# Predicting Responses to Checkpoint Inhibitors in Lymphoma: Are We Up to the Standards of Solid Tumors?

Clinical Medicine Insights: Oncology Volume 14: 1-13 © The Author(s) 2020 Article reuse guidelines: sagepub.com/journals-permissions DOI: 10.1177/1179554920976366 (S)SAGE

## Ah-Reum Jeong<sup>1</sup>, Edward D Ball<sup>2</sup> and Aaron Michael Goodman<sup>2</sup>

<sup>1</sup>Division of Hematology and Oncology, Department of Medicine, University of California, San Diego, La Jolla, CA, USA. <sup>2</sup>Division of Blood and Marrow Transplantation, Department of Medicine, University of California, San Diego, La Jolla, CA, USA.

ABSTRACT: Treatment of cancer has transformed with the introduction of checkpoint inhibitors. However, the majority of solid tumor patients do not respond to checkpoint blockade. In contrast, the response rate to programmed cell death 1 (PD-1) blockade in relapsed/refractory classical Hodgkin lymphoma (cHL) is 65% to 84% which is the highest among all cancers. Currently, checkpoint inhibitors are only approved for cHL and primary mediastinal B-cell lymphoma as the responses to single-agent checkpoint blockade in other hematologic malignancies is disappointingly low. Various established biomarkers such as programmed cell death 1 ligand 1 (PD-L1) protein surface expression, mismatch repair (MMR) status, and tumor mutational burden (TMB) are routinely used in clinical decision-making in solid tumors. In this review, we will explore these biomarkers in the context of hematologic malignancies. We review characteristic 9p24.1 structural alteration in cHL and primary mediastinal B-cell lymphoma (PMBCL) as a basis for response to PD-1 inhibition, as well as the role of antigen presentation pathways. We also explore the reported frequencies of MMR deficiency in various hematologic malignancies and investigate TMB as a predictive marker.

KEYWORDS: Lymphoma, hematological malignancies, checkpoint blockade, biomarker, 9p24.1, PD-L1, PD-L1, CTLA-4, MHC, TMB, MSI, MMR

RECEIVED: March 23, 2020. ACCEPTED: October 28, 2020.

TYPE: Review Article

FUNDING: The author(s) received no financial support for the research, authorship, and/or publication of this article.

### Introduction

T-cell inhibition through immune checkpoints, notably through cytotoxic T-lymphocyte-associated antigen 4 (CTLA-4) and programmed cell death 1 (PD-1) pathways, plays a critical role in cancer evasion of the immune system. Immune checkpoints induce self-tolerance by inhibiting T-cell maturation and dampening peripheral function. In the tumor microenvironment, CTLA-4 is expressed on T-regulatory cells (Tregs) and binds to CD80 and CD86 on antigen-presenting cells (APCs) resulting in decreased co-stimulation of conventional T-cells.1 CD4+ and CD8+ T-cells become deactivated by programmed cell death 1 ligand 1 (PD-L1) and programmed cell death 1 ligand 2 (PD-L2) expressed on cancer cells.<sup>2</sup>

Inhibition of these pathways by monoclonal antibodies has revolutionized the treatment of cancer. For example, treatment of melanoma with ipilimumab and nivolumab demonstrated an overall response rate (ORR) of 50% to 60%.<sup>3,4</sup> Importantly, many of the responses were durable.<sup>5</sup> Since the first Food and Drug Administration (FDA) approval of ipilimumab for melanoma in 2010, the use of checkpoint inhibitors has expanded across multiple solid tumors including small-cell and nonsmall-cell lung cancer (NSCLC), renal cell carcinoma (RCC), hepatocellular carcinoma (HCC), head and neck cancer, urothelial cancer, esophageal cancer, gastric cancer, colorectal cancer, cervical cancer, and endometrial cancer.

DECLARATION OF CONFLICTING INTERESTS: The author(s) declared the following potential conflicts of interest with respect to the research, authorship, and/or publication of this article: A.M.G. receives consulting and speaking fees from Seattle Genetics and consulting fees from EUSA Pharma.

CORRESPONDING AUTHOR: Ah-Reum Jeong, Division of Hematology and Oncology, Department of Medicine, University of California, San Diego, 3855 Health Sciences Dr. MC0829, La Jolla, CA 92037, USA. Email: ajeong@health.ucsd.edu

Identifying biomarkers that can predict response to checkpoint blockade is of major interest because a significant proportion of patients do not benefit and experience immune-related adverse events or possibly even hyperprogressive disease.6 These biomarkers are better established in solid tumors and include PD-L1 expression measured by immunohistochemistry (IHC),<sup>7,8</sup> *PD-L1* amplification,<sup>9</sup> tumor mutational burden (TMB),<sup>10,11</sup> microsatellite instability (MSI), and mismatch repair (MMR) status<sup>12,13</sup> (Table 1). Pembrolizumab was the first tumor agonistic treatment approved by the FDA based on MSI-high (MSI-H) or mismatch-repair-deficient (dMMR) status.14 These tumors respond exquisitely to checkpoint blockade due to their high mutational burden and mutation-associated neoantigens.<sup>12</sup> Combinations of 2 or more of the above biomarkers are also being investigated.<sup>15</sup> These biomarkers, most notably PD-L1 expression and TMB, are routinely obtained in the evaluation of solid malignancies and are being incorporated in treatment decision algorithms.<sup>16</sup>

In contrast to solid malignancies, checkpoint inhibitors are only approved for the treatment of 2 hematologic malignancies, classical Hodgkin lymphoma (cHL) and primary mediastinal B-cell lymphoma (PMBCL). The response rate to single-agent PD-1 blockade in relapsed/refractory cHL is 65% to 84%<sup>22-36</sup> which is markedly higher than the response rate in any solid tumor. In contrast to cHL, most B-cell non-Hodgkin lymphomas (NHLs), multiple myelomas, and leukemias have

 $(\mathbf{\hat{n}})$ 

Creative Commons Non Commercial CC BY-NC: This article is distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 License (https://creativecommons.org/licenses/by-nc/4.0/) which permits non-commercial use, reproduction and distribution of the work without further permission provided the original work is attributed as specified on the SAGE and Open Access pages (https://us.sagepub.com/en-us/nam/open-access-at-sage).

Table 1.	Biomarkers r	outinely used	and emerging	biomarkers in	solid malignancies
----------	--------------	---------------	--------------	---------------	--------------------

BIOMARKER	DESCRIPTION	ROUTINELY USED IN CLINICAL PRACTICE?
Gene expression profiling	18 genes indicative of T-cell activation measured by RNA expression <sup>15,17</sup>	No
MSI-H/dMMR	Determined by PCR, NGS, or IHC <sup>12,13</sup>	Yes
Multiplex IHC/IF	Multiple markers are co-stained on a single slide, enabling visualization of spatial interaction of various markers <sup>18</sup>	No
PD-L1 surface expression	IHC of the tumor cells and/or immune cells is measured and reported as TPS or CPS scores $^{7,8}$	Yes
<i>PD-L1/PD-L2</i> structural variation	Amplification or translocation of genetic locus 9p24.1 identified by NGS or FISH <sup>9</sup>	No
Tumor-infiltrating lymphocytes	T-cells in the tumor microenvironment are identified by IHC <sup>19-21</sup>	No
Tumor mutational burden	Mutations per mega-base is calculated by cancer-gene panel NGS or whole exome sequencing <sup>10,11</sup>	Yes

Abbreviations: CPS, combined positive score; dMMR, mismatch repair deficient; FISH, fluorescent in-situ hybridization; IF, immunofluorescence; IHC,

immunohistochemistry; MSI-H, microsatellite instability high; NGS, next-generation sequencing; PCR; polymerase chain reaction; PD-L1, programmed cell death 1 ligand 1; PD-L2, programmed cell death 1 ligand 2; RNA, ribonucleic acid; TPS, tumor proportion score.

disappointingly low response rates to checkpoint blockade. Furthermore, unlike solid tumors, predictive biomarkers have not been clearly defined for hematologic malignancies.

Here, we review the unique immune response biomarkers reported in lymphoma and explore the literature whether any of the markers used in solid malignancies could be applied.

