



OPEN Nanoemulsification of PTX and BEZ235 inhibits colon cancer growth

Xingyao Zhu^{1,4}, Ruiqi Zhang^{1,4}, Yuhua Liu², Caiyun Yang², Qi Ding², Wenyi Gu³ & Hong Zou¹✉

Colorectal cancer is one of the most common malignant tumors, and its drug resistance poses a huge challenge and a serious threat to people's health. In previous studies, we have found that the combination of paclitaxel and BEZ235 has synergistic anti-colon cancer effects, particularly in drug-resistant colorectal cancer, demonstrating excellent anti-tumor effects. Therefore, this study aims to investigate the preparation of the nanoemulsification of paclitaxel and BEZ235 (NEs-PTX-BEZ235) and to explore the potential key parameters for its clinical translation. This study aims to provide a new therapeutic strategy for colorectal cancer, especially for drug-resistant colorectal cancer. The nanomaterials were prepared by low energy self-emulsification method, and analyzed by Malvern laser particle size analyzer. Cell drug uptake was observed under fluorescence microscope. CCK8, Western blot and flow cytometry were used to compare the anti-cancer effects and mechanisms of different experimental groups on ordinary and drug-resistant colon cancer cells. Human colon cancer primary cells were extracted to verify the anti-tumor effect of drugs. Finally, we found that NEs-PTX-BEZ235 significantly killed colon cancer cells, especially drug-resistant cells, and performed better than Nab-PTX. It may play a synergistic role in reducing drug resistance, inhibiting tumor stemness and inducing apoptosis of colon cancer cells by inhibiting drug resistance proteins.

Keywords Colon cancer, Paclitaxel, BEZ235, Nanoemulsification, Anti-tumor drug resistance

PI3K/Akt/mTOR signaling pathway is thought to be closely related to multidrug resistance in human colon cancer. It was reported that blocking of PI3K/AKT signaling pathway could enhance drug sensitivity of HCT-116/L-OHP resistant cells and reverse Pgp-mediated multidrug resistance in human colon cancer^{1,7}. Common PI3K/Akt/mTOR pathway inhibitors in the past include PI3K inhibitors Wortmannin and Alpelisib, Akt inhibitors Perifosine and Miltefosine, as well as mTOR inhibitors Rapamycin and Everolimus. BEZ235 is a newly discovered inhibitor of the PI3K/Akt/mTOR signaling pathway¹, which is well tolerated and has significant potential as a new anti-tumor drug. On the other hand, BEZ235 has a weak ability to induce apoptosis, and can trigger cell protective autophagy to offset its anti-tumor effect, so the anti-tumor effect of single drug is not ideal². Paclitaxel (PTX) is a diterpenoid alkaloid extracted from *Taxus* plants³, which can promote the apoptosis of tumor cells, inhibit the division and proliferation of tumor cells⁴, and will not produce cross-resistance with a variety of drugs, while it is effective against multi-drug-resistant tumors. It has been applied to the treatment of malignant tumors such as breast cancer, ovarian cancer and pancreatic cancer^{5,6}.

Previously, we have shown that PTX and BEZ235 have synergistic inhibitory effects on the growth of colon cancer cells and their drug-resistant cells. However, the combination of nanoemulsion loaded with PTX and BEZ235 showed more significant anti-tumor effect^{7,8}. On this basis, according to the existing optimal compatibility ratio of PTX and BEZ235, we designed a Nanoemulsification of paclitaxel and BEZ235 (NEs-PTX-BEZ235) with high efficacy, low toxicity and high patient compliance.

In this study, a low energy self-emulsification method was used to prepare a Nanoemulsification of paclitaxel and BEZ235. This emulsifier has uniform particle size distribution, stable properties, and can be fully absorbed by tumor cells. In vitro anti-tumor experiments show that this emulsifier has good effects of synergistically inhibiting tumor proliferation, promoting apoptosis, reducing tumor drug resistance and reversing tumor cell stemness, and provides a promising anti-tumor strategy for future clinical treatment of colon cancer, especially drug-resistant colon cancer.

¹Department of Pathology, The Second Affiliated Hospital of Zhejiang University School of Medicine, Zhejiang 310009, China. ²Department of Pathology, The First Affiliated Hospital, Shihezi University School of Medicine, Xinjiang 832002, China. ³Australian Institute of Bioengineering and Nanotechnology, University of Queensland, Queensland 4072, Australia. ⁴Xingyao Zhu and Ruiqi Zhang contributed equally as Co-first authors. ✉email: zouhong@zju.edu.cn

Materials and methods

Materials

Oxaliplatin-resistant (HCT116-LOHP) and cisplatin-resistant (HT29-DDP) human colon cancer cell lines were purchased from Shanghai Aurora Technology Co., LTD. HCT116 and HT29 human colon cancer cell lines were purchased from Shanghai Fuxiang Biotechnology Co., Ltd. Cisplatin and oxaliplatin, used to maintain drug resistance in two colon cancers, were purchased from Sigma in the United States. BEZ235 and PTX were purchased from Selleck Chemical, USA. Oxaliplatin and cisplatin are dissolved in saline, and PTX and BEZ235 are stored in dimethyl sulfoxide. Sources of other reagents related to cell culture are as follows: DMEM medium (Invitgen, USA), 0.25% EDTA trypsin (Solarbio, Beijing, China), fetal bovine serum (Biolnd, Israel), penicillin–streptomycin mixture (Solarbio, Beijing, China), dimethyl sulfoxide (Solarbio, Beijing, China), PBS (Shengong, Shanghai, China). CCK-8 cell proliferation toxicity assay kit (Tong Ren Chemical Research Institute, Japan) for cell proliferation, Nile Red (Solarbio, Beijing, China), FITC-labeled anti-rabbit IgG (ABCAM, USA), cell cycle assay kit (purchased from Unitech Biotechnology, China). Western blotting detected the antibody, and its concentration was PI3K (1:1000, Abcam, USA), p-Akt (1:2000, Abcam, USA), CD44 (1:2000, Abcam, USA), CD133 (1:1000, Abcam, USA), respectively. USA, Pgp(1:1000, Cst, USA), ABCC1(1:1000, Cst, USA), β -actin (1:1000, Abcam, USA); Hanks Buffer, Type IV collagenase, Tween 20 and propylene glycol were all purchased from Beijing Solarbio, China, Capryol 90 (Gattefosse, France).

Synthesis of NEs-PTX-BEZ235

PTX and BEZ235 were dissolved in DMSO, and Tween 20 and propylene glycol were first mixed into the emulsifier by vortex shaking (with a volume ratio of Tween 20 to propylene glycol of 2:1, shaken for approximately 3–5 min); then the double drug (PTX and BEZ235 were added in different concentration ratios of 1:1, 1:2, 1:4, 1:8 and 1:10) was added with Capryol 90 was added to the emulsifier and vortex shaken for 5 min; finally, the above mixture was slowly added to double distilled water and whirlpool shaken for 5 min (volume ratio of Capryol 90, emulsifier and water was 20:40:40) to form the nano complex formulation. Nano complex formulation characterized by Transmission electron microscopy (TEM). The average particle size and stability of the nano complexed formulations (1:1 ~ 1:10) were examined by a Malvern laser particle size analyzer, and the prepared nano complexed formulations were stored at 4 °C.

