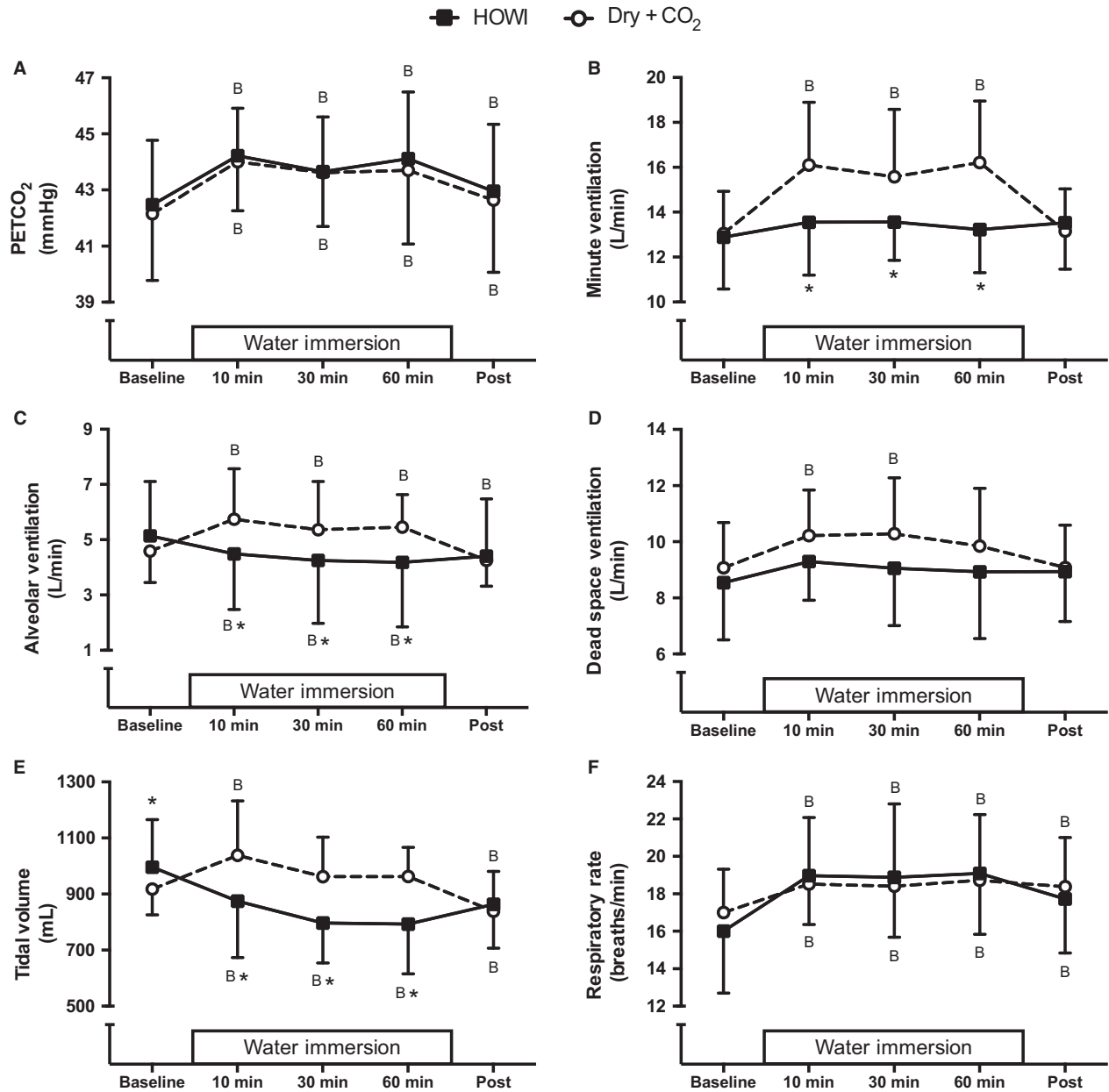


Figure 1. PETCO₂ (A), minute ventilation (B), alveolar ventilation (C), dead space ventilation (D), tidal volume (E), and respiratory rate (F) at baseline, 10, 30, 60 min, and post HOWI and Dry + CO₂. Water immersion only occurred during the HOWI visit. Values are mean ± SD. * = different from Dry + CO₂, $P < 0.050$. B = different from baseline, $P < 0.050$.

Cerebral hemodynamics

Middle cerebral artery blood velocity was greater than baseline during HOWI at 10 min ($P < 0.001$) and 30 min ($P = 0.004$). Middle cerebral artery conductance (Fig. 3B) was greater than baseline during HOWI at 10 min ($P < 0.001$) and 30 min ($P = 0.025$), while it was less than baseline during Dry + CO₂ at 30 min ($P = 0.004$) and 60 min ($P < 0.001$).

Rebreathing test

Ventilatory sensitivity to CO₂ (Fig. 4A) was greater than baseline during HOWI at 10 min ($P < 0.001$), 30 min ($P < 0.001$), 60 min ($P < 0.001$), and post ($P = 0.048$), while ventilatory sensitivity to CO₂ was not different than baseline throughout Dry + CO₂ ($P \geq 0.702$). Moreover, ventilatory sensitivity to CO₂ was higher during HOWI versus Dry + CO₂ at 10 min ($P = 0.007$), 30 min

