



Identification of Specific Oral and Gut Pathogens in Full Thickness Colon of Colitis Patients: Implications for Colon Motility

Vasudevan Dinakaran^{1†}, Sammed N. Mandape^{2†}, Kristina Shuba¹, Siddharth Pratap², Shruti S. Sakhare², Mohammad Ali Tabatabai³, Duane T. Smoot⁴, Cherae M. Farmer-Dixon¹, Lakshmyya N. Kesavalu⁵, Samuel Evans Adunyah⁶, Janet Hayes Southerland⁷ and Pandu R. Gangula^{1*}

¹ Department of ODS & Research, School of Dentistry, Meharry Medical College, Nashville, TN, United States, ² Bioinformatics Core, School of Graduate Studies/Research & School of Medicine, Meharry Medical College, Nashville, TN, United States, ³ Department of Public Health, School of Graduate Studies & Research, Meharry Medical College, Nashville, TN, United States, ⁴ Department of Internal Medicine, Division of Gastroenterology & Hepatology, Meharry Medical College, Nashville, TN, United States, ⁵ Department of Periodontology, College of Dentistry, University of Florida, Gainesville, FL, United States, ⁶ Department of Biochemistry, Cancer Biology, Neuroscience & Pharmacology, Meharry Medical College, Nashville, TN, United States, ⁷ Department of Nutrition Metabolism & Oral Surgery, University of Texas Medical Branch at Galveston, Galveston, TX, United States

OPEN ACCESS

Edited by:

Nurul-Syakima Ab Mutalib, UKM Medical Molecular Biology Institute (UMBI), Malaysia

Reviewed by:

Christopher L. Hemme, University of Rhode Island, United States Chenyang Wang, Nanjing University Medical School, China

*Correspondence:

Pandu R. Gangula pgangula@mmc.edu

[†]These authors have contributed equally to this work

Specialty section:

This article was submitted to Microbial Symbioses, a section of the journal Frontiers in Microbiology

Received: 11 September 2018 Accepted: 11 December 2018 Published: 04 January 2019

Citation:

Dinakaran V, Mandape SN, Shuba K, Pratap S, Sakhare SS, Tabatabai MA, Smoot DT, Farmer-Dixon CM, Kesavalu LN, Adunyah SE, Southerland JH and Gangula PR (2019) Identification of Specific Oral and Gut Pathogens in Full Thickness Colon of Colitis Patients: Implications for Colon Motility. Front. Microbiol. 9:3220. doi: 10.3389/fmicb.2018.03220

Impaired colon motility is one of the leading problems associated with inflammatory bowel disease (IBD). An expanding body of evidence supports the role of microbiome in normal gut function and in progression of IBD. The objective of this work is to determine whether diseased full thickness colon specimens, including the neuromuscular region (critical for colon motility function), contain specific oral and gut pathogens. In addition, we compared the differences in colon microbiome between Caucasians (CA) and African Americans (AA). Thirty-nine human full thickness colon (diseased colon and adjacent healthy colon) specimens were collected from Crohn's Colitis (CC) or Ulcerative Colitis (UC) patients while they underwent elective colon surgeries. We isolated and analyzed bacterial ribosomal RNA (rRNA) from colon specimens by amplicon sequencing of the 16S rRNA gene region. The microbiome proportions were quantified into Operational Taxonomic Units (OTUs) by analysis with Quantitative Insights Into Microbial ecology (QIIME) platform. Two hundred twenty-eight different bacterial species were identified by QIIME analysis. However, we could only decipher the species name of fifty-three bacteria. Our results show that proportion of non-detrimental bacteria in CC or UC colon samples were altered compared to adjacent healthy colon specimens. We further show, for the first time in full thickness colon specimens, that microbiome of CC and UC diseased specimens is dominated by putative oral pathogens belonging to the Phyla Firmicutes (Streptococcus, Staphylococcus, Peptostreptococcus), and Fusobacteria (Fusobacterium). In addition, we have identified patterns of differences in microbiome levels between CA and AA specimens with potential implications for health disparities research. Overall, our results suggest a significant association between oral and gut microbes in the modulation of colon motility in colitis patients.

Keywords: colitis, colon motility, nitric oxide (NO), antioxidants, oral microbiome, operational taxonomic units (OTUs), gut microbiome

INTRODUCTION

Inflammatory bowel disease (IBD) is comprised of Crohn's disease / Crohn's colitis (CC) and Ulcerative colitis (UC). The term Colitis, refers to general inflammation of the inner lining of the colon arising from numerous underlying causes including idiopathic infection, IBD (either CC or UC), ischemic colitis, allergic reactions, and/or microscopic colitis. Distally, gingivitis, and periodontal disease are chronic inflammatory gum diseases associated with orange, red, yellow, purple, and green complex bacterial infections in sub-gingival areas of oral cavity (Popova et al., 2013).

Previous studies have shown that periodontal disease (PD) is a significant risk factor and contributor to many systemic diseases, including IBD (Vavricka et al., 2013). Several factors including genetic, dietary, and environmental factors could influence the pathogenesis of microbiome (oral and gut) which in turn may increase the incidence of periodontitis and IBD (Lira-Junior and Figueredo, 2016; Agossa et al., 2017). In addition, *Porphyromonas gingivalis* known to cause PD altered the gut microbiota leading to increased gut epithelial permeability and endotoxemia, which causes systemic inflammation (Hajishengallis, 2015). In addition, many earlier studies have shown intestinal colonization of oral bacteria in the pathogenesis of IBD (Strauss et al., 2011; Atarashi et al., 2017).

Innumerable number of studies have shown that the gut microbiome including Phyla Proteobacteria, Firmicutes, and Bacteroidetes contribute to normal gut function (Mariat et al., 2009; Koliada et al., 2017; Walker et al., 2018; Zhao et al., 2018). Colon motility is mainly regulated by neuromuscular portion of the colon and this was shown to be impaired in colitis patients; putatively due to a reduction in neuronal nitric oxide (NO) synthase (nNOS) protein expression and/or neuronal degeneration (Bassotti et al., 2014; Gangula et al., 2017). Previous studies have analyzed the microbiome in feces and/or colon mucosal biopsy specimens of colitis patients (Gibson et al., 1991; Bibiloni et al., 2006). However, the relationship/interaction between oral and gut bacteria in the development and/or exacerbation of inflammatory disease in the colon (containing neuromuscular tissue) was under studied. In addition, data is limited on how oral bacteria interact with and influence the large intestinal flora, thereby contributing to colitis. Since motility of the colon is impaired in colitis patients and neuromuscular tissue play a role in the motility function (Geboes and Collins, 1998; Poli et al., 2001), we hypothesize that the interaction between oral and gut microbiome may play a significant role in the inflammatory processes associated with the development and progression of colitis seen in certain patient populations. Furthermore, we hypothesize that difference in microbiome may exist between CA and AA colitis patients, potentially contributing to health disparities in IBD.

METHODS

Ethics Statement

The participants provided both written and verbal informed consent to Collaborative Human Tissue Networking (CHTN)

Consortium to collect specimens while they underwent elective colon surgeries.

Collection of Specimens

Frozen full thickness colon specimens were obtained from Cooperative Human Tissue Networking (CHTN). Thirty-nine human full thickness colon (moderate to severe diseased colon and adjacent healthy colon) specimens were collected from CC and UC male and female patients (ages between 18 and 75 years old) while they underwent elective colon surgeries. The specimens include Ulcerative (n = 13), Crohn's (n = 13) and adjacent healthy (n = 13) specimens. Characteristics of participants included Caucasians (CA) (n = 30) and African Americans (AA) (n = 9). CC male and female patients presented with symptoms like fever, fatigue, diarrhea, blood in stool, mouth sores, abdominal cramping, and pain around the anus, reduced appetite, and weight loss. While UC male and female patients presented with additional signs like rectal pain, rectal bleeding, and inability to defecate despite urgency.

Extraction of DNA, Amplification of 16S rRNA Gene and Amplicon Sequencing

DNA extraction and microbial analysis were performed in the University of North Carolina at Chapel Hill School of Medicine Microbiome Core Facility (UNC: MC). We identified a conserved region of the 16S rRNA gene of 550 bp to amplify. This encompassed variable regions V3-V4 from the colon genomic DNA using primers 16S rRNA-F 5'-AGAGTTTGATCCTGGCTCAG-3'and 16S rRNA-R 5'-GCTGCCTCCCGTAGGAGT-3' and overhang adapter sequences appended to the primer pair for compatibility with Illumina index and sequencing adapters. Briefly, each 16SrRNA amplicon was purified using AMPure XP reagent (Beckman Coulter, Indianapolis, IN, USA). Specifically, each sample was amplified using a limited cycle PCR program, adding Illumina sequencing adapters and optional dual-index barcodes [index 1(i7) and index 2(i5)] (Illumina, San Diego, CA, USA) to the amplicon target. The final libraries were purified using AMPure XP reagent, quantified and normalized prior to pooling. The DNA library pool was denatured with NaOH, diluted with hybridization buffer and heat denatured before loading on to the MiSeq reagent cartridge and to the MiSeq instrument (Illumina). The standard Illumina paired-end 250 base pair (PE250) protocol was used for sequencing the16S rRNA amplicons (Illumina, CA, USA).

Processing of Sequence Reads

Data was analyzed and microbial proportions using Operational Taxonomic Units (OTUs) were determined using Quantitative Insights Into Microbial ecology (QIIME) pipeline (Caporaso et al., 2010a) in the Meharry Medical College Bioinformatics Core. Briefly, generated raw reads were preprocessed for adapter removal. Processed sequence reads were obtained as fastq files and were converted into fasta, quality and flow files using Mothur package (Schloss et al., 2009). The initial number of fasta sequences obtained were 31,09,793. First, the fasta files were cleaned of host reads by mapping on to 9 mm mouse

genome. Then, the primer sequences and barcode sequences were removed, demultiplexed and guality filtered. The number of high quality sequences remaining after quality filtering was 16,64,769. The OTUs were picked by *de novo* strategy. The high quality sequences were clustered at 97% identity using UCLUST inbuilt in QIIME pipeline to generate 3994 OTUs and taxonomy was assigned to OTU representative sequences using UCLUST (Edgar, 2010). The picked sequences were aligned using PvNAST aligner (Caporaso et al., 2010b). The chimeric sequences and singleton OTUs were removed using ChimeraSlayer (Haas et al., 2011). We constructed a phylogenetic tree for the sequences using FastTree version 2.1.3 (data not shown) (Price et al., 2010). Next, an OTU table was constructed and taxa were summarized using the 894 OTUs obtained from QIIME pipeline. αdiversity metrics was computed using Chao1 (abundance-based richness estimator) and Shannon analysis (diversity index) and Rarefaction plots were constructed (data not shown). β-diversity metrics was computed using weighted and unweighted Principal Coordinates Analysis (PCoA) (data not shown) (Gower, 2005). A Taxonomic Summary Bar plot showing OTUs assigned to Phyla-level taxonomy per sample was subsequently constructed (Figure 1). Bar Plots showing the relative abundance of bacteria at the Phyla-level between races, diseased tissue and healthy tissue groups is shown in Figure 3. Sample-specific sequences were deposited in the MGRAST database (accession number: b3b851ba2c6d676d343739393937332e33) and was assigned an MG-RAST project ID (mgs675214) (Keegan et al., 2016). In addition, sample-specific sequences were deposited in the NCBI (BioProject: PRJNA496071).

The pathogenic and beneficial oral and gut bacteria were identified using the NCBI Genome database (https://www.ncbi. nlm.nih.gov/genome/). This analysis was performed to assess the pathogenic and healthy bacterial proportions in human full thickness colon specimens (**Tables 1–3**).

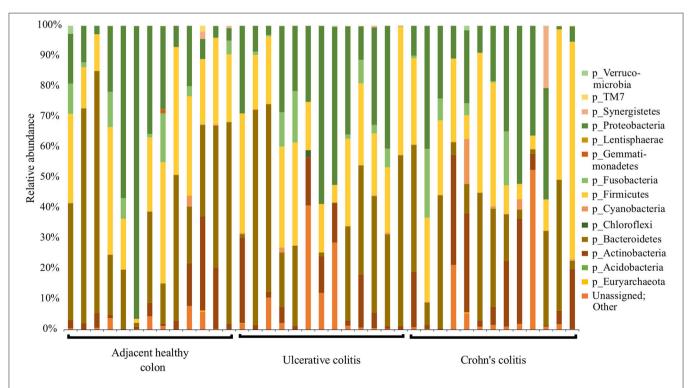
Statistical Analysis and Evaluation

Statistical analysis was performed between the healthy and diseased groups and based on race classification (n = 13 CC, n = 13 UC, n = 13 non-disease healthy patients, n = 30 CA and n = 9 AA). A non-parametric Mann-Whitney U Test *p*-value < 0.05 of bacterial 16S rRNA OTUs between the groups was considered statistically significant. IBM SPSS software package version 23 (IBM Analytics, USA) was used to conduct statistical analysis.

