

TREM-1 Modulation Strategies for Sepsis

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Siskind S, Brenner M and Wang P (2022) TREM-1 Modulation Strategies for Sepsis. Front. Immunol. 13:907387. doi: 10.3389/fimmu.2022.907387 The triggering receptor expressed on myeloid cells-1 (TREM-1) is a pattern recognition receptor, which can be upregulated in inflammatory diseases as an amplifier of immune responses. Once activated, TREM-1 induces the production and release of proinflammatory cytokines and chemokines, in addition to increasing its own expression and circulating levels of the cleaved soluble extracellular portion of TREM-1 (sTREM-1). This amplification of the inflammatory response by TREM-1 has now been considered as a critical contributor to the dysregulated immune responses in sepsis. Studies have shown that in septic patients there is an elevated expression of TREM-1 on immune cells and increased circulating levels of sTREM-1, associated with increased mortality. As a result, a considerable effort has been made towards identifying endogenous ligands of TREM-1 and developing TREM-1 inhibitory peptides to attenuate the exacerbated inflammatory response in sepsis. TREM-1 modulation has proven a promising strategy for the development of therapeutic agents to treat sepsis. Therefore, this review encompasses the ligands investigated as activators of TREM-1 thus far and highlights the development and efficacy of novel inhibitors for the treatment of sepsis and septic shock.

Keywords: sepsis, inflammation, TREM-1, shock, DAMP

INTRODUCTION

Sepsis is a complex disorder, defined by a dysregulated host response to infection leading to organ dysfunction (1, 2). The pro-inflammatory response to invading pathogens is initiated by pattern recognitions receptors (PRRs) located on the surface and intracellularly of immune and non-immune cells, that recognize pathogen-associated molecular patterns (PAMPs), molecules released from pathogens, and damage-associated molecular patterns (DAMPs), molecules released from damaged cells (3, 4). Activation of PRRs on innate immune cells initiates microbicidal and pro-inflammatory responses to contain and eliminate the invading pathogens and activates the adaptive immune response, particularly T lymphocytes (3). Toll-like receptors (TLRs) are a well-studied family of PRRs for their role in innate immunity that amplify the inflammatory response synergistically with triggering receptor expressed on myeloid cells-1 (TREM-1), a more recently characterized PRR (5–8).

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TREM-1 was first identified on circulating neutrophils and monocytes in response to lipopolysaccharide (LPS), a glycan mostly present on the surface of Gram-negative bacteria that strongly activates the host immune response (9-11). TREM-1 is part of a family of TREM receptors that also includes TREM-2, TREM-3, and TREM-like transcript-1 and 2 (TLT-1, TLT-2) (12). TREM-3 is only expressed in mice, and like TREM-1, is upregulated in response to LPS. TREM-2, on the other hand, is downregulated in response to LPS and, upon activation, attenuates the inflammatory response (12, 13). Activation of TREM-1 in neutrophils and monocytes triggers the release of pro-inflammatory cytokines and chemokines, in addition to upregulating the gene expression of TREM-1 and surface expression of cell activation markers (9). This amplification of the inflammatory response by TREM-1 has gained interest as a critical contributor to the dysregulated immune response in sepsis (4). Patients admitted to the ICU with septic shock have higher surface expression of TREM-1 on monocytes and elevated circulating levels of the cleaved soluble extracellular portion of TREM-1 (sTREM-1) (14, 15). Elevated levels of circulating sTREM-1 have also been associated with increased mortality in patients with septic shock (16, 17). Additionally, mice genetically deficient in TREM-1 had less severe disease in response to multiple pathogens without affecting their ability to clear the infection, resulting in similar pathogen loads (18). The inhibition of TREM-1 remains a promising target for development of therapeutic agents to treat sepsis. This review aims to give an overview of the role of TREM-1 and its endogenous ligands in sepsis, and the development of novel inhibitors of this pathway and their efficacy for the treatment of sepsis.

STRUCTURE AND SIGNALING PATHWAY

TREM-1 is a member of the immunoglobin superfamily, a group of cell surface receptors with related extracellular Ig-like domains (19, 20). In addition to the extracellular Ig domain, it consists of a transmembrane region with a conserved lysine residue, and a short cytoplasmic domain that does not contain a signaling motif (21, 22). Propagation of signaling is instead dependent on association with the immunoreceptor adaptor protein DNAX activation protein 12 (DAP12). TREM-1 contains a positively charged transmembrane lysine residue that associates with a negatively charged aspartate residue of DAP12 (21-24). Upon receptor activation, the immunoreceptor tyrosine-based activation motif (ITAM) of DAP12 is phosphorylated, signaling the recruitment and activation of spleen tyrosine kinase (Syk), a nonreceptor tyrosine kinase (20). Syk activates multiple downstream signal transduction pathways including the PI3K/Akt pathway, the Ras/ERK/MAPK pathway, NF-ĸB signaling, and phospholipase C phosphorylation, leading to increased intracellular calcium and proinflammatory cytokine secretion (19-22, 25).

TREM-1 activation independently triggers downstream inflammatory cascades that can synergize with TLR signaling pathways (22, 25). TLR4 activation upregulates the expression of

TREM-1, and the concomitant activation of TREM-1 and TLR4 leads to a synergistic increase in proinflammatory cytokine and chemokine release by 25-fold compared with that of TLR4 activation alone (8, 26–28). One proposed mechanism for their synergistic proinflammatory response is that TREM-1 increases the availability of TLR4 downstream signaling molecules such as MyD88, CD14, NF- κ B, and I κ B α (26). When TREM-1 s blocked, RAW cells stimulated with LPS maintain TLR4 expression but have decreased genetic expression of these signaling molecules as well as inflammatory cytokines (21, 26, 29). Understanding the structure and signaling pathway of TREM-1 remains an important focus of research to better elucidate its role in sepsis and to develop novel therapeutic targets.