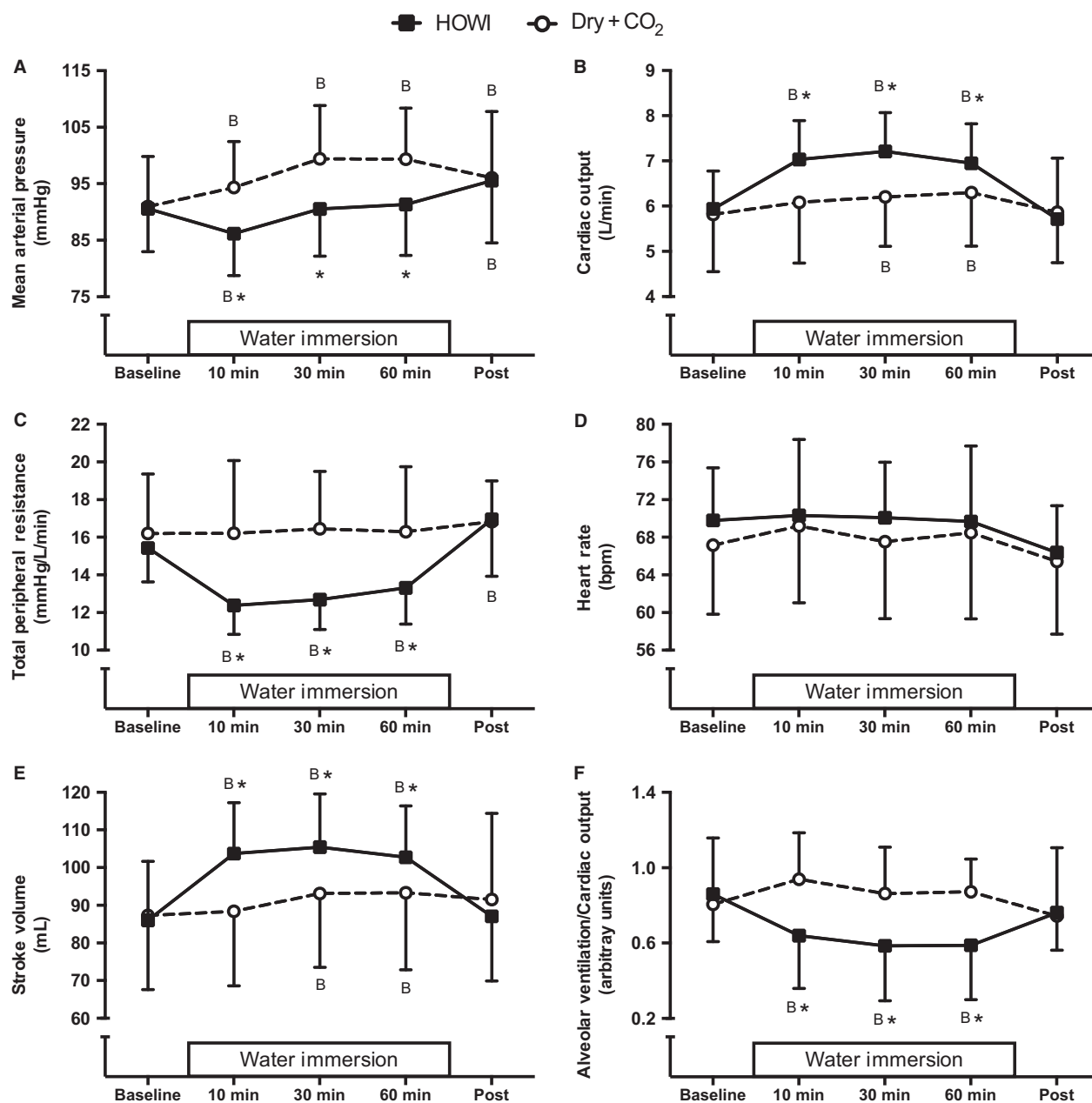


Figure 2. Mean arterial pressure (A), cardiac output (B), total peripheral resistance (C), heart rate (D), stroke volume (E), and alveolar ventilation to cardiac output ratio (F) at baseline, 10, 30, 60 min, and post HOWI and Dry + CO₂. Water immersion only occurred during the HOWI visit. Values are mean ± SD. * = different from Dry + CO₂, $P < 0.050$. B = different from baseline, $P < 0.050$.

($P = 0.040$), and 60 min ($P = 0.025$). There was not a time effect (time main effect: $P \geq 0.203$), condition effect (condition main effect: $P \geq 0.694$), or interaction effect (interaction main effect: $P \geq 0.054$) for ventilatory threshold to CO₂ (Fig. 4B) or the increase in minute ventilation over time (Fig. 4C). However, the increase in PETCO₂ over time (Fig. 4D) was lower during HOWI versus Dry + CO₂ at

10 min ($P < 0.001$), 30 min ($P < 0.001$), and 60 min ($P < 0.001$), and was greater during HOWI versus Dry + CO₂ at post ($P = 0.003$).

Cerebrovascular reactivity to CO₂ (Fig. 5) was less than baseline throughout HOWI ($P < 0.001$) and Dry + CO₂ ($P < 0.001$) but was not different between conditions (condition main effect: $P = 0.777$).

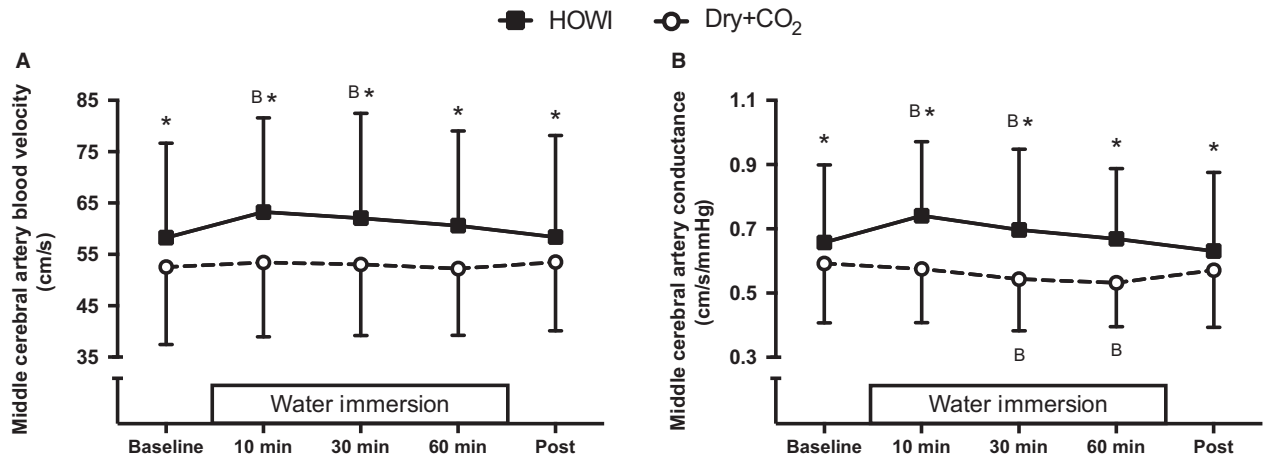


Figure 3. Middle cerebral artery blood velocity (A) and middle cerebral artery conductance (B) at baseline, 10, 30, 60 min, and post HOWI and Dry + CO₂. Water immersion only occurred during the HOWI visit. Values are mean ± SD. * = different from Dry + CO₂, *P* < 0.050. B = different from baseline, *P* < 0.050.