RESULTS

Relative Abundance Analysis

QIIME analysis showed about two hundred twenty-eight bacterial species in entire 39 specimens (**Tables 1–5**). However, non-ambiguous annotation at the species name resulted in fifty-three bacterial identifications. The dominant phyla across all samples (both diseased and healthy specimens) were Bacteroidetes (46.92%), followed by Firmicutes



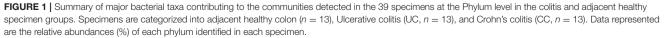


TABLE 1 | Functions and proportions of specific pathogenic Oral bacteria colonized in full thickness colon specimens.

SI. No.	Bacteria genus	Bacteria species	Proportion (%)	Bacteria phylum	Function in IBD	NCBI genome database link
	ENT HEALTHY COLON					
	Prevotella	stercorea	2.3	Bacteroidetes	Alters mucosal microbiota in the colon of patients with IBD	Н
	Prevotella	Other	0.3	Bacteroidetes	A microbial signature of Crohn's disease	GS
	Gemella	S	0.1	Firmicutes	Microbiome in New-Onset Crohn's Disease	CP
	Staphylococcus	sciuri	0.1	Firmicutes	Develops intestinal inflammation in acute and chronic colitis	I
	Staphylococcus	aureus	0.6	Firmicutes	Causes Crohn's disease	AU
	Abiotrophia	S	0.1	Firmicutes	Causes fecal microbial dysbiosis in IBD	CS
	Lactobacillus	zeae	1.9	Firmicutes	Maintains remission of ulcerative colitis	А
	Lactobacillus	s	0.4	Firmicutes	Maintains remission of ulcerative colitis	CW
	Lactococcus	S	0.7	Firmicutes	Used in the treatment of Crohn's disease	CX
0	Peptostreptococcus	anaerobius	11.6	Firmicutes	Causes dysbiosis in IBD	AW
1	Peptostreptococcus	s	0.5	Firmicutes	Causes gut microbiota dysbiosis in IBD	DR
2	Selenomonas	s	0.2	Firmicutes	Causes dysbiosis in colorectal cancer	EB
3	Eubacterium	dolichum	1.0	Firmicutes	Causes dysbiosis of the intestinal microbiota	AL
4	Fusobacterium	S	2.2	Fusobacteria	Identified from colonic biopsies of IBD patients	EN
5	Pseudomonas	alcaligenes	1.8	Proteobacteria	Identified in the gut microbiota of IBD	AX
6	Pseudomonas	S	0.1	Proteobacteria	Causes infection in Children with Early-onset Crohn's Disease	GG
7	Pseudomonas	Other	0.2	Proteobacteria	Gut microbe in children with early onset Crohn's disease	HR
8	Corynebacterium	durum	0.1	Actinobacteria	Gut microbe in IBD patients	AK
9	Corynebacterium	s	0.8	Actinobacteria	Causes experimental colitis	BI
C	Pseudoramibacter_Eubacterium	S	1.6	Firmicutes	Metabolizes Linoleic acid in the Gut	DF
ISEAS	SED COLON (ULCERATIVE COL	ITIS)				
	Prevotella	stercorea	1.0	Bacteroidetes	Alters mucosal microbiota in the colon of patients with IBD	Н
	Prevotella	S	0.3	Bacteroidetes	A microbial signature of Crohn's disease	BZ
	Prevotella	Other	0.3	Bacteroidetes	A microbial signature of Crohn's disease	GS
	Staphylococcus	aureus	0.3	Firmicutes	Causes Crohn's disease	AU
	Lactobacillus	zeae	7.6	Firmicutes	Maintains remission of ulcerative colitis	A
	Lactobacillus	S	0.3	Firmicutes	Maintains remission of ulcerative colitis	CW
	Lactococcus	S	0.6	Firmicutes	Used in the treatment of Crohn's disease	CX
	Peptostreptococcus	anaerobius	12.7	Firmicutes	Causes dysbiosis in IBD	AW
	Peptostreptococcus	S	0.3	Firmicutes	Causes gut microbiota dysbiosis in IBD	DR
0	Selenomonas	s	0.1	Firmicutes	Causes dysbiosis in colorectal cancer	EB
1	Eubacterium	dolichum	0.5	Firmicutes	Causes dysbiosis of the intestinal microbiota	AL

SI. No.	Bacteria genus	Bacteria species	Proportion (%)	Bacteria phylum	Function in IBD	NCBI genome database link
12	Pseudoramibacter_ Eubacterium	S	1.9	Firmicutes	Metabolizes Linoleic acid in the Gut	DF
3	Fusobacterium	S	3.0	Fusobacteria	Identified from colonic biopsies of IBD patients	EN
4	Pseudomonas	alcaligenes	0.4	Proteobacteria	Identified in the gut microbiota of IBD	AX
5	Pseudomonas	S	0.8	Proteobacteria	Infection in Children with Early-onset Crohn's Disease	GG
6	Aggregatibacter	S	1.4	Proteobacteria	Causes fungal microbiota dysbiosis in IBD	
7	Corynebacterium	S	1.0	Actinobacteria	Causes experimental colitis	BI
DISEAS	SED COLON (CROHN'S COLITIS)	1				
	Prevotella	tannerae	0.2	Bacteroidetes	Prevalent in colitis	F
	Prevotella	stercorea	3.3	Bacteroidetes	Alters mucosal microbiota in the colon of patients with IBD	Н
	Prevotella	melaninogenica	0.4	Bacteroidetes	Gut microbiome biomarker in ankylosing spondylitis	U
	Prevotella	Other	3.3	Bacteroidetes	A microbial signature of Crohn's disease	GS
	Gemella	S	0.1	Firmicutes	Microbiome in New-Onset Crohn's Disease	CP
	Staphylococcus	sciuri	0.1	Firmicutes	Develops intestinal inflammation in acute and chronic colitis	I
	Staphylococcus	aureus	0.5	Firmicutes	Causes Crohn's disease	AU
	Abiotrophia	S	0.2	Firmicutes	Causes fecal microbial dysbiosis in IBD	CS
	Lactobacillus	zeae	6.8	Firmicutes	Maintains remission of ulcerative colitis	А
0	Lactobacillus	reuteri	0.1	Firmicutes	Prevents colitis as a probiotic	М
1	Lactobacillus	S	0.6	Firmicutes	Maintains remission of ulcerative colitis	CW
2	Lactococcus	S	0.7	Firmicutes	Used in the treatment of Crohn's disease	CX
3	Peptostreptococcus	anaerobius	4.0	Firmicutes	Causes dysbiosis in IBD	AW
4	Peptostreptococcus	S	0.1	Firmicutes	causes gut microbiota dysbiosis in IBD	DR
5	Selenomonas	S	0.4	Firmicutes	Causes dysbiosis in colorectal cancer	EB
6	Eubacterium	dolichum	0.8	Firmicutes	Causes dysbiosis of the intestinal microbiota	AL
7	Pseudoramibacter_ Eubacterium	s	1.3	Firmicutes	Metabolizes Linoleic acid in the Gut	DF
8	Fusobacterium	S	2.4	Fusobacteria	Identified from colonic biopsies of IBD patients	EN
9	Pseudomonas	alcaligenes	0.8	Proteobacteria	Identified in the gut microbiota of IBD	AX
0	Pseudomonas	S	1.0	Proteobacteria	Infection in Children with Early-onset Crohn's Disease	GG
1	Corynebacterium	durum	0.2	Actinobacteria	Gut microbe in IBD patients	AK
2	Corynebacterium	s	0.1	Actinobacteria	Causes experimental colitis	BI
3	Pyramidobacter	piscolens	0.1	Synergistetes	Oral bacteria in IBD	Р

Specific Information of functions was adapted from NCBI Genome Database (https://www.ncbi.nlm.nih.gov/genome/).

The bacterial species that could not be identified at the genus level are mentioned as g___ and the bacterial species that could not be identified at the species level are mentioned as s___.

(27.8%), and Proteobacteria (24.5%). Most importantly, our results indicate that putative oral pathogens (belonging to mostly Phylum Firmicutes) dominated the microbiome of diseased specimens (**Figure 1**). Adjacent healthy specimens

show an increased abundance of Phylum Bacteroidetes (\sim 57%, containing mostly symbiotic and/or beneficial bacteria) population, which is altered in disease categories (**Figure 2**).

TABLE 2 | Functions and proportions of specific beneficial Gut bacteria colonized in full thickness colon specimens.

SI. No.	Bacteria genus	Bacteria species	Proportion (%)	Bacteria phylum	Function in IBD	NCBI genome database link
ADJACEI	NT HEALTHY COLON					
	Shuttleworthia	satelles	0.2	Firmicutes	Identified in the human ileum	J
2	Bifidobacterium	longum	0.1	Actinobacteria	Attenuates acute murine experimental model of IBD	Υ
3	Rhizobium	leguminosarum	0.1	Proteobacteria	Identified commensal gut microbe	AB
Ļ	Lysinibacillus	boronitolerans	12.1	Firmicutes	Identified commensal gut microbe	AT
	Alloiococcus	S	1.7	Firmicutes	Identified commensal gut microbe	CT
	Christensenella	S	2.2	Firmicutes	Identified gut microbe	DC
	Blautia	S	0.3	Firmicutes	Butyrate-producing bacterial species in Gut	DH
	Coprococcus	S	0.1	Firmicutes	Butyrate-producing bacterial species in Gut	DI
	g	S	0.7	Gemmatimonadetes	Identified commensal gut microbe	EP
0	9	S	0.7	Lentisphaerae	Normal gut microbe	EQ
1	g	S	0.4	Proteobacteria	Identified commensal gut microbe	FC
2	Comamonas	S	1.0	Proteobacteria	Identified commensal gut microbe	FL
3	Desulfovibrio	S	0.3	Proteobacteria	Sulfate reducing bacteria in IBD	FV
4	Paracoccus	Other	0.1	Proteobacteria	Identified commensal gut microbe	HH
5	Other	Other	0.2	Proteobacteria	Mucosal and fecal microbe	HP
DISEASE	D COLON (ULCERATI	VE COLITIS)				
	Lysinibacillus	boronitolerans	8.2	Firmicutes	Identified commensal gut microbe	AT
	Varibaculum	S	0.2	Actinobacteria	Identified in the gut of a premature infant	BH
	Alloiococcus	S	3.1	Firmicutes	Identified commensal gut microbe	CT
	Christensenella	S	0.6	Firmicutes	Identified gut microbe	DC
	Blautia	S	0.3	Firmicutes	Butyrate-producing bacterial species in Gut	DH
	Coprococcus	S	0.2	Firmicutes	Butyrate-producing bacterial species in Gut	DI
	g	S	0.3	Gemmatimonadetes	Identified commensal gut microbe	EP
1	g	S	0.3	Proteobacteria	Identified commensal gut microbe	FC
	Comamonas	S	2.7	Proteobacteria	Identified commensal gut microbe	FL
0	Desulfovibrio	s	0.1	Proteobacteria	Sulfate reducing bacteria in IBD	FV
1	Morganella	s	0.1	Proteobacteria	Sulfate reducing bacteria in IBD	FZ
2	g	S	0.6	TM7	Identified commensal gut microbe	
3	Other	Other	0.1	Actinobacteria	Commensal gut bacteria in IBD	GN
4	Other	Other	0.5	Proteobacteria	Adult fecal microbe	HO
DISEASE	D COLON (CROHN'S	COLITIS)				
	Akkermansia	muciniphila	0.1	Verrucomicrobia	Adheres to enterocytes and strengthens the integrity of the epithelial cell layer	S
	Bifidobacterium	longum	0.4	Actinobacteria	Attenuates acute murine experimental model of IBD	Y

SI. No.	Bacteria genus	Bacteria species	Proportion (%)	Bacteria phylum	Function in IBD	NCBI genome database link
3	Rhizobium	leguminosarum	0.2	Proteobacteria	Identified commensal gut microbe	AB
4	Anoxybacillus	kestanbolensis	0.3	Firmicutes	Identified commensal gut microbe	AD
5	Lysinibacillus	boronitolerans	11.5	Firmicutes	Identified commensal gut microbe	AT
6	g	S	0.1	Acidobacteria	Identified in human gut microbiota	BC
7	Varibaculum	S	0.1	Actinobacteria	Identified in the gut of a premature infant	BH
3	SHD-231	S	0.1	Chloroflexi	Identified in the fecal microbiome of Gout patients	CH
9	g	S	0.1	Cyanobacteria	Identified in human gut microbiota	CJ
10	Alloiococcus	S	7.1	Firmicutes	Identified commensal gut microbe	CT
1	Christensenella	S	0.1	Firmicutes	Identified gut microbe	DC
2	g	S	0.1	Firmicutes	Commensal gut bacteria in IBD	DG
13	Blautia	S	0.1	Firmicutes	Butyrate-producing bacterial species in Gut	DH
14	Coprococcus	S	0.1	Firmicutes	Butyrate-producing bacterial species in Gut	DI
15	g	S	0.3	Gemmatimonadetes	Identified commensal gut microbe	EP
16	g	S	0.3	Lentisphaerae	Normal gut microbe	EQ
17	Comamonas	S	2.4	Proteobacteria	Identified commensal gut microbe	FL
18	g	S	0.1	Proteobacteria	Identified commensal gut microbe	FT
19	Desulfovibrio	S	0.2	Proteobacteria	Sulfate reducing bacteria in IBD	FV
20	g	S	0.3	TM7	Identified commensal gut microbe	
21	Other	Other	0.2	Firmicutes	Commensal gut bacteria in IBD	HA
22	Paracoccus	Other	0.7	Proteobacteria	Identified commensal gut microbe	HH
23	Other	Other	0.1	Proteobacteria	Identified commensal gut microbe	HN
24	Other	Other	0.7	Proteobacteria	Adult fecal microbe	НО

Specific Information of functions was adapted from NCBI Genome Database (https://www.ncbi.nlm.nih.gov/genome/).

