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Review

Prostate Cancer: De-regulated Circular RNAs With Efficacy in Preclinical *In Vivo* Models

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Abstract

Therapy resistance, including castration-resistance and metastasis, remains a major hurdle in the treatment of prostate cancer. In order to identify novel therapeutic targets and treatment modalities for prostate cancer, we conducted a comprehensive literature search on PubMed to identify de-regulated circular RNAs that influence treatment efficacy in preclinical prostate cancer-related *in vivo* models. Our analysis identified 49 circular RNAs associated with various processes, including treatment resistance, transmembrane and secreted proteins, transcription factors, signaling cascades, human antigen R, nuclear receptor binding, ubiquitination, metabolism, epigenetics and other target categories. The identified targets and circular RNAs can be further scrutinized through target validation approaches. Down-regulated circular RNAs are candidates for reconstitution therapy, while up-regulated RNAs can be inhibited using small interfering RNA (siRNA), antisense oligonucleotides (ASO) or clustered regularly interspaced short palindromic repeats/CRISPR associated (CRISPR-CAS)-related approaches.

Keywords: Circ RNA-protein interaction, castrate-resistant prostate cancer (CRPC), drug delivery, metastasis, miR-sponging, siRNA, review.

Introduction

Prostate cancer is one of the most common malignancies worldwide. In 2023, 288,000 cases were diagnosed in the US, resulting in 34,700 deaths (1).

Localized prostate cancer can be cured, however, disseminated prostate cancer is associated with a poor prognosis. Treatment options include surgery, radiation, chemotherapy, androgen-deprivation therapy, and inhibition of release of gonadotrophin-releasing hormone.

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Patients with breast cancer (BRCA) gene mutations benefit from treatment with poly-ADP ribose polymerase (PARP) inhibitors. However, development of therapy resistance such as castration-resistance and metastasis to distant organs hamper current treatment of advanced prostate cancer and are associated with a poor prognosis (2-4). Due to the immune-suppressive micro-environment and low mutational burden, modest therapeutic benefit was noted in only a very small subset of patients after treatment with immune-checkpoint inhibitory antibodies (5, 6). In addition, the therapeutic potential of immune cell recruitment with bispecific antibodies and chimeric antigen receptor (CAR) T-cell therapy requires further optimization to achieve significant clinical benefits (7). Several antibodydrug conjugates (ADCs) and proteolysis targeting chimera (PROTACs) are currently evaluated in patients with prostate cancer (8-10). Despite ongoing clinical studies, there is a significant need to identify novel targets and develop new treatment modalities. Therefore, we conducted a literature search to identify de-regulated circular RNAs (circRNA) and their corresponding targets that exhibit activity in prostate cancer-related preclinical *in vivo* models. We focused on this category of circRNAs, because demonstration of efficacy in preclinical in vivo models is an important milestone in cancer drug development.

Circular RNA

Approximately 98% of human RNAs are non-coding, while only 2% encode proteins (11). Non-coding RNA can be categorized into protein synthesis-related RNAs such as ribosomal and transfer RNAs, regulatory RNAs such as linear long non-coding RNA ≥200 nucleotides (nts), small non-coding RNAs such as microRNA (miRs), small nuclear, small nucleolar RNA, and circ RNA (12). The latter subtype was first identified in plant viroids (13), later found in the human delta hepatitis virus (14), and subsequently found in mammalian cells (15). CircRNAs are highly stable and range in size from under 100 nts to over 4 kb (16). In cancer, circRNAs can exert tumor-suppressive as well as oncogenic properties and accordingly can be down- or up-

regulated (17). CircRNAs mediate a plethora of functions. both intracellularly and as exosomes. These functions include sponging of miRs, regulation of transcription, splicing, and translation through protein binding and scaffolding, interaction with RNA, and, in some cases, the encoding of proteins (18). CircRNA can be generated by direct back-splicing, intron-driven circularization, exon skipping, and debranching of intron lariats (18). Circularization generates new junctional sequences, which allow specific targeting of circRNAs (18). Up-regulated circRNAs can be inhibited using siRNA, antisense oligonucleotides (ASO), and CRISPR-CAS (19, 20). The function of down-regulated circRNAs can be restored through gene replacement therapy with expression vectors (21). Diagnostic, prognostic, and therapeutic aspects of circRNAs in prostate cancer have been summarized in several reviews (22-24). In this review, we focus on the therapeutic role of circRNAs in prostate cancer with documented efficacy in preclinical prostate cancer-related in vivo models.

Circular RNAs Involved in Therapy Resistance

Circ0004087 mediates paclitaxel (PTX) resistance through up-regulation of mitotic checkpoint ser/thr kinase BUB1. Circ0004087 (Figure 1) was aberrantly over-expressed in prostate cancer tissues (25). Knockdown of circ0004087 led to a decrease in PTX resistance in PC3 and DU145 prostate cancer cells in vitro and in vivo after subcutaneous (SQ) implantation into nude mice (25). Mechanistically, circ0004087 bound to transcription coactivator Staphylococcal nuclease-domain containing protein 1 (SND1), transactivated transcription factor MYB, and induced expression of BUB1 (25). SND1 is a multifaceted protein involved in transcriptional activation, regulation of mRNA stability, alternative splicing, ubiquitination, and can act as an oncogene in certain types of cancer (26, 27). BUB1 is involved in the spindle checkpoint mediating error-free mitosis correction (28). It has been shown that inhibition of BUB1 reduces PTX resistance in prostate cancer cells (29).

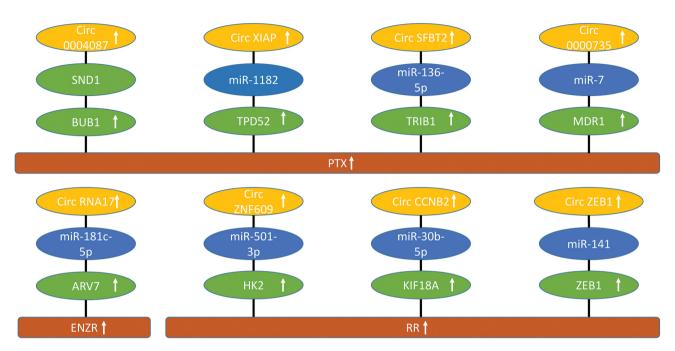


Figure 1. Circular RNAs contributing to treatment resistance of prostate cancer with efficacy in preclinical in vivo models. First line: specifies circular RNA; second line: indicates type of molecule circRNA is interacting with; third line: highlights de-regulated targets; fourth line: indicates type of resistance. Upward arrows indicate up-regulation, downward arrows indicate down-regulation. ARV7: Androgen receptor splice variant 7; BUB1: mitotic checkpoint ser/thr kinase BUB1; circCCN2: circ cyclin B2; circSFBT2: circ scm-like with four malignant brain tumor domains; circXIAP: circ X-linked inhibitor of apoptosis; circZEB1: circ zinc finger E-box binding homeobox 1; circZNF 609: circ ring finger 609; ENZR: enzalutamide resistance; HK2: hexokinase 2; KIF18A: kinesin family member 18A; MDR1: multi-drug resistance protein 1; miR: micro RNA; PTX: paclitaxel; RR: radio-resistance; SND1: staphylococcal nuclease domain enhancing protein 1; TPD52: tumor protein D52; TRIB1: tribbles homolog 1.

Circ X-linked inhibitor of apoptosis (circXIAP) mediates PTX-resistance by up-regulating tumor protein D52 (TPD52). CircXIAP (Figure 1) was up-regulated in PTX-resistant prostate cancer tissues and cell lines (30). CircXIAP was found in exosomes and its depletion in PTX-resistant DU145 and PC3 cells increased PTX-sensitivity in vitro and in nude mice (30). CircXIAP sponged miR-1182 and up-regulated TPD52 (30). The latter has been shown to be involved in protein trafficking and cytokinesis (31). TPD52 is over-expressed and amplified in prostate cancer and induces prostate cancer cell growth by transactivation of nuclear factor κ B (NF κ B) (31, 32). Furthermore, TPD52 inhibits apoptosis and promotes metastasis in preclinical prostate cancer-related models (33-35).

Circ scm-like with four malignant brain tumor domains (circSFBT2) induces PTX-resistance by up-regulating

tribbles homolog 1 (TRIB1). Exosomal circSFBT2 (Figure 1) increased PTX-resistance in prostate cancer cells *in vitro* and *in vivo* in nude mice. CircSFBT2 sponged miR-136-5p and up-regulated TRIB1 (36). The latter is a member of the SER-THR pseudokinase family, which can act as tumor suppressors as well as tumors promoters (37). The C-terminus of TRIB1 controls stability of interacting proteins through ubiquitination and proteasome-dependent degradation (37). TRIB1 is frequently over-expressed and amplified in prostate cancer and promotes cell survival by regulation of endoplasmic reticulum chaperone expression (38, 39).

Circ0000735 induces PTX-resistance by up-regulating multi-drug resistance protein 1 (MDR1). Circ0000735 (Figure 1) was up-regulated in PTX-resistant prostate cancer tissues and cells (40). Inhibition of circ0000735

increased PTX sensitivity of DU145-PTX and PC3-PTX resistant cells, both *in vitro* and *in vivo* in nude mice. Circ0000735 sponged miR-7 and up-regulated MDR1, cyclin D1, and B cell lymphoma 2 (BCL2), mediators of chemo-resistance, viability, progression, and colony formation of prostate cancer cells (2, 40-43).

Circ RNA17 inhibits enzalutamide resistance (ENZ-R) by down-regulating androgen receptor splice variant 7 (ARV7). CircRNA17 (Figure 1) was down-regulated in prostate cancer tissues and cell lines (44). CircRNA17 inhibited the expression of ARV7 in CRPC C4-2 cells, reduced proliferation and invasion, and increased sensitivity to androgen receptor (AR) antagonist enzalutamide (ENZ) in vitro and in an orthotopic nude mice model. This effect also involved miR-181c-5p (44). The underlying mechanism requires further investigation to be fully understood. ARV7 is a C-terminally truncated version of the AR, which lacks hormone binding activity (45). It has been shown that ARV7 contributes to prostate cancer progression, as well as resistance to ENZ and abiraterone (46, 47). It has also been demonstrated that ARV7 causes PTX resistance by inactivation of the spindle assembly checkpoint (48).

Circ zinc finger 609 (circZNF609) induces radio-resistance by up-regulating hexokinase 2 (HK2). CircZNF609 (Figure 1) was highly expressed in prostate cancer tissues and its knockdown in DU145 and VCaP cells decreased viability, migration, invasion, glycolysis, and increased sensitivity against radiation (49). Silencing of circZNF609 elevated radiosensitivity of DU145-related xenografts in nude mice. CircZNF609 sponged miR-501-3p and up-regulated HK2 (49). It has been shown that HK2, the rate-limiting enzyme of glycolysis, mediates radio-resistance in tumors such as cervical cancer, hepatocellular carcinoma (HCC), and laryngeal carcinoma (50-52).

Circ cyclin B2 (circCCNB2) mediates radio-resistance by up-regulating kinesin family member 18A (KIF18A). CircCCNB2 (Figure 1) was found to be up-regulated in

radiation-resistant prostate cancer tissues and cell lines (53). Knockdown of circCCNB2 increased radiosensitivity of prostate cancer cells *in vitro* and in xenografts in nude mice. CircCCNB2 inhibited autophagy of prostate cancer cells by sponging miR-30b-5p and upregulating KIF18A (54). KIF18A is a member of the kinase family of motor proteins that are associated with microtubules (54). KIF18A is correlated with radioresistance in esophageal and prostate cancer (55, 56).

Circ zinc finger E-box binding homeobox 1 (circZEB1) mediates radio-resistance by up-regulating transcription factor ZEB1. CircZEB1 (Figure 1) was up-regulated by testicular receptor 4 (TR4), and quaking (QKI). CircZEB1 sponged miR-141-3p, resulting in up-regulation of ZEB1, which mediates radio-resistance of prostate cancer cells (57). TR4 functions as a nuclear receptor in prostate cancer (58). QKI is an RNA binding protein, which is upregulated by AR in prostate cancer (59). Regarding radio-resistance, it was shown that combination therapy involving radiation therapy and metformin promoted radiosensitivity in preclinical mouse models of prostate cancer (57, 60). ZEB1 has been identified as a promoter of epithelial-mesenchymal transition (EMT) and stem cell properties in prostate cancer (61) and as a mediator of DNA damage response in breast cancer (62).

Circular RNAs Modulating Expression of Transmembrane and Secreted Proteins

Circ homeodomain interacting protein kinase 3 (circHIPK3) up-regulates metadherin (MTDH). CircHIPK3 (Figure 2) was up-regulated in prostate cancer tissues, mediated proliferation, migration, and invasion of PC3 and 22Rv1 prostate cancer cells *in vitro* and promoted growth of corresponding xenografts in nude mice. It also sponged miR-448 and up-regulated MTDH (63). The latter is also known as astrocyte elevated gene 1 (AEG1) and functions as a transmembrane protein in the plasma membrane, endoplasmic reticulum, nucleus, and nucleoli (64, 65). Plasma membrane MTDH was preclinically validated as a

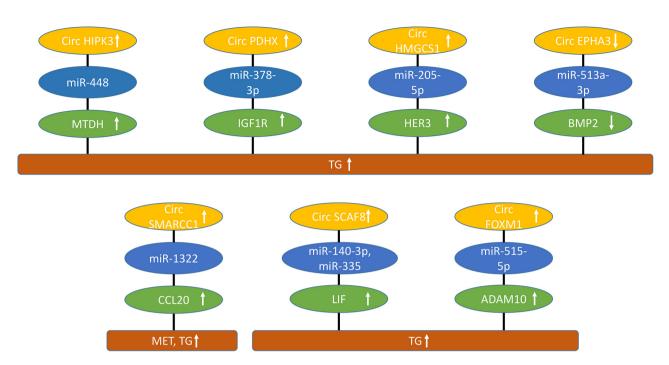


Figure 2. Circular RNAs targeting prostate cancer-related transmembrane and secreted proteins with efficacy in preclinical in vivo models. First line: specifies circRNA; second line: indicates miRs interacting with corresponding circRNA; third line: highlights de-regulated targets; fourth line: indicates impact on tumor growth and metastasis. Upward arrows indicate up-regulation, downward arrows indicate down-regulation. ADAM10: A disintegrin and metalloproteinase domain containing protein 10; BMP2: bone morphogenetic protein 2; CCL20: C-C chemokine ligand ligand 20; circFOXM1: circ forkhead box M1; circ EPA3: circ EPH receptor A3; circHIPK3: circ homeodomain interacting kinase 3; circHMGCS1: circ hydroxymethylglutaryl-CoAsynthase 1; circ PDHX: circ pyruvate dehydrogenase complex component X; circSCAF8: circ SR-related CTD-associated factor 8; circSMARCC1: circ SWI/SNF related matrix associated, actin dependent regulator of chromatin, subfamily C, member 1; HER3: human epidermal growth factor receptor 3; IGF1R: insulin-like growth factor receptor 1; LIF: leukemia inhibitory factor 8; M: metastasis; miR: micro RNA; MTDH: metadherin; TG: tumor growth.

target for the treatment of breast cancer metastasis (65) and is currently evaluated as a target in multiple types of cancers (66). In a preclinical prostate cancer *in vivo* model, it has been shown that genetic ablation of MTDH inhibits progression and metastasis (67). Furthermore, it has been demonstrated that inhibition of MTDH enhances the sensitivity of prostate cancer cells to cisplatin (68).

Circ pyruvate-dehydrogenase complex component X (circPDHX) up-regulates insulin-like growth factor 1 receptor (IGF1R). Expression of circPDHX (Figure 2) was associated with poor prognosis in patients with prostate cancer (69). Circ PDHX promoted proliferation and colony formation of 22Rv1 and PC3 prostate cancer cells in vitro and its knockdown in PC3 cells inhibited tumor

growth in nude mice. CircPDHX sponged miR-378-3p and up-regulated IGF1R (69). The latter has been considered as a target for treatment of many types of cancer (70). In prostate cancer, insulin-like growth factor 1 (IGF-1)/IGF1R interaction has been shown to promote survival and renewal, migration, spread, and resistance to radiation and castration (71, 72). Multiple agents have been developed that inhibit the IGF/IGF1R axis and evaluated in clinical studies including prostate cancer. They were all terminated, because the projected clinical endpoints were not reached and due to the unavailability of predictive biomarkers (72, 73).

Circ hydroxymethylglutaryl-CoA synthase1 (circ HMGCS1) up-regulates human epidermal growth factor receptor 3

(HER3). CircHMGCS1 (Figure 2) was over-expressed in prostate cancer tissues and promoted proliferation of DU145 and PC3 cells *in vitro*, as well as the growth of these cell lines as xenografts in nude mice. CircHMGCS1 sponged miR-205-5p and up-regulated HER3 (74). The latter is an actionable target for the treatment of advanced prostate cancer and is activated by neuregulin 1 (NRG1), which is expressed by tumor-infiltrating monomyelocytic cells (75). Several anti-HER3 drug-conjugates such as U3-1402 and AMT-562 are currently evaluated clinically for the treatment of solid tumors (75, 76). Furthermore, it has been shown that antibody-based inhibition of HER2/HER3 signaling overcomes heregulin-induced resistance to phosphoinosite-3-kinase (PI3K) inhibition in prostate cancer (77).

Circ EPH receptor A3 (circEPHA3) up-regulates bone morphogenetic protein 2 (BMP2). CircEPHA3 (Figure 2) was down-regulated in high-grade prostate cancer tissues and cell lines (78). It prevented proliferation, migration, and invasion of DU145 and PC3 cells in vitro. CircEPHA3 suppressed growth of corresponding xenografts after SQ implantation into nude mice as well as experimental metastasis after tail vein injection. It was revealed that circEPHA3 sponged miR-513a-3p, resulting in upregulation of BMP2 (78). The latter is a member of the transforming growth factor β (TGFβ) superfamily of proteins, which inhibits proliferation and survival, and modulates the tumor microenvironment (TME) by inducing differentiation of macrophages into osteoclasts (79). It has been shown that BMP2 is lost during prostate cancer progression resulting in inhibition of metastasis (80, 81). In PC3 cells, BMP2 is involved in the suppression of SMAD-1 signaling, induction of p21, retinoblastoma (RB) phosphorylation, and an increase in osteoprotogerin (OPG), which inhibits osteoclastogenesis (82).

Circ SWI/SNF-related, matrix-associated, actin-dependent regulator of chromatin subfamily C, member 1 (circSMARCC1) up-regulates C-C chemokine ligand 20 (CCL20). CircSMARCC1 (Figure 2) was up-regulated in prostate cancer tissues and correlated with Gleason score and tumor stage (83). It

accelerated proliferation, migration and invasion of DU145 and PC3 cells *in vitro*. CircSMARCC1 promoted tumor growth of prostate cancer xenografts as well as experimental metastasis in nude mice. As an underlying mechanism, sponging of miR-1322 and subsequent up-regulation of CCL20 was identified. CircSMARCC1 was associated with colonization of prostate cancer xenografts by M2 macrophages leading to tumor progression (83). The CCL20/C-C chemokine receptor 6 (CCR6) axis has been shown to promote tumor progression by enhancing proliferation and migration of tumor cells, as well as macrophage polarization in the TME (84, 85). In prostate cancer, suppression of the AR induces activation of the CCL20/CCR6 axis (86). CCR6 expression correlates with prostate cancer progression (87).