TREM-1 LIGANDS

The identification of TREM-1's endogenous ligands is crucial for studying the role of TREM-1 in the pathogenesis of sepsis. Moreover, knowing TREM-1's endogenous ligands and their structure offers critical insights to develop TREM-1-targeting pharmacological strategies to reduce sepsis hyperinflammation, such as the inhibitory peptides discussed below. Although TREM-1 was characterized in 2000, it took 14 years until the first ligand was identified. Since then, multiple ligands have been implicated as activators of TREM-1, furthering our knowledge on this receptor's role in innate immunity, sepsis, and noninfectious inflammatory diseases.

Peptidoglycan Receptor Protein 1 (PGLYRP1)

PGLYRP1 (also named Tag7) was one of the first identified ligands of TREM-1 (30). PGLYRP1 is an antimicrobial protein that is secreted from polymorphonuclear leukocyte granules in response to infection (5, 30, 31). It subsequently binds to peptidoglycan and LPS, essential components of the bacterial membrane, where it induces lethal membrane depolarization and oxidative stress to the bacteria (32–34). After it was discovered that an endogenous, unknown ligand of TREM-1 exists on bacterially activated peritoneal neutrophils, PGLYRP1 was investigated as a potential ligand (30, 35). Neutrophils were stimulated with peptidoglycan and cross-linked to sTREM-1 and, using mass spectrometry, identified that the resulting complexes contained PGLYRP1 peptides (30).

PGLYRP1 as a ligand for TREM-1 was further studied using affinity chromatography, which demonstrated conclusively that immobilized soluble TREM-1 binds PGLYRP1 (36). It has also been shown that PGLYRP1 is an activating ligand of TREM-1 on monocytes and induces the development of cytotoxic lymphocyte subpopulations in peripheral blood mononuclear cells (PBMCs) (37). In the presence of LP17, an inhibitory peptide of TREM-1 discussed further below, monocytes treated with PGLYRP1 failed to induce cytotoxic lymphocyte transformation and had reduced secretion of IL-2, thus providing further evidence that PGLYRP1 is an endogenous ligand for TREM-1 (36, 37). Interestingly, soluble PGLYRP1 alone does not activate TREM-1. Only when complexed with peptidoglycan, anchored to HEK293 cell surface, or bound to a plate, was PGLYRP1 able to elicit an inflammatory response *via* TREM-1 (30). This should be kept in mind for future research on PGLYRP1/TREM-1 interaction, as activation may only be reproducible when TREM-1 is bound to a cell surface.

Extracellular Cold Inducible RNA Binding Protein (eCIRP)

eCIRP is a DAMP that has recently been identified as an endogenous ligand of TREM-1. It was originally discovered in the blood of critically ill septic and trauma-hemorrhage surgical patients. CIRP is a ubiquitously expressed nuclear protein which has been mostly studied in macrophages, lymphocytes, and neutrophils, which release it into circulation during periods of hypoxic and mild hypothermic stress, such as in sepsis (38, 39). Mice injected with recombinant murine CIRP (rmCIRP) developed a sepsis-like acute lung injury with vascular endothelial cell damage, leukocyte infiltration and increased pro-inflammatory cytokine production (40). Using surface plasmon resonance (SPR), it was determined that rmCIRP binds to recombinant murine TREM-1 (rmTREM-1) with strong affinity, with a K_D of 11.7×10^{-8} M (41). Binding was further demonstrated between rmCIRP and surface TREM-1 on murine RAW264.7 and peritoneal macrophages using fluorescence resonance energy transfer (FRET) analysis. rmCIRP-injected mice had attenuated systemic and pulmonary inflammatory response after treatment with the TREM-1 inhibitor LP17. Additionally, mice deficient in TREM-1 had reduced serum IL-6 and IL1- β after rmCIRP injection, indicating eCIRP acts an endogenous ligand of TREM-1 (41).

High Mobility Group Box 1 (HMGB1)

HMGB1 has also been investigated as an endogenous ligand to TREM-1 (42). HMGB1 was originally identified as a nuclear DNA-binding protein that functions as a cofactor in transcription regulation. It was later found that it can be released from a variety of cells and function as a DAMP, signaling through activation of TLRs and receptor for advanced glycation end products (RAGE) to induce a proinflammatory response (43-45). More recently, using a murine model of hepatocellular carcinoma, HMGB1 released from necrotic hepatocytes was found to bind TREM-1 using immunoblotting and SPR, with a binding K_D of 35.4 x 10⁻⁶ M (42, 46). HMGB1 has also been shown to induce inflammatory responses through TREM-1 activation in THP-1 human monocytic cells, a human monocyte cell line, and upregulate TREM-1 expression on macrophages (47, 48). However, recombinant HMGB1 alone was insufficient to induce TREM-1-regulated M1 polarization, suggesting it may need coactivating molecules to fully trigger TREM-1, or that recombinant HMGB1 is different from endogenous HMGB1 (33, 49).