The bacterial species that could not be identified at the genus level are mentioned as g____ and the bacterial species that could not be identified at the species level are mentioned as

0____

Differential Expression of Microbiomes in the Colon of CA and AA Patients

Figure 3 show racial differences of various bacterial phyla in adjacent healthy, UC and CC full thickness colon specimens. The tissue specimens from Caucasians represented a significantly higher proportion (p < 0.05) of the oral pathogen, *Fusobacterium*, and gut bacteria, *Parabacteroides* (Bacteroidetes). CA specimens also showed significantly higher levels (p < 0.05) of Phyla Proteobacteria including *Citrobacter*, *Hemophilus*, *Acinetobacter*, *Pseudomonas*, and *Stenotrophomonas* as compared to AA. Whereas, the AA specimens were observed to have a significantly higher proportion (p < 0.05) of *Prevotella* (Bacteroidetes) and *Clostridia* (Firmicutes) (**Figure 3**; **Table 4**).

As depicted in **Figure 3**, the adjacent healthy colon specimens, UC and CC contained $\sim 1\%$, $\sim 7\%$ and $\sim 7\%$ of sequence reads, respectively that were un-assignable to any taxon with a larger proportion of them identified in AA Colitis patients. Other major phyla observed among these specimens also include Proteobacteria (Adjacent healthy: 23.8%; UC: 26.5% and CC: 23.1%), Actinobacteria (Adjacent healthy: 6.7%; UC: 8.1% and CC: 14.1%), Fusobacteria (Adjacent healthy: 4.2%; UC: 3.6% and CC: 4.0%), and Synergistetes (Adjacent healthy: 0.2%; UC: 0.04% and CC: 1.5%). The Phylum Proteobacteria did not show

TABLE 3 | Functions and proportions of specific pathogenic Gut bacteria colonized in full thickness colon specimens.

il. No.	Bacteria genus	Bacteria species	Proportion (%)	Bacteria phylum	Function in IBD	NCBI Genom database link
	ENT HEALTHY COLO	1				
	Ochrobactrum	S	0.1	Proteobacteria	Causes early bacterial dependent induction of inducible nitric oxide synthase (iNOS) in epithelial cells in experimental colitis	EU
	Sphingomonas	S	0.2	Proteobacteria	Tissue associated intestinal microflora	FF
	Burkholderia	S	1.2	Proteobacteria	causes dysfunction of GALT and gut flora in IBD	FI
	Acinetobacter	rhizosphaerae	0.3	Proteobacteria	Identified gut bacteria in IBD	К
	Acinetobacter	lwoffii	0.3	Proteobacteria	gut bacteria in multiple sclerosis patients	W
	Stenotrophomonas	geniculata	1.2	Proteobacteria	Identified gut bacteria in IBD	AG
	Staphylococcus	sciuri	0.1	Firmicutes	Develops intestinal inflammation in acute and chronic colitis	I
	Staphylococcus	aureus	0.6	Firmicutes	Causes Crohn's disease	AU
	Lactobacillus	zeae	1.9	Firmicutes	Maintains remission of ulcerative colitis	А
)	Lactobacillus	S	0.4	Firmicutes	Maintains remission of ulcerative colitis	CW
	Lactococcus	S	0.7	Firmicutes	Used in the treatment of Crohn's disease	CX
	Pseudomonas	alcaligenes	1.8	Proteobacteria	Identified in the gut microbiota of IBD	AX
1	Pseudomonas	S	0.1	Proteobacteria	Infection in Children with Early-onset Crohn's Disease	GG
	Pseudomonas	Other	0.2	Proteobacteria	Gut microbe in children with early onset Crohn's disease	HR
	Bacillus	S	0.2	Firmicutes	Increases cytokine levels in IBD	CL
	Bacteroides	Other	0.1	Bacteroidetes	Commensal bacteria that induces colitis	GR
	Microbacterium	maritypicum	0.1	Actinobacteria	Fecal microbiome in Obesity	V
	Eggerthella	lenta	0.8	Actinobacteria	Causes bacteremia in Crohn's disease patient	AA
	Brevundimonas	diminuta	0.1	Proteobacteria	Identified in the adult fecal microbiota of allergy patients	AO
	Propionibacterium	acnes	5.3	Actinobacteria	Intestinal microbe in Liver disease	BA
	Methanobrevibacter	S	0.7	Euryarchaeota	Identified in the gut of IBD	BB
	g	S	2.4	Acidobacteria	Identified in the gut microbiome of Type 2 Diabetes patients	BD
	g	S	1.1	Actinobacteria	Identified in gut microbiota in IBD	BF
	Actinomyces	S	0.1	Actinobacteria	Identified in Abdominopelvic actinomycosis involving the GIT	BG
	Varibaculum	S	0.2	Actinobacteria	Identified in the gut of a premature infant	BH
	Microbacterium	S	0.1	Actinobacteria	Identified in the duodenum of children with ulcerative colitis	BK
	g	S	0.2	Actinobacteria	Identified in fecal microbiota of pediatric IBD patients	BP
	Atopobium	s	0.1	Actinobacteria	Altered intestinal microbiota in Crohn's disease	BR
	Slackia	s	0.2	Actinobacteria	Human gut bacteria in Multiple Sclerosis	BS
	g	s	0.1	Bacteroidetes	Characterized in intestinal biopsies in IBD patients	СВ
	g	S	0.1	Bacteroidetes	Human gut microbe in Obesity and IBD	CC
	Cloacibacterium	S	1.1	Bacteroidetes	Identified in the rectum of human colorectal adenoma patients	CG
	g	S	0.2	Cyanobacteria	Identified in the gut microbiome of IBD patients	CI
	g	s	0.6	Firmicutes	Causes microbiota dysbiosis in IBD	CO
	g	S	0.7	Firmicutes	A microbial signature of Crohn's disease	DB
	Clostridium	S	0.9	Firmicutes	Causes infection of the gut in IBD	DE
,	Dorea	s	0.1	Firmicutes	Causes dysfunction of the intestinal microbiome in IBD	DJ
	Lachnospira	S	0.1	Firmicutes	Gut bacteria in Crohn's disease patients	DK
)	Ruminococcus	s	0.1	Firmicutes	Dominant in gut microbiome of IBD patients	DO
)	g	s	0.2	Firmicutes	Gut microbe in IBD	DP

SI. No.	Bacteria genus	Bacteria species	Proportion (%)	Bacteria phylum	Function in IBD	NCBI genom database lini
1	g	s	0.6	Firmicutes	A microbial signature of Crohn's disease	DQ
2	Anaerotruncus	s	0.2	Firmicutes	Tissue associated intestinal microflora	DT
3	Oscillospira	s	0.4	Firmicutes	Gut microbe in IBD patients	DU
Ļ	Ruminococcus	s	0.7	Firmicutes	Dominant in gut microbiome of IBD patients	DV
5	g	S	0.4	Firmicutes	Gut microbe underlying the onset of IBD	DW
6	Acidaminococcus	s	1.3	Firmicutes	Gut microbe in IBD	DX
7	Phascolarctobacterium	S	1.9	Firmicutes	Causes dysfunction of the intestinal microbiome in IBD	DZ
3	Schwartzia	s	0.5	Firmicutes	causes fecal microbial dysbiosis in IBD	EA
	g	s	0.2	Firmicutes	A microbial signature of Crohn's disease	EC
	Anaerococcus	s	1.3	Firmicutes	Microbe in Inflammatory Pouch Complications	EE
	Finegoldia	S	0.3	Firmicutes	Intestinal microbe in colorectal cancer	EF
	g	S	0.3	Firmicutes	Gut microbe in GI diseases	EI
	Bulleidia	S	0.1	Firmicutes	Fecal-associated and mucosalassociated microbiota in irritable bowel syndrome patients	EJ
	Coprobacillus	s	0.3	Firmicutes	Alters Gut Microbiota in Psoriatic Arthritis	EL
	Leptotrichia	S	0.7	Fusobacteria	Causes gut mucosal inflammation in Rheumatoid arthritis patients	EO
;	9	S	4.2	Proteobacteria	Intestinal microbe in children with severe and complicated acute viral gastroenteritis	EV
	Methylobacterium	S	0.1	Proteobacteria	Causes microbial dysbiosis in pediatric Crohn's disease	EW
	9	S	0.1	Proteobacteria	Intestinal microbe in children with severe and complicated acute viral gastroenteritis	EX
	g	S	1.2	Proteobacteria	Involved in host-microbial cross talk in IBD	FG
	Lautropia	s	0.3	Proteobacteria	causes fecal microbial dysbiosis in IBD	FJ
	g	S	0.1	Proteobacteria	Fecal and mucosa associated microbe in IBD	FK
	Citrobacter	S	0.1	Proteobacteria	Gut microbe in newly diagnosed with treatment-naïve Crohn's disease patients	FY
3	Halomonas	s	1.4	Proteobacteria	Intestinal microflora in chronic kidney disease	GB
	g	S	0.1	Proteobacteria	Microbe in colon tissue from IBD subjects	GE
	g	S	0.1	Proteobacteria	bacteria in human Ulcerative Colitis patients	GH
	Other	Other	0.1	Actinobacteria	Alters fecal microbiota in pediatric IBD patients	GO
	Other	Other	3.6	Firmicutes	gut microbe in experimental colitis	GT
	Other	Other	12.5	Firmicutes	Fecal and mucosa associated microbe in IBD	GW
	Weissella	Other	0.2	Firmicutes	Gut microbe in IBD patients	GX
)	Other	Other	0.1	Proteobacteria	Fecal and mucosa associated microbe in IBD	HL
	Other	Other	0.1	Proteobacteria	Involved in host-microbial cross talk in IBD	HM
SEA	SED COLON (ULCERATI	VE COLITIS)				
	Ochrobactrum	S	0.1	Proteobacteria	Causes early bacterial dependent induction of inducible nitric oxide synthase (INOS) in epithelial cells in experimental colitis	EU
	Delftia	s	0.1	Proteobacteria	Fecal and mucosa associated microbe in IBD	FM
	Sphingomonas	s	0.5	Proteobacteria	Tissue associated intestinal microflora	FF
	Burkholderia	s	0.2	Proteobacteria	Causes dysfunction of GALT and gut flora in IBD	FI
	Acinetobacter	 rhizosphaerae	0.7	Proteobacteria	Identified gut microbe in IBD	K
	Acinetobacter	lwoffii	0.1	Proteobacteria	Gut bacteria in multiple sclerosis patients	W
	Acinetobacter	s	0.5	Proteobacteria	Tissue associated intestinal microflora	GF
	Stenotrophomonas	geniculata	0.1	Proteobacteria	Identified gut microbe in IBD	AG
	Enterococcus	s	0.8	Firmicutes	Induces experimental IBD	CV
)	Staphylococcus	sciuri	0.0	Firmicutes	Develops intestinal inflammation in acute and	
/	Capityloooous	Soluti	0.1	1 111100100	chronic colitis	