#### 9p24.1 gene alteration

*cHL.* Chemotherapy with doxorubicin, bleomycin, vinblastine, and dacarbazine (ABVD) or escalated bleomycin, etoposide, doxorubicin, cyclophosphamide, vincristine, procarbazine, prednisolone (BEACOPP) is established first-line therapies for advanced-stage cHL. Approximately 30% of patients will relapse or be refractory to frontline therapy.<sup>37</sup> Options for relapsed patients include further cytotoxic chemotherapy, autologous stem cell transplantation, and CD30-directed antibody drug conjugate brentuximab vedotin.<sup>38</sup>

Checkpoint inhibitors are now a well-established treatment option in relapsed/refractory cHL with response rates of 65% to 84%, while its role in the frontline setting is currently being investigated (Table 2).<sup>22-36</sup> This response rate is the highest reported among all malignancies and is the result of the distinctive biology of cHL. Classical HL is characterized by few malignant Hodgkin Reed-Sternberg (HRS) cells interspersed among ample but ineffective immune infiltrate consisting of eosinophils, macrophages, and lymphocytes which produces abundant cytokines.<sup>39</sup> Despite the proinflammatory tumor microenvironment, the immune system is incapable of clearing HRS cells. Mechanisms of HRS cell immune evasion include PD-L1/PD-L2 overexpression from 9p24.1 structural alterations, mutations in antigen presentation pathways, and alterations in nuclear factor kappa-light-chain-enhancer of activated B-cells (NF-κB) signaling pathway.<sup>40</sup>

A defining characteristic of HRS cells is the 9p24.1 gene alteration/amplification, which contains genes *CD274 (PD-L1)*, *PDCD1LG2 (PD-L2)*, and *Janus Kinase 2 (JAK2)*.<sup>41,45</sup> In the phase II CheckMate 205 study evaluating single-agent nivolumab in relapsed/refractory cHL, all HRS cells had either copy number alterations (CNA) or translocations involving 9p24.1 locus.<sup>25,32,34</sup> Ten percent of tumors had polysomy, 60% had copy gain, 27% had amplification, 2% had balanced rearrangement, and 1% had unbalanced rearrangement of 9p24.1 as their highest level of genetic alteration.<sup>41</sup>

Higher level of structural variants of 9p24.1 is associated with a poor prognosis in cHL.<sup>45</sup> Increased expression of genes in this locus results in increased protein expression of PD-L1/ PD-L2 both directly and indirectly by upregulating JAK/signal transducer and activator of transcription (STAT) signaling pathway.<sup>46</sup> Expectedly, higher PD-L1/PD-L2 IHC expression measured by the product of percentage of positive malignant cells and the intensity of signal (modified PD-L1 H-score) is associated with higher response rates and progression-free survival (PFS) to anti-PD-1 therapy.<sup>32,34,41</sup>

In a post hoc analysis of cohorts B and C of CheckMate 205,<sup>41</sup> patients who had progression of disease on therapy had lower level 9p24.1 alteration (P=.006) and lower PD-L1 H-score (P=.047). Those who had lower-level 9p24.1 alteration and lower PD-L1 H-score also had lower PFS (P=.026). Interestingly, 2 patients with balanced rearrangements of PD-L2 had rapidly progressive disease. In cohort D of CheckMate 205,<sup>34</sup> a phase II trial of nivolumab combined with doxorubicin, vinblastine, and dacarbazine (AVD) in newly diagnosed advanced cHL, a greater proportion of patients in the third and fourth quartile for PD-L1 H-score had deeper and more durable responses (P=.041). Although rare in solid tumors (0.7%), *PD-L1* amplification is associated with a response rate of 66.7% to PD-1 inhibition, approaching what is seen in cHL.<sup>9</sup>

RKER					n er	-score vith .47) vith and	l ictive onth in ession ontinued)
RESPONSE RATE BY BIOMA	NR		No clear association		Cohorts B/C only: Lower-level 9p24.1 alteration associated with progression therapy ( $P$ =.006) and short PFS ( $P$ <.001)	Cohorts B/C only: Significant association of H- and degree of 9p24.1 CNA ii cells ( $P$ = .001). Lower H-score associated w progression on therapy ( $P$ =. and shorter PFS ( $P$ =.026). H-score was significantly associated with PFS in pts v >12 months between ASCT nivolumab but not in ≤12 mo	92% of CR had tumors with positive or decreased MHCl expression ( $P = .03$ ). MHCll expression was predi of PFS in those with > 12-mc of PFS in those with > 12-mc interval between ASCT and nivolumab ( $P = .014$ ) but not i $\leq 12$ months. No association between $\beta2$ -microglobulin or MHCl expreson HRS cells and PFS.
BIOMARKER ASSESSMENT	Amplification, copy gain, polysomy, disomy <sup>b</sup>	H-score⁰	Positive if at least 1% of HL cells demonstrated at least partial membrane staining with moderate or strong intensity	Intensity (0-5)	Rearrangement, amplification, copy gain, polysomy, disomy <sup>b</sup>	H-score°	Positive, decreased, or negative expression on HRS cells compared to adjacent nonmalignant inflammatory cells
PROSPECTIVE BIOMARKER	9p24.1 alteration by FISH	PD-L1 by IHC	PD-L1 by IHC	PD-L2 by IHC	9p24.1 alteration by FISH	PD-L1 by IHC	β2-microglobulin, MHC class I/II by IHC
RESPONSE RATE <sup>a</sup>	ORR 87% (66-97%) CR 17% PFS at 24wks 86%	(%CG-70)	ORR 65% (48-79%) <sup>d</sup> CR 16% (48-79%) <sup>d</sup> PFS at 52weeks 46% OS at 24weeks 100%		ORR 69% (63-75%) CR 16% PFS 14.7 months (11.3-18.5 months) OS at 1year	(%CG-00) %.76	
TREATMENT	Nivolumab 3mg/kg q2wks		Pembrolizumab 10mg/kg q2wks		Nivolumab 3mg/kg q2wks		
POPULATION	R/R		R/R after BV		R/R after: A. ASCT (n=63) B. ASCT + BV (n=80) C. BV + ASCT → ± BV (n=100)		
z	23		31		243		
STUDY	CheckMate 039 <sup>22,23</sup> Phase I		KEYNOTE-013 Armand et al <sup>26</sup> Phase lb		CheckMate 205 <sup>25,32,41</sup> Cohorts A, B, C 2016, Phase II		

Table 2. Prospective trials in classical Hodgkin lymphoma using checkpoint inhibitors.

z		POPULATION	TREATMENT	RESPONSE RATE <sup>a</sup>	PROSPECTIVE BIOMARKER	BIOMARKER ASSESSMENT	RESPONSE RATE BY BIOMARKER
5	10	R/R atter: 1. ASCT + BV (n = 69) 2. Chemo + BV	Pembrolizumab 200 mg q3wks	ORR 71.9% (65.3-77.9%) CR 27,6% (21.7-34.2%) PFS 13.7 months	PD-L1 by IHC	Intensity (0-3) and membrane staining (0%, 1-50%, or 51-100%)	щ
		(n=89) 3. ASCT (n=60)		(IT-IT-UCMONTRS) OS at 24 months 90.9%	PD-L1 in histiocytes by IHC	Intensity (0-3)	
	17	R/R	Nivolumab 3mg/kg q2wks	ORR 81.3% (54.4-96.0%) CR 25.0% PFS at 6 months 60.0% (31.8-79.7%) OS at 6 months 100%	R	R	R
G	32	R/R	BV + Nivolumab 3 mg/kg q3wks × 4 cycles	ORR 82% (70-91%) CR 61% (47-73%)	R	RN	RN
(7)	31	R/R, post-ASCT	Pembrolizumab 200 mg IV	PFS at 19 months 81%	RN	NR	NR
		consolidation	dowrs × o cycles	00-92%) OS at 19months 100%	PD-L1 by multiplex IF	R	
L)	51	Untreated advanced-stage	Nivolumab 240mg q2wks ×4 doses followed by N-AVD ×6 cycles	ORR 84% (71-93%) CR 67% (52-79%) PFS at 9months 92% (80-97%)	9p24.1 alteration by FISH	Rearrangement, amplification, copy gain, polysomy, disomy <sup>b</sup>	Positive trend between magnitude of 9p24.1 CNA and PD-L1 expression ( <i>P</i> = .067). Trend toward more favorable
				05 at 9months 98% (86-100%)	PD-L1 in HRS by IHC	Modified H-score <sup>°</sup>	response to involutinate monotherapy in patients with higher HRS cell PD-L1 expression ( $P$ =.096). Greater proportion of pts with PD-L1 expression in quartile 3 or 4 had deeper and more durable responses to N-AVD ( $P$ =.041).
0)	92	R/R	Sintilimab 200 mg IV q2wks	ORR 80.4% (70.9-88.0%) CR 34% PFS at 6 months 77.6% (66.6-85.4%)	Ш	R	R
	75	R/R after ASCT or transplant ineligible	Camrelizumab 200 mg IV q2wks	ORR 76.0 (64.7-85.1%) CR 28% PFS at 9 months 76.6% (65.1-84.8%)	Ш	R	R