Heat Shock Protein 70 kDa (Hsp70)

Along with HMGB1, the 70 kDa heat shock protein (Hsp70) was found to be released from necrotic cells and augment the

proinflammatory response through TREM-1 activation (42, 47). Under normal conditions, HSP70 is a molecular chaperone that participates in the maintenance of protein homeostasis through folding and remodeling processes (50). Normally expressed at low levels, Hsp70 becomes upregulated in pro-inflammatory states like bacterial endotoxemia (47). In addition to Hsp70's primary role of protein folding stabilization, there has been growing interest in its function as a pro-inflammatory mediator after it was found to upregulate TNF- α , IL-1 β , and IL-6 in human monocytes (51). Hsp70 was then investigated as a novel ligand for TREM-1 using LPS- and necrotic cell lysate-stimulated THP-1 cells. These cells, when treated with an anti-HSP70 antibody, had a reduction in expression of TNF- α , IL-6, and IL-8 that was further reduced by the addition of an inhibitory recombinant TREM-1 fusion chimera (47). This data suggests that Hsp70 is released from necrotic cells to aid pro-inflammatory responses in monocytes through TREM-1 activation of the cytokine expression cascade (47). Hsp70 as a ligand of TREM-1 was further studied using affinity chromatography and demonstrated to bind to sTREM-1 immobilized on a CNBr-Sepharose column as well as to TREM-1 on the monocyte surface (52). Interestingly, however, it was found that Hsp70 released from necrotic Kupffer cells along with HMGB1 in a murine model of hepatocellular carcinoma, does not directly bind to TREM-1, suggesting that HMGB1 may have a stronger affinity and compete for binding on TREM-1 (42).

Extracellular Actin

Actin is one of the most abundant proteins within cells, functioning in the form of filaments that polymerize to aid in cell morphology and motility (53). When cells undergo apoptosis and necrosis, such as during sepsis, actin is released and has deleterious effects once extracellular (53). After it was determined that an unknown ligand expressed on platelets activates TREM-1 in sepsis, further investigation using gel analysis of platelet total protein and rTREM-1 suggested the ligand to be actin (54, 55). Confocal microscopy was used to confirm the co-localization of TREM-1 and actin on RAW267.7 cells treated with LPS-stimulated platelets and LPS with recombinant actin. This co-localization was also demonstrated in vivo using a CLP model of polymicrobial sepsis. Actin and TREM-1 had increased expression and co-localized in the lungs of septic mice (55). Additionally, actin dose-dependently enhanced LPS-stimulated release of TNF-α from RAW267.7 cells and peritoneal macrophages (55). This response was blunted in cells treated with the TREM-1 inhibitor LP17 as well as in peritoneal macrophages isolated from TREM-1 knockout mice, strongly suggesting that actin activates inflammatory cells via TREM-1 (55).

IMMUNE RESPONSES IN TREM-1 KNOCKOUT MICE

Mice genetically deficient in TREM-1 are another valuable tool to help uncover the potential benefits of modulating TREM-1 for

the treatment of sepsis. To study this, multiple knockout models with different genetic modifications have been developed: TREM-1 knockout, endothelial cell-specific TREM-1 (endoTREM-1) knockout, TREM-1/3 double knockout, and TREM-like transcript-1 (TLT-1) knockout. To determine the effects of TREM-1 deficiency on the immune response to infection, TREM-1-/- mice were infected with Leishmania major (18). Compared to WT mice, TREM-1^{-/-} mice had reduced infiltration of neutrophils and decreased lesion size around the site of bacterial inoculation (18). TREM-1-/- mice also had reduced morbidity after infection with influenza A virus, measured by body weight and temperature, and decreased IL-6 in bronchoalveolar lavage (18). Importantly, while infectionrelated pathologies were improved, TREM-1^{-/-} mice had equivalent pathogen clearance of L. major, influenza virus, and Legionella pneumophila as WT mice, indicating these mice do not develop disseminated infection like TREM-1/3^{-/-} (18). Additionally, after LPS-induced septic shock, TREM-1-/- mice had decreased neutrophil extracellular trap (NET) release in the serum and lungs, which have pro-inflammatory functions that contribute to the progression of septic shock (56). Aortas and mesenteric arteries isolated from TREM-1^{-/-} mice were protected from in vitro NET-induced vascular dysfunction (possibly due to their inability to respond to the eCIRP present in NETs), maintaining normal contraction and relaxation (56-58). The involvement of TREM-1 in sepsis induced vascular dysfunction was further proven using endoTREM-1^{-/-} mice (57). After cecal ligation and puncture (CLP)-induced sepsis, endoTREM-1^{-/-} mice had restored vasorelaxation in addition to reduced serum VCAM-1 and IL-6, and prolonged 7-day survival (57).

In mice, the gene for TREM-3 is adjacent to TREM-1 and likely occurred from a duplication event, demonstrated by their high homogeneity (59). Additionally, the two receptors are both amplifiers of the immune response, and likely work synergistically (59-61). TREM-3 in humans, however, is a pseudogene and has no functional overlap with TREM-3 in mice (61). Therefore, it has been suggested that TREM-1/3^{-/-} mice might better reflect TREM-1 deficiency in humans than mice deficient in TREM-1 alone (61-63). Interestingly, although TREM-1^{-/-} mice were protected after exposure to infectious agents, TREM-1/3^{-/-} mice had worse outcomes after infection. TREM-1/3^{-/-} mice had increased mortality and bacterial dissemination after infection with Streptococcus pneumoniae and Klebsiella pneumoniae, despite infected primary TREM-1/ 3^{-1-} macrophages having decreased cytokine release (64, 65). The relevance of murine TREM-3 to human TREM-1 function should continue to be explored to better characterize which murine models best emulate human disease.

TLT-1 belongs to the TREM family, and its gene resides in the human TREM gene cluster, along with TREM-1, -2, -3, and TLT-2 (66). It is specific to platelets and megakaryocytes and, upon platelet activation, translocates to the cell surface where it plays a role in hemostasis/thrombosis (66). Additionally, a soluble fragment of TLT-1 is present in the circulation and is believed to function as an endogenous TREM-1 inhibitor (67, 68). TLT-1 knockout mice (treml-1^{-/-}) had increased gene and protein

expression of inflammatory cytokines in the lungs and plasma after CLP induces sepsis (68). Additionally, septic treml-1^{-/-} mice had increased mortality, indicating a protective role of TLT-1 in polymicrobial sepsis, likely by inhibiting TREM-1 (68).