SI. No.	Bacteria genus	Bacteria species	Proportion (%)	Bacteria phylum	Function in IBD	NCBI genome database link
1	Staphylococcus	aureus	0.6	Firmicutes	Causes Crohn's disease	AU
2	Lactobacillus	zeae	1.9	Firmicutes	Maintains remission of ulcerative colitis	А
3	Lactobacillus	S	0.4	Firmicutes	Maintains remission of ulcerative colitis	CW
1	Lactococcus	S	0.7	Firmicutes	used in the treatment of Crohn's disease	CX
5	Pseudomonas	alcaligenes	1.8	Proteobacteria	Identified in the gut microbiota of IBD	AX
6	Pseudomonas	S	0.1	Proteobacteria	Infection in Children with Early-onset Crohn's Disease	GG
7	Pseudomonas	Other	0.2	Proteobacteria	Gut microbe in children with early onset Crohn's disease	HR
3	Bacillus	S	0.1	Firmicutes	Increases cytokine levels in IBD	CL
	Bacteroides	caccae	0.1	Bacteroidetes	Identified in the gut of ulcerative colitis patients	AS
	Bacteroides	Other	0.2	Bacteroidetes	Commensal bacteria that induces colitis	GR
	Blautia	producta	0.1	Firmicutes	Gut microbe in Obesity and IBD	Ν
	Faecalibacterium	prausnitzii	0.1	Firmicutes	Gut microbe in Crohn's disease patients	0
	Microbacterium	maritypicum	0.1	Actinobacteria	Fecal microbiome in Obesity	V
	Eggerthella	lenta	5.1	Actinobacteria	Causes bacteremia in Crohn's disease patient	AA
	Propionibacterium	acnes	2.8	Actinobacteria	Intestinal microbe in Liver disease	BA
	Methanobrevibacter	S	0.3	Euryarchaeota	Identified in the gut of IBD	BB
7	g	S	1.8	Acidobacteria	Identified in the gut microbiome of Type 2 Diabetes patients	BD
	g	S	0.3	Actinobacteria	Identified in gut microbiota in IBD	BF
	Adlercreutzia	S	0.2	Actinobacteria	Causes dysbiosis in IBD patients	BQ
	Slackia	s	0.2	Actinobacteria	Alters human gut microbiome in Multiple Sclerosis	BS
	g	s	0.8	Bacteroidetes	Identified in gut microbiome of IBD patients	CA
	g	s	0.4	Bacteroidetes	Characterized in intestinal biopsies in IBD patients	CB
	g	s	0.1	Bacteroidetes	Human gut microbe in Obesity and IBD	CC
	Cloacibacterium	s	0.4	Bacteroidetes	Identified in the rectum of human colorectal adenoma patients	CG
5	g	S	0.1	Firmicutes	Causes microbiota dysbiosis in IBD	CO
	g	s	0.1	Firmicutes	Gut microbe in IBD	DA
	g	s	1.7	Firmicutes	A microbial signature of Crohn's disease	DB
	Clostridium	s	0.8	Firmicutes	Causes infection of the gut in IBD	DE
)	Dorea	S	0.1	Firmicutes	Causes dysfunction of the intestinal microbiome in IBD	DJ
)	Lachnospira	S	0.3	Firmicutes	gut bacteria in Crohn's disease patients	DK
	Ruminococcus	S	0.1	Firmicutes	Dominant in gut microbiome of IBD patients	DO
	g	S	0.3	Firmicutes	A microbial signature of Crohn's disease	DQ
	Oscillospira	s	0.1	Firmicutes	Gut microbe in IBD patients	DU
	Ruminococcus	s	0.9	Firmicutes	Dominant in gut microbiome of IBD patients	DV
	g	s	0.3	Firmicutes	Gut microbe underlying the onset of IBD	DW
	Acidaminococcus	s	1.1	Firmicutes	Gut microbe in IBD	DX
	Phascolarctobacterium	S	2.2	Firmicutes	Causes dysfunction of the intestinal microbiome in IBD	DZ
	Schwartzia	s	0.5	Firmicutes	Causes fecal microbial dysbiosis in IBD	EA
	Anaerococcus	s	0.6	Firmicutes	Microbe in Inflammatory Pouch Complications	EE
	Finegoldia	s	0.3	Firmicutes	Intestinal microbe in colorectal cancer	EF
	g	s	0.6	Firmicutes	Gut microbe in GI diseases	El
2	Bulleidia	s	0.1	Firmicutes	Fecal-associated and mucosalassociated microbiota in irritable bowel syndrome patients	EJ
3	Coprobacillus	s	0.1	Firmicutes	Alters Gut Microbiota in Psoriatic Arthritis	EL
ļ	Leptotrichia	S	0.5	Fusobacteria	Causes gut mucosal inflammation in Rheumatoid arthritis patients	EO

	NCBI genome database link
hildren with severe and al gastroenteritis	EV
nmation in IBD	FB
ciated with postoperative	/e FD
bial cross talk in IBD	FG
mental colitis	FH
al dysbiosis in IBD	FJ
sociated microbe in IBD	D FK
diagnosed with n's disease patients	FY
chronic kidney disease	e GB
cerative Colitis patients	GH
a in pediatric IBD patients	nts GO
Crohn's disease patient	nt GQ
mental colitis	GT
sociated microbe in IBD	D GW
atients	GX
biosis in pediatric Crohn's	n's HD
sociated microbe in IBD	D HL
l dependent induction of synthase (iNOS) in epithelia colitis	
estinal microflora	FF
f GALT and gut flora in IBE	IBD FI
in IBD	К
le sclerosis patients	W
estinal microflora in colitis	tis HQ
in IBD	AG
IBD	CV
lammation in acute and	I k
ase	AU
f ulcerative colitis	А
f ulcerative colitis	CW
of Crohn's disease	CX
icrobiota of IBD	AX
vith Early-onset Crohn's	GG GG
en with early onset Crohn's	nn's HR
ora of the gut	E
vels in IBD	CL
се	AJ
that induces colitis	GR
Dbesity	V
Crohn's disease patient	nt AA
fecal microbiota of allergy	

SI. No.	Bacteria genus	Bacteria species	Proportion (%)	Bacteria phylum	Function in IBD	NCBI genome database link
24	Propionibacterium	acnes	1.2	Actinobacteria	Intestinal microbe in Liver disease	BA
25	Methanobrevibacter	S	0.8	Euryarchaeota	Identified in the gut of IBD patients	BB
26	g	S	0.7	Acidobacteria	Identified in the gut microbiome of Type 2 Diabetes patients	BD
27	g	S	0.2	Actinobacteria	Identified in gut microbiota in IBD	BE
28	g	S	0.5	Actinobacteria	Identified in gut microbiota in IBD	BF
29	Microbacterium	S	0.1	Actinobacteria	Identified in the duodenum of children with ulcerative colitis	BK
30	Bifidobacterium	S	0.2	Actinobacteria	Identified in gut microbiota of IBD patients	BO
31	g	S	0.3	Actinobacteria	Identified in fecal microbiota of pediatric IBD patients	BP
32	Atopobium	S	0.2	Actinobacteria	Altered intestinal microbiota in Crohn's disease	BR
33	Slackia	S	1.3	Actinobacteria	Alters human gut microbiome in Multiple Sclerosis	BS
34	g	S	0.1	Bacteroidetes	Identified in gut microbiome of IBD patients	CA
35	g	S	0.3	Bacteroidetes	Human gut microbe in Obesity and IBD	CC
36	g	S	0.7	Firmicutes	Causes microbiota dysbiosis in IBD	CO
37	g	S	0.2	Firmicutes	Gut microbe in IBD	DA
38	g	S	0.9	Firmicutes	A microbial signature of Crohn's disease	DB
39	Clostridium	S	0.6	Firmicutes	Causes infection of the gut in IBD	DE
40	Lachnospira	s	0.6	Firmicutes	Gut bacteria in Crohn's disease patients	DK
41	Moryella	S	0.1	Firmicutes	Microbe in Inflammatory Pouch Complications	DL
42	g	s	0.1	Firmicutes	gut microbe in IBD	DP
43	g	S	0.6	Firmicutes	A microbial signature of Crohn's disease	DQ
44	Oscillospira	S	0.2	Firmicutes	Gut microbe in IBD patients	DU
45	Ruminococcus	S	0.9	Firmicutes	Dominant in gut microbiome of IBD patients	DV
46	g	s	0.2	Firmicutes	Gut microbe underlying the onset of IBD	DW
47	Acidaminococcus	S	1.0	Firmicutes	Gut microbe in IBD	DX
48	Phascolarctobacterium	s	0.6	Firmicutes	Causes dysfunction of the intestinal microbiome in IBD	DZ
49	Schwartzia	S	0.2	Firmicutes	Causes fecal microbial dysbiosis in IBD	EA
50	g	S	0.1	Firmicutes	A microbial signature of Crohn's disease	EC
51	Anaerococcus	S	0.3	Firmicutes	Microbe in Inflammatory Pouch Complications	EE
52	Finegoldia	S	0.4	Firmicutes	Intestinal microbe in colorectal cancer	EF
53	g	s	0.5	Firmicutes	Gut microbe in GI diseases	El
54	Bulleidia	S	0.2	Firmicutes	Fecal-associated and mucosalassociated microbiota in irritable bowel syndrome patients	EJ
55	Coprobacillus	S	0.3	Firmicutes	Alters Gut Microbiota in Psoriatic Arthritis	EL
56	Leptotrichia	s	0.5	Fusobacteria	Causes gut mucosal inflammation in Rheumatoid arthritis patients	EO
57	g	s	4.1	Proteobacteria	Intestinal microbe in children with severe and complicated acute viral gastroenteritis	EV
58	g	s	0.2	Proteobacteria	Microbial factor associated with postoperative Crohn's disease	FD
59	g	S	1.2	Proteobacteria	Involved in host-microbial cross talk in IBD	FG
60	Sutterella	s	0.1	Proteobacteria	Gut microbe in experimental colitis	FH
61	Lautropia	s	0.1	Proteobacteria	Causes fecal microbial dysbiosis in IBD	FJ
62	g	s	0.4	Proteobacteria	Fecal and mucosa associated microbe in IBD	FK
63	g	s	0.1	Proteobacteria	Bacteria in Mucosal and Submucosal Intestinal Tissues in Advanced Crohn's Disease	FN
64	Ralstonia	S	0.1	Proteobacteria	Microbiota in the Mucosa of Patients With Ulcerative Colitis	FP
65	Halomonas	S	0.5	Proteobacteria	Intestinal microflora in chronic kidney disease	GB

SI. No.	Bacteria genus	Bacteria species	Proportion (%)	Bacteria phylum	Function in IBD	NCBI genome database link
66	Haemophilus	S	0.7	Proteobacteria	Treatment naïve microbiome in new onset Crohn's disease	GD
67	g	S	0.1	Proteobacteria	Microbe in colon tissue from IBD subjects	GE
68	Other	Other	1.1	Actinobacteria	Alters fecal microbiota in pediatric IBD patients	GO
69	Eggerthella	Other	0.1	Actinobacteria	Causes bacteremia in Crohn's disease patient	GQ
70	Other	Other	1.7	Firmicutes	Gut microbe in experimental colitis	GT
71	Other	Other	2.2	Firmicutes	Fecal and mucosa associated microbe in IBD	GW
72	Weissella	Other	2.2	Firmicutes	Gut microbe in IBD patients	GX
73	Other	Other	0.1	Proteobacteria	Causes microbial dysbiosis in pediatric Crohn's disease	HD
74	Methylobacterium	Other	0.1	Proteobacteria	Causes gut microbial dysbiosis in pediatric Crohn's disease patients	HG
75	Other	Other	0.2	Proteobacteria	Fecal and mucosa associated microbe in IBD	HK
76	Other	Other	0.6	Proteobacteria	Fecal and mucosa associated microbe in IBD	HL

Specific Information of functions was adapted from NCBI Genome Database (https://www.ncbi.nlm.nih.gov/genome/).

The bacterial species that could not be identified at the genus level are mentioned as g___ and the bacterial species that could not be identified at the species level are mentioned as s___.

any significant difference between healthy colon specimens and diseased colon specimens (Table 4).

Bacterial Species Identified in a Significantly Higher Proportion in Diseased Colon Tissues

As shown in **Figure 3**, diseased colon specimens represented a significantly higher proportion (p < 0.05) of gut bacteria belonging to Phylum Firmicutes including *Blautia producta*, *Faecalibacterium prausnitzii*, *Anoxybacillus kestanbolensis*, *Ruminococcus gnavus*, *Eubacterium dolichum*, *Lysinibacillus boronitolerans*, and oral bacteria including *Staphylococcus sciuri*, *Staphylococcus aureus*, *Streptococcus anginosus*.

In contrast, healthy colon specimens were significantly dominated (p < 0.05) by oral bacteria belonging to Phylum Actinobacteria that includes; *Corynebacterium kroppenstedtii*, *Corynebacterium durum*. Additionally, healthy colon specimens were dominated by gut bacteria belonging to Phylum Actinobacteria that includes; *Colinsella stercoris*, *Colinsella aerofaciens*, *Kocuria rhizophila*, *Eggerthella lenta*, *Propionibacterium granulosum*, *Propionibacterium acnes*, *Actinomyces europaeus*, *Rothia dentocariosa*, and Phylum Bacteroidetes that includes; *Bacteroides fragilis*, *Bacteroides eggerthii*, *Bacteroides caccae*, *Parabacteroides distasonis* (**Figure 3**).

Alpha Diversity and Beta Diversity Analyses

Alpha diversity and beta diversity metrics were computed to analyse the diversity of bacterial species within each sample and between samples. To assess our sampling efficiency, we plotted rarefaction curves (Chao1 and Shannon) for all 39 specimens. Increased diversity (Shannon) in the diseased samples compared to control samples was observed. From the rarefaction curves, it is evident that most AA samples require additional sampling whereas Caucasian samples do not (data not shown).