Circ SR-related CTD associated factor 8 (circSCAF8) upregulates leukemia inhibitory factor (LIF). CircSCAF8 (Figure 2) was positively correlated with prostate cancer progression and was also found in urine extracellular vesicles (88). It promoted proliferation, migration, and invasion of 22Rv1 and PC3 prostate cancer cells in vitro. CircSCAF8 increased the growth of SQ and orthotopic PC3 xenografts in nude mice, as well as experimental metastasis to the lungs in RM1 murine prostate cancer cells. It sponged miRs 140-3p and -335 and up-regulated leukemia inhibitory factor (LIF). Knockdown of circSCAF8 inhibited LIF, signal transducer and activator of transcription 3 (STAT3), matrix metalloproteinases 2 and 9 (MMP2, MMP9), vascular endothelial growth factor A (VEGFA), and cyclin D1 (88). LIF represents a pleiotropic cytokine of the interleukin 6 (IL6) family, which activates oncogenic pathways such as janus kinase (JAK)/STAT3, mitogen-activated protein kinase (MAPK), ser/thr kinase AKT, and mechanistic target of rapamycin (mTOR) through interaction with the LIF receptor (LIFR). The latter signals after association with the transmembrane co-receptor gp130 (89, 90). A monoclonal antibody directed against LIF (MSC1) and a small molecule inhibiting the interaction between LIF and LIFR (EC359) are presently evaluated in clinical studies in several types of cancer (90, 91). It has

been shown in preclinical models that LIF promotes the expression of CXC-ligand 9 (CXCL9) in tumor-associated macrophages and prevents CD8+ T-cell tumor infiltration, thereby impairing anti-programmed cell death protein 1 (anti-PD1) therapy (92). It was demonstrated that LIF mediates CRPC in preclinical models (93). However, LIF signaling remains complex, since in addition to LIF, the LIFR also binds to cardiotrophin 1 (CTF1), ciliary neurotrophic factor (CNTF), and oncostatin M (OSM) (94). The identification of biomarkers indicating response to LIF-based therapy is still pending.

Circ forkhead box M1 (circFOXM1) up-regulates A disintegrin and metalloproteinase domain-containing protein 10 (ADAM10). CircFOXM1 (Figure 2) was upregulated in prostate cancer tissues and cells (95). It mediates proliferation, cell-cycle progression, migration, and invasion of prostate cancer cells and promotes tumor growth of prostate cancer-related xenografts in nude mice. CircFOXM1 sponged miR-515-5p and up-regulated ADAM10 (95). In the LNCaP prostate cancer in vivo model, dihydrotestosterone, IGF1, and epidermal growth factor (EGF) up-regulate ADAM10 (96). In prostate cancer, ADAM10 can act as an oncogenic sheddase, because ADAM10-cleaved ephrin A5 (EPHA5) mediates prostate cancer metastasis (97, 98). Furthermore, it has been shown that nuclear translocation of ADAM10 can act as a transcriptional regulatory factor via interaction with the AR and contributes to pathogenesis and progression of prostate cancer (99).

Circular RNAs that Modulate Expression of Transcription Factors

Circ0074032 up-regulates homeobox A1 (HOXA1). Circ0074032 (Figure 3) was up-regulated in prostate cancer and associated with poor prognosis (100). Its down-regulation curbed proliferation, migration, and invasion of prostate cancer cells *in vitro* and inhibited growth of corresponding xenografts in nude mice. Circ0074032 sponged miR-198 and up-regulated HOXA1 (100). HOXA1

is a member of a family of 38 members that contain a DNA binding domain, contribute to all hallmarks of cancer (101), and are also involved in the pathogenesis of prostate cancer (102). HOXA1 has been identified as a breast cancer oncogene (103). Furthermore, it has been shown that HOXA1 enhances cell proliferation and metastasis of prostate cancer cells by promoting extracellular signal-regulated kinases 1/2 (ERK1/2) and AKT signaling (104).

Circ formin 2 (circFMN2) up-regulates LIM-homeobox gene 2 (LHX2). CircFMN2 (Figure 3) was up-regulated in prostate cancer tissues and correlated with advanced tumor stage, as well as lymph node and distant metastasis (105). In DU145, PC3, and VCaP cells, circFMN2 promoted proliferation, migration, and invasion in vitro and its knockdown inhibited the growth of VCaP-derived xenografts in nude mice. CircFMN2 sponged miR-1238 and up-regulated LHX2 (105). LHX2 is a member of the LIM homeobox transcription factor family, which consists of 12 members. Each member contains two LIM-domains and a centrally located homeodomain that interacts with DNA (106-108). In a transgenic breast cancer model, it has been shown that LHX2 induces autocrine and paracrine plateletderived growth factor receptor (PDGFR) signaling (109). In prostate cancer, the mechanistic role of LHX2 remains to be elucidated in further detail.

Circ dehydrogenase/reductase 3 (circ DHRS3) up-regulates Meis homeobox 2 (MEIS 2). CircDHRS3 (Figure 3) was down-regulated in high-grade prostate cancer (110). It decreased proliferation and migration of DU145 and PC3 cells in vitro. Over-expression of circDHRS3 inhibited growth of corresponding xenografts after SQ implantation into nude mice, as well as bone- or lung metastases after intratibial or tail vein injection. CircDHRS3 sponged miR-421 and up-regulated the homeobox gene MEIS2, which acts as a transcription factor (110). Depending on the tumor type, MEIS2 can function as an oncogene or a tumor suppressor (111, 112). In prostate cancer, MEIS2 suppresses proliferation and its decreased expression correlates with poor prognosis (113). Epigenetic silencing

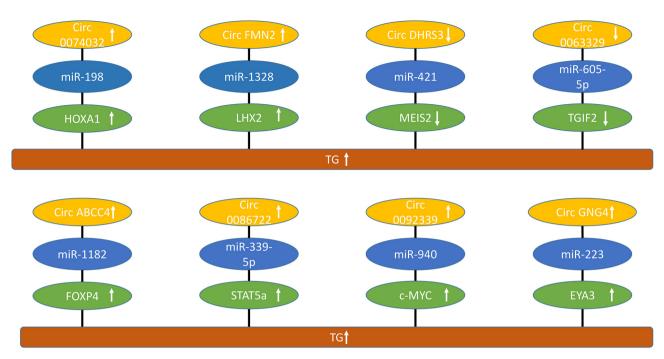


Figure 3. Circular RNAs targeting prostate cancer-related transcription factors with efficacy in preclinical in vivo models. First line: specifies circRNA; second line: indicates miR interacting with corresponding circRNA; third line: highlights de-regulated targets; fourth line: indicates impact on tumor growth and metastasis. Upward arrows indicate up-regulation, downward arrows indicate down-regulation. c-MYC: Transcription factor c-MYC; circABCC4: circ ATP-binding cassette subfamily C, member 4; circDHRS3: circ dehydrogenase/reductase 3; circFMN2: circ formin 2; circGNG4: circ G-protein subunit gamma 4; EYA3: EYA transcriptional co-activator and phosphatase 3; FOXP4: forkhead box P4; HOXA1: homeobox A1; MEIS2: meis homeobox 2; miR: micro RNA; STAT5a: signal transducer and activator of transcription 5a; TGIF2: TG interacting factor 2; TG: tumor growth.

of MEIS2 has been observed in patients with prostate cancer recurrence (114).

Circ 0063329 up-regulates TG interacting factor 2 (TGIF2). Circ 0063329 (Figure 3) was down-regulated in prostate cancer and inhibited proliferation and migration of DU145 and PC3 cells (115). It decreased growth of SQ implanted PC3 cells in nude mice. Circ0063329 sponged miR-605-5p and up-regulated TGIF2 (115). TGIFs belong to the family of TALE homeoproteins, which act as transcription factors (116). The latter can recruit histone deacetylase 1 after interaction with SMAD4 and act as a co-repressor of SMAD/TGF β signaling (117, 118). It has been shown that TGIF2 inhibits EMT in prostate cancer (119).

Circ ATP-binding cassette sub-family C member 4 (circABCC4) up-regulates transcription factor forkhead

box P4 (FOXP4). The expression of circABCC4 (Figure 3) correlated with a shorter five-year survival rate in prostate cancer patients (120). Inhibition of circABCC4 mediated reduced proliferation, migration, and invasion in DU145 and PC3 cells and decreased tumor growth of corresponding xenografts in nude mice. CircABCC4 sponged miR-1182 and up-regulated transcription factor FOXP4 (120). Forkhead proteins are categorized into subfamilies such as FOXA, FOXC, FOXM1, FOXO, and FOXP and are known to activate or repress genes by recruiting co-factors or repressors. They are commonly deregulated in cancers (121, 122). They primarily recruit histone deacetylases (121, 122). FOXPs can act as oncogenes as well as tumor suppressors (123). FOXP2 was shown to activate MET receptor tyrosine kinase signaling in prostate cancer (124). FOXP4 was associated with prostate cancer risk in Chinese men (125).

Circ0086722 up-regulates signal transducer and activator of transcription 5a (STAT5a). Circ0086722 (Figure 3) was highly expressed in prostate cancer tissues and cell lines and correlated with worse recurrence-free survival (126). Circ 0086722 promoted prostate cancer cell proliferation and progression in vitro and in vivo. Circ0086722 sponged miR-339-5p and up-regulated STAT5a (126). Seven STAT proteins have been identified, and they act as transcription factors in response to extracellular signals (127). STATs contain a src-homology 2 (SH2) domain, a DNA binding domain, and a C-terminal transcriptional activation domain (127). STAT5a is a target in prostate cancer and promotes prostate cancer growth in vitro and in vivo (128, 129). Furthermore, it was shown that STAT5a undergoes amplification during prostate cancer progression (130). Activation of STAT5A predicts early recurrence of prostate cancer in patients treated with radical prostatectomy (131, 132).

Circ0092339 up-regulates transcription factor c-MYC. Circ0092339 (Figure 3) was highly expressed in castration-resistant prostate cancer (CRPC) cells (133). It promoted proliferation of CRPC cells *in vitro* and in nude mice by sponging miR-940 and up-regulating c-MYC (133). c-MYC functions as a pleiotropic transcription factor, which affects proliferation, growth, cell differentiation, metabolism, and apoptosis (134, 135). In prostate cancer, up-regulation of c-MYC is associated with reduced overall survival, lymph node metastasis, and resistance to chemo- and radiotherapy (136). Small molecule c-MYC inhibitor OmoMYC is presently evaluated in cancer patients in clinical trials (137).

Circ G-protein subunit gamma 4 (circGNG4) up-regulates EYA transcriptional co-activator and phosphatase 3 (EYA3). CircGNG4 (Figure 3) was highly increased in prostate cancer (138). It promoted growth of LNCaP and PC3 prostate cancer cells *in vitro*, as well as the growth of PC3 xenografts in nude mice. CircGNG4 sponged miR-223 and up-regulated EYA3 (138). Four eyeless absent proteins (EYAs) have been identified. They associate with sine oculis

homeobox (SIX) transcription factors, are translocated into the nucleus, and act as transcriptional co-activators (139). EYAs exhibit tyrosine and threonine phosphatase activity located on separate domains and activate transcription of c-MYC (138, 139). In a breast cancer xenograft model, it has been shown that EYA3 partners with protein phosphatase 2A (PP2A) to induce c-MYC stabilization and metastasis (140). Furthermore, it has been demonstrated that src-phosphorylated EYA3 is a driver of cell proliferation (141).

Circular RNAs Modulating Components of Signaling Pathways

Circ0003258 up-regulates RHO quanine nucleotide exchange factor factor 5 (ARHGEF5) and insulin-like growth factor 2 mRNA binding protein 3 (IGF2BP3). Up-regulation of circ0003258 (Figure 4) was associated with tumor/ node/metastases (TNM) stage and tumor grade in prostate cancer patients (142). Circ003258 stimulated EMT in wound-healing and transwell migration assays in DU145 and PC3 cells and metastasis of DU145 cells after tail vein injection in nude mice. Circ 0003258 sponged miR-653-5p, resulting in up-regulation of ARHGEF5. Furthermore, circ0003258 interacted with IGF2BP3 to stabilize histone deacetylase 4 (HDAC4) mRNA. It has been shown that circ0003258 activated ERK signaling in prostate cancer cells (142). ARHGEF5 is a member of the RHO GTPase activating protein (RHOGAP) family and regulates small GTPase RHOA activity (143). IGF2BP3 functions as an RNA binding protein, which can stabilize selected mRNAs (144, 145). HDAC4 is one of 18 different HDACs that remove acetyl groups from histone and other non-histone proteins and which are de-regulated in prostate cancer (146, 147).

Circ midline 1 (circMID1) up-regulates YTH domain-containing protein 2 (YTHDC2). CircMID1 (Figure 4) was up-regulated in prostate cancer tissues and cell lines (148). Its down-regulation inhibited proliferation, migration, invasion, and glycolysis of DU145 and PC3 cells *in vitro* and the growth of SQ implanted PC3 cells in nude mice. CircMID1 sponged miR-330-3p and up-

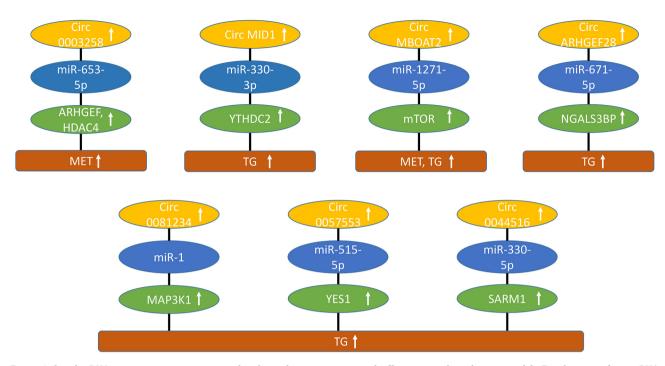


Figure 4. Circular RNAs targeting prostate cancer-related signaling components with efficacy in preclinical in vivo models. First line: specifies circRNA; second line: indicates miR interacting with corresponding circRNA; third line: high-lights de-regulated targets; fourth line: indicates impact on tumor growth and metastasis. Upward arrows indicate up-regulation, downward arrows indicate down-regulation. ARGEF5: RHO guanine nucleotide exchange factor 5; circARGEF28: circ RHO guanine nucleotide exchange factor 28; circ MBOAT2: circ membrane bound 0-acetyltransferase domain containing 2; circMID1: circ midline 1; IGF2BP3: insulin-like growth factor 2 mRNA binding protein 3; MAP3K1: mitogen-activated protein kinase kinase 1; MET: metastasis; miR: microRNA; mTOR: mechanistic target of rapamycin; NGALS3BP: galectin-3- binding protein; SARM1: sterile a and TIR motif-containing 1; TG: tumor growth; YES1: Yamaguchi sarcoma virus 1; YTHD2: YTH domain-containing protein 2.

regulated YTHDC2. YTHDC2stabilized m6A-modified IGF1R mRNA and induced AKT signaling (148). YTHDC2 functions as a m6A reader (149). It has been shown that YTHDC2 promotes growth, migration, and invasion of prostate cancer cells and its expression predicts poor outcome in prostate cancer patients (150). Furthermore, it was demonstrated that m6A RNA regulators contribute to the progression of prostate cancer (151).

Circ membrane bound O-acyltransferase domain containing 2 (circMBOAT2) up-regulates mechanistic target of rapamycin (mTOR). CircMBOAT2 (Figure 4) was associated with poor prognosis and progression of prostate cancer (152). It promoted proliferation and invasion of DU145 and PC3 prostate cancer cells *in vitro* as well as tumor growth and experimental metastasis of

PC3-derived xenografts in nude mice. CircMBOAT2 sponged miR-1271-5p and up-regulated mTOR, resulting in activation of PI3K/AKT signaling (152). The PI3K-AKT-mTOR pathway plays a role in the pro-survival resistance-mediating signaling of prostate cancer and cooperates with AR, MAPK, and wingless/integrated (WNT) signaling pathways (153-155).

CircRHO guanine nucleotide exchange factor 28 (circARHGEF28) up-regulates galectin-3-binding protein (NGALS3BP). CircARHGEF28 (Figure 4) was downregulated in prostate cancer tissues and cell lines (156). It inhibited proliferation, invasion, and migration of DU145 and PC3 prostate cancer cells *in vitro* and the growth of PC3 and 22Rv1 cells after SQ implantation in nude mice. CircARHGEF28 sponged miR-671-5p and up-regulated

NGALS3BP, inhibiting NF κ B signaling (156). NGALS3BP is a secreted, multifaceted protein composed of four domains, with seven potential N-linked glycosylation sites and three O-glycosylation sites. It inhibits activation mitogenactivated protein kinase kinase kinase 7 (MAP3K7), a negative regulator of NF κ B signaling (157, 158). It has been shown that NGALS3BP can bind to cluster of differentiation 33 (CD33)-related sialic acid-binding immunoglobulin-type of lectins (SIGLECs) to inhibit neutrophil activation (159).

Circ0081234 up-regulates mitogen-activated protein kinase kinase kinase 1 (MAP3K1). Circ0081234 (Figure 4) was found to be increased in spinal metastases compared to primary prostate cancer tissues (160). It promoted migration, invasion, and EMT in prostate cancer cells and its depletion inhibited the growth of prostate cancer xenografts in vivo. It was identified in exosomes and sponged miR-1, leading to up-regulation of MAP3K1 (160). MAP3K1 is a member of a family of 19 MAP3Ks and contains a kinase domain and a plant homeodomain motif, which functions as a ubiquitinligase (161). MAP3K1 is involved in the activation of prosurvival MAPKs (162). In prostate cancer, MAP3K1 activates AR-dependent transcription and survival (163).

Circ0057553 up-regulates proto-oncogene homolog of Yamaguchi sarcoma virus 1 (YES1). Circ 0057553 (Figure 4) was found to be up-regulated in prostate cancer tissues and cell lines (164). It promoted viability, migration, invasion, and glycolysis of DU145 and LNCaP cells in vitro and tumor growth of LNCaP cells SQ implanted into nude mice. It sponged miR-515-5p and up-regulated YES1 (164). YES1 belongs to the src family of tyrosine kinases, which control signaling pathways, cell proliferation, survival, invasiveness, and are amplified in several types of cancer (165). Several YES1 inhibitors are in clinical trials in cancer patients (165). It has been shown that YES1 mediates invasion and migration of prostate cancer cells (166) and increases their metastatic potential by phosphorylation of focal adhesion kinase (167). YES1 can also mediate PTX resistance in prostate cancer cells (168).

Circ0044516 up-regulates sterile α and TIR motif containing 1 (SARM1). Circ0044516 (Figure 4) was upregulated in prostate cancer tissues and cell lines (169). Its knockdown suppressed proliferation, migration, invasion of prostate cancer cells and inhibited growth of prostate cancer xenografts in nude mice. It sponged miR-330-5p with subsequent up-regulation of SARM1 (169). SARM1 has been identified as a multifunctional NADPase, which can function as an adaptor protein of toll-like receptors (TLRs), is highly expressed in neurons, and can induce axon degeneration and signaling (170, 171). In prostate cancer, SARM1 promotes proliferation, progression, and metastasis (172). The mechanistic underpinnings of these observations remain to be resolved.