TREM-1 BLOCKADE AS A THERAPEUTIC APPROACH

Studies using TREM-1 knockout mice have demonstrated that this receptor plays a critical role in the progression of sepsis. Armed with this knowledge, multiple receptor antagonists have been created, predominately based on the structure of TREM-1 and its ligands. Modulation of TREM-1 activation with many of these inhibitors has been demonstrated as an effective approach for attenuating sepsis severity (**Table 1**).

Soluble TREM-1 (sTREM-1)

The sTREM-1 molecule is the cleaved extracellular domain of TREM-1 that was found to be increased in the blood during sepsis (69, 73, 84, 85). In addition to being studied as a prognostic indicator of sepsis severity, it has also been proposed to act as a decoy receptor for TREM-1 by binding its ligands and reducing its activation and the subsequent pro-inflammatory cytokine release (73, 86–89) (**Figure 1**). To test this hypothesis, the gene for recombinant porcine sTREM-1 was transfected into *Escherichia coli* for cloning and expression (69). After purification, it was administered to LPS-stimulated primary porcine alveolar macrophages. Macrophages treated with sTREM-1 had reduced mRNA expression of type-I and type-II inflammatory cytokines, including TNF- α , IL-1 β , IL-2, IL-4, IL-8, IL-10, IL-12, IL-16, and IL-18 (69).

A similar method of genetic cloning was used to express and purify murine sTREM-1, which was named recombinant extracellular domain of TREM-1 (rTREM-1) (90). Mice were then infected with Streptococcus suis, a bacterium that can rapidly cause streptococcal toxic-shock-like syndrome (STSLS), followed by treatment with rTREM-1. Conversely to the in vitro study using porcine sTREM-1, mice inoculated with S. suis and treated with rTREM-1 had worse outcomes. rTREM-1 treatment was associated with increased 7-day mortality, and elevated IL1- β , TNF- α , KC, and bacterial load in both blood and peritoneal fluid (90). Interestingly, however, when rTREM-1 treatment is combined with antibiotics, mice infected with S. suis had better outcomes than with antibiotics alone (70). Mice that received combination treatment of rTREM-1 and ampicillin had a higher 7-day survival, lower serum levels of IL1- β and TNF- α , and attenuated acute lung injury compared to mice that received PBS vehicle, just ampicillin, or rTREM-1 alone (70). These data suggest that, while TREM-1 agonism may help clear some bacterial infections, it is also a key contributor to the detrimental inflammatory processes that culminate in acute lung injury. Many of the following TREM-1 inhibitory peptides developed as a treatment modality for sepsis are based on amino acid (aa) sequences found in sTREM-1, further

TABLE 1 | TREM-1 pathway inhibitory strategies.

Treatment	Sepsis Model	Cell/Animal	Effect	Ref.
sTREM-1	LPS	Macrophages	↓ mRNA of inflammatory cytokines	(69)
	S. suis	Mice	\downarrow serum TNF- α , IL-1 β , \downarrow acute lung injury, \uparrow survival (when given with antibiotics)	(70)
	intraperitoneal		, , ,, ,, ,, ,, ,, ,, ,, ,, ,, ,, ,, ,,	```
TREM-1 Fc	LPS	Monocytes	\downarrow release TNF- α , IL-1 β	(8)
fusion protein				
	P. aeruginosa	Macrophages	↓ release TNF-α, IL-1β, MCP-1	(71)
	S. pyogenes	Neutrophils	↓ release IL-6, TNF- α	(72)
	LPS intraperitoneal	Mice	\downarrow serum TNF- α , IL-1 β , \downarrow recruitment of peritoneal macrophages and neutrophils, \uparrow survival	(8)
	CLP	Mice	↑ survival	(8)
	E. coli	Mice	↑ survival	(8)
	intraperitoneal			
	S. pyogenes	Mice	↓ serum IL-6, TNF-α, ↑ survival	(72)
	intravenous			
	P. aeruginosa	Mice	↓ serum IL-1β, TNF-α, MCP-1, ↑ survival	(71)
	intraperitoneal			
LP17	LPS	Monocytes	\downarrow release TNF- α , IL-1 β	(73)
	E. coli	Neonatal leukocytes	\downarrow release TNF- α , IL-6, IL-8	(74)
	LPS intraperitoneal	Mice	↑ survival	(35)
	CLP	Mice	↓ serum TNF-α, IL-1β, ↑ survival	(73)
	S. pyogenes	Mice	↑ survival	(72)
	intravenous			
	LPS intraperitoneal	Rats	↑ hemodynamics, ↓ serum TNF-α, IL-1β	(35)
	CLP	Rats	↑ hemodynamics, ↓ TNF-α, IL-1β, IL-6, ↑ survival	(35,
				75)
	P. aeruginosa	Rats	↓ serum and broncoalveolar lavage TNF- α , IL-1 β , IL-6, lactic acidosis, hypoxia, ↑ hemodynamics, ↑	(76)
	intratracheal		survival	
GF9	LPS	Macrophages	↓ release TNF-α, IL-1β, IL-6	(77)
	LPS intraperitoneal	Mice	↓ serum TNF-α, IL-1β, IL-6, ↑ survival	(77)
SLC-TREM-1	LPS	Endothelium	\downarrow TREM-1 expression, \downarrow MCP-1, IL-8 release	(78)
	CLP	Mice	↑ survival	(78)
M3	LPS intraperitoneal	Mice	↓ serum TNF-α, IL-6, ↑ survival	(41)
	CLP	Mice	↓ serum AST, ALT, TNF-α, IL-6, ↓ acute lung injury, ↑ survival	(41)
	Cecal Slurry	Neonatal Mice	↓ serum TNF-α, IL-1β, IL-6, INFγ, ↓ cardiac and pulmonary IL-1β, IL-6, ↓ cardiac dysfunction, ↑	(79)
			survival	
N1	LPS	Mononuclear Cells/	\downarrow mRNA TNF-α, IFNγ, IL-1β, and IL-6	(80)
		Monocytes		
	LPS intrabronchial	Mice	↓ serum INFγ, IL-4, ↓ lung damage	(80)
LR17/LR12	LPS	Granulocyte	\downarrow mRNA and protein TNF- α , IL-8, IL-10, \downarrow ROS production	(23)
	LPS	Monocytes	\downarrow release TNF- α	(23)
	LPS intraperitoneal	Mice	↓ serum TNF-α, IL-6, IL-10, ↑ survival	(23)
	CLP	Mice	↓ cytokines in serum, peritoneal and bronchoalveolar fluid, liver, lung, ↓ acute lung injury, ↓ bacteria	(23,
			in spleen, blood, ↓ TREM-1 expression, ↑ survival	57)
	Cecal Slurry	Pigs	↑ cardiac function, \downarrow coagulopathy, \downarrow failure of lungs, liver, kidney, ↑survival	(68)
	LPS intravenous	Monkeys	↑ hemodynamics, ↓ serum cytokines	(81)
Nangibotide	Sepsis	Humans	\downarrow systemic IL-6, \downarrow SOFA scores, minimal side effects	(82,
				83)