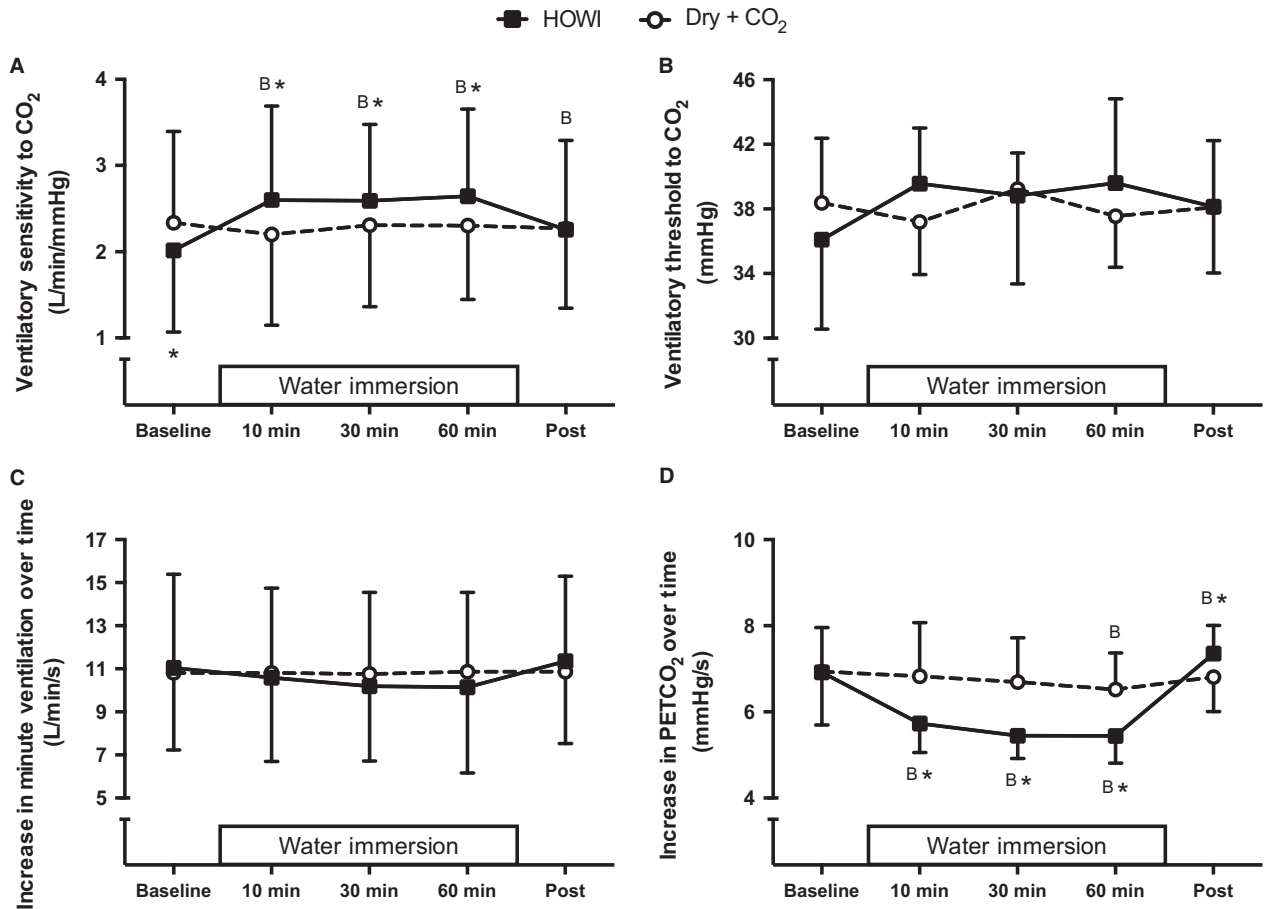


Figure 4. Ventilatory sensitivity to CO₂ (A), ventilatory threshold to CO₂ (B), increase in minute ventilation over time during the rebreathing test (C), and increase in PETCO₂ over time during the rebreathing test (D) at baseline, 10, 30, 60 min, and post HOWI or Dry + CO₂. Water immersion only occurred during the HOWI visit. Values are mean ± SD. * = different from Dry + CO₂, *P* < 0.050. B = different from baseline, *P* < 0.050.

Discussion

The novel findings of our investigation are despite a greater minute ventilation, ventilatory sensitivity to CO₂ was lower during dry conditions when breathing a hypercapnic gas to match the hypercapnia that occurs during HOWI when compared to HOWI. Moreover, cerebral perfusion was lower during dry conditions while breathing a hypercapnic gas to match the hypercapnia that occurs during HOWI when compared to HOWI, while cerebrovascular reactivity to CO₂ was not different between conditions. Thus, although acute hypercapnia increases minute ventilation, it does not augment ventilatory sensitivity to CO₂ or cerebral blood flow and it lessens cerebrovascular reactivity to CO₂. Thus, augmented ventilatory sensitivity to CO₂ and cerebral perfusion during HOWI appear to be a function of the combined effects of water immersion and not elevated PETCO₂ alone.

Ventilation

Acute hypercapnia was induced during Dry + CO₂ by adding small amounts of CO₂ to the inspirate of the breathing apparatus. Thus, inspired CO₂ levels were greater during Dry + CO₂ versus HOWI by design. Increasing PETCO₂ by raising inspired CO₂ (i.e., end tidal forcing) has been used to investigate the effects of acute hypercapnia (Slessarev et al. 2007; Koehle et al. 2009; Mark et al. 2010). It is currently unclear if the differences in inspired CO₂ contributed to the different ventilatory and hemodynamic responses that we observed in spite of tightly matching the PETCO₂ values between conditions. In this context, failure of CO₂ scrubbers increase inspired CO₂ during underwater diving while using a rebreather

system (Barlow and Macintosh 1944). Therefore, adding CO₂ to the inspirate is a valid approach to study various underwater diving scenarios. Nonetheless, future investigations should examine if other effects of water immersion (i.e., work of breathing, central hypervolemia, etc.) contribute to increase PETCO₂ during water immersion.