Circular RNAs Modulating Expression of Human Antigen R (huR)

Circ formin 2 (circFMN2) interacts with huR and represses kruppel-like factor 2 (KLF2). CircFMN2 (Figure 5) was upregulated in prostate cancer tissues and tumor-derived extracellular vesicles and induced proliferation, invasion, and migration of prostate cancer cells (173). Extracellular vesicles from prostate cancer cells also accelerated tumor growth and metastasis of prostate cancer cells in nude mice. CircFMN2 interacted with huR (174), resulting in reduced interaction with KLF2 (173, 175) and reduced KLF2 expression. Also, binding of KLF2 to the promoter of E3-ubiquitinating enzyme RNF128 was decreased, leading to its attenuated expression. It has been shown that down-regulation of RNF128 activates β-catenin/WNT signaling (176). KLF2 also inhibits the migration of prostate cancer cells by downregulation of MMP2 (177). HuR represents a multifunctional RNA binding protein, which targets mRNAs coding for oncogenes, cytokines, growth factors, and metastasis-promoting factors (174).

Circ exocyst complex component 6B (Circ EXOC6B) upregulates A kinase anchoring protein 12 (AKAP12) with

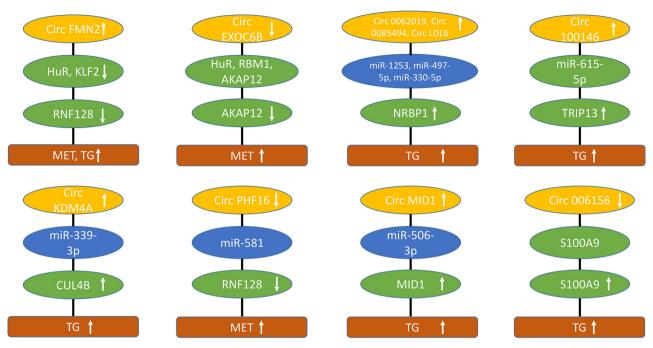


Figure 5. Circular RNAs targeting prostate cancer-related human R protein, nuclear receptor binding proteins, and ubiquitination pathway components with efficacy in preclinical in vivo models. First line: specifies circRNA; second line: indicates miRs or proteins interacting with corresponding circRNA; third line: highlights de-regulated targets; fourth line: indicates impact on tumor growth and metastasis. Upward arrows indicate up-regulation, downward arrows indicate down-regulation. AKAP12: A kinase anchoring protein 12; circEXO6B: circ exocyst complex component 6B; circFMN2: circ formin 2; circMID1: circ midline 1; circKDM4A: circ RNA lysine demethylase 4A; circPHF16: circ plant homeodomain finger protein 16; circLDL6: circ LDL receptor related protein 6; cul4B: cullin 4B; huR: human antigen R; KLF2: krüppel-like factor 2; MET: metastasis; miR: micro RNA; NRBP1: nuclear receptor binding protein 1; RNF128: ring finger protein 128; S100A9: S100 protein A9; TG: tumor growth; TRIP13: thyroid hormone receptor interacting protein 13.

the involvement of human antigen R (huR). Low circ EXOC6B (Figure 5) expression correlated with poor prognosis in prostate cancer patients compared to those with high EXOC6B expression (178). CircEXOC6B was down-regulated in prostate cancer tissues and inhibited migration and invasion of DU145 and PC3 cells. In nude mice, circEXOC6B decreased metastasis of DU145 cells after tail vein injection. Circ EXOC6B formed a ternary complex with RNA binding motif single-strand interacting protein (RBM1) (179) and AKAP12, resulting in destabilization of its mRNA (178). AKAP12 acts as a tumor suppressor gene, which can bind to protein kinase A (PKA) regulatory subunits, protein kinase C (PKC), and cytoskeletal elements, such as F-actin stress fibers, and inhibit metastasis (180-182).

Circular RNAs Affecting the Binding of Ligands to Nuclear Receptors

Circ0062019, circ0085494, and circ LDL receptor related protein 6 (circLDL6) up-regulate nuclear-receptor binding protein 1 (NRBP1). Circ0062019, circ0085494, and circLDL6 (Figure 5) were found to be up-regulated in prostate cancer tissues and cell lines (183-185). They sponged miRs-1253, -497-5p and -330-5p, respectively, resulting in up-regulation of NRBP1 (186). Down-regulation of each of these circRNAs inhibited proliferation, migration, invasion, and EMT of prostate cancer cells in vitro, whereas apoptosis was stimulated. In nude mice, down-regulation of these circRNAs inhibited growth of PC3 and DU145 prostate cancer cells after SQ implantation

(183-185). NRBP1 is a pseudokinase that functions as a multidomain adaptor protein, which is ubiquitously expressed and contains two nuclear receptor binding motifs, a putative interaction site for SH2-containing proteins, a kinase-like domain, a bipartite nuclear localization signal, and three sequences rich in glutamic acid, serine, proline, and threonine (PEST) (186), NRBP1. a 535 aa protein, is involved in mediating steroid, thyroid hormone, and vitamin-induced signaling pathways. NRBP1 has different functions in different types of cancers. In breast cancer, NRBP1 is down-regulated and inhibits proliferation through the WNT/β-catenin signaling pathway (187). In triple-negative breast cancer (TNBC), NRBP1 activates RAC1/CDC42 oncogenic signaling (188). In prostate cancer patients, NRBP1 is highly expressed and associated with increased cell proliferation and poor clinical outcomes (189). NRBP1 might be an interesting new target for the treatment of prostate cancer; however, the signaling pathways activated by NRBP1 have to be resolved in the context of target validation efforts.

Circ100146 up-regulates thyroid hormone-receptor interacting protein 13 (TRIP13). Circ100146 (Figure 5) was up-regulated in prostate cancer and its downregulation inhibited proliferation, migration, and invasion of 22Rv1 and DU145 cells in vitro (190). It sponged miR-615-5p and its down-regulation inhibited growth of 22Rv1 cells in nude mice after SQ implantation by up-regulation of TRIP13 (190). TRIP13 interacts with thyroid- and retinoid receptor, but not with glucocorticoid receptor (191). TRIP13 binds to transcriptional intermediary factor IB (TFIB), a central component of the complex of proteins necessary for transcriptional elongation (192). TRIP13 functions as an ATPase associated with diverse cellular activities (AAA+) and is involved in mitotic processes, spindle assembly checkpoint and DNA repair and may account for chromosomal instability (193). Amplification of TRIP13 has been observed in several types of cancer (193). It consists of 432 amino acids and harbors a N-terminal AAA+ substrate recognition region, which contains an

ATP binding site (193). TRIP13 is a predictor of poor prognosis in prostate cancer patients and mediates proliferation, migration, and invasion of prostate cancer cells (194). Further delineation of the TRIP13-mediated molecular interactions and identification of the pathways induced by TRIP13 will be necessary for further target validation in prostate cancer.

Circular RNAs Affecting Ubiquitination

Circ RNA lysine demethylase 4A (circKDM4A) up-regulates cullin 4B (cul4B). CircKDM4A (Figure 5) was up-regulated in serum exosomes from prostate cancer patients compared to exosomes from healthy volunteers (195). CircKDM4A promoted proliferation, migration, invasion, and inhibited apoptosis of prostate cancer cells. In nude mice, silencing of circKDM4A inhibited tumor growth of prostate cancer xenografts. Circ KDM4A sponged miR-339-3p and induced cul4B (195). The latter acts as a scaffold of the cul4B-ring E3 ligase complex, which is overexpressed in many types of tumors, regulates proteolysis, and inhibits expression of tumor suppressor genes (196). In prostate cancer, cul4B has been shown to activate the PI3K/AKT/mTOR pathway (197, 198).

Circ plant homeodomain finger protein 16 (circPHF16) upregulates ring finger protein 128 (RNF128). CircPHF16 (Figure 5) was down-regulated in prostate cancer tissues compared to corresponding normal tissues (199). In transwell and wound-healing assays, circPHF16 inhibited migration and invasion of DU145 and PC3 cells. CircPHF16 decreased metastasis of PC3 cells after tail-vein injection into nude mice (199). CircPHF16 sponged miR-581, increasing RNF128, which inhibited WNT/β-catenin signaling and EMT by targeting β -catenin (199). It has been shown that down-regulation of RNF128 activates WNT/β-catenin signaling and EMT in melanoma (200). In colorectal cancer (CRC) and hepatocellular carcinoma (HCC), RNF128 suppresses PI3K/AKT, WNT/β-catenin, and EGFR/mitogen activated kinase kinase (MEK)/ERK pathways (201, 202).

Circ midline 1 (circMID1) up-regulates MID1. CircMID1 (Figure 5) was highly expressed in prostate cancer tissues compared to benign prostate hyperplasia tissues, and in CRPC compared to AR-dependent prostate cancer (203). PC3 cells treated with exosomes derived from myeloid-derived suppressor cells up-regulated circMID1. circMID1 promoted proliferation, migration, and progression to CRPC in vitro and in nude mice. CircMID1 sponged miR-506-3p and up-regulated MID1 (203). The latter is involved in the microtubule-associated protein complex and functions as an E3-ubiquitin ligase. It is part of the tripartite motif (TRIM) subfamily of RING containing proteins and is also known as TRIM18 (204). It has been shown that MID1 is up-regulated during androgen-deprivation therapy and enhanced AKT, NFκB, and hedgehog (Hh) signaling, promoting castration resistance (205, 206).

Circ006156 blocks ubiquitinylation of S100 protein A9 (S100A9). Low levels of circ 006156 (Figure 5) were detected in prostate cancer tissues and cell lines (207). Circ006156 suppressed migration and invasion of DU145 and PC3 cells in vitro and inhibited experimental metastasis of PC3 cells to the lungs in nude mice due to up-regulation of S100A9. Circ006156 bound to S100A9 and inhibited its ubiquitinylation (207). It has been shown that S100A9 promotes growth of prostate cancer invasion by activating TLR4/NF κ B/integrin β 1/focal adhesion kinase (FAK) signaling (208). Also, S100A9 has been reported as a potential diagnostic marker for early prostate cancer, as it can distinguish between prostate cancer and benign hyperplasia, because serum levels of S100A9 are increased in prostate cancer patients (209).

Circular RNAs Affecting Metabolism

Circ phosphofructose kinase platelet (circPFKP) up-regulates inosine-5'-monophosphate dehydrogenase 2 (IMPDH2). CircPFKP (Figure 6) was increased in prostate cancer tissues compared with adjacent non-cancerous prostate tissues and expression correlated with tumor stage (210).

It promoted proliferation of prostate cancer cell *in vitro* and in nude mice after SQ implantation. CircPFKP interacted with IMPDH2 and mediated biogenesis of GTP (210, 211). IMPDH2 was identified as a marker for aggressive and advanced prostate cancer (212). Silencing of IMPDH2 reduces prostate cancer cell proliferation (213). In prostate cancer patients, enhanced expression of IMPDH2 has been shown to promote metastasis (214).

Circ pyruvate dehydrogenase complex component X (circPDHX) up-regulates acetyl-CoA synthetase long chain family member 1 (ACSL1). Circ PDHX (Figure 6) was up-regulated in prostate cancer tissues and its inhibition attenuated prostate cancer cell proliferation, migration, fatty acid metabolism, increased apoptosis, and repressed tumor growth in nude mice. It sponged miR-497-5p and up-regulated ACSL1 (215). ACSL1 is part of an enzyme family, which is commonly deregulated in cancer and is associated with poor survival (216, 217). All iso-enzymes convert free long-chain fatty acids into fatty acid acyl-CoA esters and thereby play a role in lipid biosynthesis and fatty acid degradation. They promote ungoverned cell growth, facilitate tumor invasion, and inhibit apoptosis (216, 218). ACSL1 has been shown to promote prostate cancer progression by increasing lipogenesis and fatty acid β -oxidation (218, 219).

Circ M6A RNA binding motif 33 (circ M6A RBM33) upregulates pyruvate dehydrogenase, subunit α (PDHA1). Expression of circ M6A RBM33 (Figure 6) predicted poor prognosis in prostate cancer patients (220). Silencing of circ M6A RBM33 inhibited proliferation and invasion of 22Rv1 and DU145 cells *in vitro* and decreased tumor growth of corresponding xenografts in nude mice. Depletion of circ M6A RBM33 increased sensitivity to AR inhibitors ENZ and darolutamide. Circ M6A RBM33 interacted with fragile mental retardation protein 1 (FMR1), which stabilized PDHA1 (220). FMR1 is an RNA binding protein that mediates stabilization of mRNA and enhances translation (221, 222). PDHA1 is involved in

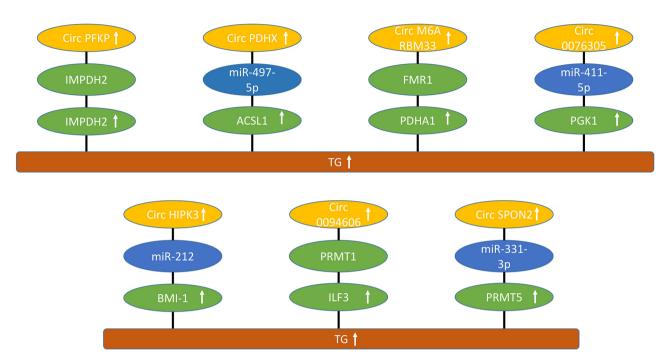


Figure 6. Circular RNAs targeting prostate cancer-related components of metabolism and epigenetic modification with efficacy in preclinical in vivo models. First line: specifies circRNA; second line: indicates miRs or proteins interacting with corresponding circRNA; third line: highlights de-regulated targets; fourth line: indicates impact on tumor growth and metastasis. Upward arrows indicate up-regulation, downward arrows indicate down-regulation.ACSL1: Acetyl-CoA-synthase long chain family member 1; BMI-1: polycomb complex protein 1; circHIPK3: circ homeodomain interacting protein kinase 3; circPDHX: circ pyruvate dehydrogenase complex component X; circPFKP: circ phosphofructo-kinase platelet; circ M6A RBM33: circ M6B RNA binding motif 33; circSPON2: circ spondin 2; FMR1: fragile mental retardation protein 1; ILF3: interleukin enhancer binding factor 3; IMPDH2: inosine-5'- monophosphate dehydrogenase 2; miR: micro RNA; PDHA1: pyruvate dehydrogenase A1, subunit α; PGK1: phosphoglycerate kinase 1; PRMT5: protein methyltransferase 5; TG: tumor growth.

mitochondrial respiration, pyruvate oxidation, and glycolysis (223, 224).

Circ0076305 up-regulates phosphoglycerate kinase 1 (PGK1). Circ0076305 (Figure 6) was highly expressed in prostate cancer tissues and its silencing triggered cell growth, migration, glycolysis, and induced apoptosis in prostate cancer. Interference with circ0076305 inhibited prostate cancer xenografts in nude mice. Circ0076305 sponged miR-411-5p and up-regulated PGK1 (225). PGK1 is a metabolic enzyme that is associated with poor prognosis, tumor growth, migration, and treatment resistance through phosphorylation of substrates (226, 227). PGK1 facilitates prostate cancer progression (226), metastasis (228) and prostate cancer induced bone formation (229).

Circular RNAs Affecting Epigenetic Modification

Circ homeodomain interacting protein kinase 3 (circHIPK3) up-regulates polycomb complex protein BMI-1. Exosomal circHIPK3 (Figure 6) was increased in the serum of prostate cancer patients (230). Exosomal circHIPK3 knockdown inhibited proliferation, migration, and invasion of 22Rv1 and DU145 cells *in vitro*. In nude mice, exosomal circ HIPK3 hampered growth of NB prostate cancer cells after SQ implantation. CircHIPK3 sponged miR-212 and up-regulated BMI-1 (230). The latter is part of the polycomb repressive complex 1 (PRC1), which inhibits transcription by trimethylation of histone H3 at lys 27 and subsequent inhibition of the RNA polymerase

pre-initiation complex by limiting the access of transcription factors to chromatin (230, 231). BMI-1 contains a helix-turn helix domain that binds to DNA, and a ring finger domain, which interacts with diverse substrates (230, 231). In prostate cancer, it has been shown that BMI-1 regulates stem cell renewal, malignant transformation, proliferation, migration, and chemoresistance (232-234).

Circ0094606 up-regulates interleukin enhancer binding factor 3 (ILF3). Circ0094606 (Figure 6) promoted proliferation, migration and EMT of prostate cancer cells and increased their growth in nude mice (235). It binds to protein arginine methyltransferase 1 (PRMT1), which methylates ILF3, a protein which stabilizes interleukin 8 (IL8) mRNA, increases the expression of IL8, and induces polarization of macrophages to the tumorigenic M2 type (235). PRMTs are involved in the epigenetic regulation of gene expression by methylating histone proteins, but they also methylate non-histone proteins (236). ILF3 has been shown to act as an RNA binding protein and mediates cytokine induced angiogenesis (237). In prostate cancer, PMRT1 has been shown to regulate AR-mediated signaling (238). Furthermore, PRMT1 functions as an epigenetic driver of prostate cancer progression (239).

Circ spondin 2 (circSPON2) up-regulates protein methyltransferase 5 (PRMT5). CircSPON2 (Figure 6) was up-regulated in prostate cancer patients and promoted proliferation of DU145 and PC3 cells in vitro and stimulated growth of PC3 xenografts in nude mice (240). It sponged miR-331-3p and up-regulated PRMT5. PRMT5is up-regulated in prostate cancer and correlated with poor progression-free survival (240). It repressed expression of calcium/calmodulin dependent protein kinase II inhibitor 1 (CAMK2N1) (240, 241). PRMT5 inhibits the transcription of tumor suppressor genes by symmetric methylation of histone 3 at lys8, and histone 4 at lys 3 (13). CAMK2N1 functions as a suppressor of prostate cancer by inhibition of AR-dependent transcription (242). Furthermore, it has been shown that

down-regulation of CAMK2N1 due to DNA methylation promotes progression of prostate cancer (243).

Circular RNAs Affecting Proteins of Other Categories

Circ sine oculis binding protein homolog (circSOBP) upregulates myosin-light chain phosphatase1 (MYPT1). CircSOBP (Figure 7) was down-regulated in prostate cancer tissues and inhibited migration, invasion of DU145 and PC3 in vitro and experimental metastasis of DU145 cells in nude mice. It sponged miR-141-3p and upregulated MYPT1 (244). The latter inhibited phospho-myosin light chain 2 (pMLC2), a mediator of invasion and metastasis (245). Myosin represents a motor protein, which is involved in muscle contraction and ATP-dependent actin based motility together with tropomyosin and troponin (245). MYPT1 is serine-threonine specific phosphatase, which dephosphorylates the light chain of myosin, which consists of three subunits (246).