CCK-8 cell viability determination

The four cell lines were cultured in DMEM medium containing 10% fetal bovine serum and 5% penicillin/streptomycin. The HT29DDP cell-resistant line and the HCT116-LOHP cell-resistant line were supplemented with cisplatin (1.5 μ g/mL) and oxaliplatin (10 μ g/mL), respectively, in the medium to maintain the corresponding resistant phenotypes of the two drug-resistant cells. Cells were inoculated on 96-well plates at a cell density of 4×10^3 cells/well. After 24 h, the original DMEM medium was removed and replaced with fresh DMEM medium containing different concentrations of BEZ235, PTX, PTX + BEZ235, NE-PTX, NE-PTX combined with BEZ235, NEs-PTX-BEZ235 (1:1, 1:2, 1:4, 1:8 and 1:10 groups) and albumin-paclitaxel (Nab-PTX), respectively. Among them, general cell lines were mixed with PTX or NE-PTX medium of different concentrations using 25 nm BEZ235. In contrast, drug-resistant cell lines were mixed with PTX or NE-PTX medium of different concentrations using 50 nm BEZ235 on 96-well plates. After 48 h of treatment, 10 μ L CCK-8 reagent was added to each well under light protection, and the 96-well plate was wrapped in aluminum foil and placed in the incubator. Two hours later, the absorbance OD value was measured at 450 nm using the BIO-RAD xMark microplate instrument, and the cell survival rate was calculated according to the following formula:

$$\text{Cell survival rate\%} = \frac{(\text{absorbance OD value of experimental group} - \text{absorbance of blank group})}{(\text{absorbance OD value of control group} - \text{absorbance of blank group})} \times 100\%$$

A combination index was calculated to determine the synergetic effect of combination treatment based on the following formula:

$$CI = \frac{\text{availability of NEs} - 1 : 10}{\text{Cell availability of NE} - \text{PTX} \times \text{cell availability of BEZ235}}$$

$CI < 1$, $= 1$ and > 1 indicated synergic, additive and antagonistic effect, respectively.

In vitro cellular uptake of drugs

NE-Nile red was prepared by encapsulating Nile red dye in nanoemulsion. Cells were seeded in 6-well plates at a cell density of 2×10^5 cells/well. After 24 h, the original medium was removed and replaced by the medium containing NE-Nile red. After 0, 12, 24 and 48 h of incubation, the plates were washed with PBS, fixed with paraformaldehyde for 15 min, washed with PBS again, and incubated with DAPI for 15 min to stain the nuclei. Then the uptake of NE-Nile red by tumor cells was observed under a fluorescence microscope.

The expression of multidrug resistance, PI3K/Akt/mTOR pathway related molecules and dry protein were detected by Western blot.

The total protein was extracted after the cells were treated with different drugs for 48 h. The protein concentration was measured with a nanodrop spectrophotometer. Proteins were separated by Bio-RAD gel electrophoresis and transferred to PVDF membrane. After blocking for 2 h at room temperature (5% bovine serum albumin), the cells were incubated with rabbit anti-human Pgp, ABCC1, PI3K, p-Akt, CD133, CD44 and mouse anti-human

β -actin antibodies overnight at 4 °C, washed at room temperature, and treated with horseradish peroxidase-conjugated anti-rabbit and mouse antibodies for 2 h, then observed and imaged by ECL.

Note: After all antibody hybridization was completed, in order to obtain clearer and more visually appealing images, we cropped and photographed the imprinting blots. Original blots have been added to the supplementary materials.

Flow cytometry for cell cycle detection experiments

$2 \times 10^5 - 1 \times 10^6$ cells were collected, washed once with PBS, and centrifuged to discard the supernatant. Precooled 80% ethanol at -20 °C was added and fixed overnight. On the day of detection, the fixed cells were centrifuged, the supernatant was discarded, washed with PBS, added with 1 ml DNA Staining solution, vortexed for 5–10 s, and incubated for 30 min at room temperature in the dark. Detection was performed on a flow cytometer.

Colon cancer primary cell extraction

Primary cells were cultured by tissue block attachment method and collagenase digestion method. The fresh tissues of human colorectal cancer were collected and stored in the preservation solution (45 ml DMEM + 5 ml penicillin–streptomycin mixture). The tissues were taken out from the ultra-clean table, fat and necrotic tissues were removed, and washed with Hank's solution. The washed tissues were cut into small pieces of 1 mm³, washed with Hank's solution again, and the supernatant was discarded. It was resuspended with preservation solution and evenly arranged at the bottom of the culture flask. After adding DMEM culture medium, the culture flask was placed with the bottom upward for 2–4 h. When there was cell growth around the tissue block, it was considered that the culture was successful. Collagenase digestion method: The tissue block was cut into small pieces of 1 mm³, washed with Hanks solution, added with type IV collagenase, and placed in a 37 °C water bath for 1 h until the tissue block was dispersed, and then the cell suspension was collected and filtered through a 200-mesh sterile cell filter. The liquid that passed through the screen was transferred to a flask that had been previously filled with DMEM medium. Cells were cultured for 2–3 weeks and then purified by trypsin digestion.

Statistical analysis

The data were analyzed by SPSS.20 statistical analysis software, and the results were presented in the form of mean \pm standard deviation. The t-test was used to compare pairwise samples, and the analysis of variance was used to compare multiple samples. The difference was considered statistically significant at $P < 0.05$.

Results

Characterization of NEs-PTX-BEZ235

In this study, nanoemulsion was successfully synthesized by the oil-in-water method and characterized by Transmission electron microscopy (TEM) (Fig. 1A). The ratios of PTX to BEZ235 in the compound were set at 1:1, 1:2, 1:4, 1:8 and 1:10. The Malvern laser particle size analyzer examined the nanoparticle size, and the results showed that the particle size distribution of the nanoemulsion (NE) system was narrow, with an average particle size of 171.1 nm and a polydispersity index (PDI) of 0.142–0.202 (Fig. 1B). The particle size and PDI were similar in alkaline DMEM medium, neutral PBS and pH5.8 buffer. During the subsequent six days of continuous measurements (Fig. 1C), we found that the distribution and particle size of NEs-PTX-BEZ235 remained in the same range without significant changes, especially in the DMEM medium, indicating good stability of NEs-PTX-BEZ235.

In vitro cellular uptake of drugs

NE-Nile red was prepared by encapsulating Nile red dye in nanoemulsion. In colon cancer cells HT29, HCT116 and colon cancer drug-resistant cells HT29-DDP, HCT116-LOHP, the uptake of NE-Nile red by tumor cells at four time points of 0, 12, 24 and 48 h was observed (Fig. 2). The results showed that NE-Nile red was gradually absorbed and taken up by colon cancer cells and drug-resistant cells over time. It was almost completely absorbed into the cytoplasm of tumor cells at 48 h, indicating that there was a good uptake of NE by colon cancer cells.

Inhibitory effects on tumor cells of single drug group, double drug combination group, nano single drug group, NE-PTX combined with BEZ235 group, NEs-PTX-BEZ235 group and albumin-paclitaxel (Nab-PTX) group

Under the same drug concentration, the effect of the compound preparation 1:10 group was better than that of the other experimental groups, and its IC₅₀ was 2.096 and 1.276 in HT29 and HCT116 cells, respectively (Fig. 3A, Table 1). 9.836 and 6.803 in the drug-resistant cell lines HT29-DDP and HCT116-LOHP, respectively (Fig. 3B, Table 1). The combination index (CI) of NEs-PTX-BEZ235 (1:10) was calculated (Table 2), demonstrating the synergistic effect of PTX and BEZ235 in the compound preparation.