Since, outliers exhibiting different microbiome profiles were observed both in the healthy and disease groups, we performed principle coordinate analysis (PCoA analysis) and hierarchial clustering to obtain a holistic view of the microbiome profile in each sample. Two dimensional PCoA plots revealed that control samples which had similar microbiome profiles as suggested by histograms and OTU heat map clustered together (data not shown).

Pathogenic Oral and Gut Flora Abundantly Colonized in Diseased Colon Specimens

The pathogenic oral bacteria identified abundantly in diseased colon specimens as compared to healthy colon specimens were Porphyromonas, Prevotella, Gemella, Staphylococcus, Streptococcus, Abiotrophia, Granulicatella, Lactobacillus, Veillonella, Lactococcus, Peptostreptococcus, Selenomonas, Parvimonas, Eubacterium, Fusobacterium. Pseudomonas, Aggregatibacter, and Corynebacterium (Table 1).

Pathogenic gut bacteria identified abundantly in diseased colon specimens as compared to healthy colon specimens include Ochrobactrum, Delftia, Sphingomonas, Burkholderia, Acinetobacter, Stenotrophomonas, Enterococcus, Granulicatella, Staphylococcus, Streptococcus, Lactobacillus, Lactococcus, Pseudomonas, Bacillus, Campylobacter, and Bacteroides (**Table 3**).

DISCUSSION

Our study demonstrates significant perturbations among bacteria belonging to Phyla Bacteroidetes and Firmicutes in fullthickness diseased colon specimens containing neuromuscular compartment (**Figure 2**). Our studies further show that the proportion of pathogenic bacteria are higher in diseased TABLE 4 | Functions and Proportions of bacterial species identified in the full thickness human colon specimens of Caucasians and African Americans.

SI. No.	Bacteria genus	Bacteria species	Proportion (%)	Bacteria phylum	Function in IBD	NCBI genome database link
CAUCA	SIAN AMERICANS					
	Lactobacillus	zeae	6.8	Firmicutes	Maintains remission of ulcerative colitis	A
	Bacillus	thermoamylovorans	0.1	Firmicutes	A probiotic- normal flora of the gut	E
	Prevotella	tannerae	0.1	Bacteroidetes	Prevalent in colitis	F
	Collinsella	stercoris	0.0	Actinobacteria	Used for treatment of IBD	G
	Prevotella	stercorea	1.4	Bacteroidetes	Alters mucosal microbiota in the colon of patients with IBD	Н
	Staphylococcus	sciuri	0.1	Firmicutes	Develops intestinal inflammation in acute and chronic colitis	Ι
	Shuttleworthia	satelles	0.0	Firmicutes	Identified in the human ileum	J
	Acinetobacter	rhizosphaerae	1.1	Proteobacteria	Identified gut microbe in IBD	К
	Blautia	producta	0.1	Firmicutes	Gut microbe in Obesity and IBD	Ν
)	Akkermansia	muciniphila	0.1	Verrucomicrobia	Adheres to enterocytes and strengthens the integrity of the epithelial cell layer	S
1	Prevotella	melaninogenica	0.2	Bacteroidetes	Gut microbiome biomarker in ankylosing spondylitis	U
0	Acinetobacter	lwoffii	0.2	Proteobacteria	Gut bacteria in multiple sclerosis patients	W
3	Bifidobacterium	longum	0.2	Actinobacteria	Attenuates acute murine experimental model of IBD	Y
1	Eggerthella	lenta	4.3	Actinobacteria	Causes bacteremia in Crohn's disease patient	AA
5	Rhizobium	leguminosarum	0.1	Proteobacteria	Identified gut microbe in IBD patients	AB
6	Anoxybacillus	kestanbolensis	0.1	Firmicutes	Identified gut microbe in IBD patients	AD
,	Stenotrophomonas	geniculata	0.1	Proteobacteria	Identified gut microbe in IBD patients	AG
3	Corynebacterium	durum	0.1	Actinobacteria	Identified gut microbe in IBD patients	AK
,	Eubacterium	dolichum	0.8	Firmicutes	Causes dysbiosis of the intestinal microbiota	AL
)	Brevundimonas	diminuta	0.1	Proteobacteria	Identified in the adult fecal microbiota of allergy patients	AO
1	Lysinibacillus	boronitolerans	12.3	Firmicutes	Identified gut microbe in IBD patients	AT
2	Staphylococcus	aureus	0.6	Firmicutes	Causes Crohn's disease	AU
3	Peptostreptococcus	anaerobius	4.8	Firmicutes	Causes dysbiosis in IBD	AW
ļ	Pseudomonas	alcaligenes	1.0	Proteobacteria	Identified in the gut microbiota of IBD	AX
5	Propionibacterium	acnes	3.9	Actinobacteria	Intestinal microbe in Liver disease	BA
, ;	Methanobrevibacter		0.6	Euryarchaeota	Identified in the gut of IBD	BB
7	g	s s	0.8	Acidobacteria	Identified in the gut microbiome of Type 2 Diabetes patients	BD
3	g	S	0.9	Actinobacteria	Identified in gut microbiota in IBD	BF
)	9 Varibaculum	s	0.1	Actinobacteria	Identified in the gut of a premature infant	BH
)	Corynebacterium	S	0.7	Actinobacteria	Causes experimental colitis	BI
1	Microbacterium	s	0.1	Actinobacteria	Identified in the duodenum of children with ulcerative colitis	BK
2	Bifidobacterium	S	0.1	Actinobacteria	identified in gut microbiota of IBD patients	во
3	g	s	0.2	Actinobacteria	Identified in fecal microbiota of pediatric IBD patients	BP
1	9 Adlercreutzia	s	0.2	Actinobacteria	Causes dysbiosis in IBD patients	BQ
5	Atopobium	s	0.1	Actinobacteria	altered intestinal microbiota in Crohn's disease	BR
5	Slackia	s	0.7	Actinobacteria	Alters human gut microbiome in Multiple Sclerosis	BS
7	Prevotella		0.1	Bacteroidetes	A microbial signature of Crohn's disease	BZ
3		s	0.1	Bacteroidetes	Identified in gut microbiome of IBD patients	CA
	g	s	0.4	Bacteroidetes		CB
)	g	s	0.2	Bacteroidetes	Characterized in intestinal biopsies in IBD patients Human out microbe in Obesity and IBD	
)	g	s			, ,	CC
1	Cloacibacterium	S	0.7	Bacteroidetes	Identified in the rectum of human colorectal adenoma patients	CG
2	SHD-231	S	0.1	Chloroflexi	Identified in the fecal microbiome of Gout patients	CH
3	g	S	0.1	Cyanobacteria	Identified in the gut microbiome of IBD patients	CI

SI. No.	Bacteria genus	Bacteria species	Proportion (%)	Bacteria phylum	Function in IBD	NCBI genome database link
14	Calothrix	S	0.1	Cyanobacteria	Identified gut microbe in IBD patients	CK
5	Bacillus	s	0.5	Firmicutes	Increases cytokine levels in IBD	CL
5	g	s	0.6	Firmicutes	Causes microbiota dysbiosis in IBD	CO
7	Gemella	s	0.1	Firmicutes	Microbiome in New-Onset Crohn's Disease	CP
3	Abiotrophia	s	0.1	Firmicutes	Causes fecal microbial dysbiosis in IBD	CS
9	Alloiococcus	s	3.4	Firmicutes	Identified gut microbe in IBD patients	CT
)	Enterococcus	s	0.4	Firmicutes	Induces experimental IBD	CV
I	Lactobacillus	s	0.5	Firmicutes	Maintains remission of ulcerative colitis	CW
2	Lactococcus	s	0.7	Firmicutes	Used in the treatment of Crohn's disease	CX
3	g	s	0.1	Firmicutes	Gut microbe in IBD	DA
1	g	s	1.1	Firmicutes	A microbial signature of Crohn's disease	DB
5	Christensenella	s	1.2	Firmicutes	Identified gut microbe	DC
6	Clostridium	s	0.9	Firmicutes	causes infection of the gut in IBD	DE
7	Pseudoramibacter_ Eubacterium	S	1.8	Firmicutes	Metabolizes Linoleic acid in the Gut	DF
3	g	s	0.1	Firmicutes	Commensal gut bacteria in IBD	DG
9	Blautia	s	0.3	Firmicutes	Butyrate-producing bacterial species in Gut	DH
)	Coprococcus	s	0.2	Firmicutes	Butyrate-producing bacterial species in Gut	DI
I	Dorea	S	0.1	Firmicutes	Causes dysfunction of the intestinal microbiome in IBD	DJ
2	Lachnospira	s	0.3	Firmicutes	Gut bacteria in Crohn's disease patients	DK
3	g	s	0.1	Firmicutes	Gut microbe in IBD	DP
Ļ	g	s	0.6	Firmicutes	A microbial signature of Crohn's disease	DQ
5	Peptostreptococcus	s	0.3	Firmicutes	Causes gut microbiota dysbiosis in IBD	DR
6	Anaerotruncus	s	0.1	Firmicutes	Tissue associated intestinal microflora	DT
,	Oscillospira	s	0.2	Firmicutes	Gut microbe in IBD patients	DU
3	Ruminococcus	s	0.4	Firmicutes	Dominant in gut microbiome of IBD patients	DV
9	g	s	0.3	Firmicutes	Gut microbe underlying the onset of IBD	DW
)	Acidaminococcus	s	1.2	Firmicutes	Gut microbe in IBD	DX
	Phascolarctobacterium	S	1.9	Firmicutes	Causes dysfunction of the intestinal microbiome in IBD	DZ
2	Schwartzia	s	0.5	Firmicutes	Causes fecal microbial dysbiosis in IBD	EA
3	Selenomonas	s	0.2	Firmicutes	Causes dysbiosis in colorectal cancer	EB
Ļ	g	s	0.1	Firmicutes	A microbial signature of Crohn's disease	EC
5	Anaerococcus	s	0.7	Firmicutes	Microbe in Inflammatory Pouch Complications	EE
6	Finegoldia	s	0.3	Firmicutes	Intestinal microbe in colorectal cancer	EF
,	g	s	0.4	Firmicutes	Gut microbe in GI diseases	El
3	Bulleidia	s	0.1	Firmicutes	Fecal-associated and mucosalassociated microbiota in irritable bowel syndrome patients	EJ
)	Coprobacillus	s	0.3	Firmicutes	Alters Gut Microbiota in Psoriatic Arthritis	EL
)	Fusobacterium	s	0.8	Fusobacteria	Identified from colonic biopsies of IBD patients	EN
	Leptotrichia	S	0.2	Fusobacteria	Causes gut mucosal inflammation in Rheumatoid arthritis patients	EO
2	g	s	0.3	Gemmatimonadetes	Identified gut microbe in IBD patients	EP
3	g	s	0.4	Lentisphaerae	Normal gut microbe	EQ
1	Ochrobactrum	S	0.1	Proteobacteria	Causes early bacterial dependent induction of inducible nitric oxide synthase (iNOS) in epithelial cells in experimental colitis	EU
5	g	S	4.7	Proteobacteria	Intestinal microbe in children with severe and complicated acute viral gastroenteritis	EV
6	g	s	0.2	Proteobacteria	Causes chronic inflammation in IBD	FB