Circ0006404 up-regulates cofilin 2 (CFL2). Circ 0006404 (Figure 7) was aberrantly up-regulated in prostate cancer (247). Interference with circ006404 inhibited proliferation and invasion of DU145 and LNCaP-AI cells, induced apoptosis *in vitro* and decreased tumor growth of DU145 xenografts in nude mice. Circ0006404 sponged miR-1299 and up-regulated CFL2, a protein involved in actin depolymerization (247, 248). CFL2 drives the cell invasive and metastatic properties of TGF β in prostate cancer (249). Over-expression of its paralog, CFL1, correlates with aggressiveness in prostate cancer (250).

Circ0004296 binds to eucaryotic translation initiation factor 4A (EIF4A). Circ0004296 (Figure 7) was decreased in prostate cancer tissues, the blood and urine in comparison to corresponding samples derived from healthy donors (251). It inhibited proliferation, invasion, migration, and EMT of DU145 and PC3 cells *in vitro*. Inhibition of circ00044296 decreased tumor growth after SQ and orthotopic implantation in nude mice as well as metastasis

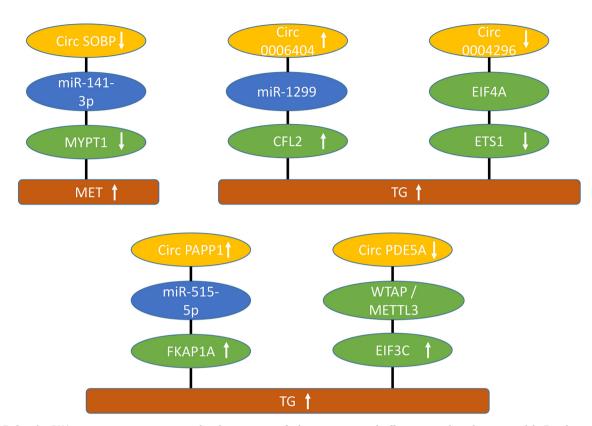


Figure 7. Circular RNAs targeting prostate cancer-related components of other categories with efficacy in preclinical in vivo models. First line: specifies circRNA; second line: indicates miRs or proteins interacting with corresponding circRNA; third line: highlights de-regulated targets; fourth line: indicates impact on tumor growth and metastasis. Upward arrows indicate up-regulation, downward arrows indicate down-regulation. CFL2: Cofilin 2; circPAPP1: circ pappalysin 1; circPDE5A: circ phosphodiesterase 5A; circSOBP: circ sine oculis binding protein homolog; FKAP1A: FKAP prolylisomerase 1A; EIF3C: eukaryotic initiation factor 3C; EIF4A: eucaryotic initiation factor 4A; ETS1: transcription factor ETS1; MET: metastasis; METTL3: M6-adenosinmethyltransferase 3; miR: micro RNA; MYPT1: myosin-light chain phosphatase 1; TG: tumor growth; WTAP: WT1 associated protein splicing regulator.

to the lungs after tail vein injection. Circ 0004296 bound to EIF4A and inhibited nuclear export of transcription factor ETS1 mRNA (251). EIF4A, a member of the DEAD-box family of proteins was shown to be involved in oncogenesis as a mediator of proliferation and cell cycle progression (252, 253). In addition, EIF4A can facilitate nucleo-cytoplasmic transport (254). ETS1 induces $TGF\beta$ signaling and promotes EMT in prostate cancer (255). ETS1 is also increased in advanced prostate cancer and induces the CRPC phenotype (256).

Circ pappalysin 1 (circPAPP1) up-regulates FKAP prolylisomerase 1A (FKAP1A). CircPAPP1 (Figure 7) was

over-expressed in prostate cancer and its knockdown inhibited cell viability, proliferation, and glycolysis, while restricting tumor growth of prostate cancer xenografts in nude mice (257). CircPAPP1 sequestered miR-515-5p and up-regulated FKAP1A, a member of the binding proteins (257).FK506 These multifunctional proteins that are involved mTOR/AKT signaling and the modification of chromatin structure due to their isomerase activity (258, 259). FKAPs can act as oncogenes as well as tumor suppressors in a context-dependent manner (258, 259). FKAP1A was shown to act as an oncogene in prostate cancer (260).

Circ phosphodiesterase 5A (circPDE5A) inhibits eucaryotic initiation factor 3C (EIF3C). CircPDE5A (Figure 7) was down-regulated in prostate cancer tissues compared to adjacent normal tissues (261). Over-expression of circPDE5A in C4-B and 22Rv-1 cells inhibited migration and invasion in vitro. 22Rv-1 cells transfected with circPDE5A led to decreased lung metastasis in an experimental metastasis model in nude mice. CircPDE5A formed a complex with WT1 associated protein splicing regulator (WTAP) and blocked N6A modification of EIF3C mRNA leading to disruption of its translation and inhibition of MAPK signaling (261). It has been shown that WTAP exerts N6A methylation by binding to N6methyltransferase 3 (METTL3) (262). EIF3C activates PI3K, AKT, NFκB, and c-MYC in PC3 prostate cancer cells (263). In addition, it has been demonstrated that N6A modification of mRNAs contributes to progression of prostate cancer (152, 264).

Technical and Disease-related Issues

Nucleic acid (NA)-related therapeutic agents can be categorized into ASO (12-30 nts), siRNAs (21-23 nts), miRs (20-24 nts), small activating RNAs (21 nts), RNA aptamers (20-100 nts), and CRISPR-CAS based agents using guide RNA (265). Corresponding agents have been approved for indications such as transerythrin-mediated amyloidosis, Duchenne muscular dystrophy, and hypercholesterolemia (266). mRNA-based agents have received great attention for vaccination against viruses and cancer (266). Regarding tackling of circRNAs in cancer with ASO, siRNA, or CRISPR-CAS related agents, several issues have to be addressed and optimized: stability, off-targets effects, immunogenicity, as well as pharmaco-kinetic and pharmaco-dynamic properties (267, 268). Polymer- or lipid-based nanoparticles and also inorganic-based delivery vehicles have contributed to delivery of NA-related agents (269). Liver delivery through the asialoglycoprotein receptor, targeting of the lungs by varying the composition of the lipids, and targeting of the blood-brain barrier are examples of recent achievements (270). Also, homing of different cell-types with designated peptides identified by phage-display related approaches have great potential with respect to delivery issues (271). The most important topic in the prostate cancer field is the treatment of castration-resistant and metastatic disease. Molecular mechanisms of resistance involve AR amplification, AR mutations, mutations in co-activators/co-repressors, androgen-independent AR activation, and intra-tumoral and alternative androgen production (272). Several internalizing receptors are potential targets for treatment of advanced prostate cancer such as: prostate-specific membrane antigen (PSMA), trophoblast surface antigen 2 (TROP2), small transmembrane epithelial antigen of the prostate 1 (STEAP1), tissue factor (TF), delta like protein 3 (DLL3), HER2, and B7 homolog 3 (B7H3) (8).

Concluding Remarks

We identified 49 de-regulated circRNAs with efficacy in preclinical prostate cancer-related in vivo models. These circRNAs belong to the following categories: therapyresistance (n=8), transmembrane and secreted (n=7), transcription factors (n=8), signaling (n=7), human antigen R (n=2), nuclear receptor binding (n=2), ubiquitination (n=4), metabolism (n=4), epigenetics (n=3), and other functions (n=5). Forty of these circRNAs exert their functions by binding to miRs, while nine bind to proteins and mediate functions such as ubiquitination, formation of scaffolding complexes and mRNAstabilization, -modification and -transport. From a clinical perspective, circRNAs and their corresponding targets associated with treatment resistance, CRPC, and metastasis are of particular importance. Notable findings include circXIAP and circSFBT2, which confer paclitaxel (PTX) resistance by targeting TPD52 and TRIB1, respectively. Similarly, circZNF609 and circZEB1, associated with radiation resistance, target HK2 and ZEB1, respectively (Figure 1). CircRNA17 which is involved in ENZ-R by targeting ARV7 has been identified as an important regulator of CRPC (Figure 1). In the category of CRPC-related circRNAs, circ0092339 was identified as an

inducer of c-MYC (Figure 3), and circMID1 (Figure 5) as an up-regulator of the ubiquitination enzyme MID1. Furthermore, nine circRNAs affect tumor growth and metastasis, or metastasis only. The up-regulated circRNAs of this category are circSMARCC (CCL6) (Figure 2), circ003258 (ARHGEF) (Figure 4), circMBOAT2 (mTOR) (Figure 4), and circFMN2 (KLF2, RNF128) (Figure 5). The down-regulated circRNAs include circ EXOC6B (AKAP12) (Figure 5), circPHF16 (RNF128) (Figure 5), circ006156 (S100A9) (Figure 5), circSOBP (MYPT1) (Figure 7) and circPDE5A (EIF3C) (Figure 7). The ranking of the identified circRNAs and their corresponding targets for the treatment of prostate cancer will require further validation experiments.

Conflicts of Interest

FB is and UHW was an employee of Roche.

Authors' Contributions

FB and UHW equally contributed to all aspects of the paper.

References

- Siegel RL, Miller KD, Wagle NS, Jemal A: Cancer statistics, 2023. CA Cancer J Clin 73(1): 17-48, 2023. DOI: 10.3322/ caac.21763
- 2 Sekhoacha M, Riet K, Motloung P, Gumenku L, Adegoke A, Mashele S: Prostate cancer review: genetics, diagnosis, treatment options, and alternative approaches. Molecules 27(17): 5730, 2022. DOI: 10.3390/molecules27175730
- 3 Hatano K, Nonomura N: Systemic therapies for metastatic castration-resistant prostate cancer: an updated review. World J Mens Health 41(4): 769-784, 2023. DOI: 10.5534/wjmh.220200
- 4 Marchetti A, Tassinari E, Rosellini M, Rizzo A, Massari F, Mollica V: Prostate cancer and novel pharmacological treatment options-what's new for 2022? Expert Rev Clin Pharmacol 16(3): 231-244, 2023. DOI: 10.1080/17512433.2023.2181783
- Marei HE, Hasan A, Pozzoli G, Cenciarelli C: Cancer immunotherapy with immune checkpoint inhibitors (ICIs): potential, mechanisms of resistance, and strategies for reinvigorating T cell responsiveness when resistance is acquired. Cancer Cell Int 23(1): 64, 2023. DOI: 10.1186/s12935-023-02902-0

- 6 Sridaran D, Bradshaw E, DeSelm C, Pachynski R, Mahajan K, Mahajan NP: Prostate cancer immunotherapy: Improving clinical outcomes with a multi-pronged approach. Cell Rep Med 4(10): 101199, 2023. DOI: 10.1016/j.xcrm.2023.101199
- 7 Zarrabi KK, Narayan V, Mille PJ, Zibelman MR, Miron B, Bashir B, Kelly WK: Bispecific PSMA antibodies and CAR-T in metastatic castration-resistant prostate cancer. Ther Adv Urol 15: 17562872231182219, 2023. DOI: 10.1177/1756287 2231182219
- 8 Sardinha M, Palma Dos Reis AF, Barreira JV, Fontes Sousa M, Pacey S, Luz R: Antibody-drug conjugates in prostate cancer: a systematic review. Cureus 15(2): e34490, 2023. DOI: 10.7759/cureus.34490
- 9 Yedla P, Babalghith AO, Andra VV, Syed R: PROTACs in the management of prostate cancer. Molecules 28(9): 3968, 2023. DOI: 10.3390/molecules28093698
- 10 Avgeris I, Pliatsika D, Nikolaropoulos SS, Fousteris MA: Targeting androgen receptor for prostate cancer therapy: From small molecules to PROTACs. Bioorg Chem 128: 106089, 2022. DOI: 10.1016/j.bioorg.2022.106089
- 11 Dong Y, Xu S, Liu J, Ponnusamy M, Zhao Y, Zhang Y, Wang Q, Li P, Wang K: Non-coding RNA-linked epigenetic regulation in cardiac hypertrophy. Int J Biol Sci 14(9): 1133-1141, 2018. DOI: 10.7150/ijbs.26215
- 12 Szczepaniak A, Bronisz A, Godlewski J: Circular RNAs-new kids on the block in cancer pathophysiology and management. Cells 12(4): 552, 2023. DOI: 10.3390/cells 12040552
- 13 Sanger HL, Klotz G, Riesner D, Gross HJ, Kleinschmidt AK: Viroids are single-stranded covalently closed circular RNA molecules existing as highly base-paired rod-like structures. Proc Natl Acad Sci USA 73(11): 3852-3856, 1976. DOI: 10.1073/pnas.73.11.3852
- 14 Wang KS, Choo QL, Weiner AJ, Ou JH, Najarian RC, Thayer RM, Mullenbach GT, Denniston KJ, Gerin JL, Houghton M: Structure, sequence and expression of the hepatitis delta (δ) viral genome. Nature 323(6088): 508-514, 1986. DOI: 10.1038/323508a0
- 15 Jeck WR, Sharpless NE: Detecting and characterizing circular RNAs. Nat Biotechnol 32(5): 453-461, 2014. DOI: 10.1038/ nbt.2890
- 16 Glažar P, Papavasileiou P, Rajewsky N: circBase: a database for circular RNAs. RNA 20(11): 1666-1670, 2014. DOI: 10.1261/rna.043687.113
- 17 Kim WR, Park EG, Lee DH, Lee YJ, Bae WH, Kim HS: The tumorigenic role of circular RNA-microRNA axis in cancer. Int J Mol Sci 24(3): 3050, 2023. DOI: 10.3390/ijms24033050
- 18 Wang H, Meng Q, Qian J, Li M, Gu C, Yang Y: Review: RNA-based diagnostic markers discovery and therapeutic targets development in cancer. Pharmacol Ther 234: 108123, 2022. DOI: 10.1016/j.pharmthera.2022.108123
- 19 Weidle UH, Hsia HE, Brinkmann U: Breast cancer: Circular RNAs mediating efficacy in preclinical *in vivo* models. Cancer

- Genomics Proteomics 20(3): 222-238, 2023. DOI: 10.21873/cgp.20377
- 20 Yang J, Meng X, Pan J, Jiang N, Zhou C, Wu Z, Gong Z: CRISPR/Cas9-mediated noncoding RNA editing in human cancers. RNA Biol 15(1): 35-43, 2018. DOI: 10.1080/ 15476286.2017.1391443
- 21 Weidle UH, Sela T, Brinkmann U, Niewoehner J: Circular RNAs with efficacy in preclinical *in vitro* and *in vivo* models of esophageal squamous cell carcinoma. Cancer Genomics Proteomics 19(3): 283-298, 2022. DOI: 10.21873/cgp. 20320
- 22 Tucker D, Zheng W, Zhang DH, Dong X: Circular RNA and its potential as prostate cancer biomarkers. World J Clin Oncol 11(8): 563-572, 2020. DOI: 10.5306/wjco.v11.i8.563
- 23 Chao F, Wang S, Zhang C, Han D, Xu G, Chen G: The emerging role of circular RNAs in prostate cancer: a systematic review. Front Cell Dev Biol 9: 681163, 2021. DOI: 10.3389/fcell. 2021.681163
- 24 Liu X, Tong Y, Xia D, Peng E, Yang X, Liu H, Ye T, Wang X, He Y, Ye Z, Chen Z, Tang K: Circular RNAs in prostate cancer: Biogenesis, biological functions, and clinical significance. Mol Ther Nucleic Acids 26: 1130-1147, 2021. DOI: 10.1016/j.omtn.2021.10.017
- 25 Chen L, Song Y, Hou T, Li X, Cheng L, Li Y, Xing Y: Circ_0004087 interaction with SND1 promotes docetaxel resistance in prostate cancer by boosting the mitosis error correction mechanism. J Exp Clin Cancer Res 41(1): 194, 2022. DOI: 10.1186/s13046-022-02404-3
- 26 Chidambaranathan-Reghupaty S, Mendoza R, Fisher PB, Sarkar D: The multifaceted oncogene SND1 in cancer: focus on hepatocellular carcinoma. Hepatoma Res 4: 32, 2018. DOI: 10.20517/2394-5079.2018.34
- 27 Cappellari M, Bielli P, Paronetto MP, Ciccosanti F, Fimia GM, Saarikettu J, Silvennoinen O, Sette C: The transcriptional coactivator SND1 is a novel regulator of alternative splicing in prostate cancer cells. Oncogene 33(29): 3794-3802, 2014. DOI: 10.1038/onc.2013.360
- 28 Kim T, Gartner A: Bub1 kinase in the regulation of mitosis. Anim Cells Syst (Seoul) 25(1): 1-10, 2021. DOI: 10.1080/ 19768354.2021.1884599
- 29 Martinez MJ, Lyles RDZ, Peinetti N, Grunfeld AM, Burnstein KL: Inhibition of the serine/threonine kinase BUB1 reverses taxane resistance in prostate cancer. iScience 26(9): 107681, 2023. DOI: 10.1016/j.isci.2023.107681
- 30 Zhang H, Li M, Zhang J, Shen Y, Gui Q: Exosomal Circ-XIAP promotes docetaxel resistance in prostate cancer by regulating miR-1182/TPD52 axis. Drug Des Devel Ther 15: 1835-1849, 2021. DOI: 10.2147/DDDT.S300376
- 31 Tennstedt P, Bölch C, Strobel G, Minner S, Burkhardt L, Grob T, Masser S, Sauter G, Schlomm T, Simon R: Patterns of TPD52 overexpression in multiple human solid tumor types analyzed by quantitative PCR. Int J Oncol 44(2): 609-615, 2014. DOI: 10.3892/ijo.2013.2200

- 32 Rubin MA, Varambally S, Beroukhim R, Tomlins SA, Rhodes DR, Paris PL, Hofer MD, Storz-Schweizer M, Kuefer R, Fletcher JA, Hsi B, Byrne JA, Pienta KJ, Collins C, Sellers WR, Chinnaiyan AM: Overexpression, amplification, and androgen regulation of TPD52 in prostate cancer. Cancer Res 64(11): 3814-3822, 2004. DOI: 10.1158/0008-5472.CAN-03-3881
- 33 Dasari C, Yaghnam DP, Walther R, Ummanni R: Tumor protein D52 (isoform 3) contributes to prostate cancer cell growth *via* targeting nuclear factor-κB transactivation in LNCaP cells. Tumour Biol 39(5): 101042831769838, 2017. DOI: 10.1177/1010428317698382
- 34 Wang R, Xu J, Mabjeesh N, Zhu G, Zhou J, Amin M, He D, Marshall FF, Zhau HE, Chung LW: PrLZ is expressed in normal prostate development and in human prostate cancer progression. Clin Cancer Res 13(20): 6040-6048, 2007. DOI: 10.1158/1078-0432.CCR-07-0640
- 35 Zhang H, Wang J, Pang B, Liang RX, Li S, Huang PT, Wang R, Chung LW, Zhau HE, Huang C, Zhou JG: PC-1/PrLZ contributes to malignant progression in prostate cancer. Cancer Res 67(18): 8906-8913, 2007. DOI: 10.1158/0008-5472.CAN-06-4214
- 36 Tan X, Song X, Fan B, Li M, Zhang A, Pei L: Exosomal circRNA Scm-like with four malignant brain tumor domains 2 (circ-SFMBT2) enhances the docetaxel resistance of prostate cancer *via* the microRNA-136-5p/tribbles homolog 1 pathway. Anticancer Drugs 33(9): 871-882, 2022. DOI: 10.1097/CAD.0000000000001365
- 37 Eyers PA, Keeshan K, Kannan N: Tribbles in the 21st century: the evolving roles of tribbles pseudokinases in biology and disease. Trends Cell Biol 27(4): 284-298, 2017. DOI: 10.1016/j.tcb.2016.11.002
- 38 Shahrouzi P, Astobiza I, Cortazar AR, Torrano V, Macchia A, Flores JM, Niespolo C, Mendizabal I, Caloto R, Ercilla A, Camacho L, Arreal L, Bizkarguenaga M, Martinez-Chantar ML, Bustelo XR, Berra E, Kiss-Toth E, Velasco G, Zabala-Letona A, Carracedo A: Genomic and functional regulation of TRIB1 contributes to prostate cancer pathogenesis. Cancers (Basel) 12(9): 2593, 2020. DOI: 10.3390/cancers12092593
- 39 Mashima T, Soma-Nagae T, Migita T, Kinoshita R, Iwamoto A, Yuasa T, Yonese J, Ishikawa Y, Seimiya H: TRIB1 supports prostate tumorigenesis and tumor-propagating cell survival by regulation of endoplasmic reticulum chaperone expression. Cancer Res 74(17): 4888-4897, 2014. DOI: 10.1158/0008-5472.CAN-13-3718
- 40 Gao Y, Liu J, Huan J, Che F: Downregulation of circular RNA hsa_circ_0000735 boosts prostate cancer sensitivity to docetaxel *via* sponging miR-7. Cancer Cell Int 20: 334, 2020. DOI: 10.1186/s12935-020-01421-6
- 41 Guo Q, Nan XX, Yang JR, Yi L, Liang BL, Wei YB, Zhu N, Hu SB, Zhang H, Luo Y, Xu YF: Triptolide inhibits the multidrug resistance in prostate cancer cells *via* the downregulation of MDR1 expression. Neoplasma 60(06): 598-604, 2014. DOI: 10.4149/neo_2013_077

