# A Review of Blood-mimicking Fluid Properties Using Doppler Ultrasound Applications

Ammar A. Oglat\*

Department of Medical Imaging, Faculty of Applied Medical Sciences, The Hashemite University, Zarqa, Jordan

## Abstract

Doppler imaging ultrasound characterization and standardization requires blood that is called blood mimicking fluid for the exam. With recognized internal properties, acoustic and physical features of this artificial blood. Both acoustical and physical merits set in the International Electrotechnical Commission (IEC) scale are determined as regular values, where the components utilized in the artificial blood preparation must have values identical to the IEC values. An artificial blood is commercially available in the medical application and may not be suitable in the mode of ultrasonic device or for rate of new imaging technique. It is sometimes qualified to have the strength to produce sound features and simulate blood configuration for particular implementations. In the current review article, appropriate artificial blood components, fluids, and measurements are described that have been created using varied materials and processes that have modified for medical applications.

Keywords: Acoustical properties, blood-mimicking fluid, Doppler ultrasound, International Electrotechnical Commission, physical properties

## INTRODUCTION

The blood is considered non-Newtonian when the viscosity relies on shear rate.<sup>[1-3]</sup> Usually, viscosity depends on the shear rate when tiny vessels of arterial were measured. Furthermore, during the movement of erythrocytes and when the ultrasound beam is scattered from every red cell, it will combine and then the Doppler ultrasound signal becomes clear. Doppler ultrasound imaging has been used widely in many clinical and per-clinical studies.<sup>[4,5]</sup> The essential drawback of Doppler signal is the noise, which is the output of different varieties of the numbers and methods demands of the sample size within the diffusing elements.<sup>[6]</sup> Besides blood flow velocity measurements, Doppler ultrasound has also been used for blood aggregation and clot formation studies.<sup>[7-10]</sup>

Diverse types of blood-mimicking fluid (BMFs) are contained in the literature and are revised by P. Hoskins *et al.* (1990). BMFs usually have a scattering particles material suspended in a fluid such as nylon, polystyrene, starch, and Sephadex.<sup>[11-14]</sup> The viscosity degree of BMF is suitable for a New-tonian fluid, so, it does not build on shear rate. Blood work as a New-tonian

Received: 10-06-2022 Revised: 01-08-2022 Accepted: 05-08-2022 Available Online: 20-10-2022

Access this article online				
Quick Response Code:	Website: www.jmuonline.org			
	<b>DOI:</b> 10.4103/jmu.jmu_60_22			

principle that deals within prime (large) vessels.<sup>[15]</sup> However, to obtain a proper artificial blood, the properties of this blood should be mimic to that of International Electrotechnical Commission (IEC) values with [Table 1].

Artificial blood, which is similar to the features of humans such as density,  $\alpha$ , scatter particles, speed of sound and viscosity are commercially available; the range of prices of the Commercial blood is different relying on the size of needed blood. Moreover, commercialized blood is not modified since it is prepared for wide markets and certain applications.<sup>[11-13,16-18]</sup>

Furthermore, Law *et al.* investigated that BMF properties were diverse via room temperature, humidity, and atmospheric conditions. via room temperature, moisture, and atmosphere pressure. Several of physical and acoustic features, and other examinations with various ways used to develop an artificial blood. The various sorts of admixture liquids and scatter

Address for correspondence: Dr. Ammar A. Oglat, Department of Medical Imaging, Faculty of Applied Medical Sciences, The Hashemite University, Zarqa, Jordan. E-mail: ammar.oglat@yahoo.com

This is an open access journal, and articles are distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 4.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.

For reprints contact: WKHLRPMedknow\_reprints@wolterskluwer.com

How to cite this article: Oglat AA. A review of blood-mimicking fluid properties using doppler ultrasound applications. J Med Ultrasound 2022;30:251-6.

materials utilized in artificial blood production are explained in this article. Artificial blood is made as a particles suspended in the fluids. The diameter of these particles is close to that of real human's RBCs, even though some have a larger diameter than the real human's RBCs.<sup>[12,13,19]</sup>

The hemodynamic styles of the artificial blood that is applied in vitro by Doppler US should have similar merits as human blood and be easy to prepare. The blood in the US is made of RBCs (erythrocytes). However, the blood is made of different components such as erythrocytes (RBCs), thrombocytes (platelets) and white blood cells (WBCs) (leukocytes). Why is that?