↓- decreased; ↑- increased.

demonstrating that sTREM-1 is effective as an anti-inflammatory mediator (88).

TREM-1 Fc Fusion Protein

TREM-1/Fc, a fusion protein consisting of the extracellular domain of mouse TREM-1 and the Fc portion of human IgG1, was developed as a TREM-1 decoy receptor that can also promote the clearance of TREM-1 ligands from the circulation *via* Fc-receptor mediated endocytosis (8) (**Figure 1**). Mice treated with TREM-1/Fc prior to LPS-induced endotoxemia had decreased serum TNF- α and IL-1 β and recruitment of peritoneal neutrophils and macrophages (8). Additionally, mice treated with TREM-1/Fc had improved 7-day survival in LPS,

CLP, and *Escherichia coli*-induced sepsis (8). These findings were reproduced in another study using intraperitoneal administration of *Pseudomonas aeruginosa* to induce sepsis (71). Treatment with TREM-1/Fc in these mice led to improved survival and a significant reduction in serum pro-inflammatory cytokines, including TNF- α , IL-1 β , INF- γ , and MCP-1 (71). Additionally, TREM-1/Fc provided protection in mice against sepsis after intravenous (i.v.) injection of *Streptococcus pyogenes* (72). Mice treated 2 hours before and 2 hours after inoculation of *S. pyogenes* had prolonged survival and decreased levels of IL-6 and TNF- α in the serum (72). In addition to its TREM-1 inhibitory effects, it is possible that the TREM-1/Fc fusion protein also enhances the phagocytic



interaction of TREM-1 with its signaling partner DAP12. (2) LR12/nangibotide, LP17, and TREM-1/Fc fusion protein act as decoy receptors and compete for binding with naturally occurring activating ligands. (3) Together with M3 and N1, LP17 also binds to TREM-1 and competitively inhibits ligand binding. M3 competitively inhibits TREM-1 binding to and activation by extracellular cold inducible RNA binding protein, while N1 inhibits TREM-1 activation by PGLYRP1 and HSP70. (4) soluble TREM-1 is generated from proteolytic cleavage of membrane bound TREM-1 by matrix metalloproteinases. Circulating soluble TREM-1 competitively binds TREM-1's ligands and prevents further activation. All inhibitors prevent the downstream signaling cascade that upregulates the translation of inflammatory cytokines and TREM-1 receptor. *Image created with BioRender*.

clearance of pathogens by bridging peptidoglycan-bound PGLYRP1 on the surface of bacteria with Fc-receptors on macrophages and neutrophils.

LP17

LP17 was one of the first peptides developed to inhibit TREM-1 (73). It has a 17-aa sequence (LQVTDSGLYRCVIYHPP) derived from on a highly conserved extracellular domain of TREM-1 in both mice and humans, and is intended to operate through a mechanism analogous to that of decoy receptors (35, 73, 76). To demonstrate its intended function, flow cytometry was used to show LP17 inhibits the binding of a fluorophore conjugatedmTREM-1/IgG1 antibody to TREM-1 on murine peritoneal exudate cells (35). There is evidence that it functions both as a direct competitive inhibitor by binding to the receptor and thus preventing its activation by the ligand, and as a decoy receptor by binding TREM-1's ligands before they are able to activate the TREM-1 receptor (4, 68) (Figure 1). Efficacy was initially demonstrated by reducing TNF- α and IL-1 β release from LP17-treated monocytes after stimulation with LPS in a dose dependent manner (73). LP17 was further tested in vivo in mice subjected to either endotoxemia or CLP-induced sepsis (73). Mice treated with LP17 had decreased levels of serum TNF-a and IL-1 β and improved survival after LPS injection and CLP (73). LP17 improved survival in endotoxemic mice when given 1 hour before or 4 hours after LPS injection (35, 73). Additionally, LP17 has been shown to improve survival in mice after i.v. inoculation with *Streptococcus pyogenes* (72).

LP17 has also been shown to be efficacious in treating septic rats. Rats treated with LP17 1 hour after LPS injection had attenuated sepsis severity, measured by improved hemodynamic parameters including mean arterial pressure, aortic blood flow, mesenteric blood flow, pH, and serum lactate (35). Additionally, these rats had lower concentrations of serum TNF- α , IL-1 β , and nitrates/nitrites (35). Rats treated with LP17 after CLP also had improved hemodynamic parameters, serum TNF- α , IL-1 β , IL-6 and nitrates/nitrites, and survival at 48 hours and 7 days (35, 75). LP17 has also been shown to be an effective treatment for sepsis secondary to *Pseudomonas aeruginosa* pneumonia in rats, resulting in with improved hemodynamic status, attenuated lactic acidosis and hypoxemia, reduced serum TNF- α , IL-1 β , and IL-6, and improved 7-day survival of the septic rats (76).