- 42 Dulińska-Litewka J, Felkle D, Dykas K, Handziuk Z, Krzysztofik M, Gąsiorkiewicz B: The role of cyclins in the development and progression of prostate cancer. Biomed Pharmacother 155: 113742, 2022. DOI: 10.1016/j.biopha.2022.113742
- 43 Wang G, Zhao D, Spring DJ, DePinho RA: Genetics and biology of prostate cancer. Genes Dev 32(17-18): 1105-1140, 2018. DOI: 10.1101/gad.315739.118
- 44 Wu G, Sun Y, Xiang Z, Wang K, Liu B, Xiao G, Niu Y, Wu D, Chang C: Preclinical study using circular RNA 17 and micro RNA 181c-5p to suppress the enzalutamide-resistant prostate cancer progression. Cell Death Dis 10(2): 37, 2019. DOI: 10.1038/s41419-018-1048-1
- 45 Zheng Z, Li J, Liu Y, Shi Z, Xuan Z, Yang K, Xu C, Bai Y, Fu M, Xiao Q, Sun H, Shao C: The crucial role of AR-V7 in enzalutamide-resistance of castration-resistant prostate cancer. Cancers (Basel) 14(19): 4877, 2022. DOI: 10.3390/cancers14194877
- 46 Antonarakis ES, Lu C, Wang H, Luber B, Nakazawa M, Roeser JC, Chen Y, Mohammad TA, Chen Y, Fedor HL, Lotan TL, Zheng Q, De Marzo AM, Isaacs JT, Isaacs WB, Nadal R, Paller CJ, Denmeade SR, Carducci MA, Eisenberger MA, Luo J: AR-V7 and resistance to enzalutamide and abiraterone in prostate cancer. N Engl J Med 371(11): 1028-1038, 2014. DOI: 10.1056/NEJMoa1315815
- 47 Zhou T, Wang S, Song X, Liu W, Dong F, Huo Y, Zou R, Wang C, Zhang S, Liu W, Sun G, Lin L, Zeng K, Dong X, Guo Q, Yi F, Wang Z, Li X, Jiang B, Cao L, Zhao Y: RNF8 up-regulates AR/ARV7 action to contribute to advanced prostate cancer progression. Cell Death Dis 13(4): 352, 2022. DOI: 10.1038/s41419-022-04787-9
- 48 Yu B, Liu Y, Luo H, Fu J, Li Y, Shao C: Androgen receptor splicing variant 7 (ARV7) inhibits docetaxel sensitivity by inactivating the spindle assembly checkpoint. J Biol Chem 296: 100276, 2021. DOI: 10.1016/j.jbc.2021.100276
- 49 Du S, Zhang P, Ren W, Yang F, Du C: Circ-ZNF609 accelerates the radioresistance of prostate cancer cells by promoting the glycolytic metabolism through miR-501-3p/HK2 axis. Cancer Manag Res 12: 7487-7499, 2020. DOI: 10.2147/CMAR. S257441
- 50 Fan L, Huang C, Li J, Gao T, Lin Z, Yao T: Long non-coding RNA urothelial cancer associated 1 regulates radioresistance via the hexokinase 2/glycolytic pathway in cervical cancer. Int J Mol Med 42(4): 2247-2259, 2018. DOI: 10.3892/ijmm. 2018.3778
- 51 Zheng Y, Zhan Y, Zhang Y, Zhang Y, Liu Y, Xie Y, Sun Y, Qian J, Ding Y, Ding Y, Fang Y: Hexokinase 2 confers radio-resistance in hepatocellular carcinoma by promoting autophagy-dependent degradation of AIMP2. Cell Death Dis 14(8): 488, 2023. DOI: 10.1038/s41419-023-06009-2
- 52 Zhong JT, Zhou SH: Warburg effect, hexokinase-II, and radioresistance of laryngeal carcinoma. Oncotarget 8(8): 14133-14146, 2017. DOI: 10.18632/oncotarget.13044
- 53 Cai F, Li J, Zhang J, Huang S: Knockdown of Circ_CCNB2 sensitizes prostate cancer to radiation through repressing

- autophagy by the miR-30b-5p/KIF18A axis. Cancer Biother Radiopharm 37(6): 480-493, 2022. DOI: 10.1089/cbr. 2019.3538
- 54 Rath O, Kozielski F: Kinesins and cancer. Nat Rev Cancer 12(8): 527-539, 2012. DOI: 10.1038/nrc3310
- 55 Qian LX, Cao X, Du MY, Ma CX, Zhu HM, Peng Y, Hu XY, He X, Yin L: KIF18A knockdown reduces proliferation, migration, invasion and enhances radiosensitivity of esophageal cancer. Biochem Biophys Res Commun 557: 192-198, 2021. DOI: 10.1016/j.bbrc.2021.04.020
- 56 Zhang H, Shen T, Zhang Z, Li Y, Pan Z: Expression of KIF18A is associated with increased tumor stage and cell proliferation in prostate cancer. Med Sci Monit 25: 6418-6428, 2019. DOI: 10.12659/MSM.917352
- 57 Chen D, Chou FJ, Chen Y, Tian H, Wang Y, You B, Niu Y, Huang CP, Yeh S, Xing N, Chang C: Targeting the radiation-induced TR4 nuclear receptor-mediated QKI/circZEB1/miR-141-3p/ZEB1 signaling increases prostate cancer radiosensitivity. Cancer Lett 495: 100-111, 2020. DOI: 10.1016/j.canlet. 2020.07.040
- 58 Lin SJ, Yang DR, Li G, Chang C: TR4 nuclear receptor different roles in prostate cancer progression. Front Endocrinol (Lausanne) 6: 78, 2015. DOI: 10.3389/fendo.2015.00078
- 59 Zhang K, Yan F, Lei X, Wei D, Lu H, Zhu Z, Xiang A, Ye Z, Wang L, Zheng W, Li X, Yuan J, Lu Z, Yuan J: Androgen receptor-mediated upregulation of quaking affects androgen receptor-related prostate cancer development and anti-androgen receptor therapy. Mol Med Rep 17(6): 8203-8211, 2018. DOI: 10.3892/mmr.2018.8882
- 60 Coyle C, Cafferty FH, Vale C, Langley RE: Metformin as an adjuvant treatment for cancer: a systematic review and meta-analysis. Ann Oncol 27(12): 2184-2195, 2016. DOI: 10.1093/annonc/mdw410
- 61 Chaves LP, Melo CM, Saggioro FP, Reis RBD, Squire JA: Epithelial-mesenchymal transition signaling and prostate cancer stem cells: emerging biomarkers and opportunities for precision therapeutics. Genes (Basel) 12(12): 1900, 2021. DOI: 10.3390/genes12121900
- 62 Zhang P, Wei Y, Wang L, Debeb BG, Yuan Y, Zhang J, Yuan J, Wang M, Chen D, Sun Y, Woodward WA, Liu Y, Dean DC, Liang H, Hu Y, Ang KK, Hung MC, Chen J, Ma L: ATM-mediated stabilization of ZEB1 promotes DNA damage response and radioresistance through CHK1. Nat Cell Biol 16(9): 864-875, 2014. DOI: 10.1038/ncb3013
- 63 Liu DC, Song LL, Li XZ, Liang Q, Zhang ZG, Han CH: Circular RNA circHIPK3 modulates prostate cancer progression *via* targeting miR-448/MTDH signaling. Clin Transl Oncol 23(12): 2497-2506, 2021. DOI: 10.1007/s12094-021-02650-5
- 64 Sutherland HG, Lam YW, Briers S, Lamond AI, Bickmore WA: 3D3/lyric: a novel transmembrane protein of the endoplasmic reticulum and nuclear envelope, which is also present in the nucleolus. Exp Cell Res 294(1): 94-105, 2004. DOI: 10.1016/j.yexcr.2003.11.020

- 65 Brown DM, Ruoslahti E: Metadherin, a cell surface protein in breast tumors that mediates lung metastasis. Cancer Cell 5(4): 365-374, 2004. DOI: 10.1016/s1535-6108(04)000 79-0
- 66 Dhiman G, Srivastava N, Goyal M, Rakha E, Lothion-Roy J, Mongan NP, Miftakhova RR, Khaiboullina SF, Rizvanov AA, Baranwal M: Metadherin: A therapeutic target in multiple cancers. Front Oncol 9: 349, 2019. DOI: 10.3389/ fonc.2019.00349
- 67 Wan L, Hu G, Wei Y, Yuan M, Bronson RT, Yang Q, Siddiqui J, Pienta KJ, Kang Y: Genetic ablation of metadherin inhibits autochthonous prostate cancer progression and metastasis. Cancer Res 74(18): 5336-5347, 2014. DOI: 10.1158/0008-5472.CAN-14-1349
- 68 Wei YB, Guo Q, Gao YL, Yan B, Wang Z, Yang JR, Liu W: Repression of metadherin inhibits biological behavior of prostate cancer cells and enhances their sensitivity to cisplatin. Mol Med Rep 12(1): 226-232, 2015. DOI: 10.3892/mmr.2015.3357
- 69 Mao Y, Li W, Hua B, Gu X, Pan W, Chen Q, Xu B, Lu C, Wang Z: Circular RNA_PDHX promotes the proliferation and invasion of prostate cancer by sponging MiR-378a-3p. Front Cell Dev Biol 8: 602707, 2021. DOI: 10.3389/fcell.2020.602707
- 70 Werner H, Bruchim I: The insulin-like growth factor-I receptor as an oncogene. Arch Physiol Biochem 115(2): 58-71, 2009. DOI: 10.1080/13813450902783106
- 71 Liu G, Zhu M, Zhang M, Pan F: Emerging role of IGF-1 in prostate cancer: a promising biomarker and therapeutic target. Cancers (Basel) 15(4): 1287, 2023. DOI: 10.3390/cancers15041287
- 72 Holly JMP, Biernacka K, Perks CM: The role of insulin-like growth factors in the development of prostate cancer. Expert Rev Endocrinol Metab 15(4): 237-250, 2020. DOI: 10.1080/17446651.2020.1764844
- 73 Tufail M, Wu C: Targeting the IGF-1R in prostate and colorectal cancer: reasons behind trial failure and future directions. Ther Deliv 13(3): 167-186, 2022. DOI: 10.4155/tde-2021-0060
- 74 Li J, Cao X, Chu T, Lin K, Chen L, Lv J, Tan Y, Chen M, Li M, Wang K, Zheng Q, Li D: The circHMGCS1-miR-205–5p-ErBB3 axis mediated the Sanggenon C-induced anti-proliferation effects on human prostate cancer. Pharmacol Res 187: 106584, 2023. DOI: 10.1016/j.phrs.2022.106584
- 75 Gil V, Miranda S, Riisnaes R, Gurel B, D'Ambrosio M, Vasciaveo A, Crespo M, Ferreira A, Brina D, Troiani M, Sharp A, Sheehan B, Christova R, Seed G, Figueiredo I, Lambros M, Dolling D, Rekowski J, Alajati A, Clarke M, Pereira R, Flohr P, Fowler G, Boysen G, Sumanasuriya S, Bianchini D, Rescigno P, Aversa C, Tunariu N, Guo C, Paschalis A, Bertan C, Buroni L, Ning J, Carreira S, Workman P, Swain A, Califano A, Shen MM, Alimonti A, Neeb A, Welti J, Yuan W, de Bono J, PCF/SU2C International Prostate Cancer Dream Team: HER3 is an actionable target in advanced prostate cancer. Cancer Res

- 81(24): 6207-6218, 2021. DOI: 10.1158/0008-5472.CAN-21-3360
- 76 Weng W, Meng T, Pu J, Ma L, Shen Y, Wang Z, Pan R, Wang M, Chen C, Wang L, Zhang J, Zhou B, Shao S, Qian Y, Liu S, Hu W, Meng X: AMT-562, a novel HER3-targeting antibody-drug conjugate, demonstrates a potential to broaden therapeutic opportunities for HER3-expressing tumors. Mol Cancer Ther 22(9): 1013-1027, 2023. DOI: 10.1158/1535-7163.MCT-23-0198
- 77 Poovassery JS, Kang JC, Kim D, Ober RJ, Ward ES: Antibody targeting of HER2/HER3 signaling overcomes heregulin-induced resistance to PI3K inhibition in prostate cancer. Int J Cancer 137(2): 267-277, 2015. DOI: 10.1002/ijc.29378
- 78 Feng H, Deng Z, Peng W, Wei X, Liu J, Wang T: Circular RNA EPHA3 suppresses progression and metastasis in prostate cancer through the miR-513a-3p/BMP2 axis. J Transl Med 21(1): 288, 2023. DOI: 10.1186/s12967-023-04132-4
- 79 Li TT, Lai YW, Han X, Niu X, Zhang PX: BMP2 as a promising anticancer approach: functions and molecular mechanisms. Invest New Drugs 40(6): 1322-1332, 2022. DOI: 10.1007/s10637-022-01298-4
- 80 Horvath LG, Henshall SM, Kench JG, Turner JJ, Golovsky D, Brenner PC, O'Neill GF, Kooner R, Stricker PD, Grygiel JJ, Sutherland RL: Loss of BMP2, Smad8, and Smad4 expression in prostate cancer progression. Prostate 59(3): 234-242, 2004. DOI: 10.1002/pros.10361
- 81 Feeley BT, Gamradt SC, Hsu WK, Liu N, Krenek L, Robbins P, Huard J, Lieberman JR: Influence of BMPs on the formation of osteoblastic lesions in metastatic prostate cancer. J Bone Miner Res 20(12): 2189-2199, 2005. DOI: 10.1359/JBMR. 050802
- 82 Brubaker KD, Corey E, Brown LG, Vessella RL: Bone morphogenetic protein signaling in prostate cancer cell lines. J Cell Biochem 91(1): 151-160, 2004. DOI: 10.1002/icb.10679
- 83 Xie T, Fu DJ, Li ZM, Lv DJ, Song XL, Yu YZ, Wang C, Li KJ, Zhai B, Wu J, Feng NH, Zhao SC: CircSMARCC1 facilitates tumor progression by disrupting the crosstalk between prostate cancer cells and tumor-associated macrophages *via* miR-1322/CCL20/CCR6 signaling. Mol Cancer 21(1): 173, 2022. DOI: 10.1186/s12943-022-01630-9
- 84 Kadomoto S, Izumi K, Mizokami A: The CCL20-CCR6 axis in cancer progression. Int J Mol Sci 21(15): 5186, 2020. DOI: 10.3390/ijms21155186
- 85 Mantovani A, Allavena P, Marchesi F, Garlanda C: Macrophages as tools and targets in cancer therapy. Nat Rev Drug Discov 21(11): 799-820, 2022. DOI: 10.1038/s41573-022-00520-5
- 86 Kano H, Izumi K, Hiratsuka K, Toriumi R, Nakagawa R, Aoyama S, Kamijima T, Shimada T, Naito R, Kadomoto S, Iwamoto H, Yaegashi H, Kawaguchi S, Nohara T, Shigehara K, Kadono Y, Saito Y, Nakagawa-Goto K, Yoshioka K, Nakata H, Lin WJ, Mizokami A: Suppression of androgen receptor

- signaling induces prostate cancer migration *via* activation of the CCL20-CCR6 axis. Cancer Sci 114(4): 1479-1490, 2023. DOI: 10.1111/cas.15683
- 87 Ghadjar P, Loddenkemper C, Coupland SE, Stroux A, Noutsias M, Thiel E, Christoph F, Miller K, Scheibenbogen C, Keilholz U: Chemokine receptor CCR6 expression level and aggressiveness of prostate cancer. J Cancer Res Clin Oncol 134(11): 1181-1189, 2008. DOI: 10.1007/s00432-008-0403-5
- 88 He T, Tao W, Zhang LL, Wang BY, Li K, Lu HM, Tang GJ, He YD, Li LY: CircSCAF8 promotes growth and metastasis of prostate cancer through the circSCAF8-miR-140-3p/miR-335-LIF pathway. Cell Death Dis 13(6): 517, 2022. DOI: 10.1038/s41419-022-04913-7
- 89 Zhang C, Liu J, Wang J, Hu W, Feng Z: The emerging role of leukemia inhibitory factor in cancer and therapy. Pharmacol Ther 221: 107754, 2021. DOI: 10.1016/j.pharmthera. 2020.107754
- 90 Viswanadhapalli S, Dileep KV, Zhang KYJ, Nair HB, Vadlamudi RK: Targeting LIF/LIFR signaling in cancer. Genes Dis 9(4): 973-980, 2021. DOI: 10.1016/j.gendis.2021.04.003
- 91 Borazanci E, Schram AM, Garralda E, Brana I, Vieito Villar M, Spreafico A, Oliva M, Lakhani NJ, Hoffman K, Hallett RM, Maetzel D, Hua F, Hilbert J, Giblin P, Anido J, Kelly A, Vickers PJ, Wasserman R, Seoane J, Siu LL, Hyman DM, Hoff DV, Tabernero J: Phase I, first-in-human study of MSC-1 (AZD0171), a humanized anti-leukemia inhibitory factor monoclonal antibody, for advanced solid tumors. ESMO Open 7(4): 100530, 2022. DOI: 10.1016/j.esmoop.2022.100530
- 92 Pascual-García M, Bonfill-Teixidor E, Planas-Rigol E, Rubio-Perez C, Iurlaro R, Arias A, Cuartas I, Sala-Hojman A, Escudero L, Martínez-Ricarte F, Huber-Ruano I, Nuciforo P, Pedrosa L, Marques C, Braña I, Garralda E, Vieito M, Squatrito M, Pineda E, Graus F, Espejo C, Sahuquillo J, Tabernero J, Seoane J: LIF regulates CXCL9 in tumor-associated macrophages and prevents CD8(+) T cell tumor-infiltration impairing anti-PD1 therapy. Nat Commun 10(1): 2416, 2019. DOI: 10.1038/s41467-019-10369-9
- 93 Liu YN, Niu S, Chen WY, Zhang Q, Tao Y, Chen WH, Jiang KC, Chen X, Shi H, Liu A, Li J, Li Y, Lee YC, Zhang X, Huang J: Leukemia inhibitory factor promotes castration-resistant prostate cancer and neuroendocrine differentiation by activated ZBTB46. Clin Cancer Res 25(13): 4128-4140, 2019. DOI: 10.1158/1078-0432.CCR-18-3239
- 94 Christianson J, Oxford JT, Jorcyk CL: Emerging perspectives on leukemia inhibitory factor and its receptor in cancer. Front Oncol 11: 693724, 2021. DOI: 10.3389/fonc.2021.693724
- 95 Liu GX, Zheng T, Zhang Y, Hao P: CircFOXM1 silencing represses cell proliferation, migration and invasion by regulating miR-515-5p/ADAM10 axis in prostate cancer. Anticancer Drugs 33(1): e573-e583, 2022. DOI: 10.1097/CAD.0000000000001183
- 96 McCulloch DR, Akl P, Samaratunga H, Herington AC, Odorico DM: Expression of the disintegrin metalloprotease,