The components of human blood are responsible for significant sides of the Doppler signal. Real blood consists (RBCs) of a suspension material (erythrocytes), thrombocytes (platelets), and WBCs (leukocytes). Because of comparable low numbers of thrombocytes (250,000-350,000) and WBCs (6000-11,000), so, it is supposed that RBCs (5,000,000-6,000,000) are accountable for the scattering of the blood. The normal diameter of the RBCs is 7  $\mu$ , and this still less than the wavelength ( $\Lambda$ ) of ultrasound (0.2–0.5 mm). Hence, the sole RBC works like a spot scatter, whose mutual effect is indicated to as Rayleigh-scattering. The blood pulse echo (PE) size is small in comparison to that obtained by the reflection during the interfaces of tissue.<sup>[20,21]</sup> Based on this view, the preferable liquid to employ is the blood itself. In contrast, there are a specific drawback in applying blood and its components, such as the chance of biohazard. RBCs are easily damaged in vitro due to the ending date of blood is limited (about 120 days). the practice *in vivo* is not ethical and not safe, and this limits the utilization of artificial blood as a standard fluid in several studies of quality monitor. Thus, the concern should be taken to minimize the hazard.<sup>[22]</sup>

# Physical Properties of Blood-Mimicking Fluid Density

Density is recognized as one of the fundamental factors in BMF because it defines the quantity of mass per unit volume. The particles components applied in the preparation of the artificial blood must be remain suspended (with no float or precipitate) inside a mixture fluid, because it is good to stay neutrally energetic, though at lower velocities. The particle density should be approximately  $1.05 \pm 0.04$  g/cm<sup>3</sup> (1.01-1.09 g/cm<sup>3</sup>) close to the human blood density as IEC values.<sup>[11-14,16,22]</sup>

The density of particles should permit them to be suspended in the liquid, especially when the particles are flowing along the tube. Some researchers reported that the basic issues linked to the particle densities happened when the density is less or much larger than the liquid density. For instance, Figure 1 shows polystyrene scatter particles precipitated in the bottom because its density  $(1.050 \cdot g/cm^3)$  is greater than mixture fluid  $(1.030 \cdot g/cm^3)$ .<sup>[6,11,23]</sup> However, several previous studies measured the density of BMF via pycnometer tool.<sup>[6,11,12,14,18,19,22-26]</sup>



**Figure 1:** Appearances of the BMF samples in glass jars after 24H. Liquid density: 1.030·g/cm<sup>3</sup>, particle diameter: 5  $\mu$ . particles settles on the bottom of glycerin and water-soluble silicone oil.<sup>[15]</sup> BMF: Blood-mimicking fluid

Table 1:	: Specif	ications	of the	blood-n	nimickin	ıg fluid
defined	as the	Internat	ional E	lectrote	chnical	Commission
standar	<b>d</b> <sup>[6]</sup>					

Acoustical and physical properties of BMF	Values
Viscosity (×10 <sup>-3</sup> Pas)	$4.0{\pm}0.4$
Attenuation (dB/cm/MHz)	≤0.1
Acoustic speed (m/s)	1570±30
Density (×10 <sup>3</sup> kg/m <sup>3</sup> )	$1.050{\pm}0.040$
BMF Blood-mimicking fluid	

### Particles' size and their concentrations

Human red cells are concave in shape on both sides [Figure 2]. Particles that are used in BMF are mostly spherical and with a diameter in the range between 5 and 7  $\mu$ m or 7 and 8  $\mu$ m to mimic real human red cells.<sup>[27]</sup> For instant, the particle diameters of polystyrene microspheres are 5–30  $\mu$ m,<sup>[14,23,25,28]</sup> 5–20  $\mu$ m for nylon,<sup>[11-13,18,19,22,24,26]</sup> and 20–70  $\mu$ m for Sephadex.<sup>[6]</sup>

#### Viscosity

The viscosity of the fluid is a fundamental feature of a liquid; it is linked to the inner friction by the force and it is against the proportional movement between layers gliding past one another. The blood viscosity is considered non-Newtonian in vivo in the small arteries and Newtonian in vitro because it is anomalous.<sup>[29]</sup> The main factors that influenced non-Newtonian blood viscosity include temperature, RBCs aggregation, RBCs deformability, shear rate, plasma viscosity, and the hematocrit. Newtonian fluids are fluids that show a fixed viscosity and do not depend on the flow rate and the previous parameters. In other words, Newtonian fluids have a direct relationship between shear rate and shear stress. For example, glycerol, H<sub>2</sub>O, blood plasma, and ethanol are Newtonian fluids.<sup>[30]</sup> The viscosity of the real blood has a direct influence on the vessel speed, especially in small vessels. The velocity increases with an increasing of fluid viscosity, and this happens during preturbulence. And it has the merit of a flow exam target.<sup>[31]</sup>