In addition to the studies in adult rodents, LP17 has been considered as a potential therapeutic agent for neonatal sepsis (74). In an *in vitro* study, leukocytes isolated from umbilical cord blood from full term human neonates and treated with LP17 had

decreased production of TNF- α , IL-6, and IL-8 after exposure to *Escherichia coli* (74).

М3

M3 is another inhibitory peptide of TREM-1 that was developed to specifically inhibit activation by its novel ligand, eCIRP (41). Its 7-aa sequence (RGFFRGG) was designed based on an area of homology between PGLYRP1 and CIRP (30, 41). Using a FRET assay, M3 was shown to dramatically abrogate CIRP's binding to TREM-1 in both murine peritoneal macrophages and RAW264.7 cells (41) (**Figure 1**). To test this peptide *in vivo*, mice were subjected to LPS-endotoxemia and those treated with M3 had decreased levels of serum TNF- α and IL-6, and improved 7-day survival (41). M3 was also protective in mice with CLP-sepsis, measured by decreased levels of serum AST, ALT, TNF- α and IL-6, and attenuated severity of sepsis-associated acute lung injury (41, 91). Mice treated with intraperitoneal (i.p.) M3 at the time of CLP or 90 minutes later also showed a 10-day survival benefit (41).

Like LP17, M3 has also been shown to be effective in treating sepsis in neonates (79). Using a cecal slurry model of sepsis, neonatal mice treated with i.p. M3 had attenuated disease severity, as measured by a reduction in serum IL-6, TNF- α , IL-1 β , and IFN- γ , and improved pulmonary and cardiac inflammation (79). Additionally, M3-treated neonates had improved cardiac function, measured by cardiac output and left-ventricular end diastolic diameter. Importantly, neonatal mice treated with M3 at either the time of cecal slurry injection or 2 hours later had improved 7-day survival (79). Although M3 is a novel peptide requires further studies, it shows promise in treating sepsis in both neonates and adults.

N1

N1 is a 10-aa inhibitory peptide based on the N-terminal aa sequence (aa 77-86) of PGLYRP1 (80). Using affinity chromatography, N1 was shown to bind sTREM-1 immobilized on CNBr-activated Sepharose (80). Using immunoblotting, N1 was also shown to bind TREM-1 on the surface of monocytes (80) (Figure 1). Monocytes and lymphocytes exposed to PGLYRP1 and Hsp70, two previously mentioned ligands of TREM-1, had decreased LDH release in the presence of N1. Additionally, N1 treated cells had decreased mRNA expression of TNF- α , IFN γ , IL-1 β , and IL-6 after exposure to LPS (80). In mice with acute lung injury after bronchial instillation of LPS and α -galactosylceramide, i.v. treatment with N1 protected against the resultant cytokine storm, leading to decreased serum levels of INFy and IL-4. These mice also had reduced histologically evaluated pulmonary inflammation (80). N1 is another novel peptide that effectively inhibits TREM-1 induced inflammation, but requires additional studies looking at its efficacy in a polymicrobial model of sepsis to better evaluate it as a treatment modality.

GF9

Moving away from the strategy of preventing ligands from interacting with TREM-1, GF9 is a ligand-independent

peptide (GLLSKSLVF) derived from murine TREM-1's transmembrane region, designed using the Signaling Chain HOmoOligomerization (SCHOOL) model (92). It functions by inhibiting the interaction between TREM-1 and its signaling partner, DAP12 (77, 92) (Figure 1). Using a mouse model of endotoxemia, the efficacy of i.p. GF9 was studied, administered in its free form, and carried within high density lipoproteins (HDL). Synthetic HDL was modified to target delivery and uptake by macrophages, therefore delivering the GF9 directly to the cell of interest. Mice subjected to endotoxemia had higher survival after pretreatment with GF9 both in free form and incorporated into macrophage-targeted HDL-like particles. However, mice required a dose of 25mg/kg of free GF9 to see a survival benefit compared to vehicle, where a significantly lower dose of 5mg/kg of HDL bound GF9 was able to achieve the same survival benefit. Additionally, incorporation of GF9 into HDL prolonged the peptide's half-life, which together with its targeted macrophage delivery could contribute to its reduced effective dose (77).

SLC-TREM-1

TREM-1 sneaking ligand construct (SLC-TREM-1) is another protein that inhibits the interaction of TREM-1 with DAP12 (Figure 1), but it was specifically designed to target the endothelium (78). It is composed of 3 portions: an E-selectin targeting domain that binds to the surface of endothelial cells, Pseudomonas aeruginosa exotoxin A to facilitate translocation from the endosomal vesicular system into the cytosol, and a 7-aa sequence (LSKSLVF) derived from the transmembrane region of TREM-1, and actually contained within GF9 (77, 78, 93). Endothelial cells stimulated with LPS in vitro had decreased TREM-1 expression and activation after treatment with SLC-TREM-1 (78). Additionally, treatment with this protein intraperitoneally improved the 10-day survival in mice subjected to CLP (78). GF9 and SLC-TREM-1 demonstrate that targeting the interaction of TREM-1 with DAP12 is a viable TREM-1 inhibitory strategy with potential for drug development in sepsis.