(Continued)

ESPONSE RATE BY BIOMARKER	œ	MB was 10 in 1 cHL patient with A, but no statistics able to be ilculated.	œ	œ	CR, complete response; FISH, AVD nivolumeh dovernition
H	Z	e PF Ca	Z	Z	ber alteration; C
BIOMARKER ASSESSMENT	Ч	Foundation one	R	R	ma; CNA, copy num
PROSPECTIVE BIOMARKER	NR	TMB by NGS	R	R	assical Hodgkin lympho
RESPONSE RATE <sup>a</sup>	ORR 87.1% (77.0- 93.9%) CR 62.9% PFS at 9months 74.5%	ORR 14.5% CR 0%	Nivo/Ipi: cHL (n= 31) ORR 74% (55-88%) CR 23% (10-41%) B-NHL (n= 16) ORR 19% (4-46%) CR 6% (0-30%) Nivo/Liri Nivo/Liri ORR 76% (53-92%) CR 24% (8-47%) CR 24% (8-47%) B-NHL (n= 32) ORR 13% (4-29%) CR 3% (0-16%)	BV + Ipi ORR 76% (53-92%) CR 57% (34-78%) BV + Nivo ORR 89% (65-99%) CR 61% (36-83%) BV + Ipi/Nivo ORR 82% (60-95%) CR 73% (50-89%)	3V, brentuximab vedotin; cHL, cl immunohistochamistry: MHC n
TREATMENT	Tislelizumab 200 mg IV q3wks	Lenalidomide + ipilimumab dose escalation from 1.5-3.0 mg/kg	Nivolumab 3 mg/ kg + ipilimumab 1 mg/kg q3wks or nivolumab 3 mg/ kg + lirilumab 3 mg/kg q4wks	Brentuximab 1.8 mg/ kg + ipilimumab 3 mg/kg or nivolumab 3 mg/kg or nivolumab 3 mg/kg and ipilimumab 1 mg/kg	الـ, B-cell non-Hodgkin lymphoma; E معند: ٦٦ أنسميا محمصيني المرك
POPULATION	R/R after ASCT or transplant ineligible	Advanced metastatic malignancy without standard therapy	R/R lymphoid malignancies (cHL, B-NHL, T-NHL, MM)	R/R	is stem cell transplant; B-NF
z	20	7	137	64	autologou
STUDY	Song et al <sup>35</sup> Phase II	Sakamuri et al⁴² Phase I <sup>e</sup>	Armand et al <sup>43</sup> Phase Ib	Diefenbach et al <sup>44</sup> Phase I/II	Abbreviations: ASCT,

incressent in-situ inyurutation; htts, hoogkin treed-steriberg; ir, immunoituorescence; IHC, immunonistochemistry; MHC, major instocompatibility complex; MM, multiple myeloma; N-AVU, involumab, doxorubicin, vinblastine, dacarbazine; NGS, next-generation sequencing; NR, nonreported; ORR, overall response; rate; OS, overall survival; PD-L1, programmed cell death 1 ligand 1; PFS, progression-free survival; PR, partial response; R/R, relapsed/refractory; TMB, tumor mutational burden; T-NHL, T-cell non-Hodgkin lymphoma.

<sup>e95%</sup> confidence interval unless otherwise indicated. <sup>b</sup>Polysomy was defined as target: control signal ratio of 1:1 but more than 2 copies per probe, copy gain as signal ratio greater than 1:1 but less than 3:1, and amplification as signal ratio 3:1 or greater. <sup>c</sup>Modified H-score was calculated by multiplying percentage of PD-L1-positive HRS cells (0-100%) and the average intensity of PD-L1 staining (0 to 3+), and ranges from score 0 to 300. <sup>d90%</sup> confidence interval.

Table 2. (Continued)

PD-L1 H-score may be a surrogate marker of level of 9p24.1 gene alteration. A major limitation is that above studies classified PD-L1 H-score to quartiles instead of defining an absolute cut off value for high level of PD-L1 expression. Further studies will need to demonstrate the predictive value of absolute PD-L1 H-score in cHL to anti-PD-1 therapy.

*PMBCL*. PMBCL is a subtype of diffuse B-cell lymphoma (DLBCL) which has similar genetic signatures and clinical characteristics to cHL. The most commonly used frontline therapies for PMBCL are dose-adjusted rituximab, etoposide, prednisone, vincristine, cyclophosphamide, doxorubicin (DA-R-EPOCH) or rituximab, cyclophosphamide, doxorubicin, vincristine, and prednisone (R-CHOP) followed by mediastinal radiation. Autologous transplant, brentuximab vedotin, or checkpoint inhibitors are used for relapsed/refractory disease.<sup>47</sup>

Alterations of 9p24.1 have been reported in 63% to 75% of cases of PMBCL.<sup>46,48,49</sup> Interestingly, there is larger change in transcript abundance of PD-L2 than PD-L1, correlating with higher PD-L2 surface expression.<sup>49</sup> In one series, PD-L1 IHC was positive in 53% while PD-L2 was positive in 78% of lymphoma cells.<sup>50</sup> The significance of this finding is unknown. There are also several care reports of chromosomal rearrangements involving 9p24.1 locus, which result in high surface expression of PD-L1.<sup>51</sup>

Similar to cHL, PMBCL is very sensitive to PD-L1 inhibition, with ORR of 45% to 73% (Table 3).<sup>33,52,53</sup> KEYNOTE-017,<sup>52</sup> a phase II study of pembrolizumab monotherapy in 53 patients with relapsed refractory PMBCL, demonstrated an ORR of 45% with complete response (CR) of 13%. Patients who achieved CR had durable response at a median follow-up of 12.5 months: 76% of patients who responded had response durations greater than 12 months. Ninety-five percent of patients had 9p24.1 structural variation, including 4 patients with translocations. The degree of CNA was significantly associated with PD-L1 protein expression, which was significantly associated with PFS. Although limited to one prospective study, PD-L1 H-score is also a promising biomarker in patients with PMBCL predicting response to checkpoint inhibition.

*DLBCL*. DLBCL is the most common type of aggressive non-Hodgkin lymphoma and is typically treated with R-CHOP. Approximately 30% to 40% of patients will relapse or be refractory to frontline chemoimmunotherapy, and 20% to 50% of those can be cured with autologous stem cell transplantation if there disease is chemosensitive.<sup>67,68</sup> Outcomes are poor for patients with chemorefractory disease.<sup>69</sup> Recently, chimeric antigen receptor (CAR) T-cell therapy was approved for the treatment of this population with 40% of patients achieving durable remissions.<sup>70</sup> Unfortunately, the majority of patients are not able to receive CAR T-cell due to cost, availability, and/or performance status.

Response rates to single-agent PD-1 blockade in DLBCL have been disappointingly low. Unlike cHL and PMBCL, there is a low incidence (10-27%) of structural variations of PD-L1/PD-L2 in DLBCL.<sup>54,71-73</sup> Interestingly, these genetic alterations are more commonly seen in activated B-cell-like (ABC)/nongerminal center B (GCB)-type DLBCL, a subtype of DLBCL associated with a poor prognosis.<sup>72</sup> Tumors with 9p24.1 amplification had similar genomic profiles to PMBCL.<sup>73</sup> In a cohort of 1253 patients with DLBCL, PD-L1 protein expression was significantly associated with non-GCB subtype and Epstein-Barr virus (EBV) positivity and showed a trend toward inferior overall survival.74 A meta-analysis demonstrated that 27% to 61% of DLBCL had PD-L1 expression, which was significantly associated with poor prognosis.75 Checkpoint blockade in DLBCL has been of great interest; however, the response rate in unselected patients has only been 9% to 36% (Table 3).54,59,62 Given the association between 9p24.1 structural alteration and response to PD-1 inhibition seen in cHL and PMBCL, future studies should focus on DLBCL tumors with these alterations.