NEs-PTX-BEZ235 can significantly reduce the expression of PI3K/Akt/mTOR pathway related proteins p-Akt and PI3K in colon cancer cells and drug-resistant cells

BEZ235 is an inhibitor of PI3K/Akt/mTOR signaling pathway, and blocking PI3K/AKT signaling pathway can enhance the drug sensitivity of drug-resistant human colon cancer cells^{1,9}. In this study, Western blot was used to detect and compare the effects of different drug groups on the expression of PI3K and p-Akt in colon cancer cells and drug-resistant cells. As a result of detection in colon cancer cells HT29, HCT116, HT29-DDP, and HCT116-LOHP (Fig. 4A–B), The expressions of PI3K and p-Akt in the BEZ235 single drug group, PTX single drug group, PTX + BEZ235 group, NE-PTX group, NE-PTX combined with BEZ235 group and the NEs-PTX-BEZ235 groups with different double drug ratios are all down-regulated compared with the negative control group, and

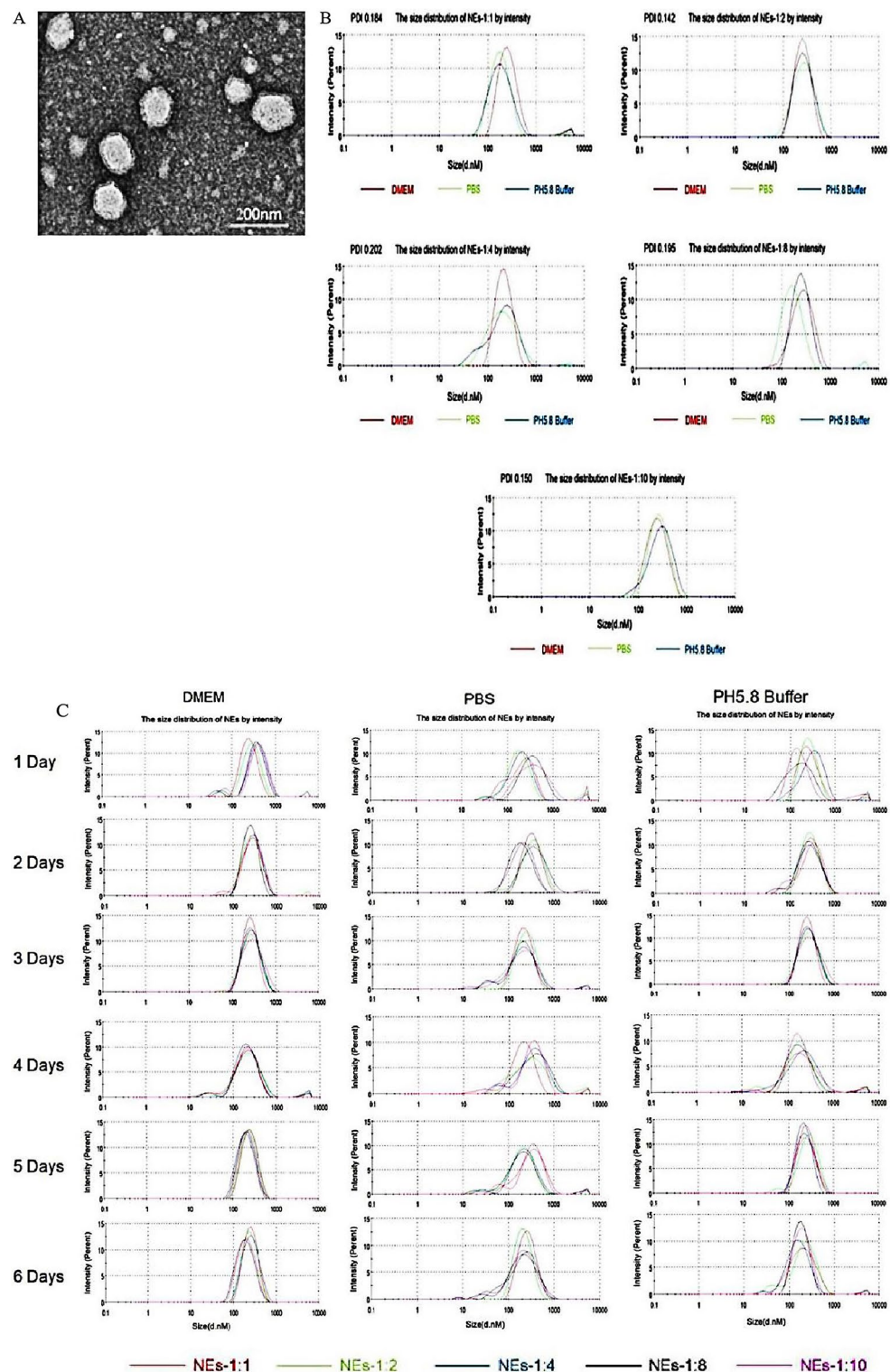


Fig. 1. Characterization of NEs-PTX-BEZ235. **A.** The synthesized NEs-PTX-BEZ235 was observed to be spherical with a smooth surface through TEM. **B.** The average particle size of 153.7 ± 2.16 nm. NEs-PTX-BEZ235 (1:1 ~ 1:10) has a narrow particle size distribution with a polydispersity index between 0.142 and 0.202, and has similar particle size and PDI in DMEM medium, PBS and pH 5.8 buffer; **C.** The particle size distribution of nanometer compound preparation (1:1 ~ 1:10) was measured for 6 consecutive days. It was found that the particle size did not change significantly in alkaline, neutral and acidic media, especially in alkaline DMEM medium.

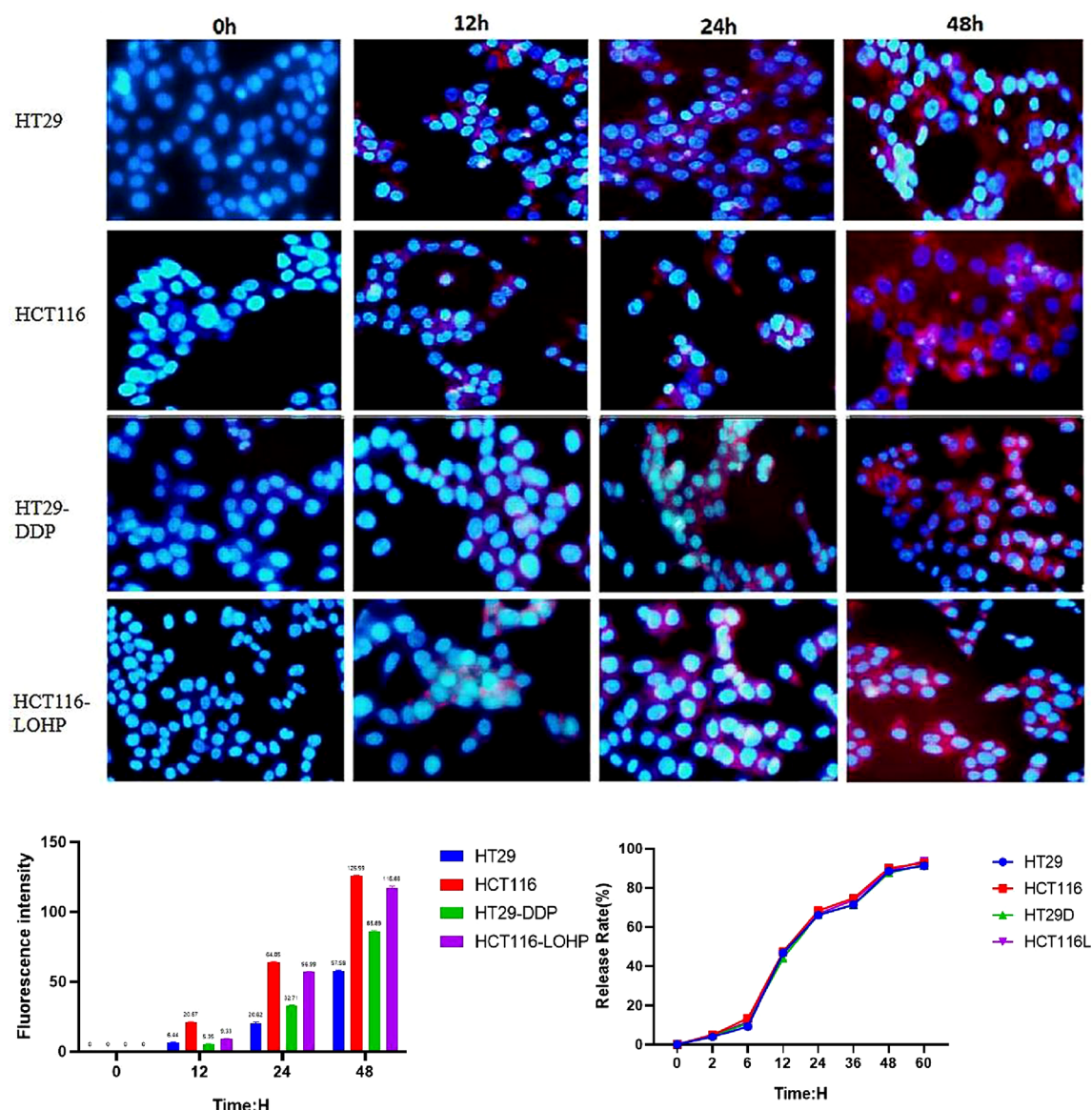
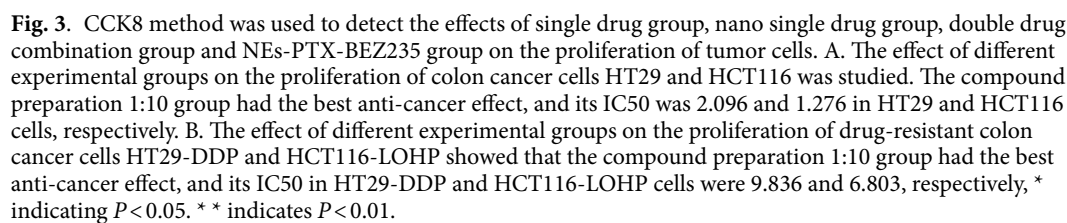