Figure 2: Erythrocyte surface and cross-section<sup>[19]</sup>

In another study, they found that the dependent on shear rate happen slightly.<sup>[32,33]</sup> Pedley (1980) stated that blood with an amount of viscosity tested under conditions of maximum shear rate (4 mPa. s) is considered Newtonian in large vessels. The viscosity measurements of human blood with high shear rate were reported as 3.5–4.5 mPa. s at 37°C and with different shear rate 2.25–4.5 mPa.s.<sup>[31,34]</sup> Moreover, it found that the viscosities of fluids relied on their molecular weight and are proportional to each other.<sup>[14]</sup>

Changes of shear rate depend on several conditions, for example, the size of vessels and blood stream velocity. However, several researchers measured and calculated the kinematic viscosity and then moved it to viscosity of mixture fluid in the BMF by U-tube viscometers.<sup>[12,14,23,25,28]</sup>

# ACOUSTICAL FEATURES OF ARTIFICIAL BLOOD Sound velocity (SS) and attenuation coefficient ( $\alpha$ )

The sound velocity in artificial blood and tissue is usually 1540 m/s.<sup>[11-14,17,35]</sup> The acoustic velocity in the BMF should be similar (the same ranges to the tubes (Vessel Mimic material [VMM] and tissue) to avoid refraction problems.<sup>[12]</sup> This problem can be produced with applying tubes and large velocity.<sup>[36]</sup> For example, Ramnarine et al. (1998) investigated that the velocity of blood in draft IEC 1685 standard was reported to be  $1570 \pm 30$  m/s. This huge domain allows the velocity to correspond wall vessel, artificial blood, and the tissue of a flow test object. However, the human blood measurements were done by measure the rate of sound velocity and  $\alpha$ ,<sup>[37]</sup> and found that the  $\alpha$  of the BMF must be <0.1 dB/ cm MHz, and this recommended from the draft of IEC 1685 standard.<sup>[35]</sup> The range of attenuation coefficient for real blood is  $<0.1 \times 10 - 4 \times$  f. Therefore, to decrease inhomogeneity of the velocity range into the tube (VMM), the  $\alpha$  of the BMF should be minimal.

A lot of medical researchers have tested both the acoustic velocity and  $\alpha$  of the artificial blood by technique called PE signal.<sup>[6,11,12,14,18,22-25,38-40]</sup> Through a comprehensive previous research study, the speed of sound,  $\alpha$  were measured and

calculated through the fluid and the solid samples by calculating the time of flight (Tof) of the reflection signal wave pulse using Equations 1, 2a, and 2b, respectively:<sup>[11,41,42]</sup>

$$SS = \frac{2(d)}{t} \tag{1}$$

where SS is the velocity of sound of the sample, d is depth of the sample (distance), t is the Tof of the reflection signal wave pulse.

$$\alpha = \frac{1}{-D} \ln \frac{Ap2}{Ap1}$$
(2 a)

$$\alpha = \frac{2x(dB/cm)}{-D} \ln \frac{Ap2}{Ap1}$$
(2 b)

where  $\alpha$  is the sample attenuation coefficient, *-D* is the variation in the sample depth in mm, Ap<sub>1</sub> is the signal amplitude (amplitude of transmitted wave), Ap2 is the power signal amplitude (amplitude of received wave), and dB/cm is a constant value and equal to 8.686.

#### Effect of particles distribution on the velocity profile

Saffman (1956) found that when RBCs diffuse in the vessels, the direction of the particles will be moved toward the center and this because of its force. Kenwright et al. (2015) also reported that it is not proper to apply huge particles in tiny vessels because the particles have no ability to take over a tiny part of the diameter and which may have effects on particles diffusing and the sound profile. Though, the influence cannot be noticed. Furthermore, it is important to use small particles and identical to RBSc.[11,14,18] To ensure that the particles concentrations are increasing even for the tiny volume in the narrowing focused beam. It provides a proper blood and helpful at greater frequencies (non-Rayleigh scattering rise because of the large diameter compared to wavelength for large particles). Moreover, to produce an artificial blood that is good in tiny vessels. However, one of the major drawbacks of applying large particles in tiny vessels is the particles aggregation (clotting) inside the flow filed and then producing flow narrowing and stenosis.[6,43]