During Dry + CO₂, we accurately matched the increase in PETCO₂ that occurred during HOWI. This small, but significant, increase in PETCO₂ allowed us to investigate the effect of acute hypercapnia alone on the ventilatory sensitivity to CO₂ and cerebral perfusion. Despite that minute ventilation was unchanged throughout HOWI, the increase in PETCO₂ during Dry + CO₂ stimulated the chemoreceptors and increased minute ventilation. Similarly, alveolar ventilation was lower during HOWI versus Dry + CO₂. Previous findings indicate that a reduced alveolar ventilation may contribute to hypercapnia during hyperbaria (Salzano et al. 1984). To our knowledge, our investigation is the first to demonstrate alveolar hypoventilation during HOWI, as this has only previously been observed during investigations at depth (Salzano et al. 1984; Mummery et al. 2003; Cherry et al. 2009). The concomitant increases in ventilation and PETCO₂ during Dry + CO₂ suggest a shift of the respiratory operating point (i.e., minute ventilation vs. PETCO₂) (Miyamoto et al. 2014). Similarly, thermoneutral HOWI induces a shift of the respiratory operating point, and this is in agreement with previous investigations (Miyamoto et al. 2014; Sackett et al. 2017, 2018). The chemoreflex feedback system can be divided into two subsystems: i) the central controller and ii) the peripheral plant (Berger et al. 1977; Miyamoto et al. 2014). The central controller is described as the relation between minute ventilation and arterial CO₂ pressure, such that minute ventilation increases linearly as a function of arterial CO₂ pressure (i.e., ventilatory sensitivity to CO₂) (Berger et al. 1977). On the other hand, the peripheral plant is described as the relation between arterial CO₂ pressure and minute ventilation, such that arterial CO₂ pressure decreases asymptotically as a function of minute ventilation (Berger et al. 1977). In this context, a shift of the respiratory operating point appears to be driven by the central controller during HOWI, while it is driven by the peripheral plant during Dry + CO₂ (Ogoh et al. 2008; Miyamoto et al. 2014). However, the mechanisms underpinning these discrepancies are not clear. It has also been proposed that an increase in dead space ventilation might contribute to hypercapnia during water immersion (Salzano et al. 1984; Mummery et al. 2003; Cherry et al. 2009). Our data do not indicate an increase in dead space ventilation during HOWI despite a decrease in tidal volume and an increase in respiratory rate during HOWI. These changes in breathing pattern are thought to be a result of increased respiratory work during water immersion (Salzano et al. 1984;

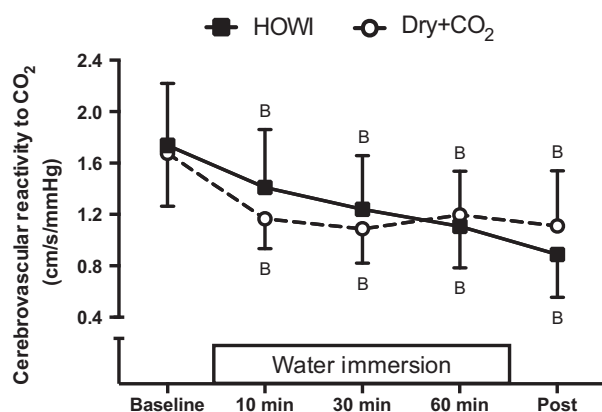


Figure 5. Cerebrovascular reactivity to CO₂ at baseline, 10, 30, 60 min, and post HOWI or Dry + CO₂. Water immersion only occurred during the HOWI visit. Values are mean ± SD. B = different from baseline, $P < 0.050$.

Moon et al. 2009; Ray et al. 2010). Such increases in respiratory work during HOWI may have prevented elevations in minute ventilation. To this end, an increase in tidal volume may be possible in the absence of increased respiratory work (i.e., Dry + CO₂) (Sackett et al. 2017, 2018). This was evident during Dry + CO₂ as increases in tidal volume and respiratory rate were achieved in the absence of increased respiratory work. Further investigations should elucidate if an increase in respiratory work causes a decrease in tidal volume during HOWI and subsequently contributes to hypercapnia during HOWI.

Hemodynamics

Mean arterial pressure increased throughout Dry + CO₂ but not during the HOWI visit until post HOWI and it was lower during HOWI versus Dry + CO₂ throughout water immersion. HOWI induced increases in cardiac output that were accompanied by decreases in total peripheral resistance, while Dry + CO₂ induced increases in cardiac output, without a change in total peripheral resistance. The reduction in total peripheral resistance during HOWI is most likely a result of the water-skin temperature gradient (Pendergast et al. 2015; Sackett et al. 2017, 2018) and attenuated sympathetic nerve activity during water immersion (Mano et al. 1985; Miwa et al. 1996). It is not clear why Dry + CO₂ did not appear to induce vasodilation (i.e., a reduction in total peripheral resistance). The effects of acute hypercapnia during dry conditions on vascular responses and sympathetic nerve activity (i.e., neurovascular transduction) are currently not known. The increase in stroke volume during HOWI and Dry + CO₂ was the primary cause for the rise in cardiac output in both conditions. While this typically occurs during HOWI due to the hydrostatic pressure induced central hypervolemia and elevated venous return (Arborelius et al. 1972), it appears that an increase in arterial CO₂ alone augments stroke volume and results in a modest increase in cardiac output. It is possible that increases in minute ventilation during Dry + CO₂ enhanced venous return and therefore stroke volume through the thoracic pump (Taylor and Groeller 2008). Moreover, despite previous findings (Sackett et al. 2018), we found a reduction in the alveolar ventilation to cardiac output ratio during HOWI, which might contribute to increases in PETCO₂ by impairing gas exchange (Derion et al. 1992; Moon et al. 2009). Previous work has demonstrated that minute ventilation is unchanged (Sackett et al. 2017, 2018) or reduced (Cherry et al. 2009) during water immersion. If we assume the typical rise in central blood volume during water immersion (Arborelius et al. 1972), these prior findings also suggest that a reduced alveolar ventilation to cardiac output ratio occurs

during HOWI. Along these lines, pulmonary blood flow is augmented during water immersion, which may contribute to increases in CO₂ pressure (Moon et al. 2009). Although unlikely, we cannot confirm that pulmonary blood flow was not elevated during Dry + CO₂.

Cerebral hemodynamics

Previous investigations indicate that middle cerebral artery blood velocity is augmented during HOWI (Carter et al. 2014). Middle cerebral artery blood velocity reflects total cerebral blood flow and is correlated with changes in PETCO₂ (Hida et al. 1996; Garbin et al. 1997; Barrett et al. 2001). Our data indicate that middle cerebral artery blood velocity is not altered during Dry + CO₂ and lower during Dry + CO₂ versus HOWI, which suggests that an increase in middle cerebral artery blood velocity during HOWI is due to the combined effects of water immersion (i.e., elevated PETCO₂, central hypervolemia, increased work of breathing, etc.). Carter et al. suggested that an increase in middle cerebral artery blood velocity during water immersion is due to increases in blood pressure (Carter et al. 2014). However, they used a colder water temperature than we did (~30°C vs. ~35°C), which would likely cause differences in total peripheral resistance between studies. Thermoneutral water immersion (~35°C) reduces muscle sympathetic activity in the tibial nerve (Mano et al. 1985; Miwa et al. 1996), which likely contributes to increases in systemic blood flow occurring secondary to reductions in peripheral vascular resistance (Arborelius et al. 1972). However, it is currently unknown how cerebral sympathetic and parasympathetic nerves (Edvinsson et al. 1993) contribute to cerebral blood flow during water immersion.