Significant PD-L1/PD-L2 gene arrangement has also been observed in extranodal subtypes of DLBCL-primary testicular lymphoma (PTL) and primary central nervous system lymphoma (PCNSL). In one series, 54% of PTL had 9p24.1 CNA and 4% had translocation involving PD-L1, while 52% of EBV-negative PCNSL had 9p24.1 CNA alterations and 6% had translocation involving PD-L2.76 Structural variants of PD-L1 and PD-L2 correlated with surface expression of respective proteins.<sup>76</sup> The surprisingly high rate of PD-L1 and PD-L2 alteration seen on PTL and PCNSL suggest that these subsets of DLBCL may respond favorably to checkpoint inhibitors. In fact, a 5-patient case series of relapsed/refractory PCNSL and PTL who were treated with nivolumab demonstrated a response in all patients, 4 with CR and 1 with partial response.<sup>77</sup> Many of these responses were durable, ongoing at 13 to 17 months follow-up. There are several studies currently underway investigating the role of checkpoint inhibitors in relapsed refractory PTL and PCNSL (NCT02857426, NCT03770416, NCT03798314, and NCT04022980).

*Gray zone lymphoma*. Gray zone lymphoma (GZL) presents with features resembling both cHL and PMBCL. It can often present with mediastinal mass and possess morphological and immunophenotypical features of both lymphomas.<sup>78</sup> Although the optimal treatment strategy is not well established, it is treated similarly to aggressive large B-cell lymphoma.<sup>79</sup>

The frequency of 9p24.1 alteration is not clearly defined in this rare entity; however, appears to be in significant proportion of tumors. One small series of 33 patients reported *JAK2* and *PDL2* alterations in 55% of cases.<sup>78</sup> Whether these patients will respond to checkpoint blockade is unclear. A case series of 3 patients in relapsed/refractory GZL demonstrated complete

STUDY	z	POPULATION	TREATMENT	RESPONSE RATE <sup>a</sup>	PREDICTIVE BIOMARKER	BIOMARKER ASSESSMENT	RESPONSE RATE BY BIOMARKER
Primary mediastina.	I B-cell I	ymphoma (PMBCL)					
KEYNOTE-170 Armand et al <sup>52</sup> Phase II	53	R/R PMBCL	Pembrolizumab 200mg q3wks	ORR 45% CR 13% PFS 5.5 (2.8-12.1)months	9p24.1 alteration by FISH	Amplification, copy gain, polysomy, disomy relative copy loss <sup>b</sup>	ORR ( <i>P</i> =.22): ORR 25% for H score of 0 ORR 42% for H score of 1-99 ORR 64% for Acore of 1-99
					PD-L1 by IHC	Modified H-score <sup>c</sup>	PFS (P=.29): 1-99: HR 0.51 (0.21-1.26; P=.14) P=.011)
KEYNOTE-013 Zinzani et al <sup>s2,53</sup> Phase Ib	21	R/R PMBCL	Pembrolizumab 10 mg/kg q2wks or 200 mg q3wks	ORR 48% CR 33% PFS 10.4 months (3.4-not reached) OS 31.4 months (4.9-not reached)	RN	H	R
CheckMate 436 Zinzani et al <sup>33</sup> Phase II	30	R/R PMBCL	Nivolumab 240 mg and BV 1.8mg/kg q3wks	ORR 73% (54-88%) CR 37% PFS at 6 months 63.5% (42.5-78.6%) OS at 6months 86.3% (67.5-94.6%)	RN	R	R
Diffuse large B-cell	lymphor	ma (DLBCL)					
Ansell et al <sup>54</sup> Phase II	121	R/R DLBCL after ASCT or ineligible for ASCT	Nivolumab 3 mg/kg q2wks	ASCT failed (n= 87): ORR 9%, CR 3% PFS 1.9months, OS 12.2months ASCT ineligible (n=34):	9p24.1 gene alteration by FISH	Amplification, copy gain, polysomy, disomy relative copy loss <sup>b</sup>	One of 3 who achieved CR had high-level amplification, one had normal copy
				PFS 1.4 months, OS 5.8 months	PD-L1 by IHC	Modified H-score <sup></sup>	nave specimen for analysis.
Follicular Lymphom	a (FL)						
Westin et al <sup>55</sup> Phase II	30	Relapsed FL	Rituximab + pidilizumab <sup>d</sup> 3mg/kg q4wks	ORR 66% CR 52% PFS 18.8months (14.7-not reached)	Gene expression by RNA	RNA expression of 41 genes including PD-1 and CXCR5 expression was used to distinguish between effector T-cells vs follicular helper T-cells.	More follicular helper T-cells and fewer effector T-cells within the tumor predicted less tumor shrinkage and shorter PFS (median 12.7 vs not reached (6.5-21.6) months).
							(Continued)

÷

Table 3. (Continued)

STUDY	z	POPULATION	TREATMENT	RESPONSE RATE <sup>a</sup>	PREDICTIVE BIOMARKER	BIOMARKER ASSESSMENT	RESPONSE RATE BY BIOMARKER
Multiple hematolog	iic maligr	ıancies					
Ansell et al <sup>56</sup> Phase I	18	R/R NHL	lpilimumab 3 mg/kg q4wks × 4 cycles	ORR 11% CR 5.5%	NR	NR	RN
Armand et al <sup>57</sup> Phase II	66	DLBCL and PMBCL after ASCT	Pidilizumab <sup>d</sup> 1.5 mg/kg q42d×3 cycles	PFS at 16 0.72 (0.60-0.82) months <sup>e</sup> OS at 16 0.85 (0.74-0.92) months <sup>e</sup>	NR	щ	щ
Davids et al <sup>58</sup> Phase l/lb	28	R/R leukemia, lymphoma, MM, MDS, MPN after allogeneic HSCT	lpilimumab 3 or 10 mg/kg q3wks×4 cycles	3 mg/kg dose (n=6) ORR 0% 10 mg/kg dose (n=22) ORR 32%, CR 23%	Soluble MICA by ELISA	Levels measured by ELISA	Low pretreatment levels of sMICA positively predicted a clinical response ( <i>P</i> =.006).
Lesokhin et al <sup>ss</sup> Phase Ib	81	R/R B-cell lymphoma, TCL, and MM	Nivolumab 1 or 3 mg/kg q2wks	FL (n=10): ORR 40% DLBCL (n=11): ORR 36% TCL (n=23): ORR 17% MM (n=27): ORR 4%	9p24.1 alteration by FISH	Amplification, copy gain, polysomy, disomy relative copy loss <sup>b</sup>	One with mycosis fungoides had high PD-L2, achieved PR. One patient with DLBCL had low level PD-L1/
					PD-L1, PD-L2 by IHC	Percentage, intensity (0-3), and subcellular localization of staining on malignant cells and tumor microenvironment	P.DLz polysoning and rad moderate intensity PD-L1 in 2% of cells and had SD.
Khouri et al <sup>60</sup> Phase II	17	Persistent relapsed lymphoid disease after allogeneic or autologous HSCT	Lenalidomide + ipilimumab 3mg/kg	After Allo HSCT (N = 10) ORR 70%, CR 40% PFS at 12 months 56% OS at 20.5 months 90% After ASCT (N = 7) ORR 86%, CR 86% PFS at 12 months 100% OS at 17 months 100%	Ř	Ř	R
Tuscano et al <sup>61</sup> Phase I	33	R/R NHL ineligible for curative intent therapy	Rituximab + ipilimumab 1 mg/kg or 3mg/kg q3wks×4 cycles then q12wks	ORR 24% CR 6% PFS 2.6 (1.6-4.6) months In FL (n = 13): ORR 54%, CR 15% PFS 5.6 (1.6-18.4) months	щ	щ	щ
							(Continued)