#### **Backscattering pattern of particles**

One of the critical merits of an ability suspension in the artificial blood is constancy and the US backscatter. During utilizing the backscatter of artificial blood in a Doppler application, the object should be entirely known, stable, and reproducible.<sup>[17,39]</sup> On the other hand, other researchers, Ramnarine *et al.* (2001), Ramnarine *et al.*, and Yang and Zhu (2010), indicated that penetration and sensibility measurements are crucial since it should be identical backscatter from the artificial blood and real blood. This backscatter should be known by the draft IEC 1685.<sup>[11,13]</sup> The relationship between backscattered power and the particles is proportional (linearity relationship). Thus, when particles clot, the backscattered power increase. This influence is recognized for blood.<sup>[11,19]</sup>

Fast Fourier Transformation (FFT) produces a frequency scope performance of an amplitude obtained in the time

range. An immediate measurement and calculation of the separate FFT would be very impressive. Via agents of FFT, it is probable to resolve a time unsteady signal wave into the frequency ranges contained herein and then measure the backscatter power. However, in the past five decades, the FFT was measured and calculated manually by several algorithms, and it was taken more efforts and consuming time for the backscatter power measurements.<sup>[44]</sup> Currently, there are signal wave processing instrumentations that permit the quick and ease calculation by FFT, like the situation utilized in the current project.<sup>[11,17,45]</sup>

#### Walled carotid artery phantoms

Walled phantoms for DU studies are made up of TMM, BMF, and VMM with acoustic properties close to soft tissues, blood, and vessel wall, respectively.<sup>[46-50]</sup> This means that for a walled phantom, the BMF flows through the VMM. Flow phantoms developed initially were made up of straight tubes designed using shrinking tubes, tapers, or rods.<sup>[51-53]</sup>

# DOPPLER ULTRASONOGRAPHY OF BLOOD

Doppler technology is a principle that can increase the estimation, diagnostic, and controlling pathologies in blood flow and good for research applications.<sup>[54-57]</sup> The movement of RBCs away from the transducer is specified by the decline or rise in the ultrasonic frequency. In the Doppler spectrum, evaluation of blood flow in a particular vessel is done by putting a sample-box pointer (sample gate) on the lumen (center) of the vessel, and the spectrum that shows the changes in the velocities through time in cooperation with pulses result from cardiac cycles (motor pump). Moreover, an angle pointer can be applied to show the angle of insonation with the flow direction.<sup>[58-60]</sup>

Doppler US is a technique used to measure blood speed and flow.<sup>[13,61]</sup> For example, Ginther and Utt (2004) studied the effect Doppler shift of moving red blood cells at a velocity of 1 m/s toward the vibration pulses transmitted by a US probe, 5 MHz is the US frequency of the probe, and the angle of insonation is °0. The 5 MHz is equal to  $5 \times 10^{-6}$  Hz. By applying the Doppler shift formula (Equation. 3). The Doppler-shift is positive and approximately is 6493.5 Hz. The Doppler-shift frequencies are in the field of humans audible frequencies and thus can be heard when operated through a speaker.

$$\Delta F_{T} = 2 (f V \cos(C)) (3)$$

where,  $\Delta F_T$ : Shift frequency (Doppler frequency), f<sub>o</sub>: US probe frequency (Hz), V: Blood velocity, Cos  $\Theta$ : Cosine angle of insonation, C: Speed sound in S. T which is equal to 1540 m/s.

Doppler signal size based on many factors. First, blood speed: as speed increases, the Doppler signal also increases. Second, the angle of insonation: when the beam of US has much aligned toward the direction of the flow, the Doppler signal will rise (the angle of insonation between the US beam and the flow direction becomes smaller).

# QUANTITATIVE DOPPLER TECHNIQUE Patterns of blood flow

Measuring the blood flow out of the cardiac and major vessels is done by Doppler tool. For the target of knowing the flow principle in the vessels, it is necessary to understand the variations between the main types of flow, regular (laminar) flow, and disturbed (turbulent) flow. However, when the flow pass through soft equivalent (parallel) lines in the vessel arteries and the RBCs in a region are moving at nearly with the same velocity and same direction, this indicates that this flow is laminar (Figure 2.14 (a)). Whereas, when the flow passes through obstruction or disruption of the normal flow in the vessel arteries, and the RBCs in a region are moving at disorganized velocity and confused direction, this indicates that this flow is turbulent <sup>[62,63]</sup> [Figure 3].