SI. No.	Bacteria genus	Bacteria species	Proportion (%)	Bacteria phylum	Function in IBD	NCBI genom database linl
37	g	S	0.3	Proteobacteria	Identified gut microbe in IBD patients	FC
8	9	s	0.1	Proteobacteria	Microbial factor associated with postoperative Crohn's disease	FD
9	Sphingomonas	S	0.3	Proteobacteria	Tissue associated intestinal microflora	FF
)	g	S	1.7	Proteobacteria	Involved in host-microbial cross talk in IBD	FG
	Sutterella	S	0.1	Proteobacteria	gut microbe in experimental colitis	FH
2	Burkholderia	S	1.2	Proteobacteria	Causes dysfunction of GALT and gut flora in IBD	FI
3	Lautropia	S	0.2	Proteobacteria	Causes fecal microbial dysbiosis in IBD	FJ
1	g	S	0.4	Proteobacteria	Fecal and mucosa associated microbe in IBD	FK
5	Comamonas	S	2.5	Proteobacteria	Identified gut microbe in IBD patients	FL
6	Delftia	S	0.1	Proteobacteria	Fecal and mucosa associated microbe in IBD	FM
7	Desulfovibrio	s	0.2	Proteobacteria	Sulfate reducing bacteria in IBD	FV
3	Citrobacter	s	0.2	Proteobacteria	Gut microbe in newly diagnosed with treatment-naïve Crohn's disease patients	FY
9	Halomonas	S	0.9	Proteobacteria	Intestinal microflora in chronic kidney disease	GB
00	Aggregatibacter	s	0.6	Proteobacteria	Causes fungal microbiota dysbiosis in IBD	GC
01	Haemophilus	S	0.3	Proteobacteria	Treatment naïve microbiome in new onset Crohn's disease	GD
02	Pseudomonas	S	0.8	Proteobacteria	Infection in Children with Early-onset Crohn's Disease	GG
03	g	S	0.2	Proteobacteria	Bacteria in human Ulcerative Colitis patients	GH
04	g	s	0.1	TM7	No role in IBD	
05	Other	Other	0.5	Actinobacteria	Alters fecal microbiota in pediatric IBD patients	GO
06	Eggerthella	Other	0.1	Actinobacteria	Causes bacteremia in Crohn's disease patient	GQ
07	Bacteroides	Other	0.1	Bacteroidetes	Commensal bacteria that induces colitis	GR
08	Prevotella	Other	0.7	Bacteroidetes	A microbial signature of Crohn's disease	GS
09	Other	Other	3.2	Firmicutes	Gut microbe in experimental colitis	GT
10	Other	Other	7.5	Firmicutes	Fecal and mucosa associated microbe in IBD	GW
11	Weissella	Other	1.5	Firmicutes	Gut microbe in IBD patients	GX
12	Other	Other	0.1	Firmicutes	Commensal gut bacteria in IBD	HA
13	Other	Other	1.1	Proteobacteria	Causes microbial dysbiosis in pediatric Crohn's disease	HD
14	Paracoccus	Other	0.3	Proteobacteria	Identified gut microbe in IBD patients	HH
15	Other	Other	0.1	Proteobacteria	Fecal and mucosa associated microbe in IBD	HK
16	Other	Other	0.7	Proteobacteria	Fecal and mucosa associated microbe in IBD	HL
17	Other	Other	0.1	Proteobacteria	Identified gut microbe in IBD patients	HN
18	Other	Other	0.5	Proteobacteria	Adult fecal microbe	HO
	AN AMERICANS					
	Lactobacillus	zeae	1.1	Firmicutes	Maintains remission of ulcerative colitis	A
	Prevotella	stercorea	4.6	Bacteroidetes	Alters mucosal microbiota in the colon of patients with IBD	Н
	Shuttleworthia	satelles	0.4	Firmicutes	Identified in the human ileum	J
	Acinetobacter	rhizosphaerae	0.2	Proteobacteria	Identified gut microbe in IBD patients	K
	Microbacterium	maritypicum	0.2	Actinobacteria	Fecal microbiome in Obesity	V
	Acinetobacter	lwoffii	0.5	Proteobacteria	Gut bacteria in multiple sclerosis patients	W
	Eggerthella	lenta	0.7	Actinobacteria	Causes bacteremia in Crohn's disease patient.	AA
	Stenotrophomonas	geniculata	2.0	Proteobacteria	Identified gut microbe in IBD patients	AG
	Corynebacterium	durum	0.1	Actinobacteria	Identified gut microbe in IBD patients	AK
0	Eubacterium	dolichum	0.5	Firmicutes	Causes dysbiosis of the intestinal microbiota	AL
1	Brevundimonas	diminuta	0.1	Proteobacteria	Identified in the adult fecal microbiota of allergy patients	AO

SI. No.	Bacteria genus	Bacteria species	Proportion (%)	Bacteria phylum	Function in IBD	NCBI genom database lin
12	Lysinibacillus	boronitolerans	5.0	Firmicutes	Identified gut microbe in IBD patients	AT
13	Staphylococcus	aureus	0.2	Firmicutes	Causes Crohn's disease	AU
4	Peptostreptococcus	anaerobius	24.9	Firmicutes	Causes dysbiosis in IBD	AW
5	Pseudomonas	alcaligenes	1.1	Proteobacteria	Identified in the gut microbiota of IBD	AX
5	Propionibacterium	acnes	0.6	Actinobacteria	Intestinal microbe in Liver disease	BA
7	Methanobrevibacter	s	0.4	Euryarchaeota	Identified in the gut of IBD	BB
3	g	s	0.2	Acidobacteria	Identified in human gut microbiota	BC
9	g	S	4.3	Acidobacteria	Identified in the gut microbiome of Type 2 Diabetes patients	BD
)	g	s	0.3	Actinobacteria	Identified in gut microbiota in IBD	BE
	Varibaculum	s	0.6	Actinobacteria	Identified in the gut of a premature infant	BH
	Corynebacterium	s	0.5	Actinobacteria	Causes experimental colitis	BI
	Arthrobacter	s	0.2	Actinobacteria	Fecal microflora in chronic IBD patients	BL
	Slackia	s	0.2	Actinobacteria	Alters human gut microbiome in Multiple Sclerosis	BS
	Chryseobacterium	s	0.3	Bacteroidetes	Fecal and mucosa associated microbe in IBD	CF
	Cloacibacterium	S	0.2	Bacteroidetes	Identified in the rectum of human colorectal adenoma patients	CG
	g	s	0.2	Cyanobacteria	Identified in human gut microbiota	CJ
	Bacillus	S	2.7	Firmicutes	Increases cytokine levels in IBD	CL
	Alloiococcus	s	6.0	Firmicutes	Identified gut microbe in IBD patients	CT
	Lactobacillus	S	0.2	Firmicutes	Maintains remission of ulcerative colitis	CW
	Lactococcus	s	0.3	Firmicutes	Used in the treatment of Crohn's disease	CX
	g	 S	0.1	Firmicutes	Gut microbe in IBD	DA
	9	s	1.3	Firmicutes	A microbial signature of Crohn's disease	DB
	Christensenella	s	0.2	Firmicutes	Identified gut microbe	DC
	Clostridium	s	0.4	Firmicutes	causes infection of the gut in IBD	DE
	Pseudoramibacter_Eubac		0.9	Firmicutes	Metabolizes Linoleic acid in the Gut	DF
	Lachnospira	s	0.3	Firmicutes	Gut bacteria in Crohn's disease patients	DK
	g	s	0.3	Firmicutes	A microbial signature of Crohn's disease	DQ
	9— Peptostreptococcus	s	0.4	Firmicutes	Causes gut microbiota dysbiosis in IBD	DR
	Oscillospira	s	0.2	Firmicutes	Gut microbe in IBD patients	DU
	Ruminococcus	s	2.2	Firmicutes	Dominant in gut microbiome of IBD patients	DV
	g	s	0.4	Firmicutes	Gut microbe underlying the onset of IBD	DW
	9 Acidaminococcus	s	1.0	Firmicutes	Gut microbe in IBD	DX
	Phascolarctobacterium	s	0.5	Firmicutes	Causes dysfunction of the intestinal microbiome in IBD	DZ
	Schwartzia	S	0.1	Firmicutes	Causes fecal microbial dysbiosis in IBD	EA
	Selenomonas	s	0.5	Firmicutes	Causes dysbiosis in colorectal cancer	EB
	Anaerococcus		0.5	Firmicutes	Microbe in Inflammatory Pouch Complications	EE
	Finegoldia	S	0.5	Firmicutes	Intestinal microbe in colorectal cancer	EF
	0	S	0.4	Firmicutes	Gut microbe in Gl diseases	EF
	9 Bulleidia	s s	0.1	Firmicutes	Fecal-associated and mucosalassociated microbiota in irritable bowel syndrome patients	EJ
	Fusobacterium	S	8.4	Fusobacteria	Identified from colonic biopsies of IBD patients	EN
	Leptotrichia	s	1.7	Fusobacteria	Causes gut mucosal inflammation in Rheumatoid arthritis patients	EO
	g	S	0.7	Gemmatimonadetes	Identified gut microbe in IBD patients	EP
3	9 Ochrobactrum	ss	0.1	Proteobacteria	Causes early bacterial dependent induction of inducible nitric oxide synthase (iNOS) in epithelial cells in experimental colitis	EU
5	g	s	1.5	Proteobacteria	Intestinal microbe in children with severe and complicated acute viral gastroenteritis	EV

SI. No.	Bacteria genus	Bacteria species	Proportion (%)	Bacteria phylum	Function in IBD	NCBI genome database link
56	g	S	0.1	Proteobacteria	Involved in host-microbial cross talk in IBD	FG
57	Sutterella	s	0.2	Proteobacteria	Gut microbe in experimental colitis	FH
58	Burkholderia	S	0.4	Proteobacteria	Causes dysfunction of GALT and gut flora in IBD	FI
59	Lautropia	S	0.2	Proteobacteria	Causes fecal microbial dysbiosis in IBD	FJ
60	Comamonas	s	0.4	Proteobacteria	ia Identified gut microbe in IBD patients	
51	Ralstonia	S	0.2	Proteobacteria	Microbiota in the Mucosa of Patients With Ulcerative Colitis	FP
62	Bilophila	S	0.1	Proteobacteria	Causes irritable bowel syndrome	FU
3	Desulfovibrio	S	0.1	Proteobacteria	Sulfate reducing bacteria in IBD	FV
64	Halomonas	s	0.7	Proteobacteria	Intestinal microflora in chronic kidney disease	GB
5	g	s	0.1	Proteobacteria	Microbe in colon tissue from IBD subjects	GE
6	Acinetobacter	s	0.7	Proteobacteria	Tissue associated intestinal microflora	GF
7	Pseudomonas	S	0.1	Proteobacteria	Infection in Children with Early-onset Crohn's Disease	GG
8	g	s	0.8	TM7	No role in IBD	
9	Other	Other	0.3	Actinobacteria	Alters fecal microbiota in pediatric IBD patients	GO
0	Eggerthella	Other	0.1	Actinobacteria	Causes bacteremia in Crohn's disease patient	GQ
1	Bacteroides	Other	0.7	Bacteroidetes	Commensal bacteria that induces colitis	GR
2	Prevotella	Other	3.2	Bacteroidetes	A microbial signature of Crohn's disease	GS
3	Other	Other	2.8	Firmicutes	Gut microbe in experimental colitis	GT
4	Paenibacillus	Other	0.1	Firmicutes	Gut microbe in a healthy infant	GU
5	Other	Other	2.2	Firmicutes	Fecal and mucosa associated microbe in IBD	GW
6	Paracoccus	Other	0.1	Proteobacteria	Identified gut microbe in IBD patients	HH
7	Other	Other	0.6	Proteobacteria	Fecal and mucosa associated microbe in IBD	HL
8	Other	Other	0.1	Proteobacteria	Adult fecal microbe	HO
9	Other	Other	0.4	Proteobacteria	Mucosal and fecal microbe	HP
0	Acinetobacter	Other	2.2	Proteobacteria	Tissue associated intestinal microflora in colitis patients	HQ
31	Pseudomonas	Other	0.2	Proteobacteria	Gut microbe in children with early onset Crohn's disease	HR

Specific Information of functions was adapted from NCBI Genome Database (https://www.ncbi.nlm.nih.gov/genome/).

The bacterial species that could not be identified at the genus level are mentioned as g____ and the bacterial species that could not be identified at the species level are mentioned as s____.

compared to adjacent healthy colon specimens. We suggest that pathogenic bacteria belonging to these two phyla have a greater impact on colon motility function in colitis patients (**Tables 1**, **3**). Although the incidence of IBD is increasing among African Americans (AA), the underlying causes are completely unknown (Sofia et al., 2014). Our study further highlight a significant disparity in bacterial dysbiosis among AA compared to CA colitis patients (**Figure 3**).

CA specimens had significantly higher levels of *Fusobacterium*, *Parabacteroides*, *Citrobacter*, *Haemophilus*, *Acinetobacter*, *Pseudomonas*, and *Stenotrophomonas*. *Fusobacterium nucleatum* is known to have a well-characterized role in the oral cavity. We have determined that *Fusobacterium* can be recovered from human full thickness colon specimens and this could indicate their ability to survive and proliferate inside host cells. *Parabacteroides* was found to be dominant in the acute phase of IBD in CA patients. *Citrobacter* is an epithelial cell adherent pathogen and can subvert inflammation in colitis.

Pseudomonas interacts with the mucosal layer of colon and disrupts the mucosal barrier integrity leading to colitis in CA patients.

The AA specimens had significantly higher levels of *Prevotella* and *Clostridia. Prevotella* augments T-helper cells mediated colon mucosal inflammation by activating Toll-like receptor 2 leading to production of T-helper cells polarizing cytokines by antigen-presenting cells, including interleukins. In addition, *Prevotella* induce epithelial cells to produce interleukins and cytokines that can promote recruitment of neutrophils and mucosal T-helper cell immune responses. *Prevotella* can mediate inflammation of the mucosa leading to the circulation of bacteria, bacterial products and other inflammatory mediators. *Prevotella* could augment release of inflammatory mediators from immune cells and various stromal cells in colitis in AA patients. *Clostridium* can disrupt gut immune dormancy and cause infectious colitis in AA patients. Collectively, our data suggest that the presence of pathogenic bacteria in AA

TABLE 5 | Proportions of bacterial species of unknown function colonized in full thickness colon of colitis patients.