Fig. 2. With the increase of time, NE was gradually taken up by tumor cells, and almost completely entered the cells after 48 h. In PBS medium, the release rate of nanoemulsions in four tumor cells is basically the same.

the down-regulation effect of the NEs-1: 10 group is the most obvious. The difference was statistically significant ($P < 0.05$), indicating that the inhibitory effect of NEs-1: 10 group was better than that of other experimental groups (Supplementary Table 1,2).

NEs-PTX-BEZ235 preparation can significantly reduce the expression of CD44 and CD133 in colon cancer cells.

Within the tumor cell population, there are specific subsets known as cancer stem cells (CSCs) or tumor initiating cells (TICs)^{17,18}. These cells have malignant biological potentials such as self-renewal, multi-directional differentiation, unlimited proliferation and tumor reconstruction, and are considered to be the root of tumorigenesis, progression, invasion and metastasis, chemoradiotherapy resistance and recurrence^{19,20}. BEZ235 can inhibit the growth of colon cancer stem cells^{1,10}. In order to further explore the possible mechanism of tumor inhibition by drugs, Western blot was used to detect and compare the effects of different drug groups on the expression of stemness related proteins CD133 and CD44 in colon cancer cells and drug-resistant cells. As a result, in the colon cancer cells HT29, HCT116, HT29-DDP, and HCT116-LOHP (Fig. 4C–D). Compared with the negative control group, the expressions of CD133 and CD44 in the BEZ235 single drug group, PTX single drug group, PTX combined with BEZ235 group, NE-PTX group, NE-PTX combined with BEZ235 group and NEs-PTX-BEZ235 groups with different double drug ratios were all down-regulated, and the down-regulation effect of the NEs-1: 10 group was the most obvious. The difference was statistically significant ($P < 0.05$), indicating



Treatment	IC50 (nM)			
	HT29	HCT116	HT29-DDP	HCT116-LOHP
PTX	14.64	12.58	24.29	23.6
NE-PTX	3.778	3.347	15.82	14.94
BEZ235	174	146.8	517.8	416.1
NE-BEZ235	122.9	104.8	270.9	263.4
PTX + BEZ235	5.146	5.141	16.18	17.18
NE-PTX + BEZ235	3.228	3.285	10.14	10.1
NEs-1:1	4.531	2.79	17.21	11.06
NEs-1:2	2.733	1.444	11.32	9.514
NEs-1:4	2.856	1.517	13.41	9.818
NEs-1:8	2.648	1.483	13.87	8.551
NEs-1:10	2.096	1.276	9.836	6.803
Nab-PTX	11.32	11.21	36.36	32.94

Table 1. The IC50 (nM) of different drug groups in four tumor cells.

Cell	Concentration(nM)		Cell survival rate (%)			*CI
	BEZ235	PTX	BEZ235	PTX	NEs-1:10	
HT29	25	1	0.9	0.83	0.66	0.88
HCT116	25	1	0.9	0.81	0.58	0.80
	NE-BEZ235	NE-PTX	NE-BEZ235	NE-PTX	NEs-1:10	
HT29-DDP	50	10	0.89	0.65	0.45	0.78
HCT116-LOHP	50	10	0.85	0.6	0.46	0.90

Table 2. Combination index. $*CI = \frac{\text{availability of NEs-1:10}}{\text{Cell availability of NE-PTX} \times \text{cell availability of BEZ235}}$ $CI < 1, = 1 \text{ and } > 1$ indicated synergic, additive and antagonistic effect, respectively.

that the inhibitory effect of NEs-1: 10 group was better than that of other experimental groups (Supplementary Table 3,4).

NEs-PTX-BEZ235 can significantly reduce the expression of Pgp and ABCC1 protein in drug-resistant cells.

Chemotherapy for colon cancer often leads to the mutation of some genes in the tumor, or stimulates normal cells to produce some chemical factors suitable for tumor growth, thus causing multidrug resistance¹¹. ABCC1/MRP1 and ABCB1/Pgp are energy-dependent drug efflux pumps with an ATP-binding site and a drug-binding site. ATP can be hydrolyzed by binding to ABCC1 and Pgp to provide energy. This energy is used to pump out the drug entering the cell, resulting in the failure of the intracellular drug concentration to reach an effective level, thus causing drug resistance and leading to chemotherapy failure^{8,21}. In order to explore the mechanism of PTX and BEZ235 on drug-resistant colon cancer cells, we compared the expression of P-glycoprotein (Pgp) and multidrug resistance-associated protein (ABCC1) in different treatment groups by Western blotting. It was found that in HT29DDP and HCT116-LOHP (Fig. 4E), Compared with the negative control group, the expressions of Pgp and ABCC1 in the BEZ235 single drug group, PTX single drug group, PTX combined with BEZ235 group, NE-PTX group, NE-PTX combined with BEZ235 group and NEs-PTX-BEZ235 groups with different double drug ratios are all down-regulated, and the down-regulation effect of the NEs-1: 10 group is the most obvious. The difference was statistically significant ($P < 0.05$), indicating that the inhibitory effect of NEs-1: 10 group was better than that of other experimental groups. (Supplementary Table 5).

Flow cytometry was used to analyze and compare the effects of different proportions of NEs-PTX-BEZ235 on the cell cycle of colon cancer cells and their drug-resistant cells.