To know the type of flow, for instance, turbulent or laminar flow, the Reynolds number ( $R_e$ ) must be calculated by measuring the entrance length ( $L_o$ ) of VMM by following Equation 4,<sup>[12,16,64]</sup> and it must be <2100.

$$L_0 = 0.04 \times D \times R_e (4)$$

where D is the diameter of the VMM, and  $R_e$  is the Reynolds number and its unit-less that specified by this Equation (5):

$$R_{e}^{=} \frac{dvD}{m}$$
(5)

where d is the BMF density (kg/m<sup>3</sup> or g/cm<sup>3</sup>), v is the BMF mean velocity (cm/sec), D is the vessel diameter (mm or cm), and m is the BMF viscosity (mPa. s).

## Waveform Doppler indices (peak systolic velocity, end diastolic velocity, and time average mean velocity) of blood

The pulse index (PI) and resistive index (RI) can be applied to represent both the elasticity and resistance of downstream blood vessel. The best way to calculate the PI is via subtracting the end diastolic velocity (EDV) from the peak systolic velocity (PSV), then dividing by the time average mean velocity Equation 6; while the RI is calculated utilizing the PSV as the denominator or divisor Equation 7.<sup>[63,65,66]</sup> Measurements of PSV, EDV, and systolic–diastolic (S, D, or S/D) velocity ratio are significant since the PSV is the primary Doppler parameter to be abnormal in stenosis.<sup>[61]</sup>



Figure 3: Schematic illustrations and photographs of the (a) laminar and (b) turbulent flow  $^{\rm [43]}$ 

$$PI = \frac{(PSV - EDV)}{TAVm}$$
(6)

$$RI = \frac{(PSV - EDV)}{PSV}$$
(7)

A normal PSV range in Common Carotid Artery (CCA) should be nearly 60-100 cm/s or 78-118 cm/s or at least to be <125 cm/s; above this range, there may be some stenosis or obstruction occur.<sup>[65,67,68]</sup> A normal EDV range in CCA should be nearly between 20 and 32 cm/s, the RI between 0.72 and 0.84 cm/s, and PI 0.98–1.94 cm/s.<sup>[65,69]</sup> The ordinary normal speed of CCA is between 30 and 40 cm/s or 30 and 45 cm/s.<sup>[70]</sup>

# CONCLUSION

This review article provided brief but comprehensive knowledge on the materials and processes of preparing an artificial blood for DU measurements. Several artificial blood components have been mentioned with diverse features such as attenuation coefficient, viscosity of liquid, particle concentration, the sound velocity, density, and backscatter particles. The components properties guarantee easy to prepare, fit to alter their acoustic features, and is qualified to Produce a normal distribution of scatters.

# Financial support and sponsorship Nil.

#### **Conflicts of interest**

There are no conflicts of interest.

## REFERENCES

- Nichols W, O'Rourke M, Vlachopoulos C. McDonald's Blood Flow in Arteries: Theoretical, Experimental and Clinical Principles. London: CRC Press; 2011.
- Oglat AA, Matjafri MZ, Suardi N, Oqlat MA, Abdelrahman MA, Oqlat AA. A review of medical Doppler ultrasonography of blood flow in general and especially in common carotid artery. J Med Ultrasound 2018;26:3-13.
- Oglat AA, Matjafri MZ, Suardi N, Oqlat MA, Abdelrahman MA, Oqlat AA, *et al.* Chemical items used for preparing tissue-mimicking material of wall-less flow phantom for Doppler ultrasound imaging. J Med Ultrasound 2018;26:123-7.
- Huang CC, Su TH, Shih CC. High-resolution tissue Doppler imaging of the zebrafish heart during its regeneration. Zebrafish 2015;12:48-57.
- Shamloo A, Boroumand A, Ebrahimi S, Kalantarnia F, Maleki S, Moradi H. Modeling of an ultrasound system in targeted drug delivery to abdominal aortic aneurysm: A patient-specific *in silico* study based on ligand-receptor binding. IEEE Trans Ultrason Ferroelectr Freq Control 2022;69:967-74.
- Hoskins PR, Loupas T, McDicken WN. A comparison of the Doppler spectra from human blood and artificial blood used in a flow phantom. Ultrasound Med Biol 1990;16:141-7.
- Huang CC, Wang SH. Assessment of blood coagulation under various flow conditions with ultrasound backscattering. IEEE Trans Biomed Eng 2007;54:2223-30.
- Huang CC, Wang SH. Statistical variations of ultrasound signals backscattered from flowing blood. Ultrasound Med Biol 2007;33:1943-54.
- Huang CC, Wang SH, Tsui PH. *In vitro* study on assessment of blood coagulation and clot formation using Doppler ultrasound. Japanese J Appl Phys 2005;44:8727.
- 10. Huang CC, Tsui PH, Wang SH. Detection of coagulating blood under