Rebreathing test

Similar to previous findings (Sackett et al. 2018), our data indicate that ventilatory sensitivity to CO₂ was augmented during HOWI and greater than during Dry + CO₂, since ventilatory sensitivity to CO₂ was not altered during Dry + CO₂. Thus, elevated PETCO₂ alone does not appear to contribute to the elevated ventilatory sensitivity to CO₂ during HOWI. The ventilatory threshold to CO₂ represents the minimum PETCO₂ that causes activation of the central chemoreceptors (Read 1967). To this end, a rightward shift in the ventilatory threshold to CO₂ indicates a higher PETCO₂ for the same minute ventilation and could contribute to increases in ventilatory sensitivity to CO₂, as previous work indicates (Sackett et al. 2018). However, we did not observe a rightward shift in the ventilatory threshold to CO₂ during either visit in the current investigation. The reason for these discrepant findings are not inherently clear. Previous findings indicate that the rate of rise of PETCO₂

during a rebreathing test is attenuated during water immersion (Chang and Lundgren 1995; Sackett *et al.* 2018) which is indicative of an increase in the CO₂ storage capacity of the body (Fowle and Campbell 1964). Chang and Lundgren suggested that an elevated tissue perfusion during HOWI contributes to a greater CO₂ redistribution to the tissues (i.e., muscle and fat) and may be related to the increase in PETCO₂ (Chang and Lundgren 1995). The present data also indicate that a slower increase in PETCO₂ over time occurs during HOWI. Thus, it appears that a slower increase in PETCO₂ over time might contribute to increases in ventilatory sensitivity to CO₂ during HOWI. It is currently unclear why PETCO₂ increases at a slower rate during HOWI.

We found that cerebrovascular reactivity to CO₂ was lower than baseline during both HOWI and Dry + CO₂ but it was not different between conditions. These data indicate that increases in PETCO₂, as a result of HOWI or breathing hypercapnic gas, blunts cerebrovascular reactivity to CO₂. Previous findings indicate that minor reductions in cerebral blood flow attenuate cerebrovascular reactivity to CO₂ and increase ventilatory sensitivity to CO₂, while further reductions in cerebral blood flow cause a greater decrease in cerebrovascular reactivity to CO₂ (Chapman *et al.* 1979). Our data indicate that reduced cerebrovascular reactivity to CO₂ during HOWI may increase cerebral perfusion and that this occurs at the same time that ventilatory sensitivity to CO₂ is elevated. It is unlikely, however, that these changes in cerebrovascular reactivity to CO₂ associated with acute hypercapnia are causally mediating changes in ventilatory sensitivity to CO₂ because decreases in cerebrovascular reactivity to CO₂ were not accompanied by increases in ventilatory sensitivity to CO₂ during Dry + CO₂. It should be noted that rebreathing tests are considered 'ventilatory independent', such that changes in ventilation during rebreathing do not appear to be dependent on changes in cerebrovascular reactivity to CO₂ (Ainslie and Duffin 2009). Although this effect is often negligible (Pandit *et al.* 2007), future investigations using steady state methods are required to elucidate causation between cerebrovascular reactivity to CO₂ and ventilatory sensitivity to CO₂, especially during HOWI.

Perspectives

PETCO₂ increases during water immersion despite augmented ventilatory sensitivity to CO₂ (Sackett *et al.* 2018). Moreover, an elevated PETCO₂ increases an underwater diver's risk for several hypercapnia related symptoms. These symptoms include breathlessness, headaches, and dizziness and enhance the likelihood of CO₂ toxicity and unexpected loss of consciousness while diving (Fothergill *et al.* 1998). Collectively, these symptoms

and conditions can lead to life-threatening situations in both recreational and working underwater divers. Lundgren demonstrated that the static lung load during HOWI mimics that of underwater diving at depth in the upright position while using an underwater breathing apparatus (Lundgren and Miller 1999). However, previous reports indicate dose dependent elevations in end tidal and/or arterial CO₂ pressures with depth and exercise (Lanphier and Bookspan 1999; Cherry *et al.* 2009). It is therefore possible that water immersion at depth or during exercise may further enhance end tidal and/or arterial CO₂ pressures and contribute to alterations in ventilatory sensitivity to CO₂ and cerebrovascular reactivity to CO₂. Furthermore, water immersion induces similar ventilatory and hemodynamic alterations that occur during microgravity, including an elevated PETCO₂ (Nagatomo *et al.* 2014; Watenpaugh 2016). To this end, immersion is often used as a model of microgravity (Watenpaugh 2016). CO₂ pressure is elevated during space flight as a result of increases in the ambient partial pressure of CO₂ in the spacecraft (Law *et al.* 2010). Consequently, flight crew members commonly report symptoms such as headaches and nausea, which are associated with hypercapnia (Law *et al.* 2010). Previous reports indicate that these symptoms may hinder the mission outcomes by contributing to fatigue, lethargy, and confusion (Carr 2006). Therefore, identifying the mechanisms that underpin the elevated CO₂ pressure and changes in ventilatory sensitivity and cerebrovascular reactivity are important to develop countermeasures to prevent increases in CO₂ pressure during water immersion and space flight. Current evidence indicates that respiratory muscle training normalizes ventilatory sensitivity to CO₂ during dry conditions in subjects with low or high baseline ventilatory sensitivity to CO₂ (Pendergast *et al.* 2006) and could therefore be an intervention to reduce the risk of CO₂ toxicity.