RESPONSE RATE BY BIOMARKER	In DLBCL, ORR was 63% in positive PD-L1 vs ORR was 19% in negative PD-L1 ( $P$ = .065). In DLBCL, all who had CR who had reportable PD-L1 values had positive PD-L1 ( $P$ = .028).	The relative expression of MHCII on PAX5+ tumor cells was increased in postprogression samples ( <i>P</i> = .04). No corresponding increase in LAG3 expression on infiltrating T-cells.	R		NR	NR	fuse large B-cell lymphoma; ELISA, 3, lymphocyte activating 3; MHC, major
BIOMARKER ASSESSMENT	Positive if expression was ≽5% in malignant cells	Protein expression by IHC	Н		NR	NR	nokine receptor 5; DLBCL, dif immunohistochemistry; LAG
PREDICTIVE BIOMARKER	РD-L1 by IHC	MHC class II and LAG3 by IHC	щ		NR	NR	CR5, C-X-C motif chen em cell transplant; IHC, non-Hodokin lympho
RESPONSE RATE <sup>a</sup>	CLL/SLL (n=36): ORR 61%, CR 0% PFS NE, OS not reached FL (n=40): ORR 33%, CR 10% PFS 9.1 (3.1-14.0) months OS not reached DLBCL (n=45): ORR 36%, CR 16% PFS 2.6 (1.9-7.6) months OS 13.5 months (6.5-NE) Richter's (n=20): ORR 65%, CR 10% PFS 5.0 months (2.4-NE) OS 10.3 months (4.8-NE)	PFS at 18.5months 58% (34-76%)	ORR 29% CR 3.6% PFS 3.7 (1.6-8) months OS 21.4 months (4.2-not reached)		ORR 9.1% CR 0%	ORR 12% CR 8%	soytic leukemia; CR, complete response; CX a; HR, hazard ratio; HSCT, hematopoletic st ndiferativa neonlasm: NF not estimation IS
TREATMENT	lbrutinib + nivolumab 3mg/kg q2wks	Pembrolizumab 200mg q3wks×8 cycles	Nivolumab 0.5-3mg/kg q2 weeks		Ipilimumab 3 mg/kg then 1.5mg/kg q4wks × 4 cycles	Ipilimumab 0.1-3 mg/kg dose escalation	entuximab vedotin; CLL, chronic lympho situ hybridization; FL, follicular lymphom s- MM mutrine mvaloma - MPN malona
POPULATION	R/R CLL, SLL, FL, DLBCL	R/R DLBCL and PMBCL after ASCT as maintenance	Relapsed hematologic malignancies after allogeneic HSCT	olid malignancies <sup>f</sup>	Progression after antitumor vaccine treatment	Relapsed malignancy after allogeneic HSCT	em cell transplant; BV, br ay; FISH, fluorescent in-
z	141	50	58	ic and so	4	25	ologous ste orbent assé vr MDS m
STUDY	Younes et al⁰² Phase I/IIa	Frigault et al <sup>63</sup> Phase II	Davids et al <sup>64</sup> Phase I	Multiple hematolog	O'Mahony et al <sup>65</sup> Phase II	Bashey et al <sup>66</sup> Phase I	Abbreviations: ASCT, aut enzyme-linked immunosc histocompatibility complex

BCA confidence interval unless otherwise indented on the invertingent with inverted on the survive receptored with non-hodgkin lymphoma; NH, non-hodgkin lymphoma; SM, soluble major histocompatibility complex class I polypeptide-related sequence A; TCL, T-cell lymphoma.

<sup>b</sup>Polysomy was defined as target: control signal ratio of 1:1 but more than 2 copies per probe, copy gain as signal ratio greater than 1:1 but less than 3:1, and amplification as signal ratio 3:1 or greater. <sup>c</sup>Modified H-score was calculated by multiplying percentage of PD-L1-positive malignant cells (0-100%) and the average intensity of PD-L1 staining (0 to 3+), and ranges from score 0 to 300. <sup>c</sup>Target of pidilizumab is currently unclear.

Number of patients indicated are those with hematologic malignancies, and excluding patients with solid malignancies.

Table 3. (Continued)

and durable responses in all 3 patients.<sup>80</sup> A clinical trial investigating the role of checkpoint inhibitors in GZL is ongoing (NCT03255018).

#### MHC expression

A key step in T-cell activation is antigen presentation on major histocompatibility complex (MHC) molecules. Any disturbance in this pathway may decrease the tumor neoantigen presentation and T-cell activation. Defects in antigen presentation have been proposed as a mechanism of immune evasion in both lymphoma and solid tumors.<sup>81</sup>

Beta-2-microglobulin (B2M), a subunit of MHC class I molecule, plays a crucial role in transport of MHC class I molecules to the cell surface. B2M is mutated in 39% of cHL patients.<sup>40</sup> MHC class I expression is decreased to negative in 79% of cases of HRS cells, and is associated with inferior survival.82 Interestingly, MHC class II expression is also significantly decreased in 67% of malignant cells. However, unlike MHC class I, this was not associated with inferior outcomes. The same group later reported in a post hoc analysis of CheckMate-205,41 92% of tumors that achieved CR had no expression of B2M and MHC class I, suggesting that activation of MHC class I is not critical for a response to PD-1 inhibitor. MHC class II expression in the available samples were negative in 29% of patients, decreased in 32% of patients, and positive in 39% of patients, and interestingly, its expression was predictive of response to anti-PD-1 therapy. Notably, B2M mutation is seen in approximately 30% to 40% of tumors of PMBCL but approximately 10% in DLBCL.48,83 Overall, these findings propose that responses to PD-1 inhibition in cHL may rely on CD4<sup>+</sup> T-cell activation.

In addition to B2M, several other genes involved in antigen presentation are frequently mutated in cHL and PMBCL. Deletion of 6p21.32, the locus for *MHCI* and *MHCII* genes, decreases surface expression of MHC class II, and is recurrently found in both cHL (43%)<sup>40</sup> and PMBCL (32%),<sup>83</sup> while it is only rarely found (12%) in DLBCL.<sup>84</sup> Structural variants of the class II MHC transactivator (CIITA) is a frequent gene fusion partner in lymphoma, and its alteration is found in 15% to 38% of PMBCL, 15% of cHL, and <5% of DLBCL.<sup>48,83,85</sup>

The above findings suggest that MHC class I and/or II downregulation may be a resistance mechanism to PD-1/PD-L1 blockade in cHL. Whether the activity of CD4<sup>+</sup> T-cell or CD8<sup>+</sup> T-cell alone or both are implicated should be further studied. In melanoma, decrease in MHC class I expression by B2M mutation has been shown to be a resistance mechanism to PD-1 therapy.<sup>86</sup> Selected population of patients may require combination approach with PD-1 and CTLA-4 blockade or CAR T-cell therapy that bypass T-cells receptor and MHC interaction.

### TMB

Cancer usually develops due to acquired somatic mutations. High nonsynonymous mutational burden leads to cancer-associated neoantigen presentation that may be more readily recognized by the immune system. High TMB correlates with increased response to PD-1/PD-L1 blockade, especially in melanoma and NSCLC.<sup>10,11,87,88</sup> When analyzing patients who received immunotherapy across different cancer types, 20% of patients with low (1-5 mutations/mb) to intermediate TMB (6-19 mutations/mb) had response to immunotherapy while 58% of patients with high TMB (>20 mutations/mb) had responses.<sup>10</sup> As such, TMB is a useful marker in predicting response to checkpoint inhibitors in solid malignancies.

Unlike solid malignancies, TMB in hematologic malignancies is not clearly defined, only reported in few series. Median TMB of cHL is 7.66 mutations/mb with notably higher TMB in EBV-negative cHL, level similar to NSCLC.<sup>40</sup> Median TMB of PMBCL is 7.0 mutations/mb.<sup>83</sup> Although not characterized by each lymphoma subtype, another study demonstrated that B-cell NHL had higher mutational burden compared to other hematological malignancies (TMB of cHL was not reported).<sup>89</sup> The relationship of TMB in hematologic malignancies to response to checkpoint inhibition has not been established. Studies characterizing the mutational load across hematologic malignancies are urgently warranted.

#### MSI and MMR

MSI-High or dMMR tumors have defects in DNA repair which result in higher mutational load. Similar to tumors with high TMB, there is an increase in cancer-associated neoantigen presentation leading to increased recognition by the T-cells. Unselected tumors with MSI-High or dMMR status have very high response rates to checkpoint inhibitiors<sup>12,13</sup> which subsequently led to the FDA approval of pembrolizumab based on MSI status agnostic of tumor type. MSI and MMR status have not been defined in various hematologic malignancies, but deficient status is seen in approximately 9% in cHL,<sup>40</sup> 8% in PMBCL,<sup>83</sup> and 0.3% to 3.2% in DLBCL.<sup>84,90</sup> Their association with response to checkpoint blockade is currently unknown in lymphoma as no patients with lymphoma were represented in the trials involving MSI-H/dMMR cancers.