In the flow cytometry results of colon cancer cells HT29 and HCT116 (Fig. 5A–B, Table 3) and colon cancer drug-resistant cells HT29-DDP (Figure) and HCT116-LOHP (Fig. 5C–D, Table 3), it was found that, compared with the untreated control group, NEs-PTX-BEZ235 groups could induce G2/M phase arrest of cells, thereby promoting apoptosis of tumor cells. Among them, the NEs-1: 10 group had the best apoptosis-promoting effect, and the difference was statistically significant ($P < 0.05$) (Fig. 5E), and the effect was better at high concentration, which also indicated that the induction of G2/M phase arrest by nano-compound drugs on colon cancer cells and their drug-resistant cells was concentration-dependent.

NEs-PTX-BEZ235 has significant inhibitory effect on primary colon cancer cells

The primary cells of colorectal cancer were extracted from human colorectal cancer specimens and purified by trypsin digestion after successful culture, and the extracted cells were tested by flow cytometry. The CCK-8

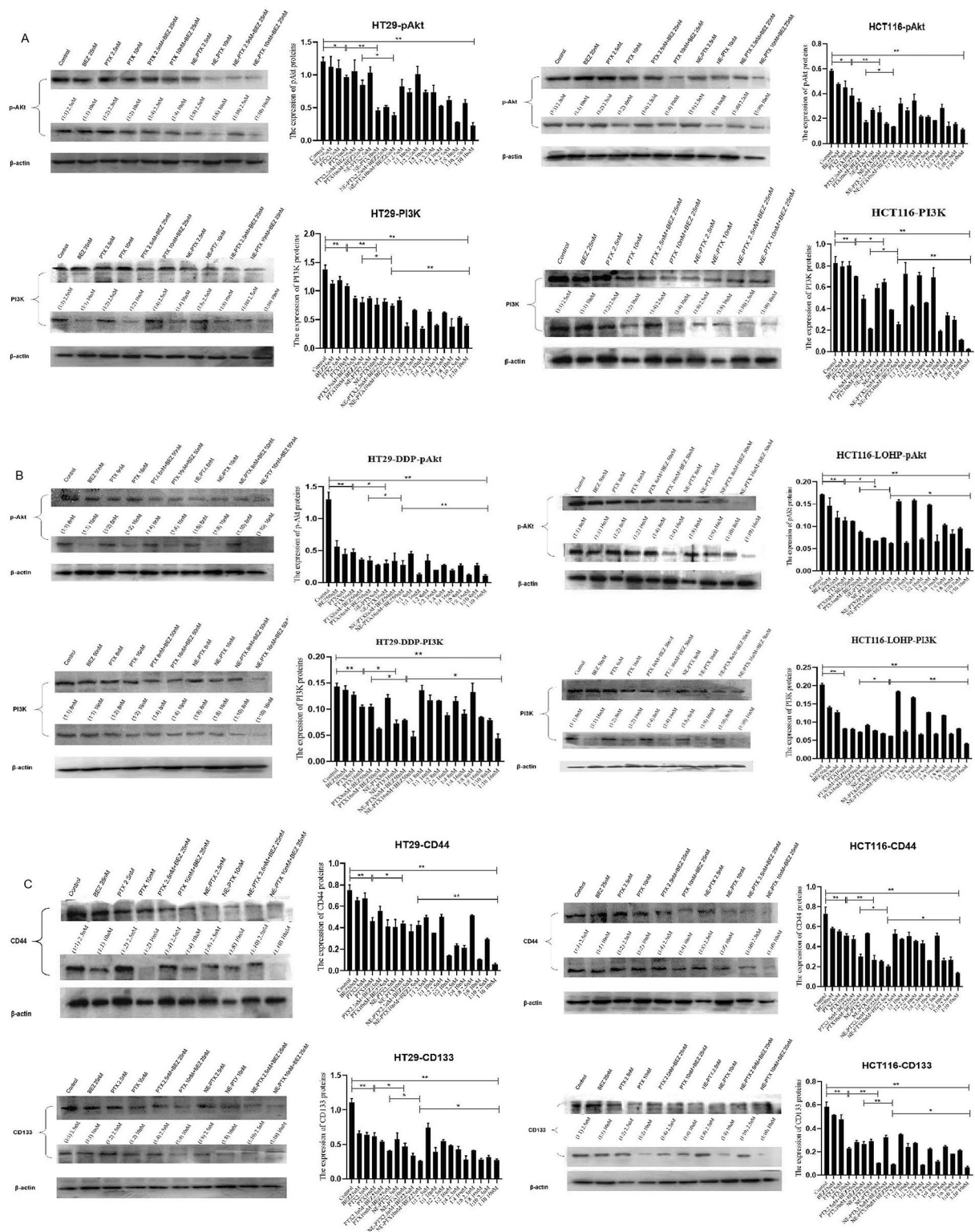
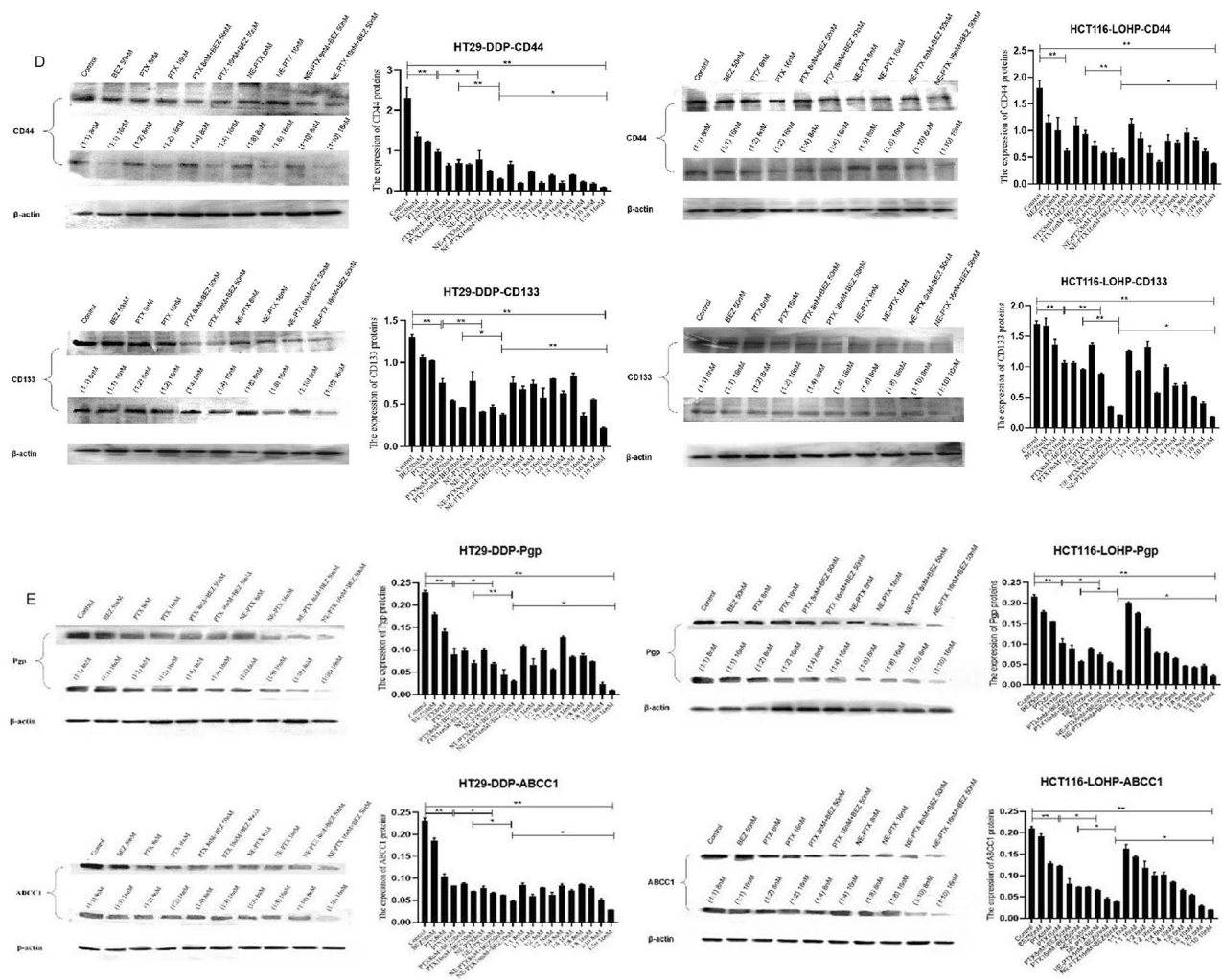