Considerations

Our investigation has several considerations worth noting. First, HOWI induced a relatively small increase in PETCO₂. This small increase in PETCO₂ during HOWI may not be considered clinically significant (Moloney *et al.* 2001). However, it was apparent that the increases in PETCO₂ were sensed by the chemoreceptors during Dry + CO₂, evidenced by the rise in minute ventilation. Along these lines, we used PETCO₂ as a marker of arterial CO₂ pressure since previous reports demonstrate that PETCO₂ and arterial CO₂ pressure are not significantly different during water immersion (Dunworth *et al.* 2017). However, PETCO₂ may underestimate arterial CO₂ pressure during instances of increased dead space (i.e., water

immersion) (Liu *et al.* 1995; Williams and Babb 1997). Thus, using PETCO₂ as a surrogate of arterial CO₂ pressure is a conservative approach, such that calculated differences in dead space ventilation and alveolar ventilation to cardiac output ratio would be exacerbated if we were to observe a higher value for arterial CO₂ pressure via direct measurements. It is also possible that increases in PETCO₂ during water immersion may be caused by other factors such as, alveolar ventilation to perfusion mismatching. Second, we recruited a convenience sample for the current investigation and therefore the subjects were not restricted to divers or nondivers. Regular underwater divers have attenuated ventilatory sensitivity to CO₂ compared to nondivers during dry conditions (Pendergast *et al.* 2006; Earing *et al.* 2014). To this end, several subjects were certified underwater divers ($n = 5$) but only a few reported to be regular divers ($n = 2$). Despite this, all subjects who were underwater divers had not been diving within 1 month of experimental testing. However, it is currently not known if divers and nondivers have similar ventilatory and cerebrovascular control during water immersion. Moreover, we did not determine if subjects had an undiagnosed patent foramen ovale. Subjects with a patent foramen ovale may have impaired gas exchange (Lovering *et al.* 2011), which could contribute to elevated PETCO₂ during water immersion. Third, subjects were seated upright throughout both experimental visits. While this was part of our experimental design in an attempt to eliminate any effects of posture, it should be noted that venous pooling may have occurred in the lower extremities during Dry + CO₂ but not during HOWI. Thus, we cannot be certain that venous pooling in the legs did not contribute to our findings. Fourth, we found that ventilatory sensitivity to CO₂ was greater during HOWI versus Dry + CO₂ at baseline. The reproducibility of ventilatory sensitivity to CO₂ has been shown to be good across a few hours and days (Scamman and Ghoneim 1983; Sullivan and Yu 1984). It is unclear why we found this baseline difference as we controlled for factors that might contribute to day to day variation in ventilatory sensitivity to CO₂ (i.e., fasting for 2 h, abstaining from exercise, alcohol, and caffeine for 12 h, and being euhydrated) and both visits were performed within one week. Finally, we used transcranial Doppler to measure middle cerebral artery blood velocity during our investigation. Although recent evidence indicates that middle cerebral artery blood velocity is indicative of total cerebral blood flow (Hida *et al.* 1996; Garbin *et al.* 1997; Barrett *et al.* 2001), it is not known if this is the case during water immersion. In this regard, we were unable to measure changes in cerebral vessel diameter, which may have occurred during the experimental visits due to hypercapnia and/or HOWI (Carter *et al.* 2014).

Conclusions

In sum, although minute ventilation is greater, ventilatory sensitivity to CO₂ is lower during dry conditions while breathing hypercapnic gas to match the elevated PETCO₂ that occurs during HOWI when compared to HOWI. Meanwhile, cerebral perfusion is lower, while cerebrovascular reactivity to CO₂ is not different during dry conditions while breathing a hypercapnic gas to match the elevated PETCO₂ that occurs during HOWI when compared to HOWI. Augmented ventilatory sensitivity to CO₂ and cerebral perfusion during HOWI appears to be a function of the integrative physiological changes that occur during water immersion and not elevated PETCO₂ (i.e., acute hypercapnia) alone. Therefore, it appears that small increases in PETCO₂ are not physiologically important with regards to modifying ventilatory sensitivity to CO₂ and cerebral perfusion during HOWI.

Acknowledgments

We thank the subjects for participating in our study. We also thank David R. Pendergast, EdD for his assistance with study design and data interpretation.

Conflict of Interest

There are no competing interests to report.

References

- Ainslie, P. N., and J. Duffin. 2009. Integration of cerebrovascular CO₂ reactivity and chemoreflex control of breathing: mechanisms of regulation, measurement, and interpretation. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 296:R1473–R1495.
- Ainslie, P. N., A. Barach, K. J. Cummings, C. Murrell, M. Hamlin, and J. Hellemans. 2007. Cardiorespiratory and cerebrovascular responses to acute poikilocapnic hypoxia following intermittent and continuous exposure to hypoxia in humans. *J. Appl. Physiol.* 102:1953–1961.
- Arborelius, M. Jr, U. I. Ballidin, B. Lilja, and C. E. Lundgren. 1972. Hemodynamic changes in man during immersion with the head above water. *Aerosp. Med.* 43:592–598.
- Armstrong, L. E., J. A. Soto, F. T. Jr Hacker, D. J. Casa, S. A. Kavouras, and C. M. Maresh. 1998. Urinary indices during dehydration, exercise, and rehydration. *Int. J. Sport Nutr.* 8:345–355.
- Barlow, H. B., and F. C. Macintosh. 1944. Shallow water black-out: report to the Sub-Committee on Underwater Physiology of the Royal Naval Personnel Research Committee. *Diving Hyperb. Med.* 43:109–112.
- Barrett, K. M., R. H. Ackerman, G. Gahn, J. M. Romero, and M. Candia. 2001. Basilar and middle cerebral artery reserve: a comparative study using transcranial Doppler and breath-holding techniques. *Stroke* 32:2793–2796.