LR17/LR12

LR17/LR12 are the most studied inhibitory peptides of TREM-1. LR-17 is a 17-aa peptide (LQEEDAGEYGCMVDGAR) based on a highly conserved sequence between TREM-1 and TREM-like transcript-1 (TLT-1), a membrane bound protein found on the surface of activated platelets (23, 68, 94) (Figure 1). After demonstrating structural similarities between TREM-1 and TLT-1 using crystallographic approaches, soluble TLT-1 (sTLT-1) was studied as a modulator of the inflammatory response and shown to decrease human neutrophil production of TNF-a, IL-6, and IL-8 after LPS stimulation (23, 94). LR17 was subsequently shown to protect mice from LPS endotoxemia (23). Mice treated with LR17 before or after LPS administration had improved 10-day survival and decreased serum proinflammatory cytokines (23). This peptide also has a protective effect in mice with CLP-sepsis (23). Mice treated with LR17 had decreased sepsis severity demonstrated by lower levels of IL-6, TNF-a, and IL-10 in serum, peritoneal and bronchoalveolar

fluid, and IL-6 and TNF-a in the liver, and lung (23). LP17 also ameliorated sepsis-induced acute lung injury, reduced bacterial load in the spleen and blood, and an improved 7-day survival (23).

It was discovered that just 12 aa accounted for the LP17's anti-inflammatory effects, and a novel peptide containing this sequence (LQEEDAGEYGCM) was created and named LR12 (4, 68). After confirming TREM-1's expression on vascular endothelial cells, the effects of LR12 on vascular reactivity during sepsis was explored (57). Using both CLP-sepsis and LPS endotoxemia, it was determined that LR12 prevents the TREM-1 upregulation induced in the murine aortas and mesenteric arteries of mice subjected to these models (57). Additionally, LR12 treatment protected mice from sepsis-induced vascular dysfunction, measured by improved vascular contractility, and attenuated IL-6, TNF- α , and IL-10 expression as well as NOS and COX signaling pathway activation in murine aortas and mesenteric arteries (57).

The effects of LR12 during sepsis were further studied in adult minipigs to better characterize its beneficial properties as a therapeutic agent (68). Polymicrobial sepsis was induced using a cecal slurry model. Pigs treated with LR12 had less cardiovascular failure, measured by MAP, vasopressor use, and cardiac index (68). Additionally, they had decreased coagulopathy and organ failure in the lungs, liver, and kidney. LR12 improved 24-hour survival in septic pigs from 40% to 100% (68). The effects of LR12 was further explored in nonhuman primates using an LPS model of endotoxemia (81). Monkeys received an i.v. bolus of LPS followed by a continuous infusion of LR12 or placebo for 8 hours. Monkeys treated with LR12 were protected from hypotension and neutropenia and had reduced plasma cytokine concentrations (81). One month after LR12 administration, no side effects were noted in the treated animals (81). In addition to i.v. infusion, a sustained release implant was tested as a novel administration route for LR12 in rats (95). An in situ poly-lactide-co-glycolide (PGLA) or poly-lactide (PLA) implant was used to deliver LR12 in a dimerized formulation to diminish peptide degradation and allow sustained release (95). In healthy rats, these in situ forming implants delivered a therapeutic concentration of LR12 for 7 days, introducing an effective method of administrating this otherwise short-lived peptide (95).

Nangibotide

LR12 is the first TREM-1 inhibitor to have reached the clinical stage. It is currently being studied in Phase 2 clinical trials of sepsis, COVID-19, and acute myocardial infarction under the name Inotrem (nangibotide) (16, 82, 83, 96–98). Nangibotide is an immune modulator targeting the inflammatory response amplification maintained by TREM-1 (82). In Phase 1 trials, continuous i.v. administration of nangibotide was safe and well tolerated at doses up 6 mg/kg/h for 7 hours and 45 mins following a 15-minute loading dose of 5 mg/kg, with few adverse events noted after 28 days of followup (83). In a Phase 2a multicenter, randomized, double-blind, placebo-controlled clinical trial, patients received a continuous infusion of 0.3, 1.0, or 3.0 mg/kg/h of nangibotide within 24 hours of the diagnosis of

septic shock (82). Treatment was continued until the patient was off vasopressors for 12 hours, or for a maximum of 5 days. There was no difference in adverse events, tolerability, inflammatory biomarkers and clinical efficacy between study groups (82). Treatment with nangibotide, however, was associated with a decrease in the serum levels of IL-6 (82). In a subgroup analysis, patients with high sTREM-1 treated with nangibotide had decreased SOFA scores after treatment (82, 98), suggesting that sTREM-1 could be a potential biomarker to predict which septic patients respond to nangibotide. A pivotal Phase 2b trial, looking at the efficacy, safety, and tolerability of nangibotide in patients with septic shock is currently underway (16, 97). Patients are given two doses of the peptide for 3-5 days, depending on vasopressor requirements. The primary endpoint being evaluated is total SOFA score changes from baseline to day 5 in all patients, and will include a subgroup analysis of patients with elevated sTREM-1 baseline levels (16). Larger studies are still needed to investigate the efficacy of nangibotide in attenuating sepsis severity, and to evaluate sTREM-1 as a potential biomarker of patient response to nangibotide.

SUMMARY AND PERSPECTIVES

Over the last two decades, extensive research has revealed TREM-1's critical role as a mediator of inflammation in sepsis and other diseases. Substantial work has been done to identify its endogenous ligands and develop inhibitors to be used as potential therapeutic agents for a range of diseases. There are now a variety of inhibitory molecules, predominately oligopeptides, that have shown promise as a treatment for sepsis using various preclinical sepsis models in a number of species. These peptides are capable of targeting TREM-1 activation at various points within its signaling cascade, from acting as a decoy receptor and binding up its endogenous ligands, to binding the extracellular domain and competitively inhibiting the binding of endogenous ligands, to interrupting the transmembrane association of TREM-1 with DAP12. Beyond oligopeptides, other modulators of the TREM-1 pathway have been studied in non-infectious inflammatory diseases (99). Researchers in Sweden have developed an anti-TREM-1 antibody that reduces secretion of proinflammatory cytokines from lamina propria cells isolated from patients with inflammatory bowel disease (99). Targeted monoclonal antibody treatment has been effective in treating a variety of inflammatory diseases, and TREM-1 targeted antibodies may be a beneficial approach to sepsis drug development. Future research could benefit from investigating this novel anti-TREM-1 antibody in septic preclinical models.