SI. No.	Bacteria genus	Bacteria species	Proportion (%)	Bacteria phylum	Function in IBD	NCBI genome database link
ADJACEN	NT HEALTHY COLON					
1	Micrococcus	luteus	0.01	Actinobacteria	No role in IBD	X
2	Arthrobacter	s	0.01	Actinobacteria	No role in IBD	BL
3	Propionicimonas	s	0.01	Actinobacteria	No role in IBD	BN
4	Paludibacter	s	0.01	Bacteroidetes	No role in IBD	BW
5	Chryseobacterium	S	0.02	Bacteroidetes	No role in IBD	CF
6	Calothrix	S	0.03	Cyanobacteria	No role in IBD	CK
7	Novosphingobium	S	0.02	Proteobacteria	No role in IBD	FE
DISEASE	D COLON (ULCERATIV	E COLITIS)				
1	Micrococcus	luteus	0.01	Actinobacteria	No role in IBD	Х
2	Arthrobacter	S	0.02	Actinobacteria	No role in IBD	BL
3	Propionicimonas	S	0.1	Actinobacteria	No role in IBD	BN
4	Paludibacter	S	0.03	Bacteroidetes	No role in IBD	BW
5	Chryseobacterium	S	0.1	Bacteroidetes	No role in IBD	CF
6	Calothrix	S	0.1	Cyanobacteria	No role in IBD	CK
7	Novosphingobium	S	0.04	Proteobacteria	No role in IBD	FE
DISEASE	D COLON (CROHN'S C	OLITIS)				
1	Micrococcus	luteus	0.02	Actinobacteria	No role in IBD	Х
2	Arthrobacter	S	0.2	Actinobacteria	No role in IBD	BL
3	Propionicimonas	S	0.02	Actinobacteria	No role in IBD	BN
4	Paludibacter	S	0.02	Bacteroidetes	No role in IBD	BW
5	Chryseobacterium	S	0.3	Bacteroidetes	No role in IBD	CF
6	Calothrix	S	0.1	Cyanobacteria	No role in IBD	CK
7	Novosphingobium	S	0.01	Proteobacteria	No role in IBD	FE

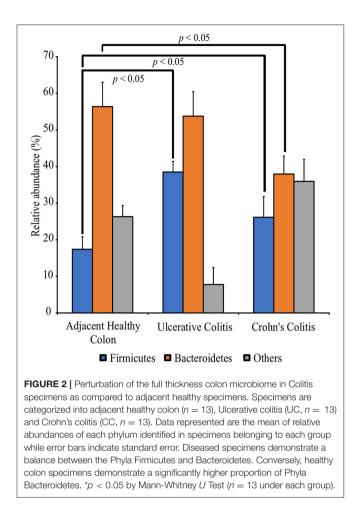
Specific Information of functions was adapted from NCBI Genome Database (https://www.ncbi.nlm.nih.gov/genome/).

The bacterial species that could not be identified at the genus level are mentioned as g___ and the bacterial species that could not be identified at the species level are mentioned as s___.

full thickness diseased specimens could adversely affect colon motility.

Additionally, our data in UC and CC specimens show the presence of several orange (Prevotella, Peptostreptococcus, Eubacterium, Fusobacterium, and *Campylobacter*), red (Porphyromonas), purple (Veillonella), and yellow (Streptococcus) complex putative oral pathogens known to cause gingivitis and periodontitis among IBD patients (Tables 1, 3). Previous studies using mucosal biopsies and feces have shown that gut microbiota in bowel diseases is characterized by an increase in certain phyla such as Proteobacteria, Firmicutes, genus Bifidobacterium, as well as a reduction in the amounts of genera Ruminococcus, Clostridia and (in some cases) Faecalibacterium (Lane et al., 2017; Nishida et al., 2018). However, none of the earlier studies using feces have shown a shift in the balance between Phyla Bacteroidetes and Firmicutes among UC or CC patients; even though this was observed in healthy individuals (Mariat et al., 2009; Koliada et al., 2017). In contrary to our results, one study using mucosal biopsies has shown a significantly decreased Firmicutes to Bacteroidetes ratio in both UC and CC compared with controls (Kabeerdoss et al., 2015). Collectively, our data suggest that the putative oral pathogens found in diseased colon specimens may modulate the proportion of non-detrimental gut bacteria, thus potentially worsening the condition of the colon in colitis patients.

Oral bacterial species like Porphyromonas, Peptostreptococcus, Eubacterium, Fusobacterium, Streptococcus salivarius, S. mitis, S. bovis, Veillonella spp., Staphylococcus aureus, S. epidermidis, and Campylobacter spp. can convert nitrate to nitrite. A large amount of bioactive NO is found in the gastrointestinal tract, generated by dietary sources and by conversion of anaerobic bacteria in the oral cavity, or by anaerobic reaction with nitrate in the colon by *Escherichia coli* spp. The entero-salivary nitrate conversion pathway provides a rich source of bioactive NO and nitrate-reducing bacteria, such as Veillonella. In this pathway, nitrate is obtained by the salivary gland and is then concentrated in the saliva. Various facultative anaerobic bacteria on the top of the tongue effectively reduces nitrate to nitrite. The bacteria then use the nitrate and the nitrite as electron acceptors in their respiration process. This also helps the host in the first steps of converting nitrate to NO. The salivary nitrate then reaches the systemic circulation, various enzymatic reactions occur leading to reduction to NO, and other reactive nitrogen intermediates. The oral cavity plays an important role the production of nitric oxide, and specifically, employs the nitrate-nitrite-NO pathway in the oral cavity. It is well known that oral cavity bacteria can migrate to the colon. Taken together, our data suggest that the



putative oral pathogens found in diseased colon specimens may survive by exploiting the nitrate-nitrite-NO pathway to modulate the proportion of non-detrimental gut bacteria, thus potentially worsening the condition of colon in colitis patients (**Figure 4**).

Previous studies suggest that enteric neurons and smooth muscle mediated gut motility is impaired in colitis patients (Snape et al., 1991; Vermillion et al., 1993). IBD associated gut inflammation affects the morphological and functional changes in the myenteric/enteric nervous system (ENS) and nitric oxide (NO) synthesis (Takahashi, 2003; Kono et al., 2004). Experimental studies have also shown that gut bacteria have a role in oxidative stress induced gut inflammation by controlling metabolic endotoxemia in obese mice (Cani et al., 2008). We have shown that polybacterial oral infection decrease the expression of nNOS and NRF2-phase II enzymes in the gut and this could lead to impaired colon motility (Gangula et al., 2015; Walker et al., 2018).

Some of the gut bacteria we have identified in the full thickness colon specimens in the present study, including *Bacteroides*, *Prevotella*, *Pseudomonas*, etc., have been identified in colon mucosal biopsies in earlier studies (Bibiloni et al., 2006). These bacteria evoke inflammatory responses affecting the innermost lining of colon. Many specific beneficial bacteria, including members of *Bacteroides* and *Prevotella* groups, *C. coccoides*, and

Lactic acid bacteria were known to be decreased in colitis patients (Gibson et al., 1991). Specimens used in prior studies were colon mucosal biopsies or stool samples; but not full thickness colon specimens (Gibson et al., 1991; Bibiloni et al., 2006). Full-thickness colon consists of four layers of tissue including mucosa, submucosa, muscularis, and serosa.

Novel to this research design, full thickness colon specimens were obtained because colitis patients often experience colon motility abnormalities (Snape et al., 1991; Annese et al., 1997; Vrees et al., 2002). Several lines of evidence suggest that nitrergic neurons that releases NO via nNOS are known to play a pivotal role in colon motility (Kono et al., 2004; Winston et al., 2013). Previous studies have demonstrated that nitrergic neurons are degenerated in colitis (Onori et al., 2005; Sung et al., 2006). Recent studies from our laboratory indicate that nNOS, as well as antioxidants (NRF2 regulated-Phase II enzymes) protein expression are down-regulated in diseased colon specimens (Myers et al., 2014; Gangula et al., 2017). Furthermore, our previous studies demonstrated that polybacterial infection led to a decrease in nNOS, NRF2 and antioxidants protein expression in the colon tissues (Gangula et al., 2015). In addition, studies have shown that NO may play homeostatic role in gut inflammation (Kolios et al., 2004). Taken together, our data suggest that elevated levels of oral and gut pathogens in diseased colon full thickness specimens could contribute to impaired nNOS-NO-NRF2-Phase II system and colon motility abnormalities in IBD patients (Figure 4).

To our knowledge, our study is the first to report the presence of several microbiota of unknown function in IBD including *Micrococcus luteus*, Chloracidobacteria, *Arthrobacter*, *Propionicimonas*, *Paludibacter*, *Chryseobacterium*, *Calothrix*, and *Novosphingobium* (**Table 5**). These new microbiota members have not been identified in mucosal/fecal specimens in previous studies, suggesting that these bacteria are primarily colonized in the neuromuscular compartment. Additional studies are warranted to characterize the novel bacteria and investigate their specific role in colon motility and constipation in IBD patients.

In summary, this study have identified specific bacterial pathogens potentially associated with colon motility in IBD patients. The observations showed that some putative oral pathogens belonging to the Phyla Firmicutes (Streptococcus, Staphylococcus, Peptostreptococcus), and Fusobacteria (Fusobacterium) dominated in the microbiomes of CC and UC diseased specimens and might involve the modulation of colon motility in IBD.

STUDY LIMITATIONS

The limitations of the study include the smaller sample size across disease and race groups making this as a preliminary study. In spite of the limitations in sample size and the fact that some of the identified bacteria were not significantly altered in colitis specimens, we were still able to observe differences in the microbiome between CA and AA colitis patients. This could be due to amplicon sequencing of a shorter conserved region of 16S rRNA gene instead of in depth shotgun sequencing.

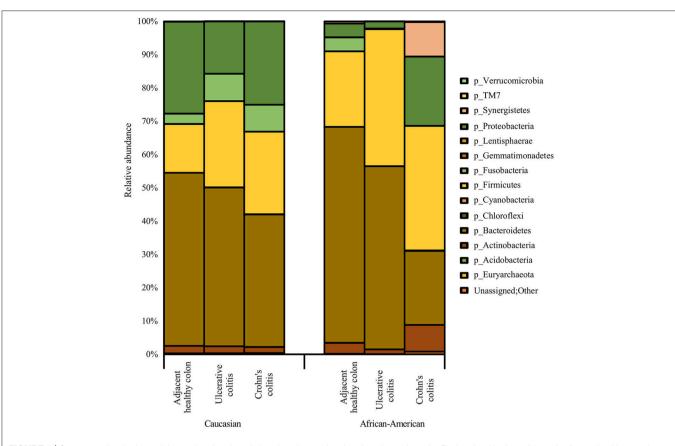
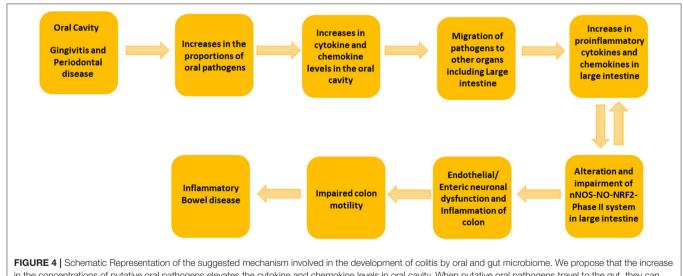


FIGURE 3 | Summary of major bacterial taxa showing the relative abundance of oral and gut bacteria at the Phylum level in the colitis and adjacent healthy specimen groups under each race. Data represented are the mean of relative abundances of each Phyla detected in samples belonging to each group. The dominant phyla across all samples (both diseased and healthy specimens) were Bacteroidetes, followed by Firmicutes and Proteobacteria. Other major phyla observed among these specimens also include Actinobacteria, Fusobacteria, and Synergistetes. The Phylum Proteobacteria did not show any significant difference between healthy colon specimens and diseased colon specimens. A larger proportion of unassigned bacteria (0.3%) was identified in AA Crohn's Colitis patients compared to other groups.



in the concentrations of putative oral pathogens elevates the cytokine and chemokine levels in oral cavity. When putative oral pathogens travel to the gut, they can colonize locally and lead to the elevated levels of proinflammatory cytokines. This can effect on nNOS-NO-NRF2-Phase II system in the large intestine and could lead to colon dysmotility and colitis.

Moreover, we did not profile the oral microbiome from oral specimens (dental plaque, etc.) in the same IBD patients from whom full thickness colon specimens were collected. Finally, host-microbiome interaction studies are needed to better discern specific roles of the oral and gut bacteria in the development of colitis. Future studies are aimed to collect oral and fecal specimens therefore a comparative experiments in regards to changes in microbiome, along with specific key proteins will be conducted from the same patient.

AUTHOR CONTRIBUTIONS

VD, SM, KS, SP, SS, PG, and MT have contributed both for data analysis and manuscript preparation. DS, CF-D, LK, SA, and JS have contributed in manuscript preparation.