- ADAM-10, in prostate cancer and its regulation by dihydrotestosterone, insulin-like growth factor I, and epidermal growth factor in the prostate cancer cell model LNCaP. Clinical Cancer Research 10(1): 314-323, 2004. DOI: 10.1158/1078-0432.ccr-0846-3
- 97 Rosenbaum D, Saftig P: New insights into the function and pathophysiology of the ectodomain sheddase A Disintegrin And Metalloproteinase 10 (ADAM10). FEBS J 291(13): 2733-2766, 2024. DOI: 10.1111/febs.16870
- 98 Cai C, Zhang M, Liu L, Zhang H, Guo Y, Lan T, Xu Y, Ma P, Li S: ADAM10-cleaved ephrin-A5 contributes to prostate cancer metastasis. Cell Death Dis 13(5): 453, 2022. DOI: 10.1038/s41419-022-04893-8
- 99 Arima T, Enokida H, Kubo H, Kagara I, Matsuda R, Toki K, Nishimura H, Chiyomaru T, Tatarano S, Idesako T, Nishiyama K, Nakagawa M: Nuclear translocation of ADAM-10 contributes to the pathogenesis and progression of human prostate cancer. Cancer Sci 98(11): 1720-1726, 2007. DOI: 10.1111/j.1349-7006.2007.00601.x
- 100 Feng C, Wang Q, Deng L, Peng N, Yang M, Wang X: Hsa_circ_0074032 promotes prostate cancer progression through elevating homeobox A1 expression by serving as a microRNA-198 decoy. Andrologia 54(1): e14312, 2022. DOI: 10.1111/and.14312
- 101 Brotto DB, Siena ÁDD, de Barros II, Carvalho SDCES, Muys BR, Goedert L, Cardoso C, Plaça JR, Ramão A, Squire JA, Araujo LF, Silva WAD Jr: Contributions of HOX genes to cancer hallmarks: Enrichment pathway analysis and review. Tumour Biol 42(5): 101042832091805, 2020. DOI: 10.1177/1010428320918050
- 102 Cantile M, Franco R, Schiavo G, Procino A, Cindolo L, Botti G, Cillo C: The HOX genes network in uro-genital cancers: mechanisms and potential therapeutic implications. Curr Med Chem 18(32): 4872-4884, 2011. DOI: 10.2174/092986711797535182
- 103 Belpaire M, Taminiau A, Geerts D, Rezsohazy R: HOXA1, a breast cancer oncogene. Biochim Biophys Acta Rev Cancer 1877(4): 188747, 2022. DOI: 10.1016/j.bbcan.2022.188747
- 104 Wang H, Liu G, Shen D, Ye H, Huang J, Jiao L, Sun Y: HOXA1 enhances the cell proliferation, invasion and metastasis of prostate cancer cells. Oncol Rep 34(3): 1203-1210, 2015. DOI: 10.3892/or.2015.4085
- 105 Shan G, Shao B, Liu Q, Zeng Y, Fu C, Chen A, Chen Q: circFMN2 sponges miR-1238 to promote the expression of LIM-homeobox gene 2 in prostate cancer cells. Mol Ther Nucleic Acids 21: 133-146, 2020. DOI: 10.1016/j.omtn.2020.05.008
- 106 Matthews JM, Lester K, Joseph S, Curtis DJ: LIM-domain-only proteins in cancer. Nat Rev Cancer 13(2): 111-122, 2013. DOI: 10.1038/nrc3418
- 107 Wang X, He C, Hu X: LIM homeobox transcription factors, a novel subfamily which plays an important role in cancer (Review). Oncol Rep 31(5): 1975-1985, 2014. DOI: 10.3892/or.2014.3112

- 108 Hobert O, Westphal H: Functions of LIM-homeobox genes. Trends Genet 16(2): 75-83, 2000. DOI: 10.1016/s0168-9525(99)01883-1
- 109 Kuzmanov A, Hopfer U, Marti P, Meyer-Schaller N, Yilmaz M, Christofori G: LIM-homeobox gene 2 promotes tumor growth and metastasis by inducing autocrine and paracrine PDGF-B signaling. Mol Oncol 8(2): 401-416, 2014. DOI: 10.1016/j.molonc.2013.12.009
- 110 Dai X, Chen X, Chen W, Ou Y, Chen Y, Wu S, Zhou Q, Yang C, Zhang L, Jiang H: CircDHRS3 inhibits prostate cancer cell proliferation and metastasis through the circDHRS3/miR-421/MEIS2 axis. Epigenetics 18(1): 2178802, 2023. DOI: 10.1080/15592294.2023.2178802
- 111 Girgin B, Karadağ-Alpaslan M, Kocabaş F: Oncogenic and tumor suppressor function of MEIS and associated factors. Turk J Biol 44(6): 328-355, 2020. DOI: 10.3906/biy-2006-25
- 112 Smith JE, Afonja O, Yee HT, Inghirami G, Takeshita K: Chromosomal mapping to 15ql4 and expression analysis of the human MEIS2 homeobox gene. Mamm Genome 8(12): 951-952, 1997. DOI: 10.1007/s003359900621
- 113 Chen JL, Li J, Kiriluk KJ, Rosen AM, Paner GP, Antic T, Lussier YA, Vander Griend DJ: Deregulation of a Hox protein regulatory network spanning prostate cancer initiation and progression. Clin Cancer Res 18(16): 4291-4302, 2012. DOI: 10.1158/1078-0432.CCR-12-0373
- 114 Nørgaard M, Haldrup C, Bjerre MT, Høyer S, Ulhøi B, Borre M, Sørensen KD: Epigenetic silencing of MEIS2 in prostate cancer recurrence. Clin Epigenetics 11(1): 147, 2019. DOI: 10.1186/s13148-019-0742-x
- 115 Lv D, Cen S, Yang S, Zou Z, Zhong J, Pan Z, Deng N, Li Y, Wu K, Wang J, Liu P: Hsa_circ_0063329 inhibits prostate cancer growth and metastasis by modulating the miR-605-5p/tgif2 axis. Cell Cycle 22(9): 1101-1115, 2023. DOI: 10.1080/15384101.2023.2174658
- 116 Hu Y, Yu H, Shaw G, Renfree MB, Pask AJ: Differential roles of TGIF family genes in mammalian reproduction. BMC Dev Biol 11: 58, 2011. DOI: 10.1186/1471-213X-11-58
- 117 Lo RS, Wotton D, Massagué J: Epidermal growth factor signaling *via* Ras controls the Smad transcriptional corepressor TGIF. EMBO J 20(1-2): 128-136, 2001. DOI: 10.1093/emboj/20.1.128
- 118 Melhuish TA, Gallo CM, Wotton D: TGIF2 interacts with histone deacetylase 1 and represses transcription. J Biol Chem 276(34): 32109-32114, 2001. DOI: 10.1074/jbc.M103377200
- 119 Zhiping C, Shijun T, Linhui W, Yapei W, Lianxi Q, Qiang D: MiR-181a promotes epithelial to mesenchymal transition of prostate cancer cells by targeting TGIF2. Eur Rev Med Pharmacol Sci 21(21): 4835-4843, 2017.
- 120 Huang C, Deng H, Wang Y, Jiang H, Xu R, Zhu X, Huang Z, Zhao X: Circular RNA circABCC4 as the ceRNA of miR-1182 facilitates prostate cancer progression by promoting FOXP4 expression. J Cell Mol Med 23(9): 6112-6119, 2019. DOI: 10.1111/jcmm.14477

- 121 Myatt SS, Lam EW: The emerging roles of forkhead box (Fox) proteins in cancer. Nat Rev Cancer 7(11): 847-859, 2007. DOI: 10.1038/nrc2223
- 122 Bach DH, Long NP, Luu TT, Anh NH, Kwon SW, Lee SK: The dominant role of forkhead box proteins in cancer. Int J Mol Sci 19(10): 3279, 2018. DOI: 10.3390/ijms19103279
- 123 Kim JH, Hwang J, Jung JH, Lee HJ, Lee DY, Kim SH: Molecular networks of FOXP family: dual biologic functions, interplay with other molecules and clinical implications in cancer progression. Mol Cancer 18(1): 180, 2019. DOI: 10.1186/s12943-019-1110-3
- 124 Zhu X, Chen C, Wei D, Xu Y, Liang S, Jia W, Li J, Qu Y, Zhai J, Zhang Y, Wu P, Hao Q, Zhang L, Zhang W, Yang X, Pan L, Qi R, Li Y, Wang F, Yi R, Yang Z, Wang J, Zhao Y: FOXP2 confers oncogenic effects in prostate cancer. Elife 12: e81258, 2023. DOI: 10.7554/eLife.81258
- 125 Liu M, Shi X, Wang J, Xu Y, Wei D, Zhang Y, Yang K, Wang X, Liang S, Chen X, Yang F, Sun L, Zhu X, Zhao C, Zhu L, Tang L, Zheng C, Yang Z: Association of FOXP4 gene with prostate cancer and the cumulative effects of rs4714476 and 8q24 in Chinese men. Clin Lab 61(10/2015): 1491-1499, 2015. DOI: 10.7754/clin.lab.2015.150313
- 126 Deng W, Zhou X, Zhu K, Chen R, Liu X, Chen L, Jiang H, Hu B, Zeng Z, Cheng X, Yao Z, Nie J, Xiong S, Zhang C, Guo J, Fu B, Wang G: Novel circular RNA circ_0086722 drives tumor progression by regulating the miR-339-5p/STAT5A axis in prostate cancer. Cancer Lett 533: 215606, 2022. DOI: 10.1016/j.canlet.2022.215606
- 127 Verhoeven Y, Tilborghs S, Jacobs J, De Waele J, Quatannens D, Deben C, Prenen H, Pauwels P, Trinh XB, Wouters A, Smits EL, Lardon F, van Dam PA: The potential and controversy of targeting STAT family members in cancer. Semin Cancer Biol 60: 41-56, 2020. DOI: 10.1016/j.sem cancer.2019.10.002
- 128 Liao Z, Lutz J, Nevalainen MT: Transcription factor Stat5a/b as a therapeutic target protein for prostate cancer. Int J Biochem Cell Biol 42(2): 186-192, 2010. DOI: 10.1016/j.biocel.2009.11.001
- 129 Dagvadorj A, Kirken RA, Leiby B, Karras J, Nevalainen MT: Transcription factor signal transducer and activator of transcription 5 promotes growth of human prostate cancer cells *in vivo*. Clin Cancer Res 14(5): 1317-1324, 2008. DOI: 10.1158/1078-0432.CCR-07-2024
- 130 Haddad BR, Gu L, Mirtti T, Dagvadorj A, Vogiatzi P, Hoang DT, Bajaj R, Leiby B, Ellsworth E, Blackmon S, Ruiz C, Curtis M, Fortina P, Ertel A, Liu C, Rui H, Visakorpi T, Bubendorf L, Lallas CD, Trabulsi EJ, McCue P, Gomella L, Nevalainen MT: STAT5A/B gene locus undergoes amplification during human prostate cancer progression. Am J Pathol 182(6): 2264-2275, 2013. DOI: 10.1016/j.ajpath.2013.02.044
- 131 Li H, Zhang Y, Glass A, Zellweger T, Gehan E, Bubendorf L, Gelmann EP, Nevalainen MT: Activation of signal transducer and activator of transcription-5 in prostate cancer predicts

- early recurrence. Clin Cancer Res 11(16): 5863-5868, 2005. DOI: 10.1158/1078-0432.CCR-05-0562
- 132 Mirtti T, Leiby BE, Abdulghani J, Aaltonen E, Pavela M, Mamtani A, Alanen K, Egevad L, Granfors T, Josefsson A, Stattin P, Bergh A, Nevalainen MT: Nuclear Stat5a/b predicts early recurrence and prostate cancer-specific death in patients treated by radical prostatectomy. Hum Pathol 44(3): 310-319, 2013. DOI: 10.1016/j.humpath.2012.06.001
- 133 Li H, Yang Y, Yu J, Zhang B, Chen X, Zhu S, Niu Y, Shang Z: Hsa_Circ_0092339 acts as a molecular sponge in castration-resistant prostate cancer *via* the Hsa-Mir-940/ *C-MYC* axis. Epigenomics 14(13): 823-836, 2022. DOI: 10.2217/epi-2022-0111
- 134 Schuhmacher M, Staege MS, Pajic A, Polack A, Weidle UH, Bornkamm GW, Eick D, Kohlhuber F: Control of cell growth by c-Myc in the absence of cell division. Curr Biol 9(21): 1255-1258, 1999. DOI: 10.1016/s0960-9822(99)80507-7
- 135 Llombart V, Mansour MR: Therapeutic targeting of "undruggable" MYC. EBioMedicine 75: 103756, 2022. DOI: 10.1016/j.ebiom.2021.103756
- 136 Faskhoudi MA, Molaei P, Sadrkhanloo M, Orouei S, Hashemi M, Bokaie S, Rashidi M, Entezari M, Zarrabi A, Hushmandi K, Mirzaei S, Gholami MH: Molecular landscape of c-Myc signaling in prostate cancer: A roadmap to clinical translation. Pathol Res Pract 233: 153851, 2022. DOI: 10.1016/j.prp.2022.153851
- 137 Beaulieu ME, Martínez-Martín S, Kaur J, Castillo Cano V, Massó-Vallés D, Foradada Felip L, López-Estévez S, Serrano Del Pozo E, Thabussot H, Soucek L: Pharmacokinetic analysis of omomyc shows lasting structural integrity and long terminal half-life in tumor tissue. Cancers (Basel) 15(3): 826, 2023. DOI: 10.3390/cancers15030826
- 138 Xu S, Lian Z, Zhang S, Xu Y, Zhang H: CircGNG4 promotes the progression of prostate cancer by sponging miR-223 to enhance EYA3/c-myc expression. Front Cell Dev Biol 9: 684125, 2021. DOI: 10.3389/fcell.2021.684125
- 139 Tadjuidje E, Hegde RS: The Eyes Absent proteins in development and disease. Cell Mol Life Sci 70(11): 1897-1913, 2013. DOI: 10.1007/s00018-012-1144-9
- 140 Zhang L, Zhou H, Li X, Vartuli RL, Rowse M, Xing Y, Rudra P, Ghosh D, Zhao R, Ford HL: Eya3 partners with PP2A to induce c-Myc stabilization and tumor progression. Nat Commun 9(1): 1047, 2018. DOI: 10.1038/s41467-018-03327-4
- 141 Ionescu AE, Mentel M, Munteanu CVA, Sima LE, Martin EC, Necula-Petrareanu G, Szedlacsek SE: Analysis of EYA3 phosphorylation by Src kinase identifies residues involved in cell proliferation. Int J Mol Sci 20(24): 6307, 2019. DOI: 10.3390/ijms20246307
- 142 Yu YZ, Lv DJ, Wang C, Song XL, Xie T, Wang T, Li ZM, Guo JD, Fu DJ, Li KJ, Wu DL, Chan FL, Feng NH, Chen ZS, Zhao SC: Hsa_circ_0003258 promotes prostate cancer metastasis by complexing with IGF2BP3 and sponging miR-653-5p. Mol Cancer 21(1): 12, 2022. DOI: 10.1186/s12943-021-01480-x

- 143 Zandvakili I, Lin Y, Morris JC, Zheng Y: Rho GTPases: Anti- or pro-neoplastic targets? Oncogene 36(23): 3213-3222, 2017. DOI: 10.1038/onc.2016.473
- 144 Liu X, Chen J, Chen W, Xu Y, Shen Y, Xu X: Targeting IGF2BP3 in cancer. Int J Mol Sci 24(11): 9423, 2023. DOI: 10.3390/ijms24119423
- 145 Mancarella C, Scotlandi K: IGF2BP3 from physiology to cancer: novel discoveries, unsolved issues, and future perspectives. Front Cell Dev Biol 7: 363, 2020. DOI: 10.3389/fcell.2019.00363
- 146 Cuttini E, Goi C, Pellarin E, Vida R, Brancolini C: HDAC4 in cancer: A multitasking platform to drive not only epigenetic modifications. Front Mol Biosci 10: 1116660, 2023. DOI: 10.3389/fmolb.2023.1116660
- 147 Abbas A, Gupta S: The role of histone deacetylases in prostate cancer. Epigenetics 3(6): 300-309, 2008. DOI: 10.4161/epi.3.6.7273
- 148 Ding Y, Wang M, Yang J: Circular RNA midline-1 (circMID1) promotes proliferation, migration, invasion and glycolysis in prostate cancer. Bioengineered 13(3): 6293-6308, 2022. DOI: 10.1080/21655979.2022.2037367
- 149 Hsu PJ, Zhu Y, Ma H, Guo Y, Shi X, Liu Y, Qi M, Lu Z, Shi H, Wang J, Cheng Y, Luo G, Dai Q, Liu M, Guo X, Sha J, Shen B, He C: Ythdc2 is an N(6)-methyladenosine binding protein that regulates mammalian spermatogenesis. Cell Res 27(9): 1115-1127, 2017. DOI: 10.1038/cr.2017.99
- 150 Song J, You G, Yin X, Zhu G, Wang W, Yu Y, Zhu J: Overexpression of YTHDC2 contributes to the progression of prostate cancer and predicts poor outcomes in patients with prostate cancer. J Biochem Mol Toxicol 37(4): e23308, 2023. DOI: 10.1002/jbt.23308
- 151 Wu Q, Xie X, Huang Y, Meng S, Li Y, Wang H, Hu Y: N6-methyladenosine RNA methylation regulators contribute to the progression of prostate cancer. J Cancer 12(3): 682-692, 2021. DOI: 10.7150/jca.46379
- 152 Shi J, Liu C, Chen C, Guo K, Tang Z, Luo Y, Chen L, Su Y, Xu K: Circular RNA circMBOAT2 promotes prostate cancer progression *via* a miR-1271-5p/mTOR axis. Aging (Albany NY) 12(13): 13255-13280, 2020. DOI: 10.18632/aging.103432
- 153 Shorning BY, Dass MS, Smalley MJ, Pearson HB: The PI3K-AKT-mTOR pathway and prostate cancer: at the crossroads of AR, MAPK, and WNT signaling. Int J Mol Sci 21(12): 4507, 2020. DOI: 10.3390/ijms21124507
- 154 Pungsrinont T, Kallenbach J, Baniahmad A: Role of PI3K-AKT-mTOR pathway as a pro-survival signaling and resistance-mediating mechanism to therapy of prostate cancer. Int J Mol Sci 22(20): 11088, 2021. DOI: 10.3390/ijms222011088
- 155 Chen H, Zhou L, Wu X, Li R, Wen J, Sha J, Wen X: The PI3K/AKT pathway in the pathogenesis of prostate cancer. Front Biosci (Landmark Ed) 21(5): 1084-1091, 2016. DOI: 10.2741/4443
- 156 Guo K, Shi J, Tang Z, Lai C, Liu C, Li K, Li Z, Xu K: Circular RNA circARHGEF28 inhibited the progression of prostate cancer