steady flow by statistical analysis of backscattered signals. IEEE Trans Ultrason Ferroelectr Freq Control 2007;54:435-42.

- Ramnarine KV, Nassiri DK, Hoskins PR, Lubbers J. Validation of a new blood-mimicking fluid for use in Doppler flow test objects. Ultrasound Med Biol 1998;24:451-9.
- Zhou X, Kenwright DA, Wang S, Hossack JA, Hoskins PR. Fabrication of two flow phantoms for Doppler ultrasound imaging. IEEE Trans Ultrason Ferroelectr Freq Control 2017;64:53-65.
- Grand-Perret V, Jacquet JR, Leguerney I, Benatsou B, Grégoire JM, Willoquet G, *et al.* A novel microflow phantom dedicated to ultrasound microvascular measurements. Ultrason Imaging 2018;40:325-38.
- Tanaka K, Yoshida T, Sato K, Kondo T, Yasukawa K, Miyamoto N, *et al.* Blood-mimicking fluid for testing ultrasonic diagnostic instrument. Japanese J Appl Phys 2012;51(7S):07GF18. 1-12
- Pedley TJ. The Fluid Mechanics of Large Blood Vessels. Vol. 1. Cambridge: Cambridge University Press; 1980.
- Kenwright DA, Laverick N, Anderson T, Moran CM, Hoskins PR. Wall-less flow phantom for high-frequency ultrasound applications. Ultrasound Med Biol 2015;41:890-7.
- Ramnarine KV, Anderson T, Hoskins PR. Construction and geometric stability of physiological flow rate wall-less stenosis phantoms. Ultrasound Med Biol 2001;27:245-50.
- Thorne ML, Poepping TL, Rankin RN, Steinman DA, Holdsworth DW. Use of an ultrasound blood-mimicking fluid for Doppler investigations of turbulence *in vitro*. Ultrasound Med Biol 2008;34:1163-73.
- Law YF, Johnston KW, Routh HF, Cobbold RS. On the design and evaluation of a steady flow model for Doppler ultrasound studies. Ultrasound Med Biol 1989;15:505-16.
- Burns PN. The physical principles of Doppler and spectral analysis. J Clin Ultrasound 1987;15:567-90.
- Oglat AA, Suardi N, Matjafri MZ, Oqlat MA, Abdelrahman MA, Oqlat AA. A review of suspension-scattered particles used in blood-mimicking fluid for Doppler ultrasound imaging. J Med Ultrasound 2018;26:68-76.
- Samavat H, Evans JA. An ideal blood mimicking fluid for Doppler ultrasound phantoms. J Med Phys 2006;31:275-8.
- Yoshida T, Tanaka K, Sato K, Kondo T, Yasukawa K, Miyamoto N, *et al.* Blood-mimicking fluid for the Doppler test objects of medical diagnostic instruments. 2012 IEEE International Ultrasonics Symposium, IEEE.
- 24. Oates CP. Towards an ideal blood analogue for Doppler ultrasound phantoms. Phys Med Biol 1991;36:1433-42.
- Kimme-Smith C, Hussain R, Duerinckx A, Tessler F, Grant E. Assurance of consistent peak-velocity measurements with a variety of duplex Doppler instruments. Radiology 1990;177:265-72.
- Raine-Fenning NJ, Nordin NM, Ramnarine KV, Campbell BK, Clewes JS, Perkins A, *et al.* Determining the relationship between three-dimensional power Doppler data and true blood flow characteristics: An *in-vitro* flow phantom experiment. Ultrasound Obstet Gynecol 2008;32:540-50.
- Hodgson DR, McKeever KH, McGowan CM. The Athletic Horse: Principles and Practice of Equine Sports Medicine. Germany: Elsevier Health Sciences; 2014.
- Tanaka K, Yoshida T, Sato K, Kondo T, Yasukawa K, Miyamoto N, *et al.* Blood-mimicking fluid for testing ultrasonic diagnostic instrument. Japanese J Appl Phys 2012;51(7S):07GF18.
- Yalcin O, Ortiz D, Williams AT, Johnson PC, Cabrales P. Perfusion pressure and blood flow determine microvascular apparent viscosity. Exp Physiol 2015;100:977-87.
- Mandal M. Rheology of blood: Biophysical significance, measurement, pathophysiology and pharmcologic therapy. World J Pharm Pharm Sci 2016;5:2165-84.
- Wells RE Jr., Merrill EW. Influence of flow properties of blood upon viscosity-hematocrit relationships. J Clin Invest 1962;41:1591-8.
- Ercan M, Koksal C. The relationship between shear rate and vessel diameter. Anesth Analg 2003;96:307.
- Papaioannou TG, Stefanadis C. Vascular wall shear stress: Basic principles and methods. Hellenic J Cardiol 2005;46:9-15.
- Elblbesy MA, Hereba AT. Computation of the coefficients of the power law model for whole blood and their correlation with blood parameters. Appl Phys Res 2016;8:1.