#### Conclusion

Although checkpoint inhibitors have changed the treatment paradigm for numerous solid malignancies, their benefit in hematologic malignancies has been limited to cHL and PMBCL. The genetic alterations of 9p24.1 locus containing PD-L1 and PD-L2 gene and antigen presentation pathways is the basis for checkpoint blockade response in cHL and PMBCL. However, there is still significant subset of patients with cHL and PMBCL who do not benefit from checkpoint



Figure 1. Proposed algorithm for anti-PD-1 inhibitor use in (A) cHL and PMBCL, and (B) in DLBCL and GZL. cHL indicates classical Hodgkin lymphoma; DLBCL, diffuse large B-cell lymphoma; dMMR, mismatch repair deficient; GZL, gray zone lymphoma; IHC, immunohistochemistry; MHCII, major histocompatibility complex class II; MSI-H, microsatellite instability high; PD-1, programmed cell death 1; PD-L1, programmed cell death 1 ligand 1; PMBCL, primary mediastinal B-cell lymphoma; R/R, relapsed/refractory; TMB, tumor mutational burden.

blockade. Furthermore, patients with DLBCL and other indolent lymphomas largely do not benefit from PD-1 inhibitors, although a small subset of patients have durable response. It is therefore imperative that appropriate patients are selected based on available biomarkers and to understand resistance mechanisms.

A proposed algorithm for treatment decisions involving PD-1 inhibitors in relapsed or refractory lymphoma is presented in Figure 1. Classical HL or PMBCL with low PD-L1 H-score (although absolute score cutoff is to be determined) may predict a suboptimal response to checkpoint inhibition. Furthermore, an absence of MHC class II expression may be a possible resistance mechanism. However, standardized ways to measure MHC class II expression in the clinic are currently lacking. Combination chemoimmunotherapy or alternative therapies such as CAR T-cell could overcome checkpoint blockade resistance.

Checkpoint inhibitors may be reasonable considerations in NHL with high TMB (particularly non-GCB/ABC subtype of DLBCL), high PD-L1 IHC expression, MSI/dMMR, or 9p24.1 structural alteration. Compared to cHL or PMBCL, NHLs exhibit lower PD-L1 surface expression and preliminary evidence of low TMB. Gene expression profiling has demonstrated that the majority of NHLs lack an inflamed gene signature, a characteristic that may be needed for successful checkpoint therapy.<sup>91</sup> Investigating these mechanisms will help explore ways to overcome resistance.

There are several areas of future interest. Structural alterations in 9p24.1, TMB, MSI/MMR status are not characterized for other types of NHLs or myeloid malignancies, and absolute

cut-off value of the PD-L1 H-score should be established and validated in different lymphoma subtypes in prospective studies. Promising data suggest checkpoint blockade is efficacious for relapse postallogeneic stem cell transplantation.<sup>58,66</sup> However, as Davids and colleagues demonstrated, increased risk of immune-related adverse events and fatal graft versus host disease is a serious concern in this setting and biomarkerbased approaches may help better select appropriate patients.<sup>64</sup> The effect of PD-1/PD-L1 inhibition is also thought to synergize with CAR-T therapy by decreasing T-cell exhaustion. However data are limited to case series which report significant off-target immune effects.92 Alternatively, preclinical data suggest that CAR-T cells engineered to secrete PD-1 blocking single change variable fragments may provide equal or increased efficacy but decreased toxicity.93 Finally, checkpoint blockade may sensitize lymphoma to subsequent therapy. Carreau and colleagues showed that the overall response to subsequent therapy after checkpoint inhibition was 62% in 81 patients with relapsed refractory Hodgkin lymphoma94 and 51% in 59 patients with relapsed refractory non-Hodgkin lymphoma.95

With further studies, we are hopeful that biomarkers including PD-L1 expression, TMB, and MSI status will be used to best-select patients with hematologic malignancies for treatment with checkpoint blockade who are most likely to benefit.

### **Author Contributions**

A.J and A.M.G drafted the manuscript. E.D.B and A.M.G reviewed and edited the manuscript. All authors approve the final manuscript.

#### **ORCID** iD

Aaron Michael Goodman D https://orcid.org/0000-0002 -6196-9427

#### REFERENCES

- Rowshanravan B, Halliday N, Sansom DM. CTLA-4: a moving target in immunotherapy. *Blood.* 2018;131:58-67.
- Boussiotis VA. Molecular and biochemical aspects of the PD-1 checkpoint pathway. N Engl J Med. 2016;375:1767-1778.
- Postow MA, Chesney J, Pavlick AC, et al. Nivolumab and ipilimumab versus ipilimumab in untreated melanoma. N Engl J Med. 2015;372:2006-2017.
- Wolchok JD, Kluger H, Callahan MK, et al. Nivolumab plus ipilimumab in advanced melanoma. N Engl J Med. 2013;369:122-133.
- Larkin J, Chiarion-Sileni V, Gonzalez R, et al. Five-year survival with combined nivolumab and ipilimumab in advanced melanoma. N Engl J Med. 2019;381: 1535-1546.
- Kato S, Goodman A, Walavalkar V, Barkauskas DA, Sharabi A, Kurzrock R. Hyperprogressors after immunotherapy: analysis of genomic alterations associated with accelerated growth rate. *Clin Cancer Res.* 2017;23:4242-4250.
- Borghaei H, Paz-Ares L, Horn L, et al. Nivolumab versus docetaxel in advanced nonsquamous non-small-cell lung cancer. NEngl J Med. 2015;373:1627-1639.
- Rosenberg JE, Hoffman-Censits J, Powles T, et al. Atezolizumab in patients with locally advanced and metastatic urothelial carcinoma who have progressed following treatment with platinum-based chemotherapy: a single-arm, multicentre, phase 2 trial. *Lancet*. 2016;387:1909-1920.
- Goodman AM, Piccioni D, Kato S, et al. Prevalence of PDL1 amplification and preliminary response to immune checkpoint blockade in solid tumors. JAMA Oncol. 2018;4:1237-1244.
- Goodman AM, Kato S, Bazhenova L, et al. Tumor mutational burden as an independent predictor of response to immunotherapy in diverse cancers. *Mol Cancer Ther.* 2017;16:2598-2608.
- 11. Yarchoan M, Hopkins A, Jaffee EM. Tumor mutational burden and response rate to PD-1 inhibition. *N Engl J Med.* 2017;377:2500-2501.
- 12. Le DT, Durham JN, Smith KN, et al. Mismatch repair deficiency predicts response of solid tumors to PD-1 blockade. *Science*. 2017;357:409-413.
- Le DT, Uram JN, Wang H, et al. PD-1 blockade in tumors with mismatchrepair deficiency. N Engl J Med. 2015;372:2509-2520.
- FDA Grants Accelerated Approval to Pembrolizumab for First Tissue/Site Agnostic Indication. Silver Spring, MD: Food and Drug Administration; 2017.
- 15. Cristescu R, Mogg R, Ayers M, et al. Pan-tumor genomic biomarkers for PD-1 checkpoint blockade-based immunotherapy. *Science*. 2018;362:eaar3593.
- Ettinger DS, Aisner DL, Wood DE, et al. NCCN guidelines insights: nonsmall cell lung cancer, version 5.2018. J Natl Compr Canc Netw. 2018;16: 807-821.
- Ott PA, Bang YJ, Piha-Paul SA, et al. T-cell-inflamed gene-expression profile, programmed death ligand 1 expression, and tumor mutational burden predict efficacy in patients treated with pembrolizumab across 20 cancers: KEY-NOTE-028. J Clin Oncol. 2019;37:318-327.
- Giraldo NA, Nguyen P, Engle EL, et al. Multidimensional, quantitative assessment of PD-1/PD-L1 expression in patients with Merkel cell carcinoma and association with response to pembrolizumab. *J Immunother Cancer.* 2018;6:99.
- Badalamenti G, Fanale D, Incorvaia L, et al. Role of tumor-infiltrating lymphocytes in patients with solid tumors: can a drop dig a stone? *Cell Immunol*. 2019;343:103753.
- Linette GP, Carreno BM. Tumor-infiltrating lymphocytes in the checkpoint inhibitor era. Curr Hematol Malig Rep. 2019;14:286-291.
- Zboralski D, Hoehlig K, Eulberg D, Fromming A, Vater A. Increasing tumorinfiltrating T cells through inhibition of CXCL12 with NOX-A12 synergizes with PD-1 blockade. *Cancer Immunol Res.* 2017;5:950-956.
- Ansell S, Armand P, Timmerman JM, et al. Nivolumab in patients (Pts) with relapsed or refractory classical Hodgkin lymphoma (R/R cHL): clinical outcomes from extended follow-up of a phase 1 study (CA209-039). *Blood.* 2015;126:583-583.
- Ansell SM, Lesokhin AM, Borrello I, et al. PD-1 blockade with nivolumab in relapsed or refractory Hodgkin's lymphoma. NEngl J Med. 2015;372:311-319.
- Armand P, Chen YB, Redd RA, et al. PD-1 blockade with pembrolizumab for classical Hodgkin lymphoma after autologous stem cell transplantation. *Blood*. 2019;134:22-29.
- Armand P, Engert A, Younes A, et al. Nivolumab for relapsed/refractory classic Hodgkin lymphoma after failure of autologous hematopoietic cell transplantation: extended follow-up of the multicohort single-arm phase II CheckMate 205 trial. J Clin Oncol. 2018;36:1428-1439.