- *via* the miR-671-5p/LGALS3BP/NF-κB axis. Cancer Sci 114(7): 2907-2919, 2023. DOI: 10.1111/cas.15820
- 157 Capone E, Iacobelli S, Sala G: Role of galectin 3 binding protein in cancer progression: a potential novel therapeutic target. J Transl Med 19(1): 405, 2021. DOI: 10.1186/s12967-021-03085-w
- 158 Hong CS, Park MR, Sun EG, Choi W, Hwang JE, Bae WK, Rhee JH, Cho SH, Chung IJ: Gal-3BP negatively regulates NF-κB signaling by inhibiting the activation of TAK1. Front Immunol 10: 1760, 2019. DOI: 10.3389/fimmu.2019.01760
- 159 Läubli H, Alisson-Silva F, Stanczak MA, Siddiqui SS, Deng L, Verhagen A, Varki N, Varki A: Lectin galactoside-binding soluble 3 binding protein (LGALS3BP) is a tumor-associated immunomodulatory ligand for CD33-related Siglecs. J Biol Chem 289(48): 33481-33491, 2014. DOI: 10.1074/jbc. M114.593129
- 160 Zhang G, Liu Y, Yang J, Wang H, Xing Z: Inhibition of circ_0081234 reduces prostate cancer tumor growth and metastasis *via* the miR-1/MAP 3 K1 axis. J Gene Med 24(8): e3376, 2022. DOI: 10.1002/jgm.3376
- 161 Suddason T, Gallagher E: A RING to rule them all? Insights into the Map3k1 PHD motif provide a new mechanistic understanding into the diverse roles of Map3k1. Cell Death Differ 22(4): 540-548, 2015. DOI: 10.1038/cdd.2014.239
- 162 Hirano T, Shino Y, Saito T, Komoda F, Okutomi Y, Takeda A, Ishihara T, Yamaguchi T, Saisho H, Shirasawa H: Dominant negative MEKK1 inhibits survival of pancreatic cancer cells. Oncogene 21(38): 5923-5928, 2002. DOI: 10.1038/sj.onc. 1205643
- 163 Abreu-Martin MT, Chari A, Palladino AA, Craft NA, Sawyers CL: Mitogen-activated protein kinase kinase kinase 1 activates androgen receptor-dependent transcription and apoptosis in prostate cancer. Mol Cell Biol 19(7): 5143-5154, 1999. DOI: 10.1128/MCB.19.7.5143
- 164 Zhang Y, Shi Z, Li Z, Wang X, Zheng P, Li H: Circ_0057553/miR-515-5p regulates prostate cancer cell proliferation, apoptosis, migration, invasion and aerobic glycolysis by targeting YES1. Onco Targets Ther 13: 11289-11299, 2020. DOI: 10.2147/OTT.S272294
- 165 Garmendia I, Redin E, Montuenga LM, Calvo A: YES1: A novel therapeutic target and biomarker in cancer. Mol Cancer Ther 21(9): 1371-1380, 2022. DOI: 10.1158/1535-7163.MCT-21-0958
- 166 Zhao S, Jie C, Xu P, Diao Y: MicroRNA-140 inhibits prostate cancer cell invasion and migration by targeting YES protooncogene 1. J Cell Biochem 121(1): 482-488, 2020. DOI: 10.1002/jcb.29231
- 167 Chatterji T, Varkaris AS, Parikh NU, Song JH, Cheng CJ, Schweppe RE, Alexander S, Davis JW, Troncoso P, Friedl P, Kuang J, Lin SH, Gallick GE: Yes-mediated phosphorylation of focal adhesion kinase at tyrosine 861 increases metastatic potential of prostate cancer cells. Oncotarget 6(12): 10175-10194, 2015. DOI: 10.18632/oncotarget.3391

- 168 Chen L, Cao H, Feng Y: MiR-199a suppresses prostate cancer paclitaxel resistance by targeting YES1. World J Urol 36(3): 357-365, 2018. DOI: 10.1007/s00345-017-2143-0
- 169 Wu Y: Circ_0044516 enriches the level of SARM1 as a miR-330-5p sponge to regulate cell malignant behaviors and tumorigenesis of prostate cancer. Biochem Genet 60(4): 1346-1361, 2022. DOI: 10.1007/s10528-021-10160-w
- 170 Waller TJ, Collins CA: Multifaceted roles of SARM1 in axon degeneration and signaling. Front Cell Neurosci 16: 958900, 2022. DOI: 10.3389/fncel.2022.958900
- 171 Sarkar A, Kumari N, Mukherjee P: The curious case of SARM1: Dr. Jekyll and Mr. Hyde in cell death and immunity? Hyde in cell death and immunity? FEBS J 290(2): 340-358, 2023. DOI: 10.1111/febs.16256
- 172 Yan K, Hou L, Liu T, Jiao W, Ma Q, Fang Z, Zhang S, Song D, Liu J, Gao X, Fan Y: IncRNA OGFRP1 functions as a ceRNA to promote the progression of prostate cancer by regulating SARM1 level *via* miR-124-3p. Aging (Albany NY) 12(10): 8880-8892, 2020. DOI: 10.18632/aging.103007
- 173 Huang S, Zhao J, Yu H, Chen G: Mechanism of tumor-derived extracellular vesicles in prostatic cancer progression through the circFMN2/KLF2/RNF128 axis. Apoptosis 28(9-10): 1372-1389, 2023. DOI: 10.1007/s10495-023-01872-y
- 174Wang J, Guo Y, Chu H, Guan Y, Bi J, Wang B: Multiple functions of the RNA-binding protein HuR in cancer progression, treatment responses and prognosis. Int J Mol Sci 14(5): 10015-10041, 2013. DOI: 10.3390/ijms 140510015
- 175 Turpaev KT: Transcription factor KLF2 and its role in the regulation of inflammatory processes. Biochemistry (Mosc) 85(1): 54-67, 2020. DOI: 10.1134/S0006297920010058
- 176 Zhu Y, Gan Y, Zou R, Sha H, Lu Y, Zhang Y, Feng J: RNF128 suppresses the malignancy of colorectal cancer cells via inhibition of Wnt/ β -catenin signaling. Am J Transl Res 13(12): 13567-13578, 2021.
- 177 Wang B, Liu M, Song Y, Li C, Zhang S, Ma L: KLF2 inhibits the migration and invasion of prostate cancer cells by downregulating MMP2. Am J Mens Health 13(1): 155798 8318816907, 2019. DOI: 10.1177/1557988318816907
- 178 Zhang C, Wang S, Chao F, Jia G, Ye X, Han D, Wei Z, Liu J, Xu G, Chen G: The short inverted repeats-induced circEXOC6B inhibits prostate cancer metastasis by enhancing the binding of RBMS1 and HuR. Mol Ther 31(6): 1705-1721, 2023. DOI: 10.1016/j.ymthe.2022.08.006
- 179 Zhang W, Sun Y, Bai L, Zhi L, Yang Y, Zhao Q, Chen C, Qi Y, Gao W, He W, Wang L, Chen D, Fan S, Chen H, Piao HL, Qiao Q, Xu Z, Zhang J, Zhao J, Zhang S, Yin Y, Peng C, Li X, Liu Q, Liu H, Wang Y: RBMS1 regulates lung cancer ferroptosis through translational control of SLC7A11. J Clin Invest 131(22): e152067, 2021. DOI: 10.1172/JCI152067
- 180 Li H: Physiologic and pathophysiologic roles of AKAP12. Sci Prog 105(3): 368504221109212, 2022. DOI: 10.1177/ 00368504221109212

- 181 Wu X, Wu T, Li K, Li Y, Hu TT, Wang WF, Qiang SJ, Xue SB, Liu WW: The mechanism and influence of AKAP12 in different cancers. Biomed Environ Sci 31(12): 927-932, 2018. DOI: 10.3967/bes2018.127
- 182 Gelman IH: Suppression of tumor and metastasis progression through the scaffolding functions of SSeCKS/Gravin/AKAP12. Cancer Metastasis Rev 31(3-4): 493-500, 2012. DOI: 10.1007/s10555-012-9360-1
- 183 Ren X, Cheng J, Zhu M, Chen X, Jiang M, Hu X, Lu Y: Circular RNA circ_0062019 exerts oncogenic properties in prostate cancer *via* mediating miR-1253/NRBP1 axis. Andrologia 54(3): , 2022. DOI: 10.1111/and.14343
- 184 Cao C, Sun G, Le K, Xu Q, Liu C: The circular RNA Circ_0085494 regulates prostate cancer progression through NRBP1/miR-497-5p axis. Biochem Genet 61(5): 1775-1790, 2023. DOI: 10.1007/s10528-023-10341-9
- 185 Qin L, Sun X, Zhou F, Liu C: CircLRP6 contributes to prostate cancer growth and metastasis by binding to miR-330-5p to up-regulate NRBP1. World J Surg Oncol 19(1): 184, 2021. DOI: 10.1186/s12957-021-02287-2
- 186 Hooper JD, Baker E, Ogbourne SM, Sutherland GR, Antalis TM: Cloning of the cDNA and localization of the gene encoding human NRBP, a ubiquitously expressed, multidomain putative adapter protein. Genomics 66(1): 113-118, 2000. DOI: 10.1006/geno.2000.6167
- 187 Wei H, Wang H, Ji Q, Sun J, Tao L, Zhou X: NRBP1 is downregulated in breast cancer and NRBP1 overexpression inhibits cancer cell proliferation through Wnt/ β -catenin signaling pathway. Onco Targets Ther 8: 3721-3730, 2015. DOI: 10.2147/OTT.S89779
- 188 Yang X, Cruz MI, Nguyen EV, Huang C, Schittenhelm RB, Luu J, Cowley KJ, Shin SY, Nguyen LK, Lim Kam Sian TCC, Clark KC, Simpson KJ, Ma X, Daly RJ: The pseudokinase NRBP1 activates Rac1/Cdc42 *via* P-Rex1 to drive oncogenic signalling in triple-negative breast cancer. Oncogene 42(11): 833-847, 2023. DOI: 10.1038/s41388-023-02594-w
- 189 Ruiz C, Oeggerli M, Germann M, Gluderer S, Stocker H, Andreozzi M, Thalmann GN, Cecchini MG, Zellweger T, Stürm S, Koivisto PA, Helin HJ, Gelmann EP, Glass AG, Gasser TC, Terracciano LM, Bachmann A, Wyler S, Bubendorf L, Rentsch CA: High NRBP1 expression in prostate cancer is linked with poor clinical outcomes and increased cancer cell growth. Prostate 72(15): 1678-1687, 2012. DOI: 10.1002/pros. 22521
- 190 Zeng L, Liu YM, Yang N, Zhang T, Xie H: Hsa_circRNA_100146 promotes prostate cancer progression by upregulating TRIP13 *via* sponging miR-615-5p. Front Mol Biosci 8: 693477, 2021. DOI: 10.3389/fmolb.2021.693477
- 191 Lee JW, Choi HS, Gyuris J, Brent R, Moore DD: Two classes of proteins dependent on either the presence or absence of thyroid hormone for interaction with the thyroid hormone receptor. Mol Endocrinol 9(2): 243-254, 1995. DOI: 10.1210/mend.9.2.7776974

- 192 Baniahmad A, Ha I, Reinberg D, Tsai S, Tsai MJ, O'Malley BW: Interaction of human thyroid hormone receptor beta with transcription factor TFIIB may mediate target gene derepression and activation by thyroid hormone. Proc Natl Acad Sci USA 90(19): 8832-8836, 1993. DOI: 10.1073/pnas.90.19.8832
- 193 Lu S, Qian J, Guo M, Gu C, Yang Y: Insights into a crucial role of TRIP13 in human cancer. Comput Struct Biotechnol J 17: 854-861, 2019. DOI: 10.1016/j.csbj.2019.06.005
- 194 Dong L, Ding H, Li Y, Xue D, Li Z, Liu Y, Zhang T, Zhou J, Wang P: TRIP13 is a predictor for poor prognosis and regulates cell proliferation, migration and invasion in prostate cancer. Int J Biol Macromol 121: 200-206, 2019. DOI: 10.1016/j.ijbiomac.2018.09.168
- 195 Huang G, Jiang Z, Zhu W, Wu Z: Exosomal circKDM4A induces CUL4B to promote prostate cancer cell malignancy in a miR-338-3p-dependent manner. Biochem Genet 61(1): 390-409, 2023. DOI: 10.1007/s10528-022-10251-2
- 196Li Y, Wang X: The role of cullin4B in human cancers. Exp Hematol Oncol 6: 17, 2017. DOI: 10.1186/s40164-017-0077-2
- 197 Gu Z, You Z, Yang Y, Ding R, Wang M, Pu J, Chen J: Inhibition of microRNA miR-101-3p on prostate cancer progression by regulating Cullin 4B (CUL4B) and PI3K/AKT/mTOR signaling pathways. Bioengineered 12(1): 4719-4735, 2021. DOI: 10.1080/21655979.2021.1949513
- 198 Ma T, Chen H, Wang P, Yang N, Bao J: Downregulation of lncRNA ZEB1-AS1 represses cell proliferation, migration, and invasion through mediating PI3K/AKT/mTOR signaling by miR-342-3p/CUL4B axis in prostate cancer. Cancer Biother Radiopharm 35(9): 661-672, 2020. DOI: 10.1089/cbr. 2019.3123
- 199 Ding L, Lin Y, Chen X, Wang R, Lu H, Wang H, Luo W, Lu Z, Xia L, Zhou X, Li G, Cheng S: circPHF16 suppresses prostate cancer metastasis *via* modulating miR-581/RNF128/Wnt/β-catenin pathway. Cell Signal 102: 110557, 2023. DOI: 10.1016/j.cellsig.2022.110557
- 200 Wei CY, Zhu MX, Yang YW, Zhang PF, Yang X, Peng R, Gao C, Lu JC, Wang L, Deng XY, Lu NH, Qi FZ, Gu JY: Downregulation of RNF128 activates Wnt/β-catenin signaling to induce cellular EMT and stemness *via* CD44 and CTTN ubiquitination in melanoma. J Hematol Oncol 12(1): 21, 2019. DOI: 10.1186/s13045-019-0711-z
- 201 Zhuang Y, Liu P, Zhan Y, Kong D, Tian F, Zhao P: RING finger protein 128 (RNF128) regulates malignant biological behaviors of colorectal cancer cells *via* PI3K/AKT signaling pathway. Cell Biol Int 46(10): 1604-1611, 2022. DOI: 10.1002/cbin.11835
- 202 Bai XS, Zhang C, Peng R, Jiang GQ, Jin SJ, Wang Q, Ke AW, Bai DS: RNF128 promotes malignant behaviors *via* EGFR/MEK/ERK pathway in hepatocellular carcinoma. Onco Targets Ther 13: 10129-10141, 2020. DOI: 10.2147/OTT.S269606

- 203 Gao F, Xu Q, Tang Z, Zhang N, Huang Y, Li Z, Dai Y, Yu Q, Zhu J: Exosomes derived from myeloid-derived suppressor cells facilitate castration-resistant prostate cancer progression *via* S100A9/circMID1/miR-506-3p/MID1. J Transl Med 20(1): 346, 2022. DOI: 10.1186/s12967-022-03494-5
- 204 Baldini R, Mascaro M, Meroni G: The MID1 gene product in physiology and disease. Gene 747: 144655, 2020. DOI: 10.1016/j.gene.2020.144655
- 205 Winter J, Basilicata MF, Stemmler MP, Krauss S: The MID1 protein is a central player during development and in disease. Front Biosci (Landmark Ed) 21(3): 664-682, 2016. DOI: 10.2741/4413
- 206 Köhler A, Demir U, Kickstein E, Krauss S, Aigner J, Aranda-Orgillés B, Karagiannidis AI, Achmüller C, Bu H, Wunderlich A, Schweiger MR, Schaefer G, Schweiger S, Klocker H, Schneider R: A hormone-dependent feedback-loop controls androgen receptor levels by limiting MID1, a novel translation enhancer and promoter of oncogenic signaling. Mol Cancer 13: 146, 2014. DOI: 10.1186/1476-4598-13-146
- 207 Zhang Y, Liu F, Feng Y, Xu X, Wang Y, Zhu S, Dong J, Zhao S, Xu B, Feng N: CircRNA circ_0006156 inhibits the metastasis of prostate cancer by blocking the ubiquitination of S100A9. Cancer Gene Ther 29(11): 1731-1741, 2022. DOI: 10.1038/s41417-022-00492-z
- 208 Lv Z, Li W, Wei X: S100A9 promotes prostate cancer cell invasion by activating TLR4/NF- κ B/integrin β 1/FAK signaling. Onco Targets Ther 13: 6443-6452, 2020. DOI: 10.2147/OTT.S192250
- 209 Hermani A, Hess J, De Servi B, Medunjanin S, Grobholz R, Trojan L, Angel P, Mayer D: Calcium-binding proteins S100A8 and S100A9 as novel diagnostic markers in human prostate cancer. Clin Cancer Res 11(14): 5146-5152, 2005. DOI: 10.1158/1078-0432.CCR-05-0352
- 210 Wang S, Chao F, Zhang C, Han D, Xu G, Chen G: Circular RNA circPFKP promotes cell proliferation by activating IMPDH2 in prostate cancer. Cancer Lett 524: 109-120, 2022. DOI: 10.1016/j.canlet.2021.10.021
- 211 Kofuji S, Sasaki AT: GTP metabolic reprogramming by IMPDH2: unlocking cancer cells' fuelling mechanism. J Biochem 168(4): 319-328, 2020. DOI: 10.1093/jb/mvaa085
- 212 Wieczorek P, Bałut-Wieczorek M, Jasinski M, Szabłoński W, Antczak A: Inosine monophosphate dehydrogenase 2 as a marker of aggressive and advanced prostate cancer. Cent European J Urol 71(4): 399-403, 2018. DOI: 10.5173/ceju. 2018.1696
- 213 Wahab NA, D Dardar H, Yunus R, M Zainudin Z, M Mokhtar N: Silencing of hepsin and inosine 5-monophosphate dehydrogenase 2 by siRNA reduces prostate cancer cells proliferation. Malays J Pathol 44(1): 29-38, 2022.
- 214 Zhou L, Xia D, Zhu J, Chen Y, Chen G, Mo R, Zeng Y, Dai Q, He H, Liang Y, Jiang F, Zhong W: Enhanced expression of IMPDH2 promotes metastasis and advanced tumor progression in