- Berger, A. J., R. A. Mitchell, and J. W. Severinghaus. 1977. Regulation of respiration (third of three parts). *N. Engl. J. Med.* 297:194–201.
- Bonde-Petersen, F., L. Schultz-Pedersen, and N. Dragsted. 1992. Peripheral and central blood flow in man during cold, thermoneutral, and hot water immersion. *Aviat. Space Environ. Med.* 63:346–350.
- Carr, C. E. 2006. Impact of moderate elevations in CO₂ on astronauts during long-duration spaceflight on the International Space Station. Internal Rep.
- Carter, H. H., A. L. Spence, C. J. Pugh, P. Ainslie, L. H. Naylor, and D. J. Green. 2014. Cardiovascular responses to water immersion in humans: impact on cerebral perfusion. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 306:R636–R640.
- Casio. 2018. Keisan online calculator: vital capacity <http://keisan.n.casio.com/has10/SpecExec.cgi?path=02500000.Health%2F05000000.Health%20Check%2F10000500.Vital%20Capacity%2Fdefault.xml&charset=utf-8>.
- Chang, L. P., and C. E. Lundgren. 1995. CO₂ chemosensitivity during immersion in humans. *Chin. J. Physiol.* 38:7–12.
- Chapman, R. W., T. V. Santiago, and N. H. Edelman. 1979. Effects of graded reduction of brain blood flow on chemical control of breathing. *J. Appl. Physiol.* 47:1289–1294.
- Cherry, A. D., I. F. Forkner, H. J. Frederick, M. J. Natoli, E. A. Schinazi, J. P. Longphre, et al. 2009. Predictors of increased PaCO₂ during immersed prone exercise at 4.7 ATA. *J. Appl. Physiol.* 106:316–325.
- Craig, A. B. Jr, and M. Dvorak. 1966. Thermal regulation during water immersion. *J. Appl. Physiol.* 21:1577–1585.
- Derion, T., H. J. Guy, K. Tsukimoto, W. Schaffartzik, R. Prediletto, D. C. Poole, et al. 1992. Ventilation-perfusion relationships in the lung during head-out water immersion. *J. Appl. Physiol.* 72:64–72.
- Dunworth, S. A., M. J. Natoli, M. Cooter, A. D. Cherry, D. F. Peacher, J. F. Potter, et al. 2017. Hypercapnia in diving: a review of CO₂ retention in submersed exercise at depth. *Undersea Hyperb. Med.* 44:191–209.
- Earing, C. M., D. J. McKeon, and H. P. Kubis. 2014. Divers revisited: the ventilatory response to carbon dioxide in experienced scuba divers. *Respir. Med.* 108:758–765.
- Edvinsson, L., E. T. MacKenzie, and J. McCulloch. 1993. Cerebral blood flow and metabolism. Raven Press, New York.
- Fan, J. L., J. D. Cotter, R. A. Lucas, K. Thomas, L. Wilson, and P. N. Ainslie. 1985. Human cardiorespiratory and cerebrovascular function during severe passive hyperthermia: effects of mild hypohydration. *J. Appl. Physiol.* 105:2008.
- Farhi, L. E., L. E. Farhi, D. Linnarsson, and D. Linnarsson. 1977. Cardiopulmonary readjustments during graded immersion in water at 35°C. *Respir. Physiol.* 30:35–50.
- Fothergill, D. M., W. F. Taylor, and D. E. Hyde. 1998. Physiologic and perceptual responses to hypercarbia during warm- and cold-water immersion. *Undersea Hyperb. Med.* 25:1–12.
- Fowle, A. S., and E. J. Campbell. 1964. The immediate carbon dioxide storage capacity of man. *Clin. Sci.* 27:41–49.
- Garbin, L., F. Habetswallner, and A. Clivati. 1997. Vascular reactivity in middle cerebral artery and basilar artery by transcranial Doppler in normals subjects during hypoxia. *Ital. J. Neurol. Sci.* 18:135–137.
- Hida, W., Y. Kikuchi, S. Okabe, H. Miki, H. Kurosawa, and K. Shirato. 1996. CO₂ response for the brain stem artery blood flow velocity in man. *Respir. Physiol.* 104:71–75.
- Koehle, M. S., L. V. Giles, A. N. Curtis, M. L. Walsh, and M. D. White. 2009. Performance of a compact end-tidal forcing system. *Respir. Physiol. Neurobiol.* 167:155–161.
- Kumar, P., and N. R. Prabhakar. 2012. Peripheral chemoreceptors: function and plasticity of the carotid body. *Compr. Physiol.* 2:141–219.
- Lambertsen, C. J. 1977. Human tolerance to He, Ne, and N₂ at respiratory gas densities equivalent to He-O₂ breathing at depths to 1200, 2000, 3000, 4000, and 5000 feet of sea water. *Aviat. Space Environ. Med.* 48:843–855.
- Lanphier, E. H., and J. Bookspan. 1999. Carbon dioxide retention. Pp. 211–236 in J. N. Miller, C. E. Lundgren, eds. *The lung at depth*. Marcel Dekker, New York, NY.
- Law, J., S. Watkins, and D. Alexander. 2010. In-flight carbon dioxide exposures and related symptoms: association, susceptibility, and operational implications. NASA Johnson Space Center 1–21.
- Levitzky, M. G. 2013. *Pulmonary physiology*. McGraw-Hill Professional Publishing, New York, NY.
- Liu, Z., F. Vargas, D. Stansbury, S. A. Sasse, and R. W. Light. 1995. Comparison of the end-tidal arterial PCO₂ gradient during exercise in normal subjects and in patients with severe COPD. *Chest* 107:1218–1224.
- Lovering, A. T., M. K. Stickland, M. Amann, M. J. O'Brien, J. S. Hokanson, and M. W. Eldridge. 2011. Effect of a patent foramen ovale on pulmonary gas exchange efficiency at rest and during exercise. *J. Appl. Physiol.* 110:1354–1361.
- Lundgren, C. E., and J. N. Miller. 1999. Immersion effects. Pp. 91–128 in J. N. Miller and C. E. Lundgren, eds. *The lung at depth*. Marcel Dekker, New York, NY.
- MacKay, C. M., R. J. Skow, M. M. Tymko, L. M. Boulet, M. H. Davenport, C. D. Steinback, et al. 2016. Central respiratory chemosensitivity and cerebrovascular CO₂ reactivity: a rebreathing demonstration illustrating integrative human physiology. *Adv. Physiol. Educ.* 40:79–92.
- Mano, T., S. Iwase, Y. Yamazaki, and M. Saito. 1985. Sympathetic nervous adjustments in man to simulated weightlessness induced by water immersion. *J. UOEH* 7 (Suppl):215–227.
- Mark, C. I., M. Slessarev, S. Ito, J. Han, J. A. Fisher, and G. B. Pike. 2010. Precise control of end-tidal carbon dioxide and oxygen improves BOLD and ASL cerebrovascular reactivity measures. *Magn. Reson. Med.* 64:749–756.
- McSwain, S. D., D. S. Hamel, P. B. Smith, M. A. Gentile, S. R. Srinivasan, J. N. Meliones, et al. 2010. End-tidal and arterial carbon dioxide measurements correlate across all levels of physiologic dead space. *Respir. Care* 55:288–293.