- Armand P, Shipp MA, Ribrag V, et al. Programmed death-1 blockade with pembrolizumab in patients with classical Hodgkin lymphoma after brentuximab vedotin failure. J Clin Oncol. 2016;34:3733-3739.
- Chen R, Zinzani PL, Fanale MA, et al. Phase II study of the efficacy and safety of pembrolizumab for relapsed/refractory classic Hodgkin lymphoma. J Clin Oncol. 2017;35:2125-2132.
- Chen R, Zinzani PL, Lee HJ, et al. Pembrolizumab in relapsed or refractory Hodgkin lymphoma: 2-year follow-up of KEYNOTE-087. *Blood.* 2019;134: 1144-1153.
- Herrera AF, Moskowitz AJ, Bartlett NL, et al. Interim results of brentuximab vedotin in combination with nivolumab in patients with relapsed or refractory Hodgkin lymphoma. *Blood.* 2018;131:1183-1194.
- Maruyama D, Hatake K, Kinoshita T, et al. Multicenter phase II study of nivolumab in Japanese patients with relapsed or refractory classical Hodgkin lymphoma. *Cancer Sci.* 2017;108:1007-1012.
- Shi Y, Su H, Song Y, et al. Safety and activity of sintilimab in patients with relapsed or refractory classical Hodgkin lymphoma (ORIENT-1): a multicentre, single-arm, phase 2 trial. *Lancet Haematol.* 2019;6:e12-e19.
- Younes A, Santoro A, Shipp M, et al. Nivolumab for classical Hodgkin's lymphoma after failure of both autologous stem-cell transplantation and brentuximab vedotin: a multicentre, multicohort, single-arm phase 2 trial. *Lancet Oncol.* 2016;17:1283-1294.
- Zinzani PL, Santoro A, Gritti G, et al. Nivolumab combined with brentuximab vedotin for relapsed/refractory primary mediastinal large B-cell lymphoma: efficacy and safety from the phase II CheckMate 436 study. J Clin Oncol. 2019;37:3081-3089.
- Ramchandren R, Domingo-Domenech E, Rueda A, et al. Nivolumab for newly diagnosed advanced-stage classic Hodgkin lymphoma: safety and efficacy in the phase II CheckMate 205 study. J Clin Oncol. 2019;37:1997-2007.
- Song Y, Gao Q, Zhang H, et al. Treatment of relapsed or refractory classical Hodgkin lymphoma with the anti-PD-1, tislelizumab: results of a phase 2, single-arm, multicenter study. *Leukemia*. 2020;34:533-542.
- Song Y, Wu J, Chen X, et al. A single-arm, multicenter, phase ii study of camrelizumab in relapsed or refractory classical Hodgkin lymphoma. *Clin Cancer Res.* 2019;25:7363-7369.
- Stephens DM, Li H, Schoder H, et al. Five-year follow-up of SWOG S0816: limitations and values of a PET-adapted approach with stage III/IV Hodgkin lymphoma. *Blood.* 2019;134:1238-1246.
- Mehta-Shah N, Bartlett NL. Management of relapsed/refractory classical Hodgkin lymphoma in transplant-ineligible patients. *Blood.* 2018;131:1698-1703.
- Greaves P, Clear A, Owen A, et al. Defining characteristics of classical Hodgkin lymphoma microenvironment T-helper cells. *Blood*. 2013;122:2856-2863.
- Wienand K, Chapuy B, Stewart C, et al. Genomic analyses of flow-sorted Hodgkin Reed-Sternberg cells reveal complementary mechanisms of immune evasion. *Blood Adv.* 2019;3:4065-4080.
- Roemer MGM, Redd RA, Cader FZ, et al. Major histocompatibility complex class II and programmed death ligand 1 expression predict outcome after programmed death 1 blockade in classic Hodgkin lymphoma. J Clin Oncol. 2018;36:942-950.
- Sakamuri D, Glitza IC, Betancourt Cuellar SL, et al. Phase I dose-escalation study of anti-CTLA-4 antibody ipilimumab and lenalidomide in patients with advanced cancers. *Mol Cancer Ther.* 2018;17:671-676.
- Armand P, Lesokhin A, Borrello I, et al. A phase 1b study of dual PD-1 and CTLA-4 or KIR blockade in patients with relapsed/refractory lymphoid malignancies [published online ahead of print June 29, 2020]. *Leukemia*. doi:10.1038/ s41375-020-0939-1.
- 44. Diefenbach CS, Hong F, Ambinder RF, et al. Ipilimumab, nivolumab, and brentuximab vedotin combination therapies in patients with relapsed or refractory Hodgkin lymphoma: phase 1 results of an open-label, multicentre, phase 1/2 trial. *Lancet Haematol.* 2020;7:e660-e670.
- Roemer MG, Advani RH, Ligon AH, et al. PD-L1 and PD-L2 genetic alterations define classical Hodgkin lymphoma and predict outcome. *J Clin Oncol.* 2016;34:2690-2697.
- 46. Green MR, Monti S, Rodig SJ, et al. Integrative analysis reveals selective 9p24.1 amplification, increased PD-1 ligand expression, and further induction via JAK2 in nodular sclerosing Hodgkin lymphoma and primary mediastinal large B-cell lymphoma. *Blood*. 2010;116:3268-3277.
- Takyar J, Raut M, Borse R, Balakumaran A, Sehgal M. Relapsed/refractory primary mediastinal large B-cell lymphoma: a structured review of epidemiology, treatment guidelines and real-world treatment practices. *Expert Rev Hematol.* 2020;13:275-287.
- Mottok A, Hung SS, Chavez EA, et al. Integrative genomic analysis identifies key pathogenic mechanisms in primary mediastinal large B-cell lymphoma. *Blood.* 2019;134:802-813.
- Shi M, Roemer MG, Chapuy B, et al. Expression of programmed cell death 1 ligand 2 (PD-L2) is a distinguishing feature of primary mediastinal (thymic)

large B-cell lymphoma and associated with PDCD1LG2 copy gain. Am J Surg Pathol. 2014;38:1715-1723.