255

- Browne JE, Watson AJ, Hoskins PR, Elliott AT. Validation of a sensitivity performance index test protocol and evaluation of colour Doppler sensitivity for a range of ultrasound scanners. Ultrasound Med Biol 2004;30:1475-83.
- Sato M, Ishida H, Konno K, Komatsuda T, Furukawa K, Yamada M, et al. Analysis of refractive artifacts by reconstructed three-dimensional ultrasound imaging. J Med Ultrason (2001) 2006;33:11-6.
- Duck FA. Physical Properties of Tissues: A Comprehensive Reference Book. USA: Academic Press; 2013.
- Boote EJ, Zagzebski JA. Performance tests of Doppler ultrasound equipment with a tissue and blood-mimicking phantom. J Ultrasound Med 1988;7:137-47.
- 39. Yang P, Zhu H. Influence of Transducer Focus Position and Signal Length in Backscatter Coefficient Measurement for Blood Mimicking Fluid. In Biomedical Engineering and Computer Science (ICBECS), 2010 International Conference on, IEEE; 2010.
- Oliveira PA, Silva RM, Morais GC, Alvarenga AV, Costa- Félix RP. Speed of sound as a function of temperature for ultrasonic propagation in soybean oil. J Phys Conf Ser 2016;733:012040.
- Halim MA, Buniyamin N, Mohamad Z. Improving Intramuscular Fat Measurement by Considering the Thickness of Protective Layer in Ultrasonic Transducer. In: Electrical, Electronics and System Engineering (ICEESE), 2014 International Conference on, IEEE; 2014.
- Kenwright DA, Sadhoo N, Rajagopal S, Anderson T, Moran CM, Hadoke PW, *et al.* Acoustic assessment of a Konjac–carrageenan tissue-mimicking material aT 5–60 MHZ. Ultrasound Med Biol 2014;40:2895-902.
- 43. McDicken WN. A versatile test-object for the calibration of ultrasonic Doppler flow instruments. Ultrasound Med Biol 1986;12:245-9.
- Cooley JW, Tukey JW. An algorithm for the machine calculation of complex Fourier series. Math Comput 1965;19:297-301.
- Rufo MM, Jiménez A, Paniagua JM. Comparative Study of Different Methods to Assess Ultrasonic Velocities of Waves in a Liquid Medium;2014.
- 46. Ammar AO, Matjafri M, Suardi N, Oqlat MA, Oqlat AA, Abdelrahman MA, *et al.* Characterization and construction of a robust and elastic wall-less flow phantom for high pressure flow rate using Doppler ultrasound applications. Nat Eng Sci 2018;3:359-77.
- 47. Oglat AA, Matjafri MZ, Suardi N, Abdelrahman MA, Oqlat MA, Oqlat AA. A new scatter particle and mixture fluid for preparing blood mimicking fluid for wall-less flow phantom. J Med Ultrasound 2018;26:134-42.
- 48. Oglat AA, Matjafri M, Suardi N, Oqlat MA, Oqlat AA, Abdelrahman M. A new blood mimicking fluid using propylene glycol and their properties for a flow phantom test of medical doppler ultrasound. Int J Chem Pharm Technol 2017;2(5):220-31.
- 49. Oglat AA, Matjafri MZ, Suardi N, Abdelrahman MA, Oqlat MA, Oqlat AA, *et al.* Acoustical and physical characteristic of a new blood mimicking fluid phantom. J Phys Conf Ser 2018;1083:012010.
- Dakok KK, Matjafri MZ, Suardi N, Oglat AA, Nabasu SE. A blood-mimicking fluid with cholesterol as scatter particles for wall-less carotid artery phantom applications. J Ultrason 2021;21:e219-24.
- 51. Alshipli M, Sayah MA, Oglat AA. Compatibility and validation of a recent developed artificial blood through the vascular phantom using Doppler ultrasound color- and motion-mode techniques. J Med

Ultrasound 2020;28:219-24.