- Minson, C. T., J. R. Halliwill, T. M. Young, and M. J. Joyner. 2000. Influence of the menstrual cycle on sympathetic activity, baroreflex sensitivity, and vascular transduction in young women. *Circulation* 101:862–868.
- Miwa, C., T. Mano, M. Saito, S. Iwase, T. Matsukawa, Y. Sugiyama, et al. 1996. Ageing reduces sympatho-suppressive response to head-out water immersion in humans. *Acta Physiol. Scand.* 158:15–20.
- Miyamoto, T., D. M. Bailey, H. Nakahara, S. Ueda, M. Inagaki, and S. Ogoh. 2014. Manipulation of central blood volume and implications for respiratory control function. *Am. J. Physiol. Heart Circ. Physiol.* 306:H1669–H1678.
- Moloney, E. D., J. L. Kiely, and W. T. McNicholas. 2001. Controlled oxygen therapy and carbon dioxide retention during exacerbations of chronic obstructive pulmonary disease. *Lancet* 357:526–528.
- Moon, R. E., A. D. Cherry, B. W. Stolp, and E. M. Camporesi. 2009. Pulmonary gas exchange in diving. *J. Appl. Physiol.* 106:668–677.
- Motulsky, H. J., and R. E. Brown. 2006. Detecting outliers when fitting data with nonlinear regression - a new method based on robust nonlinear regression and the false discovery rate. *BMC Bioinformatics* 7:123.
- Mummery, H. J., B. W. Stolp, G. deL Dear, P. O. Doar, M. J. Natoli, A. E. Boso, et al. 2003. Effects of age and exercise on physiological dead space during simulated dives at 2.8 ATA. *J. Appl. Physiol.* 94:507–517.
- Nagatomo, F., M. Kouzaki, and A. Ishihara. 2014. Effects of microgravity on blood flow in the upper and lower limbs. *Aerosp. Sci. Technol.* 34:20–23.
- Nattie, E., and A. Li. 2012. Central chemoreceptors: locations and functions. *Compr. Physiol.* 2:221–254.
- Ogoh, S., N. Hayashi, M. Inagaki, P. N. Ainslie, and T. Miyamoto. 2008. Interaction between the ventilatory and cerebrovascular responses to hypo- and hypercapnia at rest and during exercise. *J. Physiol.* 586:4327–4338.
- Pandit, J. J., R. M. Mohan, N. D. Paterson, and M. J. Poulin. 2007. Cerebral blood flow sensitivities to CO₂ measured with steady-state and modified rebreathing methods. *Respir. Physiol. Neurobiol.* 159:34–44.
- Pendergast, D. R., P. Lindholm, J. Wylegala, D. Warkander, and C. E. Lundgren. 2006. Effects of respiratory muscle training on respiratory CO₂ sensitivity in SCUBA divers. *Undersea Hyperb. Med.* 33:447–453.
- Pendergast, D. R., R. E. Moon, J. J. Krasney, H. E. Held, and P. Zamparo. 2015. Human physiology in an aquatic environment. *Compr. Physiol.* 5:1705–1750.
- Prabhakar, N. R., and Y. J. Peng. 2004. Peripheral chemoreceptors in health and disease. *J. Appl. Physiol.* 96:359–366.
- Ray, A. D., D. R. Pendergast, and C. E. Lundgren. 2010. Respiratory muscle training reduces the work of breathing at depth. *Eur. J. Appl. Physiol.* 108:811–820.
- Read, D. J. 1967. A clinical method for assessing the ventilatory response to carbon dioxide. *Australas Ann. Med.* 16:20–32.
- Rebuck, A. S. 1976. Measurement of ventilatory response to CO₂ by rebreathing. *Chest* 70:118–121.
- Sackett, J. R., Z. J. Schlader, S. Sarker, C. L. Chapman, and B. D. Johnson. 2017. Peripheral chemosensitivity is not blunted during 2 h of thermoneutral head out water immersion in healthy men and women. *Physiol. Rep.* 5:1–13.
- Sackett, J. R., Z. J. Schlader, M. C. O’Leary, C. L. Chapman, and B. D. Johnson. 2018. Central chemosensitivity is augmented during 2 h of thermoneutral head-out water immersion in healthy men and women. *Exp. Physiol.* 103:714–727.
- Salzano, J. V., E. M. Camporesi, B. W. Stolp, and R. E. Moon. 1984. Physiological-responses to exercise at 47 and 66 ATA. *J. Appl. Physiol.* 57:1055–1068.
- Scamman, F. L., and M. M. Ghoneim. 1983. How often may a Read rebreathing test be repeated? *Clin. Physiol.* 3:487–490.
- Slessarev, M., J. Han, A. Mardimae, E. Prisman, D. Preiss, G. Volgyesi, et al. 2007. Prospective targeting and control of end-tidal CO₂ and O₂ concentrations. *J. Physiol.* 581:1207–1219.
- Sramek, P., M. Simeckova, L. Jansky, J. Savlikova, and S. Vybiral. 2000. Human physiological responses to immersion into water of different temperatures. *Eur. J. Appl. Physiol.* 81:436–442.
- Sullivan, T. Y., and P. L. Yu. 1984. Reproducibility of CO₂ response curves with ten minutes separating each rebreathing test. *Am. Rev. Respir. Dis.* 129:23–26.
- Taylor, N. A., and H. Groeller, eds. 2008. *Physiological bases of human performance during work and exercise.* Churchill Livingstone Elsevier, New York, NY.
- Watenpaugh, D. E. 2016. Analogs of microgravity: head-down tilt and water immersion. *J. Appl. Physiol.* 120:904–914.
- Watenpaugh, D. E., B. Pump, P. Bie, and P. Norsk. 2000. Does gender influence human cardiovascular and renal responses to water immersion? *J. Appl. Physiol.* 89: 621–628.
- Wesseling, K. H., J. R. Jansen, J. J. Settels, and J. J. Schreuder. 1993. Computation of aortic flow from pressure in humans using a nonlinear, three-element model. *J. Appl. Physiol.* 74:2566–2573.
- West, J. B. 2012. *Respiratory physiology: the essentials.* Wolters Kluwer Health/Lippincott Williams & Wilkins, Philadelphia, p. viii, 200 p.
- Williams, J. S., and T. G. Babb. 1997. Differences between estimates and measured PaCO₂ during rest and exercise in older subjects. *J. Appl. Physiol.* 83:312–316.
- Xie, A., J. B. Skatrud, R. Khayat, J. A. Dempsey, B. Morgan, and D. Russell. 2005. Cerebrovascular response to carbon dioxide in patients with congestive heart failure. *Am. J. Respir. Crit. Care Med.* 172:371–378.