- Panjwani PK, Charu V, DeLisser M, Molina-Kirsch H, Natkunam Y, Zhao S. Programmed death-1 ligands PD-L1 and PD-L2 show distinctive and restricted patterns of expression in lymphoma subtypes. *Hum Pathol.* 2018;71:91-99.
- Twa DD, Chan FC, Ben-Neriah S, et al. Genomic rearrangements involving programmed death ligands are recurrent in primary mediastinal large B-cell lymphoma. *Blood.* 2014;123:2062-2065.
- Armand P, Rodig S, Melnichenko V, et al. Pembrolizumab in relapsed or refractory primary mediastinal large B-cell lymphoma. J Clin Oncol. 2019:37: 3291-3299.
- Zinzani PL, Ribrag V, Moskowitz CH, et al. Safety and tolerability of pembrolizumab in patients with relapsed/refractory primary mediastinal large B-cell lymphoma. *Blood.* 2017;130:267-270.
- Ansell SM, Minnema MC, Johnson P, et al. Nivolumab for relapsed/refractory diffuse large B-cell lymphoma in patients ineligible for or having failed autologous transplantation: a single-arm, phase II study. J Clin Oncol. 2019;37: 481-489.
- Westin JR, Chu F, Zhang M, et al. Safety and activity of PD1 blockade by pidilizumab in combination with rituximab in patients with relapsed follicular lymphoma: a single group, open-label, phase 2 trial. *Lancet Oncol.* 2014;15:69-77.
- Ansell SM, Hurvitz SA, Koenig PA, et al. Phase I study of ipilimumab, an anti-CTLA-4 monoclonal antibody, in patients with relapsed and refractory B-cell non-Hodgkin lymphoma. *Clin Cancer Res.* 2009;15:6446-6453.
- 57. Armand P, Nagler A, Weller EA, et al. Disabling immune tolerance by programmed death-1 blockade with pidilizumab after autologous hematopoietic stem-cell transplantation for diffuse large B-cell lymphoma: results of an international phase II trial. *J Clin Oncol.* 2013;31:4199-4206.
- Davids MS, Kim HT, Bachireddy P, et al. Ipilimumab for patients with relapse after allogeneic transplantation. N Engl J Med. 2016;375:143-153.
- Lesokhin AM, Ansell SM, Armand P, et al. Nivolumab in patients with relapsed or refractory hematologic malignancy: preliminary results of a phase Ib study. J Clin Oncol. 2016;34:2698-2704.
- Khouri IF, Fernandez Curbelo I, Turturro F, et al. Ipilimumab plus lenalidomide after allogeneic and autologous stem cell transplantation for patients with lymphoid malignancies. *Clin Cancer Res.* 2018;24:1011-1018.
- Tuscano JM, Maverakis E, Groshen S, et al. A phase I study of the combination of rituximab and ipilimumab in patients with relapsed/refractory B-cell lymphoma. *Clin Cancer Res.* 2019;25:7004-7013.
- Younes A, Brody J, Carpio C, et al. Safety and activity of ibrutinib in combination with nivolumab in patients with relapsed non-Hodgkin lymphoma or chronic lymphocytic leukaemia: a phase 1/2a study. *Lancet Haematol.* 2019; 6:e67-e78.
- Frigault MJ, Armand P, Redd RA, et al. PD-1 blockade for diffuse large B-cell lymphoma after autologous stem cell transplantation. *Blood Adv.* 2020; 4:122-126.
- Davids MS, Kim HT, Costello C, et al. A multicenter phase 1 study of nivolumab for relapsed hematologic malignancies after allogeneic transplantation. *Blood*. 2020;135:2182-2191.
- O'Mahony D, Morris JC, Quinn C, et al. A pilot study of CTLA-4 blockade after cancer vaccine failure in patients with advanced malignancy. *Clin Cancer Res.* 2007;13:958-964.
- Bashey A, Medina B, Corringham S, et al. CTLA4 blockade with ipilimumab to treat relapse of malignancy after allogeneic hematopoietic cell transplantation. *Blood.* 2009;113:1581-1588.
- Gisselbrecht C, Glass B, Mounier N, et al. Salvage regimens with autologous transplantation for relapsed large B-cell lymphoma in the rituximab era. J Clin Oncol. 2010;28:4184-4190.
- Philip T, Guglielmi C, Hagenbeek A, et al. Autologous bone marrow transplantation as compared with salvage chemotherapy in relapses of chemotherapy-sensitive non-Hodgkin's lymphoma. *N Engl J Med.* 1995;333:1540-1545.
- Crump M, Neelapu SS, Farooq U, et al. Outcomes in refractory diffuse large B-cell lymphoma: results from the international SCHOLAR-1 study. *Blood.* 2017;130:1800-1808.
- Neelapu SS, Locke FL, Bartlett NL, et al. Axicabtagene ciloleucel CAR T-cell therapy in refractory large B-cell lymphoma. N Engl J Med. 2017;377:2531-2544.

- Godfrey J, Tumuluru S, Bao R, et al. PD-L1 gene alterations identify a subset of diffuse large B-cell lymphoma harboring a T-cell-inflamed phenotype. *Blood*. 2019;133:2279-2290.
- Georgiou K, Chen L, Berglund M, et al. Genetic basis of PD-L1 overexpression in diffuse large B-cell lymphomas. *Blood.* 2016;127:3026-3034.
- Wang Y, Wenzl K, Manske MK, et al. Amplification of 9p24.1 in diffuse large B-cell lymphoma identifies a unique subset of cases that resemble primary mediastinal large B-cell lymphoma. *Blood Cancer J.* 2019;9:73.
- Kiyasu J, Miyoshi H, Hirata A, et al. Expression of programmed cell death ligand 1 is associated with poor overall survival in patients with diffuse large B-cell lymphoma. *Blood*. 2015;126:2193-2201.
- Zeng Q, Liu Z, Liu T. Prognostic value and clinicopathological characteristics of PD-L1 overexpression in non-Hodgkin lymphoma: a meta-analysis. *BMC Cancer.* 2020;20:59.
- Chapuy B, Roemer MG, Stewart C, et al. Targetable genetic features of primary testicular and primary central nervous system lymphomas. *Blood*. 2016;127: 869-881.
- Nayak L, Iwamoto FM, LaCasce A, et al. PD-1 blockade with nivolumab in relapsed/refractory primary central nervous system and testicular lymphoma. *Blood.* 2017;129:3071-3073.
- Eberle FC, Salaverria I, Steidl C, et al. Gray zone lymphoma: chromosomal aberrations with immunophenotypic and clinical correlations. *Mod Pathol.* 2011;24:1586-1597.
- Kritharis A, Pilichowska M, Evens AM. How I manage patients with grey zone lymphoma. Br J Haematol. 2016;174:345-350.
- Melani C, Major A, Schowinsky J, et al. PD-1 blockade in mediastinal grayzone lymphoma. N Engl J Med. 2017;377:89-91.
- McGranahan N, Rosenthal R, Hiley CT, et al. Allele-specific HLA loss and immune escape in lung cancer evolution. *Cell*. 2017;171:1259-1271.e1211.
- Roemer MG, Advani RH, Redd RA, et al. Classical Hodgkin lymphoma with reduced beta2M/MHC class I expression is associated with inferior outcome independent of 9p24.1 Status. *Cancer Immunol Res.* 2016;4:910-916.
- Chapuy B, Stewart C, Dunford AJ, et al. Genomic analyses of PMBL reveal new drivers and mechanisms of sensitivity to PD-1 blockade. *Blood.* 2019;134: 2369-2382.
- Chapuy B, Stewart C, Dunford AJ, et al. Molecular subtypes of diffuse large B cell lymphoma are associated with distinct pathogenic mechanisms and outcomes. *Nat Med.* 2018;24:679-690.
- Steidl C, Shah SP, Woolcock BW, et al. MHC class II transactivator CIITA is a recurrent gene fusion partner in lymphoid cancers. *Nature*. 2011;471:377-381.
- Zaretsky JM, Garcia-Diaz A, Shin DS, et al. Mutations associated with acquired resistance to PD-1 blockade in melanoma. *N Engl J Med.* 2016;375:819-829.
- Rizvi NA, Hellmann MD, Snyder A, et al. Cancer immunology. Mutational landscape determines sensitivity to PD-1 blockade in non-small cell lung cancer. *Science*. 2015;348:124-128.
- Alexandrov LB, Nik-Zainal S, Wedge DC, et al. Signatures of mutational processes in human cancer. *Nature*. 2013;500:415-421.
- Galanina N, Bejar R, Choi M, et al. Comprehensive genomic profiling reveals diverse but actionable molecular portfolios across hematologic malignancies: implications for next generation clinical trials. *Cancers (Basel)*. 2018;11:11.
- Tian T, Li J, Xue T, Yu B, Li X, Zhou X. Microsatellite instability and its associations with the clinicopathologic characteristics of diffuse large B-cell lymphoma. *Cancer Med.* 2020;9:2330-2342.
- 91. Kline J, Godfrey J, Ansell SM. The immune landscape and response to immune checkpoint blockade therapy in lymphoma. *Blood*. 2020;135:523-533.
- 92. Kambhampati S, Gray L, Fakhri B, et al. Immune-related adverse events associated with checkpoint inhibition in the setting of CAR T cell therapy: a case series. *Clin Lymphoma Myeloma Leuk*. 2020;20:e118-e123.
- Rafiq S, Yeku OO, Jackson HJ, et al. Targeted delivery of a PD-1-blocking scFv by CAR-T cells enhances anti-tumor efficacy in vivo. *Nat Biotechnol.* 2018; 36:847-856.
- Carreau NA, Pail O, Armand P, et al. Checkpoint blockade treatment may sensitize Hodgkin lymphoma to subsequent therapy. *Oncologist.* 2020;25:878.
- Carreau NA, Armand P, Merryman RW, et al. Checkpoint blockade treatment sensitises relapsed/refractory non-Hodgkin lymphoma to subsequent therapy. *Br J Haematol.* 2020;191:44-51.