Fig. 4. (A–B) Effects of different experimental groups on PI3K/Akt/mTOR pathway related proteins in tumor cells. * means $P < 0.05$, ** means $P < 0.01$. (C–D) Effects of different experimental groups on tumor cell stem Protein. * means $P < 0.05$, ** means $P < 0.01$. (E) Effects of different experimental groups on drug-resistant protein of colon cancer drug-resistant cells. * means $P < 0.05$, ** means $P < 0.01$.



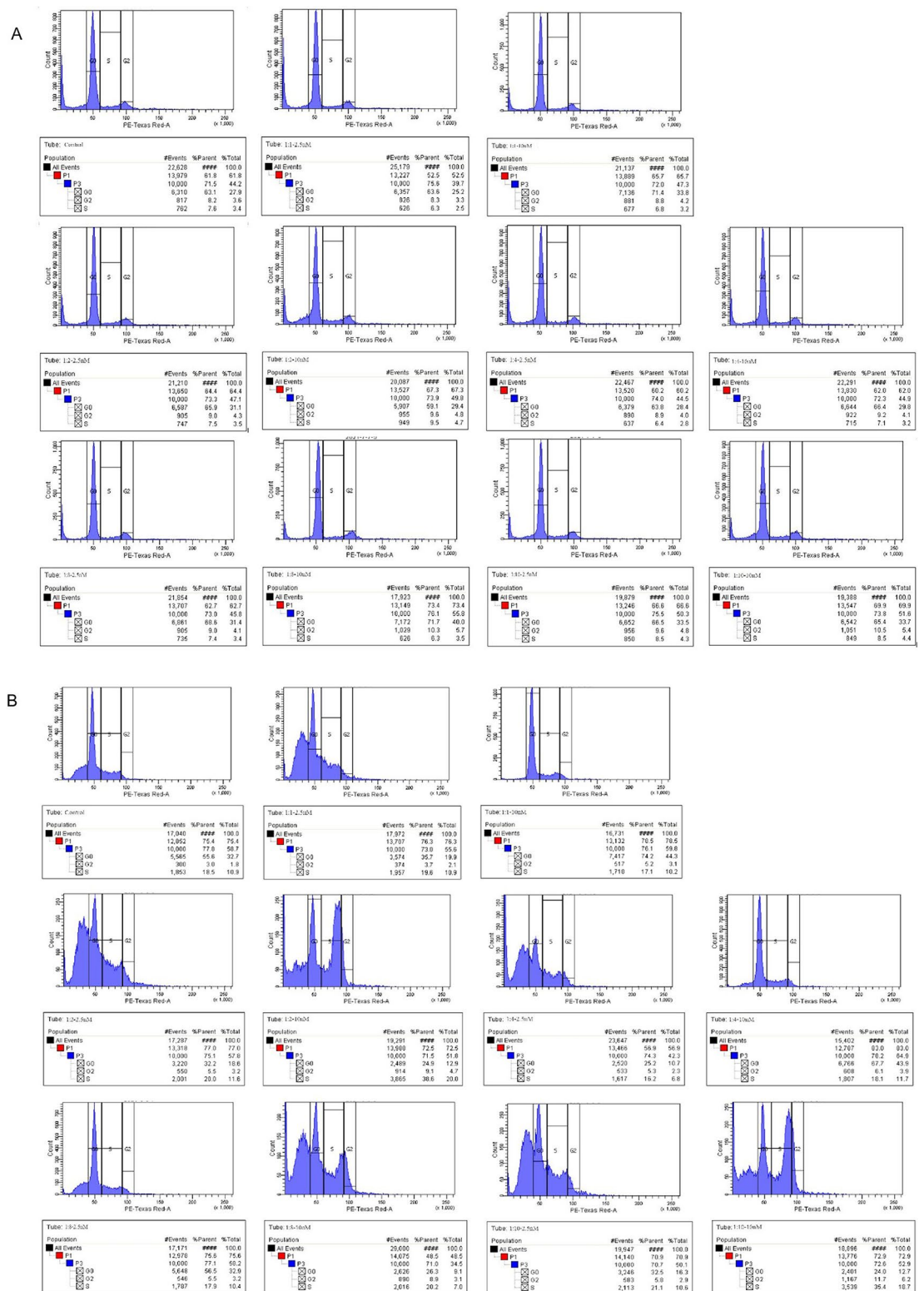


Fig. 5. Flow cytometry was used to analyze and compare the effects of different proportions of NEs-PTX-BE2235 on the cell cycle of tumor cells. In HT29 (A) and HCT116 (B), HT29-DDP (C) and HCT116-LOHP (D), different compound preparations could induce G2/M phase arrest of tumor cells, and the effect of NEs-1:10 group was the most obvious. E. Comparison of the percentage of cells in G2/M phase after treatment between different administration groups, * and ** indicate $P < 0.05$ and $P < 0.01$, respectively, compared with the control group.

increase the solubility of poorly soluble drugs, enhancing their stability and bioavailability^{22,23}. In this study, NEs were used to deliver a combination of PTX and BE2235, with the aim of overcoming PTX resistance and