Although many TREM-1 inhibitory peptides have shown promise in preclinical models, there are currently no FDA approved treatments for sepsis. Strategies targeting inflammatory mediators like TNF- α and IL-1 β were not effective in clinical trials for sepsis, despite efficacy in experimental models (100). Further, drotrecogin alpha, the only treatment to receive FDA approval for the treatment of sepsis was later withdrawn from the market (101). This recombinant human activated protein C did not reduce mortality in septic patients at 28 days in a follow-up phase 3 international, randomized, controlled trial, and is no longer approved as a treatment for sepsis (101). There are likely many barriers to creating effective sepsis treatments, including poor translation between animal models and human disease, heterogeneity of the patient population selected for clinical trials, and the overall heterogeneity of the etiology of the disease (102). The physiologic response to sepsis in mice differs from humans, and mice are also more resistant to endotoxin and resilient to infection. Further, many treatments tested in murine models are given before, during, or shortly after infectious exposure. Conversely, patients often present for treatment days after infection, with sepsis developing over time. Additionally, in preclinical models, treatments are often tested on sepsis from a single, well-defined source of infection, where clinical trials often include patients who present with sepsis from a wide variety of etiologies. The host response has great variation based on the location of infectious insult and inciting organism species. Patients also have a variety of specific characteristics, like sex, age, comorbidities, and medications that can all effect response to an experimental treatment. In that regard, clinical trials of sepsis treatments would possibly benefit from selecting patients according to the source of infection and infectious agent.

LR12, a 12-aa peptide, has been studied extensively in mice, rats, pigs, and nonhuman primates as a treatment for sepsis and has been shown to be effective and safe when administered as an i.v. bolus, continuous infusion, and an *in situ* forming implant. It is currently in phase 2b clinical trials under the name nangibotide and has so far been demonstrated to be safe and

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apparently effective in Phase-1 and Phase-2a studies. This peptide shows significant promise as a treatment for septic patients. It has demonstrated efficacy in seven preclinical models and in four different species, and has been administered up to 24 hours after onset of infection, addressing key issues of translatability and time of treatment in patients. Further, in the current trial, investigators are identifying patients with elevated levels of plasma sTREM-1 to evaluate whether these patients are better responder and, consequently, are more likely to benefit from treatment. By identifying a subgroup of patients who best respond to this treatment, administration can be personalized and efficacy can be better assessed. Sepsis has been called a "graveyard for pharmaceutical companies". Targeting the TREM-1 activation pathway, however, shows great promise and may finally become the first safe and effective therapeutic strategy to treat septic patients.

AUTHOR CONTRIBUTIONS

SS and MB conceived the original idea. SS prepared the figures and wrote the manuscript. PW and MB revised and edited the manuscript. PW supervised the project. All authors contributed to the article and approved the submitted version.

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GLOSSARY

aa	Amino acid
AKT	Protein kinase B
ALI	Acute lung injury
ALT	Alanine aminotransferase
AST	Aspartate aminotransferase
BAL	Bronchoalveolar lavage
BMDN	Bone marrow-derived neutrophils
CIRP	Cold-inducible RNA-binding protein
CLP	Cecal ligation and puncture
DAMP	Damage-associated molecular pattern
DAP12	DNAX activation protein 12
eCIRP	Extracellular cold-inducible RNA-binding protein
FRK	Extracellular signal-regulated kinase
FRFT	Eluorescence resonance energy transfer
HDI	High density lipoprotein
HMGB1	High mobility group box 1
	70 kDa baat shack protein
igG	Immunogiobulin G
Іква	Nuclear factor of kappa light polypeptide gene enhancer in B-cells
	inhibitor alpha
IL.	Interleukin
IP	Intraperitoneal
ITAM	Immunoreceptor tyrosine-based activation motif
IV	Intravenous
KO	Knockout
LDH	Lactate dehydrogenase
LPS	Lipopolysaccharides
MAP	Mean arterial pressure
MAPK	Mitogen-activated protein kinase
MCP	Macrophage chemoattractant protein
MMP	Metalloproteinases
mRNA	Messenger ribonucleic acid
MyD88	Myeloid differentiation primary response 88
NÉT	Neutrophil extracellular trap
NF-κB	Nuclear factor kappa-light-chain-enhancer of activated B cells
PAMPs	Pathogen-associated molecular patterns
PBMC	Peripheral blood mononuclear cell
PBS	Phosphate buffered saline
PGLA	Poly-lactide-co-alycolide
PGI VRP1	Pentidoalycan recognition protein 1
	Poptidoglycan
	Phoenbatidulinosital 2 kinasa
	Phosphalidyiii lositoi 5-kii lase
	Polymorphonuclear leukocyte
	Pattern recognition receptor
RAGE	Receptor for advanced glycation end products
RAS	Reticular activating system
rm	Recombinant murine
SCHOOL	Signaling chain homooligomerization
SLC-	Triggering receptor expressed on myeloid cells-1 sneaking ligand
TREM-1	construct
SPR	Surface plasmon resonance
sTREM-1	Soluble TREM-1
STSLS	Streptococcal toxic-shock-like syndrome
Syk	spleen tyrosine kinase
TLR	Toll-like receptor
TLT-1	Triggering receptor expressed on myeloid cells-1-like transcript 1
TNF-α	Tumor necrosis factor-a
TREM-1	Triggering receptor expressed on myeloid cells-1
WT	Wild-type.