- patients with prostate cancer. Clin Transl Oncol 16(10): 906-913, 2014. DOI: 10.1007/s12094-014-1167-9
- 215 Chen J, Xie Q, Miao W, Fan J, Zhou X, Li M: CircPDHX promotes prostate cancer cell progression *in vitro* and tumor growth *in vivo via* miR-497-5p/ACSL1 axis. Biochem Biophys Res Commun 620: 35-41, 2022. DOI: 10.1016/j.bbrc.2022.06.012
- 216 Tang Y, Zhou J, Hooi SC, Jiang YM, Lu GD: Fatty acid activation in carcinogenesis and cancer development: Essential roles of long-chain acyl-CoA synthetases. Oncol Lett 16(2): 1390-1396, 2018. DOI: 10.3892/ol.2018.8843
- 217 Rossi Sebastiano M, Konstantinidou G: Targeting long chain acyl-CoA synthetases for cancer therapy. Int J Mol Sci 20(15): 3624, 2019. DOI: 10.3390/ijms20153624
- 218 Quan J, Bode AM, Luo X: ACSL family: The regulatory mechanisms and therapeutic implications in cancer. Eur J Pharmacol 909: 174397, 2021. DOI: 10.1016/j.ejphar.2021. 174397
- 219 Ma Y, Zha J, Yang X, Li Q, Zhang Q, Yin A, Beharry Z, Huang H, Huang J, Bartlett M, Ye K, Yin H, Cai H: Long-chain fatty acyl-CoA synthetase 1 promotes prostate cancer progression by elevation of lipogenesis and fatty acid beta-oxidation. Oncogene 40(10): 1806-1820, 2021. DOI: 10.1038/s41388-021-01667-y
- 220 Zhong C, Long Z, Yang T, Wang S, Zhong W, Hu F, Teoh JY, Lu J, Mao X: M6A-modified circRBM33 promotes prostate cancer progression *via* PDHA1-mediated mitochondrial respiration regulation and presents a potential target for ARSI therapy. Int J Biol Sci 19(5): 1543-1563, 2023. DOI: 10.7150/ijbs.77133
- 221 Siomi H, Siomi MC, Nussbaum RL, Dreyfuss G: The protein product of the fragile X gene, FMR1, has characteristics of an RNA-binding protein. Cell 74(2): 291-298, 1993. DOI: 10.1016/0092-8674(93)90420-u
- 222 Hu Y, Gao Q, Ma S, Yu P, Ding S, Yao X, Zhang Z, Lu S, Lu M, Zhang J, Wang Y, Qian X, Zhong J: FMR1 promotes the progression of colorectal cancer cell by stabilizing EGFR mRNA in an m(6)A-dependent manner. Cell Death Dis 13(11): 941, 2022. DOI: 10.1038/s41419-022-05391-7
- 223 Sperl W, Fleuren L, Freisinger P, Haack TB, Ribes A, Feichtinger RG, Rodenburg RJ, Zimmermann FA, Koch J, Rivera I, Prokisch H, Smeitink JA, Mayr JA: The spectrum of pyruvate oxidation defects in the diagnosis of mitochondrial disorders. J Inherit Metab Dis 38(3): 391-403, 2015. DOI: 10.1007/s10545-014-9787-3
- 224 Liu Z, Yu M, Fei B, Fang X, Ma T, Wang D: miR-21-5p targets PDHA1 to regulate glycolysis and cancer progression in gastric cancer. Oncol Rep 40(5): 2955-2963, 2018. DOI: 10.3892/or.2018.6695
- 225 Ding X, Sun J, Zhang X: Circ_0076305 facilitates prostate cancer development *via* sponging miR-411-5p and regulating PGK1. Andrologia 54(6): e14406, 2022. DOI: 10.1111/and.14406
- 226 He Y, Luo Y, Zhang D, Wang X, Zhang P, Li H, Ejaz S, Liang S: PGK1-mediated cancer progression and drug resistance. Am J Cancer Res 9(11): 2280-2302, 2019.

- 227 Fu Q, Yu Z: Phosphoglycerate kinase 1 (PGK1) in cancer: A promising target for diagnosis and therapy. Life Sci 256: 117863, 2020. DOI: 10.1016/j.lfs.2020.117863
- 228 Chen JY, Xu LF, Hu HL, Wen YQ, Chen D, Liu WH: MiRNA-215-5p alleviates the metastasis of prostate cancer by targeting PGK1. Eur Rev Med Pharmacol Sci 24(2): 639-646, 2020. DOI: 10.26355/eurrev_202001_20040
- 229 Jung Y, Shiozawa Y, Wang J, Wang J, Wang Z, Pedersen EA, Lee CH, Hall CL, Hogg PJ, Krebsbach PH, Keller ET, Taichman RS: Expression of PGK1 by prostate cancer cells induces bone formation. Mol Cancer Res 7(10): 1595-1604, 2009. DOI: 10.1158/1541-7786.MCR-09-0072
- 230 Tang Y, Liu J, Li X, Wang W: Exosomal circRNA HIPK3 knockdown inhibited cell proliferation and metastasis in prostate cancer by regulating miR-212/BMI-1 pathway. J Biosci 46: 69, 2021.
- 231 Xu J, Li L, Shi P, Cui H, Yang L: The crucial roles of Bmi-1 in cancer: implications in pathogenesis, metastasis, drug resistance, and targeted therapies. Int J Mol Sci 23(15): 8231, 2022. DOI: 10.3390/ijms23158231
- 232 Gautam N, Kaur M, Kaur S: Structural assembly of Polycomb group protein and Insight of EZH2 in cancer progression. J Cancer Res Ther 17(2): 311-326, 2021. DOI: 10.4103/ jcrt.JCRT_1090_19
- 233 Bansal N, Bartucci M, Yusuff S, Davis S, Flaherty K, Huselid E, Patrizii M, Jones D, Cao L, Sydorenko N, Moon YC, Zhong H, Medina DJ, Kerrigan J, Stein MN, Kim IY, Davis TW, DiPaola RS, Bertino JR, Sabaawy HE: BMI-1 targeting interferes with patient-derived tumor-initiating cell survival and tumor growth in prostate cancer. Clin Cancer Res 22(24): 6176-6191, 2016. DOI: 10.1158/1078-0432.CCR-15-3107
- 234 Yu J, Lu Y, Cui D, Li E, Zhu Y, Zhao Y, Zhao F, Xia S: miR-200b suppresses cell proliferation, migration and enhances chemosensitivity in prostate cancer by regulating Bmi-1. Oncol Rep 31(2): 910-918, 2014. DOI: 10.3892/or.2013.2897
- 235 Zhang Y, Wang K, Yang D, Liu F, Xu X, Feng Y, Wang Y, Zhu S, Gu C, Sheng J, Hu L, Xu B, Lu Y, Feng N: Hsa_circ_0094606 promotes malignant progression of prostate cancer by inducing M2 polarization of macrophages through PRMT1-mediated arginine methylation of ILF3. Carcinogenesis 44(1): 15-28, 2023. DOI: 10.1093/carcin/bgac091
- 236 Hwang JW, Cho Y, Bae GU, Kim SN, Kim YK: Protein arginine methyltransferases: promising targets for cancer therapy. Exp Mol Med 53(5): 788-808, 2021. DOI: 10.1038/s12276-021-00613-y
- 237 Vrakas CN, Herman AB, Ray M, Kelemen SE, Scalia R, Autieri MV: RNA stability protein ILF3 mediates cytokine-induced angiogenesis. FASEB J 33(3): 3304-3316, 2019. DOI: 10.1096/fj.201801315R
- 238 Tang S, Sethunath V, Metaferia NY, Nogueira MF, Gallant DS, Garner ER, Lairson LA, Penney CM, Li J, Gelbard MK, Alaiwi SA, Seo JH, Hwang JH, Strathdee CA, Baca SC, AbuHammad S, Zhang X, Doench JG, Hahn WC, Takeda DY, Freedman ML, Choi

- PS, Viswanathan SR: A genome-scale CRISPR screen reveals PRMT1 as a critical regulator of androgen receptor signaling in prostate cancer. Cell Rep 38(8): 110417, 2022. DOI: 10.1016/j.celrep.2022.110417
- 239 Grypari IM, Logotheti S, Zolota V, Troncoso P, Efstathiou E, Bravou V, Melachrinou M, Logothetis C, Tzelepi V: The protein arginine methyltransferases (PRMTs) PRMT1 and CARM1 as candidate epigenetic drivers in prostate cancer progression. Medicine (Baltimore) 100(36): e27094, 2021. DOI: 10.1097/MD.00000000000027094
- 240 Yao B, Zhu S, Wei X, Chen MK, Feng Y, Li Z, Xu X, Zhang Y, Wang Y, Zhou J, Tang N, Ji C, Jiang P, Zhao SC, Qin C, Feng N: The circSPON2/miR-331-3p axis regulates PRMT5, an epigenetic regulator of CAMK2N1 transcription and prostate cancer progression. Mol Cancer 21(1): 119, 2022. DOI: 10.1186/s12943-022-01598-6
- 241 Girardot M, Hirasawa R, Kacem S, Fritsch L, Pontis J, Kota SK, Filipponi D, Fabbrizio E, Sardet C, Lohmann F, Kadam S, Ait-Si-Ali S, Feil R: PRMT5-mediated histone H4 arginine-3 symmetrical dimethylation marks chromatin at G + C-rich regions of the mouse genome. Nucleic Acids Res 42(1): 235-248, 2014. DOI: 10.1093/nar/gkt884
- 242 Wang T, Guo S, Liu Z, Wu L, Li M, Yang J, Chen R, Liu X, Xu H, Cai S, Chen H, Li W, Xu S, Wang L, Hu Z, Zhuang Q, Wang L, Wu K, Liu J, Ye Z, Ji JY, Wang C, Chen K: CAMK2N1 inhibits prostate cancer progression through androgen receptor-dependent signaling. Oncotarget 5(21): 10293-10306, 2014. DOI: 10.18632/oncotarget.2511
- 243 Peng W, Feng H, Pang L, Zhang J, Hao Y, Wei X, Xia Q, Wei Z, Song W, Wang S, Liu J, Chen K, Wang T: Downregulation of CAMK2N1 due to DNA hypermethylation mediated by DNMT1 that promotes the progression of prostate cancer. J Oncol 2023: 4539045, 2023. DOI: 10.1155/2023/4539045
- 244 Chao F, Song Z, Wang S, Ma Z, Zhuo Z, Meng T, Xu G, Chen G: Novel circular RNA circSOBP governs amoeboid migration through the regulation of the miR-141-3p/MYPT1/p-MLC2 axis in prostate cancer. Clin Transl Med 11(3): e360, 2021. DOI: 10.1002/ctm2.360
- 245 Trivedi DV, Nag S, Spudich A, Ruppel KM, Spudich JA: The myosin family of mechanoenzymes: from mechanisms to therapeutic approaches. Annu Rev Biochem 89: 667-693, 2020. DOI: 10.1146/annurev-biochem-011520-105234
- 246 Naydenov NG, Lechuga S, Huang EH, Ivanov AI: Myosin motors: novel regulators and therapeutic targets in colorectal cancer. Cancers (Basel) 13(4): 741, 2021. DOI: 10.3390/cancers13040741
- 247 Li P, Wang Z, Li S, Wang L: Circ_0006404 accelerates prostate cancer progression through regulating miR-1299/CFL2 signaling. Onco Targets Ther 14: 83-95, 2021. DOI: 10.2147/OTT.S277831
- 248 Xu J, Huang Y, Zhao J, Wu L, Qi Q, Liu Y, Li G, Li J, Liu H, Wu H: Cofilin: a promising protein implicated in cancer metastasis

- and apoptosis. Front Cell Dev Biol 9: 599065, 2021. DOI: 10.3389/fcell.2021.599065
- 249 Collazo J, Zhu B, Larkin S, Martin SK, Pu H, Horbinski C, Koochekpour S, Kyprianou N: Cofilin drives cell-invasive and metastatic responses to TGF-β in prostate cancer. Cancer Res 74(8): 2362-2373, 2014. DOI: 10.1158/0008-5472.CAN-13-3058
- 250 Lu LI, Fu NI, Luo XU, Li XY, Li XP: Overexpression of cofilin 1 in prostate cancer and the corresponding clinical implications. Oncol Lett 9(6): 2757-2761, 2015. DOI: 10.3892/ol.2015.3133
- 251 Mao S, Zhang W, Yang F, Guo Y, Wang H, Wu Y, Wang R, Maskey N, Zheng Z, Li C, Ma W, Zhang J, Yan Y, Yao X: Hsa_circ_0004296 inhibits metastasis of prostate cancer by interacting with EIF4A3 to prevent nuclear export of ETS1 mRNA. J Exp Clin Cancer Res 40(1): 336, 2021. DOI: 10.1186/s13046-021-02138-8
- 252 Ye J, She X, Liu Z, He Z, Gao X, Lu L, Liang R, Lin Y: Eukaryotic initiation factor 4A-3: a review of its physiological role and involvement in oncogenesis. Front Oncol 11: 712045, 2021. DOI: 10.3389/fonc.2021.712045
- 253 Zhu Y, Ren C, Yang L: Effect of eukaryotic translation initiation factor 4A3 in malignant tumors. Oncol Lett 21(5): 358, 2021. DOI: 10.3892/ol.2021.12619
- 254 Chan CC, Dostie J, Diem MD, Feng W, Mann M, Rappsilber J, Dreyfuss G: eIF4A3 is a novel component of the exon junction complex. RNA 10(2): 200-209, 2004. DOI: 10.1261/rna.5230104
- 255 Rodgers JJ, McClure R, Epis MR, Cohen RJ, Leedman PJ, Harvey JM, Australian Prostate Cancer BioResource, Thomas MA, Bentel JM: ETS1 induces transforming growth factor β signaling and promotes epithelial-to-mesenchymal transition in prostate cancer cells. J Cell Biochem 120(1): 848-860, 2019. DOI: 10.1002/jcb.27446
- 256 Smith AM, Findlay VJ, Bandurraga SG, Kistner-Griffin E, Spruill LS, Liu A, Golshayan AR, Turner DP: ETS1 transcriptional activity is increased in advanced prostate cancer and promotes the castrate-resistant phenotype. Carcinogenesis 33(3): 572-580, 2012. DOI: 10.1093/carcin/bgs007
- 257 Wang G, Zhao H, Duan X, Ren Z: CircRNA pappalysin 1 facilitates prostate cancer development through miR-515-5p/FKBP1A axis. Andrologia 53(11): e14227, 2021. DOI: 10.1111/and.14227
- 258 Romano S, D'Angelillo A, Romano MF: Pleiotropic roles in cancer biology for multifaceted proteins FKBPs. Biochim Biophys Acta 1850(10): 2061-2068, 2015. DOI: 10.1016/j.bbagen.2015.01.004
- 259 Hausch F, Kozany C, Theodoropoulou M, Fabian AK: FKBPs and the Akt/mTOR pathway. Cell Cycle 12(15): 2366-2370, 2013. DOI: 10.4161/cc.25508
- 260 Zhang Y, Zhang D, Lv J, Wang S, Zhang Q: LncRNA SNHG15 acts as an oncogene in prostate cancer by regulating miR-

- 338-3p/FKBP1A axis. Gene 705: 44-50, 2019. DOI: 10.1016/j.gene.2019.04.033
- 261 Ding L, Wang R, Zheng Q, Shen D, Wang H, Lu Z, Luo W, Xie H, Ren L, Jiang M, Yu C, Zhou Z, Lin Y, Lu H, Xue D, Su W, Xia L, Neuhaus J, Cheng S, Li G: circPDE5A regulates prostate cancer metastasis via controlling WTAP-dependent N6-methyladenisine methylation of EIF3C mRNA. J Exp Clin Cancer Res 41(1): 187, 2022. DOI: 10.1186/s13046-022-02391-5
- 262 Fan Y, Li X, Sun H, Gao Z, Zhu Z, Yuan K: Role of WTAP in cancer: from mechanisms to the therapeutic potential. Biomolecules 12(9): 1224, 2022. DOI: 10.3390/biom 12091224
- 263 Hu J, Luo H, Xu Y, Luo G, Xu S, Zhu J, Song D, Sun Z, Kuang Y: The prognostic significance of EIF3C gene during the tumorigenesis of prostate cancer. Cancer Invest 37(4-5): 199-208, 2019. DOI: 10.1080/07357907.2019.1618322
- 264 Lothion-Roy J, Haigh DB, Harris AE, Metzler VM, Alsaleem M, Toss MS, Kariri Y, Ntekim A, Robinson BD, Khani F, Gudas LJ, Allegrucci C, James VH, Madhusudan S, Mather M, Emes RD, Archer N, Fray RG, Rakha E, Jeyapalan JN, Rutland CS, Mongan NP, Woodcock CL: Clinical and molecular significance of the RNA m(6)A methyltransferase complex in prostate cancer. Front Genet 13: 1096071, 2023. DOI: 10.3389/fgene. 2022.1096071
- 265 Fu J, Dong H, Wu J, Jin Y: Emerging progress of RNA-based antitumor therapeutics. Int J Biol Sci 19(10): 3159-3183, 2023. DOI: 10.7150/ijbs.83732
- 266 Lin L, Su K, Cheng Q, Liu S: Targeting materials and strategies for RNA delivery. Theranostics 13(13): 4667-4693, 2023. DOI: 10.7150/thno.87316
- 267 Weidle UH, Birzele F: Triple-negative breast cancer: Identification of circRNAs with efficacy in preclinical *in vivo* models. Cancer Genomics Proteomics 20(2): 117-131, 2023. DOI: 10.21873/cgp.20368
- 268 Weidle UH, Nopora A: Up-regulated circular RNAs in colorectal cancer: new entities for therapy and tools for identification of therapeutic targets. Cancer Genomics Proteomics 20(2): 132-153, 2023. DOI: 10.21873/cgp.20369
- 269 Chen D, Liu X, Lu X, Tian J: Nanoparticle drug delivery systems for synergistic delivery of tumor therapy. Front Pharmacol 14: 1111991, 2023. DOI: 10.3389/fphar.2023.1111991
- 270 Steffens RC, Wagner E: Directing the way-receptor and chemical targeting strategies for nucleic acid delivery. Pharm Res 40(1): 47-76, 2023. DOI: 10.1007/s11095-022-03385-w
- 271 Ruoslahti E: Molecular ZIP codes in targeted drug delivery. Proc Natl Acad Sci USA 119(28): e2200183119, 2022. DOI: 10.1073/pnas.2200183119
- 272 Crowley F, Sterpi M, Buckley C, Margetich L, Handa S, Dovey Z: A review of the pathophysiological mechanisms underlying castration-resistant prostate cancer. Res Rep Urol 13: 457-472, 2021. DOI: 10.2147/RRU.S264722