- Shalbi SM, Oglat AA, Albarbar B, Elkut F, Qaeed MA, Arra AA. A brief review for common Doppler ultrasound flow phantoms. J Med Ultrasound 2020;28:138-42.
- 53. Oglat AA, Matjafri M, Suardi N, Oqlat MA, Abdelrahman MA, Oqlat AA, *et al.* Measuring the acoustical properties of fluids and solid materials via dealing with a-SCAN (GAMPT) Ultrasonic. 2018 Journal of Physics: Conference Series, IOP Publishing.
- Oglat AA, Alshipli M, Sayah MA, Ahmad MS. Artifacts in diagnostic ultrasonography, 44, 4. 2020.
- 55. Dakok K, Matjafri M, Suardi N, Oglat AA, Nabasu SE. Determination of the effect of glucose and cholesterol on the flow velocity of Blood Mimicking Fluid in a Common Carotid Artery Wall-less Phantom. J Positive Sch Psychol 2022;6:394-405.
- Dakok K, Matjafri M, Suardi N, Oglat AA, Sirisena UAI. Influences of Body Composition on Carotid Artery Structure and function in Adult Humans. J Positive Sch Psychol 2022;6:406-17.
- Oglat AA, Dheyab MA. Performance evaluation of ultrasonic imaging system (Part I). J Med Ultrasound 2021;29:258-63.
- Ginther O, Utt MD. Doppler ultrasound in equine reproduction: Principles, techniques, and potential. J Equine Vet Sci 2004;24:516-26.
- Dakok KK, Matjafri MZ, Suardi N, Oglat AA, Nabasu SE. A review of carotid artery phantoms for Doppler ultrasound applications. J Med Ultrasound 2021;29:157-66.
- Athamnah SI, Oglat AA, Fohely F. Diagnostice breast elastography estimation from Doppler imaging using central difference (CD) and least-squares (LS) algorithms. Biomed Signal Process Control 2021;68:102667.
- Mehra S. Role of duplex Doppler sonography in arterial stenoses. J Indian Acad Clin Med 2010;11:294-9.
- Kisslo JA, Adams DB. Principles of Doppler Echocardiography and the Doppler Examination# 1. London: Ciba-Geigy; 1987.
- Chavhan GB, Parra DA, Mann A, Navarro OM. Normal Doppler spectral waveforms of major pediatric vessels: specific patterns. Radiographics 2008;28:691-706.
- 64. Zhou X, Xia C, Khan F, Corner GA, Huang Z, Hoskins PR. Investigation of ultrasound-measured flow rate and wall shear rate in wrist arteries using flow phantoms. Ultrasound Med Biol 2016;42:815-23.
- Chen SY, Hsu HY. Analysis of Doppler blood flow waveform of cerebral arteries and common abnormal findings. J Med Ultrasound 2014;22:3-6.
- McNaughton DA, Abu-Yousef MM. Doppler US of the liver made simple. Radiographics 2011;31:161-88.
- 67. Gerhard-Herman M, Gardin JM, Jaff M, Mohler E, Roman M, Naqvi TZ, et al. Guidelines for noninvasive vascular laboratory testing: A report from the American Society of Echocardiography and the Society for Vascular Medicine and Biology. Vasc Med 2006;11:183-200.
- Slovut DP, Romero JM, Hannon KM, Dick J, Jaff MR. Detection of common carotid artery stenosis using duplex ultrasonography: A validation study with computed tomographic angiography. J Vasc Surg 2010;51:65-70.
- Mikkonen RH, Kreula JM, Virkkunen PJ. Peak systolic velocity, resistance index and pulsatility index. Variations in measuring a pre-recorded videotape. Acta Radiol 1997;38:598-602.
- 70. Lee W. General principles of carotid Doppler ultrasonography. Ultrasonography 2014;33:11-7.