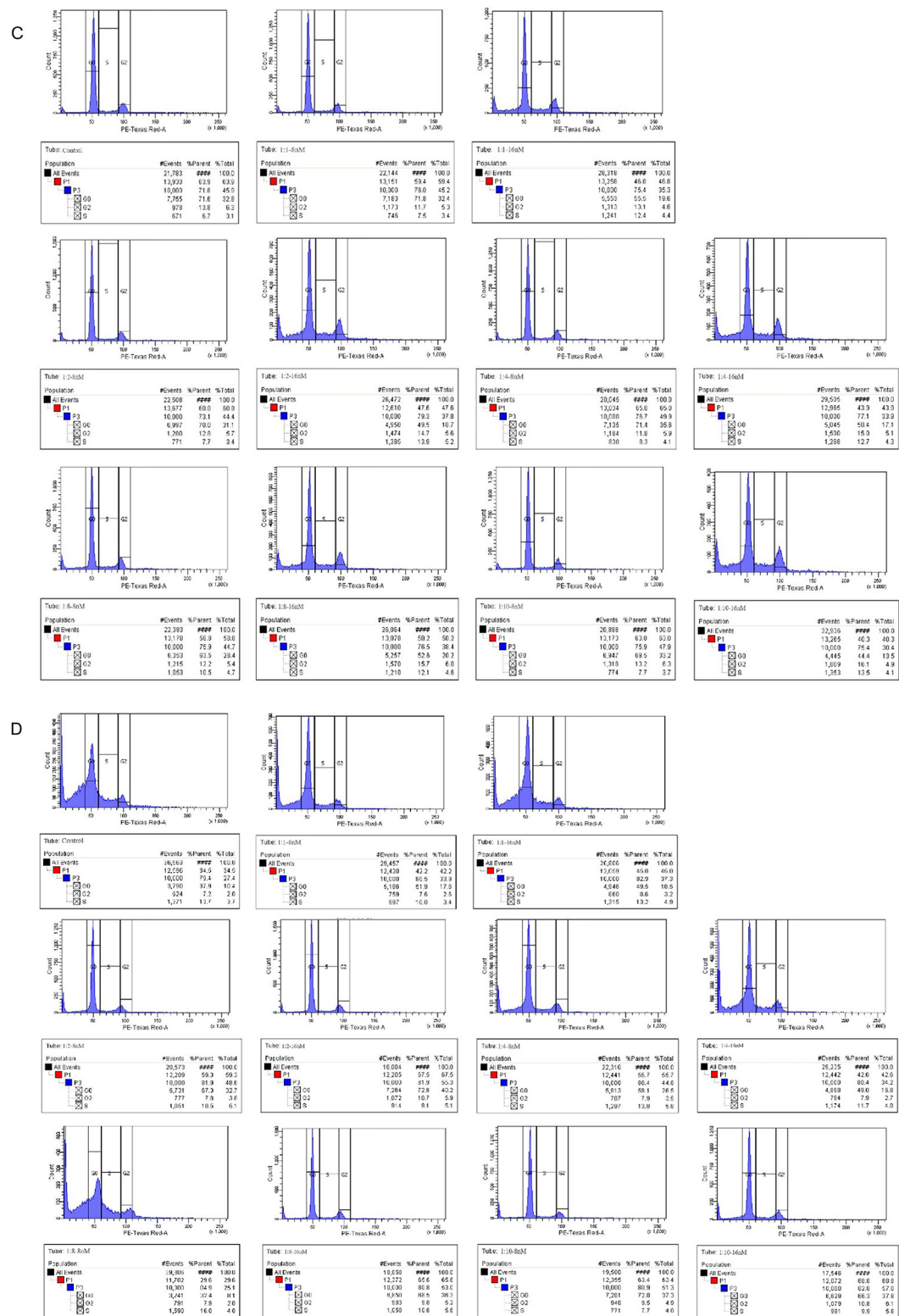


Figure 5. (continued)

enhancing its antitumor activity. The research results demonstrate that this nanoemulsion formulation (NES-PTX-BEZ235) exhibits superior antitumor effects in both colon cancer cell lines and their drug-resistant counterparts. By optimizing the drug concentration ratio (1:10), NES-PTX-BEZ235 not only effectively reduced the IC₅₀ value but also demonstrated significantly better efficacy than clinically commonly used nab-paclitaxel (Nab-PTX), suggesting potential advantages of this formulation in overcoming drug resistance.

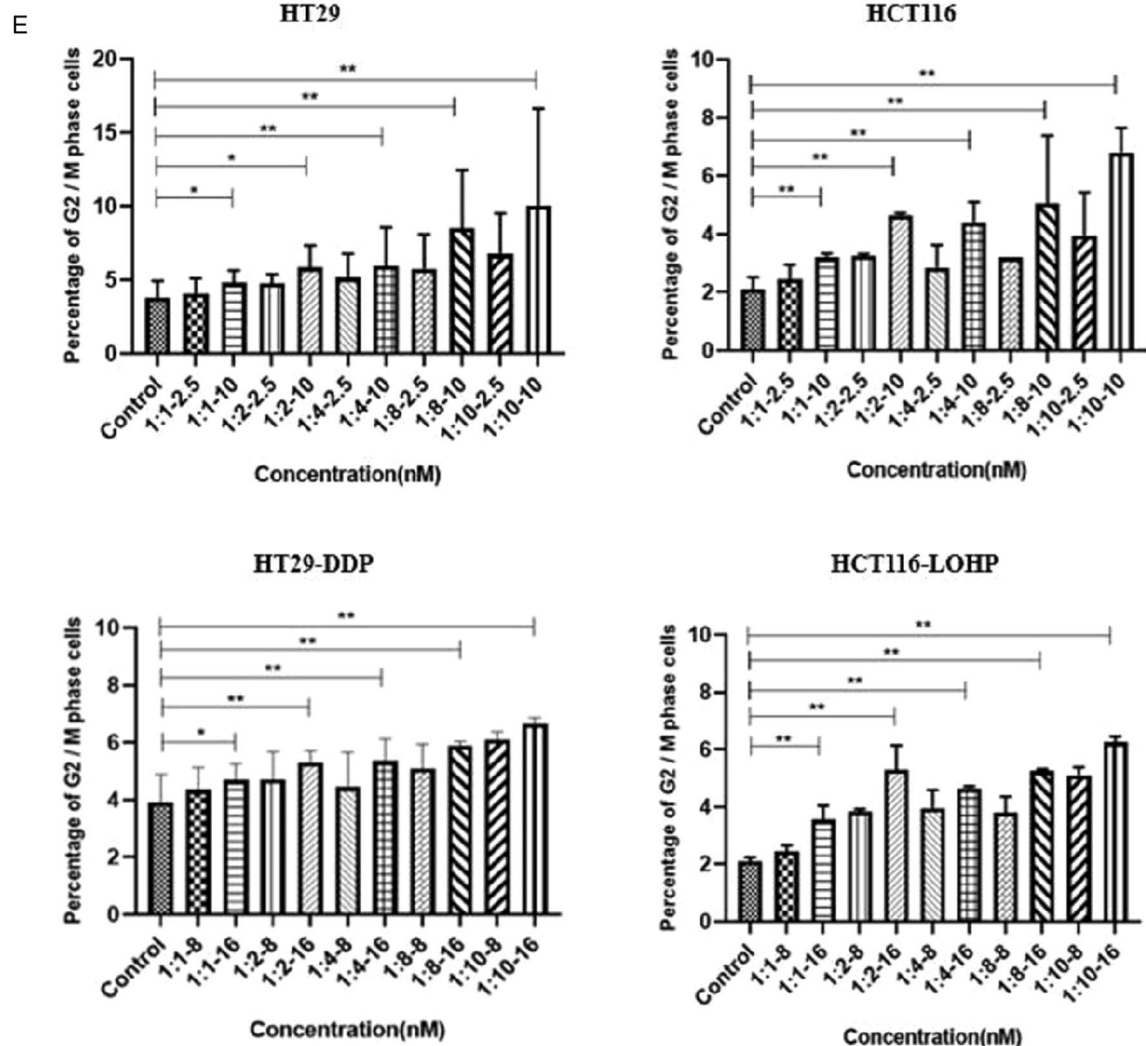


Figure 5. (continued)

Percentage of G2/M phase cells (%)					
Treatment	Cell		Treatment	Cell	
	HT29	HCT116		HT29-DDP	HCT116-LOHP
Control	3.70	2.07	Control	3.43	2.50
NEs(1:1)-2.5 nM	4.10	2.43	NEs(1:1)-8 nM	3.93	3.00
NEs(1:1)-10 nM	4.83	3.20	NEs(1:1)-16 nM	4.57	4.10
NEs(1:2)-2.5 nM	5.07	3.43	NEs(1:2)-8 nM	4.27	4.17
NEs(1:2)-10 nM	6.27	4.83	NEs(1:2)-16 nM	5.43	5.63
NEs(1:4)-2.5 nM	5.37	3.07	NEs(1:4)-8 nM	4.47	4.30
NEs(1:4)-10 nM	6.27	4.50	NEs(1:4)-16 nM	5.50	5.30
NEs(1:8)-2.5 nM	6.00	3.43	NEs(1:8)-8 nM	4.87	4.43
NEs(1:8)-10 nM	8.60	5.33	NEs(1:8)-16 nM	5.97	6.03
NEs(1:10)-2.5 nM	7.60	4.47	NEs(1:10)-8 nM	5.53	5.50
NEs(1:10)-10 nM	10.87	7.07	NEs(1:10)-16 nM	6.53	7.00

Table 3. Percentage of G2/M phase cells treated with tumor cells in different experimental groups.