REFERENCES

- Agossa, K., Dendooven, A., Dubuquoy, L., Gower-Rousseau, C., Delcourt-Debruyne, E., and Capron, M. (2017). Periodontal manifestations of inflammatory bowel disease: emerging epidemiologic and biologic evidence. J. Periodontal Res. 52, 313–324. doi: 10.1111/jre.12422
- Annese, V., Bassotti, G., Napolitano, G., Usai, P., Andriulli, A., and Vantrappen, G. (1997). Gastrointestinal motility disorders in patients with inactive Crohn's disease. *Scand. J. Gastroenterol.* 32, 1107–1117.
- Atarashi, K., Suda, W., Luo, C., Kawaguchi, T., Motoo, I., Narushima, S., et al. (2017). Ectopic colonization of oral bacteria in the intestine drives TH1 cell induction and inflammation. *Science* 358, 359–365. doi: 10.1126/science.aan4526
- Bassotti, G., Antonelli, E., Villanacci, V., Salemme, M., Coppola, M., and Annese, V. (2014). Gastrointestinal motility disorders in inflammatory bowel diseases. *World J. Gastroenterol.* 20, 37–44. doi: 10.3748/wjg.v20.i1.37
- Bibiloni, R., Mangold, M., Madsen, K. L., Fedorak, R. N., and Tannock, G. W. (2006). The bacteriology of biopsies differs between newly diagnosed, untreated, Crohn's disease and ulcerative colitis patients. *J. Med. Microbiol.* 55, 1141–1149. doi: 10.1099/jmm.0.46498-0
- Cani, P. D., Bibiloni, R., Knauf, C., Waget, A., Neyrinck, A. M., Delzenne, N. M., et al. (2008). Changes in gut microbiota control metabolic endotoxemiainduced inflammation in high-fat diet-induced obesity and diabetes in mice. *Diabetes* 57, 1470–1481. doi: 10.2337/db07-1403
- Caporaso, J. G., Bittinger, K., Bushman, F. D., DeSantis, T. Z., Andersen, G. L., and Knight, R. (2010a). PyNAST: a flexible tool for aligning sequences to a template alignment. *Bioinform*. 26, 266–267. doi: 10.1093/bioinformatics/btp636
- Caporaso, J. G., Kuczynski, J., Stombaugh, J., Bittinger, K., Bushman, F. D., Costello, E. K., et al. (2010b). QIIME allows analysis of high-throughput community sequencing data. *Nat. Methods* 7, 335–336. doi: 10.1038/nmeth.f.303
- Edgar, R. C. (2010). Search and clustering orders of magnitude faster than BLAST. *Bioinformatics* 26, 2460–2461. doi: 10.1093/bioinformatics/btq461
- Gangula, P., Ravella, K., Chukkapalli, S., Rivera, M., Srinivasan, S., Hale, A., et al. (2015). Polybacterial periodontal pathogens alter vascular and gut BH₄/nNOS/NRF2-phase II enzyme expression. *PLoS ONE* 10:e0129885. doi: 10.1371/journal.pone.0129885.
- Gangula, P. R., Smoot, D. T., Izban, M., Ballard, B., and Adunyah, S. (2017). "Expression of NO and NRF2 enzymes in Human Colitis," in *Proceedings of the RCMI Translational Science 2017 Conference* (Washington, DC), 78.
- Geboes, K., and Collins, S. (1998). Structural abnormalities of the nervous system in Crohn's disease and ulcerative colitis. *Neurogastroenerol. Motil.* 10, 189–202.
- Gibson, G. R., Cummings, J. H., and Macfarlane, G. T. (1991). Growth and activities of sulphate-reducing bacteria in gut contents of healthy subjects and patients with ulcerative colitis. *FEMS Microbiol. Lett.* 86, 103–111. doi: 10.1111/j.1574-6968.1991.tb04799.x

ACKNOWLEDGMENTS

The authors sincerely thank University of North Carolina at Chapel Hill School of Medicine- Microbiome Core Facility (UNC; MC) for performing amplicon sequencing. We thank CHTN for providing diseased and healthy specimens. We further thank dental student doctors Mr. A. Arab and J. Hodges for their initial contribution on this project. The Meharry Bioinformatics Core is funded in part by NIH grants MD007593 and MD007586. SA is supported by NIMHD R-CTR grant No. 54544MD007593 and NCI grant No. 5454CA163069. This work is supported in part by 1SC1GM121282 funded to PG. In addition, Research reported in this publication was supported by the National Institute of General Medical Sciences of the National Institutes of Health under Award Number SG1GM121282 (PG).

- Gower, J. C. (2005). "Principal coordinates analysis," in *Encyclopedia of Biostatistics*, eds P. Armitage and T. Colton, (John Wiley & Sons Ltd). doi: 10.1002/0470011815
- Haas, B. J., Gevers, D., Earl, A. M., Feldgarden, M., Ward, D. V., Giannoukos, G., et al. (2011). Chimeric 16S rRNA sequence formation and detection in Sanger and 454-pyrosequenced PCR amplicons. *Genome Res.* 21, 494–504. doi: 10.1101/gr.112730.110
- Hajishengallis, G. (2015). Periodontitis: from microbial immune subversion to systemic inflammation. *Nat. Rev. Immunol.* 15, 30–44. doi: 10.1038/ nri3785
- Kabeerdoss, J., Jayakanthan, P., Pugazhendhi, S., and Ramakrishna, B. S. (2015). Alterations of mucosal microbiota in the colon of patients with inflammatory bowel disease revealed by real time polymerase chain reaction amplification of 16S ribosomal ribonucleic acid. *Indian J Med Res.* 142, 23–32. doi: 10.4103/0971-5916.162091
- Keegan, K. P., Glass, E. M., and Meyer, F. (2016). MG-RAST, a metagenomics service for analysis of microbial community structure and function. *Methods Mol. Biol.* 1399, 207–233. doi: 10.1007/978-1-4939-3369-3_13
- Koliada, A., Syzenko, G., Moseiko, V., Budovska, L., Puchkov, K., Perederiy, V., et al. (2017). Association between body mass index and Firmicutes/Bacteroidetes ratio in an adult Ukrainian population. BMC Microbiol. 17:120. doi: 10.1186/s12866-017-1027-1
- Kolios, G., Valatas, V., and Ward, S. G. (2004). Nitric oxide in inflammatory bowel disease: a universal messenger in an unsolved puzzle. *Immunology* 113, 427–437. doi: 10.1111/j.1365-2567.2004.01984.x
- Kono, T., Chisato, N., Ebisawa, Y., Asama, T., Sugawara, M., Ayabe, T., et al. (2004). Impaired nitric oxide production of the myenteric plexus in colitis detected by a new bioimaging system. J. Surg. Res. 117, 329–336. doi: 10.1016/j.jss.2003.11.004
- Lane, E. R., Zisman, T. L., and Suskind, D. L. (2017). The microbiota in inflammatory bowel disease: current and therapeutic insights. J. Inflamm. Res. 10, 63–73. doi: 10.2147/JIR.S116088
- Lira-Junior, R., and Figueredo, C. M. (2016). Periodontal and inflammatory bowel diseases: is there evidence of complex pathogenic interactions? World J. Gastroenterol. 22, 7963–7972. doi: 10.3748/wjg.v22.i35.7963
- Mariat, D., Firmesse, O., Levenez, F., Guimaraes, V. D., Sokol, H., Dore, J., et al. (2009). The Firmicutes/Bacteroidetes ratio of the human microbiota changes with age. *BMC Microbiol.* 9:123. doi: 10.1186/1471-2180-9-123
- Myers, J. N., Schaffer, M. W., Korolkova, O. Y., Williams, A. D., Gangula, P. R., and M'Koma, A. E. (2014). Implications of the colonic deposition of free hemoglobin-α chain: a previously unknown tissue by-product in inflammatory bowel disease. *Inflamm. Bowel Dis.* 20, 1530–1547. doi: 10.1097/MIB.00000000000144
- Nishida, A., Inoue, R., Inatomi, O., Bamba, S., Naito, Y., and Andoh, A. (2018). Gut microbiota in the pathogenesis of inflammatory bowel disease. *Clin. J. Gastroenterol.* 11, 1–10. doi: 10.1007/s12328-017-0813-5

- Onori, L., Aggio, A., D'Alo, S., Muzi, P., Cifone, M. G., Mellillo, G., et al. (2005). Role of nitric oxide in the impairment of circular muscle contractility of distended, uninflamed mid-colon in TNBS-induced acute distal colitis in rats. *World J. Gastroenterol.* 11, 5677–5684. doi: 10.3748/wjg.v11.i36.5677
- Poli, E., Lazzeretti, M., Grandi, D., Pozzoli, C., and Coruzzi, G. (2001). Morphological and functional alterations of the myenteric plexus in rats with TNBS-induced colitis. *Neurochem. Res.* 26, 1085–1093. doi: 10.1023/A:1012313424144
- Popova, C., Dosseva-Panova, V., and Panov, V. (2013). Microbiology of periodontal diseases. a review. *Biotechnol. Biotechnol. Equip.* 27, 3754–3759. doi: 10.5504/BBEQ.2013.0027
- Price, M. N., Dehal, P. S., and Arkin, A. P. (2010). FastTree 2-approximately maximum-likelihood trees for large alignments. *PLoS ONE* 5:e9490. doi: 10.1371/journal.pone.0009490.
- Schloss, P. D., Westcott, S. L., Ryabin, T., Hall, J. R., Hartmann, M., Hollister, E. B., et al. (2009). Introducing mothur: open-source, platformindependent, community-supported software for describing and comparing microbial communities. *Appl. Environ. Microbiol.* 75, 7537–7541. doi: 10.1128/AEM.01541-09
- Snape, W. J. Jr., Williams, R., and Hyman, P. E. (1991). Defect in colonic smooth muscle contraction in patients with ulcerative colitis. *Am. J. Physiol.* 261, G987–G991. doi: 10.1152/ajpgi.1991.261.6.G987
- Sofia, M. A., Rubin, D. T., Hou, N., and Pekow, J. (2014). Clinical presentation and disease course of inflammatory bowel disease differs by race in a large tertiary care hospital. *Dig Dis Sci.* 59, 2228–2235. doi: 10.1007/s10620-014-3160-0
- Strauss, J., Kaplan, G. G., Beck, P. L., Rioux, K., Panaccione, R., Devinney, R., et al. (2011). Invasive potential of gut mucosa-derived *Fusobacterium nucleatum* positively correlates with IBD status of the host. *Inflamm. Bowel Dis.* 17, 1971–1978. doi: 10.1002/ibd.21606
- Sung, T. S., La, J. H., Kim, T. W., and Yang, I. S. (2006). Alteration of nitrergic neuromuscular transmission as a result of acute experimental colitis in rat. J. Vet. Sci. 7, 143–150. doi: 10.4142/jvs.2006.7.2.143
- Takahashi, T. (2003). Pathophysiological significance of neuronal nitric oxide synthase in the gastrointestinal tract. J. Gastroenterol. 38, 421–430. doi: 10.1007/s00535-003-1094-y

- Vavricka, S. R., Manser, C. N., Hediger, S., Vogelin, M., Scharl, M., Biedermann, L., et al. (2013). Periodontitis and gingivitis in inflammatory bowel disease: a case-control study. *Inflamm. Bowel. Dis.* 19, 2768–2777. doi: 10.1097/01.MIB.0000438356.84263.3b
- Vermillion, D. L., Huizinga, J. D., Riddell, R. H., and Collins, S. M. (1993). Altered small intestinal smooth muscle function in Crohn's disease. *Gastroenterol.* 104, 1692–1699.
- Vrees, M. D., Pricolo, V. E., Ptenti, F. M., and Cao, W. (2002). Abnormal motility in patients with ulcerative colitis: the role of inflammatory cytokines. *Arch. Surg.* 137, 439–445. doi: 10.1001/archsurg.137.4.439
- Walker, M. Y., Pratap, S., Southerland, J. H., Farmer-Dixon, C. M., Lakshmyya, K., and Gangula, P. R. (2018). Role of oral and gut microbiome in nitric oxide-mediated colon motility. *Nitric Oxide* 73, 81–88. doi: 10.1016/j.niox.2017.06.003
- Winston, J. H., Li, Q., and Sarna, S. K. (2013). Paradoxical regulation of ChAT and nNOS expression in animal models of Crohn's colitis and ulcerative colitis. Am. J. Physiol. Gastrointest. Liver Physiol. 305, G295–302. doi: 10.1152/ajpgi.00052.2013
- Zhao, L., Huang, Y., Lu, L., Yang, W., Huang, T., Lin, Z., et al. (2018). Saturated long-chain fatty acid-producing bacteria contribute to enhanced colonic motility in rats. *Microbiome* 6:107. doi: 10.1186/s40168-018-0492-6

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 Dinakaran, Mandape, Shuba, Pratap, Sakhare, Tabatabai, Smoot, Farmer-Dixon, Kesavalu, Adunyah, Southerland and Gangula. This is an openaccess article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.