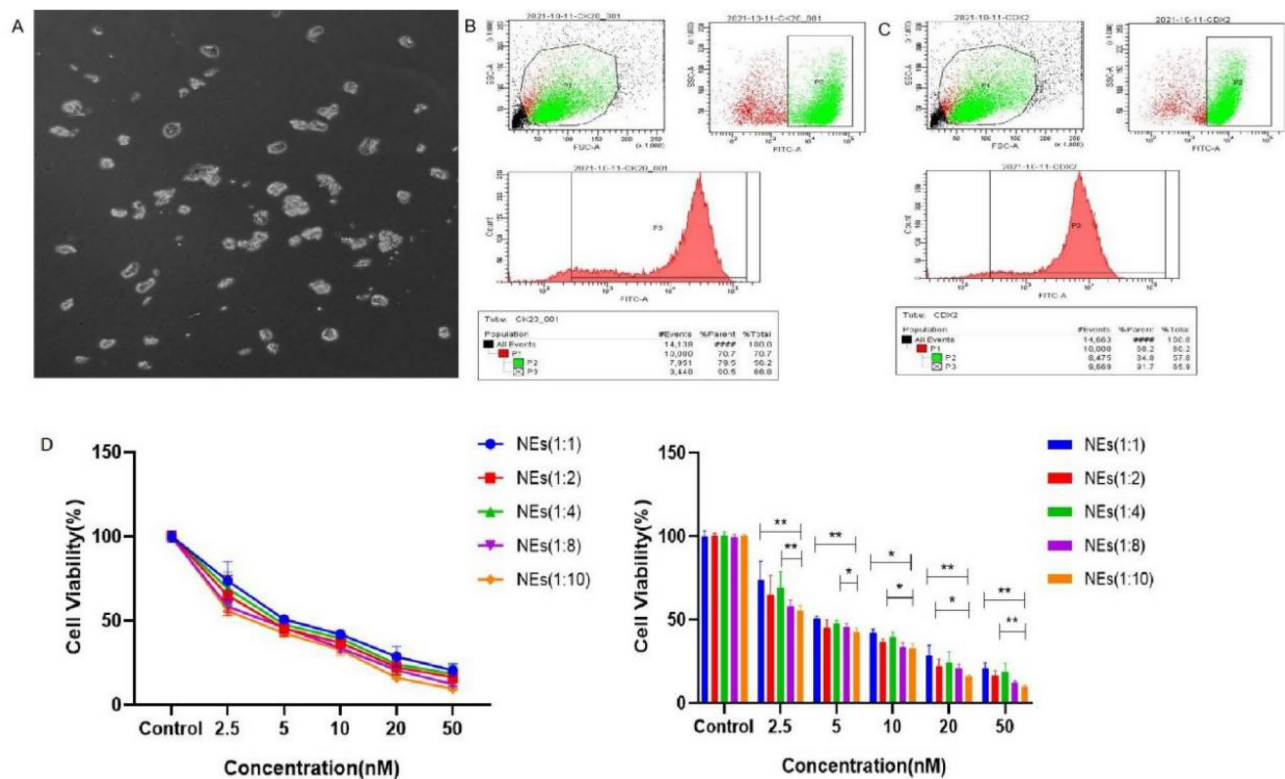


Fig. 6. The primary colon cancer cells were treated with different proportions of NEs-PTX-BEZ235 with gradient concentration, and the effects of different groups on the proliferation of primary colon cancer cells were detected and compared. **(A)** Extracted human colon cancer primary cells (100×); **(B)** The positive rate of CK20 was 90.5% by flow cytometry. The positive rate of CDX2 was 91.7% by flow cytometry. The IC₅₀ of different proportions of NEs-PTX-BEZ235 on primary human colon cancer cells were 5.371 nM, 3.813 nM, 4.475 nM, 3.857 nM and 3.471 nM, respectively. * indicates $P < 0.05$, ** indicates $P < 0.01$.

Mechanism studies revealed that NEs-PTX-BEZ235 can enhance the sensitivity of colon cancer cells through multiple mechanisms. On the one hand, this combination formulation blocks the pro-proliferative and anti-apoptotic effects of the PI3K/Akt/mTOR signaling pathway in tumor cells by significantly inhibiting the expression of related proteins (PI3K and p-Akt) in this pathway. On the other hand, BEZ235 can downregulate the expression of drug resistance-related proteins ABCB1 and Pgp, thereby reducing drug efflux and increasing the intracellular accumulation of PTX. In addition, this study found that NEs-PTX-BEZ235 can induce G2/M phase arrest in colon cancer cells and promote cell apoptosis through a Caspase-8-dependent pathway. The synergistic effects of these mechanisms may be key reasons for the enhanced cytotoxicity and overcoming of drug resistance exhibited by this formulation in colon cancer cells. Compared with previous studies, this study further validates the potential of the nanoemulsion-loaded combination strategy of PTX and BEZ235 in the treatment of colon cancer.

Previous studies have shown that MEK inhibitors combined with PTX can enhance the killing effect on colon cancer cells. Building on this, the current study further optimized the delivery of PTX using nanoemulsion technology and combined it with the synergistic sensitizing effect of BEZ235, achieving stronger antitumor effects at lower doses. Compared with traditional PTX formulations, the advantages of NEs-PTX-BEZ235 lie in improved solubility and bioavailability of PTX, while also reducing systemic toxicity, offering a potentially better treatment option in clinical applications.

However, this study still has certain limitations. Firstly, it is primarily based on in vitro cell models and has not yet conducted in vivo animal experiments to verify its biodistribution, pharmacokinetic characteristics, and long-term safety. Secondly, although this study revealed that NEs-PTX-BEZ235 may function by inhibiting the PI3K/Akt/mTOR pathway, downregulating drug resistance proteins, and promoting cell apoptosis, other possible mechanisms for overcoming drug resistance, such as autophagy and gene expression regulation, still need further exploration. Therefore, future studies will further evaluate the therapeutic effects of NEs-PTX-BEZ235 in animal models and explore broader mechanisms of overcoming drug resistance, laying a more solid foundation for its clinical translation.

In summary, this study confirms the significant antitumor activity of NEs-PTX-BEZ235 in colon cancer cells, reveals its potential mechanism of action, and provides new research ideas for overcoming PTX resistance. This strategy is expected to become a new direction in the treatment of colon cancer drug resistance and provide an important basis for further preclinical studies.

Data availability

All data generated or analysed during this study are included in this published article [and its supplementary information files].

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Author contributions

X.Z.: Data curation, Formal analysis, Methodology, Validation, Writing—original draft. R.Z.: Methodology, Resources, Writing—original draft. Y.L.: Data curation. C.Y.: Data curation. Q.D.: Data curation. W.G.: Supervision. H.Z.: Conceptualization, Formal analysis, Funding acquisition, Project administration.

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Declarations

Competing interests

The authors declare no competing interests.

Ethical approval

The experiment has been approved by the Human Research Ethics Committee of the Second Affiliated Hospital, Medical College of Zhejiang University.

Additional information

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Correspondence and requests for materials should be addressed to H.Z.

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